

Electronic Supplementary Information (ESI)

## Heterologous expression of bacterial natural product biosynthetic pathways

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**Table S1. Selected examples of heterologous expression of bacterial BGCs from 2013 to 2018**

<sup>§</sup>Method: Heterologous expression method/s employed. Plas, Self-replicating plasmid; Hom, homologous recombination with insertion locus indicated in brackets; Tnps, Transposition; Int, Phage integration. *S. Streptomyces*; *Sti. Stigmatella*; *Ps. Pseudomonas*. <sup>†</sup>Yield: Yields relative to the native producer,  $\infty$  indicates the compound was unable to be isolated from the native host. Actual yield from heterologous host in brackets. Comments: Promoters listed replaced the native promoter of the cluster. Where yields from the native producer were not reported in the expression paper, the approximate levels were determined from the literature

Compound	Native host	Size	Cloning method	Heterologous host	Method <sup>§</sup>	Yield <sup>†</sup>	Comments *	Ref.
<b>Known</b>								
Abyssomicin / Neoabyssomicin 5802	<i>S. koyangensis</i> SCSIO	62.9	PAC library	<i>S. coelicolor</i> M1152	Int	n.d.	Native cluster	1
Actinoallolides	<i>Actinoallomurus fulvus</i> K09-0307	53	Cosmid library	<i>S. coelicolor</i> M1152	Int	n.d.	<i>P ermE*</i> promoter	2
Actinorhodin	<i>S. coelicolor</i> M145	26	Fosmid library	<i>S. albus</i> J1074	Int	n.d.	Gene deletion of <i>pfk</i> gene encoding phosphofructokinase, overexpression of <i>crp</i> gene encoding cAMP receptor protein from <i>S. coelicolor</i>	3
Aloesaponarin II	<i>Frankia</i> sp. CcI3		Fosmid library	<i>S. albus</i> Del14	Int	n.d.	Engineered heterologous host	4
Alterochromides (A 22)	<i>Pseudoalteromonas piscicida</i> JCM 20779	34	TAR	<i>E. coli</i> BL21	Plas	n.d.	<i>Pseudoalteromonas luteoviolacea</i> 2ta16 coproduced, KBr supplemented in media	5
Amicoumacin A 24	<i>Bacillus subtilis</i> 1779	47.4	TAR	<i>B. subtilis</i> JH6421	Hom ( <i>amyE</i> )	similar	<i>lac</i> repressor <i>lacI</i> and an IPTG-inducible promoter, <i>sfp</i> (PPTase) co-expression	6
Alpiniamides	<i>S. sp.</i> 2014/011-12	47.6	TAR	<i>S. lividans</i> TK24 and <i>S. albus</i> Del14	Int	n.d.	Native cluster	7
Avermectins	<i>S. avermitilis</i> ATCC 31267	81	BAC library	<i>S. lividans</i>	Int	n.d.	Native cluster	8
Bacillomycin	<i>B. amyloliquefaciens</i> FZB42	37.2	LLHR	<i>B. subtilis</i> 1A751	Hom ( <i>amyE</i> )	n.d.	Sfp (PPTase) rescued in host	9
Bengamides	<i>Myxococcus virescens</i> ST200611	25	Cosmid library	<i>Myxococcus xanthus</i> DK1622	Int	similar	Native cluster	10
Bhimamycin A	<i>Frankia</i> sp. CcI3		BAC library	<i>S. albus</i> Del14	Int	n.d.	Engineered heterologous host	4
Bicyclomycin	<i>Ps. aeruginosa</i> SCV20265 / <i>S.</i>	7	PCR + Gibson assembly	<i>Pseudomonas fluorescens</i> SBW25 / <i>S.</i>	Plas / Int	n.d.	Synthetic <i>P tac</i> promoter / <i>P ermE*</i> promoter	11

	<i>cinnamoneus</i> DSM 41675	<i>coelicolor</i> M1146/1152						
BM1157/BM1300	<i>Lactobacillus crustorum</i> MN047	0.2	PCR and ligation	<i>E. coli</i> BL21	Plas	n.d.	T7 promoter	<sup>12</sup>
Brasilicardin	<i>Nocardia terpenica</i> IFM 0406	13.1	Fosmid library	<i>Amycolatopsis japonicum</i>	Int	n.d.	<i>P ermE*</i> promoter	<sup>13</sup>
Bottromycin A2 <b>7</b>	<i>S. sp.</i> BC16019	16	Cosmid library	<i>S. albus</i> J1074 and <i>S. lividans</i> TK24	Int	5–50 fold higher	Optimized the expression of genes in operons and created a library of modified gene clusters	<sup>14</sup>
Cacibiocin A/B	<i>Catenulispora acidiphila</i> DSM 44928	16.6	Cosmid library	<i>S. coelicolor</i> M1152	Int	↑ 12 fold (60 mg L <sup>-1</sup> )	Native cluster	<sup>15</sup>
Chaxamycin	<i>S. leeuwenhoekii</i>	80.2	PAC library	<i>S. coelicolor</i> M1152	Int	n.d.	<i>P ermE</i> promoter	<sup>16</sup>
Chuangxinmycin <b>25</b>	<i>Actinoplanes tsinanensis</i>	11	LLHR	<i>Streptomyces</i> spp	Int	n.d.	Strong artificial promoter Virolle-a1-14	<sup>17</sup>
Clethramycin/mediomycin A	<i>S. mediocidicus</i> ATCC23936	180	BAC library	<i>S. lividans</i> TK24	Int	n.d.	<i>P ermE</i> promoter	<sup>18</sup>
Corallopyronin A <b>16</b> Corallopyronin B <b>17</b>	<i>Corallococcus coralloides</i> B035	65	Cosmid library	<i>Myxococcus xanthus</i> DK1622	Int	↑ ca. 30 fold (37 mg L <sup>-1</sup> )	<i>P npII</i> promoter and media optimization	<sup>19</sup>
Cosmomycin (C <b>23</b> )	<i>S. sp.</i> CNT-302	54	TAR	<i>S. coelicolor</i> M512	Int	n.d.	Native cluster	<sup>20</sup>
Dawenol <b>20</b>	<i>Sti. aurantiaca</i> DW4/3-1	21	Subcloning via plasmid recovery	<i>Myxococcus xanthus</i> DK1622	Int	n.d.	Native cluster	<sup>21</sup>
6-Deoxyerythronolide B	<i>Saccharopolyspora erythraea</i>	30	Cosmid library	<i>Bacillus subtilis</i> JK13	Int	↓ 2.6 µg L <sup>-1</sup>	Optimized RBS, <i>P acoA</i> promoter, deletion of the <i>prpBD</i> operon responsible for propionyl-CoA degradation, feeding propionate precursor, <i>sfp</i> (PPTase) co-expression	<sup>22</sup>
Desotamides	<i>S. scopoliridis</i> SCSIO ZJ46	39	Cosmid library	<i>S. coelicolor</i> M1152	Int	n.d.	Native promoter	<sup>23</sup>
2,4-Diacetylphloroglucinol	<i>Ps. protegens</i> Pf-5 / <i>Ps. sp.</i> G22	3.8	PCR and ligation	<i>Pseudomonas</i> sp. WS5	Plas	↑ 7-fold (12–14 µg m L <sup>-1</sup> )	<i>P lac</i> promoter	<sup>24</sup>
Disorazol A1 Disorazol A2	<i>Sorangium cellulosum</i> So ce12	58	BAC library	<i>Myxococcus xanthus</i> DK1622	Tpns	↑ 12-fold (0.7 mg L <sup>-1</sup> )	<i>P tet</i> promoter, synthetic <i>P cp25</i> promoter	<sup>25</sup>

