

Synthesis of artemisinin derived glycoconjugates inspired by click chemistry

Lakshmi Goswami,^{‡ab} Sayantan Paul,^{‡ab} Tharun K. Kotammagari^{ab}
and Asish K. Bhattacharya^{*ab}

^aDivision of Organic Chemistry, CSIR-National Chemical Laboratory, Dr. Homi Bhabha Road, Pune-411 008, India; E-mail: ak.bhattacharya@ncl.res.in

^bAcademy of Scientific and Innovative Research (AcSIR), Dr. Homi Bhabha Road, Pune-411 008, India

Supporting Information

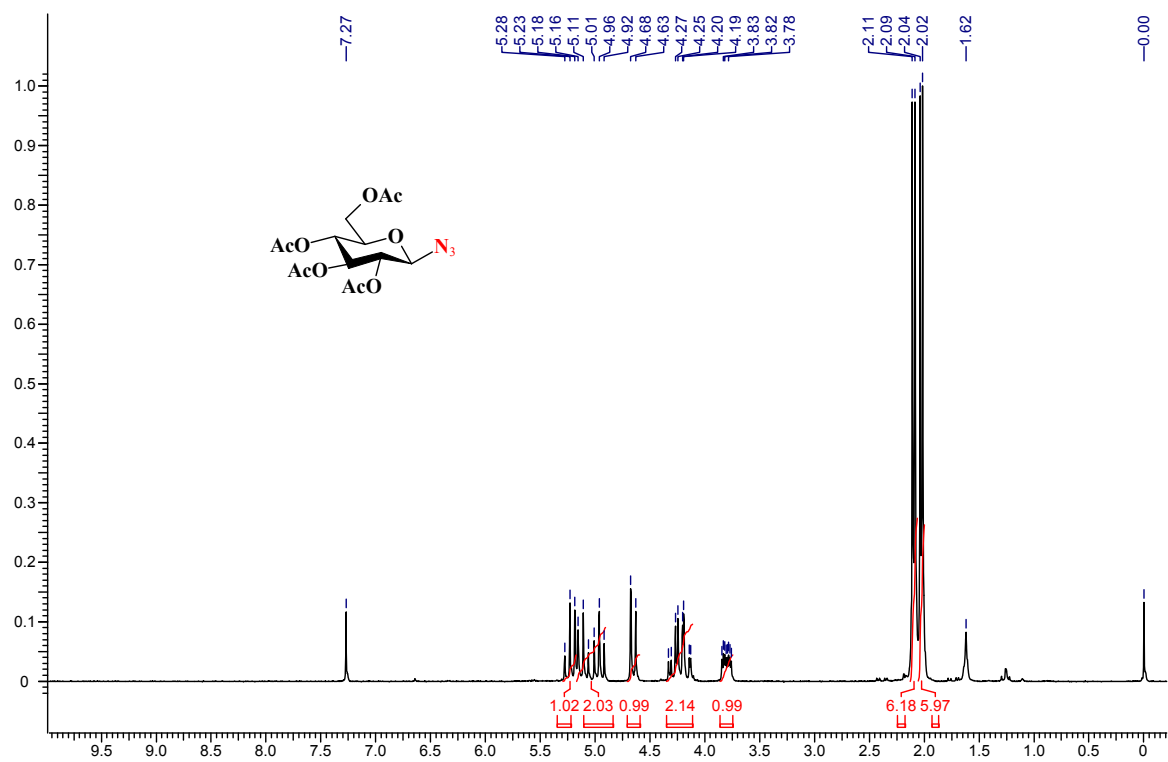
Spectra of all the synthesised compounds

Representative procedure for the synthesis of tetra-*O*-acetyl azido glucose (**8a**):

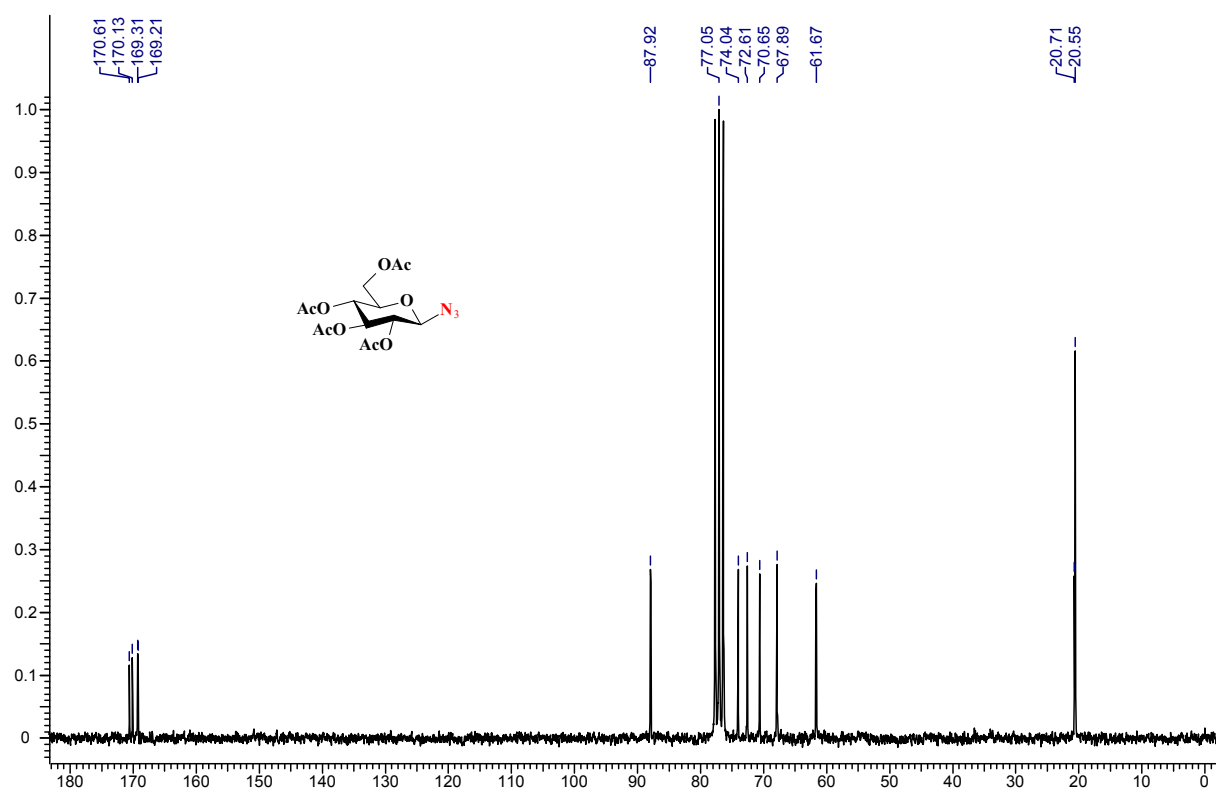
To a solution of D-glucose (10.0 g, 55.5 mmol, 1.0 equiv.) in dry pyridine (100 mL) was cooled to 0 °C. Acetic anhydride (100 mL) was added dropwise to the stirred reaction mixture at 0 °C. After 6 h when the reaction was complete, the reaction mixture was poured to an ice-water mixture which was then extracted with ethyl acetate (3 X 100 mL) and dried over anhydrous sodium sulphate. The reaction mixture was coevaporated with toluene to furnish a semi-solid which was used as such for the bromination without further purification. Pentacyclated glucose (6 g) was dissolved in dichloromethane (20 mL) and HBr in acetic acid (33% w/w, 10 mL) was added dropwise. After 2 hours, the whole reaction mixture was azeotroped with toluene to furnish tetra-*O*-acetyl α -bromo glucose. This was utilized in the next step without any further purification. To a solution of tetra-*O*-acetyl α -bromo glucose (2.0 g, 4.86 mmol, 1.0 equiv.) in dry DMF (10 mL), sodium azide was added (0.14 g, 5.84 mmol, 1.2 eq.) and the reaction was allowed to stir at room temperature for 16 h. After completion of the reaction, the reaction mixture was then diluted with water (50 mL) and extracted with EtOAc (100 mL). The organic layer was dried over anhydrous sodium sulphate and evaporated to dryness. The residue was purified by column chromatography (SiO₂, EtOAc/petroleum ether 1:2) to furnish pure **8a**.

Spectral data of tetra-*O*-acetyl azido glucose (**8a**): Colorless solid; m.p. 128-130 °C; $[\alpha]_D^{25}$ -29.7 (c 1.0, CHCl₃); ¹H NMR (200 MHz, CHLOROFORM-*d*) δ_H : 2.03 (d, *J* = 4.30 Hz, 6H), 2.10 (d, *J* = 5.1 Hz, 6H), 3.75-3.86 (m, 1H), 4.11-4.35 (m, 2H), 4.65 (d, *J* = 8.8 Hz, 1H) (anomeric proton), 4.90-5.17 (m, 2H), 5.17-5.29 (m, 1H); ¹³C NMR (50 MHz, CHLOROFORM-*d*) δ_C : 20.6, 20.7, 61.7, 67.9, 70.6, 72.6, 74.0, 87.9, 169.2, 169.3, 170.1, 170.6; LCMS (ESI): 396 (M+Na)⁺.

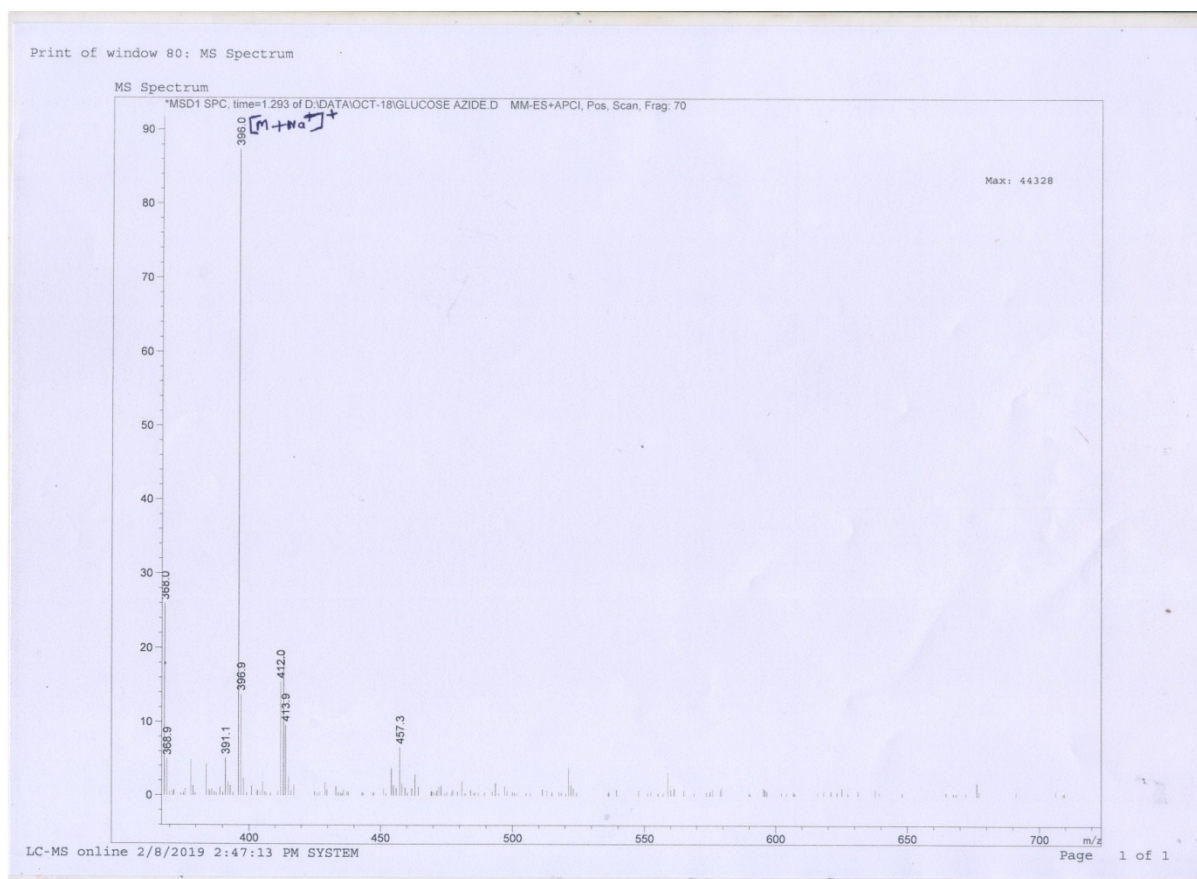
Ref.: M. A. Maier, C. G. Yannopoulos, N. Mohamed, A. Roland, H. Fritz, V. Mohan, G. Just and M. Manoharan, *Bioconjugate Chem*, 2003, **14**, 18-29.



