

Supporting material

Figures

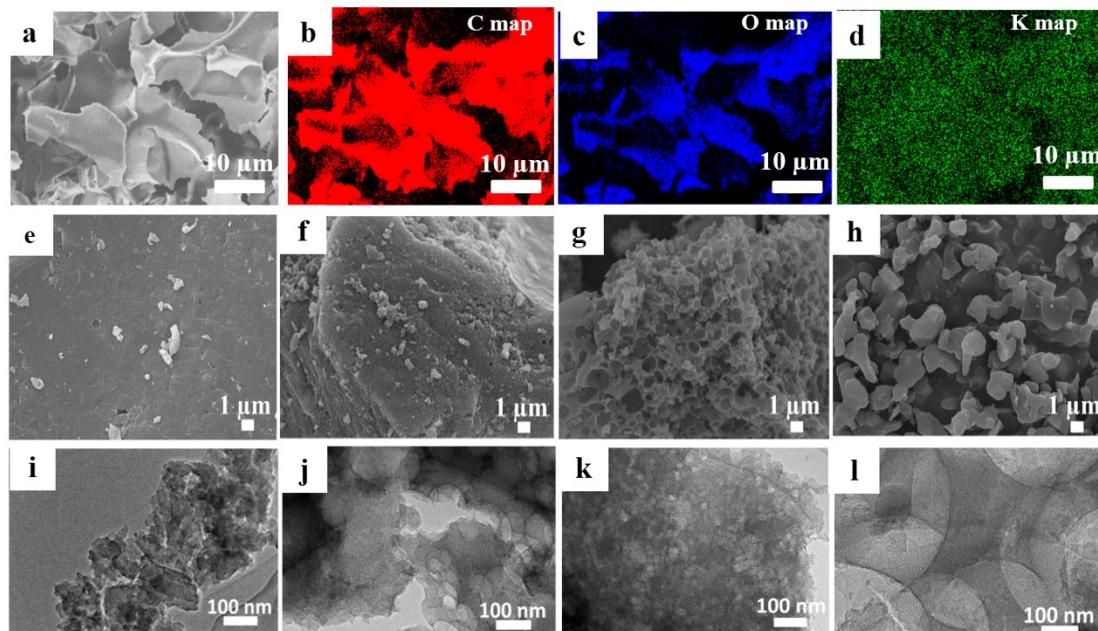


Figure S1. (a) SEM image and EDS (b) C, (c) O, and (d) K mapping of aT . Typical SEM images of (e) TC, (f) aTC-0.2, (g) aTC-0.4, and (h) aTC-0.8. Typical TEM images of (i) TC, (j) aTC-0.2, (k) aTC-0.4, and (l) aTC-0.8.

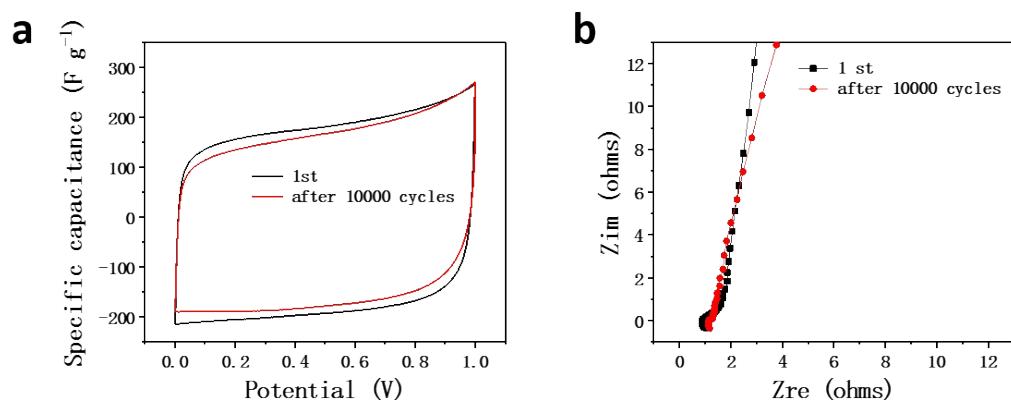


Figure S2. Electrochemical behavior of aTC-0.6 before and after 10000 cycles in 1 M H₂SO₄. (a) CV curves at 100 mV s⁻¹ and (b) EIS.