Enterocin A	<i>Enterococcus faecium</i> T136	0.5	PCR and ligation	<i>Lactobacillus</i> spp.	Plas	↑ 4.9-fold (9.3 µg mg⁻¹ cell dry weight)	Signal peptide SP <sub>usp45</sub> fused to EntA, inducible promoter	26
Enterocin	<i>Salinispora pacifica</i> CNT-150	18	TAR	<i>S. coelicolor</i> M1146 / <i>S. lividans</i> TK23	Int	n.d.	Native cluster	27
Epothilone A 9 Epothilone B 10	<i>Sorangium cellulosum</i> So ce90	56	Cosmid library + LCHR assembly	Burkholderiales sp. DSM 7029	Tnps	↑ 75-fold (0.3 mg L⁻¹)	<i>P<sub>Tn5-kan</sub>/P<sub>tet</sub></i> promoter, introduction of propionyl-CoA carboxylase (PCC) encoding genes from <i>Streptomyces coelicolor</i> A3(2), introduction of rare tRNA genes	28
Epothilone A 9 Epothilone B 10	<i>Sorangium cellulosum</i> So0157-2	56	Cosmid library + LCHR assembly	<i>Myxococcus xanthus</i>	Int	↑ (21.8 mg L⁻¹)	Tandem repeat engineering on the original promoter <i>P<sub>epo</sub></i>	29, 30
FK506	<i>S. tsukubaensis</i> NRRL 18488	83.5	PAC library	<i>S. coelicolor</i> M1146	Int	n.d.	<i>ermE*</i> promoter, overexpression of the LuxR regulatory gene <i>fkbN</i>	31
Fostriecin	<i>S. pulveraceus</i> ATCC 31906	48.4	Cosmid library + LCHR assembly	<i>S. lividans</i> TK24	(2 parts) Int and Plas	n.d.	<i>P<sub>actI</sub></i> promoter	32
FR900359 26	<i>Burkholderia crenata</i>	11.7	BAC library	<i>E. coli</i> BL21 and BAP1	Plas	n.d.	Constitutive promoter <i>pS7</i>	33
Galbonolide	<i>S. sp.</i> LZ35	11.8	PCR and ligation	<i>S. coelicolor</i> ZM12	Int	n.d.	<i>P<sub>ermE*</sub></i> promoter	34
Glidobactin	Burkholderiales sp. DSM7029	25	LLHR	<i>E. coli</i> Nissle 1917	Plas	n.d.	<i>P<sub>tet</sub>/P<sub>BAD</sub></i> promoter	35
Grecocycline	<i>S. sp.</i> Acta 1362	36	PCR + TAR assembly	<i>S. albus</i> J1074	Int	n.d.	Mutations in genes due to PCR, detect several biosynthetic intermediate	36
Haliangicin	<i>Haliangium ochraceum</i> SMP-2	47.8	Cosmid library	<i>M. xanthus</i>	Int	↑ 10-fold higher	Native cluster	37
Hapalosin 5	<i>Fischerella</i> sp. PCC 9431	23	DiPaC + SLIC	<i>E. coli</i> BAP1	Plas	n.d.	T7 promoter, <i>sfp</i> (PPTase) co-expression	38
Herbicidin	<i>S. sp.</i> L-9-10	26	Cosmid library	<i>S. albus</i> J1074	Int	n.d.	Native cluster	39
Ikarugamycin A 21	<i>S. sp.</i> Tü 6239	33	Fosmid library	<i>E. coli</i> BAP1	Plas	n.d.	T7 promoter, <i>sfp</i> (PPTase) co-expression	40
Kinamycin	<i>S. galtieri</i> Sgt26	75	BAC library	<i>S. albus</i> J1074	Int	n.d.	Native cluster	41
Kocurin 28	<i>Kocuria flava</i> HO-9041	12	PCR and Gibson	<i>S. coelicolor</i> M1146	Int	n.d.	<i>P<sub>ermE*</sub></i> promoter	42