¹ H NMR of **8a** (CDCl₃, 200 MHz)

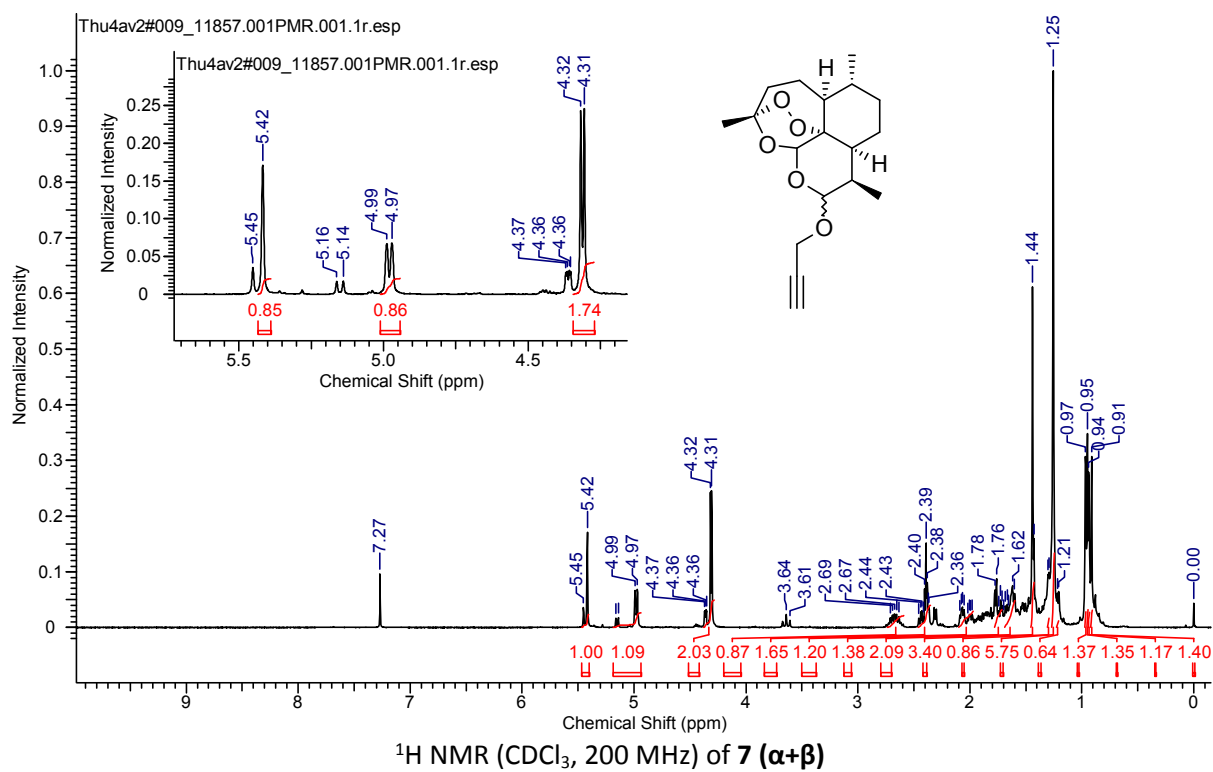


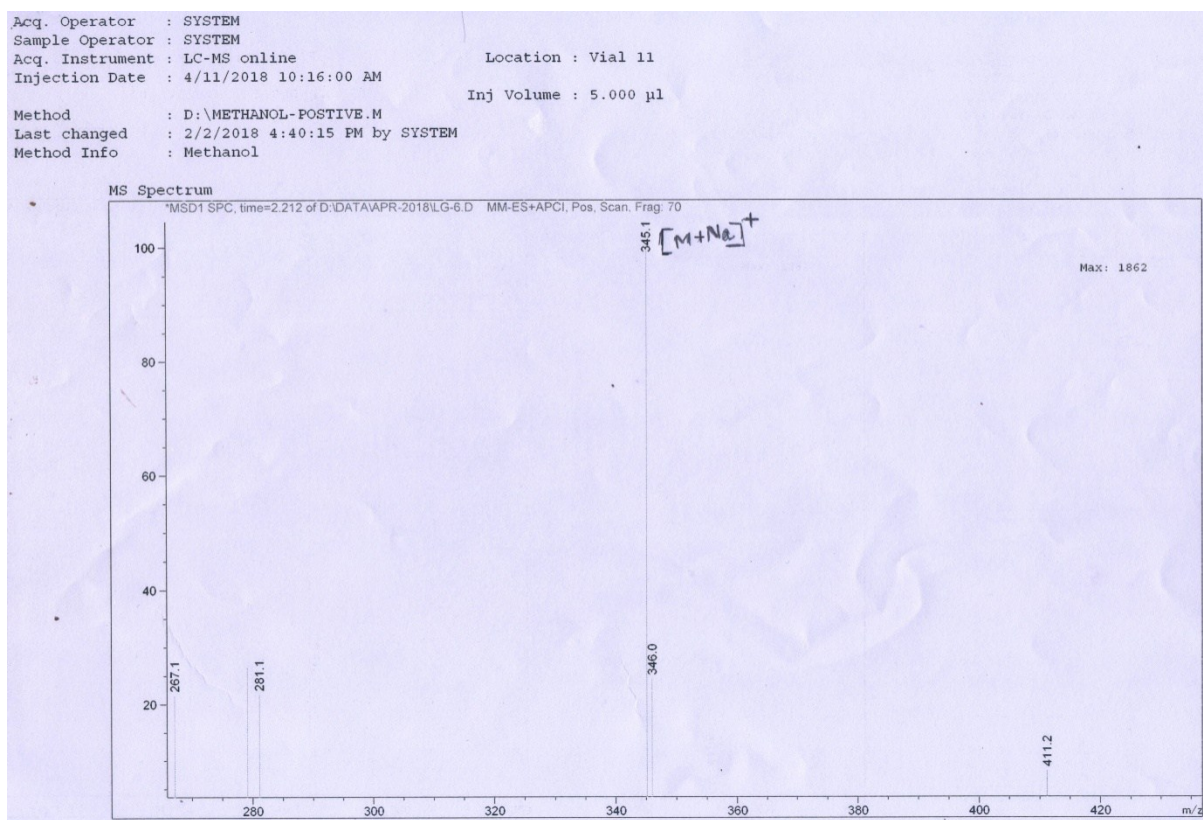
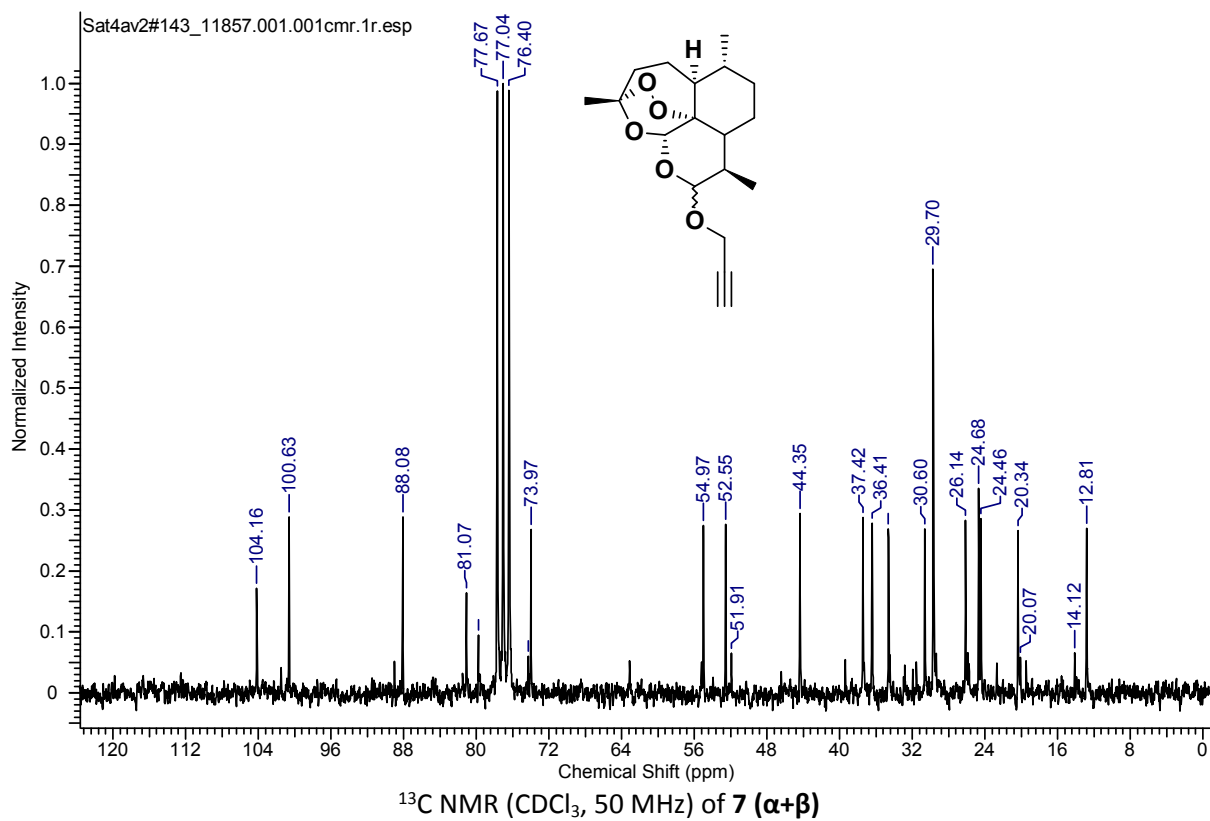
¹³ C NMR of **8a** (CDCl₃, 50 MHz)



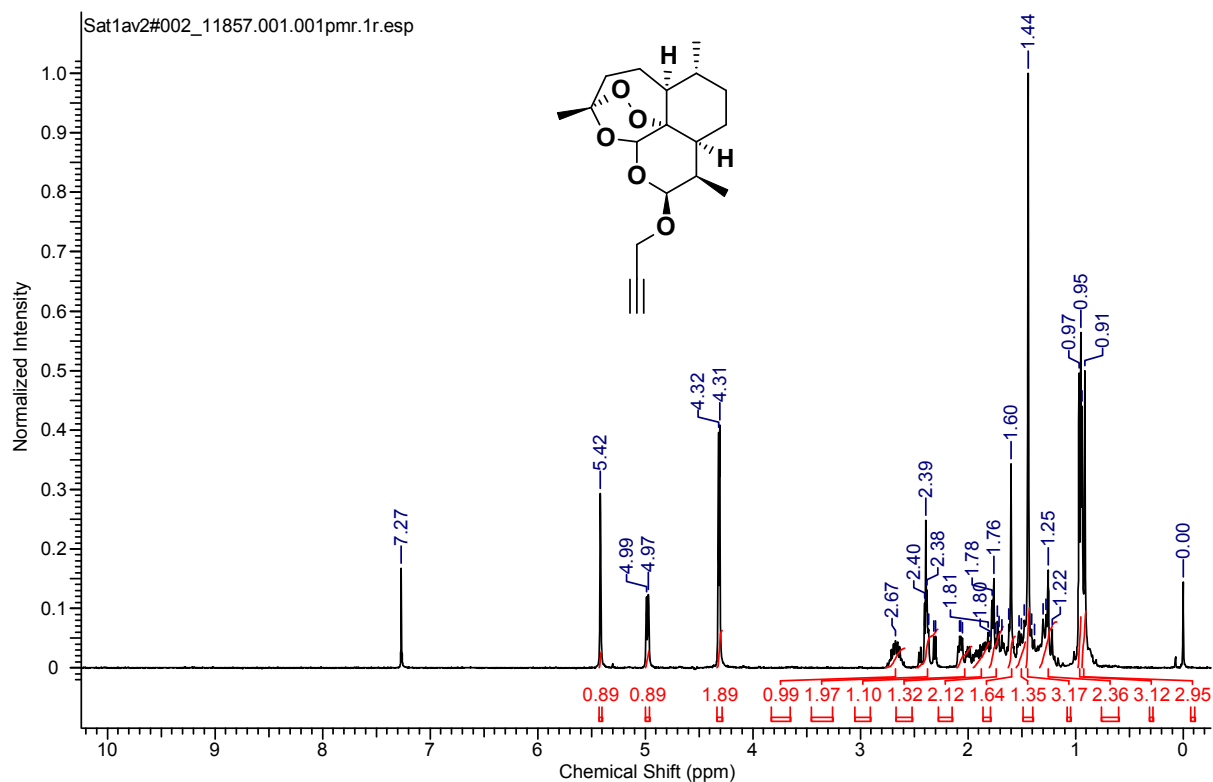
LCMS of **8a**

^1H , ^{13}C , LCMS and HRMS spectra of all the synthesized artemisinin glycoconjugates (**9a-i**)

