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#### Formal [4+2] Cycloaddition of Imines with Alkoxyisocoumarins

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### I. General information

Reagents and solvents were purchased from commercial sources and were purified by distillation or recrystallization prior to use. Purification of reaction products was carried out by flash column chromatography using EM Reagent silica gel 60 (230-400 mesh). All reactions were run under a nitrogen atmosphere unless stated otherwise. Analytical thin layer chromatography was performed on EM Reagent 0.25 mm silica gel 60  $F_{254}$  plates. Visualization was accomplished with UV light, and potassium permanganate, Dragendorff-Munier and anisaldehyde stains, followed by heating. Melting points were recorded on a Thomas Hoover capillary melting point apparatus and are uncorrected. Infrared spectra were recorded on an ATI Mattson Genesis Series FT-Infrared spectrophotometer. Proton nuclear magnetic resonance spectra (<sup>1</sup>H-NMR) were recorded on a Varian VNMRS-500 MHz instrument and are reported in ppm using solvent as an internal standard (CDCl<sub>3</sub> at 7.26 ppm). Data are reported as app = apparent, s = singlet, d = doublet, t = triplet, q = quartet, p = pentet, br = broad, dd = doublet of doublets, dt = doublet of triplets, td = triplet of doublets, ddd = doublet of doublets, ddd = doublet of doublet of doublet of doublets, dddd = doublet of doublet of doublet of doublet of doublets, m = multiplet, comp = complex; and coupling constant(s) in Hz. Protondecoupled carbon nuclear magnetic resonance spectra (<sup>13</sup>C-NMR) were recorded on a Varian VNMRS-500 MHz instrument and are reported in ppm using solvent as an internal standard (CDCl<sub>3</sub> at 77.16 ppm). Mass spectra were recorded on a Finnigan LCO-DUO mass spectrometer or on a Finnigan 2001 Fourier Transform Ion Cyclotron Resonance Mass Spectrometer. HPLC analysis was carried out on an Agilent 1100 series instrument with auto sampler and multiple wavelength detectors. Optical rotations were measured using a 1 mL cell with a 1 dm path length on a Jasco P-2000 polarimeter at 589 nm and at 20 °C. Racemic products 5a-5d were prepared using racemic 2a. Racemic products 6a-6d were prepared using catalytic 1,3-bis(3,5-Compound  $3a^2$ , homophthalic anhydride  $1^3$ , 3,4bis(trifluoromethyl)phenyl)thiourea.<sup>1</sup> dihydroisoquinoline,<sup>4</sup> 6,7-dimethoxy-3,4-dihydroisoquinoline,<sup>5</sup> 6-chloro-3,4dihydroisoquinoline,<sup>6</sup> 1-methyl-3,4-dihydroisoquinoline,<sup>7</sup> 6-methoxy-3,4-dihydroisoquinoline,<sup>8</sup> and imine precursors<sup>9</sup> were prepared according to previously published procedures. Catalysts  $2a^{10}$  and  $2b^{11}$  were prepared according to previously reported procedures.

### **II. Reaction optimization**

### Evaluation of catalysts.<sup>a</sup>





<sup>a</sup> Yields are those of chromatographically purified compounds. The ee values were determined by HPLC analysis.

### Evaluation of sulfonamide catalysts.<sup>a</sup>



<sup>a</sup> Yields are those of chromatographically purified compounds. The ee values were determined by HPLC analysis. <sup>b</sup> 30 h with PhCF<sub>3</sub> as the solvent. <sup>c</sup> 26 h with PhCF<sub>3</sub> as the solvent

## Evaluation of additives and solvents.<sup>a</sup>



0.2 mmol

**2b** (20 mol%) additive (X mol%) solvent (0.05 M), rt, 72 h



entry	additive (mol%)	solvent	yield (%)	dr	ee (%)
1	0	DLM	50	× 10-1 da	50
1	0	Phile	50	>19:1 dr	52
$2^{a}$	0	PhCF <sub>3</sub>	47	>19:1 dr	56
3	0	DCM	70	>19:1 dr	49
4	0	CHCl <sub>3</sub>	61	>19:1 dr	46
5	0	PhMe:DCM (3:1)	68	>19:1 dr	52
6	0	PhMe:DCM (9:1)	51	>19:1 dr	31
7	Acetic Acid (20)	PhMe:DCM (3:1)	50	>19:1 dr	30
8	Acetic Acid (50)	PhMe:DCM (3:1)	15	>19:1 dr	52
9	Benzoic Acid (20)	PhMe:DCM (3:1)	15	>19:1 dr	35
10	Benzoic Acid (50)	PhMe:DCM (3:1)	28	>19:1 dr	32
11	Pyridine (20)	PhMe:DCM (3:1)	80	>19:1 dr	32
12 <sup>b</sup>	Pyridine (10)	PhMe	39	>19:1 dr	50
13	Pyridine (20)	PhMe	65	>19:1 dr	50
14	Pyridine (25)	PhMe	76	>19:1 dr	50
15 <sup>b</sup>	Pyridine (50)	PhMe	34	>19:1 dr	50
16	Imidazole (20)	PhMe	65	>19:1 dr	44
17	DABCO (20)	PhMe	49	>19:1 dr	30
18	2,6-Lutidine (20)	PhMe	71	>19:1 dr	50
19 <sup>b</sup>	N,N-dimethylaniline (20)	PhMe	64	>19:1 dr	49
20 <sup>b</sup>	CuCl (20)	PhMe	13	>19:1 dr	7
21 <sup>b</sup>	H <sub>2</sub> O (100)	PhMe	0	-	-

<sup>a</sup> Yields are those of chromatographically purified compounds. The ee values were determined by HPLC analysis. <sup>b</sup> Reaction run for 24 h.

