

B(C₆F₅)₃-Catalyzed Transfer Hydrogenations of Imines with Hantzsch Ester

Qiaotian Wang,^{a,b} Jingjing Chen,^{a,b} Xiangqing Feng,^{a,b,*} and Haifeng Du^{a,b,*}

^a Beijing National Laboratory of Molecular Sciences, CAS Key Laboratory of Molecular Recognition and Function, Institute of Chemistry, Chinese Academy of Sciences, Beijing 100190, China.

^b University of Chinese Academy of Sciences, Beijing 100049, P. R. China

Supporting Information

General consideration

All air-sensitive compounds were handled under an atmosphere of argon or in a nitrogen-filled glovebox. ^1H NMR and ^{13}C NMR spectra were recorded on Bruker AV 400 at ambient temperature with CDCl_3 as solvent and TMS as internal standard. Chemical shifts (δ) were given in ppm, referenced to the residual proton resonance of TMS (0) or to the carbon resonance of the CDCl_3 (77.23). Coupling constants (J) were given in Hertz (Hz). All solvents were purified by conventional methods, distilled before use. Commercially available reagents were used without further purification. All the substrates were synthesized according to reported method.

General procedure for the metal-free catalytic transfer hydrogenation of imine

(Scheme 3): Dissolving $\text{B}(\text{C}_6\text{F}_5)_3$ (0.0010 g, 0.002 mmol) in *n*-hexane (10 mL) to give a solution of catalyst (0.0002 M). Then to a sealed-tube were added imine **5** (0.6 mmol), Hantzsch Ester (0.1820 g, 0.72 mmol) and the solution of *n*-hexane with dissolved catalyst (0.0002 M, 3.0 mL, 0.0006 mmol) in a nitrogen atmosphere glovebox. After being sealed, the resulting mixture was stirred for 48 h at 120 °C. The reaction mixture was cooled to room temperature, and the solvent was removed under reduced pressure. The crude residue was purified by flash chromatography on silica gel using *n*-hexane/DCM (20/1-50/1) as the fluent to give the desired product **7**.

General procedure for the asymmetric transfer hydrogenation of imine (Scheme 5):

To a sealed-tube were added $\text{HB}(\text{C}_6\text{F}_5)_2$ (0.0104 g, 0.03 mmol), chiral diene **8**

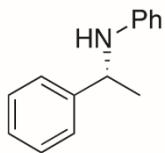
(0.0102 g, 0.015 mmol) and dry toluene (1.5 mL). The resulting mixture was stirred for 5 min at room temperature, and substrate imine **5** (0.3 mmol) and Hantzsch Ester (0.0911 g, 0.36 mmol) was added in a nitrogen atmosphere glovebox. After being sealed, the resulting mixture was stirred for 12 h at 40 °C. The reaction mixture was cooled to room temperature, and the solvent was removed under reduced pressure, and the crude residue was purified by flash chromatography on silica gel using *n*-hexane/DCM (20/1-50/1) as the fluent to give the desired product **7**.

Table S1. Optimization of reaction conditions for asymmetric transfer hydrogenations of imine **5a**.^a

Entry	Solvent	Temp. (°C)	Conv. (%) ^b	Ee (%) ^c
1	toluene	0	16	<i>rac.</i>
2	toluene	25	83	13
3	toluene	40	100	38
4	toluene	50	100	36
5	toluene	60	100	33
6	toluene	80	100	29
7	DCM	40	100	11
8	<i>n</i> -hexane	40	100	9
9	cyclohexane	40	100	24
10	<i>n</i> -pentane	40	100	32
11	<i>n</i> -heptane	40	100	27

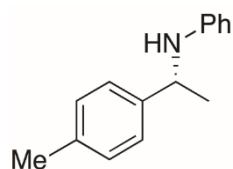
^a All the reactions were carried out with chiral diene (5 mol %), HB(C₆F₅)₂ (10 mol %), imine **5a** (0.1 mmol) and Hantzsch ester **1** (0.12 mmol) in solvent (1.0 mL) unless otherwise noted. ^b Determined by crude ¹H NMR. ^c Determined by chiral HPLC.

Characterization of products



(R)-N-(1-Phenylethyl)aniline (7a): Colorless oil; $[\alpha]_D^{20} = -6.8$ (*c* 0.84, MeOH) (38% ee) [lit.: $[\alpha]_D^{20} = -4.3$ (*c* 1.1, CHCl₃) (78% ee for *R*-isomer)]; ¹H NMR (400 MHz, CDCl₃, ppm) δ 7.35 (d, *J* = 7.6 Hz, 2H), 7.29 (dd, *J* = 7.6, 7.2 Hz, 2H), 7.24-7.17 (m, 1H), 7.07 (dd, *J* = 7.6, 7.6 Hz, 2H), 6.65-6.60 (m, 1H), 6.49 (d, *J* = 8.0 Hz, 2H), 4.46 (q, *J* = 6.8 Hz, 1H), 3.99 (brs, 1H), 1.49 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃, ppm) δ 147.6, 145.5, 129.4, 128.9, 127.1, 126.1, 117.5, 113.6, 53.7, 25.3.

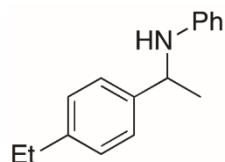
W. Li, G. Hou, M. Chang and X. Zhang, *Adv. Synth. Catal.*, 2009, **351**, 3123.



(R)-N-(1-(*p*-Tolyl)ethyl)aniline (7b): Yellow oil; $[\alpha]_D^{20} = +1.3$ (*c* 0.99, MeOH) (37% ee); ¹H NMR (400 MHz, CDCl₃) δ 7.25 (d, *J* = 8.0 Hz, 2H), 7.11 (d, *J* = 8.0 Hz, 2H), 7.09 (dd, *J* = 8.4, 7.6 Hz, 2H), 6.65-6.60 (m, 1H), 6.50 (d, *J* = 8.4 Hz, 2H), 4.45

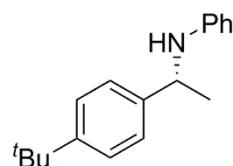
(q, $J = 6.8$ Hz, 1H), 3.98 (brs, 1H), 2.31 (s, 3H), 1.48 (d, $J = 6.8$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3 , ppm) δ 147.6, 142.5, 136.6, 129.6, 129.3, 126.0, 117.4, 113.5, 53.4, 25.3, 21.3.

K. Ye, X. Wang, C. G. Daniliuc, G. Kehr and G. Erker, *Eur. J. Inorg. Chem.*, 2016, 368.



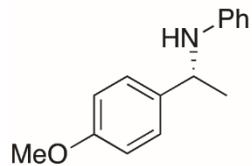
N-(1-(4-Ethylphenyl)ethyl)aniline (7c): Yellow oil; ^1H NMR (400 MHz, CDCl_3 , ppm) δ 7.26 (d, $J = 7.6$ Hz, 2H), 7.13 (d, $J = 7.6$ Hz, 2H), 7.08 (dd, $J = 7.6, 7.6$ Hz, 2H), 6.66-6.59 (m, $J = 7.2$ Hz, 1H), 6.50 (d, $J = 8.0$ Hz, 2H), 4.45 (q, $J = 6.4$ Hz, 1H), 3.97 (brs, 1H), 2.61 (q, $J = 7.6$ Hz, 2H), 1.48 (d, $J = 6.8$ Hz, 3H), 1.21 (t, $J = 7.6$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3 , ppm) δ 147.7, 143.0, 142.7, 129.4, 128.4, 126.1, 117.4, 113.56, 53.4, 28.7, 25.2, 15.8.

Y. Liu and H. Du, *J. Am. Chem. Soc.*, 2013, **135**, 6810.



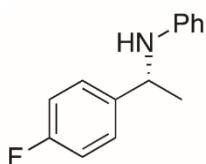
(R)-N-(1-(4-(*tert*-Butyl)phenyl)ethyl)aniline (7d): Yellow solid, m.p. 60-62°C; $[\alpha]_D^{20} = +2.1$ (*c* 0.99, CH₂Cl₂) (20% ee) [lit.: $[\alpha]_D^{20} = +5.0$ (*c* 0.50, CH₂Cl₂) (85% ee for *R*-isomer)]; ¹H NMR (400 MHz, CDCl₃, ppm) δ 7.29 (dd, *J* = 8.2, 8.2 Hz, 4H), 7.08 (dd, *J* = 8.0, 7.6 Hz, 2H), 6.66-6.60 (m, 1H), 6.52 (d, *J* = 8.0 Hz, 2H), 4.46 (q, *J* = 6.8 Hz, 1H), 3.98 (brs, 1H), 1.49 (d, *J* = 6.4 Hz, 3H), 1.29 (s, 9H); ¹³C NMR (100 MHz, CDCl₃, ppm) δ 149.9, 147.7, 142.3, 129.4, 125.8, 125.8, 117.4, 113.5, 53.2, 34.7, 31.7, 25.0.

E. Kumaran and W. K. Leong, *Organometallics*, 2012, **31**, 1068.



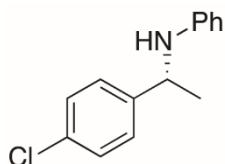
(R)-N-(1-(4-Methoxyphenyl)ethyl)aniline (7e): Yellow oil; $[\alpha]_D^{20} = -2.8$ (*c* 0.99, MeOH) (24% ee) [lit.: $[\alpha]_D^{20} = -6.2$ (*c* 0.55, MeOH) (84% ee for *R*-isomer)]; ¹H NMR (400 MHz, CDCl₃, ppm) δ 7.24 (d, *J* = 7.6 Hz, 2H), 7.11 (d, *J* = 8.0 Hz, 2H), 7.08 (dd, *J* = 8.4, 7.6 Hz, 2H), 6.66-6.59 (m, 1H), 6.51 (d, *J* = 8.4 Hz, 2H), 4.44 (q, *J* = 6.8 Hz, 1H), 3.97 (brs, 1H), 3.77 (s, 3H), 1.48 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃, ppm) δ 158.7, 147.6, 137.5, 129.3, 127.1, 117.4, 114.2, 113.5, 55.5, 53.1, 25.2.

A. V. Malkov, S. Stončius, K. N. MacDougall, A. Mariani, G. D. MacGeoch and P. Kočovský, *Tetrahedron*, 2005, **62**, 264.



(R)-N-(1-(4-Fluorophenyl)ethyl)aniline (7f): Yellow oil; $[\alpha]_D^{20} = -11.8$ (c 1.01, MeOH) (37% ee); ^1H NMR (300 MHz, CDCl_3 , ppm) δ 7.38-7.28 (m, 2H), 7.13-7.05 (m, 2H), 7.04-6.94 (m, 2H), 6.69-6.61 (m, 1H), 6.48 (d, $J = 7.8$ Hz, 2H), 4.46 (q, $J = 6.6$ Hz, 1H), 4.00 (brs, 1H), 1.49 (d, $J = 6.6$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3 , ppm) δ 162.0 (d, $J = 240.0$ Hz), 147.3, 141.2 (d, $J = 3.0$ Hz), 141.2, 129.3, 127.6 (d, $J = 8.0$ Hz), 117.7, 115.7 (d, $J = 21.0$ Hz), 113.6, 53.1, 25.4.

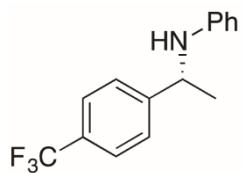
W. Pan, Y. Deng, J. He, B. Bai and H. Zhu, *Tetrahedron*, 2013, **69**, 7253.



(R)-N-(1-(4-Chlorophenyl)ethyl)aniline (7g): Yellow oil; $[\alpha]_D^{20} = +0.7$ (c 0.83, MeOH) (36% ee); ^1H NMR (400 MHz, CDCl_3 , ppm) δ 7.38-7.27 (m, 4H), 7.18-7.10 (m, 2H), 6.74-6.66 (m, 1H), 6.51 (d, $J = 8.0$ Hz, 2H), 4.47 (q, $J = 6.8$ Hz, 1H), 4.02

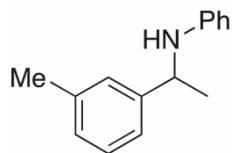
(brs, 1H), 1.50 (d, $J = 6.8$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3 , ppm) δ 147.3, 144.12, 132.7, 129.5, 129.1, 127.6, 117.8, 113.6, 53.3, 25.4.

W. Pan, Y. Deng, J. He, B. Bai and H. Zhu, *Tetrahedron*, 2013, **69**, 7253.



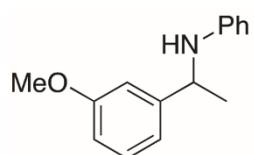
(R)-N-(1-(4-(Trifluoromethyl)phenyl)ethyl)aniline (7h): Yellow oil; $[\alpha]_D^{20} = -11.1$ (c 0.98, CH_2Cl_2) (33% ee) [lit.: $[\alpha]_D^{20} = -23.1$ (c 0.59, CH_2Cl_2) (85% ee for *R*-isomer)]; ^1H NMR (400 MHz, CDCl_3 , ppm) δ 7.57 (d, $J = 8.0$ Hz, 2H), 7.48 (d, $J = 8.4$ Hz, 2H), 7.12-7.05 (m, 2H), 6.70-6.64 (m, 1H), 6.47 (d, $J = 7.6$ Hz, 2H), 4.52 (q, $J = 6.8$ Hz, 1H), 4.04 (brs, 1H), 1.52 (d, $J = 6.8$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3 , ppm) δ 149.7, 147.0, 129.6, 129.4, 129.2, 126.4, 125.9 (q, $J = 4.0$ Hz), 123.1, 117.9, 113.5, 53.5, 29.9, 25.3, 1.2.

M. Rueping, E. Sugiono, C. Azap, T. Theissmann and M. Bolte, *Org. Lett.*, 2005, **7**, 3781.



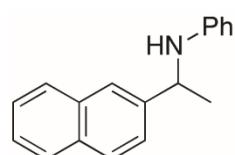
N-(1-(*m*-Tolyl)ethyl)aniline (7i): Yellow oil; ^1H NMR (400 MHz, CDCl_3 , ppm) δ 7.24-7.12 (m, 3H), 7.08 (dd, $J = 7.6, 7.6$ Hz, 2H), 7.03 (d, $J = 7.2$ Hz, 1H), 6.66-6.61 (m, 1H), 6.51 (d, $J = 8.0$ Hz, 2H), 4.44 (q, $J = 6.8$ Hz, 1H), 3.98 (brs, 1H), 2.33 (s, 3H), 1.49 (d, $J = 6.8$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3 , ppm) δ 147.6, 145.5, 138.4, 129.3, 128.7, 127.9, 126.8, 123.1, 117.4, 113.5, 53.7, 25.2, 21.8.

A. Lefranc, Z.-W. Qu, S. Grimme and M. Oestreich, *Chem. Eur. J.*, 2016, **22**, 10009.



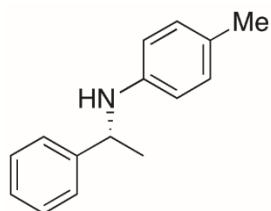
N-(1-(3-Methoxyphenyl)ethyl)aniline (7j): Yellow oil; ^1H NMR (400 MHz, CDCl_3 , ppm) δ 7.13 (dd, $J = 8.0, 7.6$ Hz, 1H), 6.98 (dd, $J = 7.6, 7.2$ Hz, 2H), 6.88-6.81 (m, 2H), 6.69-6.63 (m, 1H), 6.54 (dd, $J = 7.6, 7.2$ Hz, 1H), 6.41 (d, $J = 8.4$ Hz, 2H), 4.34 (q, $J = 6.8$ Hz, 1H), 3.90 (s, 1H), 3.65 (s, 3H), 1.39 (d, $J = 6.8$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3 , ppm) δ 160.2, 147.6, 147.45, 130.0, 129.4, 118.5, 117.6, 113.6, 112.3, 112.0, 55.4, 53.8, 25.3.

A. Lefranc, Z.-W. Qu, S. Grimme and M. Oestreich, *Chem. Eur. J.*, 2016, **22**, 10009.



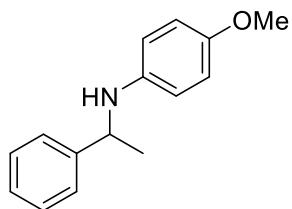
N-(1-(Naphthalen-2-yl)ethyl)aniline (7k): Yellow oil; ^1H NMR (400 MHz, CDCl_3 , ppm) δ 7.83-7.74 (m, 4H), 7.50-7.45 (m, 1H), 7.45-7.36 (m, 2H), 7.10-7.02 (m, 2H), 6.66-6.58 (m, 1H), 6.53 (d, $J = 8.0$ Hz, 2H), 4.61 (q, $J = 6.8$ Hz, 1H), 4.08 (brs, 1H), 1.55 (d, $J = 6.8$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3 , ppm) δ 147.6, 143.1, 133.9, 133.0, 129.4, 128.8, 128.1, 128.0, 126.3, 125.8, 124.7, 124.5, 117.6, 113.7, 54.0, 25.3.