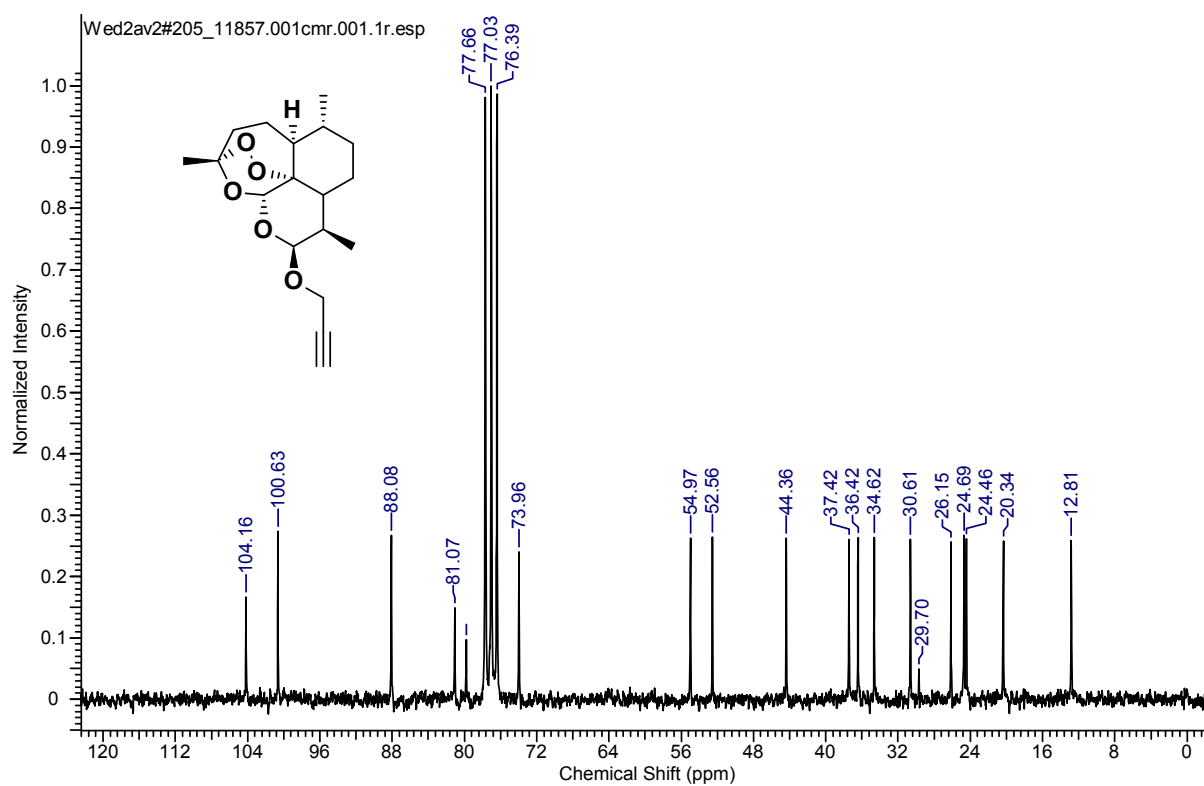




LCMS of **7**

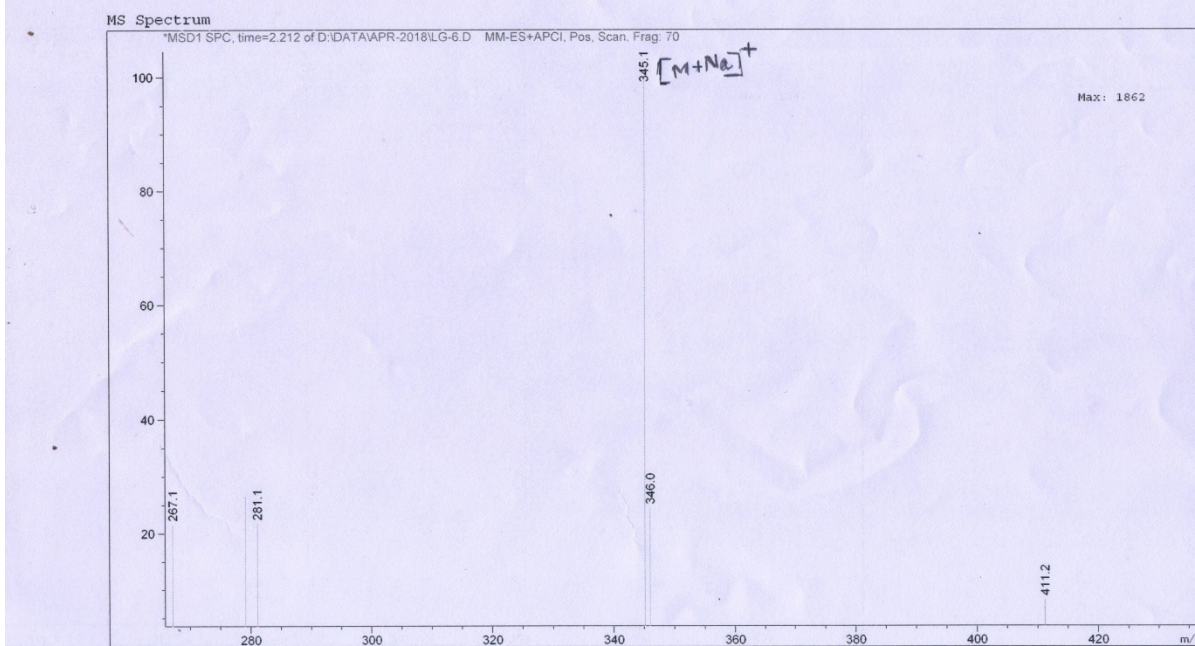


^1H NMR (CDCl_3 , 200 MHz) of **7a**

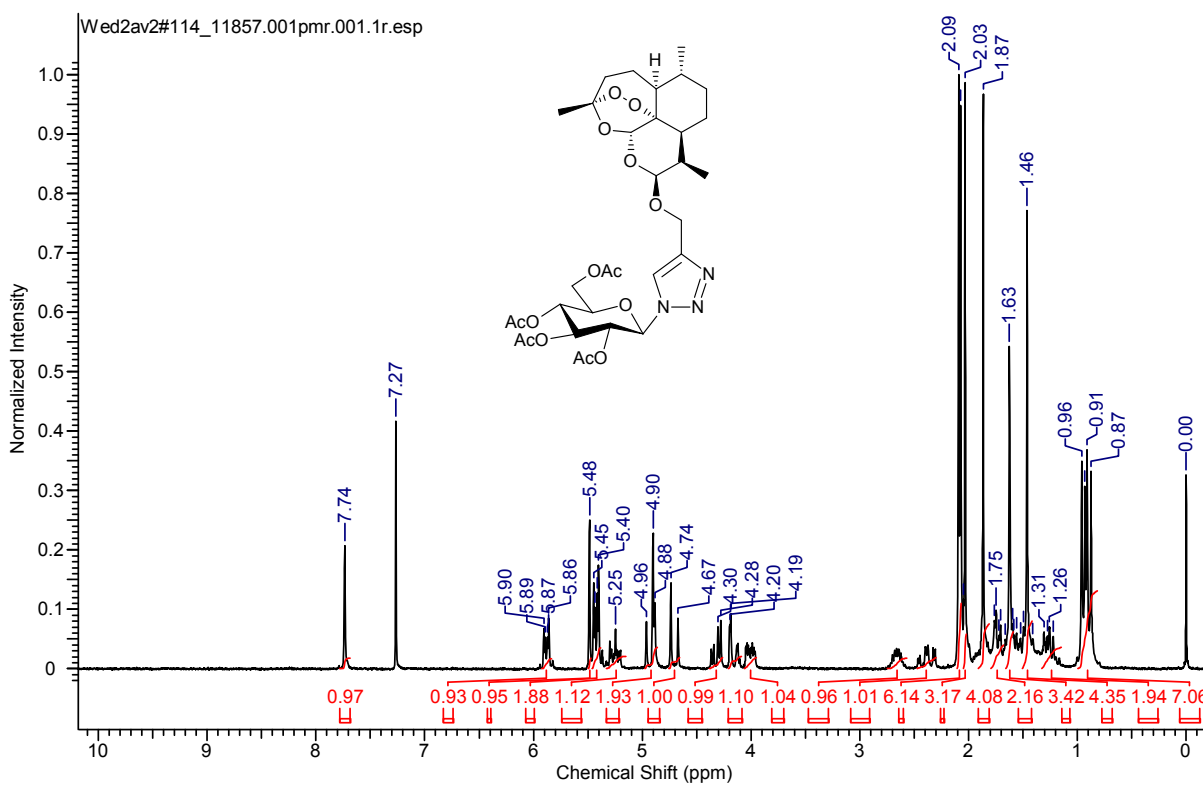


^{13}C NMR (CDCl_3 , 50 MHz) of **7a**

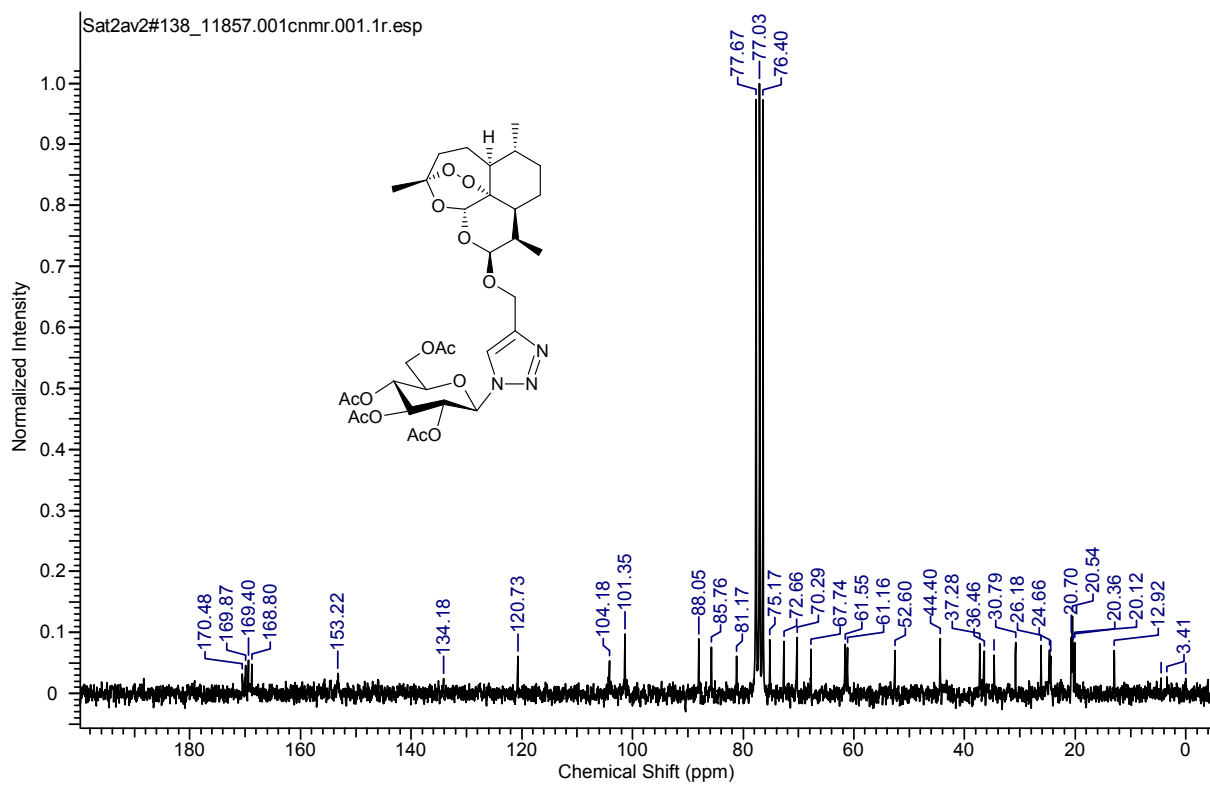
Acq. Operator : SYSTEM
 Sample Operator : SYSTEM
 Acq. Instrument : LC-MS online Location : Vial 11
 Injection Date : 4/11/2018 10:16:00 AM Inj Volume : 5.000 µl
 Method : D:\METHANOL-POSTIVE.M
 Last changed : 2/2/2018 4:40:15 PM by SYSTEM
 Method Info : Methanol