# Evaluation of other alkoxyisocoumarins. <sup>a</sup>

0.2 mmol	+ 0 RO 3 (1.2 equiv)	<b>2b</b> (20 mol%) PhMe (0.05 M), rt, 24 h			
entry	R	yield (%)	dr	ee (%)	
1	Me	50	>19:1	52	
2	Et	47	>19:1	65	
3	<i>n</i> -Pr	24	>19:1	53	
4	<i>i</i> -Pr	7	>19:1	51	
5	<i>n</i> -Bu	39	>19:1	57	
6	Bn	0	-	-	
7	CH <sub>2</sub> CH <sub>2</sub> Cl	67	>19:1	52	
8	CH <sub>2</sub> CF <sub>3</sub>	94	>19:1	18	
9	CH <sub>2</sub> CH <sub>2</sub> OCH <sub>3</sub>	0	-	-	

<sup>a</sup> Yields are those of chromatographically purified compounds. The ee values were determined by HPLC analysis.



### Evaluation of basic additives on reaction conversion.

#### **III.** Synthesis of starting materials.

#### 3-ethoxy-1H-isochromen-1-one (EIC) (3b)



To a solution of 2-(2-ethoxy-2-oxoethyl)benzoic acid<sup>12</sup> (200 mg, 1.0 mmol, 1 equiv) in DCM (3.3 mL, 0.25 M) at 0 °C was added trifluoroacetic anhydride (0.16 mL, 1.15 mmol, 1.2 equiv) in 0.5 mL DCM dropwise. The reaction mixture was stirred at 0 °C for 1 h then concentrated. The crude residue was dissolved in ethyl acetate and washed with water, sat. sodium bicarbonate, brine then dried over sodium sulfate, concentrated to afford **3b** in 83% yield;  $R_f = 0.71$  (EtOAc/hexanes 3:7 v/v); mp = 63–65 °C; IR (KBr) 3053, 2987, 2685, 2306, 1741, 1641, 1564, 1484, 1421, 1363, 1312, 1261, 1218, 1186, 1037, 1008, 896, 873, 750 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)  $\delta$  8.33–8.08 (m, 1H), 7.68–7.52 (m, 1H), 7.35–7.16 (comp, 2H), 5.56 (s, 1H), 4.20 (q, *J* = 7.1 Hz, 2H), 1.46 (t, *J* = 7.0 Hz, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  161.41, 158.90, 140.21, 135.15, 129.95, 125.49, 124.67, 117.59, 80.03, 65.31, 14.46; *m/z* (ESI–MS) 191.3 [M+H]<sup>+</sup>.

### IV. Synthesis and characterization of products

#### General Procedure A for Lewis Acid Promoted Synthesis of Alkyl Ester:

An oven-dried vial was charged with alkoxyisocoumarin (0.3 mmol, 1 equiv) and imine (0.3 mmol, 1 equiv), powdered 4 A molecular sieves (50 mg) and acetonitrile (0.75 mL, 0.4 M). The reaction was charged with boron trifluoride diethyl etherate (46  $\mu$ L, 0.33 mmol, 1.1 equiv). The reaction mixture was stirred at rt or 45 °C until the imine could no longer be detected by TLC analysis. The reaction mixture was quenched with sat. sodium bicarbonate (1 mL) and the aqueous layer extracted with diethyl ether. The combined organic layers were dried over sodium sulfate and concentrated.

The crude residue was dissolved in methanol (0.5 mL). Sodium methoxide was added (360  $\mu$ L, 25% wt. in MeOH, 1.59 mmol, 5.3 equiv) and the reaction mixture stirred for 30 minutes at room temperature. Water (2 mL) was added and the reaction mixture extracted with diethyl ether. The combined organic layers were washed with saturated sodium chloride, dried over sodium sulfate, concentrated and purified by flash column chromatography on silica gel. The resulting product was dried under high vacuum.

### General Procedure B for Asymmetric Cycloaddition to Form Methyl Ester:

An oven-dried vial was charged with homophthalic anhydride **1** (36 mg, 0.22 mmol, 1.1 equiv), catalyst **2a** (25 mg, 0.04 mmol, 0.2 equiv), powdered 4 Å molecular sieves (200 mg) and MTBE (8 mL, 0.025 M). The reaction mixture was cooled to -55 °C and then charged with the imine (0.2 mmol, 1 equiv). The reaction mixture was stirred at -55 °C until the imine could no longer

be detected by TLC analysis, then MeOH (1 mL) and trimethylsilyldiazomethane (0.2 mL, 2.0 M in diethyl ether, 0.4 mmol, 2 equiv) was added and the reaction allowed to warm to room temperature. After one hour the reaction was quenched with 5 drops of glacial acetic acid. The reaction mixture was filtered through celite, concentrated and purified by flash column chromatography on silica gel. The resulting product was dried under high vacuum.

