

Electronic Supplementary Material (ESI) for Organic & Biomolecular Chemistry. This journal is
© The Royal Society of Chemistry 2018

Electronic supplementary information (ESI)

Phenolate-induced intramolecular ring-opening cyclization of *N*-tosylaziridines: access to functionalized benzoxacycles

Runjun Devi, Jonali Das, Bipul Sarma, and Sajal Kumar Das*

Department of Chemical Sciences, Tezpur University, Napaam, Tezpur, Assam, India-784028

*E-mail: sajalkd@tezu.ernet.in

Table of contents

1.	X ray crystallography	S1-S4
2.	References.....	S4
3.	Copies of ^1H and ^{13}C NMR spectra of all new compounds.....	S5-S65

1. X-ray crystallography

X-ray crystallography: X-ray reflections were collected on a Bruker APEX-II, CCD diffractometer using Mo K α ($\lambda = 0.71073 \text{ \AA}$) radiation. Data reduction was performed using Bruker SAINT Software.¹ Intensities for absorption were corrected using SADABS. Structures are solved and refined using SHELXL-2014 with anisotropic displacement parameters for non-H atoms. Hydrogen atoms on O and N are experimentally located in all crystal structures. All C–H atoms are fixed geometrically using the HFIX command in SHELX-TL.² A check of the final CIF file using PLATON have not shown any missed symmetry.^{3,4} The crystallographic parameters for all structures are summarized in Table ESI-1. ORTEP is generated using XP-software² with 35% probability ellipsoid (Figure ESI-1) and hydrogen bond parameters are available in Table ESI-2.

Compound **12e** was crystallized (from its solution in ethyl acetate/hexane) in non-enantiogenic orthogonal space group *Pna*2₁. Crystal structure is solved and refined with two symmetry independent molecules. The ORTEP is shown in Figure ESI-1. Two 2-fold screw axis related molecules form a dimer via strong N–H···O hydrogen bonding between the amine NH of first molecule to the second molecules OH group. The dangling OH groups form O–H···O hydrogen bonds with the sulfonamide oxygen of nearby dimer extending the structure into a one dimensional molecular tapes along [001] crystallographic axis (Figure ESI-2). These tapes are connected primarily via C–H··· π interactions that completes the 3D packing of the molecules. The crystallographic data and hydrogen bond matrices are shown in Table ESI-1 and Table ESI-2.

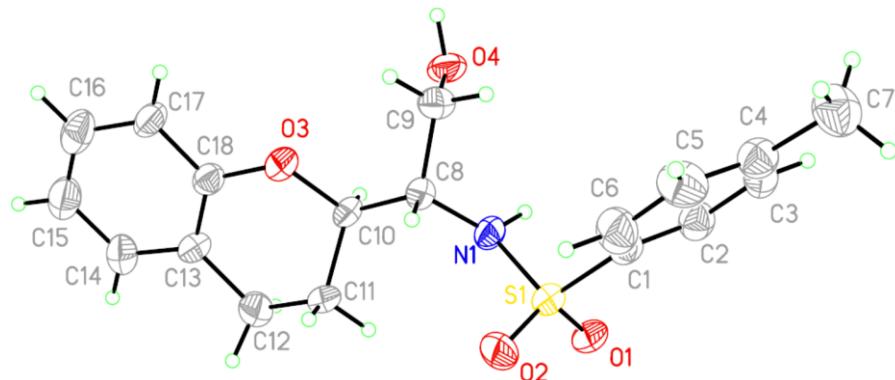


Figure ESI-1 ORTEP of chroman derivative **12e** with 35% probability ellipsoid [only one asymmetric molecule is displayed for clarity]

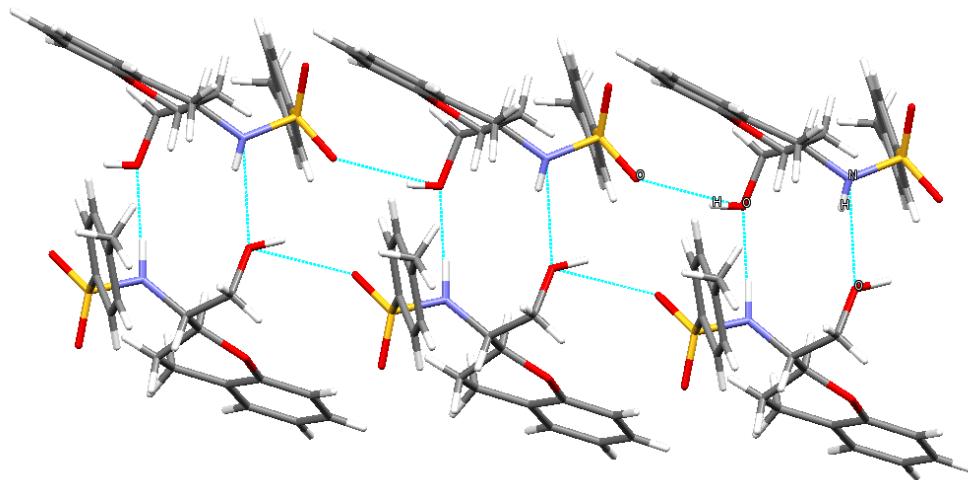


Figure ESI-2 Molecules are arranged in one dimensional tape like structure in chroman derivative **12e** via N–H···O and O–H···O hydrogen bonds along [001] crystallographic axis.

Table ESI-1: Crystallographic data of chroman derivative **12e**

Crystal Data	12e
Formula unit	C ₁₈ H ₂₁ NO ₄ S
Formula wt.	347.42
Crystal system	Orthorhombic
T [K]	100
<i>a</i> [\AA]	19.409 (11)
<i>b</i> [\AA]	22.804 (14)
<i>c</i> [\AA]	7.899 (5)
α [$^{\circ}$]	90
β [$^{\circ}$]	90
γ [$^{\circ}$]	90
Volume [\AA ³]	3496 (4)
Space group	<i>Pna</i> 2 ₁
<i>Z</i>	8
<i>D</i> _{calc} [g cm ⁻³]	1.320
μ /mm ⁻¹	0.206
Reflns. Collected	36469
Unique reflns.	7968
Observed reflns.	3221
<i>R</i> ₁ [$I > 2\sigma(I)$], <i>wR</i> ₂	0.0692; 0.1375
GOOF	0.933
Instrument	Bruker APEX-II CCD
X-ray	MoK α ; $\lambda = 0.71073$
CCDC Reference	1840365

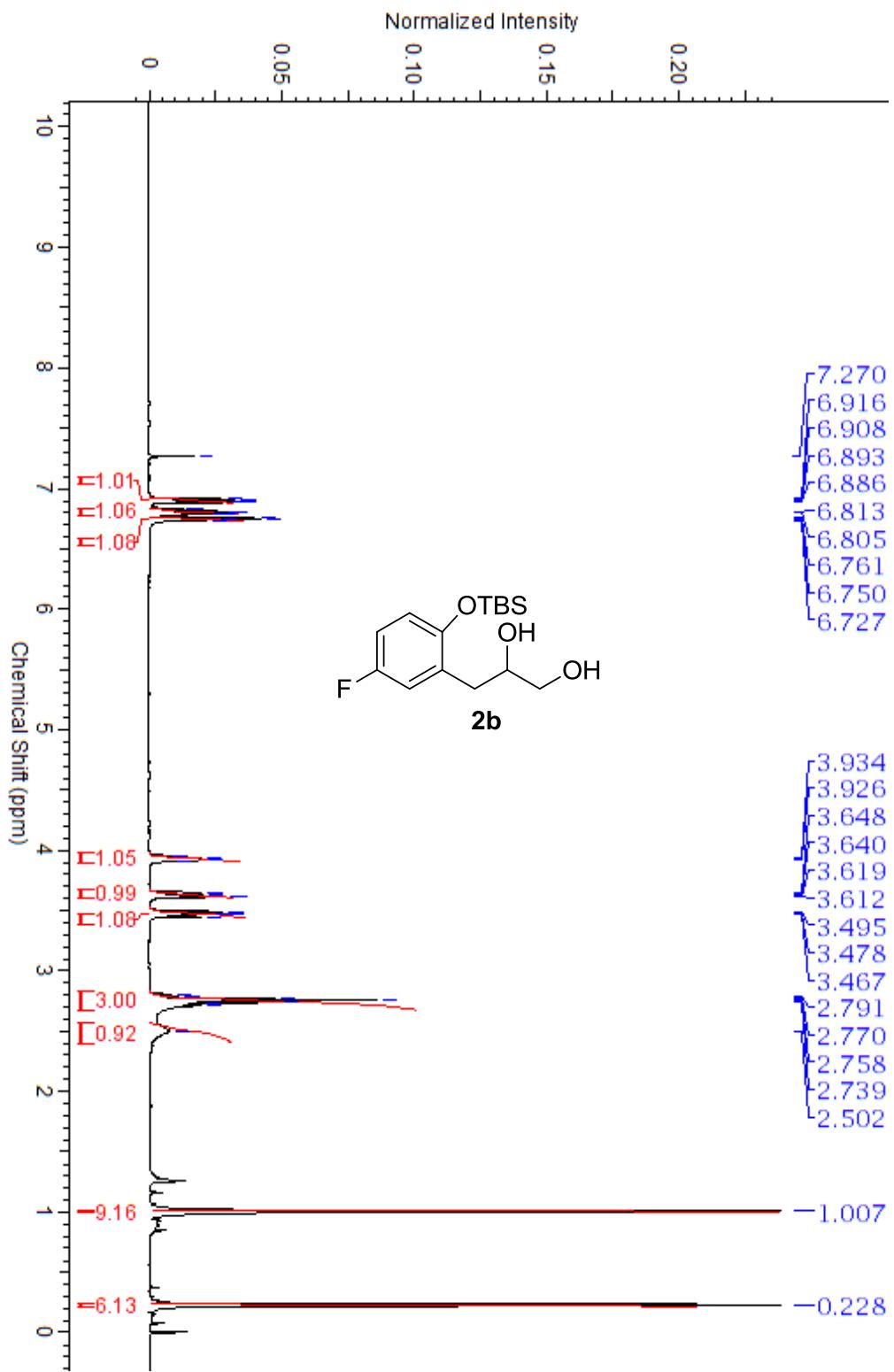
Table ESI-2: Hydrogen bond matrices of **12e**

Interaction	H···A (Å)	D···A (Å)	D–H···A (°)	Symmetry Code
N ₁ –H _{1A} ···O ₈	1.98(9)	2.957(7)	176(10)	1/2-x,-1/2+y,-1/2+z
N ₂ –H _{2A} ···O ₄	2.16(7)	2.923(7)	157(7)	1/2-x,1/2+y,1/2+z
O ₄ –H _{4A} ···O ₁	2.12(11)	2.854(6)	138(8)	x,y,-1+z
O ₄ –H _{4A} ···O ₆	2.50(10)	3.202(6)	135(8)	1/2-x,-1/2+y,-1/2+z
O ₈ –H _{8A} ···O ₆	2.14(6)	2.889(6)	161(7)	x,y,1+z
C ₃₅ –H ₃₅ ···O ₅	2.45(5)	3.369(8)	168(6)	x,y,1+z
C ₂₁ –H ₂₁ ···O ₂	2.54(5)	3.435(8)	161(6)	-
Intra C ₂ –H ₂ ···O ₁	2.60(4)	2.932(7)	102(6)	-
Intra C ₈ –H ₈ ···O ₂	2.52(6)	2.977(8)	108(4)	-
Intra C ₂₀ –H ₂₀ ···O ₅	2.58(6)	2.928(7)	103(6)	-
Intra C ₂₆ –H ₂₆ ···O ₅	2.52(6)	2.997(8)	109(4)	-

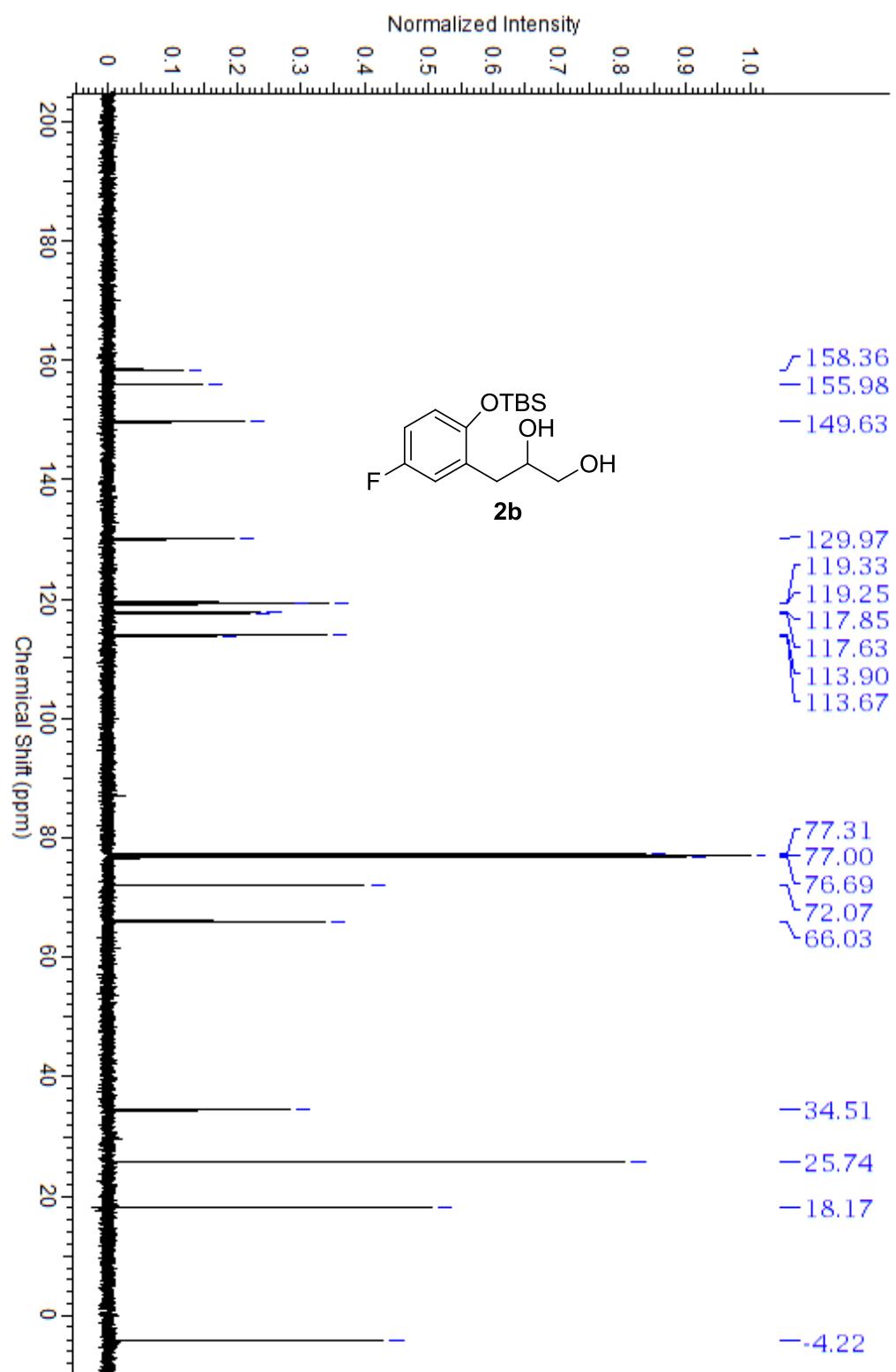
2. References

1. SAINT Plus, Bruker AXS Inc.: Madison, WI, 2008; BRUKER AXS (v 6.14).
2. Bruker AXS Inc.: Madison, WI, 2008.
3. PLATON, A Multipurpose Crystallographic Tool; A. L. Spek, Utrecht University: Utrecht, Netherland, 2002.
4. A. L. Spek, *J. Appl. Crystallogr.*, **2003**, *36*, 7–13.