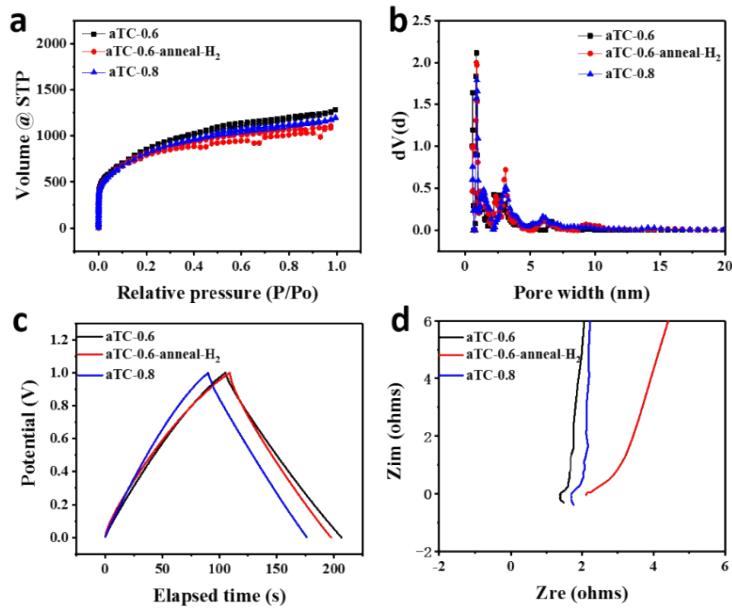


Figure S3. (a) Nitrogen adsorption-desorption isotherms of aTC-0.6, aTC-0.8 and aTC-0.6-anneal-H₂. (b) Pore size distribution of aTC-0.6, aTC-0.8 and aTC-0.6-anneal-H₂. (c) GCD plots of aTC-0.6, aTC-0.8 and aTC-0.6-anneal-H₂. (d) Nyquist plots of aTC-0.6, aTC-0.8 and aTC-0.6-anneal-H₂.

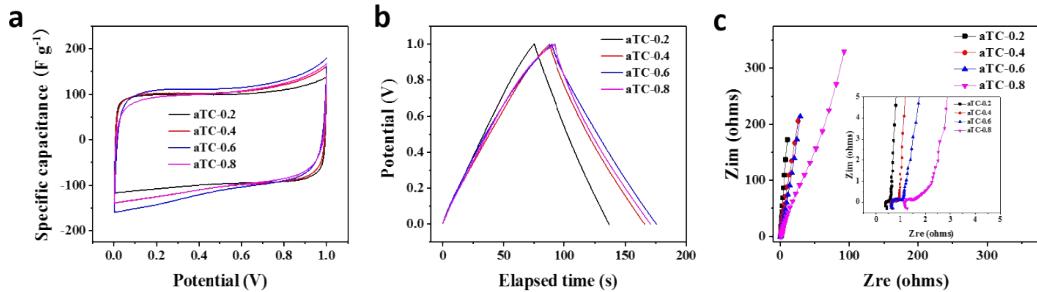


Figure S4. (a) CV curves of aTC at a scan rate of 100 mV s⁻¹ in 6 M KOH. (b) GCD curves of aTCa at a current density of 1 A g⁻¹ in 6 M KOH. (c) Nyquist plots of aTCa in 6 M KOH.