LCMS of 7a

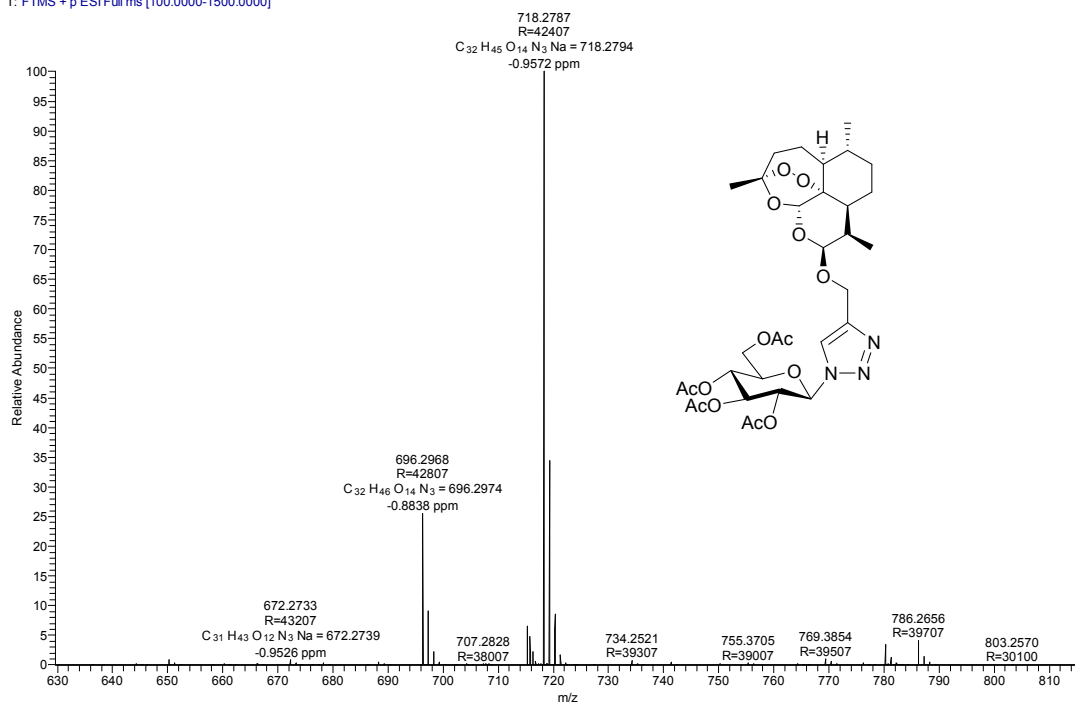


^1H NMR (CDCl_3 , 200 MHz) of 9a

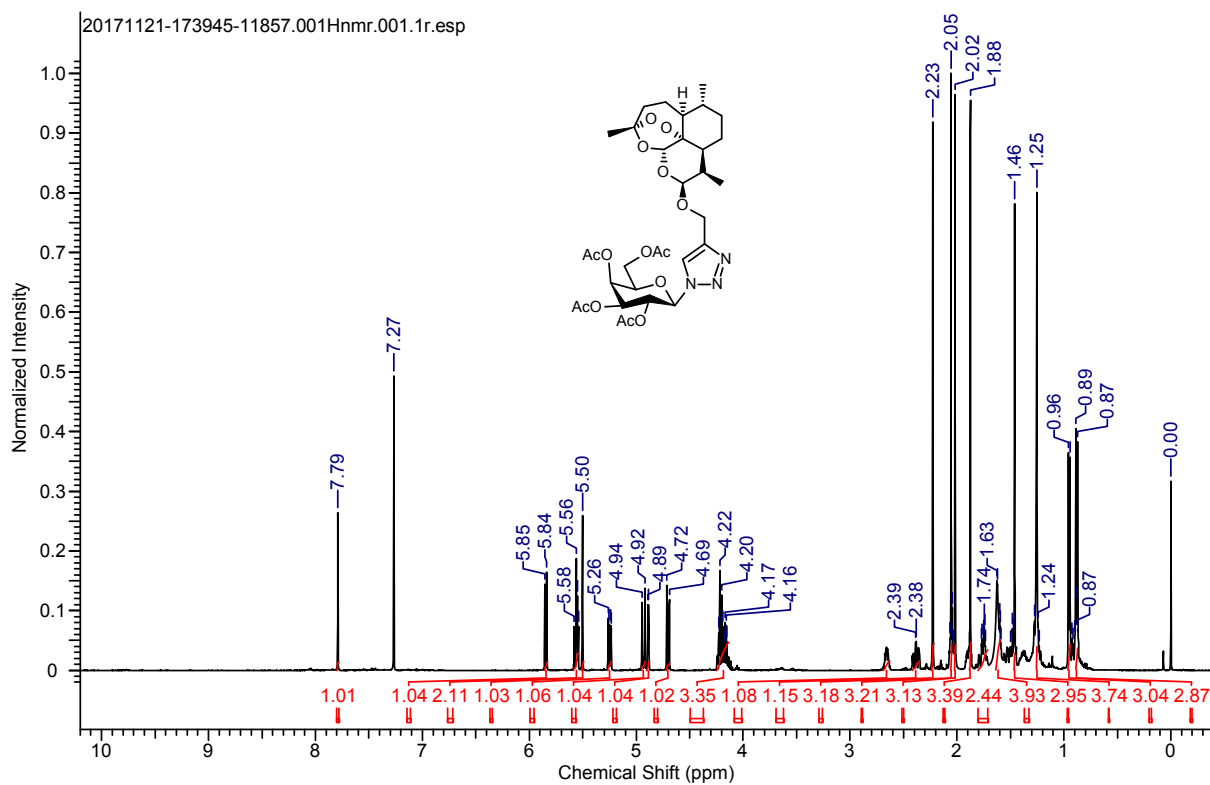


^{13}C NMR (CDCl_3 , 50 MHz) of **9a**

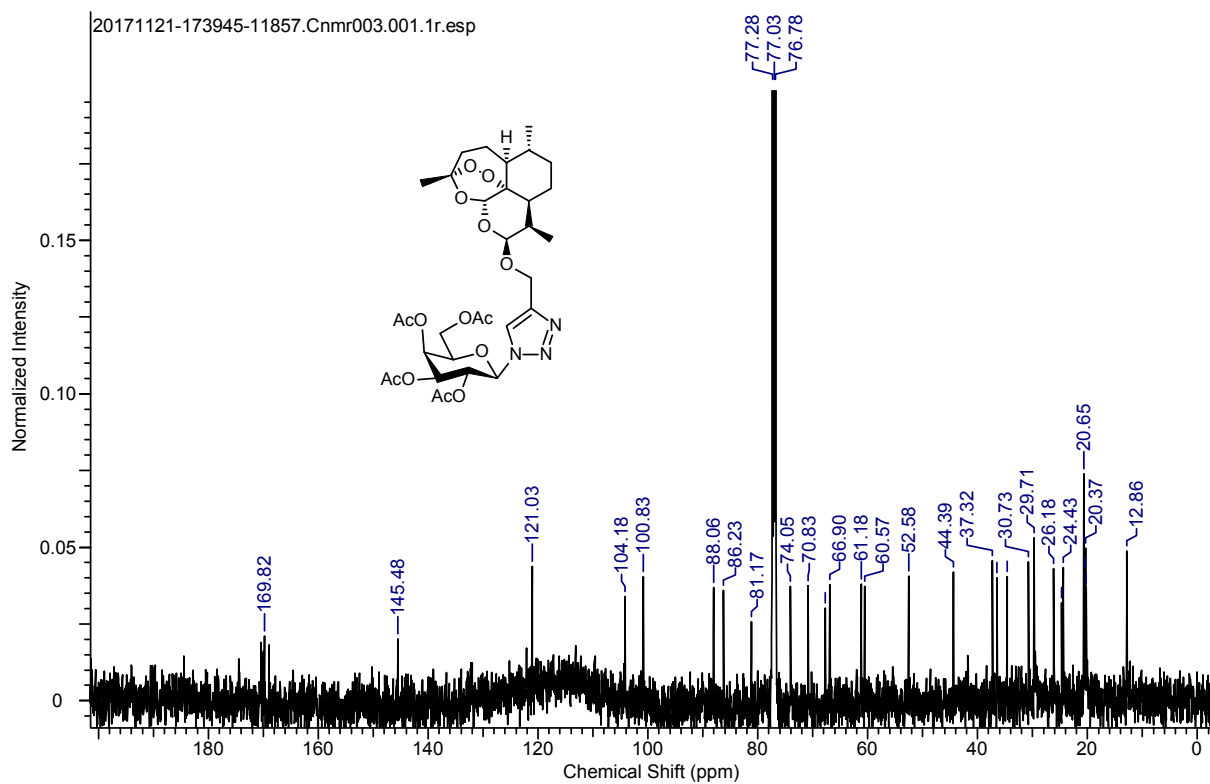
LG-BR#280 RT: 1.25 AV: 1 NL: 1.04E9
T: FTMS + p ESI Full ms [100.0000-1500.0000]