S. Guizzetti, M. Benaglia, F. Cozzi and R. Annunziata, *Tetrahedron*, 2009, **65**, 6354.



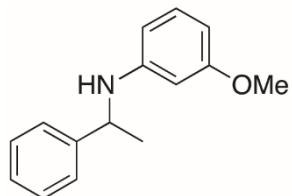
(R)-4-Methyl-N-(1-phenylethyl)aniline (7l): Yellow oil; $[\alpha]_D^{20} = -4.0$ (c 1.0, EtOAc) (16% ee) [lit.: $[\alpha]_D^{25} = +27.3$ (c 0.7, EtOAc) (91% ee for *S*-isomer)]; ^1H NMR (400 MHz, CDCl_3 , ppm) δ 7.40-7.25 (m, 4H), 7.24-7.16 (m, 1H), 6.89 (d, $J = 8.0$ Hz, 2H), 6.42 (d, $J = 8.0$ Hz, 2H), 4.44 (q, $J = 6.4$ Hz, 1H), 3.86 (brs, 1H), 2.17 (s, 3H), 1.48 (d, $J = 6.8$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3 , ppm) δ 145.7, 145.3, 129.9, 128.9, 128.5, 127.1, 126.6, 126.1, 113.7, 53.9, 25.3, 20.6.

D. Pei, Z. Wang, S. Wei, Y. Zhang and J. Sun, *Org. Lett.*, 2006, **8**, 5913.



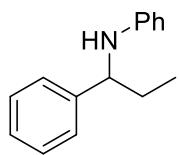
4-Methoxy-N-(1-phenylethyl)aniline (7m): Yellow oil; ^1H NMR (500 MHz, CDCl_3 , ppm) δ 7.35 (d, $J = 7.5$ Hz, 2H), 7.33-7.26 (m, 2H), 7.21 (d, $J = 6.0$ Hz, 1H), 6.68 (d, $J = 9.0$ Hz, 2H), 6.46 (d, $J = 9.0$ Hz, 2H), 4.40 (q, $J = 7.0$ Hz, 1H), 3.77 (brs, 1H), 3.67 (s, 3H), 1.48 (d, $J = 6.5$ Hz, 3H); ^{13}C NMR (126 MHz, CDCl_3 , ppm) δ 150.8, 144.5, 140.5, 127.6, 125.8, 124.8, 113.7, 113.5, 54.7, 53.2, 24.1.

I. Chatterjee and M. Oestreich, *Angew. Chem., Int. Ed.*, 2015, **54**, 1965.



3-Methoxy-N-(1-phenylethyl)aniline (7n): Yellow oil; ^1H NMR (400 MHz, CDCl_3 , ppm) δ 7.37-7.24 (m, 4H), 7.23-7.14 (m, 1H), 6.97 (dd, $J = 8.0, 8.0$ Hz, 1H), 6.20 (d, $J = 8.0$ Hz, 1H), 6.13 (d, $J = 7.6$ Hz, 1H), 6.05 (s, 1H), 4.45 (q, $J = 6.4$ Hz, 1H), 4.03 (brs, 1H), 3.65 (s, 3H), 1.47 (d, $J = 6.8$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3 , ppm) δ 161.0, 149.0, 145.5, 130.1, 128.9, 127.2, 126.1, 106.8, 102.7, 99.7, 55.2, 53.8, 25.2.

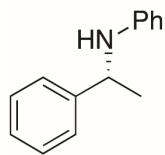
X.-Y. Liu and C.-M. Che, *Org. Lett.*, 2009, **11**, 4204.



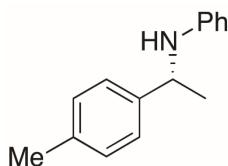
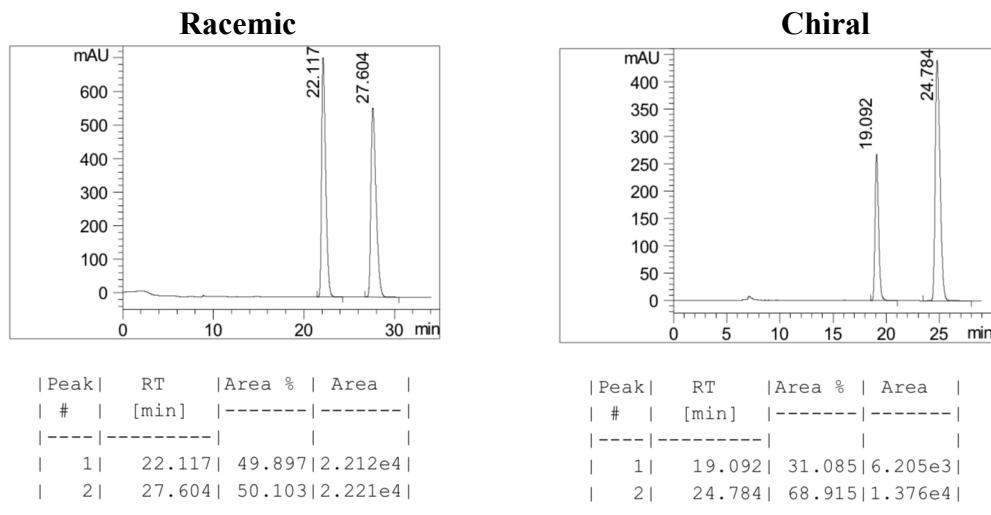
N-(1-Phenylpropyl)aniline (7o): Yellow oil; ^1H NMR (500 MHz, CDCl_3 , ppm) δ 7.37-7.24 (m, 4H), 7.23-7.17 (m, 1H), 7.10-7.02 (t, $J = 7.5$ Hz, 2H), 6.65-6.58 (m, 1H), 6.50 (d, $J = 8.0$ Hz, 2H), 4.21 (t, $J = 6.7$ Hz, 1H), 4.03 (brs, 1H), 1.80 (m, 2H), 0.94 (t, $J = 7.4$ Hz, 3H); ^{13}C NMR (126 MHz, CDCl_3 , ppm) δ 146.5, 142.9, 128.0, 127.4, 125.83, 125.4, 116.1, 112.2, 58.7, 30.6, 9.8.

I. Chatterjee and M. Oestreich, *Angew. Chem., Int. Ed.*, 2015, **54**, 1965.

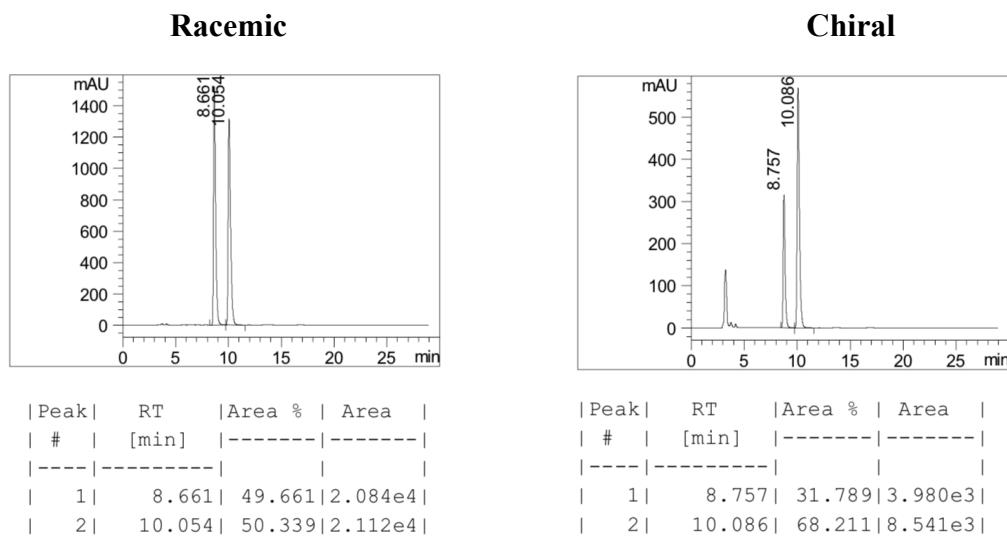
The chromatography for the determination of enantiomeric excess

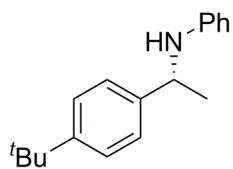


HPLC Conditions: Column: Chiralcel OD-H, Daicel Chemical Industries, Ltd.,
Eluent: Hexanes/IPA (98/2); **Flow rate:** 0.5 mL/min; **Detection:** UV 254 nm



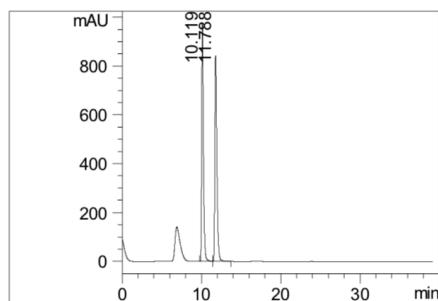
HPLC Conditions: Column: Chiralcel OD-H, Daicel Chemical Industries, Ltd.,
Eluent: Hexanes/IPA (99/1); **Flow rate:** 1.0 mL/min; **Detection:** UV 254 nm





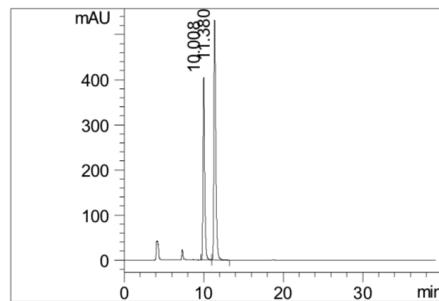
HPLC Conditions: Column: Chiralcel OD-H, Daicel Chemical Industries, Ltd.,
Eluent: Hexanes/IPA (98/2); **Flow rate:** 0.8 mL/min; **Detection:** UV 254 nm

Racemic

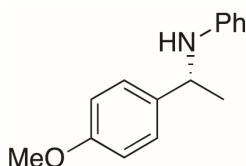


Peak	RT	Area %	Area
#	[min]		
1	10.119	49.825	1.488e4
2	11.788	50.175	1.499e4

Chiral

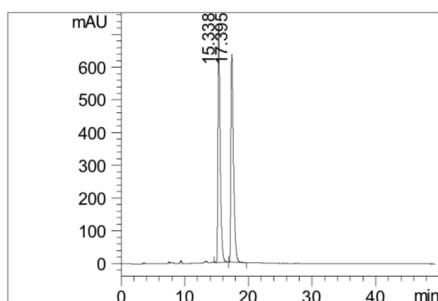


Peak	RT	Area %	Area
#	[min]		
1	10.008	39.932	5.900e3
2	11.380	60.068	8.875e3



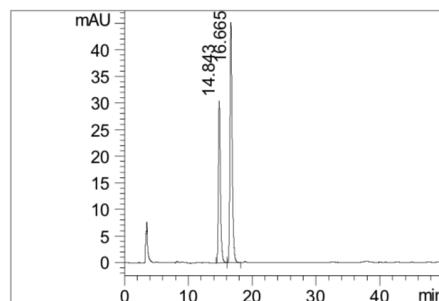
HPLC Conditions: Column: Chiralcel OD-H, Daicel Chemical Industries, Ltd.,
Eluent: Hexanes/IPA (99/1); **Flow rate:** 0.8 mL/min; **Detection:** UV 254 nm

Racemic

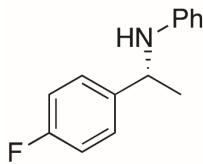


Peak	RT	Area %	Area
#	[min]		
1	15.338	50.021	1.799e4
2	17.395	49.979	1.798e4

Chiral

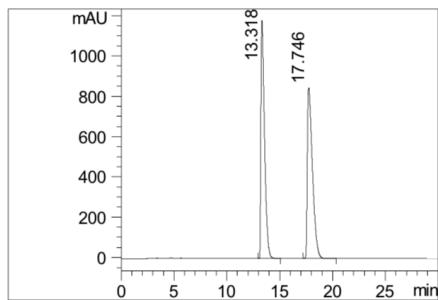


Peak	RT	Area %	Area
#	[min]		
1	14.843	38.115	673.712
2	16.665	61.885	1.094e3



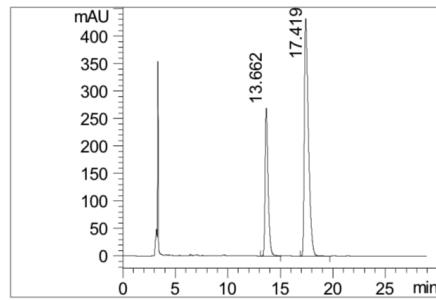
HPLC Conditions: Column: Chiralcel OD-H, Daicel Chemical Industries, Ltd.,
Eluent: Hexanes/IPA (99/1); **Flow rate:** 1.0 mL/min; **Detection:** UV 254 nm

Racemic

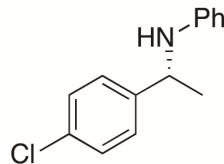


Peak #	RT [min]	Area %	Area
1	13.318	49.676	2.815e4
2	17.746	50.324	2.851e4

Chiral

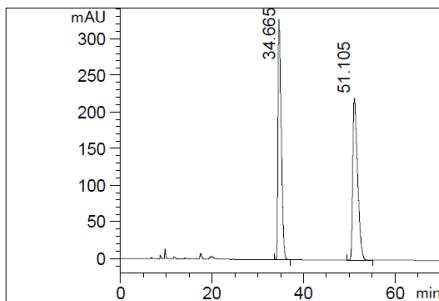


Peak #	RT [min]	Area %	Area
1	13.662	31.294	5.321e3
2	17.419	68.706	1.168e4



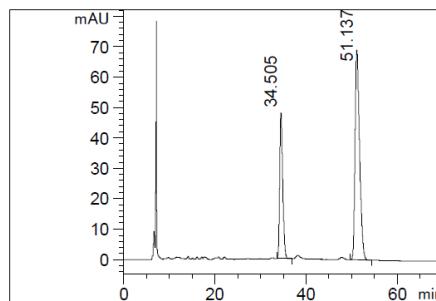
HPLC Conditions: Column: Chiralcel OD-H, Daicel Chemical Industries, Ltd.,
Eluent: Hexanes/IPA (98/2); **Flow rate:** 0.5 mL/min; **Detection:** UV 254 nm

Racemic

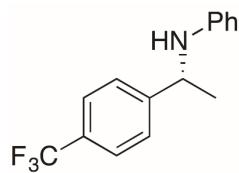


Peak #	RT [min]	Area %	Area
1	34.665	49.866	1.624e4
2	51.105	50.134	1.632e4

Chiral

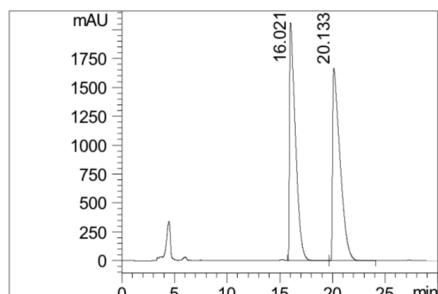


Peak #	RT [min]	Area %	Area
1	34.505	31.839	2.235e3
2	51.137	68.161	4.785e3



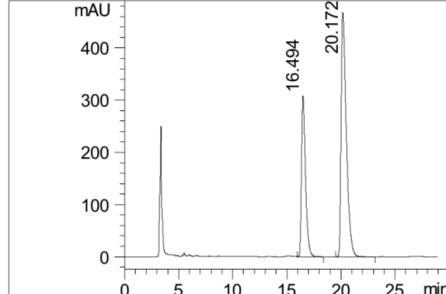
HPLC Conditions: Column: Chiralcel OD-H, Daicel Chemical Industries, Ltd.,
Eluent: Hexanes/IPA (98/2); **Flow rate:** 0.6 mL/min; **Detection:** UV 254 nm

Racemic

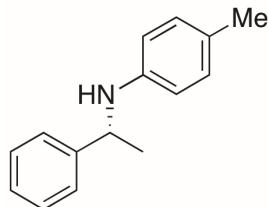


Peak	RT	Area %	Area
#	[min]	-----	-----
1	16.021	49.578	8.049e4
2	20.133	50.422	8.185e4

Chiral

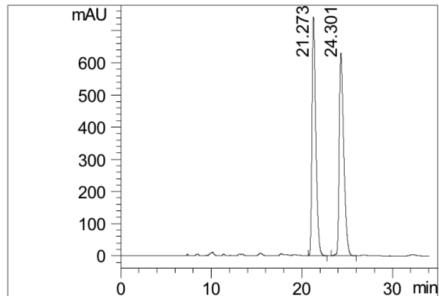


Peak	RT	Area %	Area
#	[min]	-----	-----
1	16.494	33.668	7.972e3
2	20.172	66.332	1.571e4



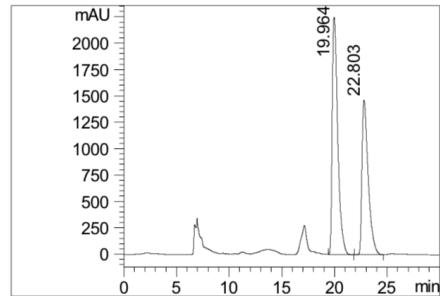
HPLC Conditions: Column: Chiralcel OD-H, Daicel Chemical Industries, Ltd.,
Eluent: Hexanes/IPA (98/2); **Flow rate:** 0.5 mL/min; **Detection:** UV 254 nm