#### General Procedure C for Asymmetric Cycloaddition to Form Alkyl Ester:

An oven-dried vial was charged with alkoxyisocoumarin **3a** or **3b** (0.24 mmol, 1.2 equiv), catalyst **2b** (22 mg, 0.04 mmol, 0.2 equiv) and trifluorotoluene (4 mL, 0.05 M). The reaction mixture was cooled to 0 °C-and then charged with imine (0.2 mmol, 1 equiv) and pyridine (4  $\mu$ L, 0.05 mmol, 0.25 equiv). The reaction mixture was stirred at 0 °C until the imine could no longer be detected by TLC analysis. The reaction was concentrated and purified by flash column chromatography in silica gel. The resulting product was dried under high vacuum.

(±)-(**3S,4S**)-methyl 1-oxo-2,3-diphenyl-1,2,3,4-tetrahydroisoquinoline-4-carboxylate (4a): Following the general procedure A, **4a** was obtained as a white solid in 63% yield (68 mg, >19:1 dr);  $R_f = 0.30$  (EtOAc/hexanes 3:7 v/v); mp = 150–153 °C; IR (KBr) 3026, 2947, 2871, 1749, 1655, 1619, 1478, 1464, 1402, 1349, 1241, 1133, 992 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)  $\delta$  8.27–8.24 (m, 1H), 7.47–7.42 (comp, 2H), 7.35–7.34 (comp, 2H), 7.34–7.32 (comp, 2H), 7.26–7.14 (comp, 7H), 5.65 (d, *J* = 1.5 Hz, 1H), 4.04 (d, *J* = 1.5 Hz, 1H), 3.74 (s, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  171.20, 163.45, 142.45, 139.21, 132.49, 132.30, 129.62, 129.50, 129.12, 128.50, 128.00, 127.10, 126.73, 126.49, 65.00, 53.02, 51.86; *m/z* (ESI–MS) 358.3 [M+H]<sup>+</sup>.



**2-(4-methoxyphenyl)-1-oxo-3-phenyl-1,2,3,4-tetrahydroisoquinoline-4carboxylate (4b):** Following the general procedure A, compound **4b** was obtained as a white solid in 61% yield (71 mg, >19:1 dr);  $R_f = 0.20$ (EtOAc/hexanes 3:7 v/v); mp: 192–194 °C; IR (KBr) 3467, 3005, 2970, 2949, 1736, 1654, 1426, 1365, 1217, 1092, 898, 528 cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.27–8.20 (m, 1H), 7.48–7.39 (comp, 2H), 7.29–7.15 (comp, 6H), 7.16–7.12 (comp, 2H), 6.85 (dd, J = 8.5, 5.5 Hz, 2H), 5.58 (d,

J = 4.2 Hz, 1H), 3.99 (d, J = 4.3 Hz, 1H), 3.76 (s, 3H), 3.74 (s, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  171.31, 163.63, 158.44, 139.30, 135.33, 132.42, 132.33, 129.69, 129.56, 128.86, 128.69, 128.54, 128.06, 126.60, 114.44, 65.27, 55.52, 53.06, 51.84; *m/z* (ESI–MS) 387.4 [M+H]<sup>+</sup>;

(±)-(3S,4S)-methyl Clock (±)-(3S,4S)-methy

7.25–7.19 (comp, 4H), 7.13 (dd, J = 7.5, 2.0 Hz, 2H), 5.67–5.51 (m, 1H), 4.04 (d, J = 1.4 Hz, 1H), 3.73 (s, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  171.11, 163.40, 140.93, 138.82, 132.69,

132.63, 132.25, 129.63, 129.25, 128.95, 128.74, 128.49, 128.17, 128.13, 126.39, 64.90, 53.05, 51.63; m/z (ESI–MS) 392.2 [M+H]<sup>+</sup>.

(±)-(3S,4S)-methyl 1-oxo-3-phenyl-2-(p-tolyl)-1,2,3,4-tetrahydroisoquinoline-4-carboxylate



(4d): Following the general procedure A, 4d was obtained as a white solid in 72% yield (81 mg, >19:1 dr);  $R_f = 0.49$  (EtOAc/hexanes 3:7 v/v); mp = 169–172 °C; IR (KBr) 3042, 2947, 1736, 1701, 1432, 1325, 1239, 1183, 1002, 855, 723 617, 457 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz) δ 8.27–8.23 (m, 1H), 7.46–7.42 (comp, 2H), 7.21 (t, J = 8.2 Hz, 6H), 7.18–7.12 (comp, 4H), 5.62 (d, J = 1.2 Hz, 1H), 4.02 (d, J = 1.4 Hz, 1H), 3.74 (s, 3H), 2.32 (s, 3H);

<sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 171.23, 163.50, 139.85, 139.27, 136.90, 132.39, 132.29, 129.77, 129.69, 129.48, 128.81, 128.64, 128.49, 127.96, 126.55, 126.53, 65.08, 53.01, 51.86, 21.17; m/z (ESI-MS) 372.1  $[M+H]^+$ .

(±)-(3S,4S)-methyl 1-oxo-2-phenyl-3-(p-tolyl)-1,2,3,4-tetrahydroisoquinoline-4-carboxylate



(4e): Following the general procedure A, 4e was obtained as a white solid in 73% yield (82 mg, >19:1 dr);  $R_f = 0.50$  (EtOAc/hexanes 3:7 v/v); mp = 144-146 °C; IR (KBr) 3247, 3235, 3029, 2953, 2613, 2534, 1925, 1641, 1495, 1343, 1121, 936, 529 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz) δ 8.36 – 8.18 (m, 1H), 7.53-7.41 (comp, 2H), 7.37-7.30 (comp, 4H), 7.26-7.22 (m, 1H), 7.20–7.17 (m, 1H), 7.08-7.01 (comp, 4H), 5.60 (d, J = 1.4 Hz, 1H),

4.00 (d, J = 1.5 Hz, 1H), 3.74 (s, 3H), 2.25 (s, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  171.31, 163.51, 142.54, 137.78, 136.23, 132.47, 132.43, 129.72, 129.56, 129.52, 129.13, 128.67, 128.53, 127.10, 126.74, 126.43, 64.86, 53.04, 52.01, 21.10; *m/z* (ESI-MS) 372.2 [M+H]<sup>+</sup>.