HRMS of **9a**

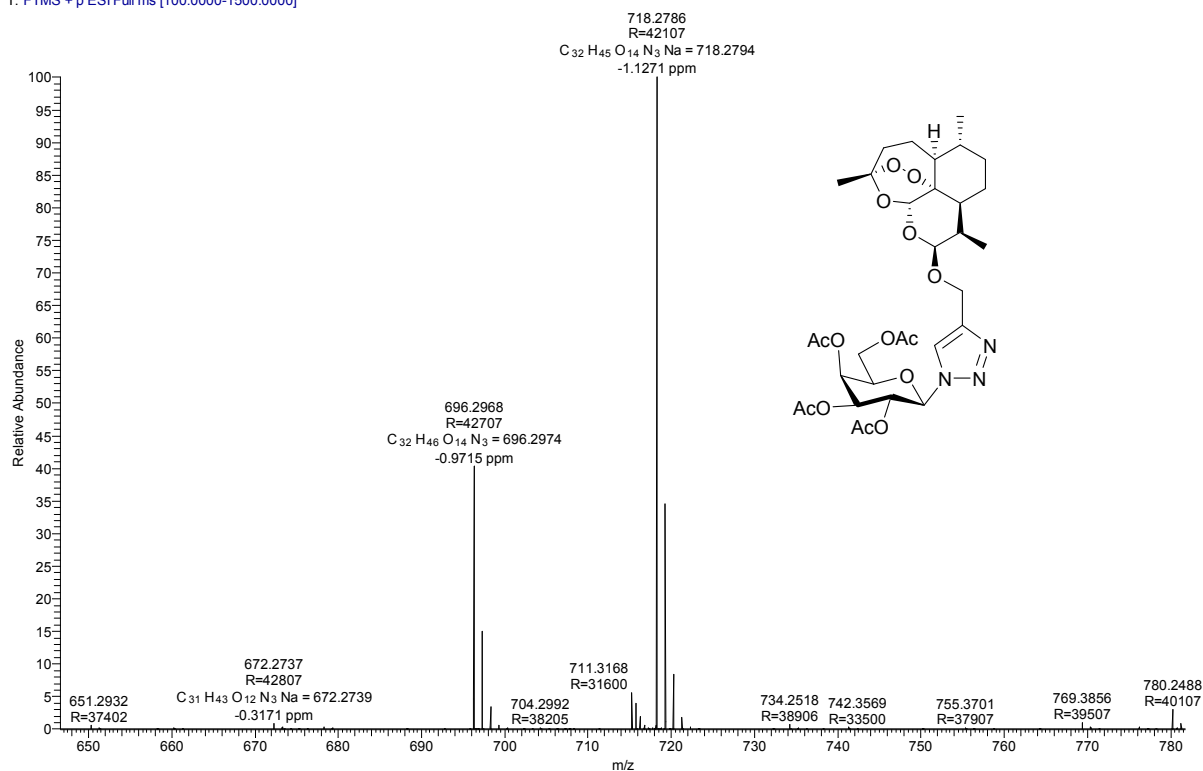


^1H NMR (CDCl_3 , 500 MHz) of **9b**

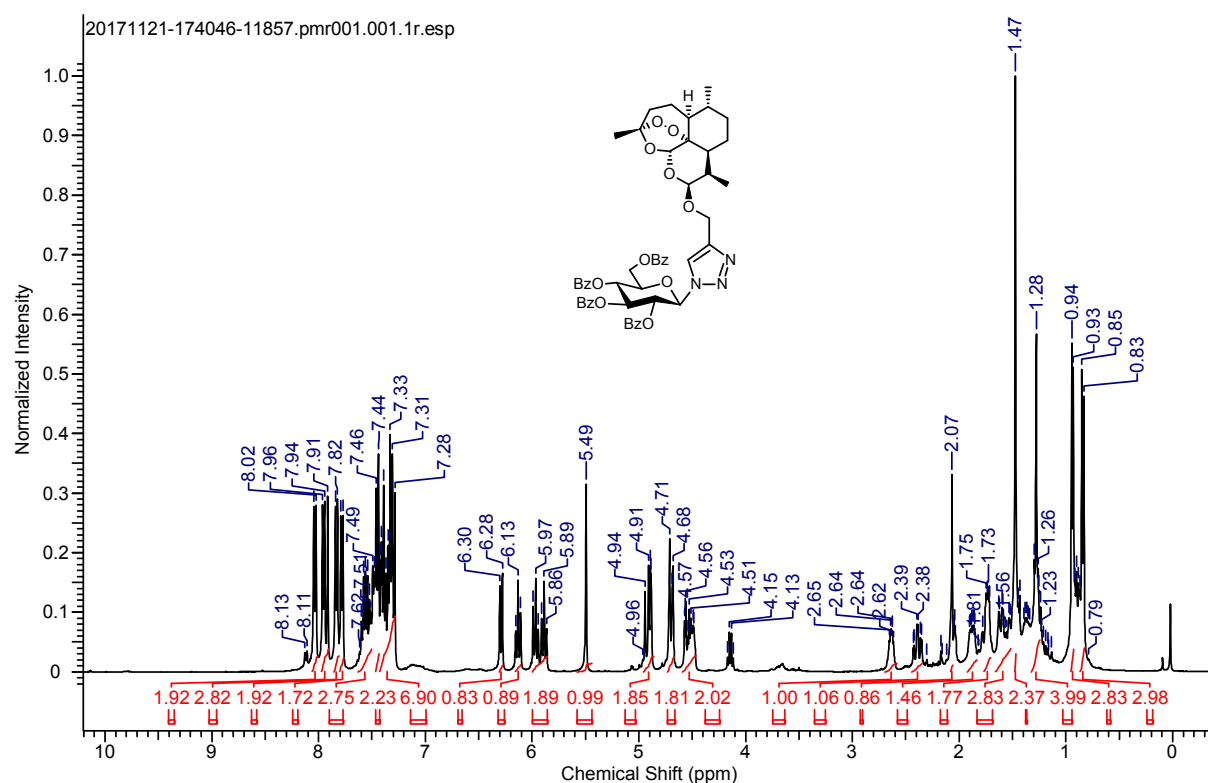


^{13}C NMR (CDCl_3 , 125 MHz) of **9b**

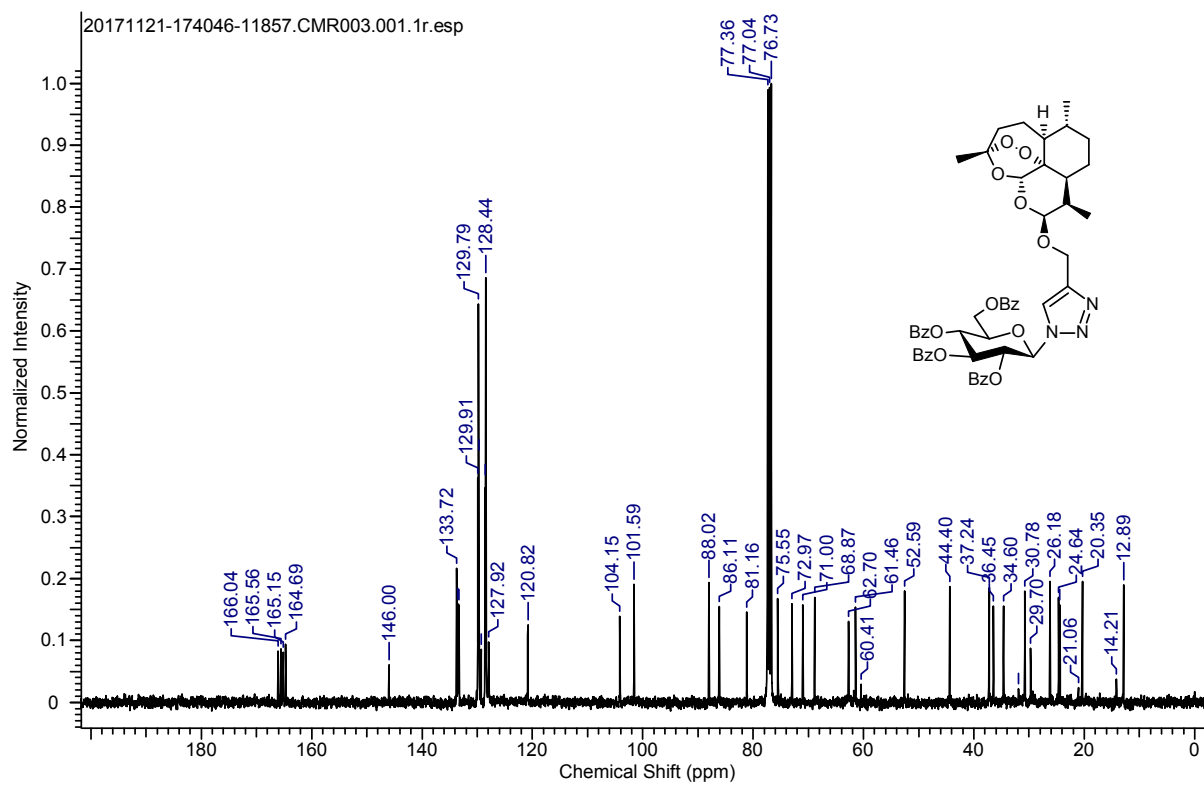
LG-10 #283 RT: 1.26 AV: 1 NL: 1.31E9
T: FTMS + p ESI Full ms [100.0000-1500.0000]



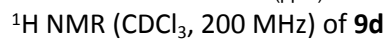
HRMS of **9b**

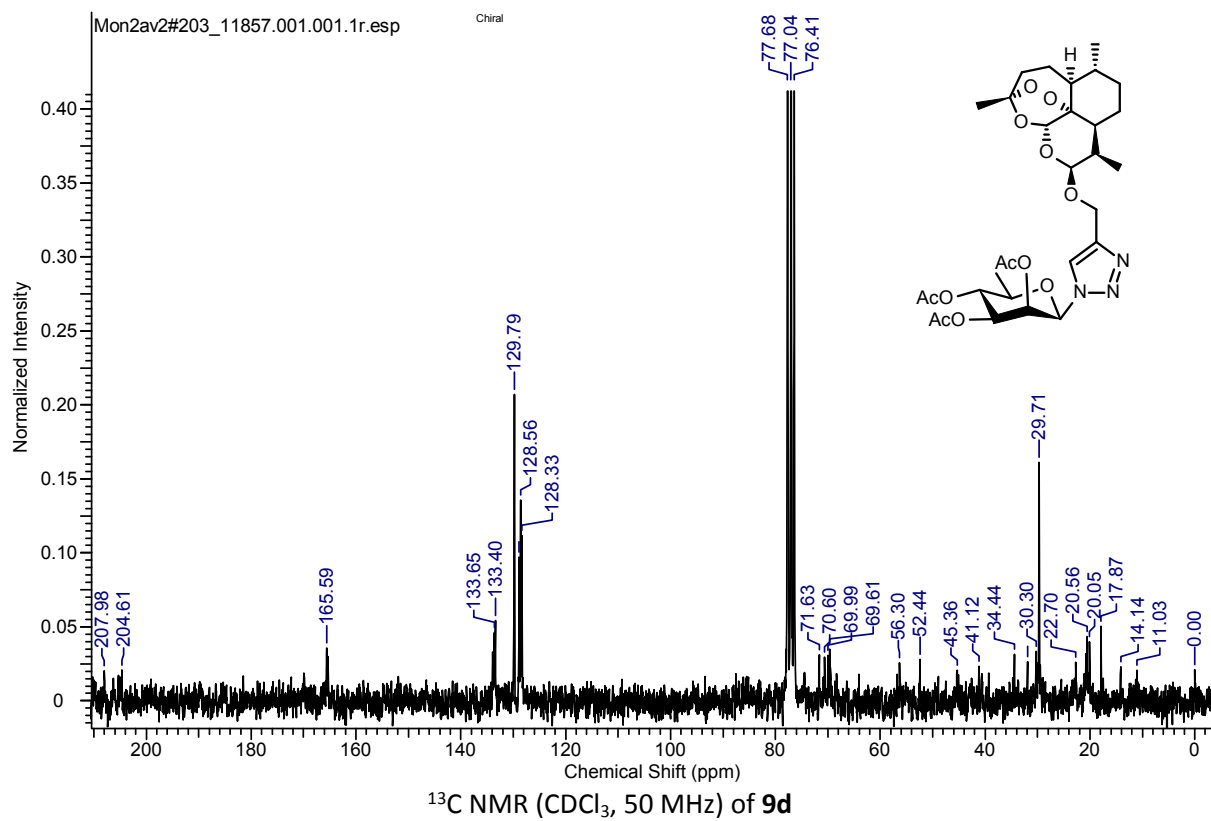


¹H NMR (CDCl₃, 400 MHz) of **9c**

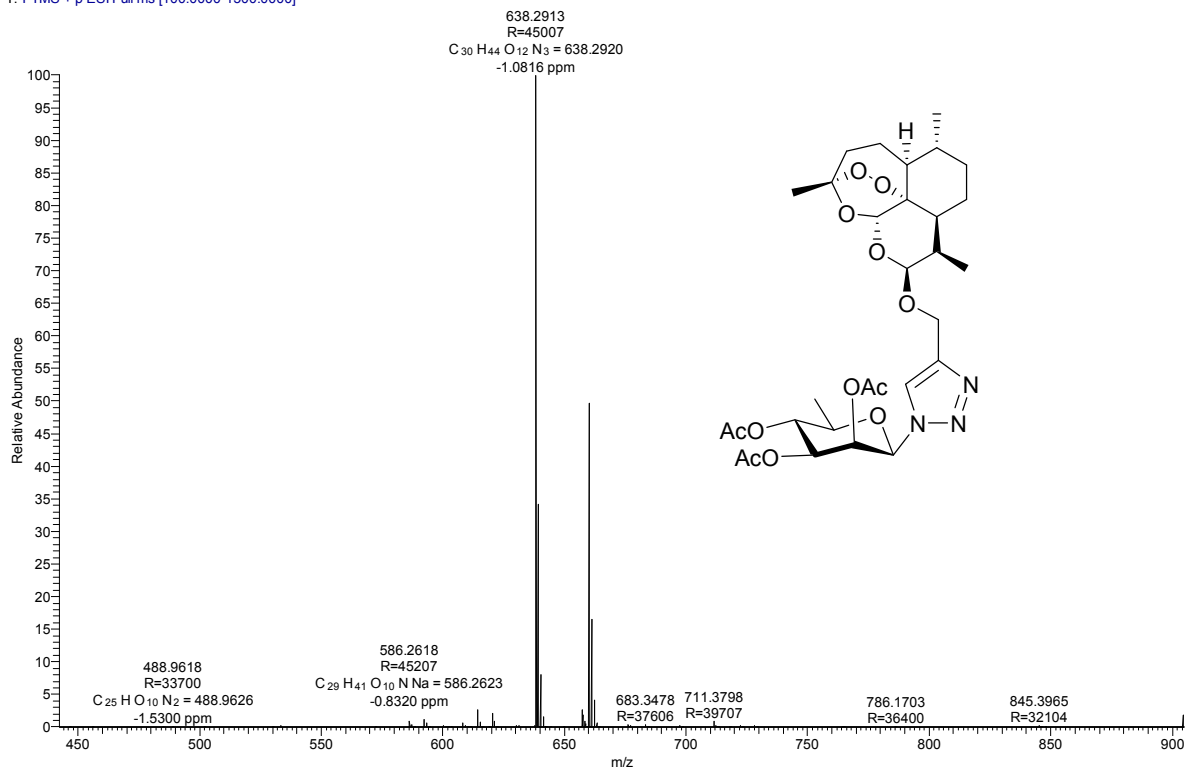


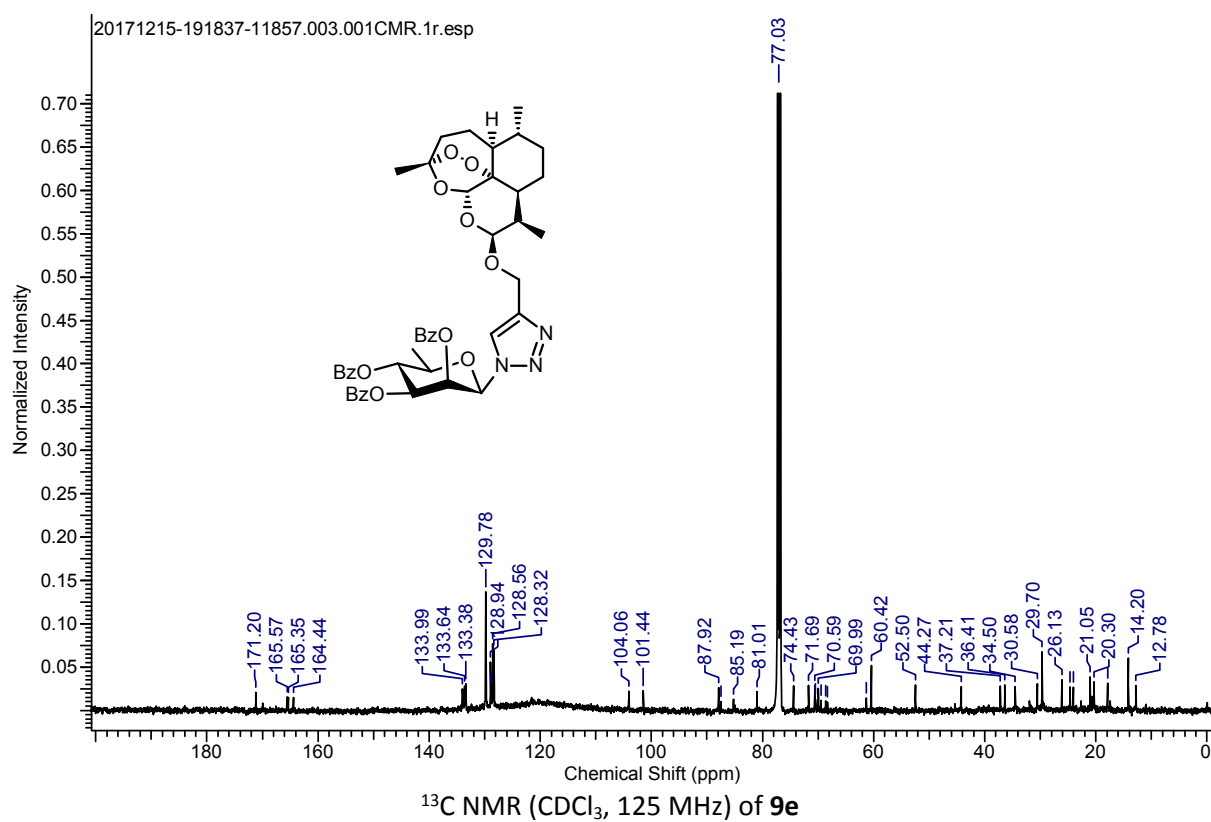
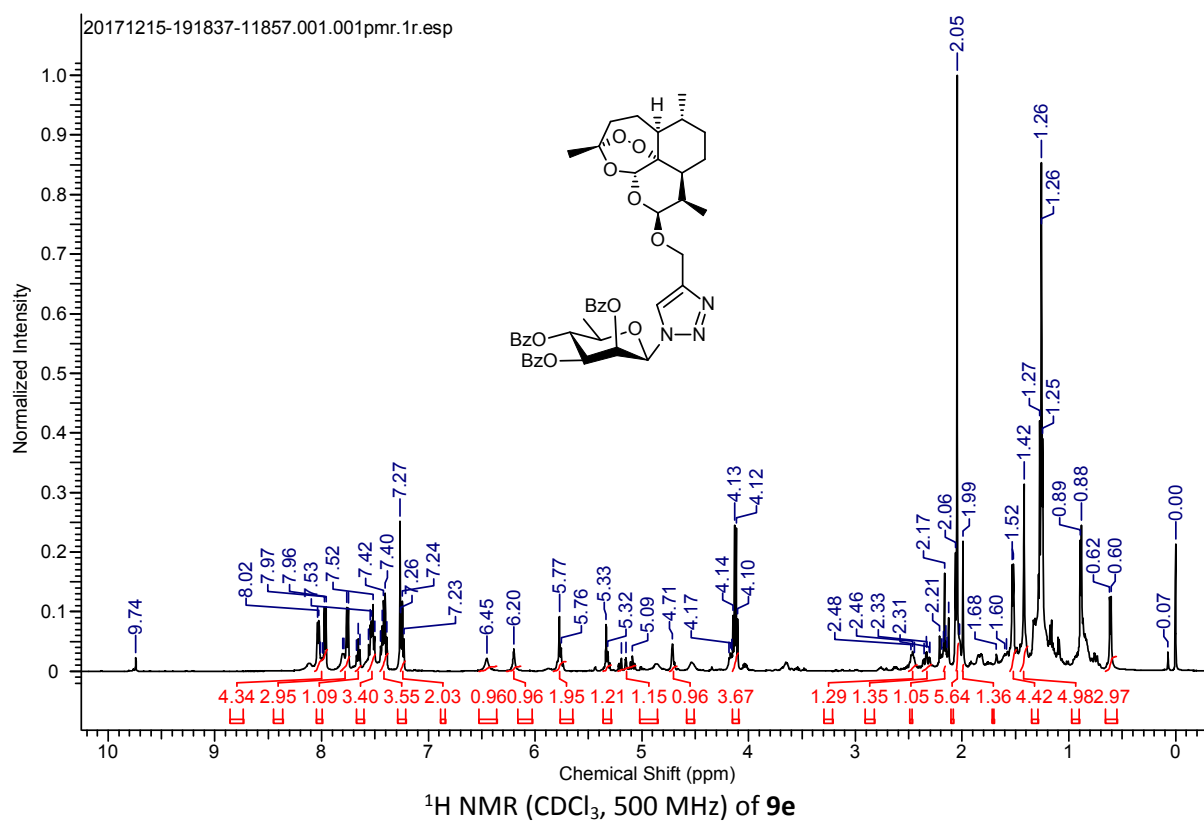
¹³C NMR (CDCl₃, 100 MHz) of **9c**