Racemic

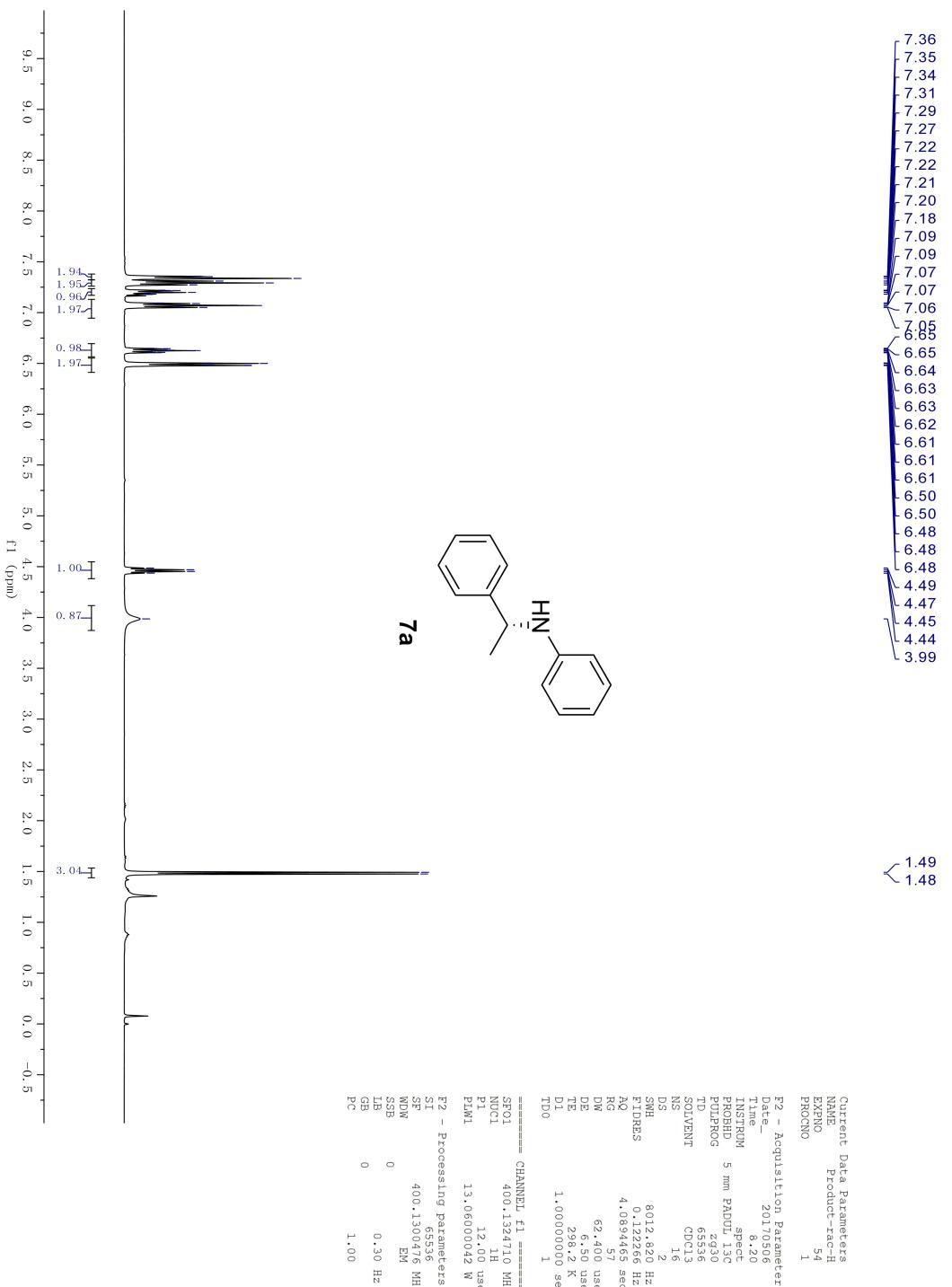


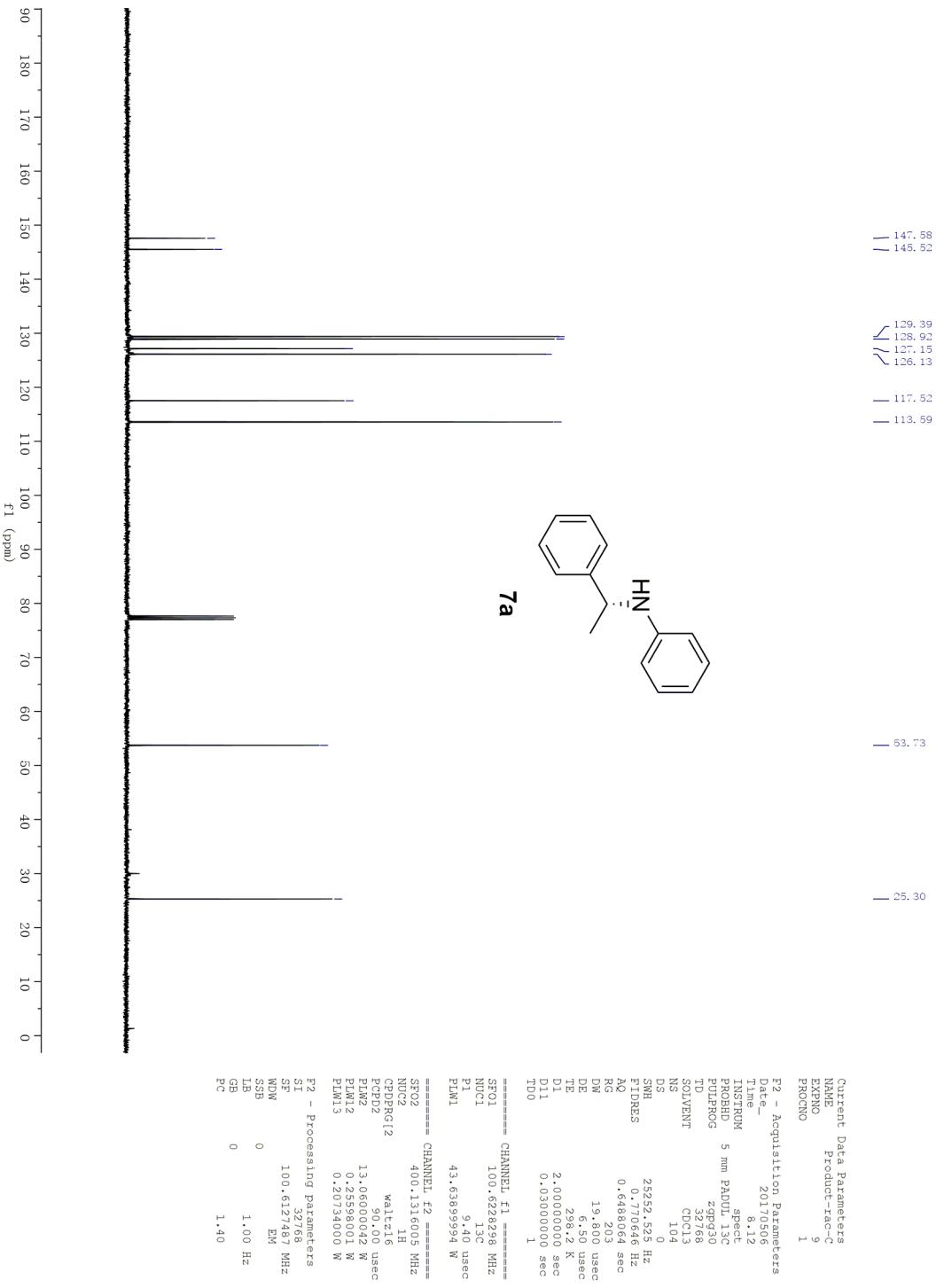
Peak	RT	Area %	Area
#	[min]	-----	-----
1	21.273	49.887	2.181e4
2	24.301	50.113	2.191e4

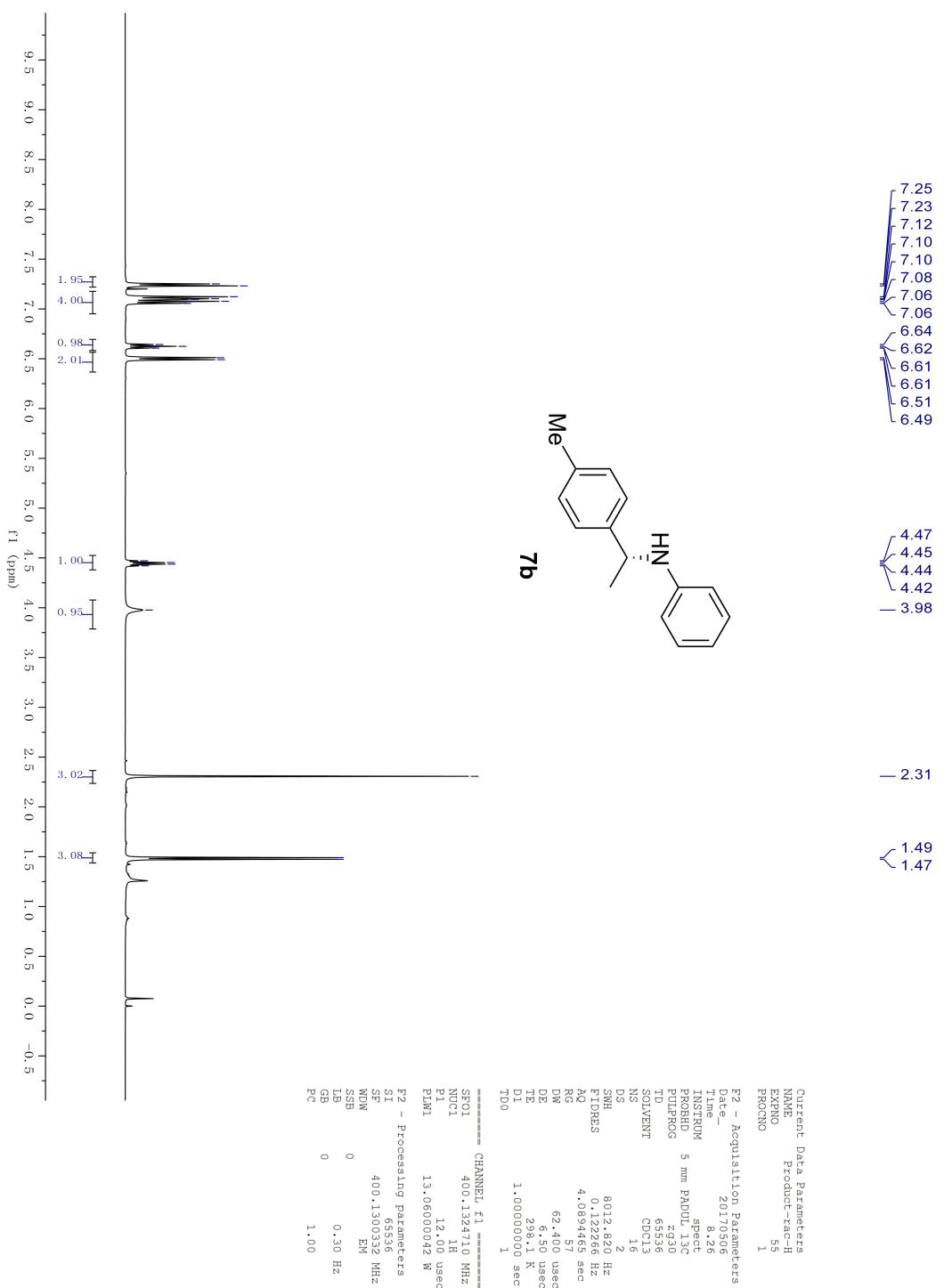
Chiral

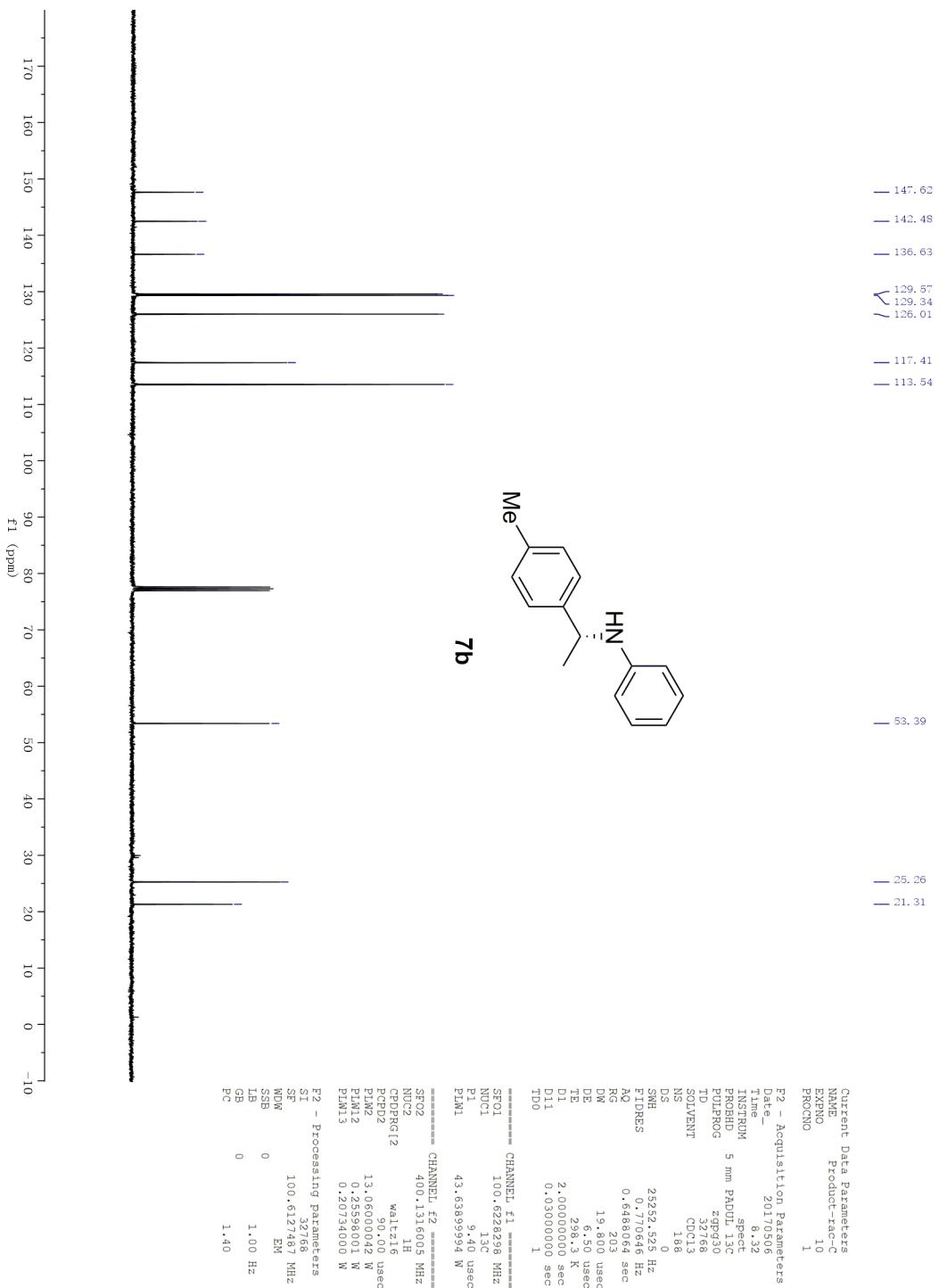


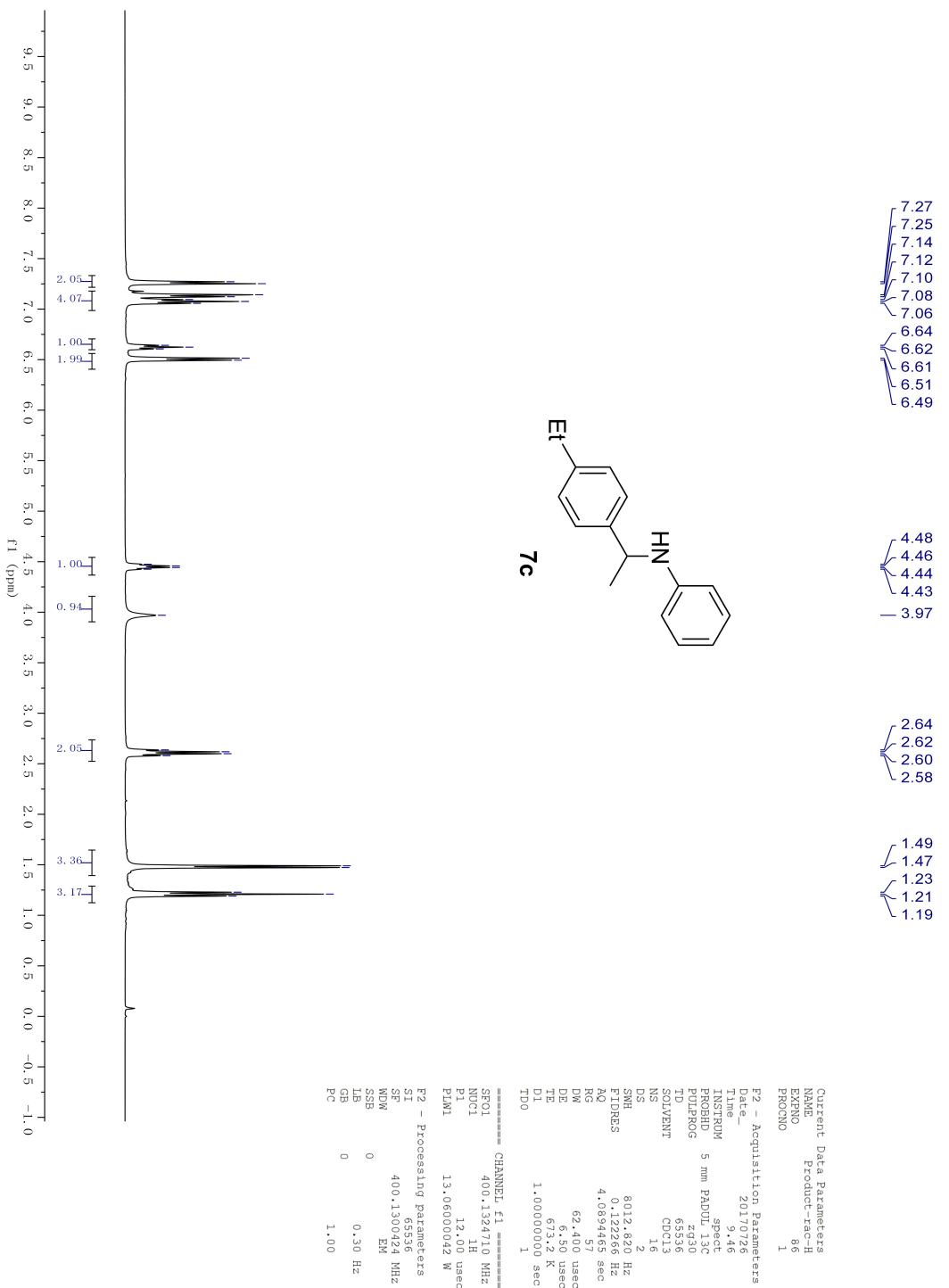
Peak	RT	Area %	Area
#	[min]	-----	-----
1	19.964	57.782	8.256e4
2	22.803	42.218	6.032e4

