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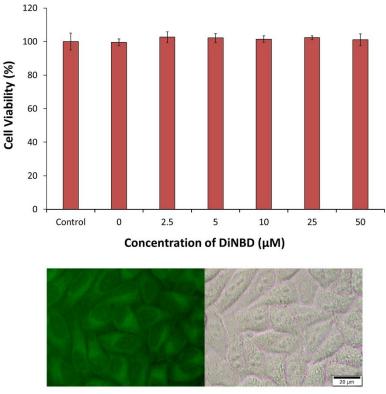
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## **Supplementary Information**

## Triple detection modes for Hg<sup>2+</sup>sensing based on NBD-fluorescent and colorimetric sensor and its potential in cell imaging

Sasiwimon Kraithong, a Rapeepat Sangsuwan, b Nattawan Worawannotai, c Jitnapa Sirirak, c Adisri Charoenpanich, d Patchanita Thamyongkit, a Nantanit Wanichachewa\*c

<sup>&</sup>lt;sup>d</sup> Department of Biology, Faculty of Science, Silpakorn University, Nakhon Pathom 73000, Thailand



Retaining of DiNBD within the cells after 24 h in growth media.

**Fig. S1.** Cell viability values (%) estimated by Prestoblue<sup>TM</sup> Cell Viability reagent versus incubation concentrations of **DiNBD**. HeLa cells were incubated with **DiNBD** (2.5–50  $\mu$ M) for 30 min (10% DMSO in PBS), and further cultured in growth media for 24 h before the cell viability assay. After 24 h culture, **DiNBD** remained within the cells but did not affect HeLa cell viability.

<sup>&</sup>lt;sup>a</sup> Department of Chemistry, Faculty of Science, Chulalongkorn University, Bangkok 10330, Thailand

<sup>&</sup>lt;sup>b</sup> Department of Chemistry, University of California, Berkeley CA 94720, USA

<sup>&</sup>lt;sup>c</sup> Department of Chemistry, Faculty of Science, Silpakorn University, Nakhon Pathom 73000, Thailand

## Cell viability assay

Cell survival of HeLa after treatment with **DiNBD** was measured using Prestoblue<sup>TM</sup> Cell Viability reagent (invitrogen, USA) according to the manufacturer's protocol. In brief, cells were seeded in 96 well plate at 20 kcells/well and incubated overnight. Cells were treated with different concentrations of **DiNBD** (10% DMSO in phenol red free DMEM) ranging from 2.5–50 μM, and two negative controls of DMEM alone or DMEM with 10% DMSO, then incubated for 30 min. Cells were washed twice followed by 24 h incubation in growth media. The media was replaced with 10% Prestoblue reagent in fresh media and incubated for 2 hours. Data values were measured as OD reading at 570 nm with reference subtraction at 600 nm. Data were expressed as mean±SD of triplicate experiments. Cell viability was calculated using the following formula: 100×[(OD<sub>570</sub> of treated sample)/(OD<sub>570</sub> of untreated sample (DMEM alone))]. Cell morphology and retaining of **DiNBD** within the cells at the maximum concentration test of 50 μM **DiNBD** was visualized under inverted phase contrast/fluorescence microscope (Olympus CKX53/ DP27-2, 100W mercury burner).