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# **Supporting Information**

# **New Journal of Chemistry**

# Covalent Attachment of a Fluorescent 'Pourbaix Sensor' onto a Polymer Bead for Sensing in Water

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# **Table of Contents**

Experimental	2
Synthesis	3
Figure S1. Reaction scheme for the synthesis of 3 and covalent attachment on the	
polymer bead to yield 4	3
Figure S2. UV-visible absorption spectra of 10 $\mu$ M 3 in 1:1 ( $\nu/\nu$ ) methanol/water at	
<b>pH</b> 3.0, 7.0 and 10.5	4
<b>Figure S3</b> . Fluorescent spectra of 10 $\mu$ M <b>3</b> in 1:1 ( $\nu/\nu$ ) methanol/water excited at 385	
nm. The input conditions are: (a) high $H^+$ and high $Fe^{3+}$ , (b) high $H^+$ , (c) high $Fe^{3+}$ ,	
and (d) neat sensor. The $H^{\scriptscriptstyle +}$ and $Fe^{3+}$ concentrations are 200 $\mu M$ and 20 $\mu M,$	
respectively	4
<b>Figure S4</b> . <sup>1</sup> H NMR spectrum of <b>3</b> in CDCl <sub>3</sub>	5
Figure S5. FTIR spectrum of 3	6
Figure S6. HRMS of compound 3	7

### **1** Experimental

### **1.1 Instrumentation**

Syntheses were performed with an IKA C-MAG HS 7 hot plate assisted by an IKA ETS-D5 temperature probe to help maintain a constant temperature during reflux. Thin-layer chromatography (TLC) was performed on pre-coated silica plates (Sigma-Aldrich) using glass capillary tubes. The spots were observed under long wavelength (365 nm) and short wavelength (254 nm) UV light from a UVGL-58 handheld lamp. Melting points were measured with a Stuart SMP11 melting point apparatus.

NMR spectra were acquired with a Bruker Avance III HD NMR spectrometer fitted with an Ascend 500 11.75 Tesla superconducting magnet and a multinuclear 5mm PABBO probe. The frequency is 500.13 MHz for <sup>1</sup>H NMR and 125.76 MHz for <sup>13</sup>C NMR. Data was analysed and processed by means of the software Topspin ver.3.2. Chemical shifts were reported in ppm downfield with respect to TMS at 0.00 ppm at 298 K. Infra-red spectra were recorded using a Shimadzu IR-Affinity-1 spectrophotometer between 4000-400 cm<sup>-1</sup>. The instrument was calibrated against 1601 cm<sup>-1</sup> polystyrene absorption peak. IR analyses were performed as KBr disks or as a thin film between NaCl plates. The high resolution mass spectrometry (HRMS) was performed by ESI-ToF outsourced to Medac Ltd (UK). http://medacltd.com/

UV-visible absorption spectra were acquired on a Jasco V-650 spectrophotometer. Quartz cells with a pathlength of 1.0 cm were used for the measurements. Parameters of the instrument were set at 0.5 nm bandwidth and 400 nm/min scan speed. Blank spectrum was performed on the neat solvent as a reference prior the measurements of the spectra. Fluorescent measurements were acquired with a Jasco FP-8300 spectrophotometer. Quartz cells with a pathlength of 1.0 cm were used for the measurements. The parameters were excitation and emission bandwidths of 2.5 nm, 50 msec response time, and a scanning speed of 200 nm min<sup>-1</sup>. The absorbance of the solutions for fluorescence was set to less than 0.1 to prevent quenching of the emission.

Photos of beads were taken with a fluorescent microscope Axiovert 40 CLF, manufactured by Carl Zeiss MicroImaging. The microscope had an ocular magnification of  $\times 10$ , and the magnification that was used was  $\times 40$ . As a source UV light, a low pressure mercury lamp was used. A UV filter with an excitation band between 450-490 nm was used. The emission of light started from 515 nm and the beam splitter was adjusted to 510 nm.