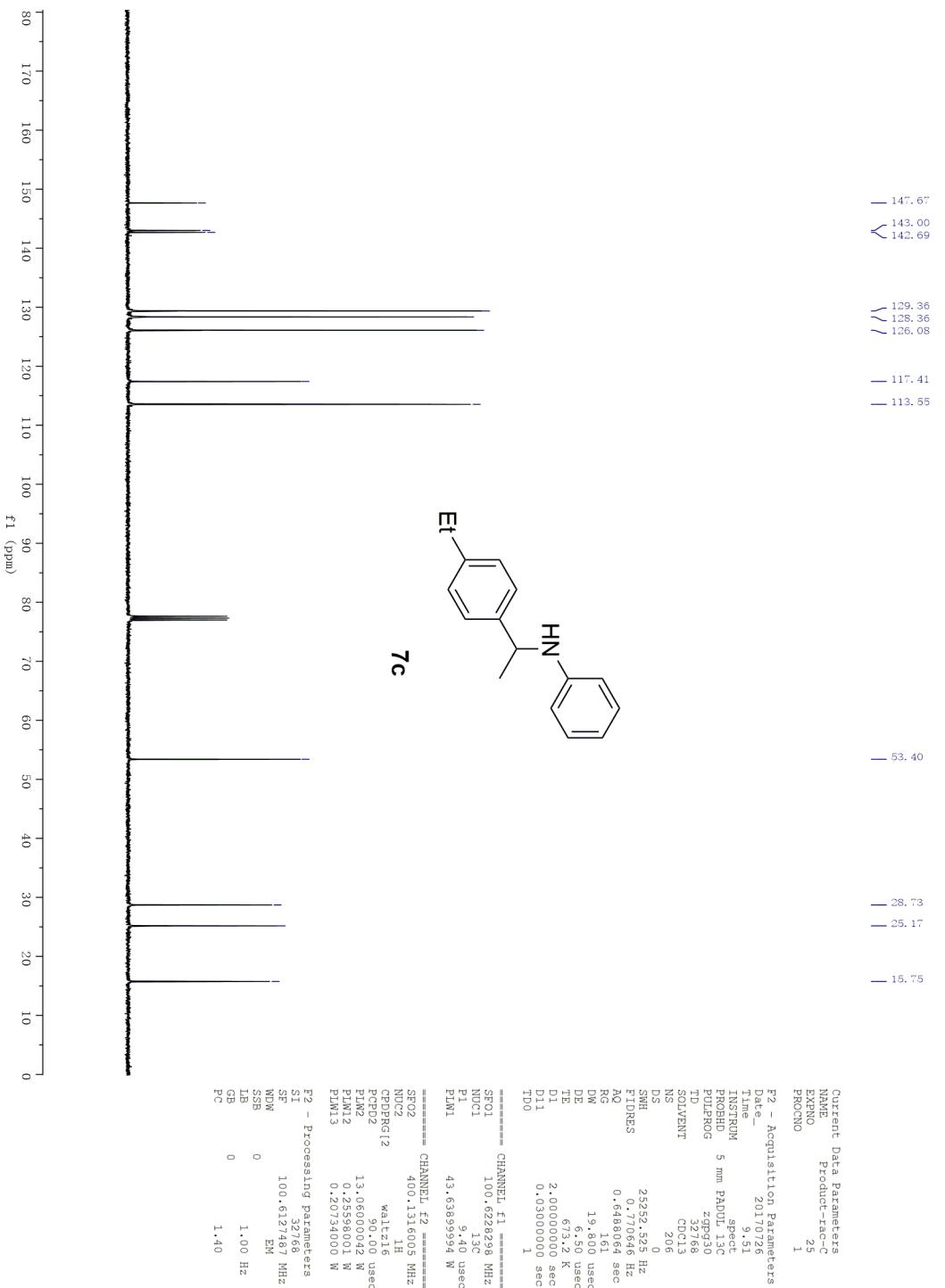


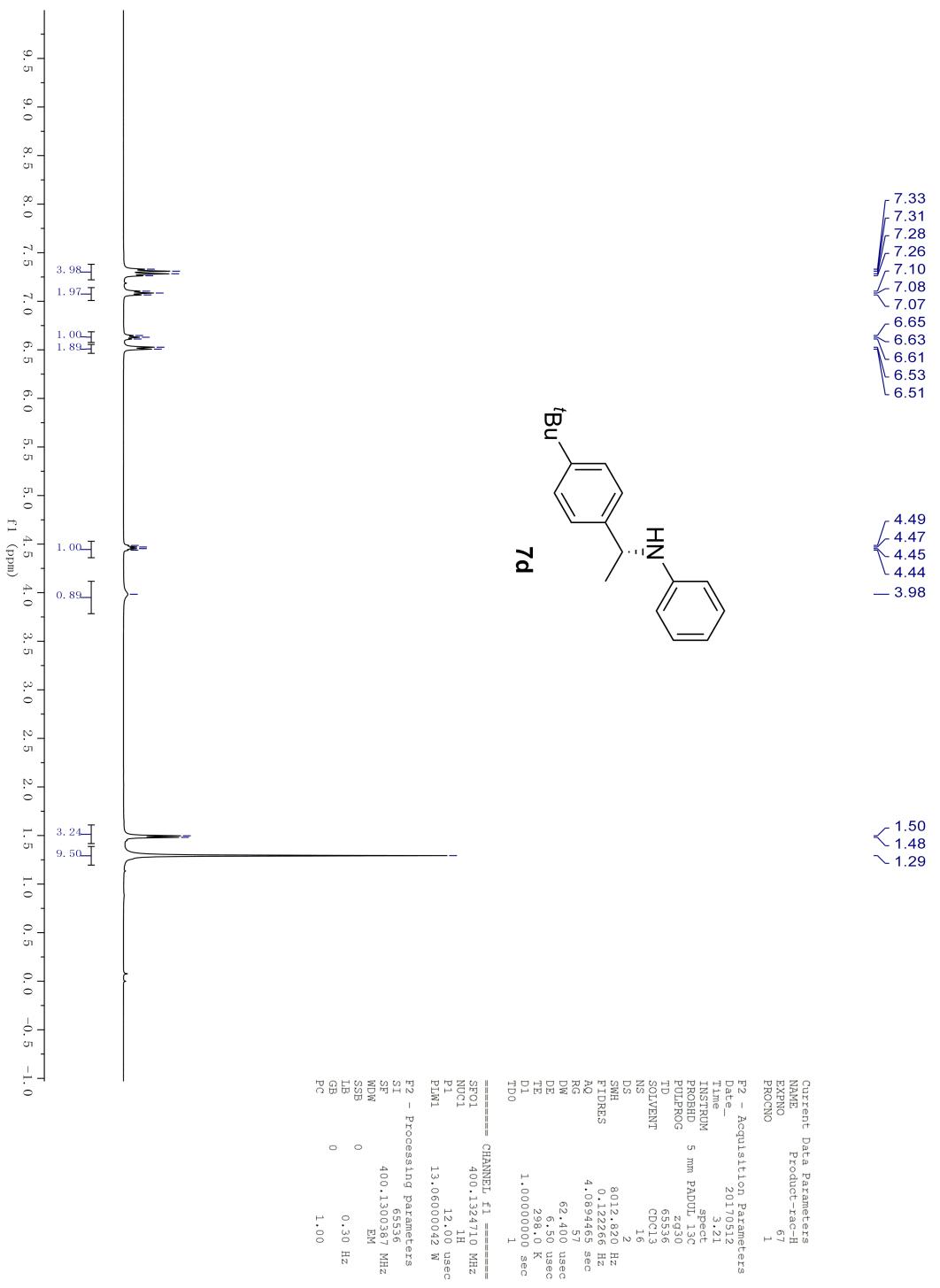


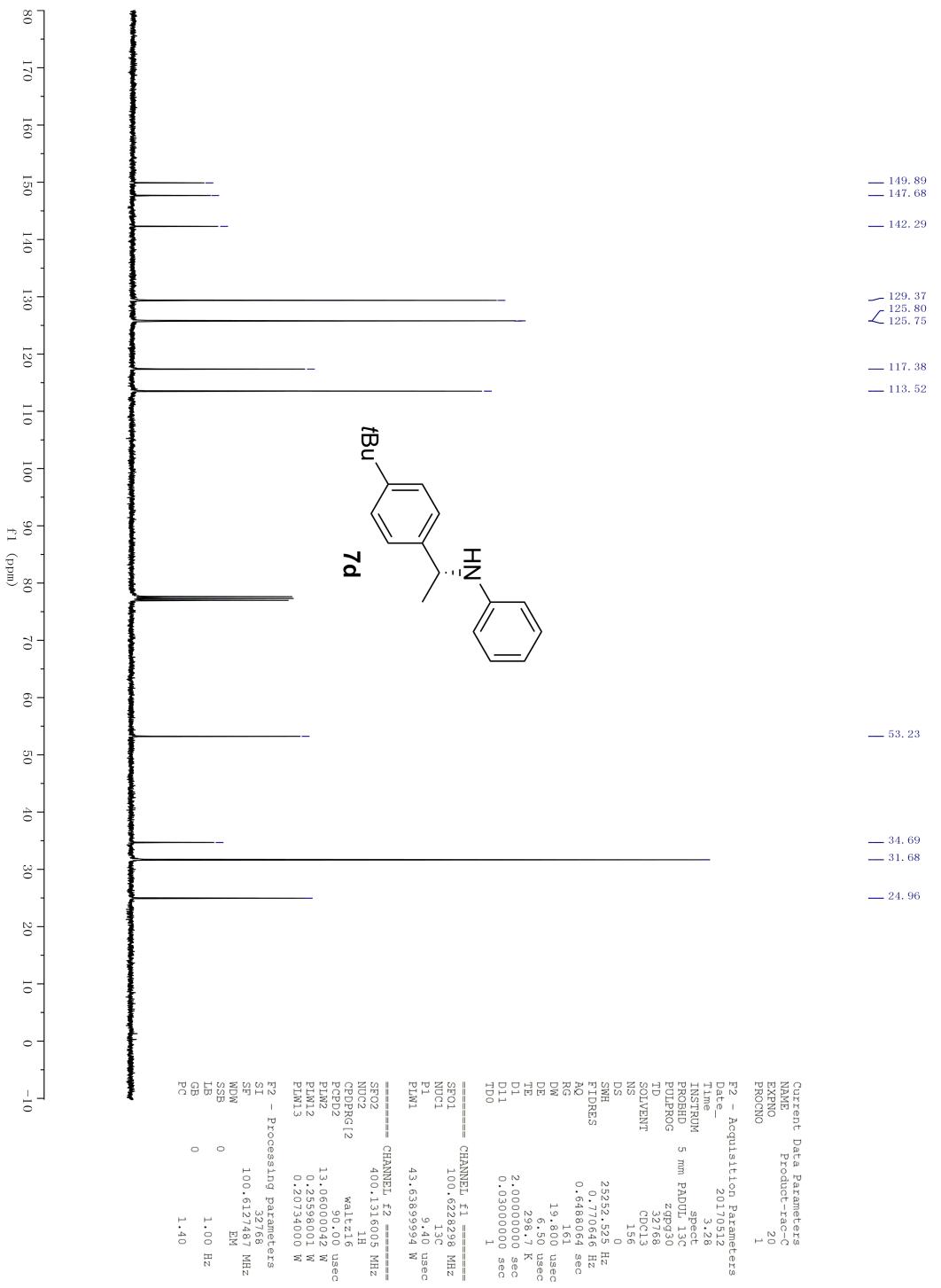


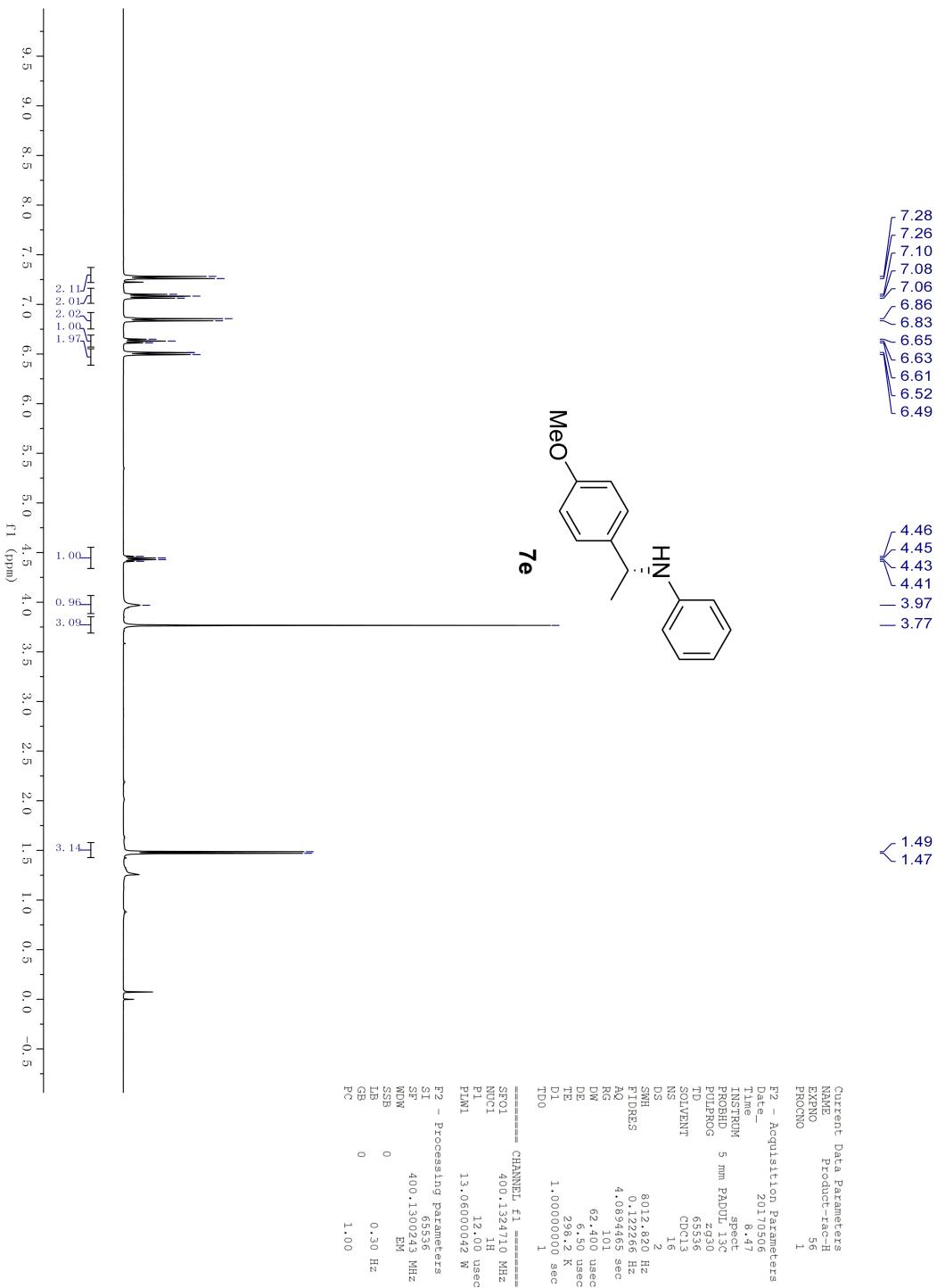


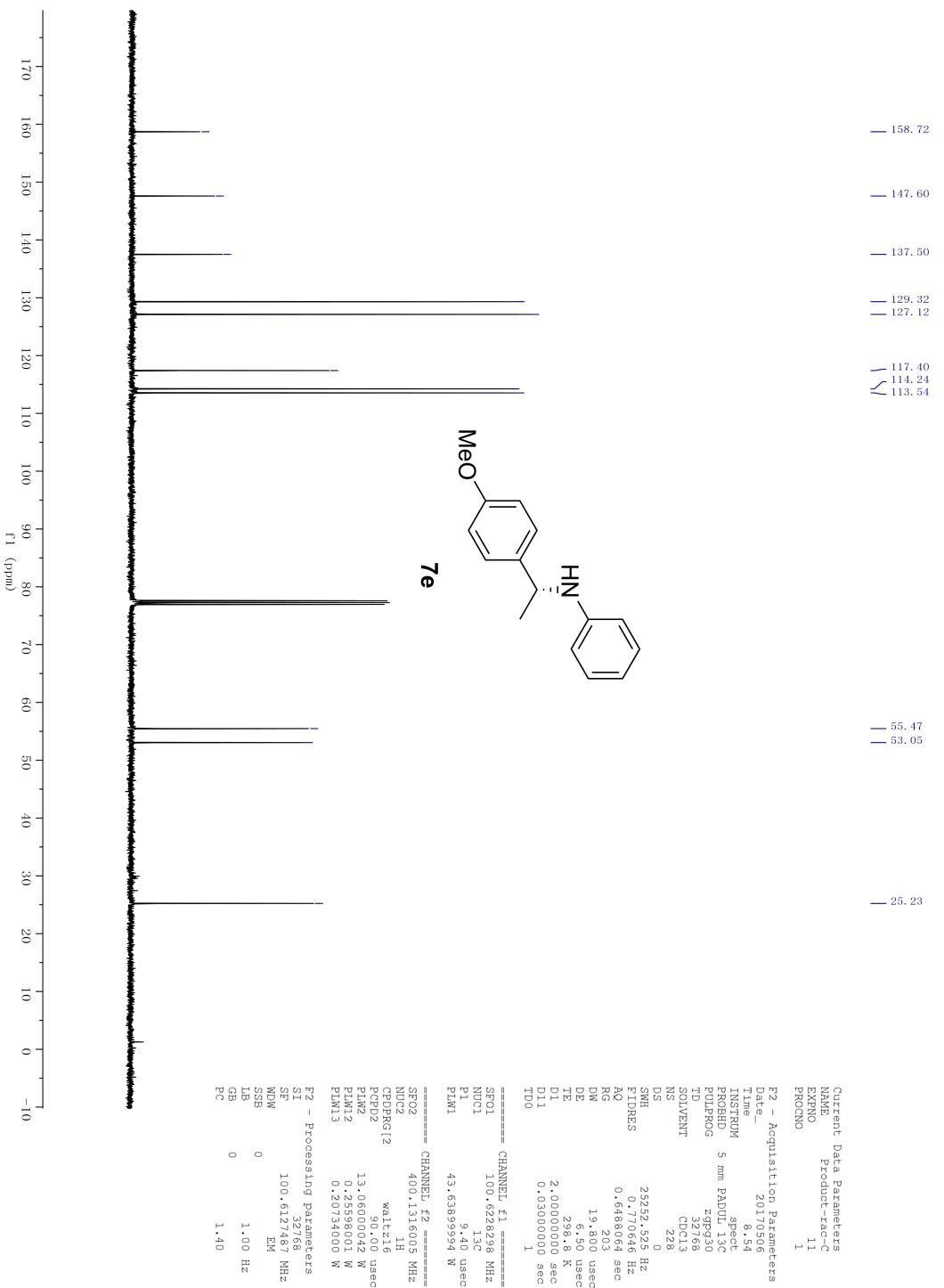


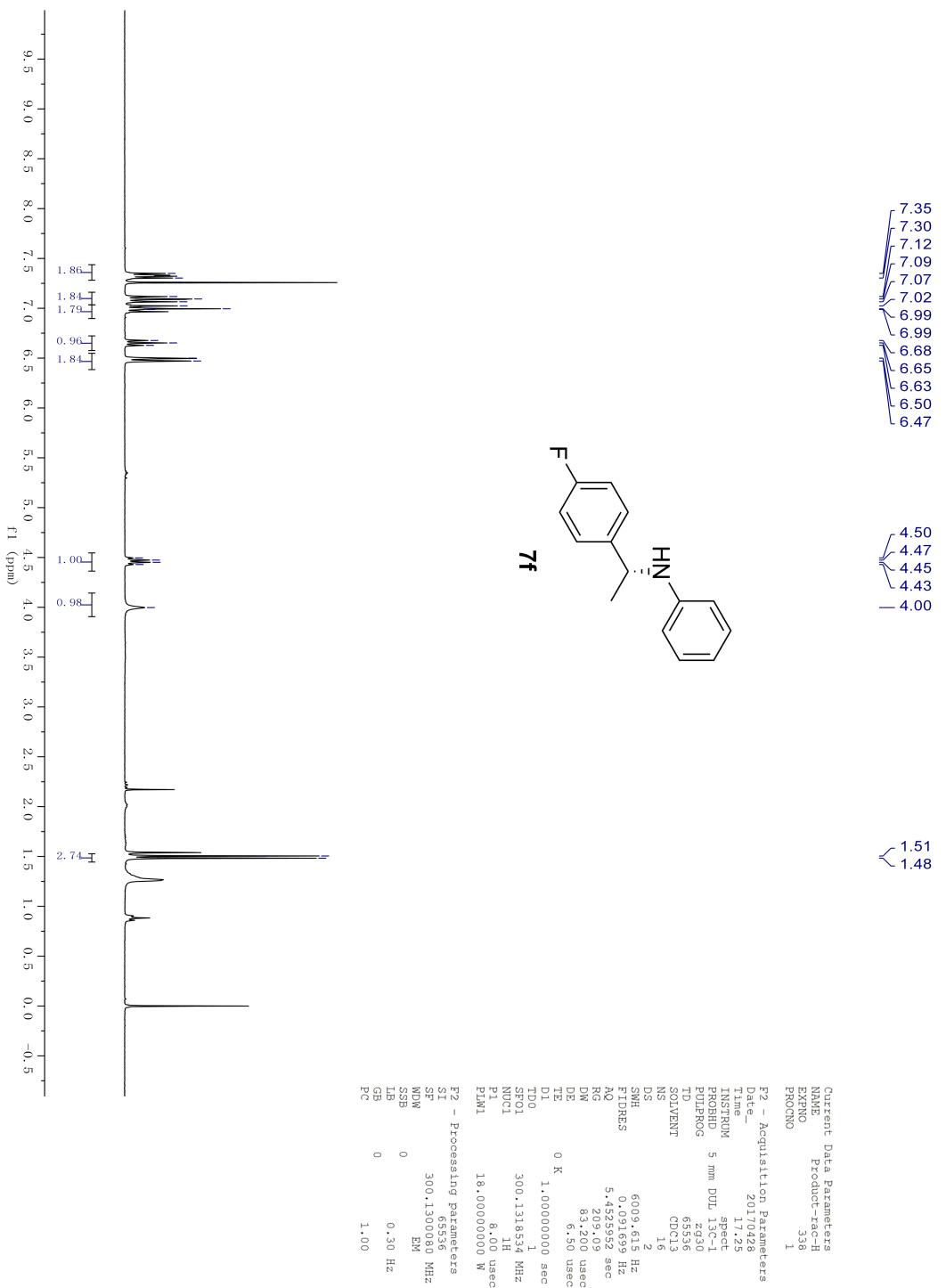


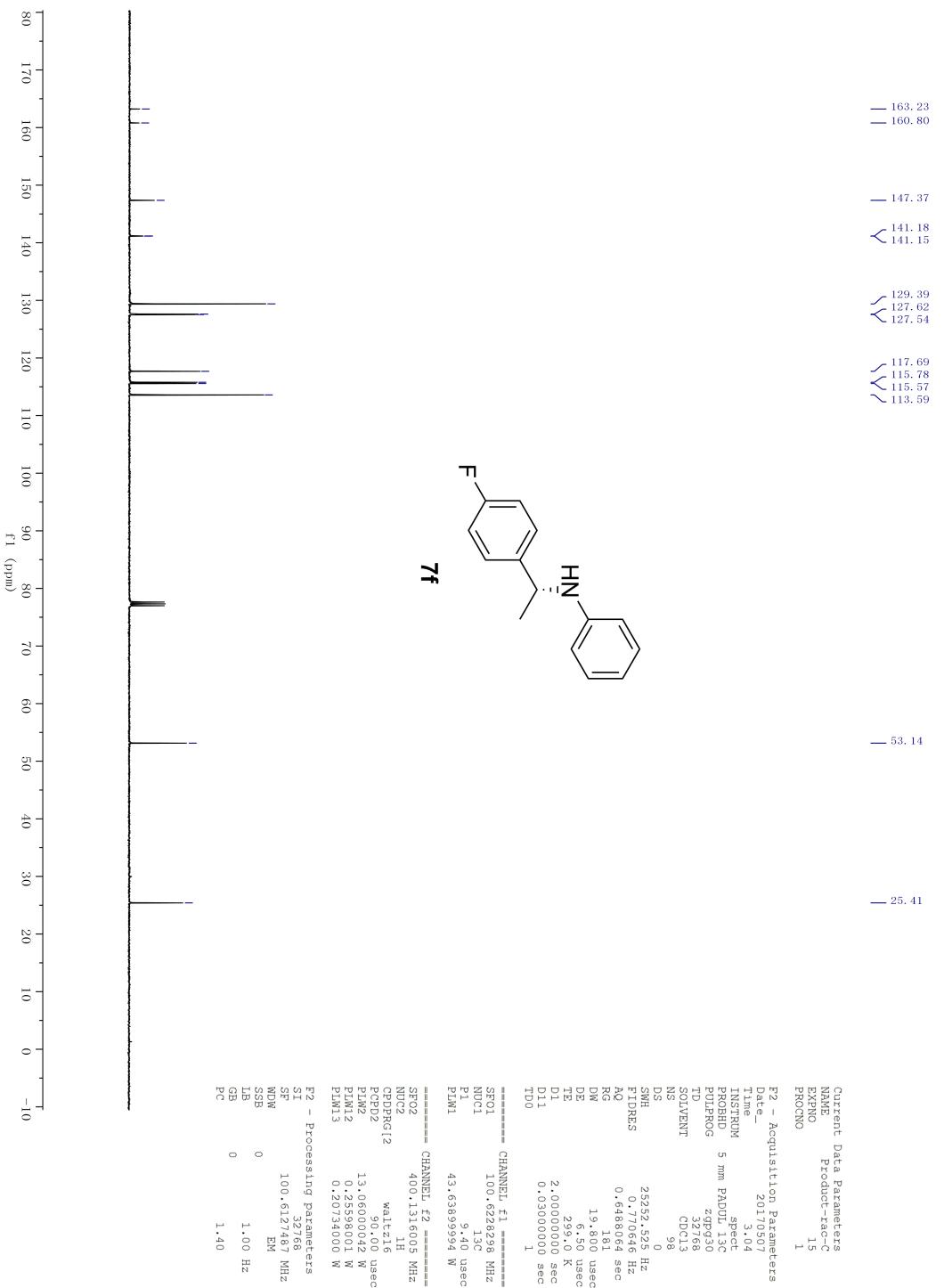


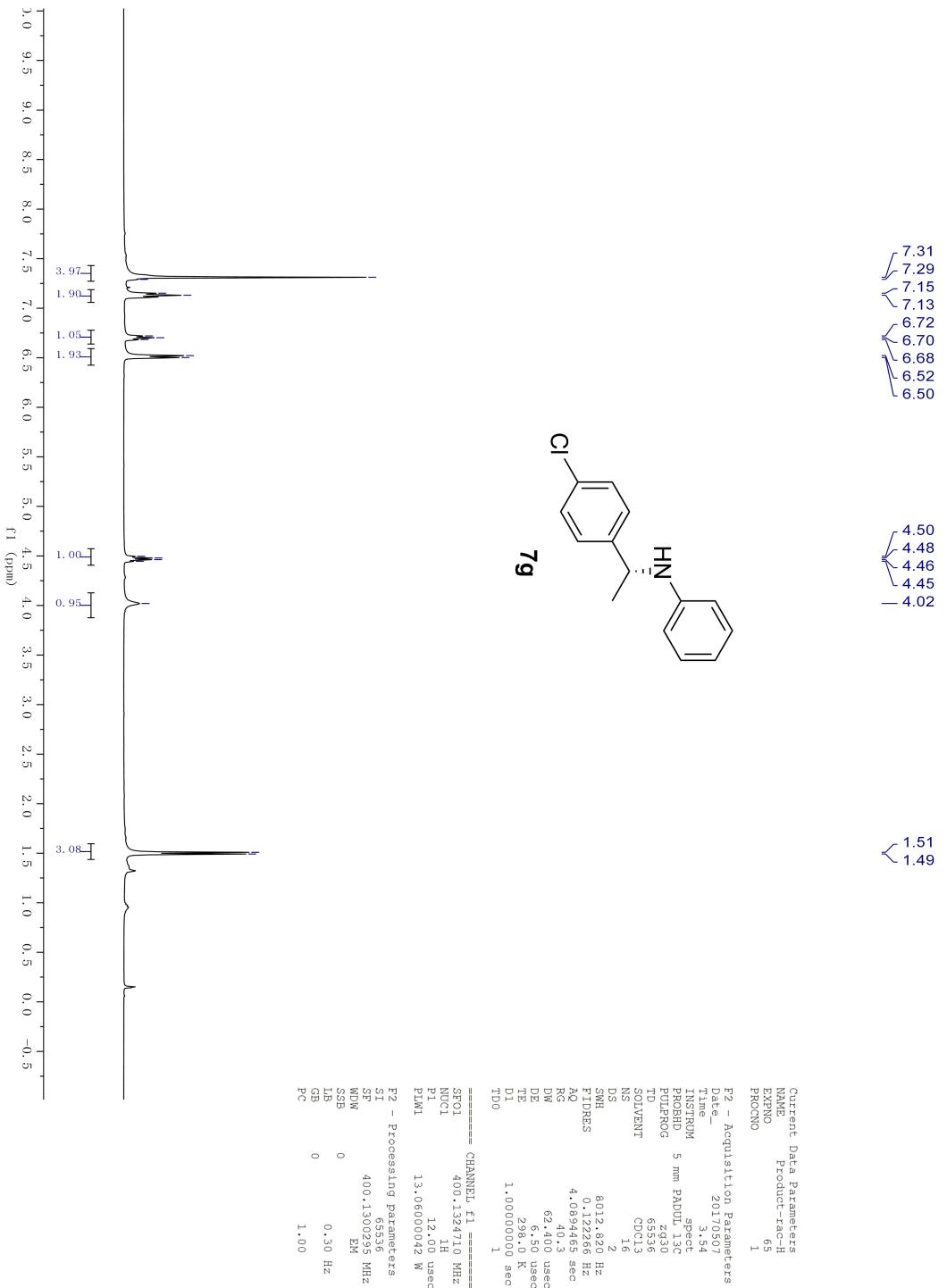


