Electronic Supplementary Information

Target-Initiated Synthesis of Fluorescent Copper Nanoparticles for Sensitive and

Label-Free Detection of Bleomycin

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Fig. S1 Measurement of fluorescence emission spectra in response to 1 μ M synthetic sequence. The synthesized sequence is used to mimic the DNAzyme-catalyzed cleavage product and performs the TdTase-assisted base extension. The sequence is 5'-GCG CCG CCG CAA AAT TCA CCA ACT AT <u>X</u>-3'. The underlined base (X) indicates the deoxyribonucleoside adenosine (A, green line), the ribonucleoside adenosine (rA, red line), the A

modified with PO₄ at the 3' terminal (A 3'-PO₄, blue line), and the rA modified with PO₄ at the 3' terminal (rA 3'-PO₄, magenta line), respectively. In the presence of the synthetic sequence without 3'-OH ends (blue line and magenta lines), low fluorescence signals are observed. In contrast, the enhanced fluorescence signals are detected in the presence of the synthetic sequences with 3'-OH ends (green line and red line) as a result of the staining of poly-T products by SYBR Gold. Moreover, the same fluorescence signals are observed in the presence of A (green line) and rA (red line). These results demonstrate that TdTase can incorporate a number of dNTPs to the 3'-OH ends of DNA and RNA.



Fig. S2 Measurement of fluorescence emission spectra under different experimental conditions. The BLM concentration is 100 nM. The 1 μ M Hp1, 4 μ M Hp2 and 10 U of TdTase were used in the experiments. In the presence of Hp1 + BLM (black line), Hp1 + BLM + TdTase (blue line), Hp2 + BLM (cyan line), Hp2 + BLM + TdTase (magenta line), Hp1 + Hp2 + BLM (green line), low fluorescence signals are observed as a result of the staining of Hp1 and Hp2 by SYBR Gold. In contrast, an enhanced fluorescence signal is detected in the presence of Hp1 + Hp2 + BLM + TdTase (red line) as a result of the staining of poly-T products by SYBR Gold. These results demonstrate that the TdTase-assisted base extension reaction can only be triggered by target BLM in the presence

of specific Hp1 and Hp2.



Fig. S3 (A) Measurement of fluorescence emission spectra in the absence of Zn^{2+} and Pb^{2+} (black line) and presence of 20 μ M Pb²⁺ (magenta line), 20 μ M Pb²⁺ + 2 U of PNK + 1 mM ATP (blue line), 10 μ M Zn²⁺ (red line), and 10 μ M Zn²⁺ + 2 U PNK + 1 mM ATP (green line), respectively. (B) Measurement of fluorescence emission spectra in the absence of Co²⁺ (blue line) and presence of 0.25 mM Co²⁺ (red line) and 0.5 mM Co²⁺ (black line), respectively. All buffer contains 10 μ M Zn²⁺, 10 mM Mg²⁺ and 100 nM Fe²⁺. (C) Measurement of fluorescence emission spectra in the absence of Mg²⁺ (blue line) and presence of 10 mM Mg²⁺ (red line) and 20 mM Mg²⁺ (black line), respectively. All buffer contains 10 μ M Zn²⁺, 0.25 mM Co²⁺, and 100 nM Fe²⁺. (D) Measurement of fluorescence emission spectra in the absence of Fe²⁺ (blue line) and presence of 100 nM Fe²⁺. (D) Measurement of fluorescence emission spectra in the absence of Fe²⁺ (blue line) and presence of 100 nM Fe²⁺. (D) Measurement of

nM. The 1 μM Hp1, 4 μM Hp2 and 10 U of TdTase were used in the experiments.



Fig. S4 Real-time monitoring of the oxidative cleavage of Hp1 induced by different-concentration BLM.

strategy	assay	requirement of	read out	linear range	LOD	ref.
	time*	labels				
electrochemical assay	~8 h	electrochemical	Turn-off	100 pM - 100	100 pM	1
		label (ferrocene)		μΜ		
electrochemical assay based on	over 16h	electrochemical	Turn-off	0.07-910 nM	0.02 nM	2
dual-amplification		label(MBA-AuNP				
		s/DA-AuNPs)				
electrogenerated	~12 h	no	Turn-off	0.1-50 pM	0.03 pM	3
chemiluminescence assay						
colorimetric assay	~20 min	no	Turn-on	25 nM - 1 μM	16 nM	4
fluorescence quenching	no data	no	Turn-off	0.09-2.0 μg/mL	40 ng /	5
					mL	
exo III-aided DNA recycling	~40 min	fluorescent label	Turn-on	0.1 nM - 1 μM	23 pM	6
amplification-based fluorescent		(FAM and BHQ)				
assay						
WS ₂ nanosheet quencher-based	~23 min	fluorescent label	Turn-on	0.5 nM - 1 μM	0.3 nM	7
fluorescent assay		(FAM)				
perylene derivative	~12 h	fluorescent label	Turn-on	5-500 nM	0.2 nM	8
quencher-based fluorescent		(FAM)				
assay						
silver nanocluster-based	~83 min	no	Turn-off	100-500 nM	54 nM	9
fluorescent assay						
enzymatic polymerization-	100 min	no	Turn-on	1 fM - 5 nM	8.1 ×	this
mediated synthesis of CuNPs-					10 ⁻¹⁶ M	work
based fluorescent assay						

Table S1. Comparison of the proposed method with the reported methods for BLM assay

*Assay time includes the preparation time. MBA-AuNPs/DA-AuNPs: 4-mercaptophenyl boronic acid-capped gold

nanoparticles and dopamine-capped gold nanoparticles. WS₂: tungsten disulfide. LOD: limit of detection.

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