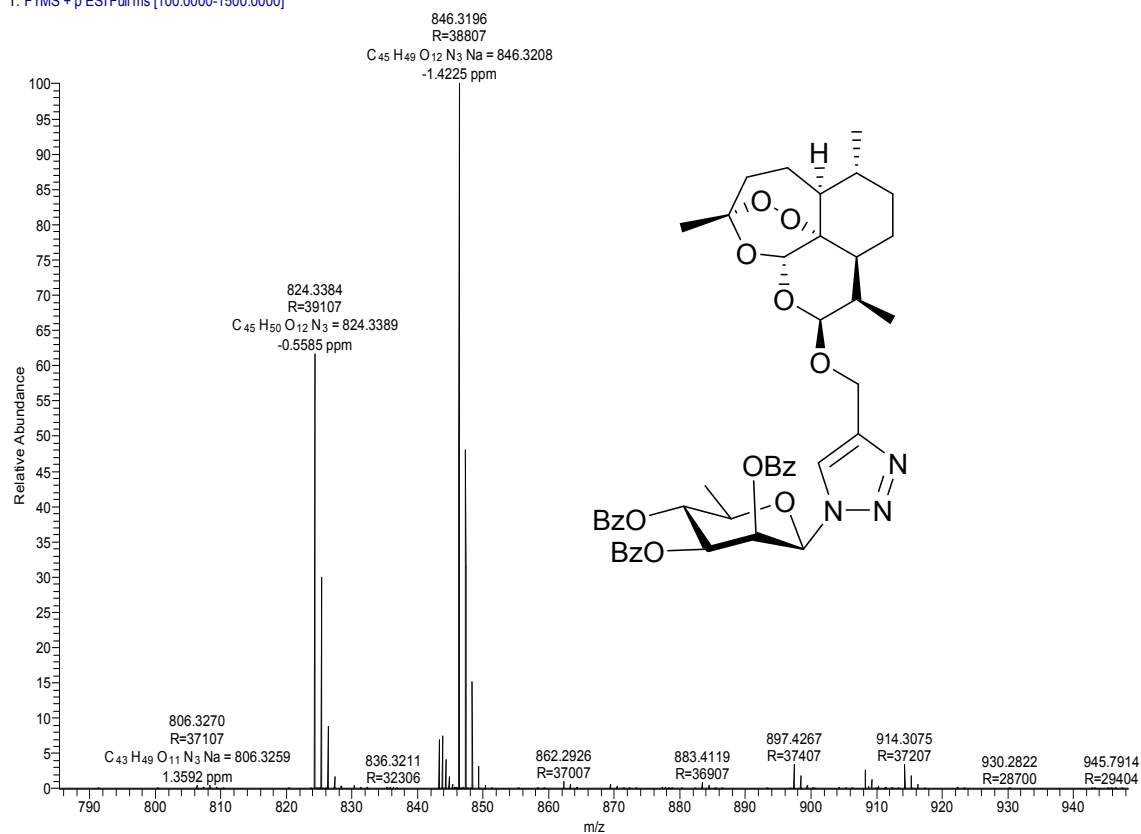


LG-12 #285 RT: 1.27 AV: 1 NL: 1.76E9
T: FTMS + p ESI Full ms [100.0000-1500.0000]



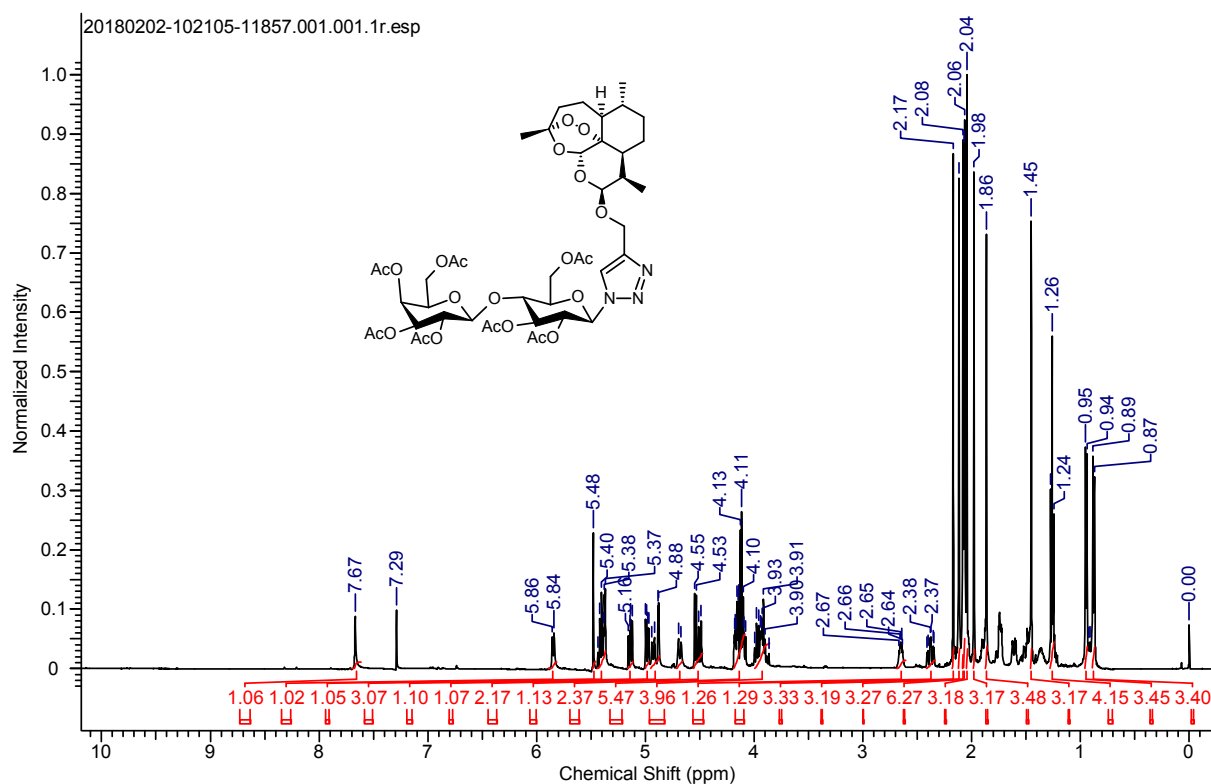


LG-13 #348 RT: 1.55 AV: 1 NL: 5.17E8
T: FTMS + p ESI Full ms [100.0000-1500.0000]