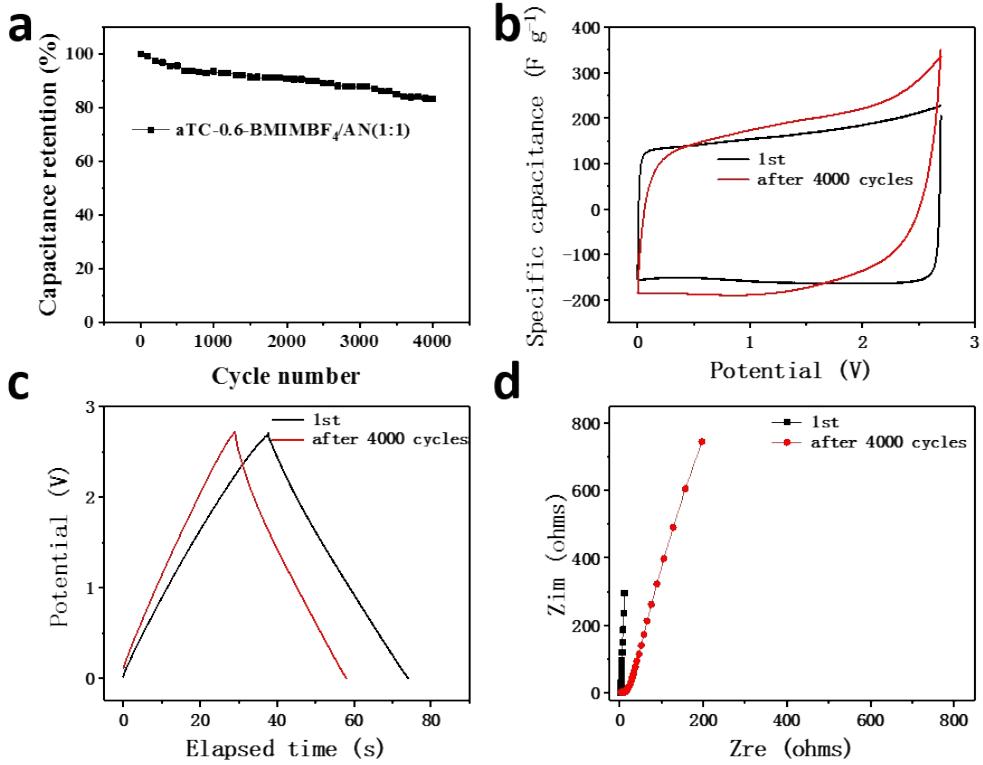


Figure S5. (a) Cycling stability of aTC-0.6 at 5 A g⁻¹ in BMIMBF₄/AN. (b) CV curves of aTC-0.6 before and after 4000 cycles. (c) GCD curves of aTC-0.6 before and after 4000 cycles. (d) Nyquist plots of aTC-0.6 before and after 4000 cycles. The inset shows the details in low-frequency region.

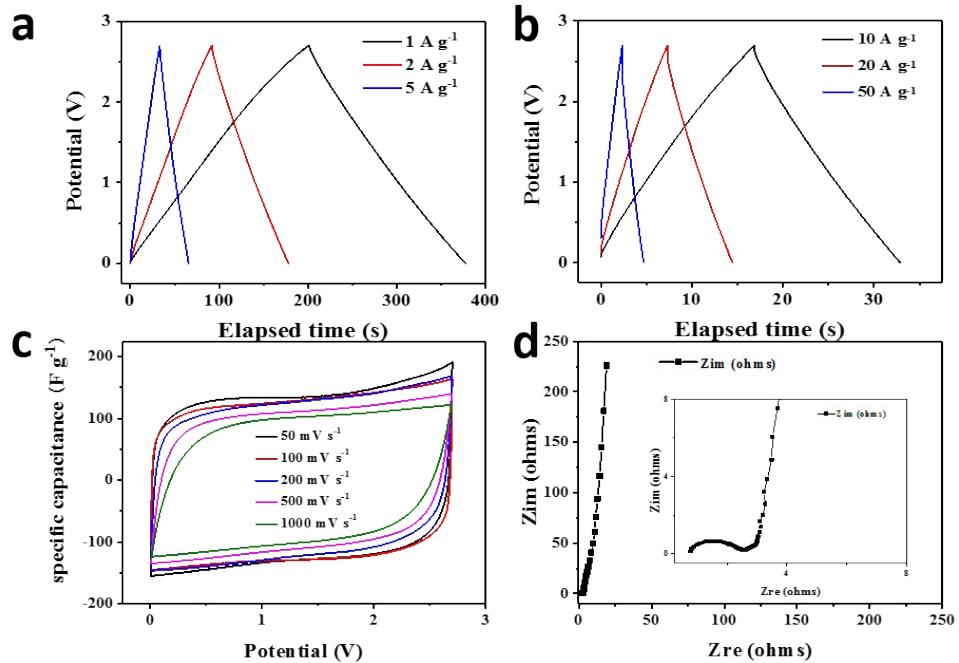


Figure S6. (a, b) GCD curves of aTC-0.6 at different current densities in 1 M TEABF₄/AN. (c) CV curves of aTC-0.6 at various scan rates in 1 M TEABF₄/AN. (d) Nyquist plots of aTC-0.6 in 1 M TEABF₄/AN.