3-(4-chlorophenyl)-1-oxo-2-phenyl-1,2,3,4-tetrahydroisoquinoline-4carboxylate (4f): Following the general procedure A, 4f was obtained as a white solid in 68% yield (80 mg, >19:1 dr);  $R_f = 0.45$  (EtOAc/hexanes 3:7 v/v); mp = 97–99 °C; IR (KBr) 3457, 3006, 2969, 1736, 1427, 1366, 1216, 899, 528 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)  $\delta$  8.24 (dd, J = 5.6, 3.6 Hz, 1H), 7.46 (dd, J = 5.7, 3.3 Hz, 2H), 7.37–7.29 (comp, 4H), 7.27–7.22 (m, 1H), 7.19 (d, J = 8.6 Hz, 2H), 7.09 (d, J = 8.4 Hz, 2H), 5.63 (d, J = 1.2 Hz, 1H), 3.98 (d, J = 1.5

Hz, 1H), 3.73 (s, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 170.93, 163.30, 142.21, 137.78, 133.93, 132.66, 132.03, 129.53, 129.42, 129.21, 129.08, 128.84, 128.57, 127.90, 127.28, 126.70, 64.45, 53.09, 51.69; m/z (ESI–MS) 392.0 [M+H]<sup>+</sup>.

 $(\pm)$ -(3S,4S)-methyl



3-(4-methoxyphenyl)-1-oxo-2-phenyl-1,2,3,4-tetrahydroisoquinoline-4carboxylate (4g): Following the general procedure A, 4g was obtained as a colorless oil in 45% yield (49 mg, >19:1);  $R_f = 0.43$  (EtOAc/hexanes 3:7 v/v); mp = 112–115 °C; IR (KBr) 3458, 3004, 2973, 1716, 1651, 1366, 1218, 1092, 901, 529 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz) δ 8.27– 8.21 (m, 1H), 7.49–7.43 (comp, 2H), 7.37–7.29 (comp, 4H), 7.26–7.21 (m, 1H), 7.21–7.18 (m, 1H), 7.05 (d, J = 8.8 Hz, 2H), 6.74 (d, J = 8.8 Hz,

2H), 5.57 (d, J = 1.5 Hz, 1H), 3.99 (d, J = 1.6 Hz, 1H), 3.74 (s, 3H), 3.72 (s, 3H); <sup>13</sup>C NMR (125) MHz, CDCl<sub>3</sub>) δ 171.30, 163.42, 159.28, 142.50, 132.50, 131.18, 129.69, 129.55, 129.14, 128.68, 128.54, 127.72, 127.13, 126.80, 114.22, 64.59, 55.30, 53.02, 52.00; *m/z* (ESI-MS) 387.9  $[M+H]^{+}$ .

(±)-(3S,4S)-methyl 1-oxo-2-phenyl-3-(pyridin-3-yl)-1,2,3,4-tetrahydroisoquinoline-4carboxylate (4h): Following the general procedure A, 4h was obtained as a colorless oil in 95% yield (101 mg, >19:1 dr);  $R_f = 0.33$  (EtOAc/hexanes 3:7 v/v); IR (KBr) 3458, 3004, 2946, 1739, 1435, 1365, 1227, 1217, 528 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)  $\delta$  8.50 (ddd, J = 4.8, 1.7, 0.9 Hz, 1H), 8.21–8.17 (m, ČO<sub>2</sub>Me 1H), 7.53 (td, J = 7.8, 1.8 Hz, 1H), 7.47 (dd, J = 8.6, 1.2 Hz, 1H), 7.39 (td, J = 5.8, 2.9 Hz, 1H), 7.35 (d, J = 8.4 Hz, 2H), 7.31–7.27 (comp, 2H), 7.25–7.22 (m, 1H), 7.22–7.19 (m, 1H), 7.08 (ddd, J = 7.6, 4.8, 1.0 Hz, 1H), 5.79 (d, J = 1.5 Hz, 1H), 4.53 (d, J = 1.6 Hz, 1H), 3.73 (s, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 171.17, 163.58, 158.43, 149.76, 142.36, 136.95, 132.64, 132.45, 129.49, 129.44, 129.09, 128.53, 126.90, 126.25, 122.59, 120.79, 66.14, 52.94, 49.84; m/z (ESI–MS) 359.5 [M+H]<sup>+</sup>.