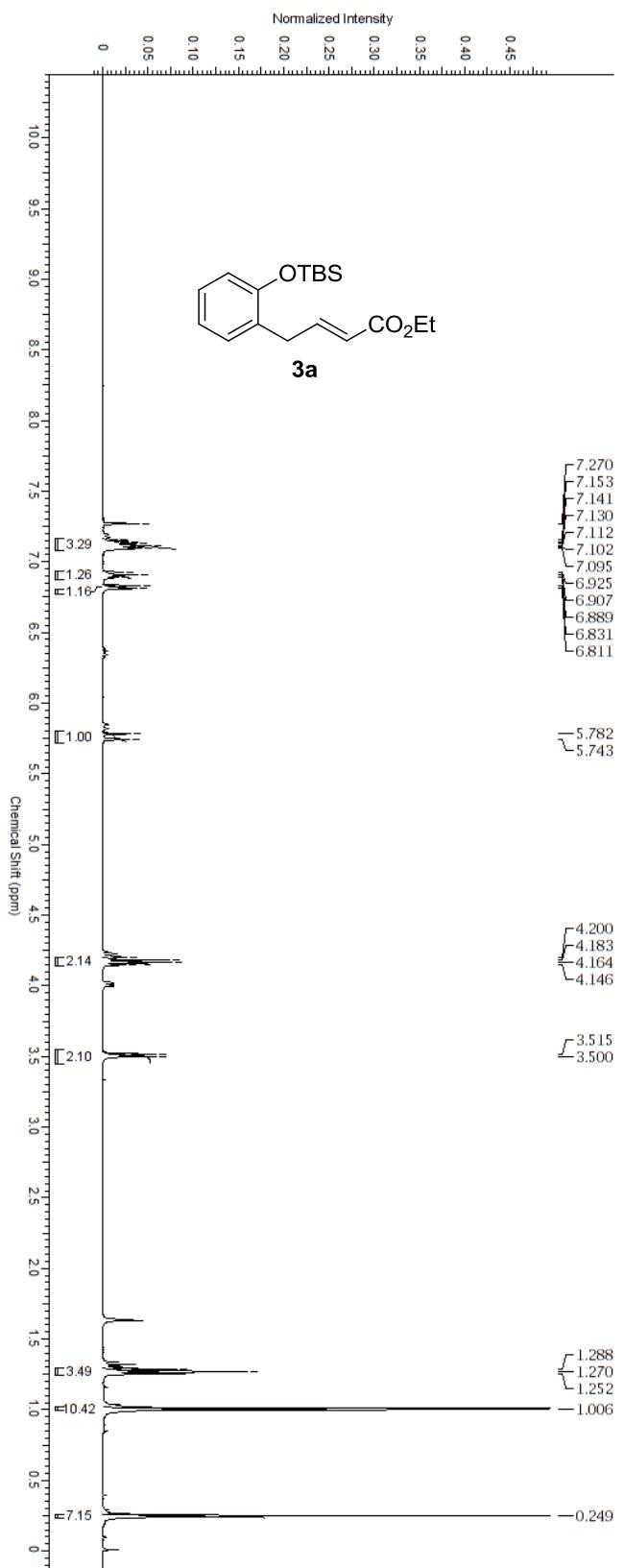
3. Copies of ¹H and ¹³C NMR spectra of all new compounds



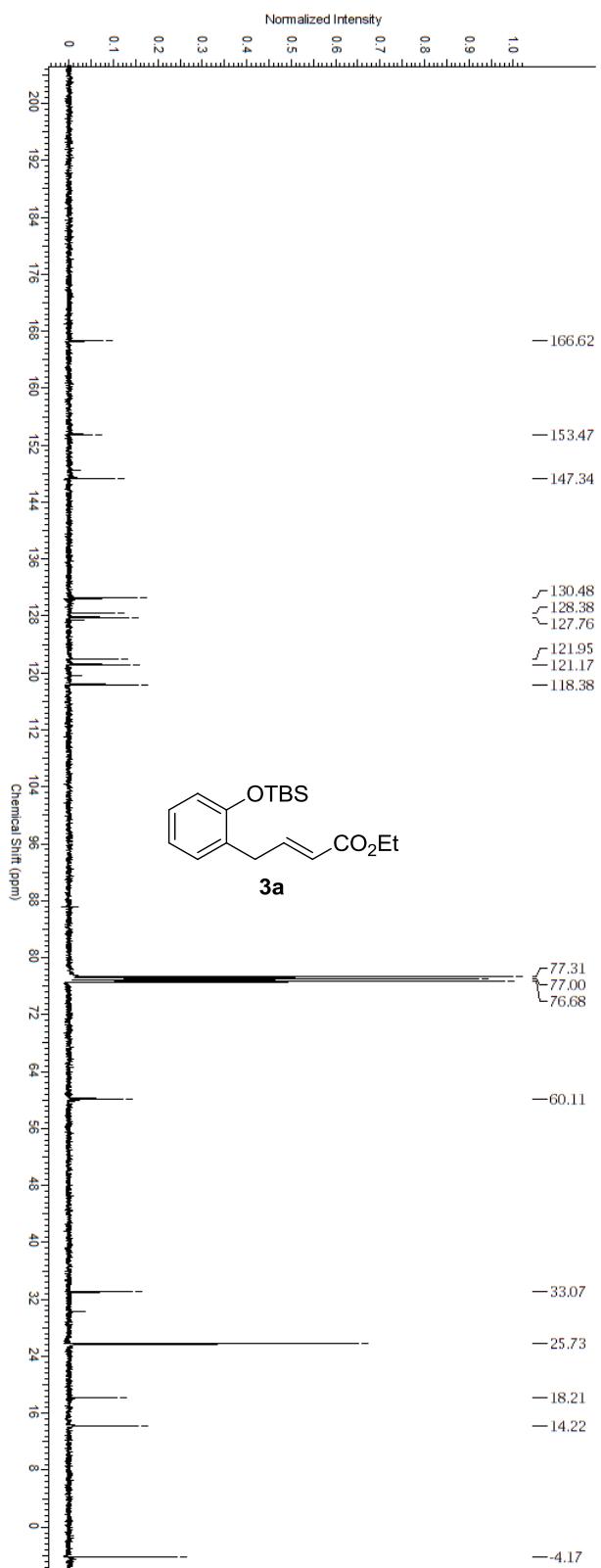
¹H NMR (400 MHz, CDCl₃) spectrum of 2b



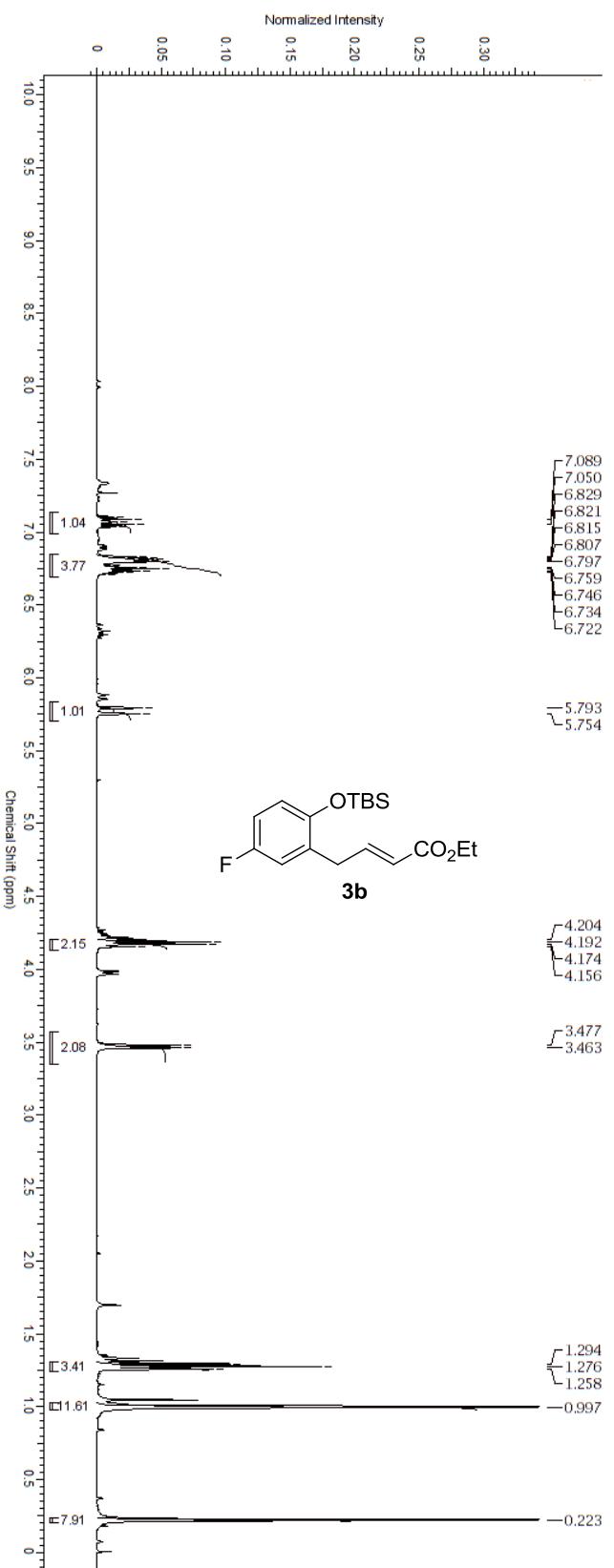
^{13}C NMR (100 MHz, CDCl_3) spectrum of 2b



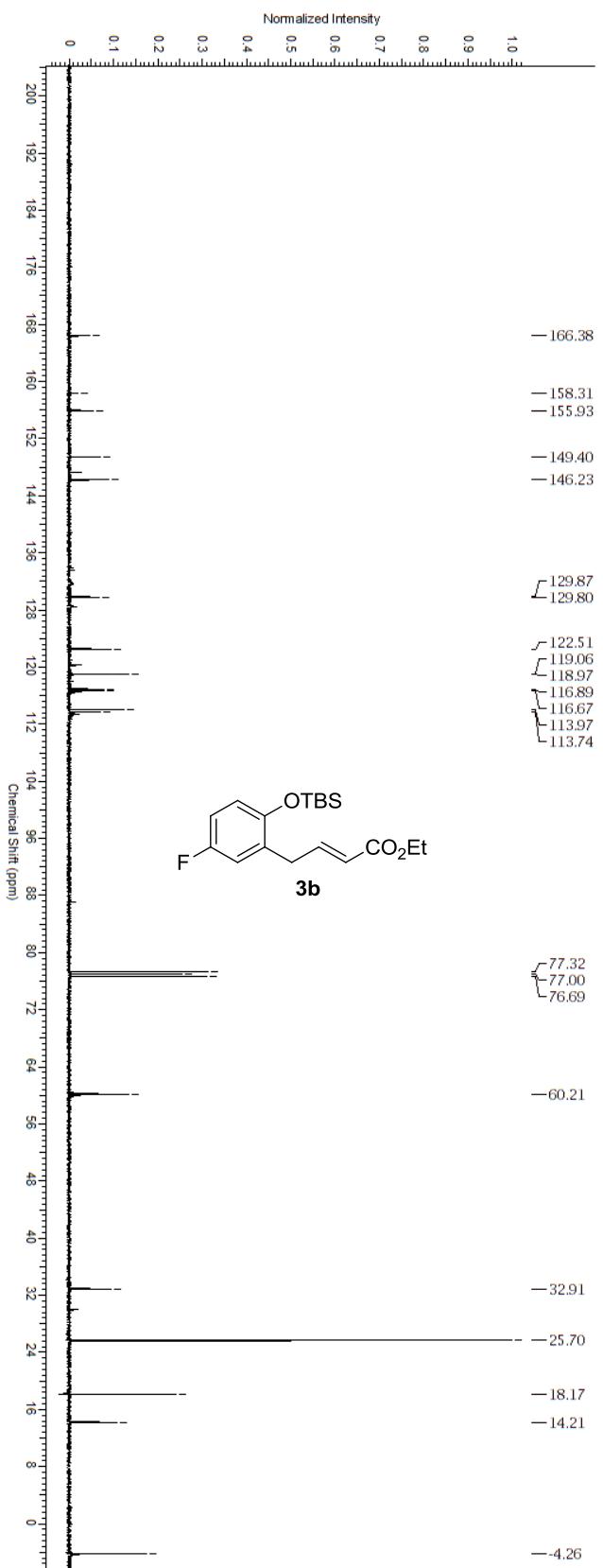
¹H NMR (400 MHz, CDCl₃) spectrum of 3a (major) and its *cis* isomer (minor); only δ values corresponding to the peaks of the *trans* isomer are reported



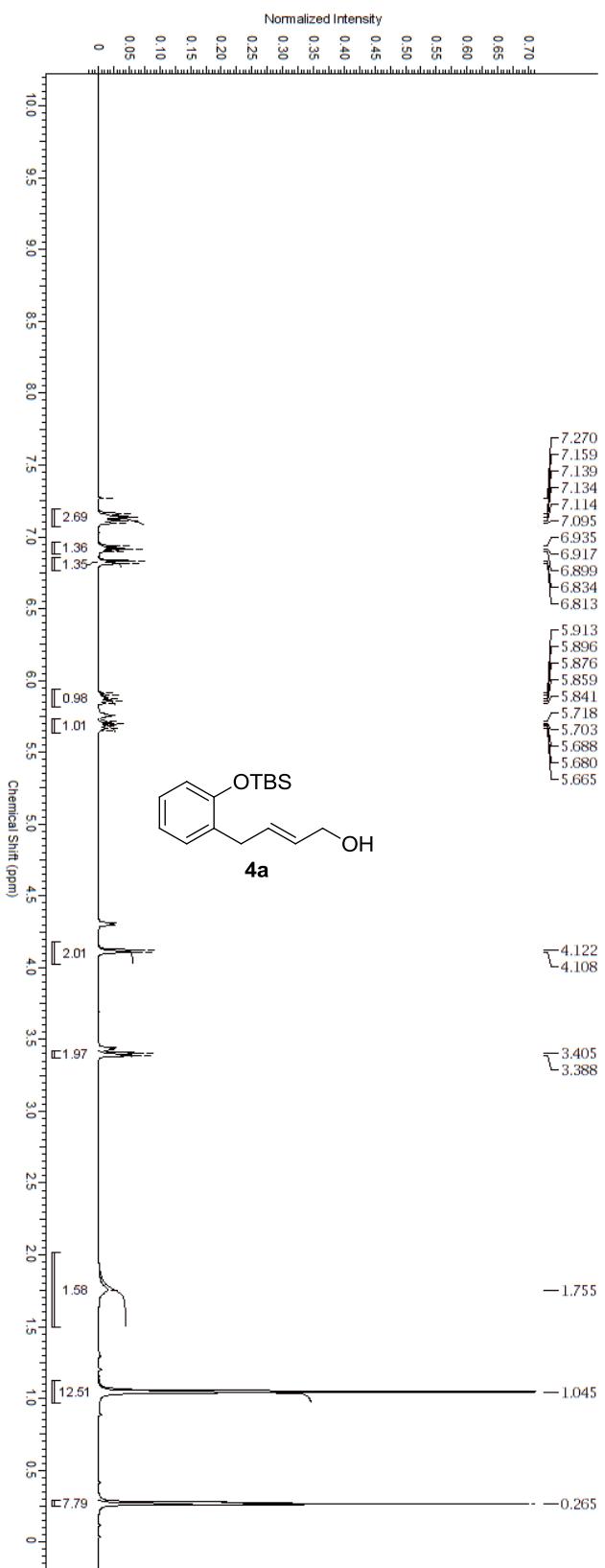
^{13}C NMR (400 MHz, CDCl_3) spectrum of 3a (major) and its *cis* isomer (minor); only δ values corresponding to the peaks of the *trans* isomer are reported