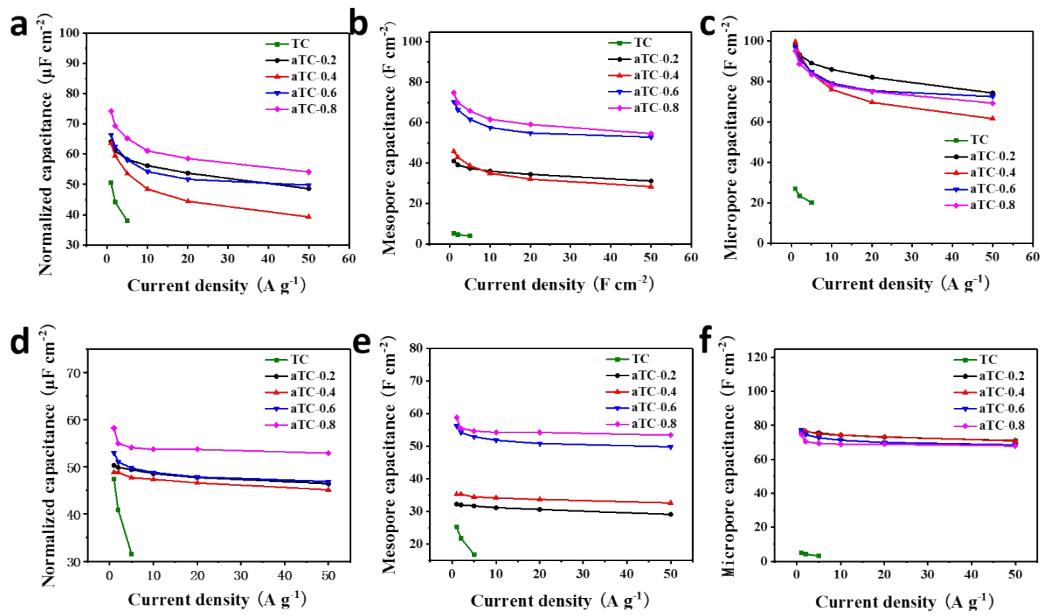


Figure S7. (a) Surface-area-normalized capacitance in 1 M H_2SO_4 . (b) Capacitance attributed to the fraction of microspores in 1 M H_2SO_4 . (c) Capacitance attributed to the fraction of mesopore in 1 M H_2SO_4 . (d) Surface-area-normalized capacitance in 1 M BMIMBF₄/AN. (e) Capacitance attributed to the fraction of microspores in 1 M BMIMBF₄/AN. (f) Capacitance attributed to the fraction of mesopore in 1 M BMIMBF₄/AN.

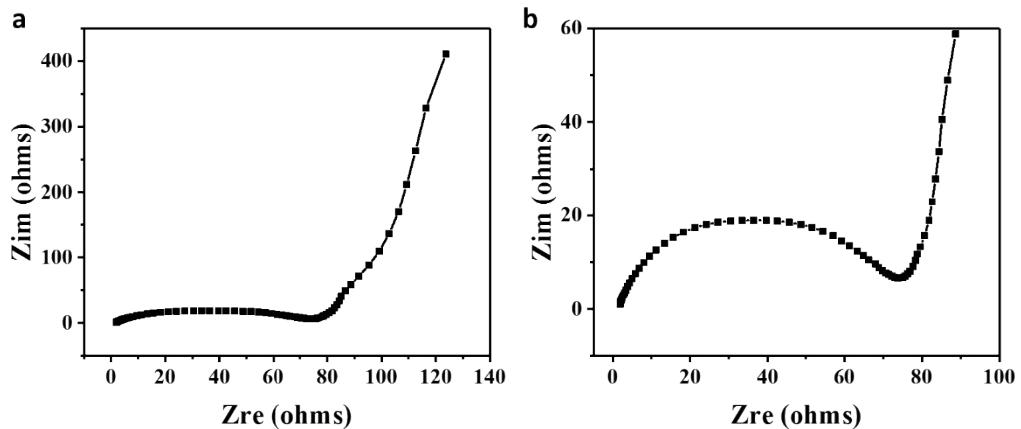


Figure S8. Nyquist plots of aTC-0.6 as LIB anode.