assembly								
Lactococcin Z	<i>Lactococcus lactis</i> QU7	5.1	PCR and ligation	<i>Lactococcus lactis</i> NZ9000	Plas	n.d.	Native cluster	43
Lasso peptides	Marine <i>Actinobacteria</i>	2–3	PCR and Gibson assembly	<i>Streptomyces</i> spp.	Int	n.d.	<i>P<sub>A127-LP</sub></i> promoter, co-expression of gene encoding <i>Streptomyces</i> antibiotic regulatory protein (SARP)	44
Lyngbyatoxin A <b>12</b>	<i>M. producens</i>	11.3	TAR/Red-ET recombineering	<i>Anabaena</i> sp. PCC 7120	Plas	↑ 13-fold	<i>P<sub>glnA</sub></i> promoter and media optimization	45
Matlystatin	<i>Actinomadura atramentaria</i> DSM 43919	24.1	Fosmid library	<i>S. coelicolor</i> M1154	Int	n.d.	Native cluster	46
Microcystin-LR [d-Asp <sup>5</sup> ]microcystin-LR	<i>Microcystis aeruginosa</i> PCC 7806	55	Fosmid library/LCHR	<i>E. coli</i> GB05-MtaA	Plas	↑ ca. 2–3 fold	<i>P<sub>tetO</sub></i> promoter, <i>mtaA</i> (PPTase) co-expression	47
Mithramycin A <b>13</b>	<i>S. argillaceus</i> ATCC12956	45	TAR	<i>S. lividans</i> TK24	Int	↑ ca. 20 fold (3 g L <sup>-1</sup> )	Deletions of secondary metabolite gene clusters from <i>S. lividans</i> TK24	48
Mycosporine-2-(4-deoxygadusolyl-ornithine)	<i>Nostoc flagelliforme</i> CCNUN1	8.19	PCR and ligation	<i>Anabaena</i> sp. PCC 7120	Plas	n.d.	Native cluster	49
Mycosporine-Ornithine / Mycosporine-Lysine	<i>Cylindrospermum stagnale</i> PCC 7417	6.23	PCR and ligation	<i>E. coli</i> BL21(DE3)	Plas	n.d.	T7 promoter, rare tRNA genes co-expressed	50
Myxopyronin A <b>14</b> Myxopyronin B <b>15</b>	<i>Myxococcus fulvus</i> Mxf50	53	Subcloning via plasmid recovery	<i>Myxococcus xanthus</i> DK1622	Int	↑ ca. 20-fold (156 mg mL <sup>-1</sup> )	<i>P<sub>npnII</sub></i> promoter and media optimization	19
Napsamycin	<i>S. sp.</i> DSM5940	38.6	Cosmid library	<i>S. coelicolor</i> M1154	Int	n.d.	Native cluster	51
Natazazole / AJ9561	<i>S. sp.</i> Tü 6176	68.4	TAR	<i>S. lividans</i> JT46	Int	n.d.	Native cluster	52
Nematophin	<i>Xenorhabdus nematophila</i> ATCC 19601	4.1	PCR + Gibson assembly	<i>E. coli</i> DH10BMtaA	Plas	~ 1 mg L <sup>-1</sup>	<i>P<sub>BAD</sub></i> promoter, precursor feeding	53
Oxytetracycline	<i>S. rimosus</i> M4018	29	Fosmid library	<i>S. venezuelae</i> WVR2006	Int	Similar (431 mg L <sup>-1</sup> )	Engineering two cluster-situated regulators (CSR) OtcR and OtrR and feeding precursor, <i>P<sub>ermE*</sub></i> and <i>kasOp*</i> promoter	54
Plantaricyclin A	<i>Lactobacillus plantarum</i> NI326	3.17	PCR and ligation	<i>Lactococcus lactis</i> NZ9000	Plas	n.d.	Native cluster	55

Pikromycin	<i>S. venezuelae</i> ATCC15439	60	Ligation-based plasmid rescue	<i>S. lividans</i> / <i>S. coelicolor</i>	Int	↑ 2.1 fold (333.7 mg L <sup>-1</sup> )	Tandem repeat of the <i>pik</i> gene cluster	56
Pyridinopyrone A 27	<i>S. albus</i> subsp. <i>Chlorinus</i> NRRL B-24108		BAC library	<i>S. albus</i> Del14	Int	n.d.	Solid medium, engineered heterologous host	4
Salinomycin	<i>S. albus</i> DSM41398	106	LLHR + LCHR	<i>S. coelicolor</i> A3(2)	Int	n.d.	Assembly of native BGC from three DNA fragments into a BAC vector	57
Septacidin 1	<i>S. fimbriatus</i> CGMCC 4.1598	24	CATCH	<i>S. albus</i> J1074	Int	n.d.	Native cluster	58
Spinosad	<i>Saccharopolyspora spinosa</i> NRRL 18395	80	BAC library	<i>S. albus</i> J1074 and <i>S. lividans</i> TK24	Int	1.46 mg L <sup>-1</sup>	<i>kasOp*</i> , <i>rpsL-TP</i> and <i>rpsL-cf</i> promoter	59
Spinosad	<i>Saccharopolyspora spinosa</i> ATCC 49460	80	Cosmid library and iterative recombination to replace erythromycin PKS genes with the spinosad BGC	<i>Saccharopolyspora erythraea</i> ATCC 40137	Int	↑ 40 fold (830 mg L <sup>-1</sup> )	Native erythromycin polyketide synthase (PKS) genes in <i>S. erythraea</i> were replaced by the assembled spinosad gene cluster, overexpression of the rhamnosyltransferase, duplication of rhamnose biosynthesis genes, UV mutagenesis, <i>sfp</i> (PPTase) co-expression	60
Streptoseomycin	<i>S. seoulensis</i> A01	76	BAC library	<i>S. chartreusis</i> 1018	Int	n.d.	Native cluster	61
Streptothrinicin	<i>S. sp.</i> strain fd1-xmd	41.8	Plasmid rescue	<i>S. coelicolor</i> M1146	Int	n.d.	Native cluster	62
Syringolin	<i>Ps. syringae</i> pv. <i>syringae</i> (Pss) B728a	22	LLHR	<i>S. coelicolor</i> M145 / <i>S. lividans</i> TK24	Tnps	n.d.	<i>P<sub>snpA</sub></i> promoter	63
Tautomycetin	<i>S. sp.</i> CK4412	80	Ligation-based plasmid rescue	<i>S. coelicolor</i> M145	Int	↑ 5.4 fold (13.31 mg L <sup>-1</sup> )	Tandem repeat of the <i>tmc</i> gene cluster	64
Telomestatin	<i>S. anulatus</i> 3533-SV4	19	BAC library	<i>S. avermitilis</i> SUKA	Int	n.d.	<i>P<sub>sav2794</sub></i> promoter	65
Telomycin	<i>S. canus</i> ATCC 12646	80.5	BAC library	<i>S. albus</i> J1074	Int	n.d.	Native promoter	66
Thaxtomins	<i>S. scabiei</i> 87.22	18	TAR	<i>S. albus</i> J1074	Int	↑10-fold (222 mg L <sup>-1</sup> )	Medium optimization	67
Thaxtomins	<i>S. scabiei</i>	18	Gibson	<i>S. albus</i>	Int	n.d.	Biotransformation step utilizing	68

