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Electronic Supplementary Information (ESI[†])

Colorimetric and "off-on" fluorescent Pd²⁺ chemosensor based on rhodamine-ampyrone conjugate: synthesis, experimental and theoretical studies along with *in vitro* applications

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Figure S1: ¹H NMR spectrum of compound L.



Figure S2: ¹³C NMR spectrum of chemosensor L.



Figure S3: Mass spectrum of chemosensor L.



Figure S4: Mass spectrum of chemosensor L-Pd²⁺ complex.



Figure S5: Change in the absorption spectrum of receptor L $[c = 4 \times 10^{-5} \text{ M}, CH_3CN/H_2O = 1 :1, v/v, 10 mM HEPES buffer, pH = 7.4) with respective metal cations (c = 4× 10⁻⁴ M, left to right- L, K⁺, Na⁺, Mg²⁺, Ca²⁺, Sr²⁺, Ba²⁺, Cr³⁺, Mn²⁺, Fe²⁺, Fe³⁺, Pd²⁺, Co²⁺, Ni²⁺, Cu²⁺, Zn²⁺, Cd²⁺, Pb²⁺, Hg²⁺, Pt²⁺, Pd⁰, Al³⁺, Ru³⁺, and Ag⁺).$



Figure S6: Fluorescence Job's plot for L with Pd^{2+} in CH_3CN/H_2O solution (8:2, v/v, 10 mM HEPES buffer, pH 7.4). ([H] = [G] = 4 × 10⁻⁵ M).



Figure S7: Fluorescence response of L ($c = 1.0 \times 10^{-5}$ M) to 1.0 equiv addition of Pd²⁺ (the red bar portion) and to the mixture of 10 equiv. of other metal ions with 1.0 equiv. of Pd²⁺ (the black bar portion, left to right- K⁺, Na⁺, Mg²⁺, Ca²⁺, Sr²⁺, Ba²⁺, Cr³⁺, Mn²⁺, Fe²⁺, Fe³⁺, Co²⁺, Ni²⁺, Cu²⁺, Zn²⁺, Cd²⁺, Pb²⁺, Hg²⁺, Pt²⁺, Pd⁰, Al³⁺, Ru³⁺, and Ag⁺).

Calculation for Limit of Detection (LOD):

The LOD of L for Pd^{2+} was determined using the following equation:

LOD = 3Sbl/S, Sb1 is the standard deviation of the blank solution; S is the slope of the calibration curve.



Figure S8: Calibration curve for fluorescence titration of L with Pd²⁺.

From the graph we get slope (S) = 6×10^6 Standard deviation (Sb1 = 23.81723) Thus, using the formula, we get the LOD = 11.9×10^{-6} M = 11.9μ M.



Figure S9: Bensei-Hildebrand plot obtained from the Fluorescence (emission calculated from 575 nm) studies. Binding constant ($K_a = 8.36 \times 10^3 \text{ M}^{-1}$) curve of sensor L with Pd²⁺ determined by fluorescence method.



Figure S10: FTIR spectra of (a) L and (b) L-Pd²⁺ complex.

Theoretical study:

Species	E(HOMO)	E(LUMO)	$\Delta E(Hartree$	$\Delta E(eV)$	$\Delta E(\text{kcal/mol})$
L	-0.18937	-0.06438	0.12499	3.401	78.43
L-Pd ²⁺	-0.35355	-0.30237	0.05118	1.3927	32.11

Table S1. HOMO-LUMO energy calculated for L1 and L1- $2Zn^{2+}$ complex using [(B3LYP/6-311G(d,p)] for L and B3LYP/LanL2DZ for L-Pd²⁺ for level of theory)

1 Hartree = 27.2116 eV, 1 Hartree = $627.5095 \text{ kcal mol}^{-1}$



Figure S11: Energy-minimized structure of L-Pd²⁺ complex (atom color: gray = C, red = O, blue = N, white = H, teal blue = Pd).

Table S2: Selected bond length and bond angles in B3LYP/LanL2DZ optimized geometry of L-Pd²⁺.

Bond	Bond length (Å)
Pd-O (rhodamine carbonyl)	2.09
Pd-N (rhodamine imino)	1.96
Pd-O (antipyrine carbonyl)	2.16
Pd-N (antipyrine inamine)	2.02



Figure S12: Molecular orbital plots of L and L–Pd²⁺.