(±)-(3S,4S)-methyl 3-(furan-3-yl)-1-oxo-2-phenyl-1,2,3,4-tetrahydroisoquinoline-4carboxylate (4i): Following the general procedure A, 4i was obtained as a yellow solid in 71% yield (74 mg, >19:1 dr);  $R_f = 0.47$  (EtOAc/hexanes 3:7 v/v); mp = 65–67 °C; IR (KBr) 3072, 3039, 1725, 1631, 1602, 1495, 1388, 1284, 1228, 1174, 1091, 1012 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz) δ 8.20 (dd, J ČO₂Me = 7.6, 1.5 Hz, 1H), 7.52 (td, J = 7.4, 1.6 Hz, 1H), 7.47 (td, J = 7.5, 1.4 Hz, 1H), 7.41–7.34 (comp. 4H), 7.32 (dd, J = 7.5, 1.2 Hz, 1H), 7.32 – 7.25 (m, 1H), 7.22 (t, J = 1.7 Hz, 1H), 7.17–7.12 (m, 1H), 5.94 (d, J = 2.8 Hz, 1H), 5.55 (d, J = 1.5 Hz, 1H), 4.00 (d, J = 1.6 Hz, 1H), 3.73 (s, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 170.88, 162.86, 143.59, 142.05, 140.24, 133.15, 132.53, 129.55, 129.39, 129.17, 128.72, 128.67, 127.26, 127.02, 124.50, 108.97, 57.84, 53.03, 50.52; m/z (ESI–MS) 348.1 [M+H]<sup>+</sup>.

(±)-(3S,4S)-methyl 3-(furan-2-yl)-1-oxo-2-phenyl-1,2,3,4-tetrahydroisoquinoline-4carboxylate (4j): Following the general procedure A, 4j was obtained as a brown oil in 77% yield (80 mg, >19:1 dr);  $R_f = 0.51$  (EtOAc/hexanes 3:7 v/v); IR (KBr) 3493, 3214, 2935, 1749, 16862 1611, 1430, 1371, 1397, 1022, 748, 530 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)  $\delta$  8.19 (dd, J = 7.6, 1.5 Hz, 1H), 7.49 ČO₂Me (td, J = 7.5, 1.5 Hz, 1H), 7.47-7.36 (comp, 5H), 7.32 (dd, J = 7.5, 1.3 Hz, 1H),7.29 (dt, J = 6.5, 1.3 Hz, 2H), 7.23 (dd, J = 1.8, 0.8 Hz, 1H), 6.18 (dd, J = 3.3, 1.8 Hz, 1H), 6.07

(d, J = 3.3 Hz, 1H), 5.68 (d, J = 1.4 Hz, 1H), 4.24 (d, J = 1.7 Hz, 1H), 3.74 (s, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 170.61, 163.10, 151.96, 142.44, 142.07, 132.97, 132.42, 129.35, 129.18, 129.14, 128.65, 128.58, 127.24, 126.81, 59.57, 53.05, 48.68; *m/z* (ESI–MS) 348.3 [M+H]<sup>+</sup>.

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(±)-(3S,4S)-methyl 1-oxo-2-phenyl-3-(thiophen-2-yl)-1,2,3,4-tetrahydroisoguinoline-4carboxylate (4k): Following the general procedure A, 4k was obtained as a yellow solid in 66% yield (72 mg, >19:1 dr);  $R_f = 0.53$  (EtOAc/hexanes 3:7 v/v); mp = 61–62 °C; IR (KBr) 3426, 3107, 3020, 2733, 2458, 2337, 1721, 1658, 1510, 1436, 1363, 1194, 702, 529 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz) δ ČO₂Me 8.22 (dd, J = 7.5, 1.7 Hz, 1H), 7.54 (td, J = 7.4, 1.7 Hz, 1H), 7.50 (td, J = 7.5, 1.5 Hz, 1H), 7.40–7.31 (comp. 5H), 7.30–7.26 (m, 1H), 5.86 (d, J = 1.6 Hz, 1H), 4.13 (d, J = 1.6

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Hz, 1H), 3.75 (s, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 170.69, 162.77, 142.43, 141.92, 132.82, 132.64, 129.93, 129.44, 129.21, 128.91, 128.83, 127.42, 127.20, 126.54, 126.35, 125.44, 61.40, 53.15, 51.62; *m/z* (ESI–MS) 364.0 [M+H]<sup>+</sup>.

(±)-(**3S,4S**)-ethyl **1-oxo-2,3-diphenyl-1,2,3,4-tetrahydroisoquinoline-4-carboxylate** (**4**): Following the general procedure A, **4**I was obtained as a white solid in 69% yield (77 mg, >19:1 dr),  $R_f = 0.32$  (EtOAc/hexanes 3:7 v/v); mp = 149–151 °C; IR (KBr) 3033, 2955, 1734, 1687, 1503, 1457, 1308, 1233, 1171, 889, 707, 519 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)  $\delta$  8.25 (dd, J = 5.8, 3.3 Hz, 1H), 7.46–7.42 (comp, 2H), 7.38–7.30 (comp, 4H), 7.25–7.15 (comp, 6H), 5.76–5.56 (m, 1H), 4.36–4.09 (comp, 2H), 4.02 (d, J = 1.3 Hz, 1H), 1.21 (t, J = 7.1 Hz, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  170.69, 163.48, 142.50, 139.33, 132.48, 132.42, 129.58, 129.43, 129.06, 128.81, 128.57, 128.45, 127.96, 127.01, 126.64, 126.48, 64.96, 62.02, 52.00, 14.16; m/z (ESI–MS) 372.2 [M+H]<sup>+</sup>.