			assembly				promiscuous tryptophan synthase (TrpS) from <i>Salmonella typhimurium</i> to produce 4-substituted L-tryptophans, and fed to a <i>S. albus</i> strain possessing the NRPS TxtA and TxtB to established a <i>de novo</i> biosynthesis of unnatural thaxtomin derivatives	
Thiolactomycin	<i>Lentzea</i> sp. ATCC 31319	13.7	PCR + Gibson assembly	<i>S. lividans</i> TK24	Int	Similar	Native cluster	69
Thiolactomycin	<i>S. pacifica</i> CNS-863	22	TAR	<i>S. coelicolor</i> M1152	Int	n.d.	Native cluster	70
Thiostreptamide S4	<i>S. sp.</i> NRRL S-4	19	TAR	<i>S. coelicolor</i> M1146	Int	n.d.	Native cluster	71
Thioviridamide	<i>S. olivoviridis</i> NA05001	16.5	PCR and ligation	<i>S. lividans</i> TK23	Int	n.d.	Native cluster	72, 73
Tilivalline	<i>Klebsiella oxytoca</i>	17	TAR	<i>E. coli</i> BL21	Plas	n.d.	$P_{tet}$ and $P_{BAD}$ promoter, tryptophanase TnaA of <i>K. oxytoca</i> , precursor feeding	74
Tilivalline	<i>Xenorhabdus indica</i>	17	TAR	<i>E. coli</i> DH10B-MtaA	Plas	n.d.	$P_{BAD}$ promoter, <i>mtaA</i> (PPTase) co-expression, precursor feeding	75
Vancoresmycin	<i>Amycolatopsis</i> sp. ST 101170	141	PAC library	<i>S. coelicolor</i> M1152	Int	n.d.	Native cluster	76
Violacein	<i>Pseudoalteromonas luteoviolacea</i> 2ta16	8	TAR	<i>Pseudomonas putida</i> KT2440 <i>Agrobacterium tumefaciens</i> LBA4404	Plas	~15 mg L <sup>-1</sup> 10 mg L <sup>-1</sup>	Native cluster	77
Vioprolides A 11	<i>Cystobacter violaceus</i> Cb vi35	56	Cosmid library + LCHR assembly	<i>Myxococcus xanthus</i> DK1622 <i>Burkholderiales</i> strain DSM7029 <i>Pseudomonas putida</i> KT2440	Hom ( <i>tetR</i> site) Tnps	↑ ca. 100 fold (500 mg L <sup>-1</sup> )	$P_{m5}$ promoter	78
YM-216391	<i>S. nobilis</i>	13	Cosmid library	<i>S. coelicolor</i> M1146-M1546	Int with multiple <i>attB</i> sites	36.4 mg L <sup>-1</sup>	One integrase-multiple <i>attB</i> sites (up to four)	79
<b>Novel</b>								
8D1-1 29, 8D1-2 30	<i>S. rochei</i>	81	LEXAS	<i>S. lividans</i> SBT5	Int	n.d.	Native cluster	80
Ambactin 44	<i>Xenorhabdus miraniensis</i> DSM	20	ExRec	<i>E. coli</i> DH10B	Plas	n.d.	<i>mtaA</i> (PPTase) co-expression, T7 promoter	81

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Burrioglumin A <b>42</b>	<i>Burkholderia glumae</i>	41	LLHR	<i>E. coli</i> GB05-MtaA	Plas	n.d.	<i>P<sub>tet</sub></i> promoter, MtaA (PPTase) in heterologous host 82
Burrioglumin B <b>43</b>	PG1						
Clifednamide C <b>53</b>	<i>S. sp.</i> JV178		PCR and ligation	<i>Streptomyces</i> sp. strain NRRL F-2890	Int	n.d.	<i>P<sub>ermE*</sub></i> promoter, <i>cftA</i> heterologous expression 83
Combamides (E <b>39</b> , F <b>40</b> )	<i>S. sp.</i> S10	15	Fosmid library	<i>Streptomyces</i> sp. SR111	Int	n.d.	<i>kasOp*</i> promoter 84
Fluostatin L <b>31</b> /difluostatin A <b>32</b>	<i>Micromonospora rosaria</i> SCSIO N160	40	Cosmid library	<i>S. coelicolor</i> YF11	Int	n.d.	Native cluster 85
Fontizine A <b>5</b>	<i>Serratia fonticola</i> DSM 4576	9.7	DiPaC	<i>E. coli</i> BAP1	Plas	n.d.	T7 promoter 86
Fralmimycin <b>8</b>	<i>Frankia alni</i> strain ACN14a		Fosmid library	<i>S. albus</i> Del14	Int	n.d.	Native cluster, engineered host 4
Lavendiol <b>46</b>	<i>S. lavandulae</i> FRI-5	55.1	Cosmid library	<i>S. avermitilis</i> SUKA22	Int	n.d.	Native cluster 87
Neothioviridamide <b>48</b>	<i>S. olivoviridis</i> NA05001	13	BAC library	<i>S. avermitilis</i> SUKA22	Int	n.d.	Native cluster 88
Nevaltophins (A <b>47</b> )	<i>Xenorhabdus</i> PB62.4 / <i>Xenorhabdus mirianensis</i> DSM17902	12.6	PCR + Gibson assembly	<i>E. coli</i> DH10B-MtaA	Plas	~ 1 mg L <sup>-1</sup>	<i>P<sub>BAD</sub></i> promoter, precursor feeding 53
Pactamides (A <b>38</b> )	<i>S. pactum</i> SCSIO 02999	14.8	Cosmid library	<i>S. lividans</i> TK64,	Int	n.d.	<i>P<sub>ermE*</sub></i> promoter 89
Pepteridines (A - B)	<i>Photobacterium luminescens</i> TT01		PCR + ligation	<i>E. coli</i> BAP1	Plas	n.d.	T7 promoter 90
Polycyclic tetramate macrolactam (SGR810-815)	<i>S. griseus</i>	18	DNA assembler	<i>S. lividans</i>	Int	n.d.	Six constitutive promoters inserted upstream of SGR genes 91
Pyxidicyclines (B <b>4</b> )	<i>Pyxidicoccus fallax</i> <i>An d48</i>	33.7	PCR + TAR assembly	<i>Myxococcus xanthus</i> DK1622, <i>Sti. aurantiaca</i> DW4/3-1	Int	n.d.	<i>P<sub>m5</sub></i> promoter, <i>P<sub>van</sub></i> promoter 92
Sevadincin <b>41</b>	<i>Paenibacillus larvae</i>	11.6	LLHR	<i>E. coli</i> GB05-MtaA	Plas	n.d.	<i>P<sub>BAD</sub></i> promoter, swap start codon from TTG to ATG along with RBS insertion, <i>mtaA</i> (PPTase) co-expression 93
Taromycin A <b>2</b>	<i>Saccharomonospora</i> sp. CNQ-490	67	TAR	<i>S. coelicolor</i> M512	Int	n.d.	Native cluster 94
Taromycin B <b>3</b>	<i>Saccharomonospora</i> sp. CNQ-490	67	TAR	<i>S. coelicolor</i> M1146	Int	n.d.	Native cluster 95
Thiotetroamides	<i>S. afghaniensis</i> NRRL 5621	29	TAR	<i>S. coelicolor</i> M1152	Int	n.d.	Native cluster 70
Violapyrones (33-37)	<i>S. somaliensis</i> SCSIO	1.2	PCR and	<i>S. coelicolor</i> M1146	Int	n.d.	<i>P<sub>gapDH</sub></i> promoter 96