Cell imaging study:

Minimum Inhibitory Concentration (MIC): We have treated probe L with both gram positive and gram-negative bacteria. After 24 hrs. of treatment probe L shows no effect on gram negative bacteria but it showed some effect on gram positive bacteria. Probe L showed bactericidal activity on 100 μ M.



Figure S13: Minimum inhibitory concentration of compound against gram positive bacteria.

Cell Survivability Assay: Probe L showed cytotoxity against MDA-MB 468 cells when treated with different concentrations 0 - 150 μ M for 24 hrs and cell survivability was determined by MTT assay. As seen in Figure S14 cell survivability decreased with increasing concentration of probe L. From the graph we also calculated LD₅₀ and found that the value was 50 μ M approx. for the probe L.



Figure S14: Cytotoxic effect of probe L on MDA-MB-468 cells. Cells were incubated with increasing concentrations probe L and its survivability was assessed by MTT assay.

SI.	Chemical Structure	Media	Pd ²⁺	Biological
No.			limit of	application
			detection	
1.	Our Work	CH ₃ CN/H ₂ O (4:1, v/v, 10 mM HEPES buffer, pH 7.4)	11.9 μM	Our chemosensor can detect intracellular Pd ²⁺ ion in MDA-MB-468 cells.
2.	R = -0 - (Tetrahedron, 2014, 70, 1997-2002)	DMF/H ₂ O (v/v, 7/1)	7.32 ppb (5.53 μM)*1	Not mentioned
3.	MeO (<i>Chem. Commun.</i> , 2008, 6339–6341)	Ethanol–water (60 : 40, v/v) solution at pH 7.2	Not mentioned	Not mentioned
4.	EtHN O NHEt (<i>Chem. Commun.</i> , 2011, 47 , 9101–9103)	EtOH-H ₂ O (1 : 1, v/v, 25 1C) at pH 7.2 (50 mM HEPES buffer)	Not mentioned	Not mentioned
5.	(<i>Dalton Trans.</i> , 2014, 43 , 4626–4630)	EtOH–H ₂ O (1 : 1, v/v)	73.8 nM (0.0738 μM) * ²	Not mentioned

Table S3. Summary of representative fluorescent probes for Pd^{2+}

6		MeOH/PBS	0.05 µM* ³	Not mentioned
0.	1	(pH = 7.4, 10		
		mM) solution		
	H S	(8:2, v:v),		
	(Sens Actuators B 2012 171-172 1277-1282)			
	(Sens. Menualors, D 2012, 111 112, 1211 1202)			
7.		HAc–NaAc	0.13 μM*4	Not mentioned
		buffer		
	N-N N	solution (pH = 4.7)		
	ΓΥΥΝ Ϊ	4.7)		
	Ĵ			
	(Sens. Actuators, B 2012, 171–172 , 508–514)			
8.		MeCN/DMSO	Not	Not mentioned
		(99:1)	mentioned	
	N.			
	NH VI			
	s N O			
	CO O NI S N CO			
	P OL NH			
	(Tetrahedron 2011, 67, 7106-7113)			
a		Ethanol-water	1 49 ×10 ⁻⁹	Not mentioned
9.		(4:1, v/v),	M	i tot mentioned
	P or V		(1.40, 10.2	
			(1.49×10^{-3})	
			μΜ)**	
	(Chem. Commun., 2013, 49 , 822-824)			

10.	(Analyst, 2017, 142, 1536–1544)	CH ₃ CN : H ₂ O (3 : 2 v/v)	1 μM* ⁶	Not mentioned
11.	(<i>Tetrahedron Lett.</i> 2012, 53 , 3459–3462)	PBS (20 mM, pH 7.4) solution containing 10% (v:v) CH ₃ CN	Not mentioned	Not mentioned

 $*^1$ Molecular weight of fluorophore is too high compared to L (our work). Thus, it is less important with respect to atom economy though its limit of detection is less than L

*² Overall yield of reported compound formation (37% in last step) is low compared to the synthesis of L.

*³ The number of steps for the synthesis of fluorescent probe is more and some of the steps are low yielding (24%-75%).

*⁴ Experiments were carried out at acidic condition (pH = 4.7). But biological pH is generally close to neutral. Thus, this method has much lower significance for Pd^{2+} contaminated cell.

*⁵ Final step is low yielding (25%).

*⁶ Yield of the final step is not high (55%). Due to large size of the molecule, it is less important in terms of atom economy too.

*¹⁻⁶ None of the tabulated methods above, except ours, were silent about their applications on living cells.