Supplementary Information

One-step synthesis of red/green dual-emission carbon dots

for ratiometric sensitive ONOO⁻ probing and cell imaging

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Methylhydrazine sulfate

Scheme S1. Structural formulas of 2,5-diaminotoluene sulfate, 2,5-diaminotoluene and methylhydrazine sulfate.

Synthetic raw material	Single/dual emission	Excitation wavelength (nm)	Emission wavelength (nm)	QY (%)	Reference
citric acid, urea, sodium fluoride	single	530	600	1.20	1
citric acid, neutral red	single	530	632	12.1	2
p-phenylenediamine	single	470	620	15.0	3
ascorbic acid, ethylene glycol	dual	365	435/538	not mentioned	4
m-phenylenediamine, sulfuric acid	dual	300	360/520	43.0 (460 nm excitation)	5
citric acid, urea	dual	380	455/520	14.0 (420 nm excitation)	6, 7
2,5-diaminotoluene sulfate	dual	380	525/603	9.00 (380 nm excitation)	this work

Table S1. Comparation of the properties of the RGDE CDs with other CDs.



Fig. S1. Effect of ethanol volume on the fluorescence intensity (603 nm) of the RGDE CDs.



Fig. S2. Effect of reaction temperature on the fluorescence intensity (603 nm) of the RGDE CDs.



Fig. S3. Effect of reaction time on the fluorescence intensity (603 nm) of the RGDE CDs.



Fig. S4. XPS full-scan spectrum of the RGDE CDs.



Fig. S5. Fluorescence excitation spectrum of the RGDE CDs.



Fig. S6. Fluorescence excitation and emission spectra of the CDs prepared through using 2,5-diaminotoluene as carbon source.



Fig. S7. Fluorescence excitation and emission spectra of the CDs prepared through using methylhydrazine sulfate as carbon source.



Fig. S8. Confocal microscopy fluorescence images of HeLa cells treated with the RGDE CDs (30 μ g/mL) in green channel, red channel and merged image of green and red channel. Scale bar is 10 μ m.



Fig. S9. Effect of light illumination time on the fluorescence intensity of the RGDE CDs.



Fig. S10. Effect of concentration of NaCl solution on the fluorescence intensity of the RGDE CDs.



Fig. S11. Effect of environmental temperature on the fluorescence intensity of the RGDE CDs.



Fig. S12. Effect of pH of buffer solution on the fluorescence intensity of the RGDE CDs.



Fig. S13. Effect of pH of buffer solution on $(F_{525}/F_{603})_0$ and $(F_{525}/F_{603})_0$. $(F_{525}/F_{603})_0$ and (F_{525}/F_{603}) were the fluorescence intensity ratios of the RGDE CDs solution in the absence and presence of 22 μ M ONOO⁻, respectively.



Fig. S14. Time-dependent fluorescence intensity of the RGDE CDs with the addition of 22 μM ONOO⁻ at room temperature.



Fig. S15. FT-IR spectrum of the RGDE CDs in the presence of $ONOO^-$.



Fig. S16. Cyclic voltammograms of the RGDE CDs in the solution state (solvent: freshly dried dimethylformamide).



Fig. S17. Cyclic voltammograms of ONOO⁻ in the solution state (solvent: freshly dried dimethylformamide).



Fig. S18. UV-vis absorption spectrum of ONOO⁻, inset was the photograph of ONOO⁻ solution under visible light.



Fig. S19. Time-resolved decays of the RGDE CDs in the absence (black line) and presence (red line) of $ONOO^-$.



Fig. S20. Cell viability of HeLa cells with different concentrations of the RGDE CDs.

Quantum yield (QY) measurements.

QY of the obtained RGDE CDs was determined by the method mentioned in our previous work.⁸ The absolute photoluminescence quantum yield can be represented simple in the equation below:

$$QY = \frac{\int L_{\text{emission}}}{\int E_{\text{solvent}} - \int E_{\text{sample}}}$$
(1)

where QY was the absolute quantum yield, L_{emission} was the fluorescence (FL) emission spectrum of the RGDE CDs sample, collected using the sphere; E_{sample} was the spectrum of the light used to excite the sample, collected using the sphere; E_{solvent} was the spectrum of the light used for excitation with only the solvent in the sphere, collected using the sphere. The solvent in this experiment was deionized water.

References

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