	ZH66	ligation						
Xenolindicin A <b>45</b>	<i>Xenorhabdus indica</i> DSM 17382	20	ExRec	<i>E. coli</i> DH10B-MtaA	Plas	n.d.	<i>mtaA</i> (PPTase) co-expression, T7 promoter	81
<b>Metage-</b> <b>nomics</b>								
Arenimycin C <b>58</b> Arenimycin D <b>59</b>	eDNA collection	40	Cosmid library + TAR assembly	<i>S. albus</i>	Int	n.d.	Native cluster	97
Arimetamycin A <b>60</b>	eDNA collection	40	Cosmid library	<i>S. albus</i>	Int	n.d.	<i>P ermE*</i> promoter	98
Arixanthomycin A <b>57</b>	eDNA collection	33	Cosmid library + TAR assembly	<i>S. albus</i>	Int	n.d.	Native cluster	99
Calixanthomycin A	eDNA collection	50	Cosmid library + TAR assembly	<i>S. albus</i>	Int	n.d.	Native cluster	97
Clarepoxcins A–E (A <b>61</b> )	eDNA collection	25	Cosmid library	<i>S. albus</i>	Int	n.d.	Native cluster	100
Divamide A <b>66</b>	eDNA collection	12	Illumina library	<i>E. coli</i> DH10β	Plas	n.d.	Native cluster	101
Hydroxysporine <b>55</b>	eDNA collection	3.2	Cosmid library	<i>S. albus</i>	Int	n.d.	Native cluster	102
Landepoxcin A <b>62</b> and B	eDNA collection	25	Cosmid library	<i>S. albus</i>	Int	n.d.	Native cluster	100
Lazarimides A and B	eDNA collection	25	Cosmid library + TAR assembly	<i>S. albus</i>	Int	n.d.	Promoter engineering	103
Malacidin A <b>64</b> Malacidin B <b>65</b>	eDNA collection	69.2	Cosmid library + TAR assembly	<i>S. albus</i> J1074	Int	n.d.	Native cluster	104
Metatricycloene <b>63</b>	eDNA collection	n.d.	Cosmid library	<i>S. albus</i>	Int	n.d.	Native cluster	105
Methylaryciarubin <b>54</b>	eDNA collection	5	Cosmid library	<i>E. coli</i> BL21	Plas	n.d.	T7 promoter, indole-3-pyruvic acid (IPA) imine synthase <i>vioA</i> co-expression	106
Polybrominated diphenyl ether	eDNA collection	n.d.	Metagenomic library	<i>Synechococcus</i> <i>elongatus</i> PCC 7942	Int	n.d.	Introduction of synthetic promoter–riboswitch system	107
Reductasporine <b>56</b>	eDNA collection	n.d.	Cosmid library	<i>S. albus</i>	Int	n.d.	Native promoter	102

## References

1. J. Tu, S. Li, J. Chen, Y. Song, S. Fu, J. Ju and Q. Li, *Microb. Cell Fact.*, 2018, **17**, 28.
2. Y. Inahashi, T. Shiraishi, A. Take, A. Matsumoto, Y. Takahashi, S. Omura, T. Kuzuyama and T. Nakashima, *J. Antibiot. (Tokyo)*, 2018, DOI: 10.1038/s41429-018-0057-8.
3. D. Kallifidas, G. Jiang, Y. Ding and H. Luesch, *Microb. Cell Fact.*, 2018, **17**, 25.
4. M. Myronovskyi, B. Rosenkranzer, S. Nadmid, P. Pujic, P. Normand and A. Luzhetskyy, *Metab. Eng.*, 2018, **49**, 316–324.
5. A. C. Ross, L. E. Gulland, P. C. Dorrestein and B. S. Moore, *ACS synthetic biology*, 2015, **4**, 414–420.
6. Y. Li, Z. Li, K. Yamanaka, Y. Xu, W. Zhang, H. Vlamakis, R. Kolter, B. S. Moore and P. Y. Qian, *Scientific reports*, 2015, **5**, 9383.
7. C. Paulus, Y. Rebets, J. Zapp, C. Ruckert, J. Kalinowski and A. Luzhetskyy, *Front. Microbiol.*, 2018, **9**, 1959.
8. Q. Deng, L. Zhou, M. Luo, Z. Deng and C. Zhao, *Synth Syst Biotechnol*, 2017, **2**, 59–64.
9. Q. Liu, Q. Shen, X. Bian, H. Chen, J. Fu, H. Wang, P. Lei, Z. Guo, W. Chen, D. Li and Y. Zhang, *Sci. Rep.*, 2016, **6**, 34623.
10. S. C. Wenzel, H. Hoffmann, J. Zhang, L. Debussche, S. Haag-Richter, M. Kurz, F. Nardi, P. Lukat, I. Kochems, H. Tietgen, D. Schummer, J. P. Nicolas, L. Calvet, V. Czepczor, P. Vrignaud, A. Muhlenweg, S. Pelzer, R. Muller and M. Bronstrup, *Angew. Chem. Int. Ed. Engl.*, 2015, **54**, 15560–15564.
11. N. M. Vior, R. Lacret, G. Chandra, S. Dorai-Raj, M. Trick and A. W. Truman, *Appl. Environ. Microbiol.*, 2018, **84**.
12. L. Yi, Y. Dang, J. Wu, L. Zhang, X. Liu, B. Liu, Y. Zhou and X. Lu, *J. Dairy Sci.*, 2016, **99**, 7002–7015.
13. P. N. Schwarz, A. Buchmann, L. Roller, A. Kulik, H. Gross, W. Wohlleben and E. Stegmann, *Biotechnol. J.*, 2018, **13**.
14. L. Horbal, F. Marques, S. Nadmid, M. V. Mendes and A. Luzhetskyy, *Metab. Eng.*, 2018, **49**, 299–315.
15. J. Zettler, H. Xia, N. Burkard, A. Kulik, S. Grond, L. Heide and A. K. Apel, *ChemBioChem*, 2014, **15**, 612–621.
16. J. F. Castro, V. Razmilic, J. P. Gomez-Escribano, B. Andrews, J. A. Asenjo and M. J. Bibb, *Appl. Environ. Microbiol.*, 2015, **81**, 5820–5831.
17. X. Xu, H. Zhou, Y. Liu, X. Liu, J. Fu, A. Li, Y. Z. Li, Y. Shen, X. Bian and Y. Zhang, *J. Nat. Prod.*, 2018, **81**, 1060–1064.
18. H. A. Iqbal, L. Low-Beinart, J. U. Obiajulu and S. F. Brady, *J. Am. Chem. Soc.*, 2016, **138**, 9341–9344.
19. H. Sucipto, D. Pogorevc, E. Luxenburger, S. C. Wenzel and R. Muller, *Metab Eng*, 2017, **44**, 160–170.