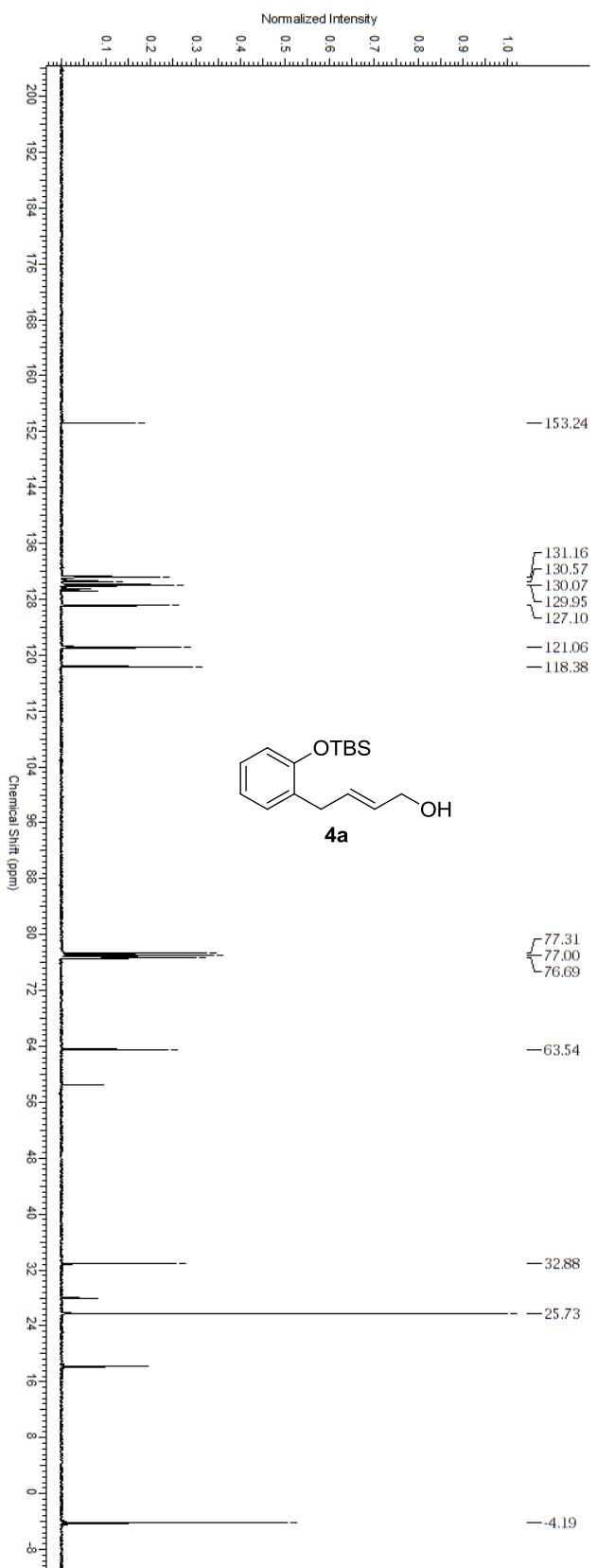
¹H NMR (400 MHz, CDCl₃) spectrum of 3b (major) and its *cis* isomer (minor); only δ values corresponding to the peaks of the *trans* isomer are reported



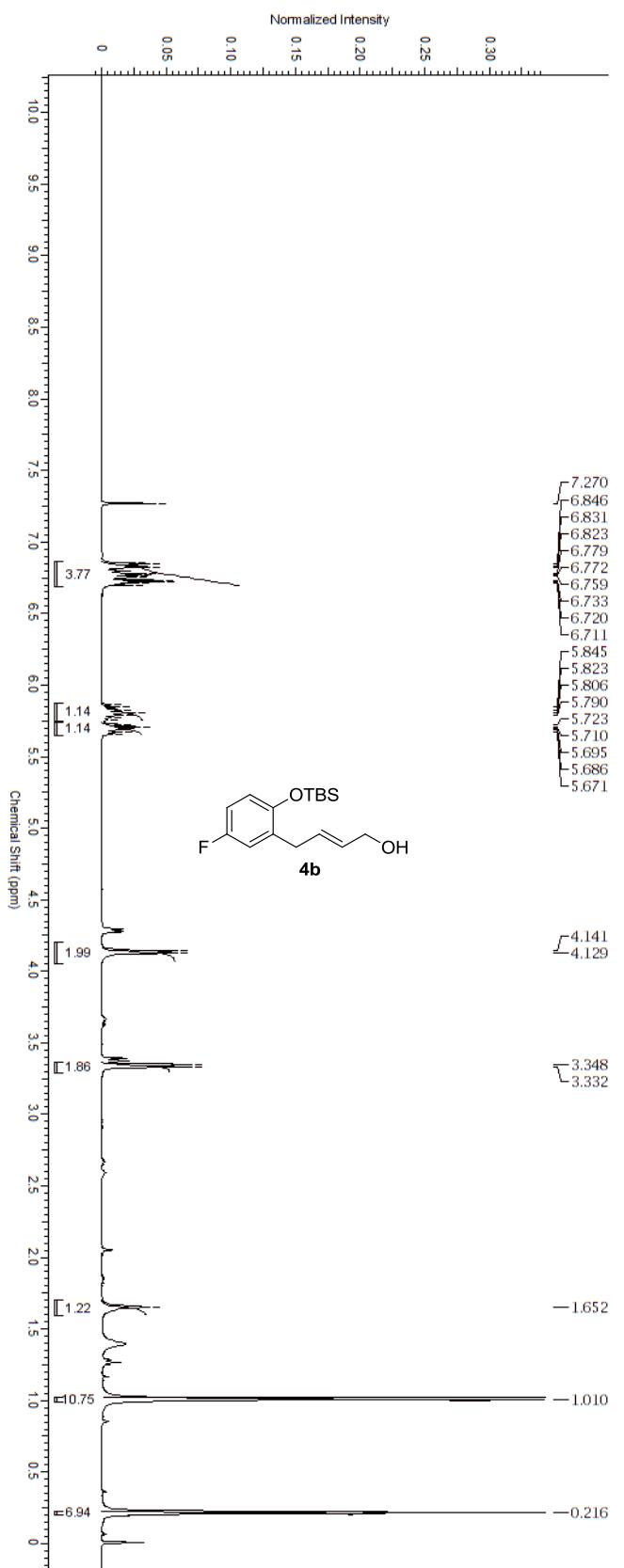
^{13}C NMR (400 MHz, CDCl_3) spectrum of 3b (major) and its *cis* isomer (minor); only δ values corresponding to the peaks of the *trans* isomer are reported



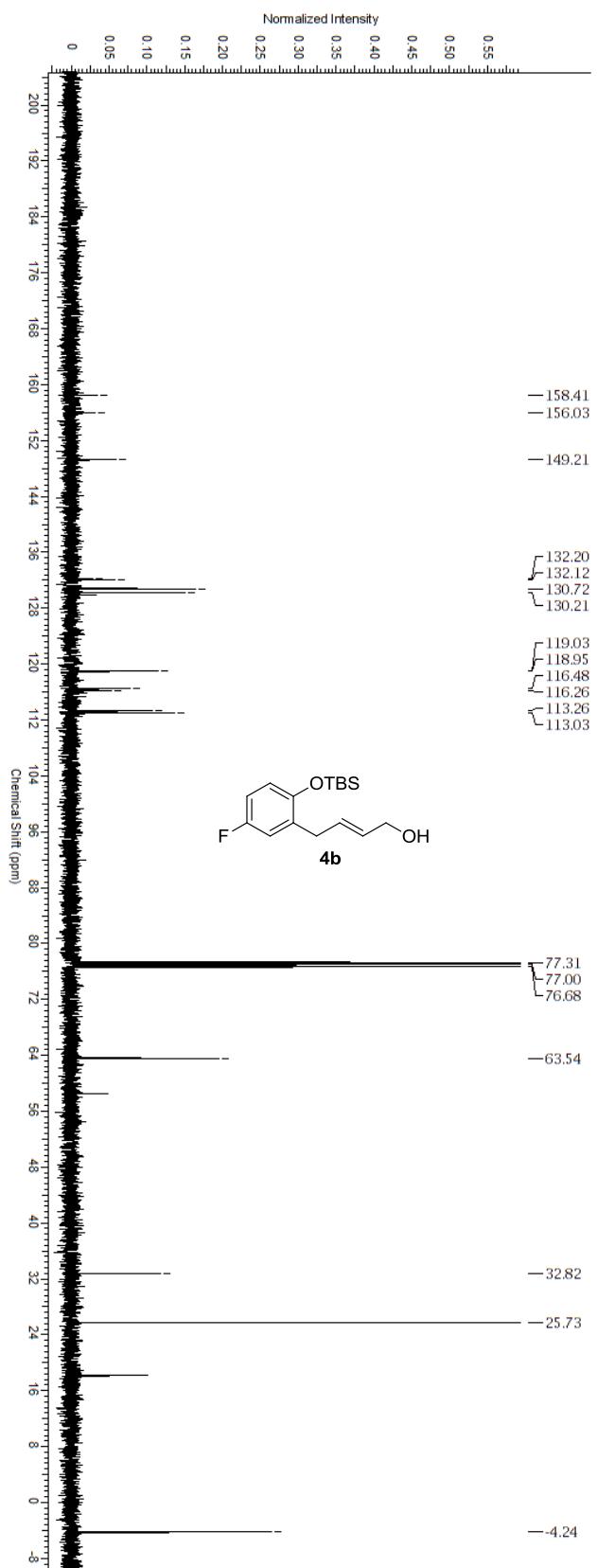
¹H NMR (400 MHz, CDCl₃) spectrum of 4a (major) and its *cis* isomer (minor); only δ values corresponding to the peaks of the *trans* isomer are reported



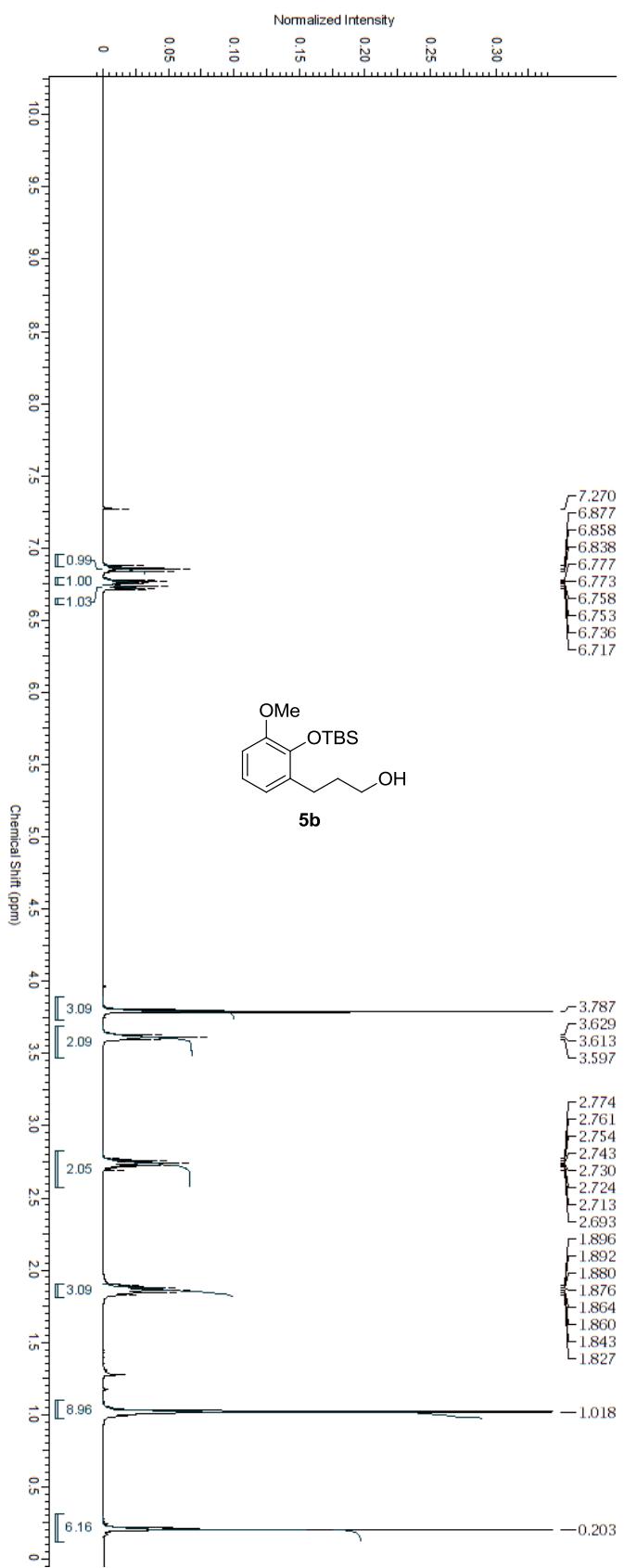
^{13}C NMR (100 MHz, CDCl_3) spectrum of **4a (major) and its *cis* isomer (minor); only δ values corresponding to the peaks of the *trans* isomer are reported**



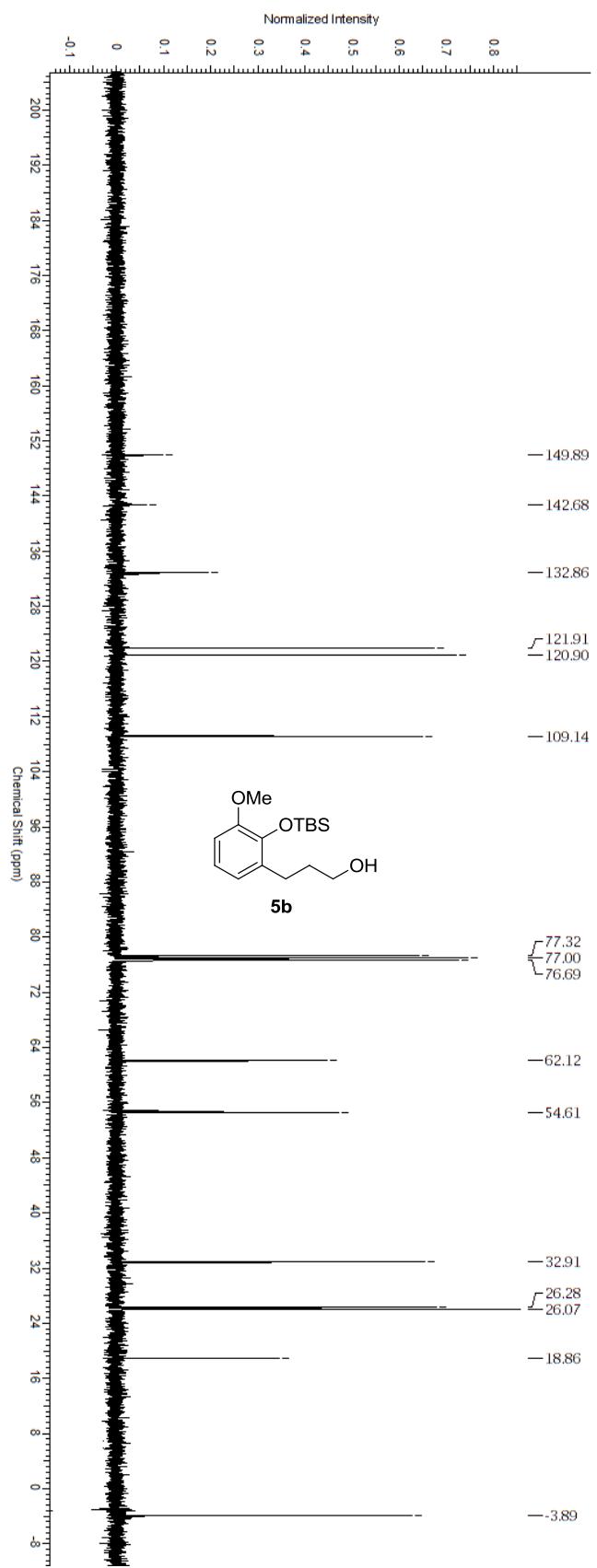
^1H NMR (400 MHz, CDCl_3) spectrum of **4b (major) and its *cis* isomer (minor); only δ values corresponding to the peaks of the *trans* isomer are reported**



