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A straightforward and versatile FeCl₃ catalyzed Friedel-Crafts *C*glycosylation process. Application to the synthesis of new functionalized *C*nucleosides.

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Chemistry

General procedures.

All organic solvents were purchased from commercial sourcesand used as received or dried using standard procedures; all chemicals were purchased fromAldrich, Merck or Alfa Aesar and used without further purification.

Analytical thin layer-chromatographies (TLC) have been performed on pre-coated silica gel plates (Kieselgel 60 F_{254} , E. Merck, Germany), and chromatograms were visualized by UV- light irradiation (254 and 360 nm), then by staining with ninhydrin,*p*-anisaldehyde or H₂SO₄/EtOH. Purificationsby column chromatography have been performed using silica gel, 100-200 mesh (Merck, Germany).

NMR spectroscopies were recorded in dry deuterated solvent (DMSO or chloroform) on a Bruker AC 200, or on a Bruker AC 500 spectrometers at 200 MHz or 400 MHz for ¹H NMR and 50 MHz for ¹³C NMR; δ is expressed in ppm related to TMS (0 ppm) as internal standard. Splitting patterns are designated as follow: s (singlet), d (doublet), t (triplet), m (multiplet), br (broad). Coupling constants (*J* values) are listed in Hertz (Hz). Mass spectra (ESI-MS) were recorded on a Bruker Daltonics Esquire 3000+, and the samples were diluted in methanol.

General procedure for the Friedel-Craft ribosylation.

To a stirred solution of the carbohydrate (1'- β -ribofuranose-1',2',3',5'-tetraacetate, 1 mmol) and the appropriate (hetero)aryle (2 mmol) in dichloromethane (8 mL), was added at room temperature and in one portion FeCl₃ (10 mol%). The resulting mixture is heated in refluxing dichloromethane and the reaction is monitored by TLC. Once the carbohydrate fully reacted (10-30 min.), 10 mL of a saturated solution of NaHCO₃ was added to quench the reaction. The mixture is extracted three times using dichloromethane (15 mL). The organic layers were combined, then dried over MgSO₄ and concentrated under reduced pressure; the crude material is purified by flash chromatography using a mixture of ethyl acetate and hexane as solvent. Importantly, samples used for the fluorescence assay have been submitted to a second purification (preparative TLC) is order to strengthen their final purities.

List of the synthesized compounds.



Characterization of compounds 4a-4k and 5a-5e



4a Rf = 0.54 (Cyclohexane/AcOEt : 1-1); ¹H NMR (400 MHz, CDCl₃) δ 6.99 (bs, 1H), 6.70-6.62 (m, 2H), 5.74 (t, J = 4.2 Hz, 1H), 5.31 (t, J = 4.5 Hz, 1H), 5.17 (d, J = 3.7 Hz, 1H), 4.40 - 4.05 (m, 3H), 3.68 (s, 3H), 3.67 (s, 3H), 2.05 (s, 3H), 2.03 (s, 3H), 1.97 (s, 3H); ¹³C NMR (50 MHz, CDCl₃) δ 170.7, 169.6, 169.5, 153.7, 150.6, 127.5, 113.6, 113.1, 111.3, 79.3, 75.4, 72.0, 70.5, 63.1, 55.8, 55.5, 20.7, 20.6, 20.5 HRMS Calcd. for C₁₉H₂₅O₉: ([M+]): 397.1420, Found: 397.14931.



Rf = 0.73 (Cyclohexane/AcOEt : 1-1); ¹H NMR (200 MHz, CDCl₃) : δ 7.71 (d, *J* = 1.6 Hz, 1H), 7.47 (dd, *J* = 8.6, *J*' = 2.3 Hz, 1H), 6.50 (d, *J* = 8.6 Hz, 1H), 5.72 (dd, *J* = 4.3, *J*' = 3.3 Hz, 1H), 5.40 (d, *J* = 3.1 Hz, 1H), 5.34 (dd, *J* = 8.1, 4.4 Hz, 1H), 4.39 (t, *J* = 2.8 Hz, 1H), 4.36 – 4.30 (m, 1H), 4.12 (dd, *J* = 12.8, *J*' = 5.5 Hz, 1H), 3.70 (s, 3H), 2.06 (s, 3H), 1.98 (s, 3H), 1.74 (s, 3H); ¹³C NMR (50 MHz, CDCl₃) δ 170.7, 169.8, 169.3, 155.7, 137.4, 136.1, 126.8, 112.0, 82.3, 76.8, 76.3, 72.5, 71.8, 63.6, 55.4, 20.8, 20.5, 20.3; HRMS Calcd. for C₁₈H₂₂O₈I: ([M+]): 492.0281, Found: 493.03539.