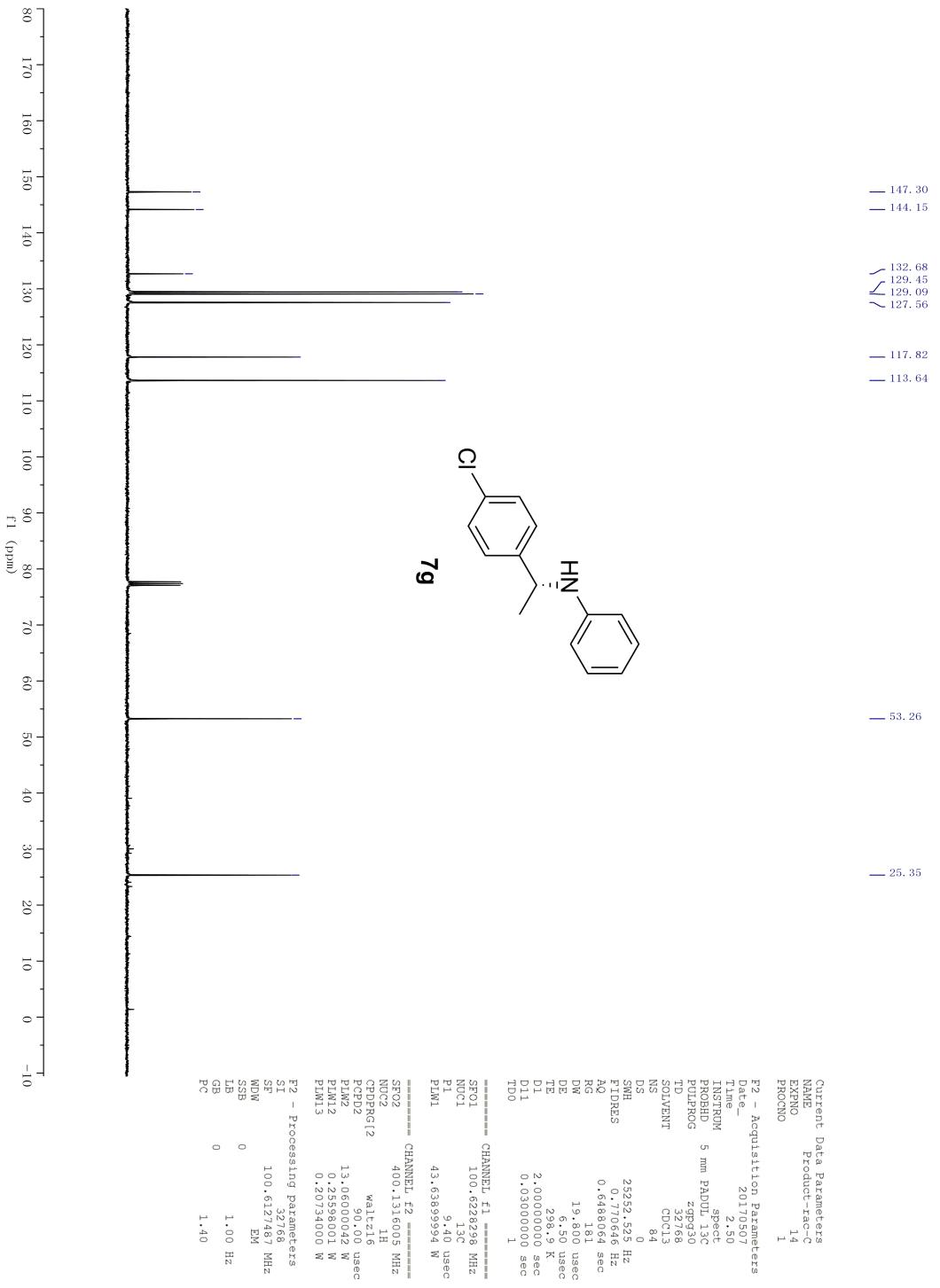




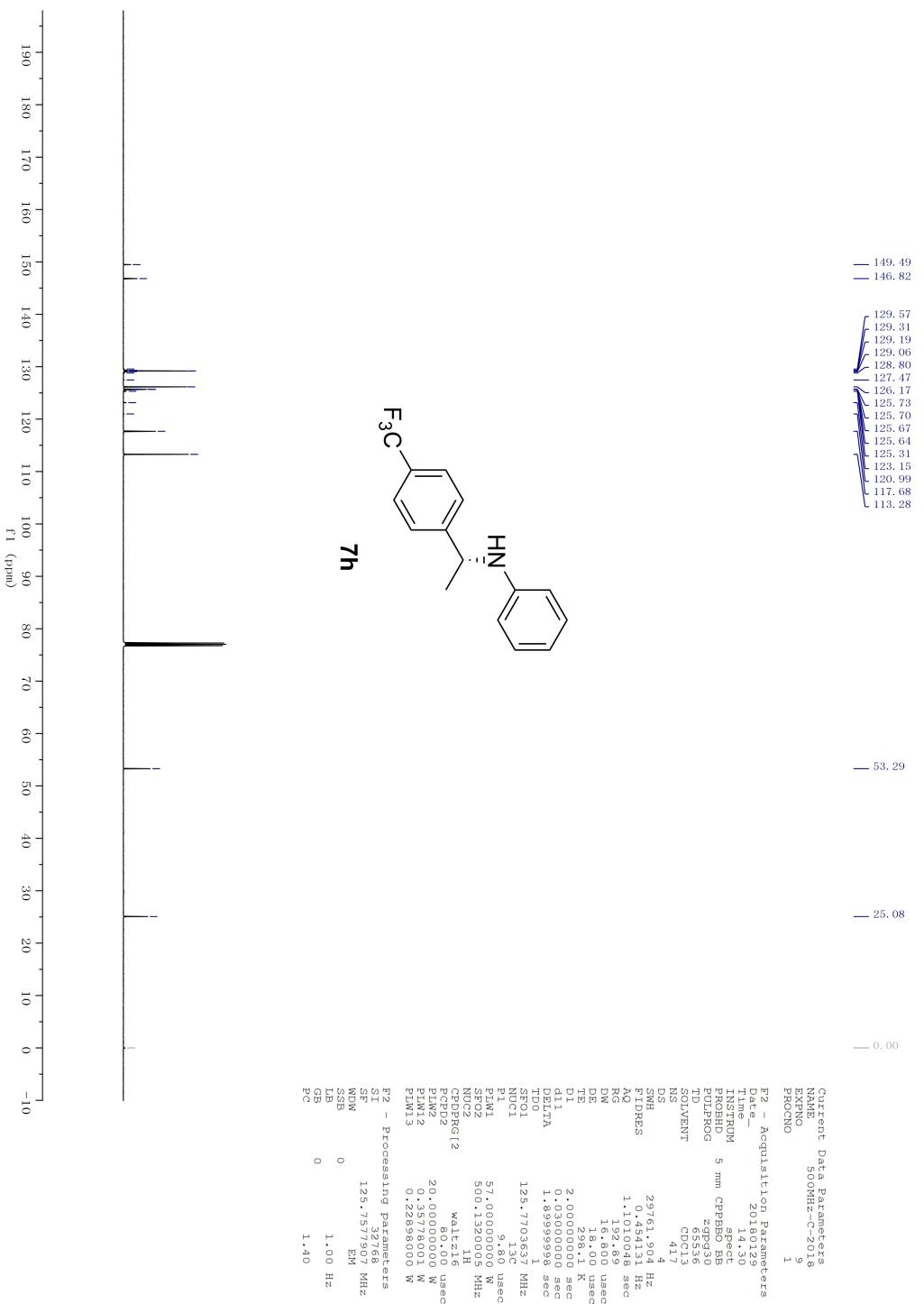


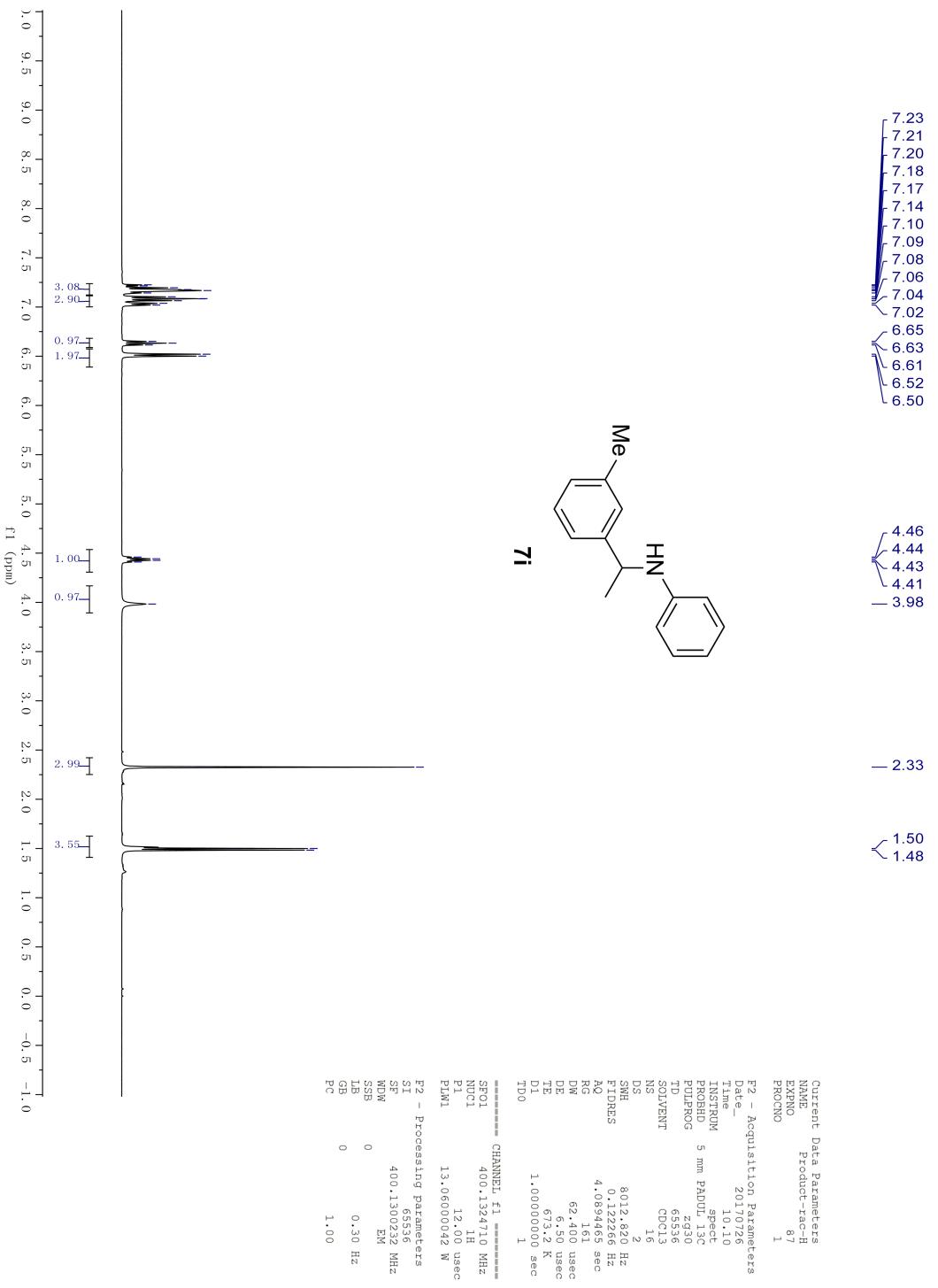


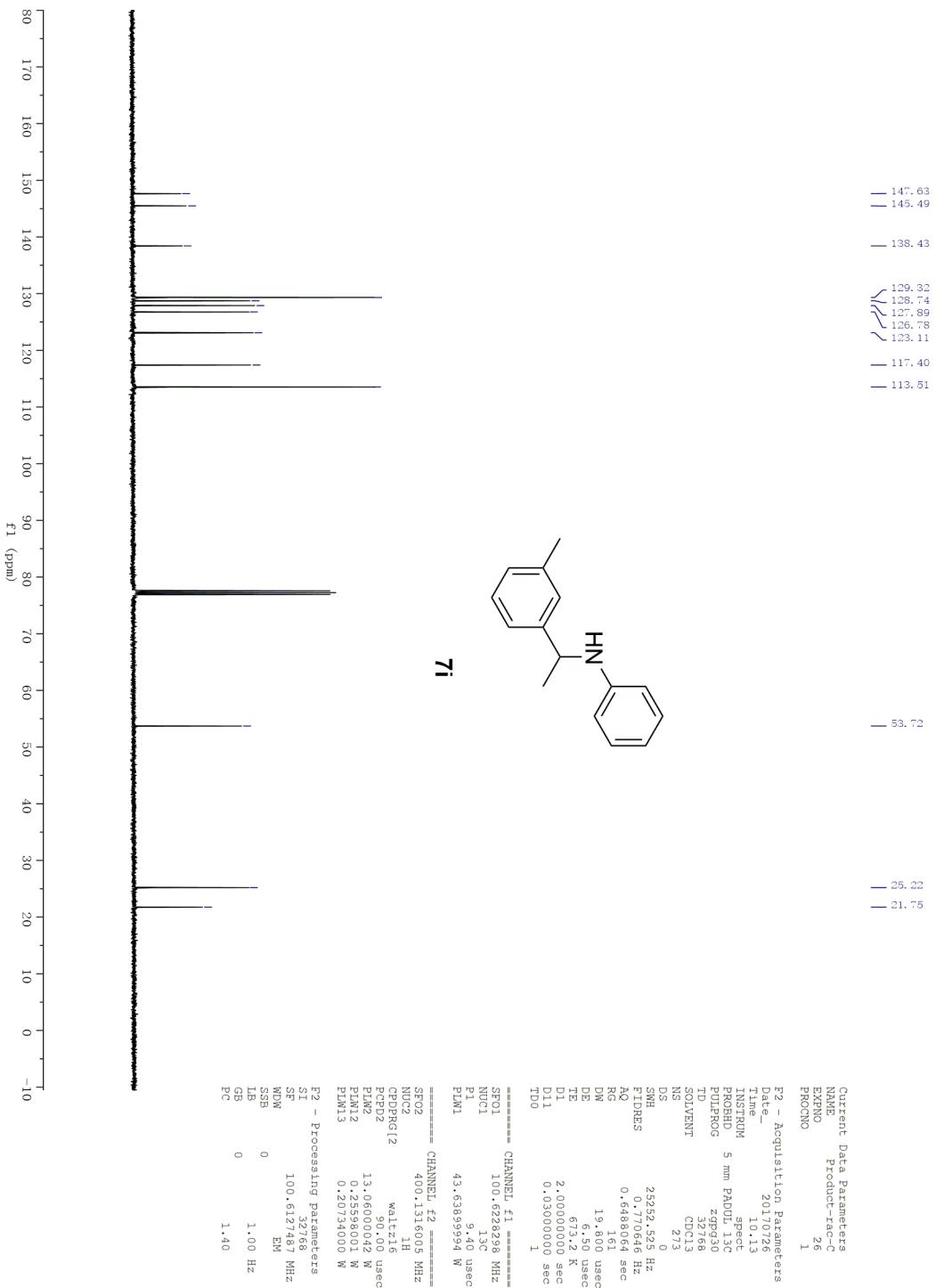


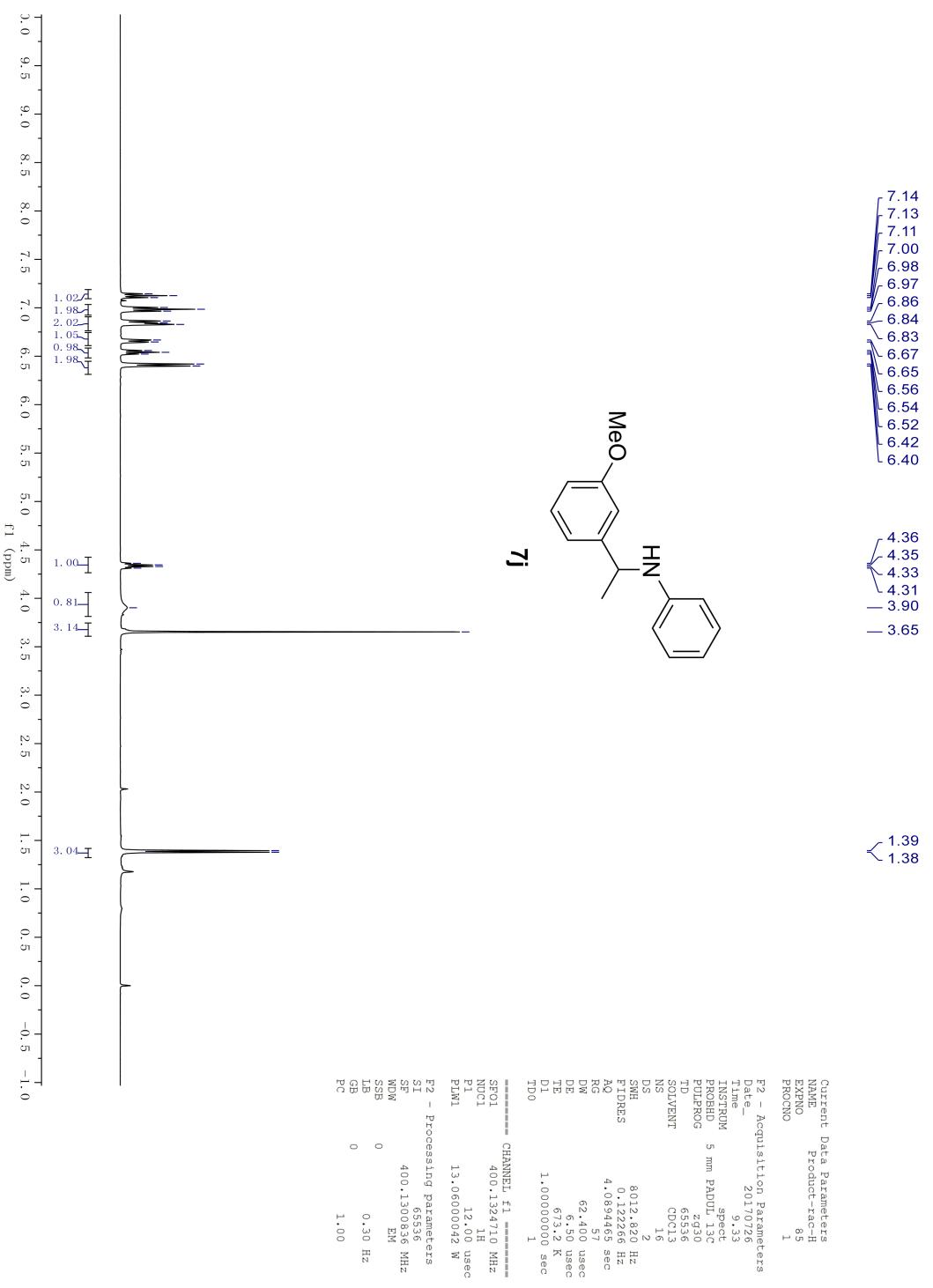


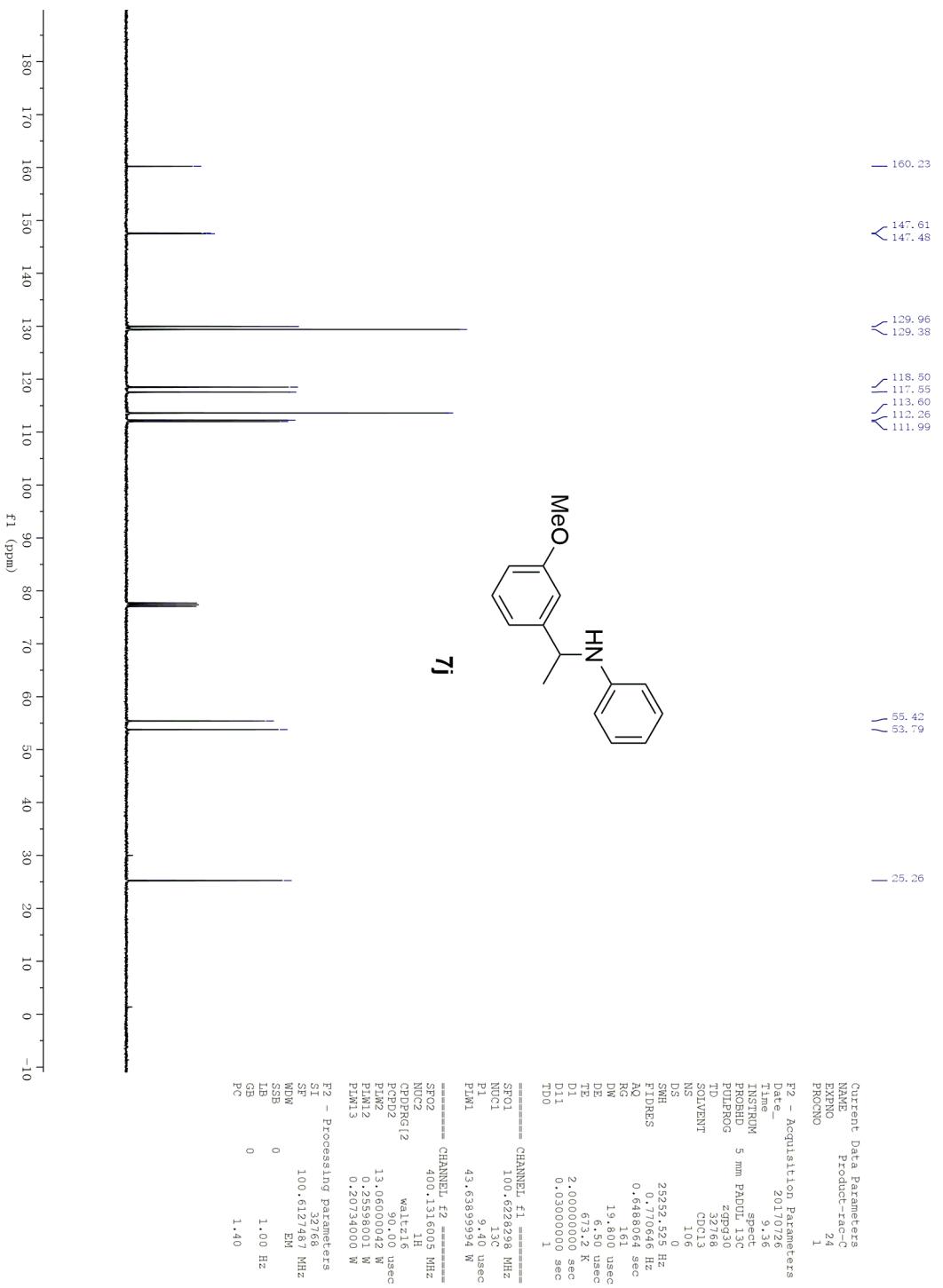


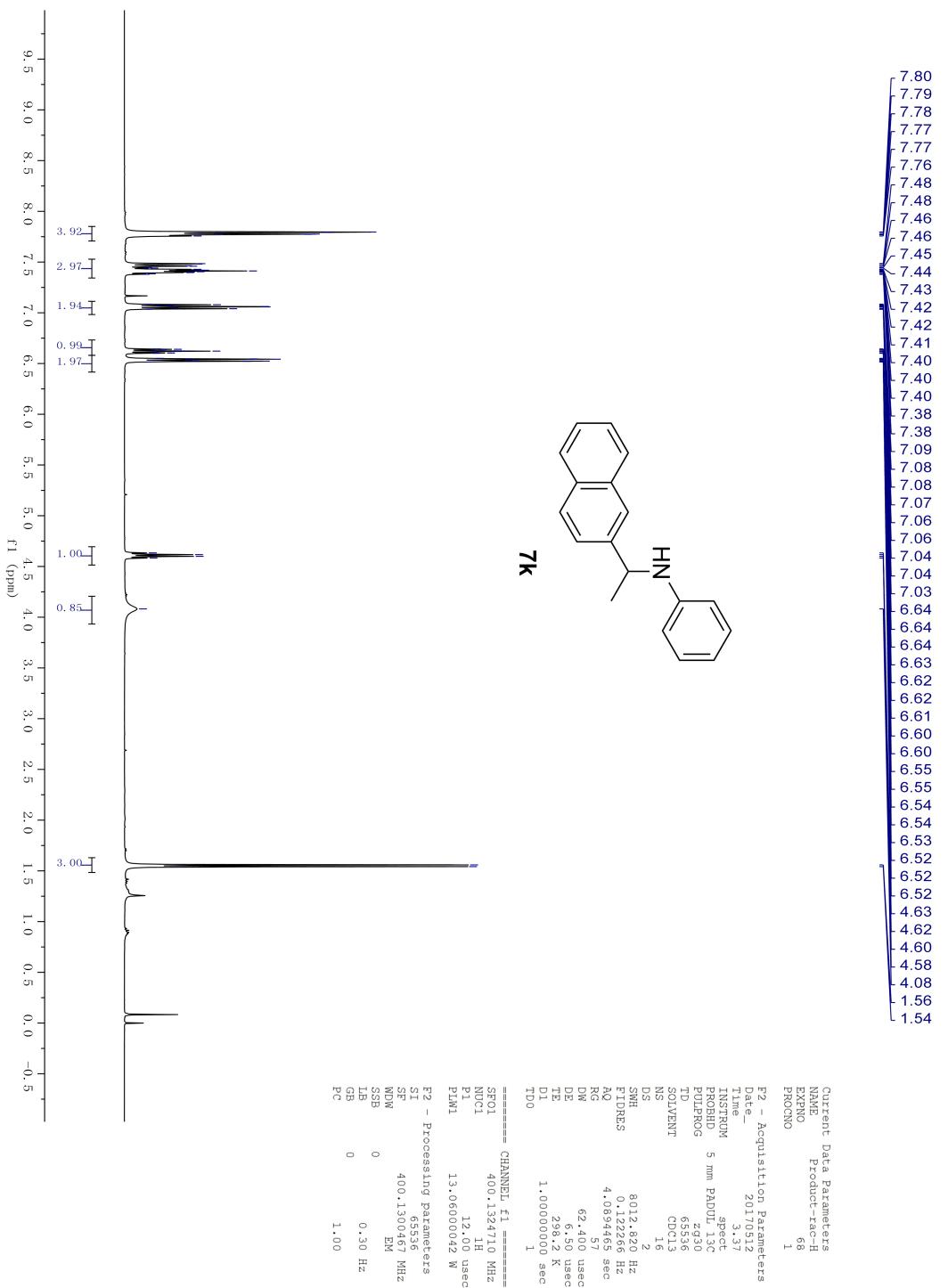


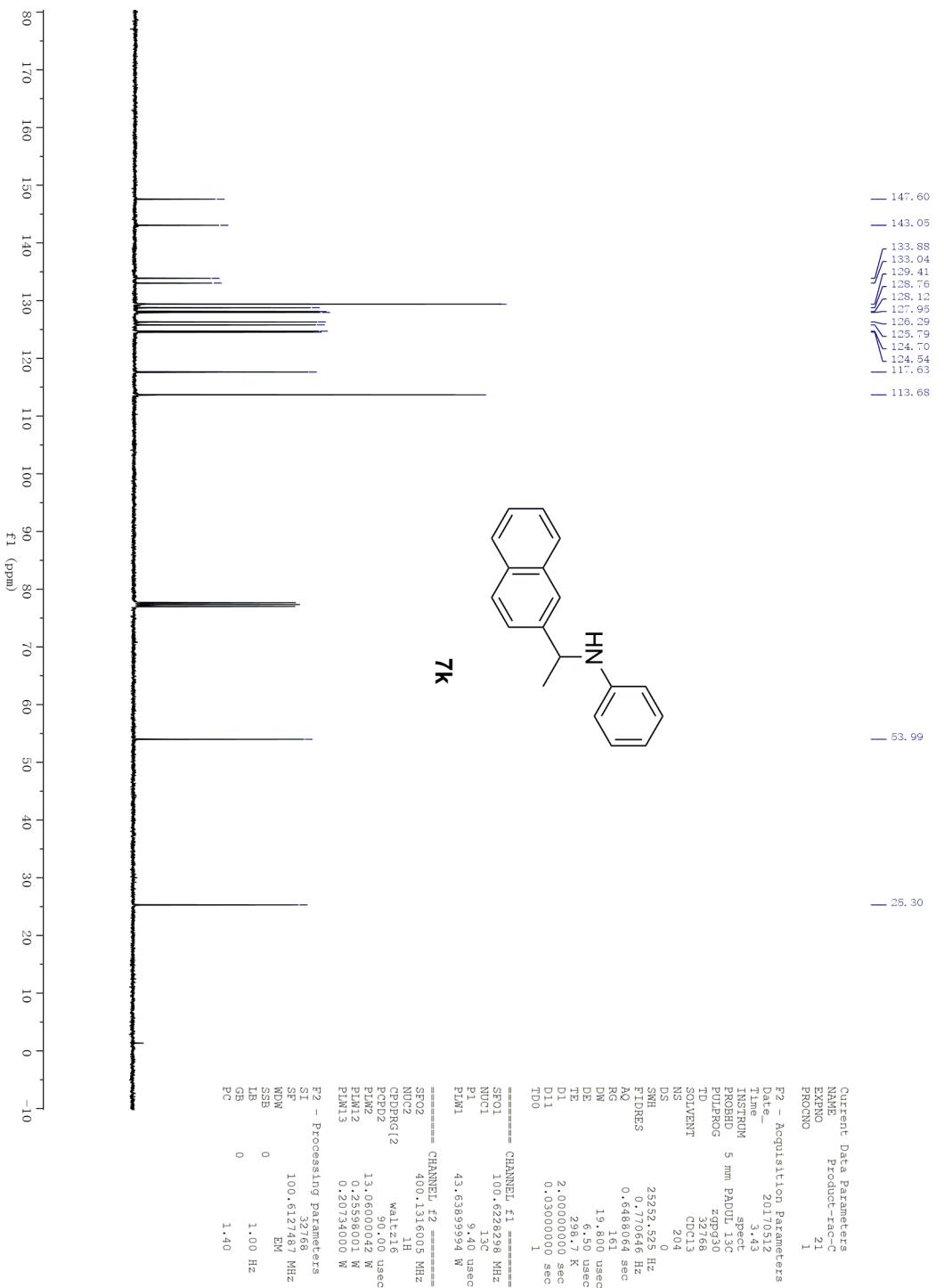


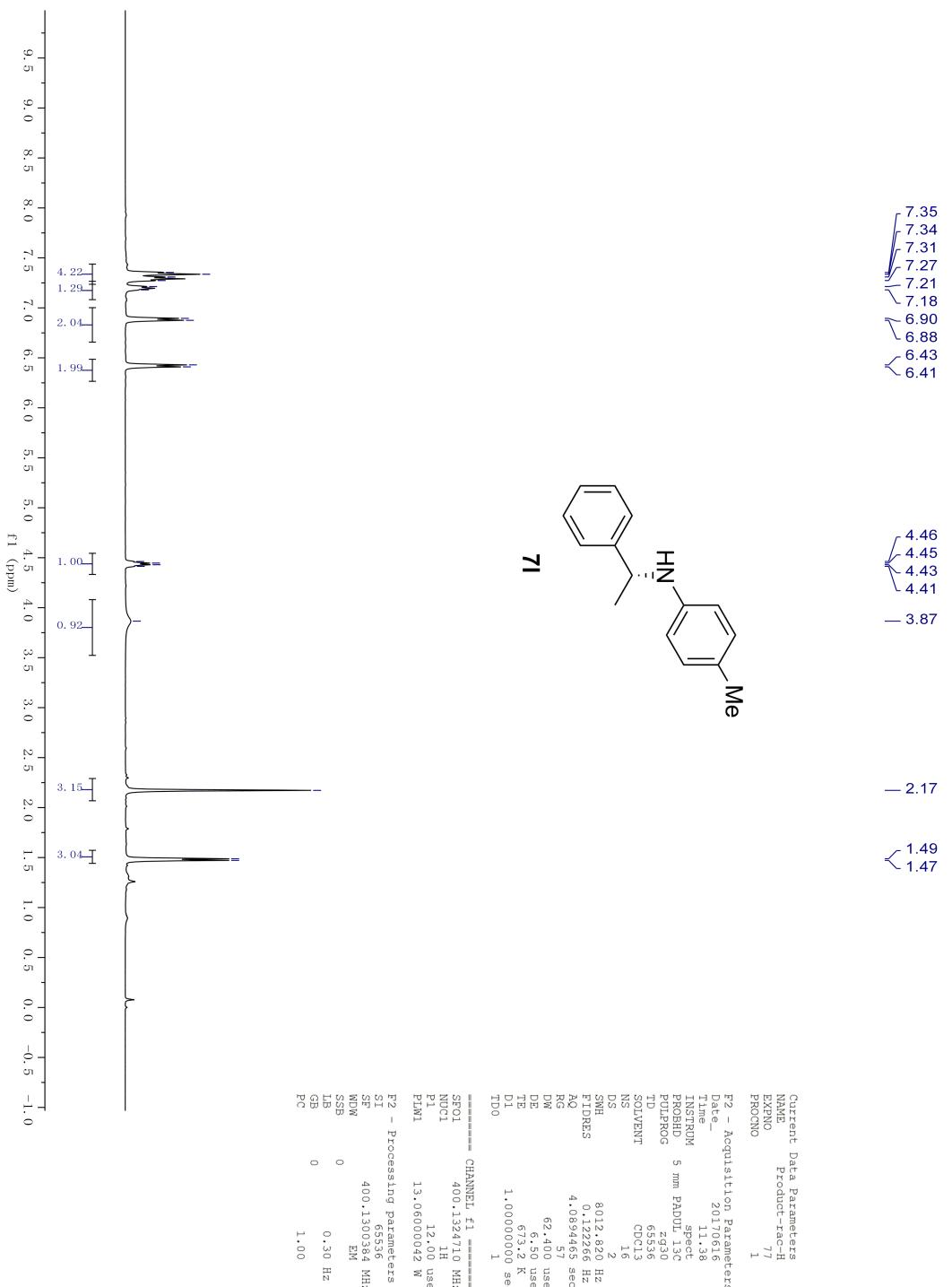