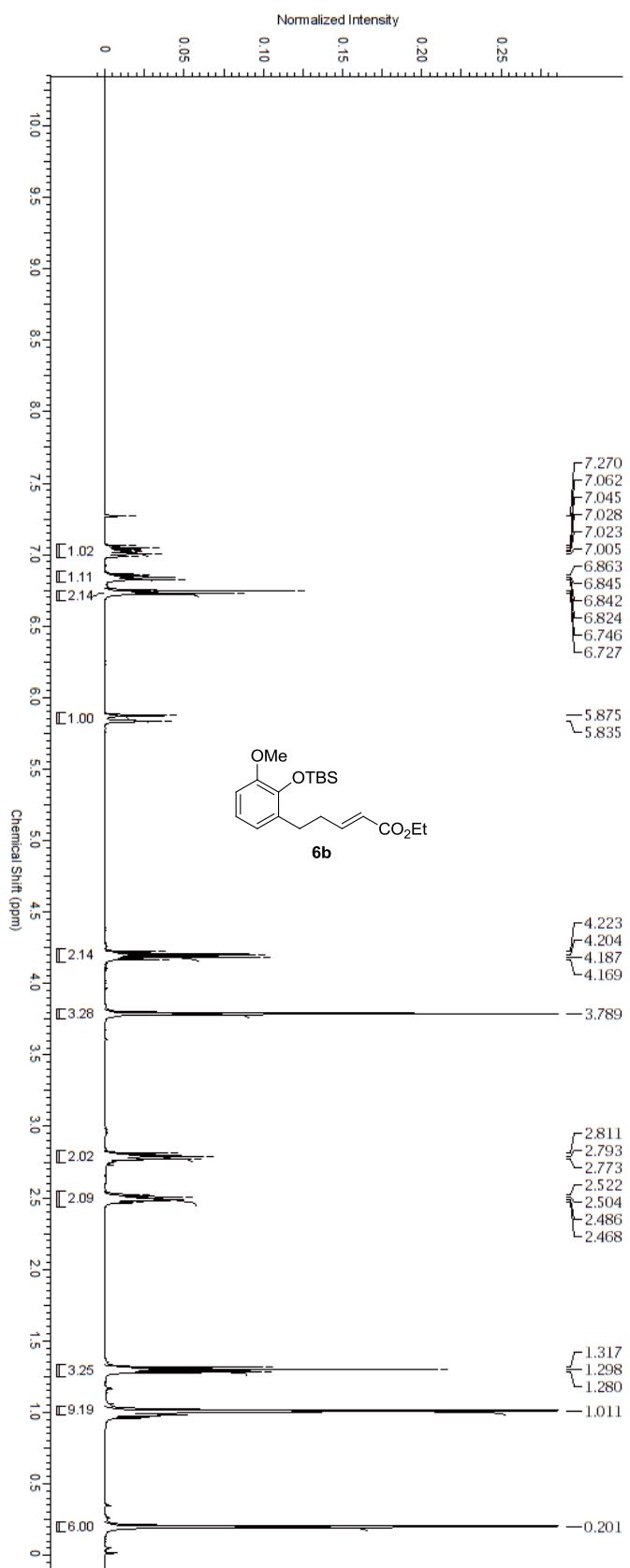
^{13}C NMR (100 MHz, CDCl_3) spectrum of **4b (major) and its *cis* isomer (minor); only δ values corresponding to the peaks of the *trans* isomer are reported**



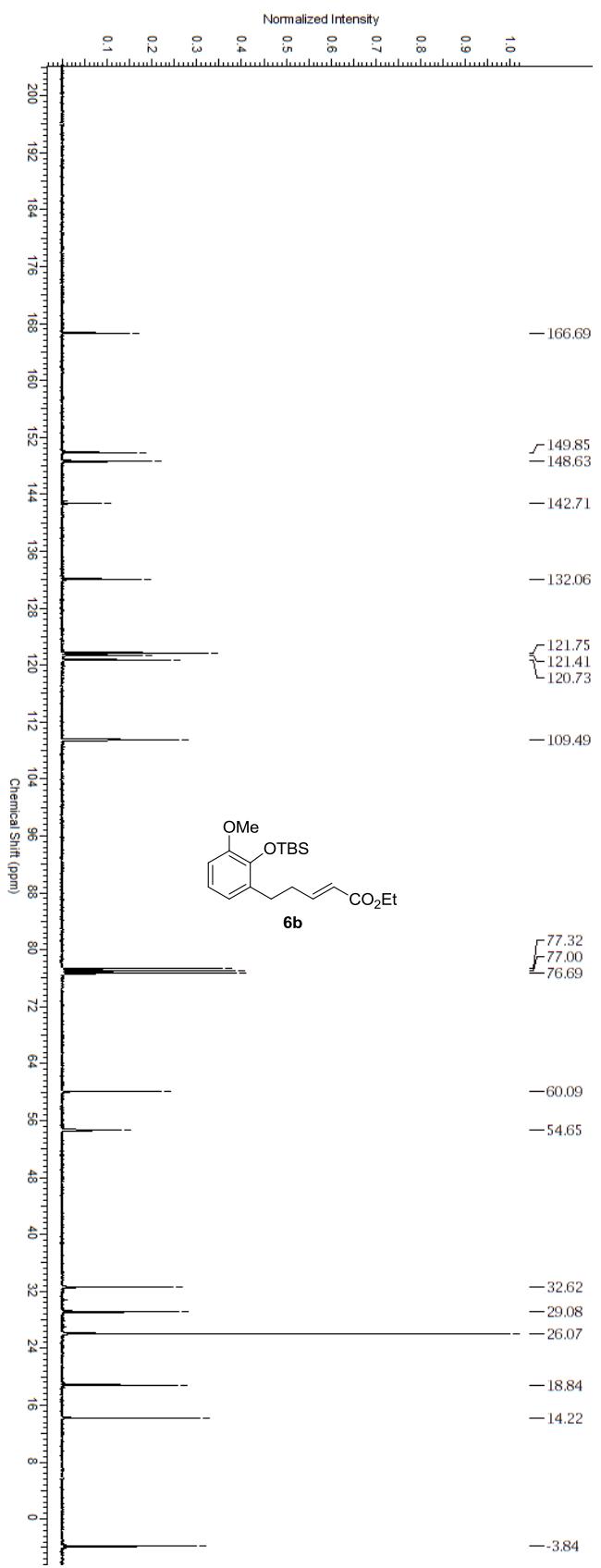
^1H NMR (400 MHz, CDCl_3) spectrum of 5b



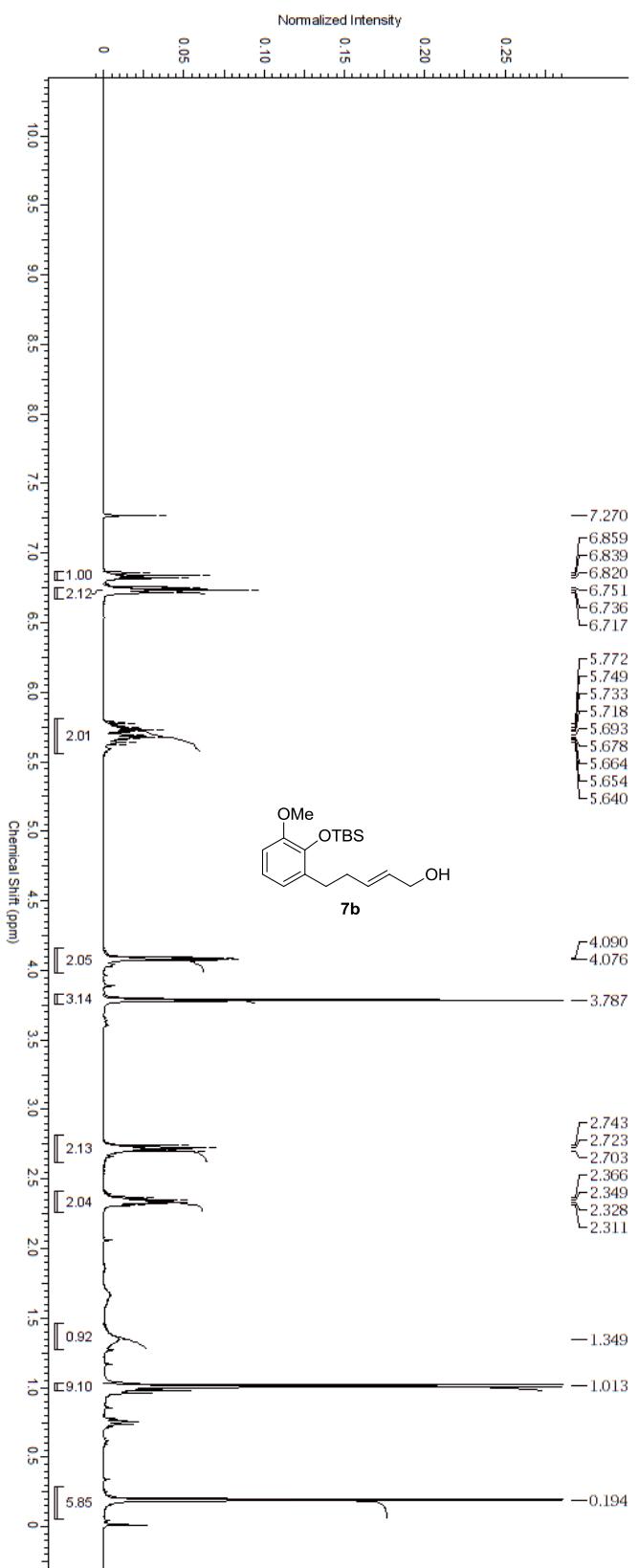
¹³C NMR (100 MHz, CDCl₃) spectrum of 5b



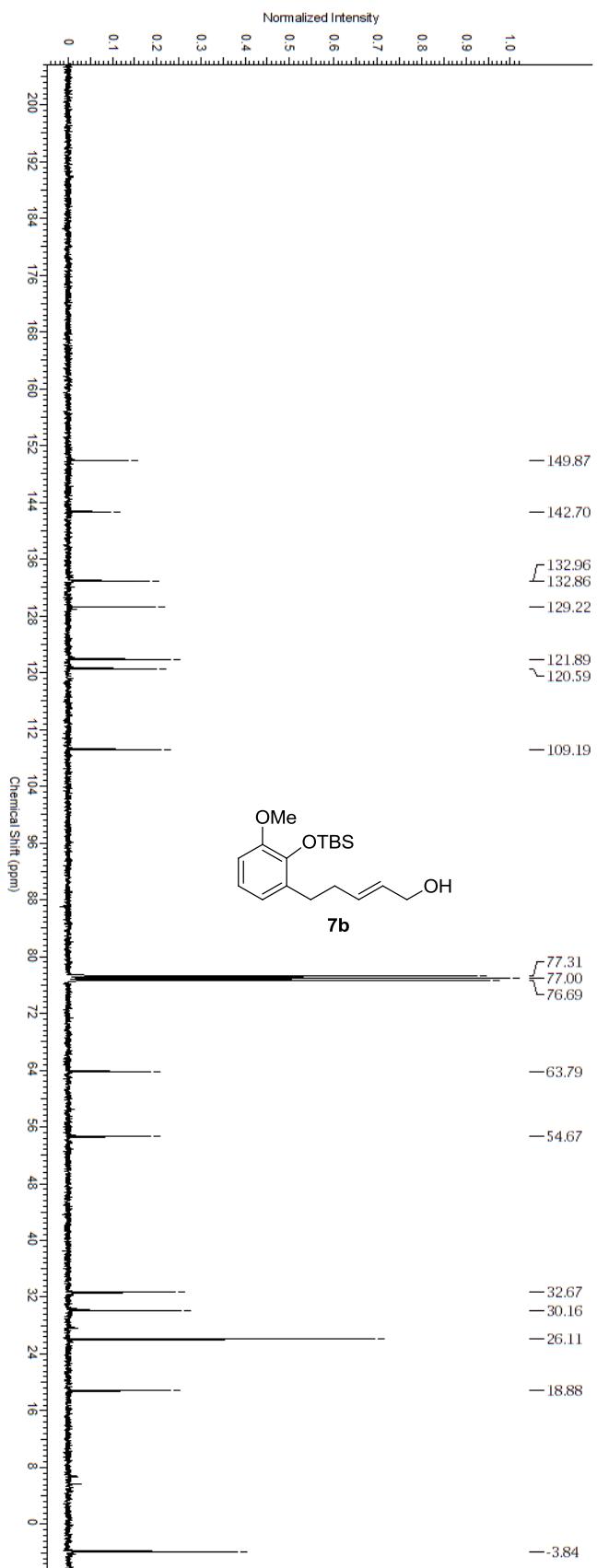
^1H NMR (400 MHz, CDCl_3) spectrum of 6b