#### Synthesis

Synthesis of *N*-Ferrocenyl-4-((4-(2-methyl-1,3-dioxo-2,3-dihydro-1H-benzo[de]isoquinolin-6-yl)piperazin-1-yl)methyl)benzoic acid **3** 

*N*-Ferrocenylmethyl-4-piperazine-1,8-naphthalimide **2** (76 mg, 0.16 mmol), 4-(bromomethyl)benzoic acid (114 mg, 0.53 mmol) and triethylamine (88 µL, 0.63 mmol) were dissolved in 4 mL of 1,2-dichloroethane and refluxed at 90 °C for 3 hours. The solution was extracted with 1,2-dichloromethane and washed with 10% aqueous NaOH. The product was recrystallised from acetone to give an orange powder in 19% yield (22 mg). m.p. > 250 °C;  $R_f$ = 0.44 (9:1 (v/v) CH<sub>2</sub>Cl<sub>2</sub>/methanol); FTIR (KBr, salt, cm<sup>-1</sup>): 3051, 2950, 2850, 1695 (C=O), 1653, 1585, 1382, 1183, 1110; MS (ES-TOF, 1.56 mV) m/z (%): 177 (59), 178 (10), 410 (100), 411 (28), 412 (5); HRMS for C<sub>32</sub>H<sub>43</sub>N<sub>4</sub>O<sub>5</sub>FeNa [M+Na] calculated 642.2481, found 642.2484.

### Attachment of **3** onto Tentagel

To a 50 mL round-bottom flask, 16 mg (0.025 mmol) of **3**, 26 mg (0.13 mol) of DCC and 100  $\mu$ L of HOBt were dissolved in 1 mL of DMF. The reaction was stirred at 40 °C for 4 hours. The beads were washed sequentially with 10 mL of DMF, 1:1 (v/v) DMF/MeOH and MeOH.

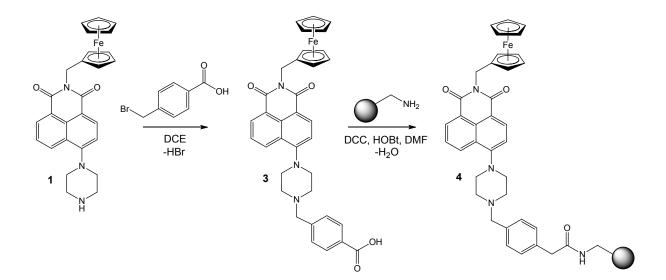
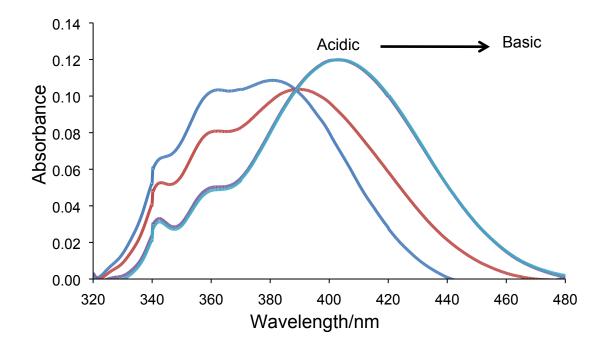
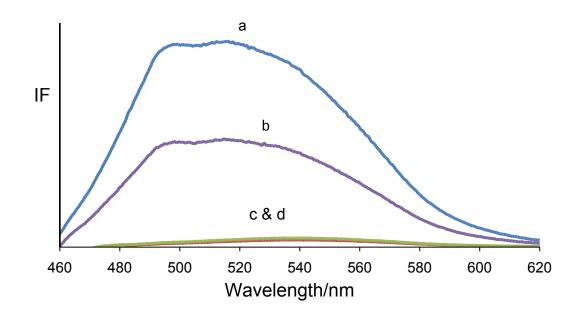


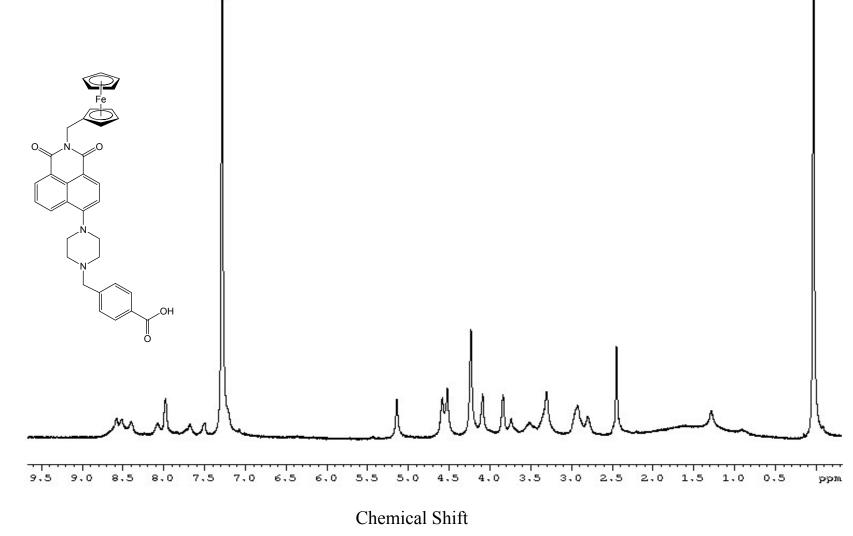
Figure S1. Reaction scheme for the synthesis of 3 and covalent attachment on the polymer bead to yield 4.