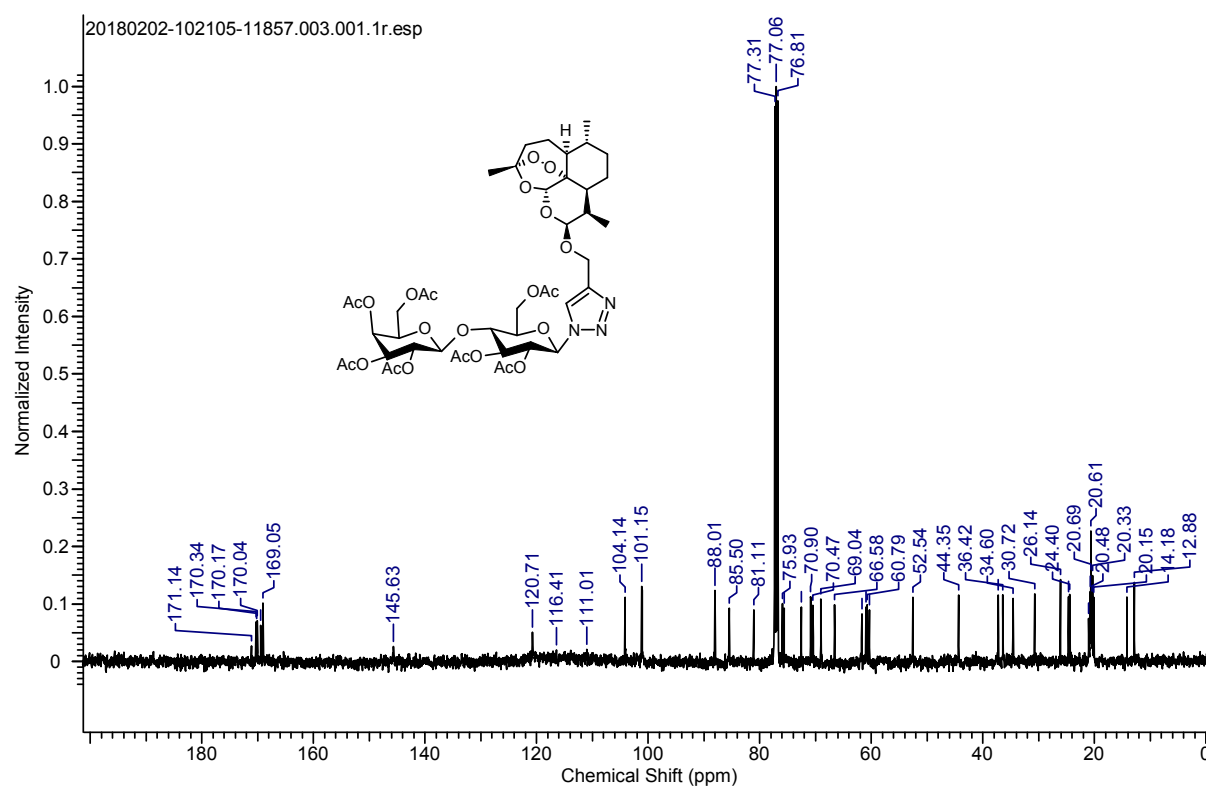


HR

MS of **9e**

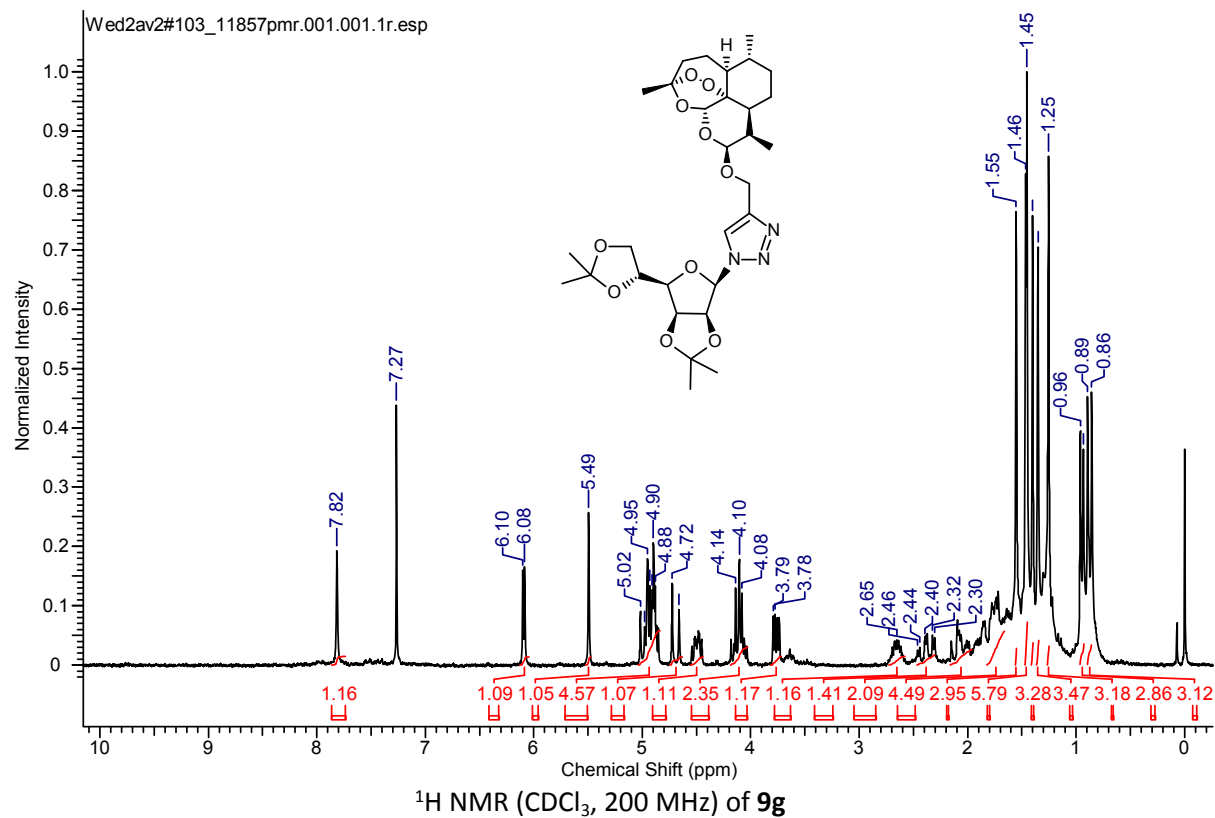
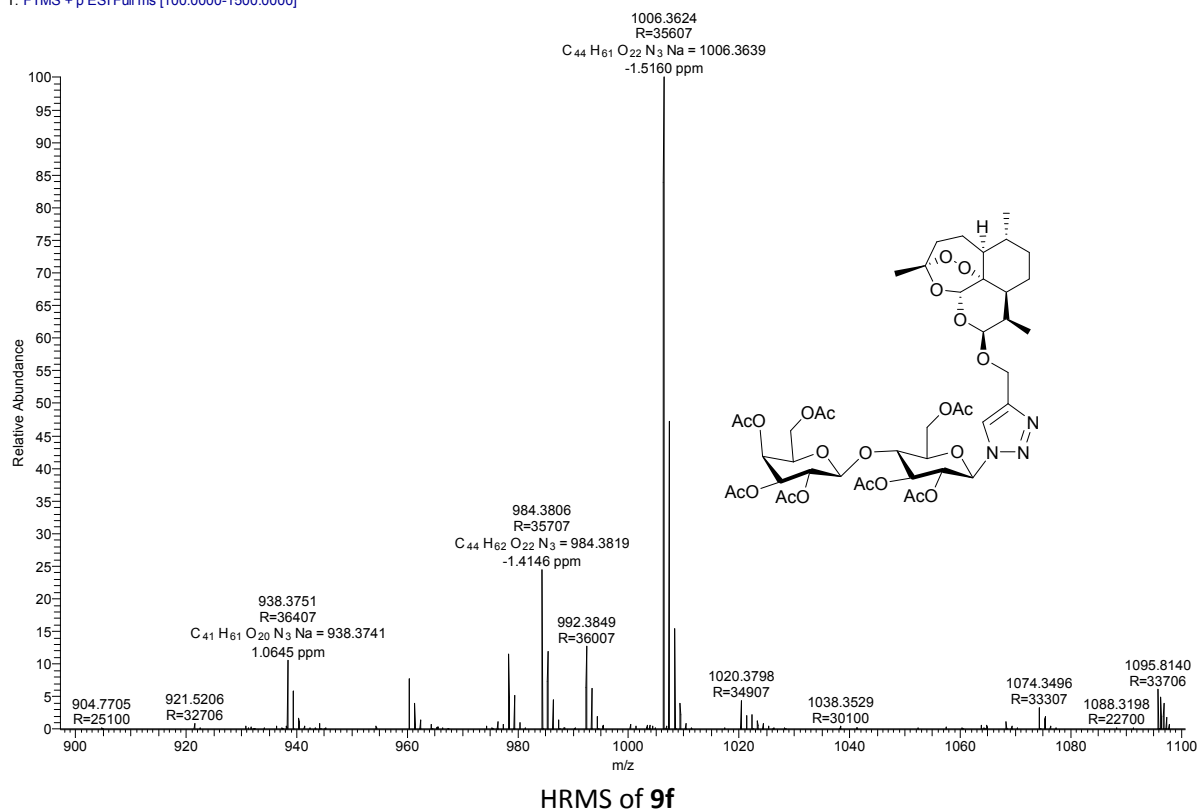


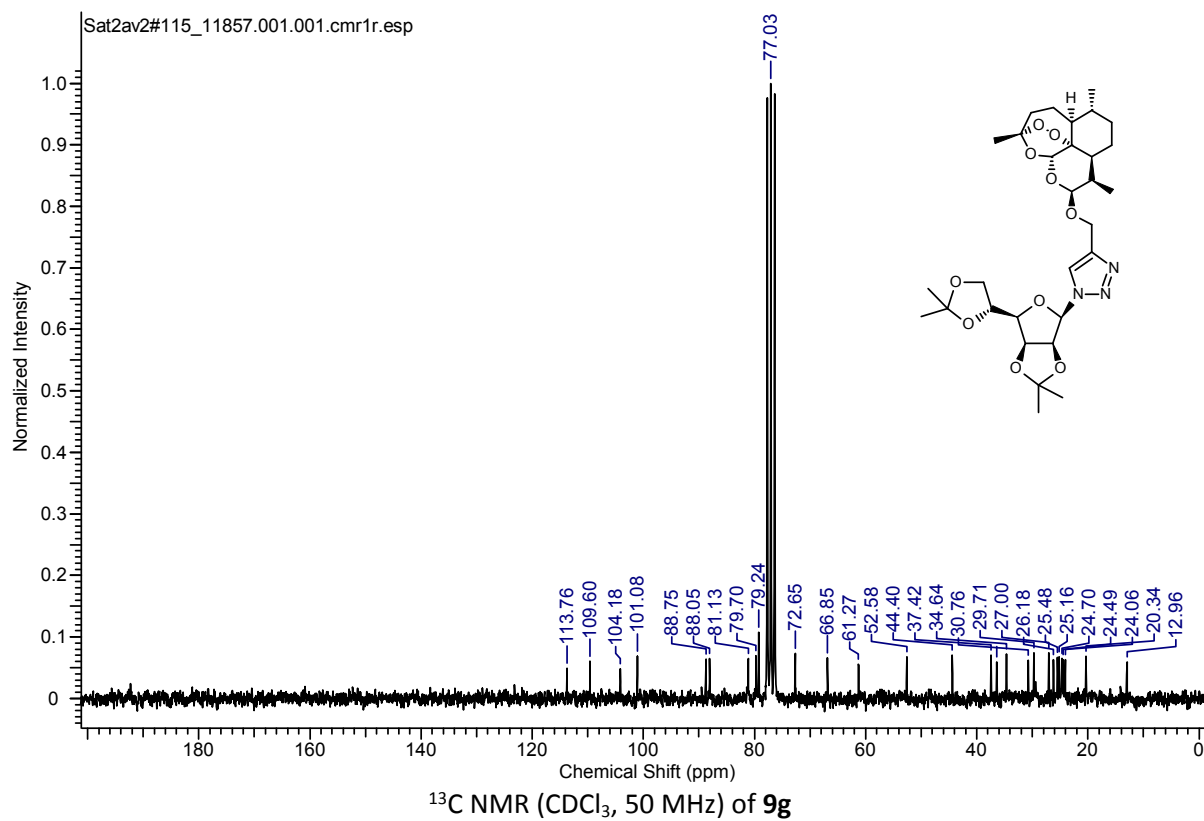
¹H NMR (CDCl₃, 500 MHz) of **9f**



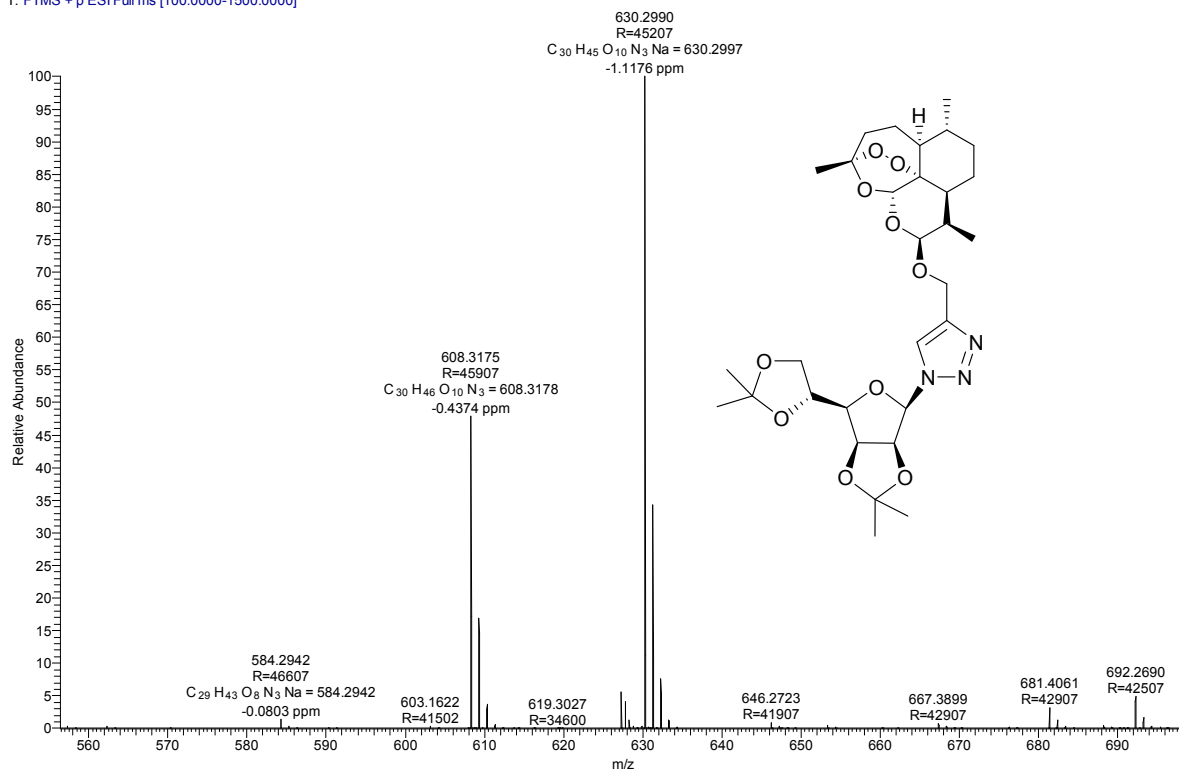
^{13}C NMR (CDCl_3 , 125 MHz) of **9f**

LG-14 #247 RT: 1.10 AV: 1 NL: 1.38E8
T: FTMS + p ESI Full ms [100.0000-1500.0000]