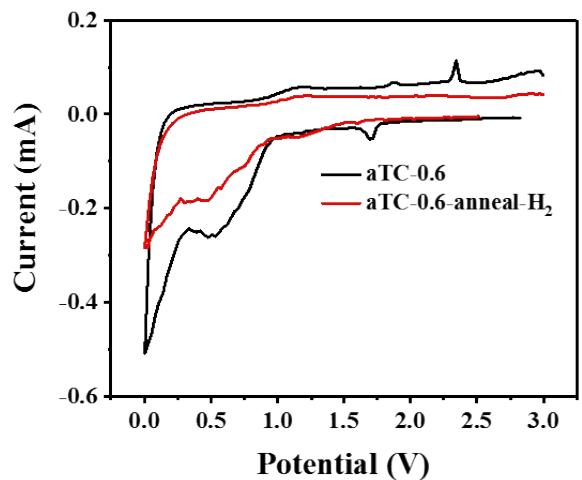


Figure S9. CV curves at 0.1 mV s^{-1} for aTC-0.6 and aTC-0.6-anneal-H₂ in 1 M LiPF₆/EC/DMC electrolyte.

Tables

Table S1. Elemental composition and porosity of aTC-0.6, aTC-0.8 and aTC-0.6-anneal-H₂.

Samples	aT	aT	aTC-0.6-
C (at.%)	95	95.	96.76%
N (at.%)	0.8	0.6	0.46%
O (at.%)	4.1	3.6	2.79%
S BET ^a (m ²)	31	24	2441
V total ^b	1.8	1.6	1.56

Table S2. Weight, thickness, diameter of supercapacitor electrodes tested in different electrolytes.

Sample	2-Electrode Weight (mg)	Electrode Diameter (mm)	Electrode Thickness (μm)	Electrode Density (g cm ⁻³)	Electrolyte
TC	1.77+1.74	6	61	1.02	1M H ₂ SO ₄
aTC-0.2	1.36+1.39	8	53	0.52	1M H ₂ SO ₄
	1.53+1.54	8	53	0.57	BMIM BF ₄ /AN (1:1)
aTC-0.4	1.42+1.43	8	63	0.45	1M H ₂ SO ₄
	1.0+1.0	7	60	0.43	BMIM BF ₄ /AN (1:1)
aTC-0.6	1.20+1.18	8	60	0.39	1M H ₂ SO ₄
	1.03+1.07	8	53	0.39	BMIM BF ₄ /AN (1:1)
aTC-0.8	1.04+1.04	8	53	0.37	1 M TEABF ₄ /AN
	1.14+1.18	8	60	0.38	1M H ₂ SO ₄
	0.88+0.88	8	52	0.34	BMIM BF ₄ /AN (1:1)

Table S3. Comparison of the supercapacitor performance of different carbon materials derived from biomass that have been published to the results in our work.

Carbon Sources	Activation Methods	Test	Electrolyte	Gravimetric capacitance (F g ⁻¹)	Ref.
Soybean curd	Thermal	2-electrode	0.5 M Na ₂ SO ₄	166 at 0.25 A g ⁻¹	1
		3-electrode	2 M KOH	215 at 0.5 A g ⁻¹	
Soybean root	KOH	2-electrode	6 M KOH	268 at 0.5 A g ⁻¹	2
			EMIMBF ₄	239 at 1 A g ⁻¹	
Bagasse waste	KOH	2-electrode	1 M Na ₂ SO ₄	44 at 0.2 A g ⁻¹	3
Corn cob	Thermal-chemical	2-electrode	0.5 M H ₂ SO ₄	210 at 0.5 A g ⁻¹	4
Shrimp shell	Template-chemical	2-electrode	6 M KOH	201 at 1 A g ⁻¹	5
Soya	NaOH	2-electrode	1 M H ₂ SO ₄	155 at 1 A g ⁻¹	6