20. C. B. Larson, M. Crusemann and B. S. Moore, *J. Nat. Prod.*, 2017, **80**, 1200–1204.
21. C. Osswald, N. Zaburannyi, C. Burgard, T. Hoffmann, S. C. Wenzel and R. Müller, *J. Biotechnol.*, 2014, **191**, 54–63.
22. J. Kumpfmüller, K. Methling, L. Fang, B. A. Pfeifer, M. Lalk and T. Schweder, *Appl. Microbiol. Biotechnol.*, 2016, **100**, 1209–1220.
23. Q. Li, Y. Song, X. Qin, X. Zhang, A. Sun and J. Ju, *J. Nat. Prod.*, 2015, **78**, 944–948.
24. J. K. Patel and G. Archana, *Appl Soil Ecol*, 2018, **124**, 34–44.
25. Q. Tu, J. Herrmann, S. Hu, R. Raju, X. Bian, Y. Zhang and R. Müller, *Sci. Rep.*, 2016, **6**, 21066.
26. J. J. Jimenez, D. B. Diep, J. Borrero, L. Gutiez, S. Arbulu, I. F. Nes, C. Herranz, L. M. Cintas and P. E. Hernandez, *Microb. Cell Fact.*, 2015, **14**, 166.
27. B. Bonet, R. Teufel, M. Crusemann, N. Ziemert and B. S. Moore, *J Nat Prod*, 2015, **78**, 539–542.
28. X. Bian, B. Tang, Y. Yu, Q. Tu, F. Gross, H. Wang, A. Li, J. Fu, Y. Shen, Y. Z. Li, A. F. Stewart, G. Zhao, X. Ding, R. Muller and Y. Zhang, *ACS Chem Biol*, 2017, **12**, 1805–1812.
29. X. J. Yue, X. W. Cui, Z. Zhang, W. F. Hu, Z. F. Li, Y. M. Zhang and Y. Z. Li, *Appl. Microbiol. Biotechnol.*, 2018, **102**, 5599–5610.
30. L. P. Zhu, X. J. Yue, K. Han, Z. F. Li, L. S. Zheng, X. N. Yi, H. L. Wang, Y. M. Zhang and Y. Z. Li, *Microb. Cell Fact.*, 2015, **14**, 105.
31. A. C. Jones, B. Gust, A. Kulik, L. Heide, M. J. Buttner and M. J. Bibb, *PLoS One*, 2013, **8**, e69319.
32. C. Su, X. Zhao, R. Qiu and L. Tang, *Pharm. Biol.*, 2015, **53**, 269–274.
33. M. Crusemann, R. Reher, I. Schamari, A. O. Brachmann, T. Ohbayashi, M. Kuschak, D. Malfacini, A. Seidinger, M. Pinto-Carbo, R. Richarz, T. Reuter, S. Kehraus, A. Hallab, M. Attwood, H. B. Schioth, P. Mergaert, Y. Kikuchi, T. F. Schaberle, E. Kostenis, D. Wenzel, C. E. Muller, J. Piel, A. Carlier, L. Eberl and G. M. Konig, *Angew. Chem. Int. Ed. Engl.*, 2018, **57**, 836–840.
34. C. Liu, J. Zhang, C. Lu and Y. Shen, *Antonie Van Leeuwenhoek*, 2015, **107**, 1359–1366.
35. X. Bian, F. Huang, H. Wang, T. Klefisch, R. Müller and Y. Zhang, *ChemBioChem*, 2014, **15**, 2221–2224.
36. O. Bilyk, O. N. Sekurova, S. B. Zotchev and A. Luzhetskyy, *PLoS One*, 2016, **11**, e0158682.

37. Y. Sun, Z. Feng, T. Tomura, A. Suzuki, S. Miyano, T. Tsuge, H. Mori, J. W. Suh, T. Iizuka, R. Fudou and M. Ojika, *Sci. Rep.*, 2016, **6**, 22091.
38. P. D'Agostino and T. A. M. Gulder, *ACS Synth Biol*, 2018, DOI: 10.1021/acssynbio.8b00151.
39. G. M. Lin, A. J. Romo, P. H. Liem, Z. Chen and H. W. Liu, *J. Am. Chem. Soc.*, 2017, **139**, 16450–16453.
40. C. J. Guo, F. Y. Chang, T. P. Wyche, K. M. Backus, T. M. Acker, M. Funabashi, M. Taketani, M. S. Donia, S. Nayfach, K. S. Pollard, C. S. Craik, B. F. Cravatt, J. Clardy, C. A. Voigt and M. A. Fischbach, *Cell*, 2017, **168**, 517–.
41. X. Liu, D. Liu, M. Xu, M. Tao, L. Bai, Z. Deng, B. A. Pfeifer and M. Jiang, *J. Nat. Prod.*, 2018, **81**, 72–77.
42. L. Linares-Otoya, V. Linares-Otoya, L. Armas-Mantilla, C. Blanco-Olano, M. Crusemann, M. L. Ganoza-Yupanqui, J. Campos-Florian, G. M. Konig and T. F. Schaberle, *Microbiology*, 2017, DOI: 10.1099/mic.0.000538.
43. G. M. Daba, N. Ishibashi, T. Zendo and K. Sonomoto, *J. Appl. Microbiol.*, 2017, **123**, 1124–1132.
44. J. Mevaere, C. Goulard, O. Schneider, O. N. Sekurova, H. Y. Ma, S. Zirah, C. Afonso, S. Rebuffat, S. B. Zotchev and Y. Y. Li, *Sci. Rep.*, 2018, **8**.
45. P. Videau, K. N. Wells, A. J. Singh, W. H. Gerwick and B. Philmus, *ACS synthetic biology*, 2016, **5**, 978–988.
46. F. Leipoldt, J. Santos-Aberturas, D. P. Stegmann, F. Wolf, A. Kulik, R. Lacret, D. Popadic, D. Keinhorster, N. Kirchner, P. Bekiesch, H. Gross, A. W. Truman and L. Kaysser, *Nat. Commun.*, 2017, **8**, 1965.
47. T. Liu, R. Mazmouz, S. E. Ongley, R. Chau, R. Pickford, J. N. Woodhouse and B. A. Neilan, *ACS Chem. Biol.*, 2017, **12**, 2021–2029.
48. R. Novakova, L. E. Nunez, D. Homerova, R. Knirschova, L. Feckova, B. Rezuchova, B. Sevcikova, N. Menendez, F. Moris, J. Cortes and J. Kormanec, *Appl. Microbiol. Biotechnol.*, 2018, **102**, 857–869.
49. J. L. Shang, Z. C. Zhang, X. Y. Yin, M. Chen, F. H. Hao, K. Wang, J. L. Feng, H. F. Xu, Y. C. Yin, H. R. Tang and B. S. Qiu, *Environ. Microbiol.*, 2018, **20**, 200–213.
50. M. Katoch, R. Mazmouz, R. Chau, L. A. Pearson, R. Pickford and B. A. Neilan, *Appl. Environ. Microbiol.*, 2016, **82**, 6167–6173.
51. X. Tang, M. Gross, Y. Xie, A. Kulik and B. Gust, *ChemBioChem*, 2013, **14**, 2248–2255.
52. C. Cano-Prieto, R. Garcia-Salcedo, M. Sanchez-Hidalgo, A. F. Brana, H. P. Fiedler, C. Mendez, J. A. Salas and C. Olano, *ChemBioChem*, 2015, **16**, 1461–1473.