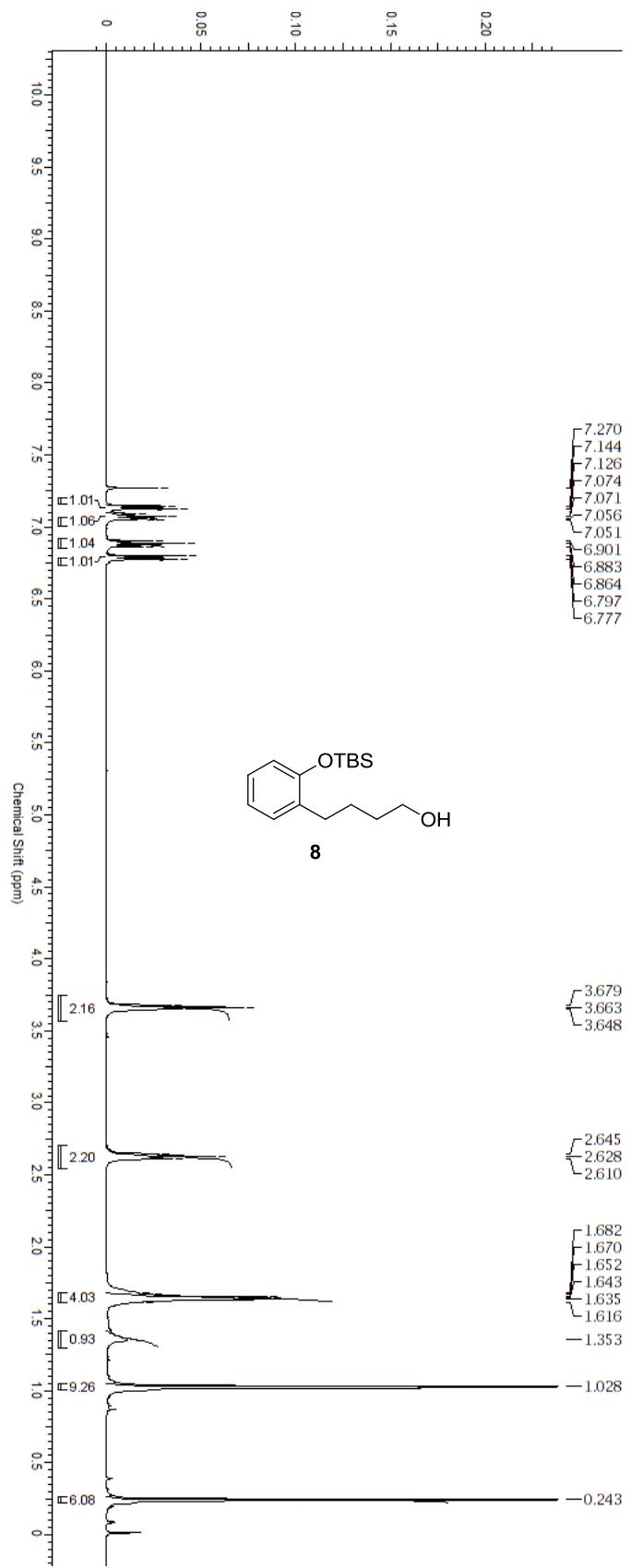
¹³C NMR (100 MHz, CDCl₃) spectrum of 6b



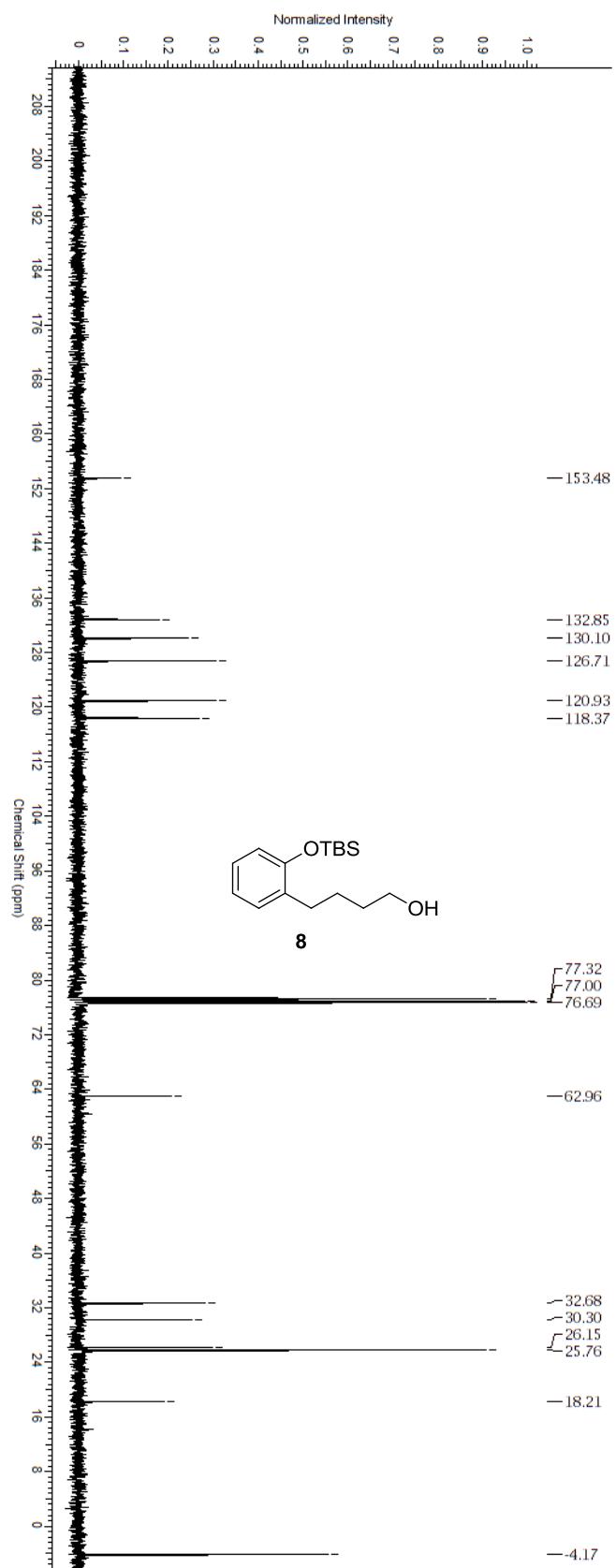
¹H NMR (400 MHz, CDCl₃) spectrum of 7b



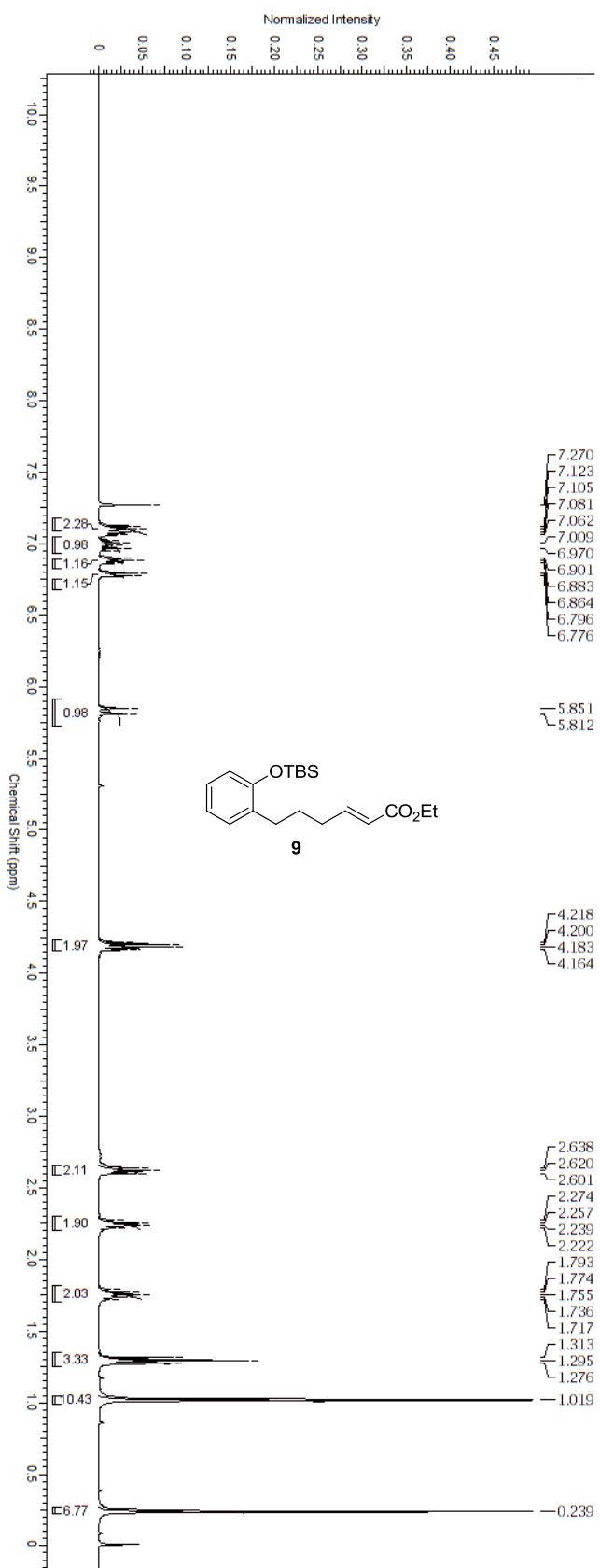
^{13}C NMR (100 MHz, CDCl_3) spectrum of **7b**



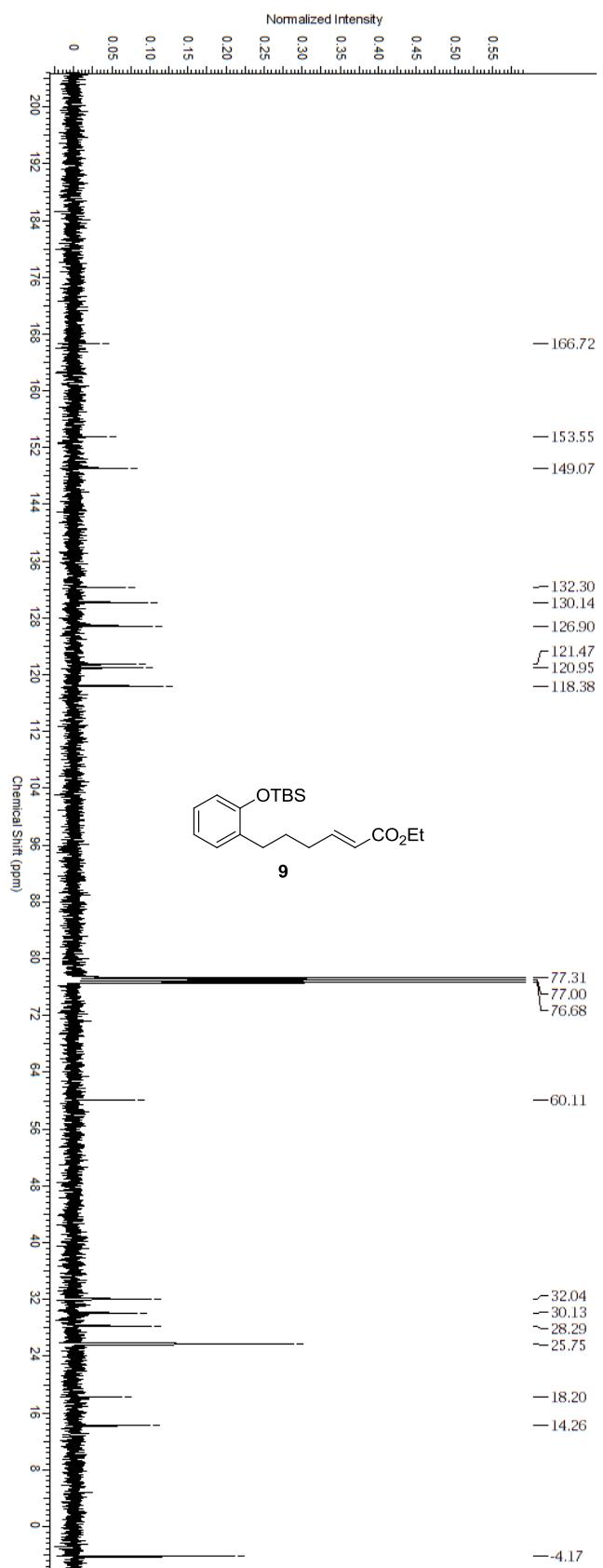
^1H NMR (400 MHz, CDCl_3) spectrum of 8



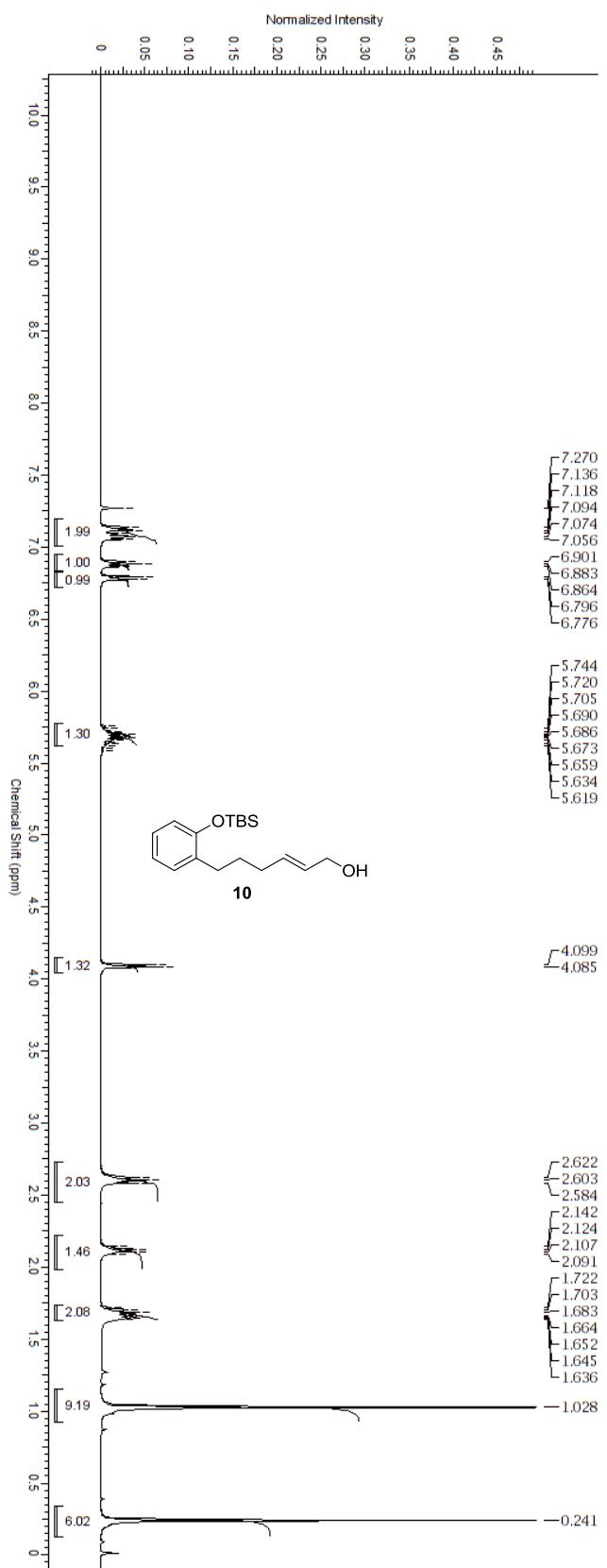
^{13}C NMR (100 MHz, CDCl_3) spectrum of 8



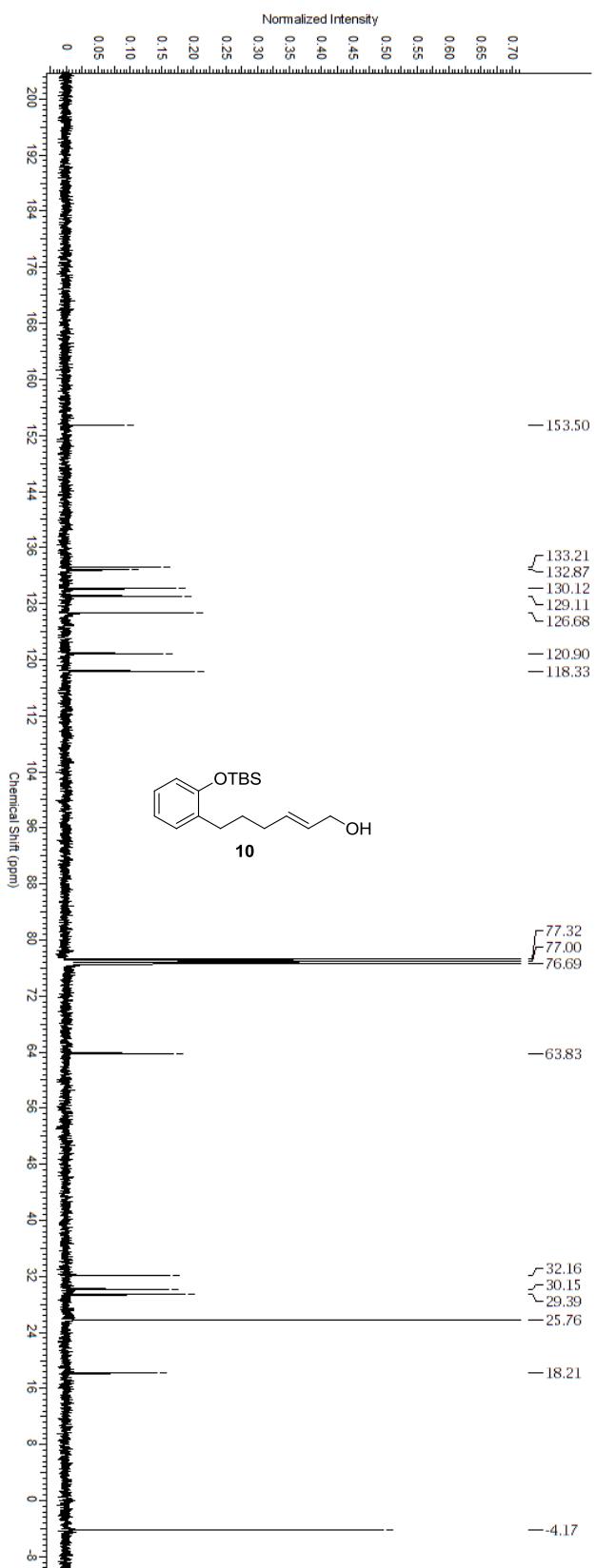
¹H NMR (400 MHz, CDCl₃) spectrum of **9**