**Figure S2**. UV-visible absorption spectra of 10  $\mu$ M **3** in 1:1 (*v*/*v*) methanol/water at pH 3.0, 7.0 and 10.5.



**Figure S3**. Fluorescent spectra of 10  $\mu$ M **3** in 1:1 ( $\nu/\nu$ ) methanol/water excited at 385 nm. The input conditions are: (a) high H<sup>+</sup> and high Fe<sup>3+</sup>, (b) high H<sup>+</sup>, (c) high Fe<sup>3+</sup>, and (d) neat sensor. The H<sup>+</sup> and Fe<sup>3+</sup> concentrations are 200  $\mu$ M and 20  $\mu$ M, respectively.



**Figure S4**. <sup>1</sup>H NMR spectrum of **3** in CDCl<sub>3</sub>.

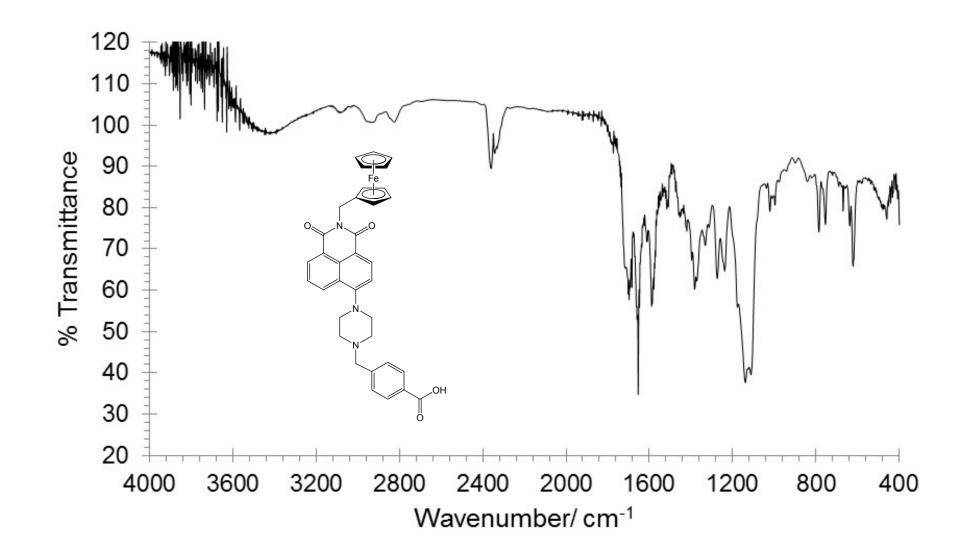


Figure S5. FTIR spectrum of 3.

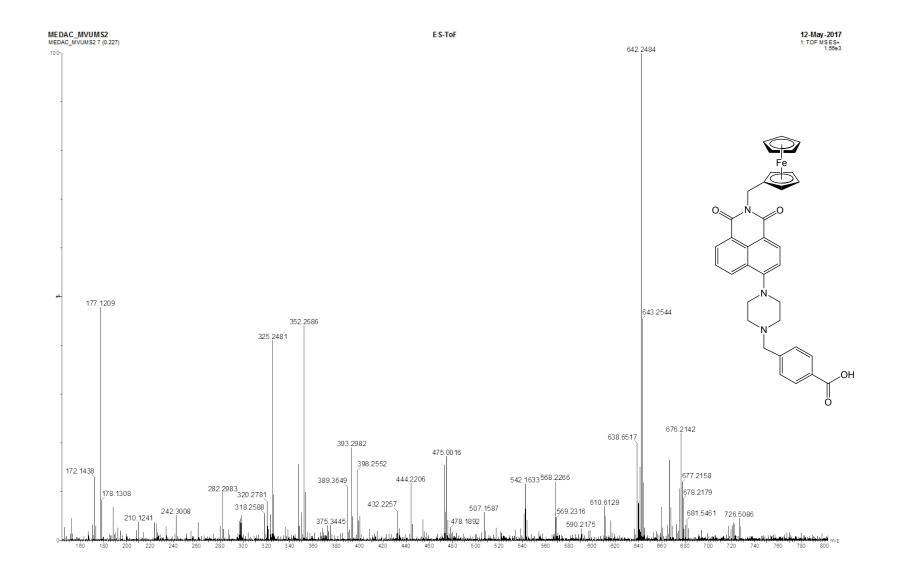


Figure S6. HRMS of compound 3.