53. X. Cai, V. L. Challinor, L. Zhao, D. Reimer, H. Adihou, P. Grun, M. Kaiser and H. B. Bode, 2017, **19**, 806–809.
54. S. Yin, Z. Li, X. Wang, H. Wang, X. Jia, G. Ai, Z. Bai, M. Shi, F. Yuan, T. Liu, W. Wang and K. Yang, *Appl. Microbiol. Biotechnol.*, 2016, **100**, 10563–10572.
55. J. Borrero, E. Kelly, P. M. O'Connor, P. Kelleher, C. Scully, P. D. Cotter, J. Mahony and D. van Sinderen, *Appl. Environ. Microbiol.*, 2018, **84**.
56. H. R. Pyeon, H. J. Nah, S. H. Kang, S. S. Choi and E. S. Kim, *Microb Cell Fact*, 2017, **16**, 96.
57. J. Yin, M. Hoffmann, X. Bian, Q. Tu, F. Yan, L. Xia, X. Ding, A. F. Stewart, R. Müller, J. Fu and Y. Zhang, *Sci. Rep.*, 2015, **5**, 15081.
58. W. Tang, Z. Guo, Z. Cao, M. Wang, P. Li, X. Meng, X. Zhao, Z. Xie, W. Wang, A. Zhou, C. Lou and Y. Chen, *Proc. Natl. Acad. Sci. U. S. A.*, 2018, **115**, 2818–2823.
59. G. Y. Tan, K. Deng, X. Liu, H. Tao, Y. Chang, J. Chen, K. Chen, Z. Sheng, Z. Deng and T. Liu, *ACS Synth. Biol.*, 2017, **6**, 995–1005.
60. J. Huang, Z. Yu, M. H. Li, J. D. Wang, H. Bai, J. Zhou and Y. G. Zheng, *Appl. Environ. Microbiol.*, 2016, **82**, 5603–5611.
61. B. Zhang, K. B. Wang, W. Wang, S. F. Bi, Y. N. Mei, X. Z. Deng, R. H. Jiao, R. X. Tan and H. M. Ge, *Org. Lett.*, 2018, **20**, 2967–2971.
62. Y. Yu, B. Tang, R. Dai, B. Zhang, L. Chen, H. Yang, G. Zhao and X. Ding, *Appl. Microbiol. Biotechnol.*, 2018, **102**, 2621–2633.
63. F. Huang, J. Tang, L. He, X. Ding, S. Huang, Y. Zhang, Y. Sun and L. Xia, *Microb. Cell Fact.*, 2018, **17**, 31.
64. H. J. Nah, M. W. Woo, S. S. Choi and E. S. Kim, *Microb Cell Fact*, 2015, **14**, 140.
65. K. Amagai, H. Ikeda, J. Hashimoto, I. Kozone, M. Izumikawa, F. Kudo, T. Eguchi, T. Nakamura, H. Osada, S. Takahashi and K. Shin-Ya, *Sci. Rep.*, 2017, **7**, 3382.
66. C. Fu, L. Keller, A. Bauer, M. Bronstrup, A. Froidbise, P. Hammann, J. Herrmann, G. Mondesert, M. Kurz, M. Schiell, D. Schummer, L. Toti, J. Wink and R. Muller, *J. Am. Chem. Soc.*, 2015, **137**, 7692–7705.
67. G. D. Jiang, Y. C. Zhang, M. M. Powell, P. L. Zhang, R. Zuo, Y. Zhang, D. Kallifidas, A. M. Tieu, H. Luesch, R. Loria and Y. S. Ding, *Appl. Environ. Microbiol.*, 2018, **84**.
68. M. Winn, D. Francis and J. Micklefield, *Angewandte Chemie International Edition*, 2018, **57**, 6830–6833.