4c Rf = 0.64 (Cyclohexane/AcOEt : 1-1) ; ¹H NMR (400 MHz, CDCl₃)δ 7.43 (d, J = 6.9 Hz, 1H), 7.23 (t, J = 7.6 Hz, 1H), 6.88 (t, J = 7.5 Hz, 1H), 6.72 (d, J = 8.1 Hz, 1H), 5.75 (t, J = 3.3 Hz, 1H), 5.39 – 4.86 (m, 2H), 4.40 – 4.18 (m, 2H), 4.13 (dd, J = 12.8, 5.6 Hz, 1H), 3.71 (s, 3H), 2.04 (s, 3H), 1.97 (s, 3H), 1.68 (s, 3H). ¹³C NMR (50 MHz, CDCl₃) δ 170.8, 169.8, 169.4, 155.8, 128.7, 127.3, 126.9, 120.0, 109.5, 79.7, 75.3, 72.7, 72.1, 63.7, 55.2, 20.8, 20.5, 20.3; HRMS Calcd. for C₁₈H₂₃O₈: ([M+]): 367.1315, Found: 397.13934.



40 Rf = 0.75(Cyclohexane/AcOEt : 1-1); ¹H NMR (200 MHz, CDCl₃) δ 7.55 (d, J = 2.6 Hz, 1H), 7.28 (dd, J = 8.7, J' = 2.5 Hz, 1H), 6.61 (d, J = 8.7 Hz, 1H), 5.73 (t, J = 3.0 Hz, 1H),

5.41 (d, J = 3.1 Hz, 1H), 5.34 (dd, J = 8.1, J' = 4.3 Hz, 1H), 4.42 – 4.21 (m, 2H), 4.12 (dd, J = 12.8, J' = 5.5 Hz, 1H), 3.70 (s, 3H), 2.06 (s, 3H), 1.98 (s, 3H), 1.74 (s, 3H); ¹³C NMR (50 MHz, CDCl₃) δ 169.8, 168.8, 168.4, 153.9, 130.3, 129.3, 125.5, 111.6, 110.4, 78.0, 75.8, 71.5, 70.9, 62.6, 54.5, 19.8, 19.5, 19.3 ; HRMS Calcd. for C₁₈H₂₂O₈Br: ([M+]): 445.0420, Found: 445.04926.



4e Rf = 0.71 (Cyclohexane/AcOEt : 1-1) ; 1H NMR (200 MHz, CDCl₃) δ 7.24 (d, J = 5.1 Hz, 1H), 6.88 (d, J = 5.3 Hz, 1H), 5.31–5.19 (m, 2H), 4.42-4.02 (m, 3H), 2.07 (s, 3H), 2.05 (s, 3H), 2.01 (s, 3H); 13C NMR (50 MHz, CDCl₃) δ 170.45, 169.68, 169.35, 136.20, 130.30, 126.04, 109.59, 79.96, 77.82 76.01, 71.56, 63.29, 20.80, 20.55, 20.44 ; HRMS Calcd. for C₁₅H₁₈O₇BrS: ([M+]): 420.9878, Found : 420.99533



4t Rf = 0.69 (Cyclohexane/AcOEt : 1-1); ¹H NMR (200 MHz, CDCl₃) δ 6.86 (d, *J* = 3.8 Hz, 1H), 6.76 (d, *J* = 3.8 Hz, 1H), 5.56 – 5.16 (m, 2H), 5.08 (d, *J* = 2.7 Hz, 1H), 4.38-4.10 (m, 3H), 2.07 (s, 3H), 2.05 (s, 3H), 2.02 (s, 3H) ; ¹³C NMR (50 MHz, CDCl₃) δ 169.5, 168.7, 168.6, 141.9, 128.7, 124.6, 111.7, 79.0, 77.7, 75.3, 70.6, 62.3, 19.8, 19.6, 19.5 ; HRMS Calcd. for C₁₈H₂₃O₈: ([M+]): 420.9878, Found: 420.99512.