(±)-(**3S,4S**)-methyl **2-benzyl-1-oxo-3-phenyl-1,2,3,4-tetrahydroisoquinoline-4-carboxylate** (**4n**): Following the general procedure A, **4n** was obtained as a white solid in 34% yield (38 mg, >19:1 dr);  $R_f = 0.66$  (EtOAc/hexanes 3:7 v/v); mp = 77– 79 °C; IR (KBr) 3066, 2954, 1737, 1657, 1606, 1453, 1411, 1269, 1237, 1164, 985 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)  $\delta$  8.26 (dd, J = 7.7, 1.6 Hz, 1H), 7.44 (td, J = 7.6, 1.4 Hz, 1H), 7.40 (td, J = 7.5, 1.6 Hz, 1H), 7.34–7.27 (comp, 5H), 7.25–7.20 (comp, 3H), 7.11–7.06 (comp, 2H), 7.03 (dd, J = 7.4, 1.3 Hz, 1H), 5.72 (d, J = 14.6Hz, 1H), 5.12 (d, J = 1.5 Hz, 1H), 3.87 (d, J = 1.4 Hz, 1H), 3.62 (d, J = 14.6 Hz, 1H), 3.38 (s, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  171.01, 164.12, 138.57, 137.14, 132.30, 132.01, 129.43, 129.08, 129.05, 129.01, 128.68, 128.60, 128.35, 128.17, 127.72, 126.44, 60.56, 52.70, 51.65, 48.98; m/z (ESI–MS) 372.4 [M+H]<sup>+</sup>.

(13S,13aS)-methyl 8-oxo-6,8,13,13a-tetrahydro-5H-isoquinolino[3,2-a]isoquinoline-13carboxylate (5a): Following the general procedure B, compound 5a was obtained as a white solid in 93% yield (57 mg, >19:1 dr);  $R_f = 0.24$ (EtOAc/hexanes 3:7 v/v); mp: 159–160 °C;  $[\alpha]_D^{20}$  –142.2 (c 0.5, CHCl<sub>3</sub>, 65% *ee*); IR (KBr) 3493, 3302, 3248, 2914, 2364, 1734, 1648, 1463, 1366, 1518, 1004, 913, 783, 744, 530 cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.15 (app dd, J

= 7.7, 1.4 Hz, 1H), 7.49 (app dd, J = 7.5, 1.4 Hz, 1H), 7.42 (t, J = 7.3 Hz, 1H), 7.25–7.19 (comp, 2H), 7.19–7.14 (comp, 3H), 5.34 (d, J = 9.0 Hz, 1H), 4.89 (dd, J = 12.2, 4.7 Hz, 1H), 4.18 (d, J = 9.0 Hz, 1H), 3.76 (s, 3H), 3.15–3.03 (comp, 2H), 2.92–2.84 (m, 1H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 172.33, 163.81, 136.87, 135.04, 134.17, 132.35, 129.44, 128.89, 128.62, 128.39, 127.84, 126.78, 126.52, 125.61, 58.07, 52.52, 51.93, 40.99, 29.88; *m/z* (ESI–MS) 309.0 [M+2H]<sup>+</sup>; HPLC: Daicel Chiralpak AD-H, *n*-hexane/*i*-PrOH = 90/10, Flow rate = 1 mL/min, UV = 254 nm, t<sub>R</sub> = 18.1 min (major) and t<sub>R</sub> = 20.2 min (minor).

(13R,13aS)-methyl



**2,3-dimethoxy-8-oxo-6,8,13,13a-tetrahydro-5H-isoquinolino[3,2a]isoquinoline-13-carboxylate (5b):** Following the general procedure B, compound **5b** was obtained as a white solid in 85% yield (65 mg, >19:1 dr);  $R_f = 0.11$  (EtOAc/hexanes 3:7 v/v); mp: 167–169 °C;  $[\alpha]_D^{-20}$  –210.2 (c 0.5, CHCl<sub>3</sub>, 64% *ee*); IR (KBr) 3490, 3476, 3117, 3012, 2732, 2345, 2321, 1726, 1520, 1418, 1250, 1111, 744 cm<sup>-1</sup>; <sup>-1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.16 (dd, J = 7.6, 1.5 Hz, 1H), 7.48 (td, J = 7.5, 1.5 Hz, 1H), 7.43

(t, J = 7.0 Hz, 1H), 7.09 (d, J = 7.6 Hz, 1H), 6.72 (s, 1H), 6.70 (s, 1H), 5.26 (d, J = 10.1 Hz, 1H), 4.97–4.89 (m, 1H), 4.08 (d, J = 10.1 Hz, 1H), 3.88 (s, 3H), 3.82 (s, 3H), 3.79 (s, 3H), 3.06–2.93 (comp, 2H), 2.81–2.74 (m, 1H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  172.65, 163.97, 148.47, 147.56, 135.51, 132.40, 129.34, 128.96, 128.62, 128.38, 126.13, 125.53, 112.02, 109.68, 57.88, 56.16, 55.99, 53.43, 52.43, 40.73, 29.77; m/z (ESI–MS) 368.1 [M+H]<sup>+</sup>; HPLC: Daicel Chiralpak AD-H, *n*-hexane/*i*-PrOH = 90/10, Flow rate = 1 mL/min, UV = 230 nm, t<sub>R</sub> = 35.8 min (minor) and t<sub>R</sub> = 42.1 min (major).