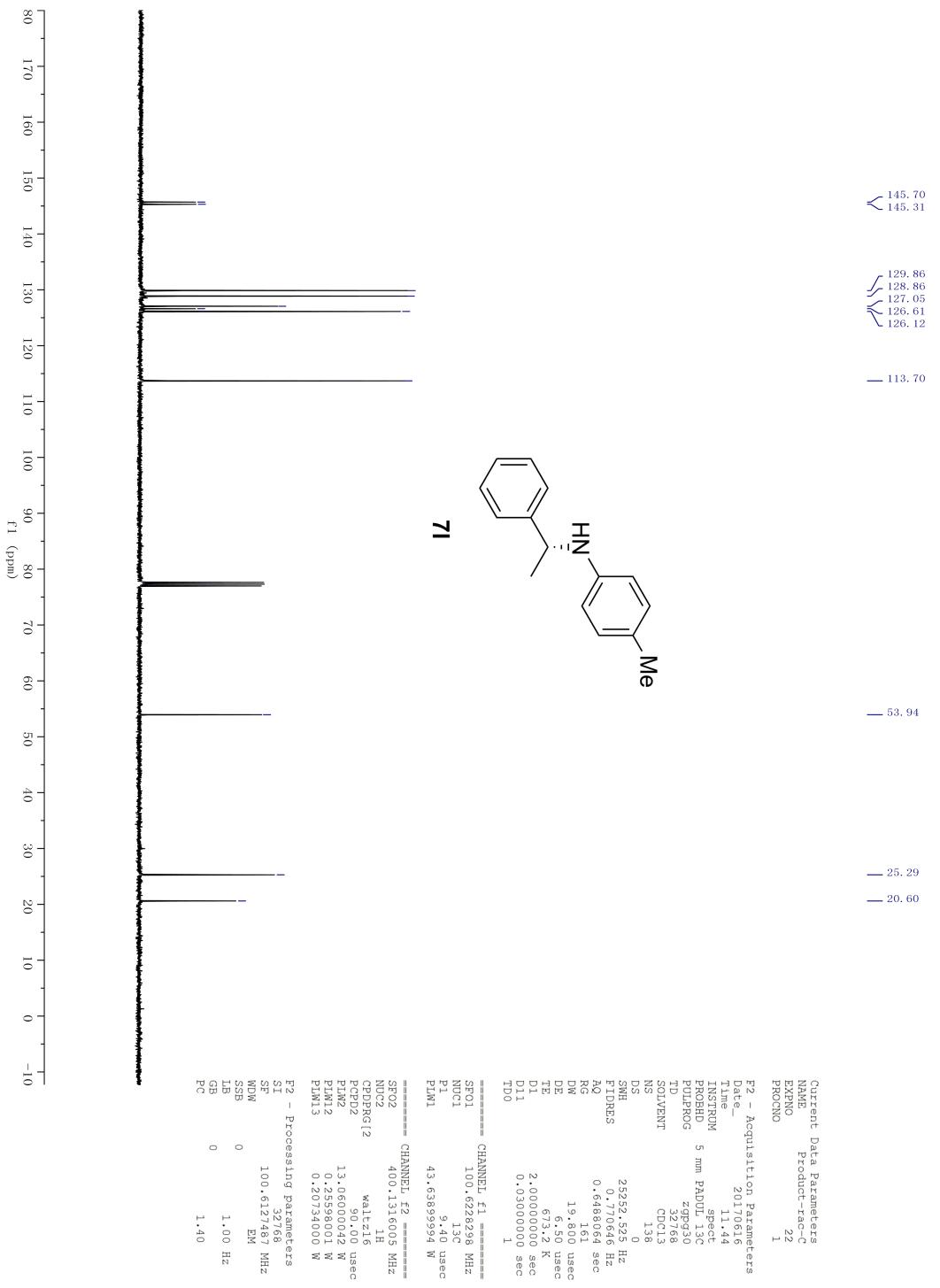


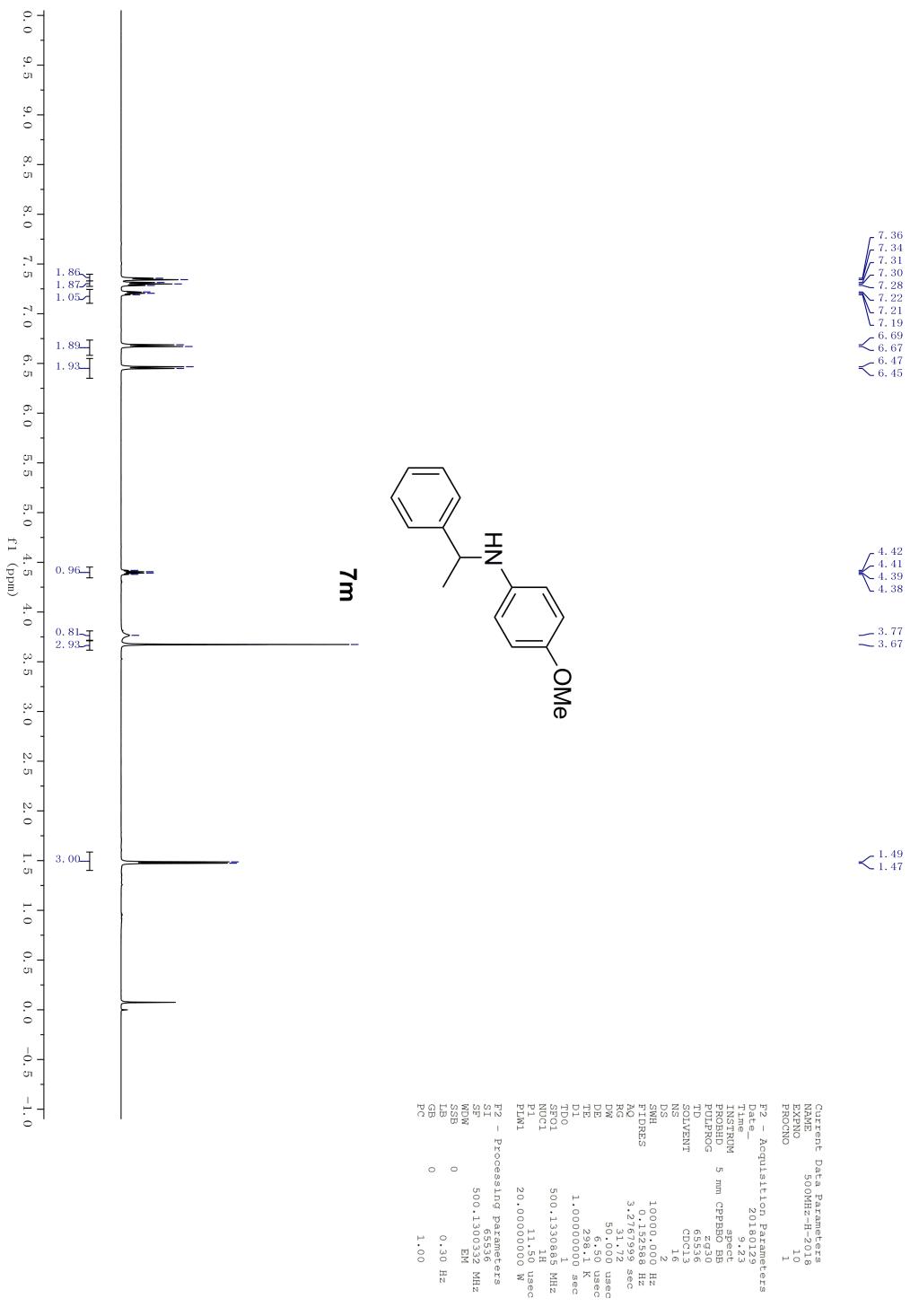


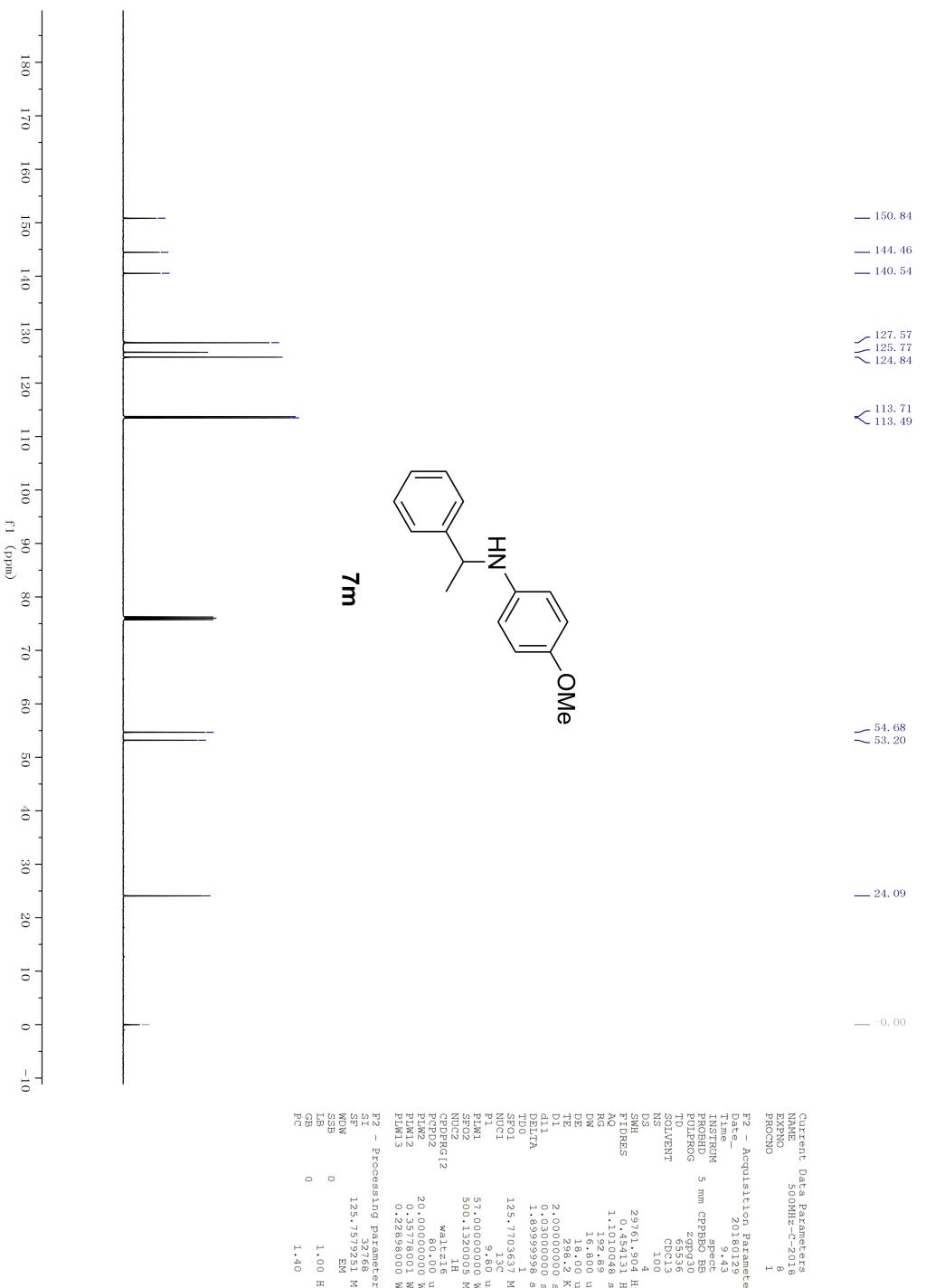


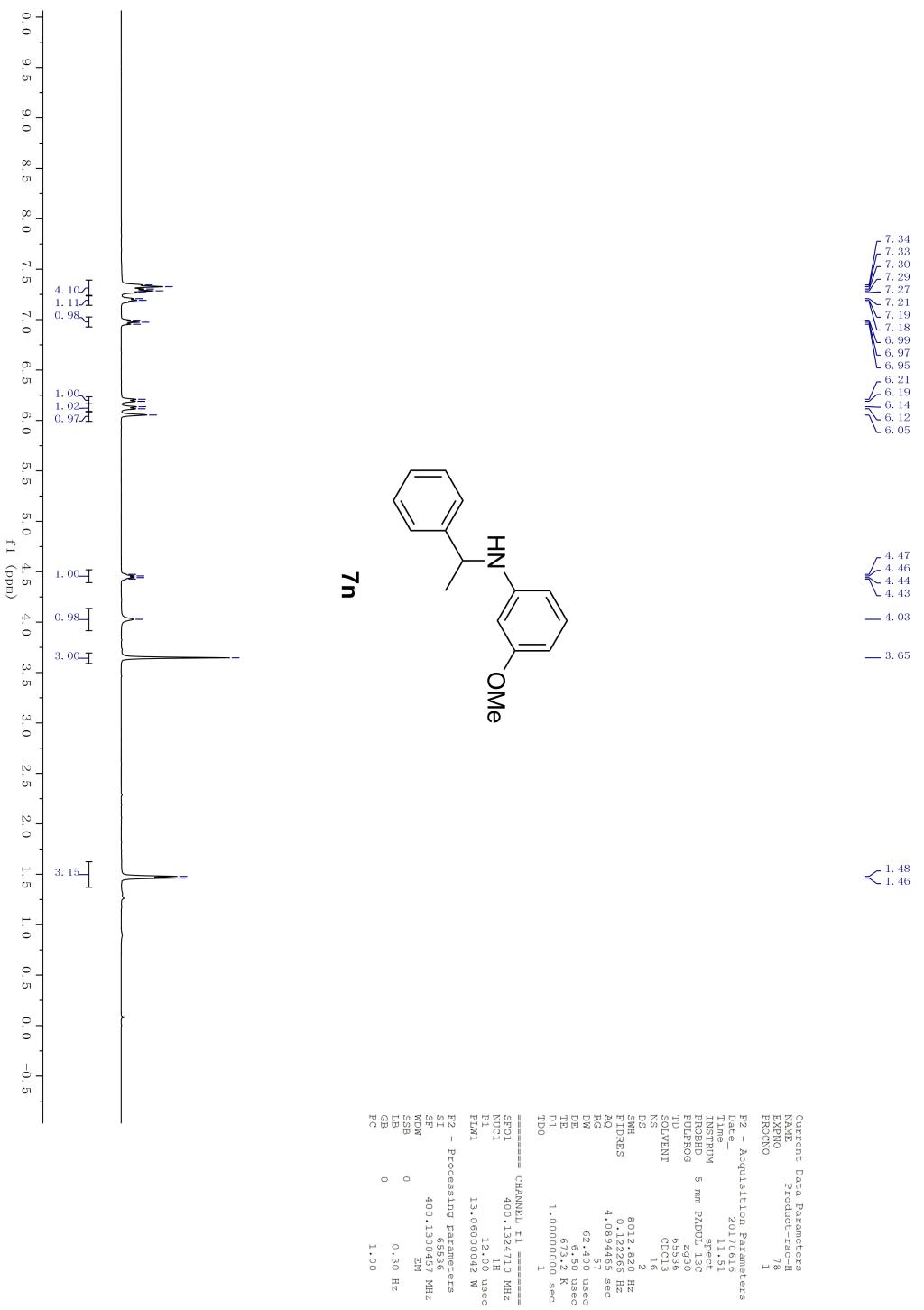


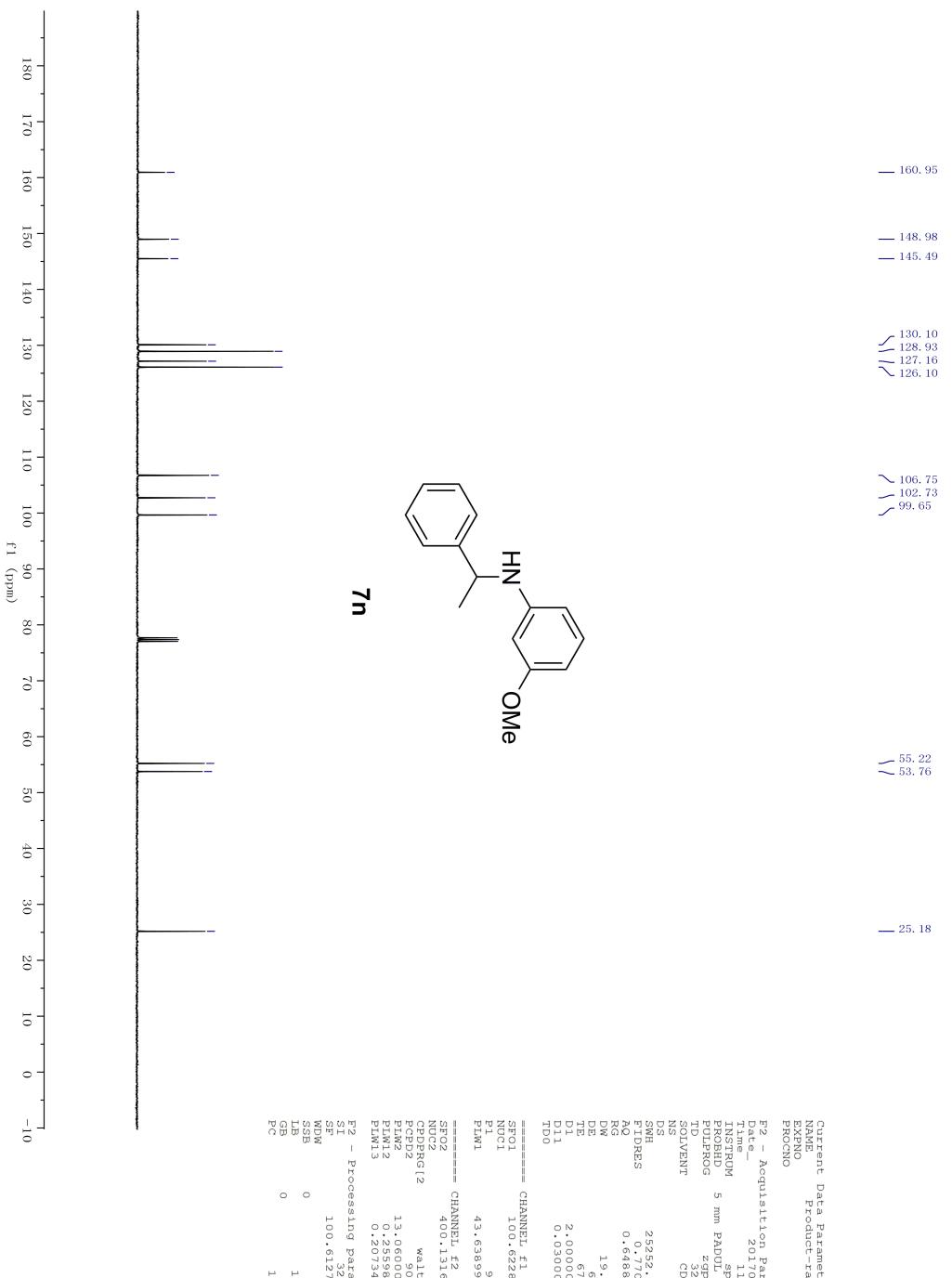










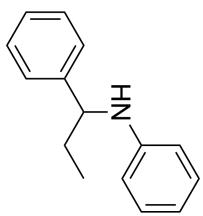


7.33
 7.31
 7.30
 7.29
 7.27
 7.21
 7.20
 7.18
 7.08
 7.06
 7.05
 6.63
 6.61
 6.60
 6.51