4g Rf = 0.72 (Cyclohexane/AcOEt : 1-1); ¹H NMR (200 MHz, CDCl₃) δ 6.69 (s, 1H), 5.21 (t, J = 4.5 Hz, 1H), 5.12–5.00 (m, 2H), 4.33 (dd, J = 11.7Hz, J' = 2.8 Hz, 1H), 4.26 – 4.19 (m, 1H), 4.13 (dd, J = 11.7 Hz, J' = 3.7 Hz, 1H), 2.07 (s, 3H), 2.06 (s, 3H), 2.04 (s, 3H), 2.02 (s, 3H); ¹³C NMR (50 MHz, CDCl₃) δ 170.5, 169.7, 169.6, 140.6, 137.0, 127.2, 109.5, 79.8, 78.7, 76.2, 71.6, 63.3, 20.8, 20.5, 20.5, 15.2; HRMS Calcd. for C₁₆H₂₀O₇BrS: 435.0035, Found: 435.0112.



4h Rf = 0.71 (Cyclohexane/AcOEt : 1-1); ¹H NMR (200 MHz, CDCl₃) δ 7.05 (d, J = 3.7 Hz, 1H), 6.68 (d, J = 3.7 Hz, 1H), 5.22 (t, J = 4.6 Hz, 1H), 5.17–5.04 (m, 2H), 4.35 (dd, J = 11.7 Hz, J' = 2.8 Hz, 1H), 4.26-4.21 (m, 1H), 4.14 (dd, J = 11.7 Hz, J' = 3.7 Hz, 1H), 2.07 (s, 3H), 2.04 (s, 3H), 2.01 (s, 3H); ¹³C NMR (50 MHz, CDCl₃) δ 170.6, 169.7, 169.6, 147.3, 136.8, 126.8, 80.0, 75.5, 76.4, 71.6, 63.3, 20.9, 20.6, 20.5; HRMS Calcd. for C₁₅H₁₈O₇IS: 467.9740, Found: 468.9813.



41 Rf = 0.60 (Cyclohexane/AcOEt : 1-1); ¹H NMR (200 MHz, CDCl₃) δ 7.33 (d, J = 5.3 Hz, 1H), 7.15 (d, J = 5.3 Hz, 1H), 5.87 (t, J = 4.3, 3.8 Hz, 1H), 5.44 – 5.07 (m, 2H), 4.45 – 4.04 (m, 5H), 2.06 (s, 3H), 2.02 (s, 3H), 1.97 (s, 3H), 1.30 (t, J = 7.1 Hz, 3H); ¹³C NMR (50 MHz, CDCl₃) δ 170.7, 169.5, 169.1, 162.7, 148.8, 128.7, 128.3, 124.3, 78.3, 77.4, 72.3, 72.2, 63.3, 60.7, 20.8, 20.4, 20.2, 14.2; HRMS Calcd. for C₁₈H₂₃O₉: 415.0985, Found: 415.10583.



4j Rf = 0.69 (Cyclohexane/AcOEt : 1-1); ¹H NMR (200 MHz, CDCl₃) δ 7.36 (s, 1H), 6.36 – 6.20 (m, 2H), 5.43 (t, J = 5.5 Hz, 1H), 5.32 (t, J = 5.3 Hz, 1H), 4.94 (d, J = 5.6 Hz, 1H), 4.33 (dd, J = 11.6, 3.1 Hz, 1H), 4.26 – 4.17 (m, 1H), 4.09 (dd, J = 11.6, 4.5 Hz, 1H), 2.03 (s, 6H), 2.00 (s, 3H) ; ¹³C NMR (50 MHz, CDCl₃) δ 170.6, 169.7, 169.6, 150.0, 143.4, 110.4, 109.4, 79.2, 76.1, 73.3, 71. 7, 63.5, 20.7, 20. 6, 20.5 ; HRMS Calcd. for C₁₅H₁₉O₈: ([M+]): 327.1002, Found: 327.10744.