Water hyacinth	KOH	2-electrode	1 M H ₂ SO ₄	53 at 0.1 A g ⁻¹	7
Wild jujube pit	KOH	2-electrode	6 M KOH	260 at 0.5 A g ⁻¹	8
Moringa oleifera stem	ZnCl ₂	2-electrode	1 M H ₂ SO ₄	280 at 0.1 A g ⁻¹	9
Polytetrafluoroethylene	Thermal-template	2-electrode	6 M KOH	201 at 0.02 A g ⁻¹	10
Cattle bones	Thermal	2-electrode	EMIMBF4	258 at 5 A g ⁻¹	11
Eucalyptus tree leaves	KHCO ₃	3-electrode 2-electrode	1 M H ₂ SO ₄ 1 M LiClO ₄ /PPC	372 at 0.5 A g ⁻¹ 71 at 2 A g ⁻¹	12
Water purifier	KOH	2-electrode	1 M H ₂ SO ₄	122.8 at 1 A g ⁻¹	13
Sorghum vinasse	KOH	2-electrode	1M TEABF ₄ /AN	164 at 1 A g ⁻¹	14
Pine tree sawdust	KOH	2-electrode	1M TEABF ₄ /AN	146 at 0.1 A g ⁻¹	15
Rice straw	KOH	2-electrode	1 M H ₂ SO ₄ 1M EMIMBF ₄ /AN	156 at 0.5 A g ⁻¹ 80 at 0.1 A g ⁻¹	16
Melia azedarach stone	KOH	2-electrode	1 M H ₂ SO ₄	232 at 1 A g ⁻¹	17
carboxymethylcellulose sodium	KNO ₃	2-electrode	1M TEABF ₄ /AN	149 at 1 A g ⁻¹	18
Corn straw	Hydrothermal and chemical activation	2-electrode	6 M KOH 1M TEABF ₄ /AN	222 at 1 A g ⁻¹ 202 at 1 A g ⁻¹	19
Bacterial cellulose	NH ₄ H ₂ PO ₄	2-electrode	EMIMBF ₄ 1 M H ₂ SO ₄	188 at 1 A g ⁻¹ 204.9 at 1 A g ⁻¹	20
Corncob	KOH	2-electrode	1 M H ₂ SO ₄	164 at 1 A g ⁻¹	21
Tofu	KOH	2-electrode	1 M H ₂ SO ₄ BMIMBF ₄ /AN (1:1) 1 M TEABF ₄ /AN	243 at 0.1 A g ⁻¹ 170 at 1 A g ⁻¹ 130 at 1 A g ⁻¹	Our work

Table S4. Comparison of the LIB performance of different carbon materials derived from biomass that have been published to the results in our work.

Carbon Sources	Activation Methods	Current/Capacity		Ref.
human hair derived carbon	KOH	50 mA g ⁻¹	~700 mA h g ⁻¹	
		100 mA g ⁻¹	~610 mA h g ⁻¹	22
		3.8 A g ⁻¹	181 mA h g ⁻¹	
Ox horns	KOH	100 mA g ⁻¹	1181 mA h g ⁻¹	
		5 A g ⁻¹	304 mA h g ⁻¹	23
Coconut oil	H ₂ SO ₄ and H ₂ O ₂	100 mA g ⁻¹	742 mA h g ⁻¹	
		10 A g ⁻¹	321 mA h g ⁻¹	24
		20 A g ⁻¹	226 mA h g ⁻¹	
silk	ZnCl ₂ and FeCl ₃	100 mA g ⁻¹	1913 mA h g ⁻¹	25

		5 A g^{-1}	523 mA h g^{-1}	
		20 A g^{-1}	261 mA h g^{-1}	
		37.2 A g^{-1}	212 mA h g^{-1}	
Duckweed	HNO_3	100 mA g^{-1}	1308 mA h g^{-1}	26
		1 A g^{-1}	630 mA h g^{-1}	
Cattle bones	Thermal	1 A g^{-1}	1230 mA h g^{-1}	11
Shrimp shell	Template-chemical	100 mA g^{-1}	1507 mAh g^{-1}	5
		500 mA g^{-1}	1014 mAh g^{-1}	
Algae-enteromorpha	KOH	100 mA g^{-1}	1709 mA h g^{-1}	27
Tofu	KOH	100 mA g^{-1}	2120 mA h g^{-1}	Our work
		1 A g^{-1}	900 mA h g^{-1}	

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