Supplementary Information

DNA-templated silver nanoclusters locate microRNAs in nuclei of gastric cancer cells

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SUPPLEMENTARY METHODS

Cell Culture

Cells were maintained at 37°C in a humidified air atmosphere containing 5% carbon dioxide in Dulbecco's Modified Eagle's Media supplemented with 10% FBS.

Establishment of MGC803 cells with over-expressed miR-101-3p

The miR-101-3p sequence in a length of 490bp was amplified from HEK293T cells and constructed into the lentivirus expression vector pCDH1-CMV-MCS-EF1a-GFP-T2A-Puroto. The primers used were miR-101-1F: CCTGAATTCATTCTAATTTAATTCAACTGG and miR-101-1R: TATGG ATCCTCAGCACAACATGGCTGCAC. The acquired vectors were transfected into HEK293T cells using GM easyTM lentivirus packaging kit (Genomeditech) to generate lentivirus. MGC803 cells were infected with the recombinant lentivirus-transducing units plus 8 μ g/mL Polybrene (Sigma). The stable colonies (MGC803(101))were obtained after selection on puromycin ¹.

RNA extraction and qRT-PCR

10⁶ cells were collected for RNA extraction according to the method described earlier². The extracted RNA was detected on NanoDrop 1000 (NanoDrop Technologies, Delaware, USA) and stored at -80°C for further use. After that, miR-101-3p, miR-16-5p and miR-19b-3p were detected by TaqMan qRT-PCR as described in our previous methods^{2, 3}, in which the spiked-in cel-miR-39 was used as an external control for normalized quantification.

SUPPLEMETARY TABLES AND FIGURES



Fig. S1 The protocol flowchart of the main steps for fluorescence *in situ* hybridization detection of miRNAs based on AgNCs/DNAs.



Fig. S2 Schematic illustration of the relationship of our previous study² with the presented study.



Fig. S3 Fluorescence spectra of the AgNCs/DNAs and their hybrids prepared in different molar ratios of DNAs : AgNO₃ : NaBH₄. **(A-C)** AgNCs/PrC (PrC) and its hybrids (PrC-h19bG) via hybridization with Hph19bG in molar ratios of DNAs:AgNO₃:NaBH₄ as 1:9:9, 1:14:14 or 1:25:25 (pH8.0) (A, λ_{ex} = 430 nm; B, λ_{ex} = 460 nm; C, λ_{ex} = 580 nm). **(D-F)** AgNCs/PrC trans (PrC tra) and its duplex (PrC-h19bG tra) via hybridization with Hph19bG trans in molar ratios of DNAs:AgNO₃:NaBH₄ as 1:14:14 or 1:25:25 (pH 6.6) (D, λ_{ex} = 430 nm; E, λ_{ex} = 460 nm; F, λ_{ex} = 580 nm). Sodium phosphate buffer (Pi) at the same pH was used as blank control.

DNAs: AgNO ₃ :	AgNCs/DNAs 或 AgNCs/DNA-HpDNAs	430nm (450-800nm)		460nm (480-800nm)		580nm (600-900nm)	
NaBH ₄		A ^a	R ^b	A ^a	R ^b	A ^a	R ^b
1:9:9	PrC	316.744	-	199.757	-	117.189	-
	PrC-h19bG	296.150	0.93	723.702	3.62	50.935	0.43
1:14:14	PrC	225.079	-	241.491	-	351.189	-
	PrC-h19bG	434.725	1.93	958.817	3.97	54.087	0.15
1:25:25	PrC	61.761	-	71.672	-	282.769	-
	PrC-h19bG	411.551	6.66	317.953	4.44	140.185	0.50
1:14:14	PrC tra	282.915	-	915.671	-	1081.408	-
	PrC-h19bG tra	539.396	1.91	497.329	0.54	352.105	0.33
1:25:25	PrC tra	63.369	-	161.540	-	154.751	-
	PrC-h19bG tra	384.306	6.06	443.783	2.75	581.519	3.76

 Table S1 Integral areas in the fluorescence spectra for AgNCs/DNAs and their hybrids prepared in different molar ratios of DNAs : AgNO3 : NaBH4

^{a,b} elucidated in Table 2.

рН	AgNCs/DNAs 或	430nm (450-800nm)		4601	ım	580nm		
				(480-800nm)		(600-900nm)		
	AgNCs/DNA-HpDNAs	A ^a	R ^b	A ^a	R ^b	A ^a	R ^b	
рН6.6	PrC	961.628	-	563.627	-	145.474	-	
	PrC-h101G	466.972	0.49	354.567	0.63	23.651	0.16	
	PrC-h16G	532.758	0.55	401.970	0.71	34.925	0.24	
	PrC-h19bG	462.761	0.48	275.550	0.49	18.767	0.13	
	PrC tra	191.670	-	213.175	-	809.598	-	
	PrC-h101G tra	1541.494	8.04	946.905	4.44	1458.880	1.80	
	PrC-h16G tra	1098.513	5.73	786.435	3.69	764.732	0.94	
	PrC-h19bG tra	1190.606	6.21	827.569	3.88	1778.794	2.20	
рН7.2	PrC	560.763	-	399.668	-	266.014	-	
	PrC-h101G	601.154	1.07	1163.578	2.91	71.696	0.27	
	PrC-h16G	552.638	0.99	625.675	1.57	128.778	0.48	
	PrC-h19bG	490.693	0.88	1008.772	2.52	48.061	0.18	
	PrC tra	144.613	-	308.033	-	972.755	-	
	PrC-h101G tra	869.546	6.01	725.160	2.35	671.586	0.69	
	PrC-h16G tra	710.756	4.91	692.310	2.25	846.998	0.87	
	PrC-h19bG tra	873.168	6.04	737.936	2.40	775.566	0.80	
рН8.0	PrC	432.245	-	309.824	-	145.396	-	
	PrC-h101G	463.907	1.07	1190.910	3.84	20.114	0.14	
	PrC-h16G	495.800	1.15	1369.513	4.42	36.534	0.25	
	PrC-h19bG	474.091	1.10	1063.243	3.43	21.037	0.14	
	PrC tra	149.375	-	470.589	-	598.073	-	
	PrC-h101G tra	791.429	5.30	689.830	1.47	216.737	0.36	
	PrC-h16G tra	760.788	5.09	716.006	1.52	211.523	0.35	
	PrC-h19bG tra	984.388	6.59	829.842	1.76	290.965	0.49	