#### (13S,13aS)-methyl



**3-chloro-8-oxo-6,8,13,13a-tetrahydro-5H-isoquinolino[3,2a]isoquinoline-13-carboxylate (5c):** Following the general procedure B, compound **5c** was obtained as a white solid in 78% yield (53 mg, >19:1 dr);  $R_f = 0.26$  (EtOAc/hexanes 3:7 v/v); mp: 80–81 °C;  $[\alpha]_D^{20}$  –64.1 (c 1.0, CHCl<sub>3</sub>, 61% *ee*); IR (KBr) 3078, 3013, 2925, 2321, 1716, 1659, 1519, 1494, 1455, 1360, 1276, 1183, 914, 873, 826, 767, 659, 627, 616, 530 cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.14 (dd, J = 7.7, 1.3 Hz, 1H), 7.49 (td, J = 7.5,

1.5 Hz, 1H), 7.43 (t, J = 7.5 Hz, 1H), 7.21 (d, J = 2.0 Hz, 1H), 7.17–7.13 (comp, 2H), 7.09 (d, J = 8.4 Hz, 1H), 5.31 (d, J = 8.9 Hz, 1H), 4.91 (d, J = 4.5 Hz, 1H), 4.15 (d, J = 8.9 Hz, 1H), 3.77 (s, 3H), 3.14–3.00 (comp, 2H), 2.89–2.80 (m, 1H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  172.16, 163.77, 138.86, 134.77, 133.76, 132.75, 132.50, 129.41, 128.95, 128.52, 128.43, 127.03, 126.90, 126.68, 57.74, 52.64, 51.76, 40.73, 29.70; m/z (ESI–MS) 343.5 [M+2H]<sup>+</sup>; HPLC: Daicel Chiralpak AS-H, *n*-hexane/*i*-PrOH = 95/05, Flow rate = 1 mL/min, UV = 230 nm, t<sub>R</sub> = 48.5 min (major).and t<sub>R</sub> = 53.9 min (minor).

#### (13S,13aS)-methyl



13a-methyl-8-oxo-6,8,13,13a-tetrahydro-5H-isoquinolino[3,2a]isoquinoline-13-carboxylate (5d): Following the general procedure B, compound 5d was obtained as a white solid in 89% yield (57 mg, >19:1 dr); R<sub>f</sub> = 0.37 (EtOAc/hexanes 3:7 v/v); mp: 113–116 °C;  $[\alpha]_D^{20}$  –205.8 (c 0.5, CHCl<sub>3</sub>, 32% *ee*); IR (KBr) 3510, 3488, 3348, 3296, 2927, 2712, 1718, 1653, 1436, 1363, 1091, 895, 773, 721, 530 cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ

8.17 (dd, J = 7.6, 1.3 Hz, 1H), 7.52–7.47 (m, 1H), 7.44 (t, J = 7.5 Hz, 1H), 7.33–7.16 (comp, 4H), 7.04 (d, J = 7.5 Hz, 1H), 5.14 (ddd, J = 12.6, 4.4, 1.8 Hz, 1H), 4.19 (s, 1H), 3.74 (s, 3H), 3.08–2.99 (m, 1H), 2.88–2.81 (comp, 2H), 1.72 (s, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  172.42, 163.09, 139.16, 136.05, 134.67, 132.43, 129.84, 128.77, 128.55, 128.16, 127.43, 126.41, 126.34, 126.00, 62.01, 57.88, 52.03, 36.77, 31.28, 21.09; m/z (ESI–MS) 322.3 [M+H]<sup>+</sup>; HPLC: Daicel Chiralpak AD-H, *n*-hexane/*i*-PrOH = 90/10, Flow rate = 1 mL/min, UV = 230 nm, t<sub>R</sub> = 8.6 min (minor) and t<sub>R</sub> = 9.9 min (major).

#### (13R,13aS)-methyl



hyl 8-oxo-6,8,13,13a-tetrahydro-5H-isoquinolino[3,2-a]isoquinoline-13carboxylate (6a): Following the general procedure C, 6a was obtained as a white solid in 72% yield (66 mg, >19:1 dr);  $R_f = 0.22$  (EtOAc/hexanes 3:7 v/v); mp = 163–166 °C;  $[\alpha]_D^{20}$ –166.1 (c 0.5, CHCl<sub>3</sub>, 74% *ee*); IR (KBr) 3501, 3319, 3254, 2919, 2364, 1727, 1644, 1458, 1371, 1525, 998, 912, 746, 531 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)  $\delta$  8.15–8.10 (m, 1H), 7.47–7.36 (comp, 2H), 7.29 (d, J

= 7.7 Hz, 1H), 7.24 – 7.10 (m, 4H), 5.20 (d, J = 4.3 Hz, 1H), 4.96 (dd, J = 11.0, 4.8 Hz, 1H), 4.17 (d, J = 4.3 Hz, 1H), 3.25 (s, 3H), 3.04–2.86 (comp, 2H), 2.79–2.68 (m, 1H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  169.45, 163.75, 136.51, 134.63, 133.29, 132.11, 129.28, 128.98, 128.82, 127.61, 127.31, 126.89, 126.23, 124.07, 56.62, 52.04, 51.30, 38.81, 28.98; m/z (ESI–MS) 307.8 [M+H]<sup>+</sup>.