4k Rf = 0.61 (Cyclohexane/AcOEt : 1-1); ¹H NMR (200 MHz, CDCl₃)δ 7.93 (s, 1H), 6.69 (s, 1H), 5.40 (t, J = 5.5 Hz, 1H), 5.28 (t, J = 5.2 Hz, 1H), 4.92 (d, J = 5.7 Hz, 1H), 4.34 (dd, J = 11.7 Hz, J' = 3.1 Hz, 1H), 4.23 (q, J = 7.1 Hz, 3H), 4.10 (dd, J = 11.7 Hz, J' = 4.2 Hz, 1H), 2.06 (s, 3H), 2.05 (s, 3H), 2.01 (s, 3H), 1.27 (t, J = 7.1 Hz, 3H); ¹³C NMR (50 MHz, CDCl₃) δ 170.6, 169.7, 169.5, 162.7, 151.5, 148.0, 120.3, 109.3, 79.5, 75. 9, 73.2, 71.4, 63.3, 60.6, 20.7, 20.5, 20.5, 14.3; HRMS Calcd. for C₁₈H₂₃O₁₀: 399.1213, Found: 399.1281.



5a Rf = 0.69 (Cyclohexane/AcOEt : 1-1); ¹H NMR (400 MHz, CDCl₃) δ 8.08 (d, J = 8.7 Hz, 1H), 7.76 (d, J = 9.1 Hz, 1H), 7.68 (dd, J = 8.2 Hz, J' = 1.3 Hz, 1H), 7.39 (t, J = 7.8 Hz,

1H), 7.25 (t, J = 7.5 Hz, 1H), 7.18 (d, J = 9.0 Hz, 1H), 5.82 (d, J = 5.7 Hz, 1H), 5.72 (t, J = 6.1 Hz, 1H), 5.52 (t, J = 6.7 Hz, 1H), 4.38 (dd, J = 11.5 Hz, J' = 2.0 Hz, 1H), 4.26–4.19 (m, 2H), 3.90 (s, 3H), 2.03 (s, 3H), 2.00 (s, 3H), 1.94 (s, 3H); ¹³C NMR (50 MHz, CDCl₃) δ 170.7, 169.7, 169.4, 156.1, 132.8, 131.3, 123.3, 128.8, 127.0, 123.5, 122.7, 116.9, 113.5, 78.9, 77.8, 74.2, 70.9, 63.6, 56.5, 20.8 (x2), 20.6; HRMS Calcd. for C₂₅H₂₇O₇: 417.1471, Found: 417.15442.



 $\begin{array}{l} \textbf{Kf} = 0.73 \ (Cyclohexane/AcOEt: 1-1); \ ^1H \ NMR \ (200 \ MHz, CDCl_3) \ \delta \ 8.48-8.40 \\ (m, 3H), \ 7.99-7.92 \ (m, 2H), \ 7.47-7.36 \ (m, 4H), \ 6.35 \ (d, \textit{J} = 8.0 \ Hz, 1H), \ 5.68-5.62 \ (m, 2H), \ 4.56 \ (dd, \textit{J} = 12.2, \textit{J}' = 2.7 \ Hz, 1H), \ 4.46 \ (dd, \textit{J} = 12.3, \textit{J}' = 3.6 \ Hz, 1H), \ 4.32 \ (m, 1H), \ 2.21 \ (s, 3H), \ 2.12 \ (s, 3H), \ 1.84 \ (s, 3H); \ ^{13}C \ NMR \ (50 \ MHz, CDCl_3) \ \delta \ 170.6, \ 169.7, \ 169.4, \ 138.8, \ 131.5, \ 130.5, \ 129.8, \ 129.5, \ 127.8, \ 127.7, \ 127.4, \ 126.2, \ 126.1, \ 124.8, \ 124.1, \ 123.4, \ 121.1, \ 81.8, \ 78.8, \ 74.7, \ 70.3, \ 63.3, \ 20.9, \ 20.6, \ 20.3; \ HRMS \ Calcd. \ for \ C_{25}H_{27}O_7; \ 437.1522, \ Found: \ 437.15958. \end{array}$