 6.49

4.22
 4.21
 4.19
 4.03

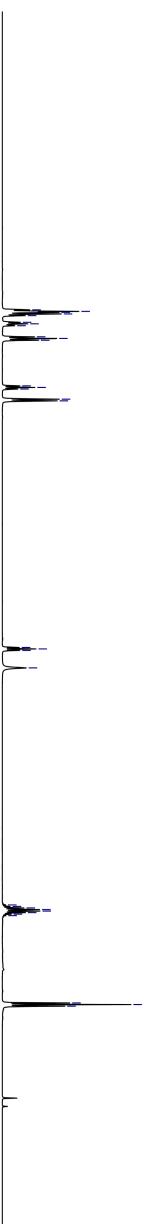
1.85
 1.84
 1.81
 1.80
 1.78
 1.77
 1.75

0.95
 0.94
 0.92



7o

0.0 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 -0.5 -1.0



Current Data Parameters
 NAME 500MHz-H-2018
 EXPNO 9
 PROCNO 1

 F2 – Acquisition Parameters
 Date 20181229
 Time 9.20
 DPPMTRIM 5 mm CPPBBSB
 PWDPRG ZG3D
 PULPROG 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 10000.000 Hz
 FIDRES 0.115384 Hz
 AQ 3.276399 sec
 RG 2.000000 sec
 DW 50.000 usec
 DE 6.50 usec
 TE 298.1 K
 D1 1.0000000 sec
 TDO 1
 SFO1 500.1330885 MHz
 NUC1 1H
 F1WL 11.50 usec
 F1ML 20.0000000 W

 F2 – Processing parameters
 SI 65536
 SF 500.1330513 MHz
 WDW 0
 EM 0
 L1B 0.30 Hz
 GB 0
 PC 1.00