(13R,13aS)-ethyl



**8-oxo-6,8,13,13a-tetrahydro-5H-isoquinolino**[**3,2-a**]isoquinoline-13carboxylate (6b): Following the general procedure C, **6b** was obtained as a white solid in 61% yield (39 mg, >19:1 dr);  $R_f = 0.23$  (EtOAc/hexanes 3:7 v/v); mp = 150–153 °C;  $[\alpha]_D^{20}$  –176.8 (c 0.5, CHCl<sub>3</sub>, 84% *ee*); IR (KBr) 2982, 2941, 2903, 2841, 2750, 1752, 1691, 1566, 1494, 1401, 1235, 1150, 1034, 981, 854, 782, 529 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)  $\delta$  8.17 (dd, *J* = 7.1, 1.8 Hz, 1H), 7.51–7.40 (comp, 2H), 7.36–7.31 (m, 1H), 7.27–7.24 (m, 1H), 7.24–7.12 (comp,

3H), 5.25 (d, J = 4.4 Hz, 1H), 5.11–4.92 (m, 1H), 4.21 (d, J = 4.4 Hz, 1H), 3.74 (q, J = 7.1 Hz, 2H), 3.11–2.90 (comp, 2H), 2.77 (d, J = 13.3 Hz, 1H), 0.80 (t, J = 7.1 Hz, 3H).; <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  169.10, 163.79, 136.56, 134.76, 133.40, 132.08, 129.29, 129.25, 128.95, 128.75, 127.53, 127.27, 126.83, 126.35, 60.98, 56.67, 51.26, 38.78, 28.99, 13.79; m/z (ESI–MS) 322.3 [M+H]<sup>+</sup>; HPLC: Daicel Chiralpak AD-H, *n*-hexane/*i*-PrOH = 90/10, Flow rate = 1 mL/min, UV = 230 nm, t<sub>R</sub> = 15.9 min (major) and t<sub>R</sub> = 27.3 min (minor).

#### (13R,13aS)-ethyl



**2,3-dimethoxy-8-oxo-6,8,13,13a-tetrahydro-5H-isoquinolino[3,2a]isoquinoline-13-carboxylate (6c):** Following the general procedure C, **6c** was obtained as a white solid in 80% yield (61 mg, >19:1 dr);  $R_f = 0.14$ (EtOAc/hexanes 3:7 v/v); mp = 164–165 °C;  $[\alpha]_D^{20}$ –316.4 (c 0.5, CHCl<sub>3</sub>, 83% *ee*); IR (KBr) 3488, 3125, 3027, 2679, 2368, 2315, 1734, 1531, 1413, 1261, 1109, 739, 528 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)  $\delta$  8.19 (dd, J =7.5, 1.3 Hz, 1H), 7.51–7.42 (comp, 2H), 7.35 (d, J = 7.3 Hz, 1H), 6.75 (s,

1H), 6.66 (s, 1H), 5.19 (d, J = 4.4 Hz, 1H), 5.08–4.94 (m, 1H), 4.20 (d, J = 4.4 Hz, 1H), 3.90 (s, 3H), 3.87 (s, 3H), 3.79 (q, J = 7.1 Hz, 2H), 3.08–2.89 (comp, 2H), 2.79–2.51 (m, 1H), 0.87 (t, J = 7.1 Hz, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  169.05, 163.77, 148.09, 134.66, 132.00, 129.31, 129.22, 128.98, 128.69, 127.42, 124.86, 111.40, 109.27, 60.91, 56.38, 56.33, 56.02, 51.16, 38.85, 28.48, 13.86; m/z (ESI–MS) 382.2 [M+H]<sup>+</sup>; HPLC: Daicel Chiralpak AD-H, *n*-hexane/*i*-PrOH = 90/10, Flow rate = 1 mL/min, UV = 230 nm, t<sub>R</sub> = 13.3 min (major) and t<sub>R</sub> = 50.9 min (minor).

#### (13R,13aS)-ethyl



**3-methoxy-8-oxo-6,8,13,13a-tetrahydro-5H-isoquinolino[3,2a]isoquinoline-13-carboxylate (6d):** Following the general procedure C, **6d** was obtained as a white solid in 64% yield (45 mg, >19:1 dr);  $R_f =$ 0.17 (EtOAc/hexanes 3:7 v/v); mp = 157–159 °C;  $[\alpha]_D^{20}$  –253.7 (c 0.5, CHCl<sub>3</sub>, 79% *ee*); IR (KBr) 3522, 3479, 3361, 3293, 2950, 2743, 1709, 1652, 1414, 1376, 1088, 901, 717, 529 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)

δ 8.19 (dd, J = 7.5, 1.5 Hz, 1H), 7.52–7.42 (comp, 2H), 7.38–7.33 (m, 1H), 7.21 (d, J = 8.6 Hz,

1H), 6.85 (dd, J = 8.6, 2.7 Hz, 1H), 6.71 (d, J = 2.6 Hz, 1H), 5.23 (d, J = 4.4 Hz, 1H), 5.01 (dd, J = 12.5, 4.2 Hz, 1H), 4.20 (d, J = 4.4 Hz, 1H), 3.85–3.74 (comp, 5H), 3.11–2.94 (comp, 2H), 2.75 (d, J = 14.8 Hz, 1H), 0.87 (t, J = 7.1 Hz, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  169.17, 163.78, 158.53, 137.91, 134.73, 132.02, 129.28, 129.21, 128.69, 127.50, 127.47, 125.38, 113.37, 113.18, 60.94, 56.24, 55.42, 51.31, 38.69, 29.25, 13.86; m/z (ESI–MS) 552.4 [M+H]<sup>+</sup>; HPLC: Daicel Chiralpak AD-H, *n*-hexane/*i*-PrOH = 90/10, Flow rate = 1 mL/min, UV = 230 nm, t<sub>R</sub> = 14.5 min (major) and t<sub>R</sub> = 25.0 min (minor).

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# V. HPLC profiles of products.







### HPLC Profile of **5b**







### HPLC Profile of **5c**







### HPLC Profile of **5d**







### HPLC Profile of 6a







### HPLC Profile of 6b







### HPLC Profile of 6c







### HPLC Profile of 6d





























































