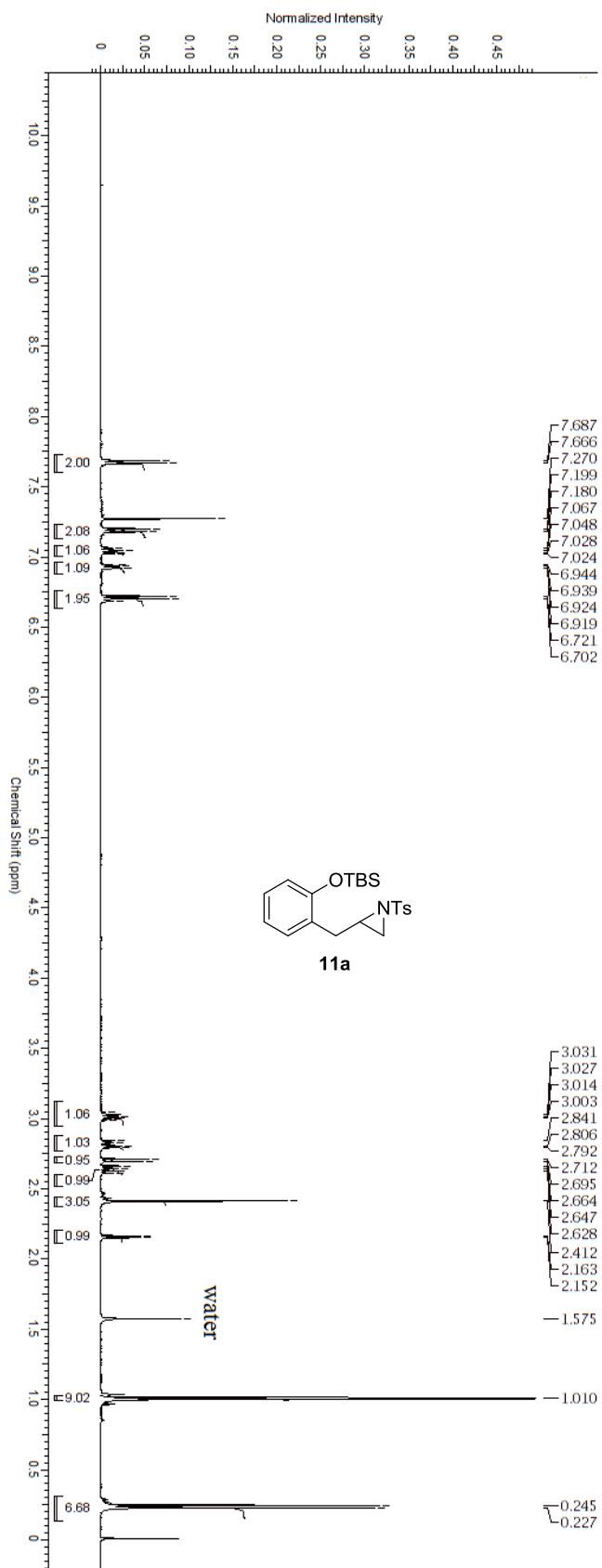
¹³C NMR (100 MHz, CDCl₃) spectrum of 9



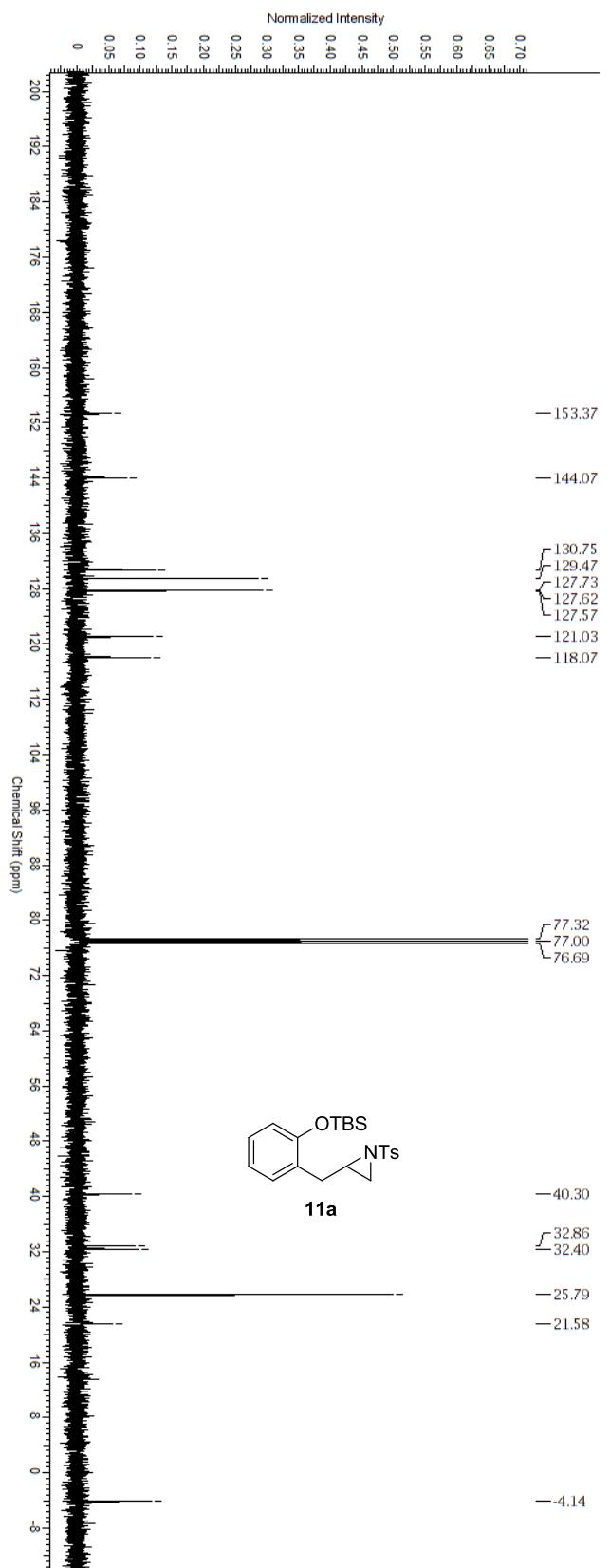
^1H NMR (400 MHz, CDCl_3) spectrum of **10**



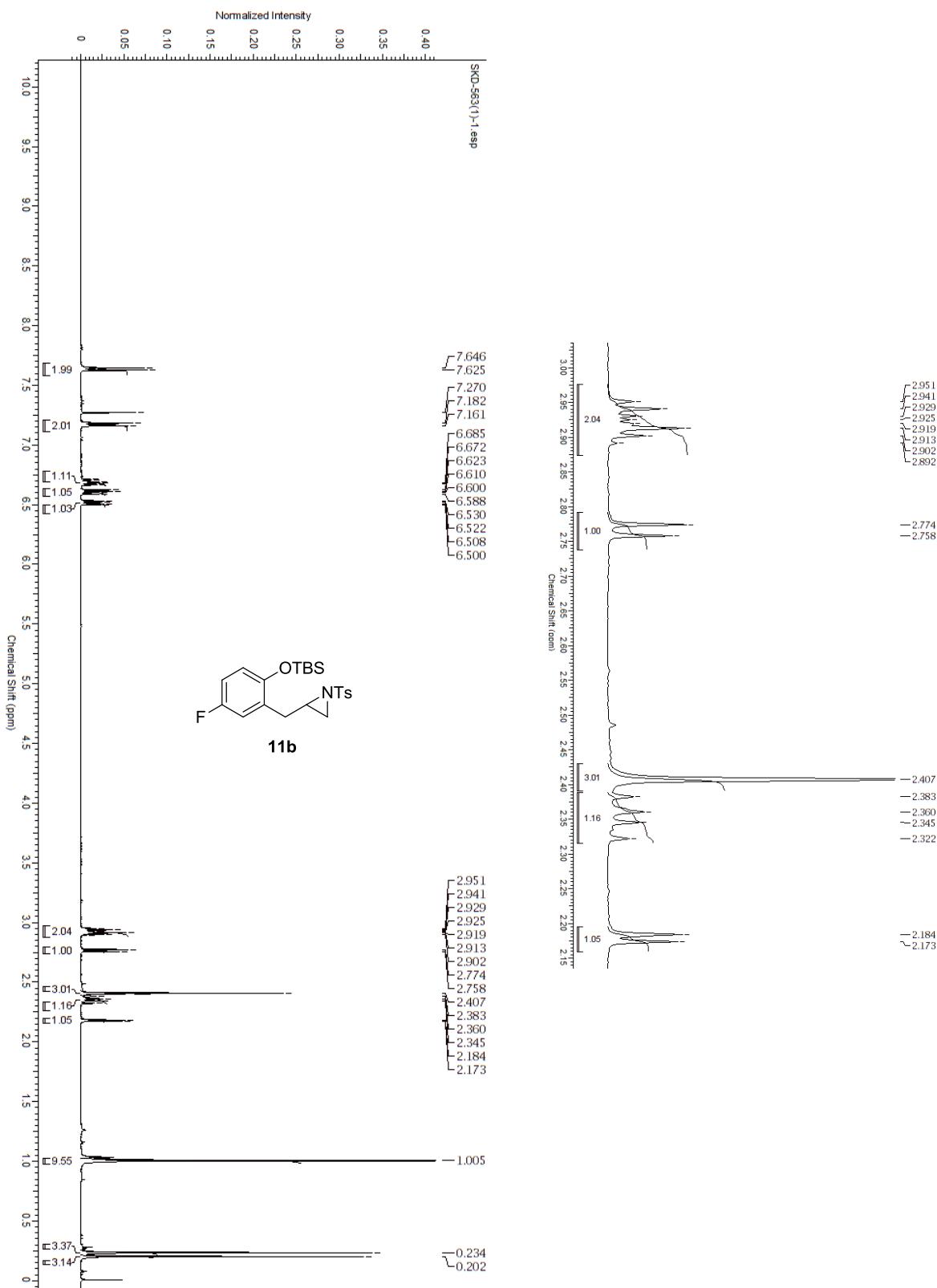
^{13}C NMR (100 MHz, CDCl_3) spectrum of **10**



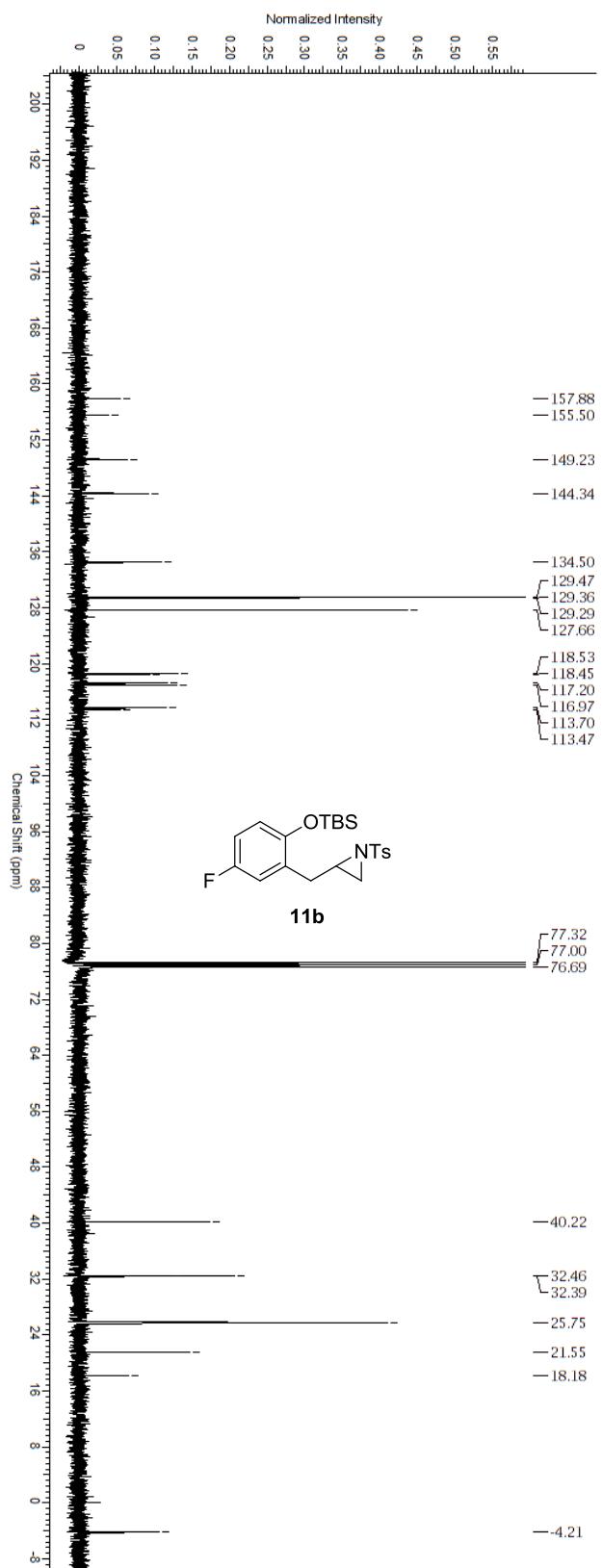
¹H NMR (400 MHz, CDCl₃) spectrum of 11a



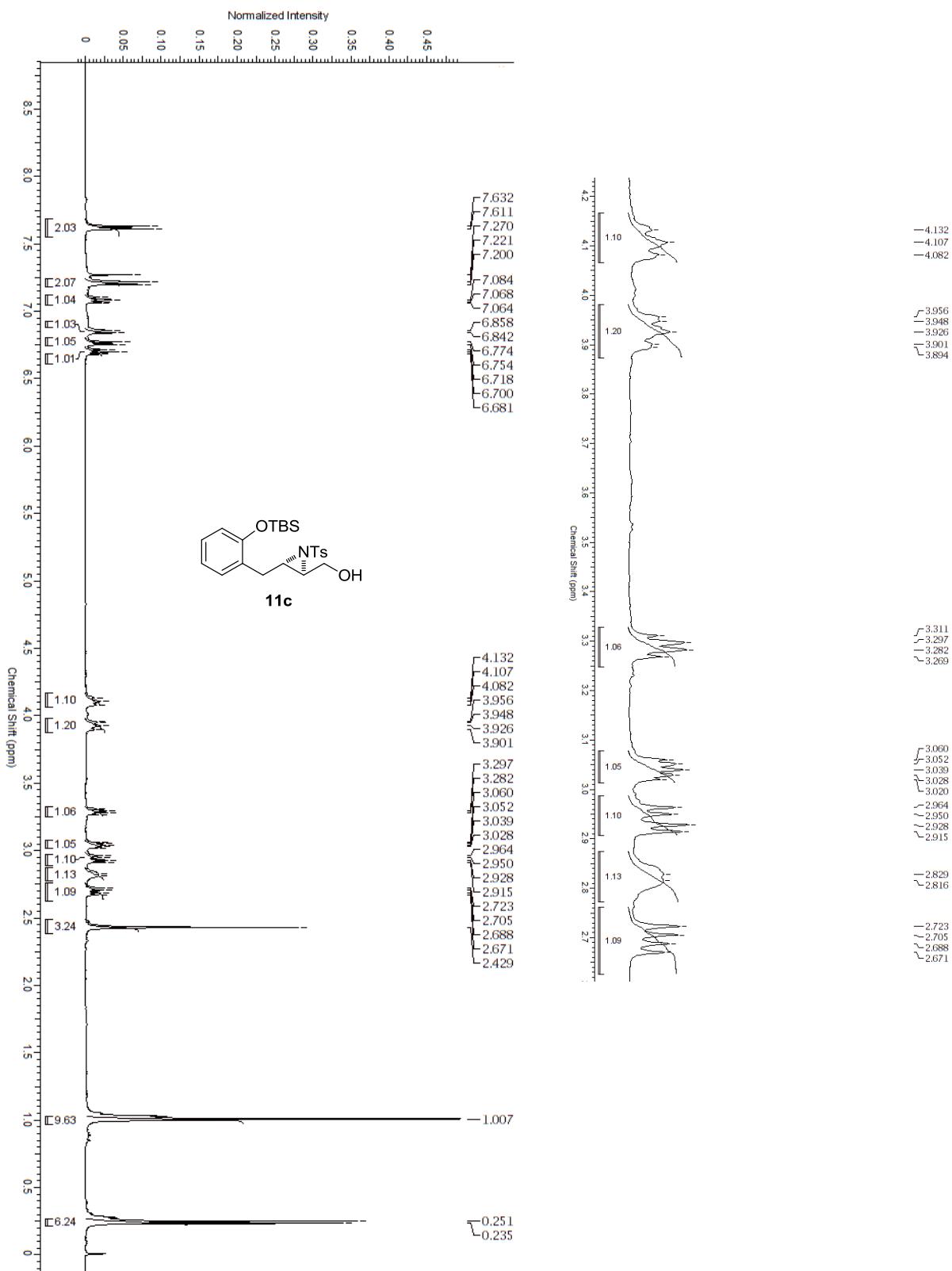
^{13}C NMR (100 MHz, CDCl_3) spectrum of **11a**