5c Rf = 0.31 (Cyclohexane/AcOEt : 8-2) ; ¹H NMR (200 MHz, CDCl₃) δ 8.32 – 8.29 (m, 1H), 8.25 – 8.20 (m, 4H), 8.15 – 8.12 (m, 1H), 8.11 – 8.07 (m, 2H), 8.06 – 8.01 (m, 1H), 6.12 (d, J = 6.5 Hz, 1H), 5.45 (m, 2H), 4.72 – 4.33 (m, 3H), 2.20 (s, 3H), 2.18 (s, 3H), 2.06 (s, 3H); ¹³C NMR (50 MHz, CDCl₃) δ 170.9, 169.9, 169.6, 131.5, 131.5, 131.1, 130.7, 128.6, 128.0, 127.9, 127.6, 126.2, 125.7, 125.4, 125.3, 124.9, 124.8, 123.2, 122.4, 80.2, 79.3, 71.9, 63.8, 53.6, 21.1, 20.9, 20.7; HRMS Calcd. for C₂₇H₂₄O₇: ([M+]): 461.1522, Found [M+Na⁺]: 483.14160.



4d Rf = 0.70 (Cyclohexane/AcOEt : 1-1); ¹H NMR (400 MHz, CDCl₃) δ 7.99 (d, J = 8.4 Hz, 1H), 7.78 (d, J = 9.2 Hz, 2H), 7.41–7.38 (m, 3H), 7.31 (t, J = 7.8 Hz, 2H), 7.07 (t, J = 7.9 Hz, 2H), 6.73 – 6.78 (m, 3H) , 6.03 (d, J = 8.5 Hz, 1H), 4.95 (dd, J = 5.5 Hz, J' = 1.9 Hz, 1H), 4.45 (t, J = 7.0 Hz, 1H), 4.14 (bs), 4.00 (d, J = 3.4 Hz, 2H), 2.00 (s, 3H) , 1.99 (s, 3H) 1.84 (s, 3H); ¹³C NMR

(50 MHz, CDCl₃) δ 170.4, 169.8, 169.0, 148.8, 137.7, 134.8, 133.2, 129.4, 129.1 (x2), 128.6, 128.3, 126.4 (x2) 126.0, 124.3, 120.1, 116.1 (x2), 89.2, 78.2, 71.0, 70.6, 63.9, 20.7, 20.4, 20.4; HRMS Calcd. for C₂₇H₂₈O₇N: 478.1788, Found: 478.1859.



5e Rf = 0.61 (Cyclohexane/AcOEt : 1-1); ¹H NMR (200 MHz, CDCl₃) δ 7.59– 7.32 (m, 2H), 7.27–7.10 (m, 2H), 6.71 (s, 1H), 5.54 (t, J = 5.4 Hz, 1H), 5.37 (t, J = 5.3 Hz, 1H), 5.08 (d, J = 5.4 Hz, 1H), 4.39 (dd, J = 11.6 Hz, J' = 3.2 Hz, 1H), 4.33 – 4.25 (m, 1H), 4.14 (dd, J = 11.7 Hz, J'= 4.4 Hz, 1H), 2.04 (s, 3H), 2.02 (s, 3H), 2.00 (s, 3H); ¹³C NMR (50 MHz, CDCl₃) δ 170.6, 169.7, 169.6, 155.2, 152.6, 127.6, 124.9, 123.0, 121.3, 111.5, 106.0, 79.6, 79.5, 73.4, 71.7, 63.4, 20.8, 20.6, 20.5. HRMS Calcd. for C₁₉H₂₁O₈: 377.1158, Found: 377.1231.

Photophysics

General procedures.

All solvents for absorption and fluorescence experiments were of spectroscopic grade. Absorption spectra were recorded on a Cary 4 spectrophotometer (Varian) using quartz cells of 1 cm path length. Fluorescence spectra were recorded on FluoroMax 4.0 spectrofluorometer (Jobin Yvon, Horiba). Wavelengths used as the excitation wavelengths are highlighted in table 3. Stock solutions of the solvatofluorochromic dyes were prepared using 1,4- doxane. The samples used for spectroscopic measurements contained $\approx 0.1\%$ v/v of the stock solvent. For fluorescence quantum yield measurements, the absorbance of the corresponding solution was kept below 0.05.