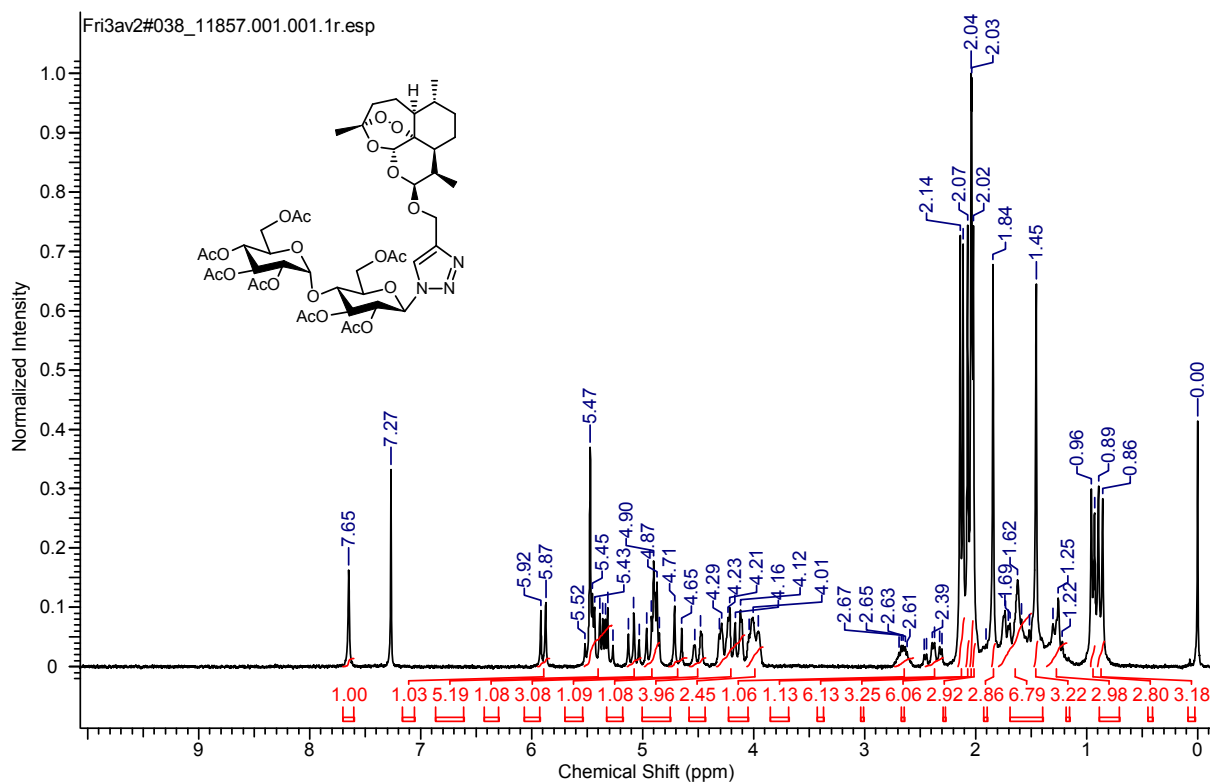




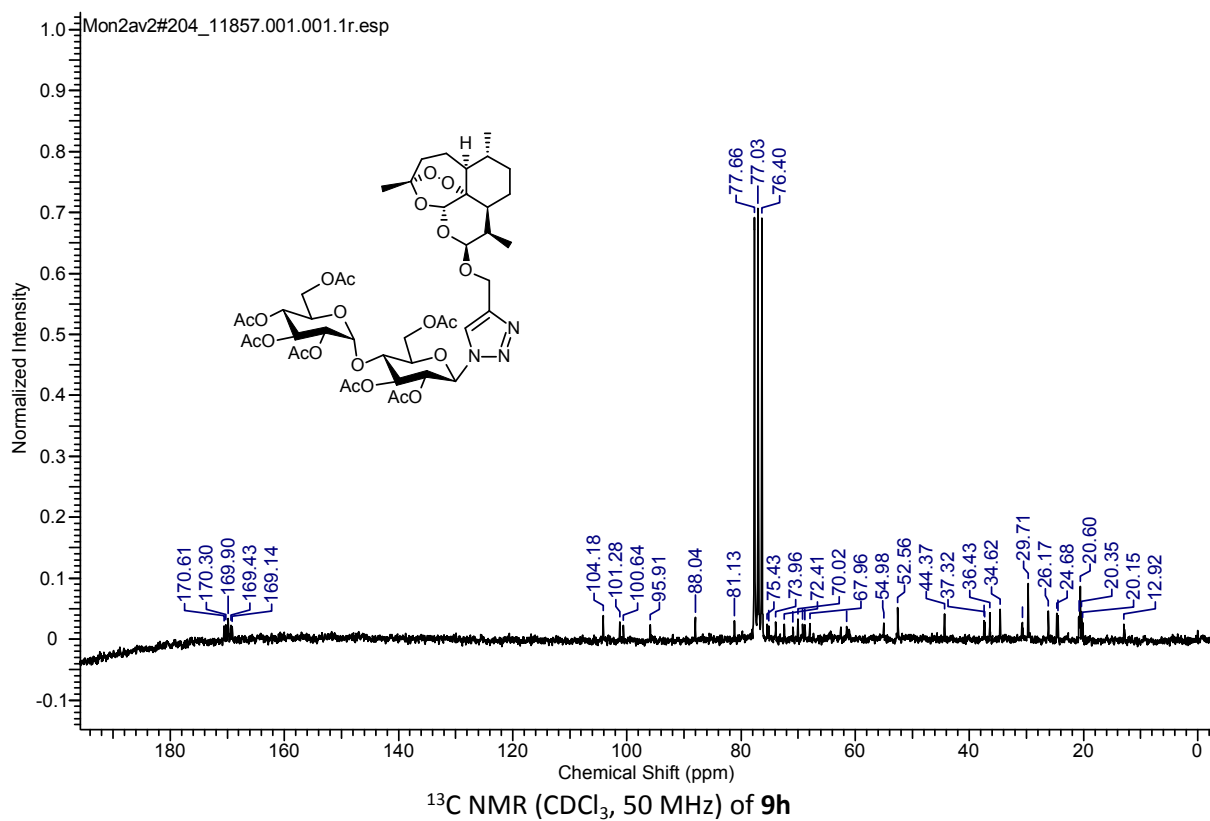
LG-17 #311 RT: 1.39 AV: 1 NL: 8.67E8
T: FTMS + p ESI Full ms [100.0000-1500.0000]



HRMS of **9g**

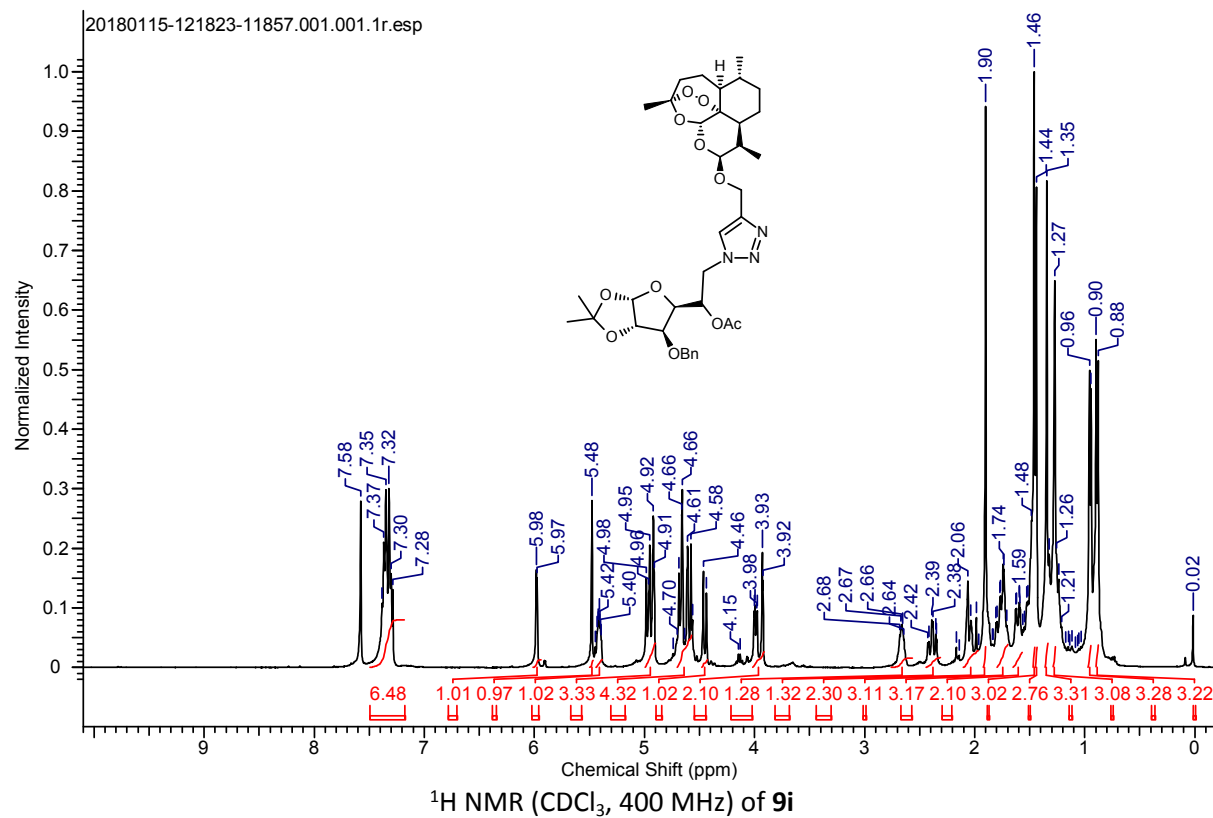
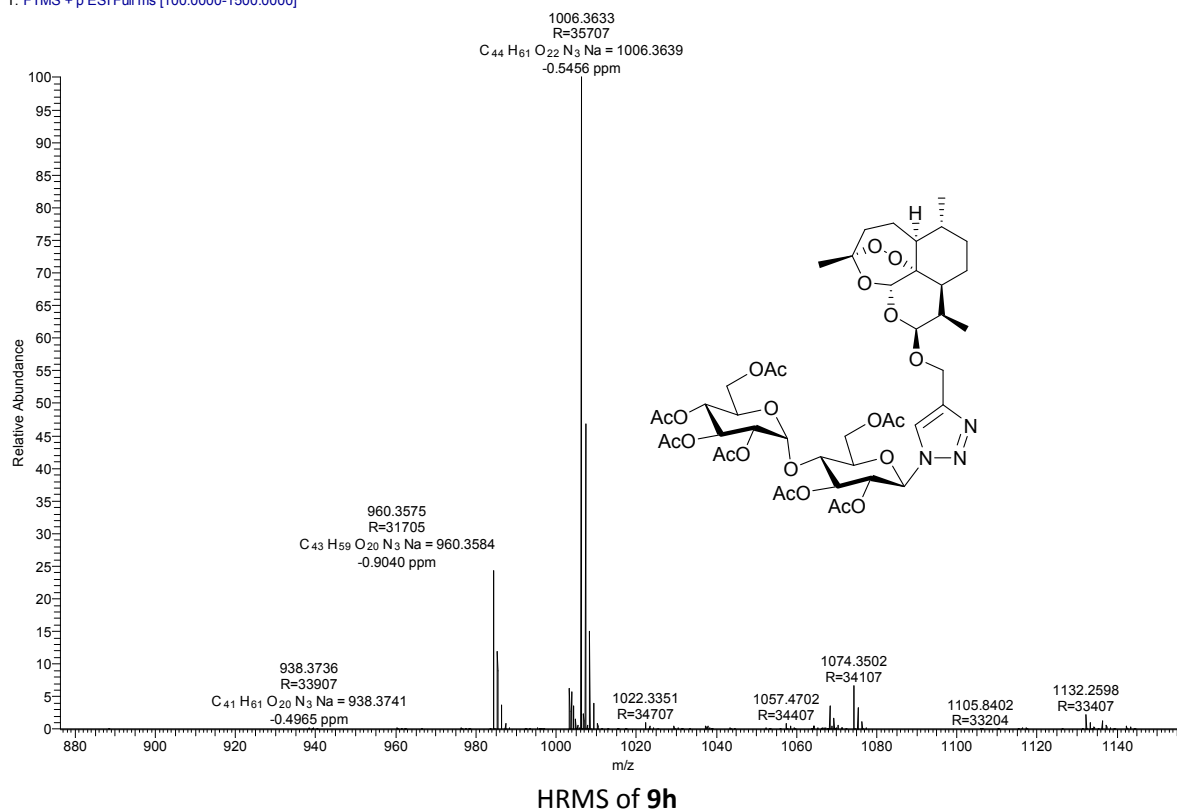


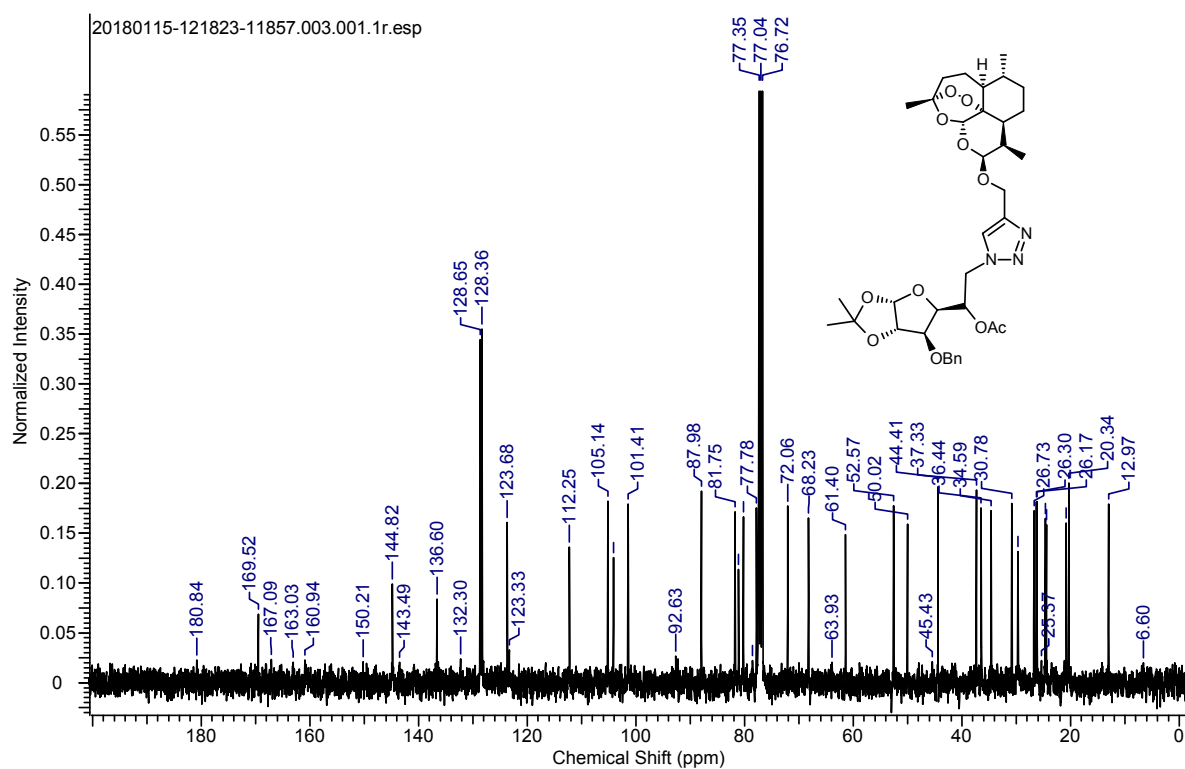
^1H NMR (CDCl_3 , 200 MHz) of **9h**



^{13}C NMR (CDCl_3 , 50 MHz) of **9h**

LG-19 #281 RT: 1.26 AV: 1 NL: 4.75E8
T: FTMS + p ESI Full ms [100.0000-1500.0000]





LG-23 #314 RT: 1.40 AV: 1 NL: 6.67E8
T: FTMS + p ESI Full ms [100.0000-1500.0000]