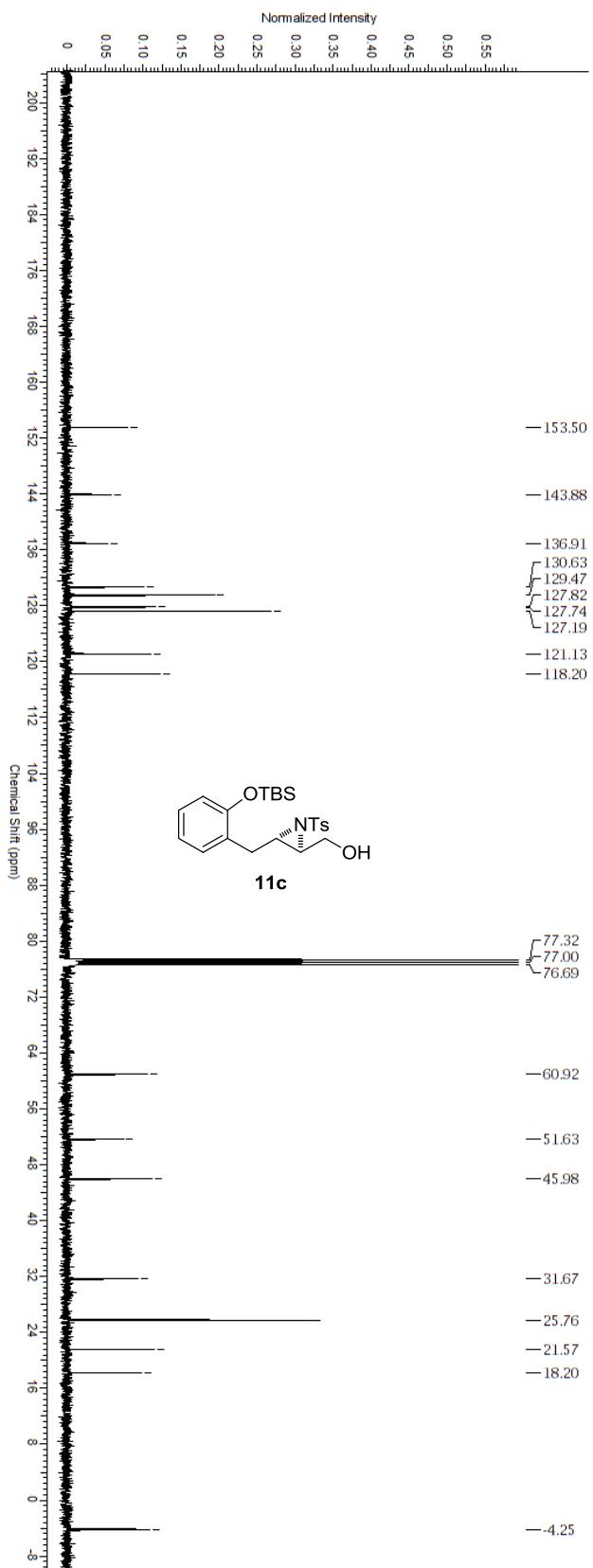
¹H NMR (400 MHz, CDCl₃) spectrum of 11b



^{13}C NMR (100 MHz, CDCl_3) spectrum of **11b**



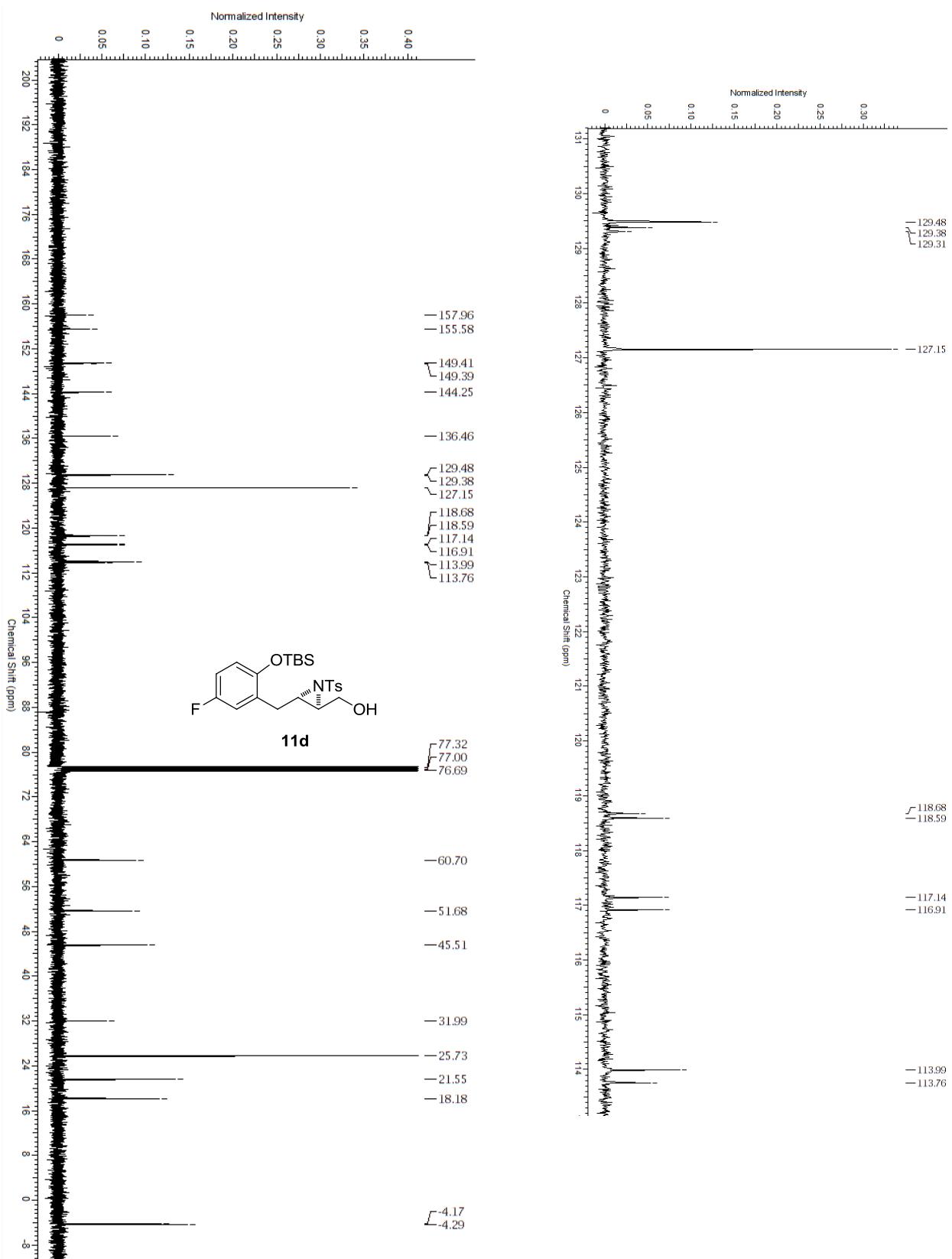
¹H NMR (400 MHz, CDCl₃) spectrum of 11c



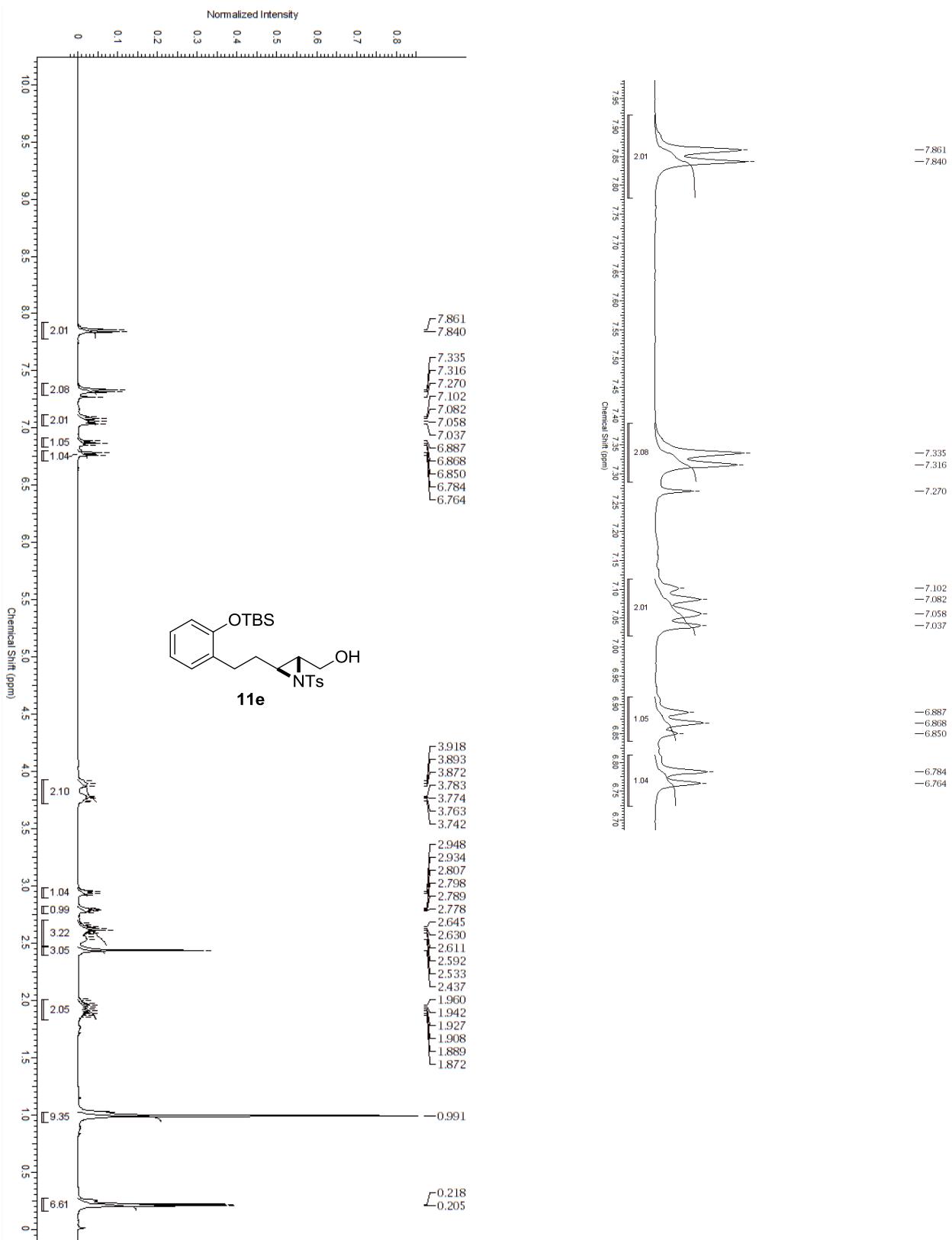
^{13}C NMR (100 MHz, CDCl_3) spectrum of 11c



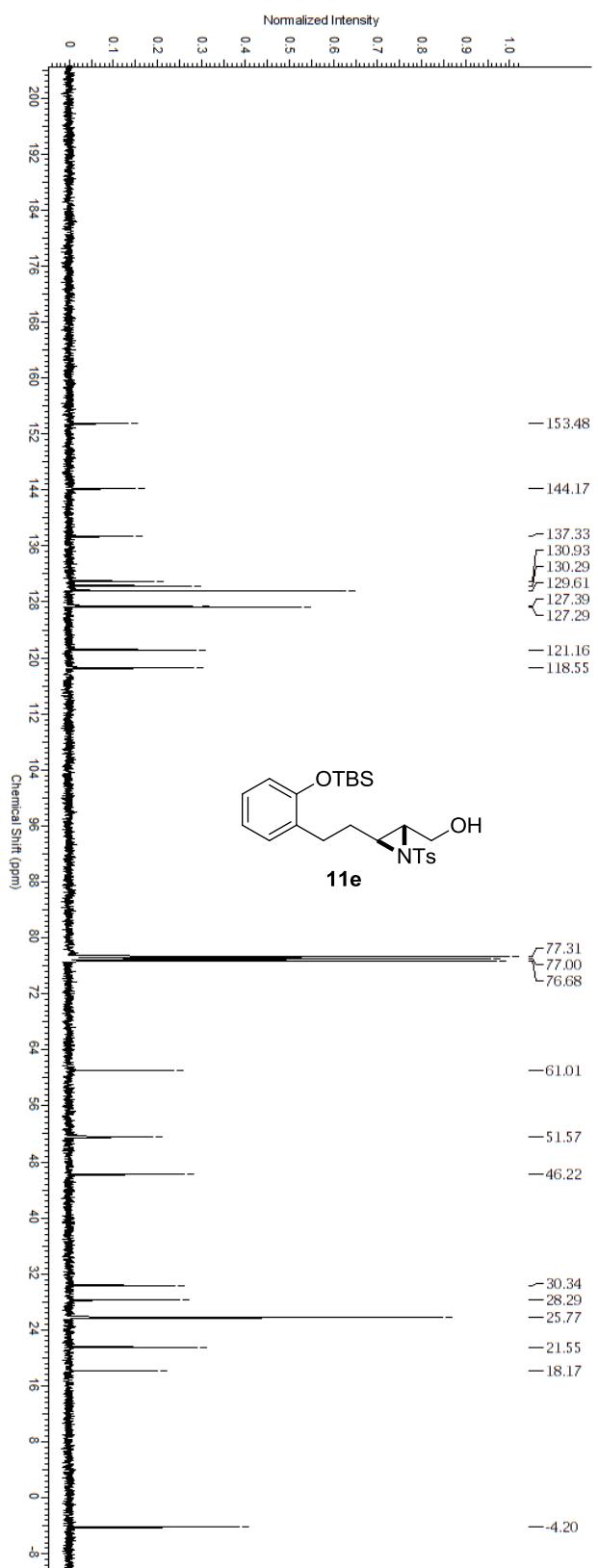
¹H NMR (400 MHz, CDCl₃) spectrum of 11d



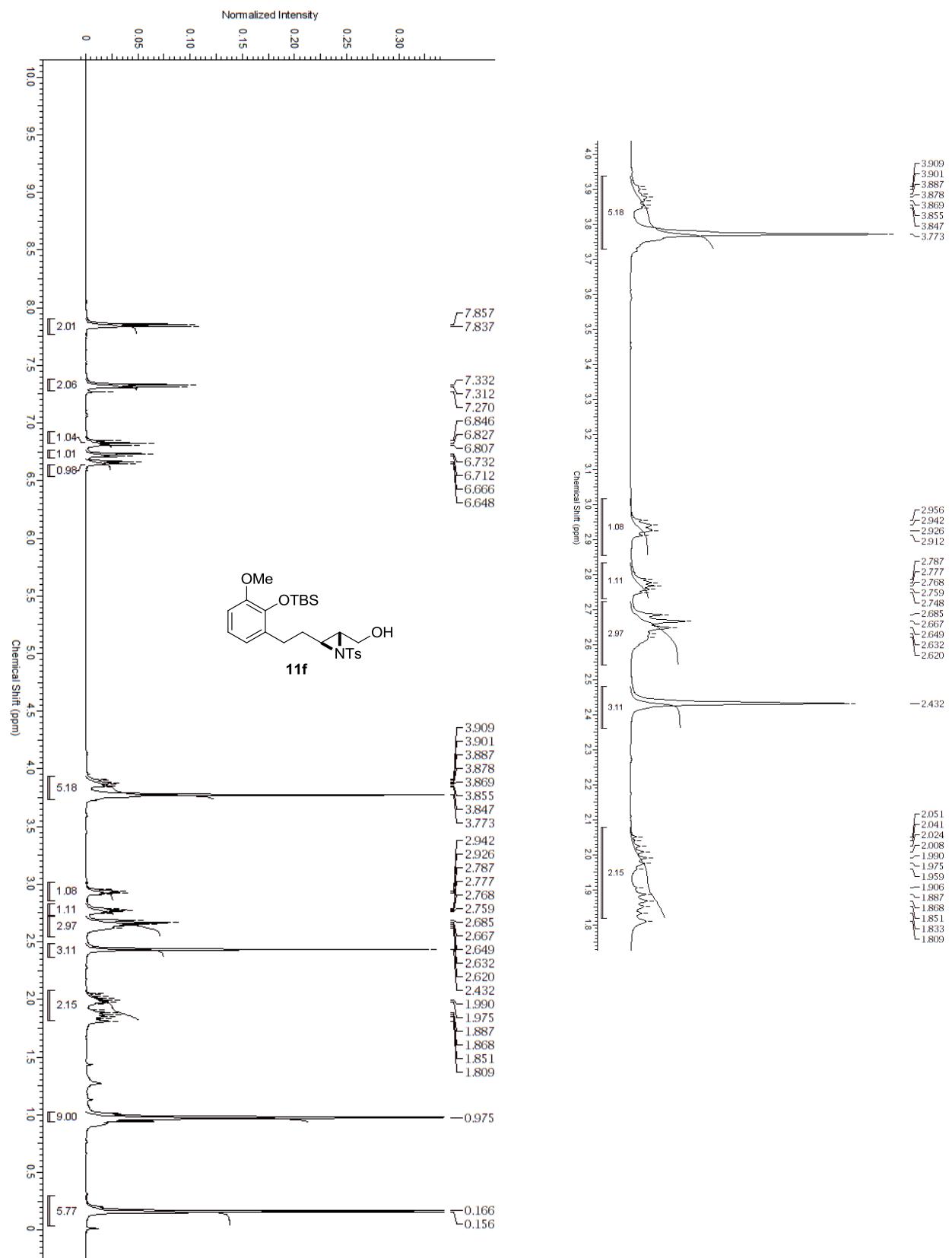
^{13}C NMR (100 MHz, CDCl_3) spectrum of 11d