69. M. E. Yurkovich, R. Jenkins, Y. Sun, M. Tosin and P. F. Leadlay, *Chem. Commun. (Camb.)*, 2017, **53**, 2182–2185.
70. X. Tang, J. Li, N. Millan-Aguinaga, J. J. Zhang, E. C. O'Neill, J. A. Ugalde, P. R. Jensen, S. M. Mantovani and B. S. Moore, *ACS Chem Biol.*, 2015, **10**, 2841–2849.
71. L. Frattaruolo, R. Lacret, A. R. Cappello and A. W. Truman, *ACS Chem. Biol.*, 2017, **12**, 2815–2822.
72. M. Izawa, T. Kawasaki and Y. Hayakawa, *Appl. Environ. Microbiol.*, 2013, **79**, 7110–7113.
73. M. Izawa, S. Nagamine, H. Aoki and Y. Hayakawa, *J. Gen. Appl. Microbiol.*, 2018, **64**, 50–53.
74. A. von Tesmar, M. Hoffmann, A. Abou Fayad, S. Huttel, V. Schmitt, J. Herrmann and R. Müller, *ACS Chem. Biol.*, 2018, **13**, 812–819.
75. H. Wolff and H. B. Bode, *PLoS One*, 2018, **13**, e0194297.
76. B. Kepplinger, S. Morton-Laing, K. H. Seistrup, E. C. L. Marrs, A. P. Hopkins, J. D. Perry, H. Strahl, M. J. Hall, J. Errington and N. E. E. Allenby, *ACS Chem. Biol.*, 2018, **13**, 207–214.
77. J. J. Zhang, X. Tang, M. Zhang, D. Nguyen and B. S. Moore, *MBio*, 2017, **8**.
78. F. Yan, D. Auerbach, Y. Chai, L. Keller, Q. Tu, S. Huttel, A. Glemser, H. A. Grab, T. Bach, Y. Zhang and R. Müller, *Angew. Chem. Int. Ed. Engl.*, 2018, DOI: 10.1002/anie.201802479.
79. L. Li, G. Zheng, J. Chen, M. Ge, W. Jiang and Y. Lu, *Metab. Eng.*, 2017, **40**, 80–92.
80. M. Xu, Y. Wang, Z. Zhao, G. Gao, S. X. Huang, Q. Kang, X. He, S. Lin, X. Pang, Z. Deng and M. Tao, *Appl. Environ. Microbiol.*, 2016, **82**, 5795–5805.
81. O. Schimming, F. Fleischhacker, F. I. Nollmann and H. B. Bode, *ChemBioChem*, 2014, **15**, 1290–1294.
82. T. Thongkongkaew, W. Ding, E. Bratovanov, E. Oueis, M. Garcia-Altares, N. Zaburannyi, K. Harmrolfs, Y. M. Zhang, K. Scherlach, R. Müller and C. Hertweck, *Acs Chemical Biology*, 2018, **13**, 1370–1379.
83. Y. Qi, E. Ding and J. A. V. Blodgett, *ACS Synth Biol.*, 2018, **7**, 357–362.
84. Y. Liu, H. Wang, R. Song, J. Chen, T. Li, Y. Li, L. Du and Y. Shen, *Org. Lett.*, 2018, **20**, 3504–3508.
85. C. Yang, C. Huang, W. Zhang, Y. Zhu and C. Zhang, *Org. Lett.*, 2015, **17**, 5324–5327.
86. C. Greunke, E. R. Duell, P. M. D'Agostino, A. Glockle, K. Lamm and T. A. M. Gulder, *Metab. Eng.*, 2018, **47**, 334–345.
87. I. G. U. Pait, S. Kitani, F. W. Roslan, D. Ulanova, M. Arai, H. Ikeda and T. Nihira, *J. Ind. Microbiol. Biotechnol.*, 2018, **45**, 77–87.

88. T. Kawahara, M. Izumikawa, I. Kozone, J. Hashimoto, N. Kagaya, H. Koiwai, M. Komatsu, M. Fujie, N. Sato, H. Ikeda and K. Shin-Ya, *J. Nat. Prod.*, 2018, **81**, 264–269.
89. S. Saha, W. Zhang, G. Zhang, Y. Zhu, Y. Chen, W. Liu, C. Yuan, Q. Zhang, H. Zhang, L. Zhang, W. Zhang and C. Zhang, *Chem. Sci.*, 2017, **8**, 1607–1612.
90. H. B. Park, C. E. Perez, K. W. Barber, J. Rinehart and J. M. Crawford, *Elife*, 2017, **6**.
91. Y. Luo, H. Huang, J. Liang, M. Wang, L. Lu, Z. Shao, R. E. Cobb and H. Zhao, *Nat. Commun.*, 2013, **4**, 2894.
92. F. Panter, D. Krug, S. Baumann and R. Müller, *Chem. Sci.*, 2018, **9**, 4898–4908.
93. Y. Tang, S. Frewert, K. Harmrolfs, J. Herrmann, L. Karmann, U. Kazmaier, L. Xia, Y. Zhang and R. Muller, *J Biotechnol*, 2015, **194**, 112–114.
94. K. Yamanaka, K. A. Reynolds, R. D. Kersten, K. S. Ryan, D. J. Gonzalez, V. Nizet, P. C. Dorrestein and B. S. Moore, *Proc Natl Acad Sci U S A*, 2014, **111**, 1957–1962.
95. K. A. Reynolds, H. Luhavaya, J. Li, S. Dahesh, V. Nizet, K. Yamanaka and B. S. Moore, *The Journal of antibiotics*, 2018, **71**, 333–338.
96. L. Hou, H. Huang, H. Li, S. Wang, J. Ju and W. Li, *Microb. Cell Fact.*, 2018, **17**, 61.
97. H. S. Kang and S. F. Brady, *J. Am. Chem. Soc.*, 2014, **136**, 18111–18119.
98. H. S. Kang and S. F. Brady, *Angew. Chem. Int. Ed. Engl.*, 2013, **52**, 11063–11067.
99. H. S. Kang and S. F. Brady, *ACS Chem. Biol.*, 2014, **9**, 1267–1272.
100. J. G. Owen, Z. Charlop-Powers, A. G. Smith, M. A. Ternei, P. Y. Calle, B. V. Reddy, D. Montiel and S. F. Brady, *Proc. Natl. Acad. Sci. U. S. A.*, 2015, **112**, 4221–4226.
101. T. E. Smith, C. D. Pond, E. Pierce, Z. P. Harmer, J. Kwan, M. M. Zachariah, M. K. Harper, T. P. Wyche, T. K. Matainaho, T. S. Bugni, L. R. Barrows, C. M. Ireland and E. W. Schmidt, *Nat. Chem. Biol.*, 2018, **14**, 179–185.
102. F. Y. Chang, M. A. Ternei, P. Y. Calle and S. F. Brady, *J. Am. Chem. Soc.*, 2015, **137**, 6044–6052.
103. D. Montiel, H. S. Kang, F. Y. Chang, Z. Charlop-Powers and S. F. Brady, *Proc. Natl. Acad. Sci. U. S. A.*, 2015, **112**, 8953–8958.
104. B. M. Hover, S. H. Kim, M. Katz, Z. Charlop-Powers, J. G. Owen, M. A. Ternei, J. Maniko, A. B. Estrela, H. Molina, S. Park, D. S. Perlin and S. F. Brady, *Nat Microbiol*, 2018, **3**, 415–422.

105. H. A. Iqbal, L. Low-Beinart, J. U. Obiajulu and S. F. Brady, *J. Am. Chem. Soc.*, 2016, **138**, 9341–9344.
106. F. Y. Chang and S. F. Brady, *ChemBioChem*, 2014, **15**, 815–821.
107. V. Agarwal, J. M. Blanton, S. Podell, A. Taton, M. A. Schorn, J. Busch, Z. Lin, E. W. Schmidt, P. R. Jensen, V. J. Paul, J. S. Biggs, J. W. Golden, E. E. Allen and B. S. Moore, *Nat. Chem. Biol.*, 2017, **13**, 537–543.