Table S2 Integral areas in the fluorescence spectra for AgNCs/DNAs and their hybrids prepared at different pH

^{a,b} elucidated in Table 2.



Fig. S4 (A-C) The absorbance spectra of the AgNCs/DNAs and their hybrids prepared at different pH(A, pH 6.6; B, pH 7.2; C, pH 8.0). (**D-F)** Photos of the solutions taken under white light in (A-C) (D, A; E, B; F, C). Subjects and No. labels were illustrated in Fig. 2. The AgNCs/DNAs probes (PrC) without hybridization at the same pH were used as controls.



Fig. S5 Effect of components in FISH on the AgNCs/DNA probe and its fluorescence enhancement. (A-C) Fluorescence spectra of AgNCs/PrC (PrC) and its hybrid AgNCs/PrC-h19bG (PrC-h19bG) with the following components: formamide(Form), SSC (SSC), yeast tRNA(Yeast), Denhardt' s solution (Den), Hybridization solution (Hyb), Tween 20 (Tw20), acetic anhydride (Ac), PFA (PFA), proteinase K (PK), Triton X-100 (Trix) (A, $\lambda_{ex} = 430$ nm; B, $\lambda_{ex} = 460$ nm; C, $\lambda_{ex} = 580$ nm). (**D**, **E**) (D) Photo and (E) image taken under UV light by Gel Image System for the solutions in (A-C). 1-12 in (D-E) corresponded to the labels from top to bottom in (A-C). Sodium phosphate buffer (Pi) at the same pH was used as blank control.



Fig. S6 Effect of the self-prepared hybridization solution in FISH on the AgNCs/DNA probe and its fluorescence enhancement. **(A-C)** Fluorescence spectra of AgNCs/PrC (PrC) and its hybrid AgNCs/PrC-h19bG (PrC-h19bG) with the hybridization solutions (A, $\lambda_{ex} = 430$ nm; B, $\lambda_{ex} = 460$ nm; C, $\lambda_{ex} = 580$ nm). **(D, E)** (D) Photo and (E) image taken under UV light by Gel Image System for the solutions in (A-C). 1-6 corresponded to the labels from top to bottom in (A-C). (-3), (-5), (+3) and (+5) were illustrated in Experimental Section. Sodium phosphate buffer (Pi) at the same pH was used as blank control.



Fig. S7 (A-C) Absorbance spectra of the solutions respectively in (A) Fig.S3, (B) Fig. S4 and (C) Fig. 3. **(D-F)** Photos taken under white light of the solutions in (A-C) (D, A; E, B; F,C). Subjects and No. labels were illustrated respectively in Fig.S4, Fig.S5 and Fig. 3. The AgNCs/DNAs probes (PrC or PrC tra) without hybridization at the same pH were used as controls.

Table S3 Integral areas in the fluorescence spectra for the AgNCs/PrC probe and its hybrids

	430 nm (450-800 nm)		460 n	m	580 nm (600-900 nm)	
Agines/Dina of			(480-800	nm)		
	A ^a	R ^b	A ^a	R ^b	A ^a	R ^b
PrC	555.595	-	368.371	-	160.561	-
PrC-h19bG	451.265	0.81	1077.577	2.93	116.439	0.73
PrC-h19bG(-3) °	634.116	1.14	1245.243	3.38	224.364	1.40
PrC-h19bG(-5) °	489.979	0.88	1242.099	3.37	421.378	2.62
PrC-h19bG(+3) °	543.431	0.98	1137.579	3.09	198.858	1.24
PrC-h19bG(+5) °	535.404	0.96	852.855	2.32	334.993	2.09

^{a,b} elucidated in Table 2. ^c the AgNCs/PrC probe and its hybrids as illustrated in Fig. S6.



Fig. S8 Quantification of miR-101-3p, miR-16-5p and miR-19b-3p in MGC803 cells and miR-101-3p over-expressed MGC803 cells (MCC803(101)) based on qRT-PCR.

REFERENCES

- B. Q. Zheng, L. H. Liang, C. M. Wang, S. L. Huang, X. Cao, R. P. Zha, L. Liu, D. S. Jia, Q. Tian, J. H. Wu, Y. W. Ye, Q. F. Wang, Z. W. Long, Y. Zhou, C. Y. Du, X. H. He and Y. Q. Shi, *Clinical Cancer Research*, 2011, **17**, 7574-7583.
- 2. J. Zhang, C. Li, X. Zhi, G. A. Ramon, Y. Liu, C. Zhang, F. Pan and D. Cui, *Analytical Chemistry*, 2016, **88**, 1294-1302.
- 3. J. Zhang, Y. Song, C. Zhang, X. Zhi, H. Fu, Y. Ma, Y. Chen, F. Pan, K. Wang, J. Ni, W. Jin, X. He, H. Su and D. Cui, *Theranostics*, 2015, **5**, 733-745.