Copies of ¹H and ¹³C NMR spectra



¹H NMR (400 MHz, CDCl₃) spectrum of **4a**.



 ^{13}C NMR (50 MHz, CDCl₃) spectrum of **4a**.



 $^1\mathrm{H}$ Noesy correlation for $\beta\text{-configuration}.$



Cosy spectrum of 4a.



Noesy spectrum of 4a.



Noesy spectrum of 4a (4 - 6 ppm).



Selective noesy spectrum of 4a (5.25 ppm, β -anomer).



¹H NMR (200 MHz, CDCl₃) spectrum of **4b**.



 ^{13}C NMR (50 MHz, CDCl₃) spectrum of **4b**.



¹H NMR (400 MHz, CDCl₃) spectrum of **4c**.



 ^{13}C NMR (50 MHz, CDCl₃) spectrum of 4c.



¹H NMR (200 MHz, CDCl₃) spectrum of **4d**.



¹³C NMR (50 MHz, CDCl₃) spectrum of **4d**.



¹H NMR (200 MHz, CDCl₃) spectrum of **4e**.



 ^{13}C NMR (50 MHz, CDCl₃) spectrum of 4e.



 1 H NMR (200 MHz, CDCl₃) spectrum of **4f**.



¹³C NMR (50 MHz, CDCl₃) spectrum of **4f**.



¹H NMR (200 MHz, CDCl₃) spectrum of 4g.



 ^{13}C NMR (50 MHz, CDCl₃) spectrum of 4g.



¹H NMR (200 MHz, CDCl₃) spectrum of **4h**.



¹³C NMR (50 MHz, CDCl₃) spectrum of **4h**.



¹H NMR (200 MHz, CDCl₃) spectrum of **4i**.



¹³C NMR (50 MHz, CDCl₃) spectrum of **4i**.



¹H NMR (200 MHz, CDCl₃) spectrum of **4j**.



 ^{13}C NMR (50 MHz, CDCl₃) spectrum of 4j.



¹H NMR (200 MHz, CDCl₃) spectrum of **4k**.



 ^{13}C NMR (50 MHz, CDCl₃) spectrum of 4k.



¹H NMR (400 MHz, CDCl₃) spectrum of **5a**.



 ^{13}C NMR (50 MHz, CDCl₃) spectrum of **5a**.



COSY spectrum of 5a.



NOESY spectrum of 5a.



¹H NMR (200 MHz, CDCl₃) spectrum of **5b**.



 ^{13}C NMR (50 MHz, CDCl₃) spectrum of **5b**.



 1 H NMR (400 MHz, CDCl₃) spectrum of **5c**.



¹³C NMR (101 MHz, CDCl₃) spectrum of **5c**.



 1 H NMR (400 MHz, CDCl₃) spectrum of **5d**.



¹³C NMR (50 MHz, CDCl₃) spectrum of **5d**.



¹H NMR (200 MHz, CDCl₃) spectrum of **5**e.



¹³C NMR (50 MHz, CDCl₃) spectrum of **5e**.

Copies of HRMS spectra



HRMS (ESI-MS) spectrum of 4a.



HRMS (ESI-MS) spectrum of 4b.



HRMS (ESI-MS) spectrum of 4c.



HRMS (ESI-MS) spectrum of 4d.



HRMS (ESI-MS) spectrum of 4e.



HRMS (ESI-MS) spectrum of 4f.



HRMS (ESI-MS) spectrum of 4g.



HRMS (ESI-MS) spectrum of 4h.



HRMS (ESI-MS) spectrum of 4i.



HRMS (ESI-MS) spectrum of 4j.



HRMS (ESI-MS) spectrum of 4k.







HRMS (ESI-MS) spectrum of **5b**.







HRMS (ESI-MS) spectrum of 5d.







Figure S1. Fluorescence emission spectra of **5c** in different concentrations in cyclohexane.