## Distinct autophagy-inducing abilities of similar-sized nanoparticles in cell culture and live *C. elegans*

Qin Wang<sup>a,b</sup>, Yanfeng Zhou<sup>b,c</sup>, Rong Fu<sup>a</sup>, Yi Zhu<sup>a</sup>, Bin Song<sup>b</sup>, Yiling Zhong<sup>b</sup>, Sicong Wu<sup>b</sup>, Yu Shi<sup>b</sup>, Yanyan

Wu<sup>b</sup>, Yuanyuan Su<sup>b</sup>, Huimin Zhang\*<sup>a</sup>, Yao He\*<sup>b</sup>

<sup>a</sup>Jiangsu Key Laboratory of Infection and Immunity, Institutes of Biology and Medical Sciences (IBMS), Soochow

University, Suzhou, Jiangsu 215123, China.

<sup>b</sup>Jiangsu Key Laboratory for Carbon-Based Functional Materials and Devices, Institute of Functional Nano and Soft

Materials (FUNSOM), Collaborative Innovation Center of Suzhou Nano Science and Technology, Soochow

University, Suzhou, Jiangsu 215123, China.

<sup>c</sup>School of Public Health, Shanghai Jiao Tong University School of Medicine, Shanghai 200025, China.

\* Corresponding authors. Telephone: 86-512-6588 0946 Fax: 86-512-6588 2846

E-mails: zhanghuimin@suda.edu.cn (H. Zhang), yaohe@suda.edu.cn (Y. He)

## Additional data:



Fig. S1 Representative confocal images of autophagosomes labeled by anti-LC3 immunostaining (a) and the

fluorescence of SiNPs (b) in SiNP-treated cells corresponding to Fig. 2.



**Fig. S2** Autophagosomes formation in HeLa cells after exposure to different doses of blue- or green-emitting SiNPs. (a-g) Representative confocal images of autophagosomes labeled by anti-LC3 immunostaining in cells treated with de-ionized H<sub>2</sub>O (a), blue-emitting SiNPs with 15 µg/mL (b), 150 µg/mL (c), 1500 µg/mL (d), or greenemitting SiNPs with 15 µg/mL (e), 150 µg/mL (f), 1500 µg/mL (g) for 24 h. (a'-g') are enlarged views of the boxed areas in (a-g), respectively. (h) Quantitative analysis of the number of LC3 puncta corresponding to the experimental groups in (a-g). The scale bar represents 25 µm. N ≥ 4 for each data set. Error bars, ± SEM; ns, p >0.05.



**Fig. S3** Autophagosomes formation in HEK293T cells after exposure to different doses of blue- or green-emitting SiNPs. (a-g) Representative confocal images of autophagosomes labeled by anti-LC3 immunostaining in cells treated with de-ionized H<sub>2</sub>O (a), blue-emitting SiNPs with 15 µg/mL (b), 150 µg/mL (c), 1500 µg/mL (d), or greenemitting SiNPs with 15 µg/mL (e), 150 µg/mL (f), 1500 µg/mL (g) for 24 h. (a'-g') are enlarged views of the boxed areas in (a-g), respectively. (h) Quantitative analysis of the number of LC3 puncta corresponding to the experimental groups in (a-g). The scale bar represents 25 µm. N ≥ 4 for each data set. Error bars, ± SEM; ns, p >0.05.



**Fig. S4** Autophagosomes formation in *C. elegans* after CDs, AuNPs, QDs, or SiNPs microinjection. (a-f) Representative confocal images of autophagosome marked by LGG-1::mCherry in worms after 6-h injection of H<sub>2</sub>O (a, negative control), starvation (b, positive control), CDs (c), AuNPs (d), QDs (e) or SiNPs (f) into *C. elegans* intestinal cells. (g) The fluorescence of SiNPs in the same region corresponding to (f). (h) Quantitative analysis of the number of mCherry::LGG-1 aggregates corresponding to the experimental groups in (a-f). N  $\geq$  4 for each data set. Error bars, ± SEM; \*\**p* < 0.01; \**p* < 0.05; ns, *p* > 0.05.



Fig. S5 Zeta potentials of CDs, AuNPs, SiNPs and QDs. Three experimental repeats for each data set. Error bars, ±

SEM.