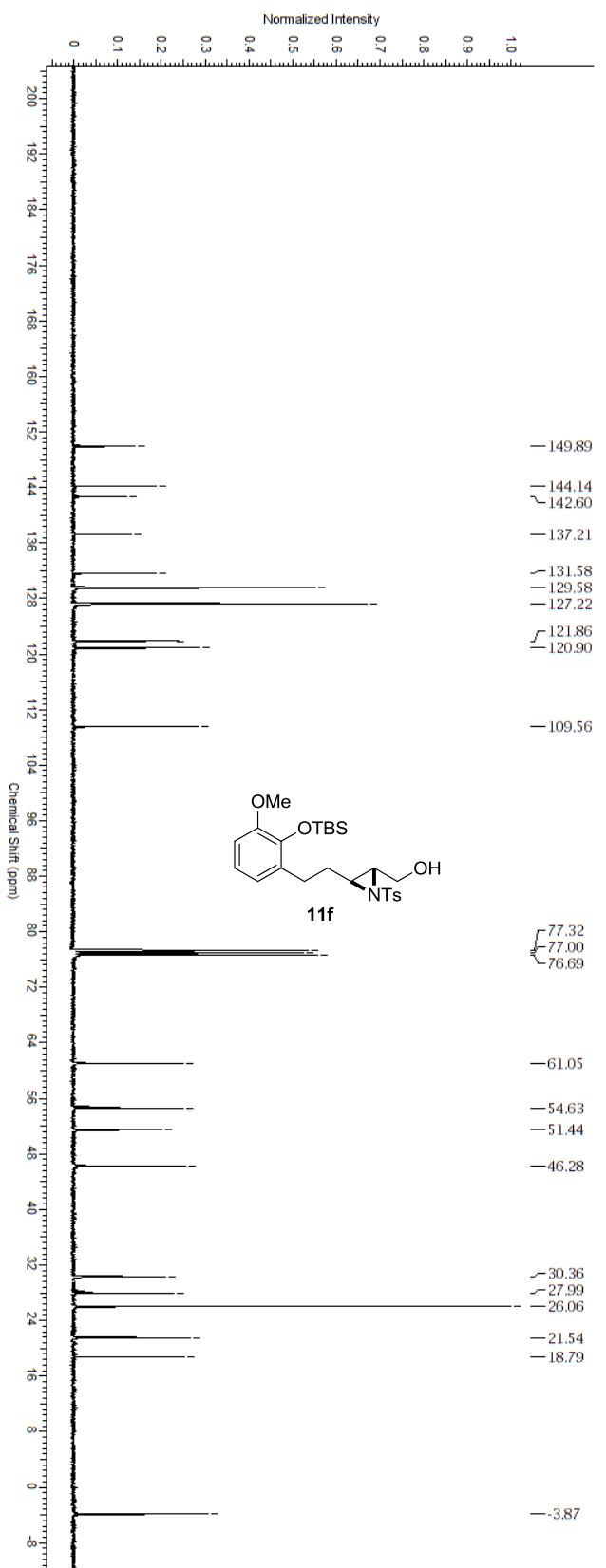
¹H NMR (400 MHz, CDCl₃) spectrum of 11e



^{13}C NMR (100 MHz, CDCl_3) spectrum of 11e



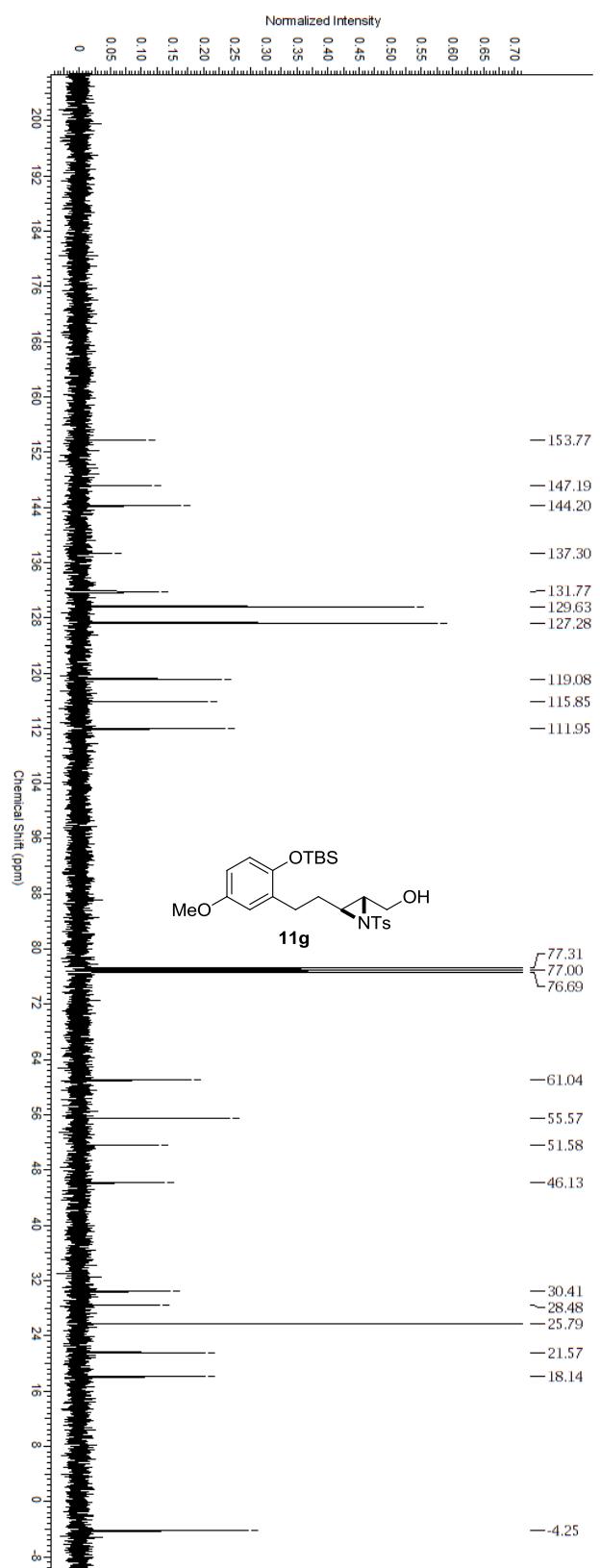
^1H NMR (400 MHz, CDCl_3) spectrum of 11f



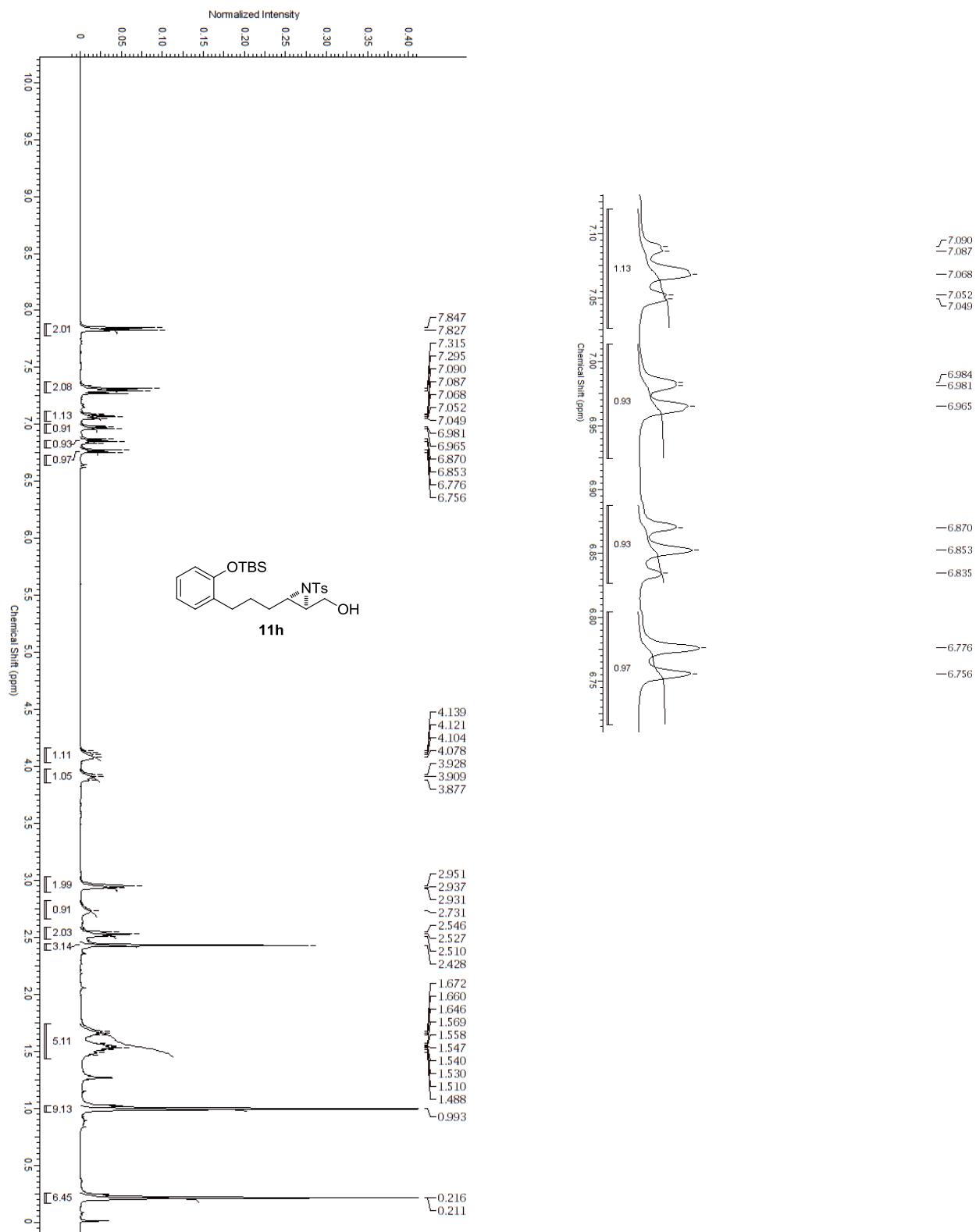
^{13}C NMR (100 MHz, CDCl_3) spectrum of **11f**



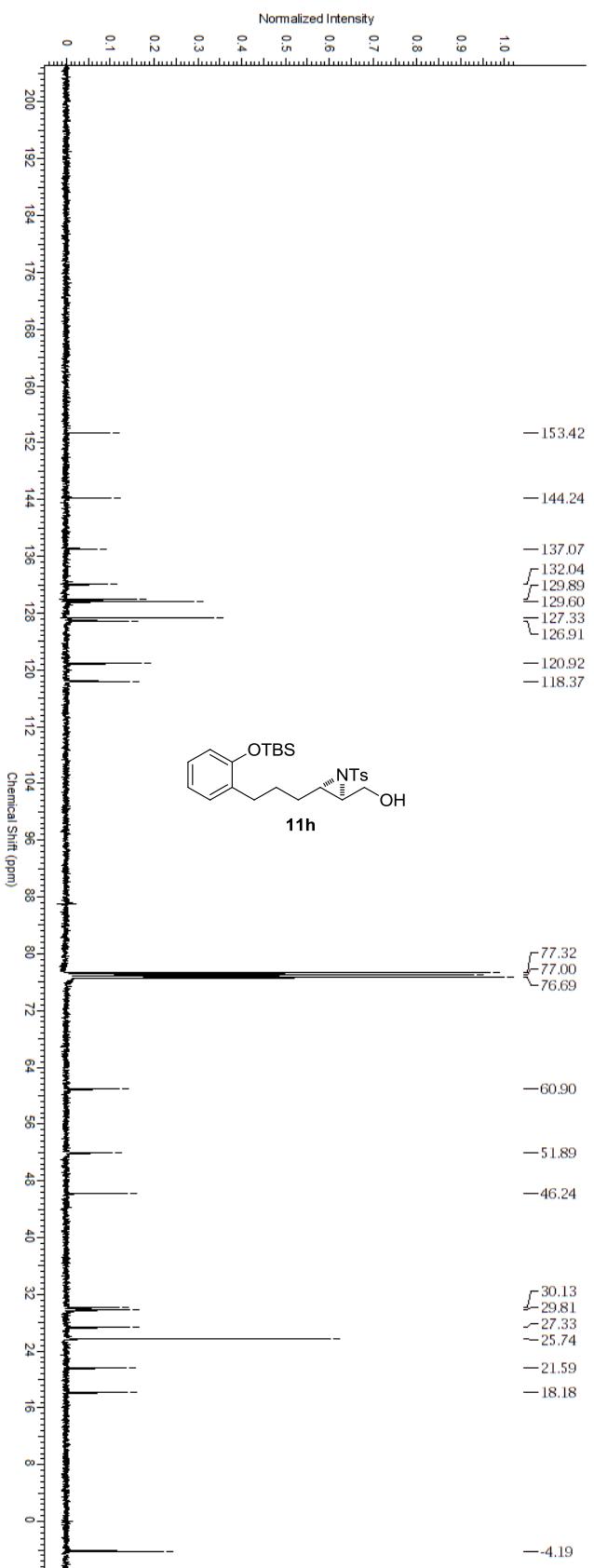
¹H NMR (400 MHz, CDCl₃) spectrum of 11g



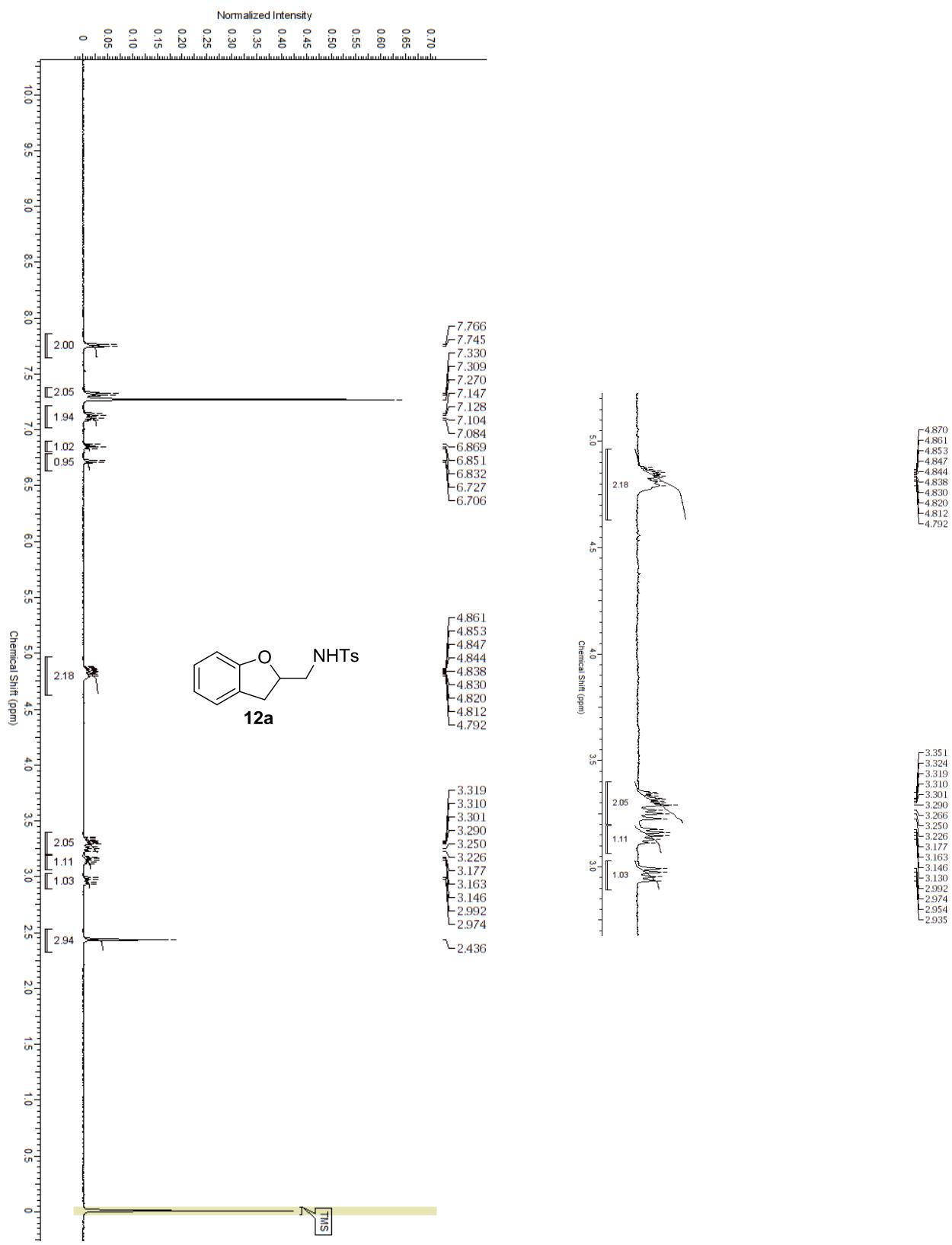
¹³C NMR (100 MHz, CDCl₃) spectrum of 11g



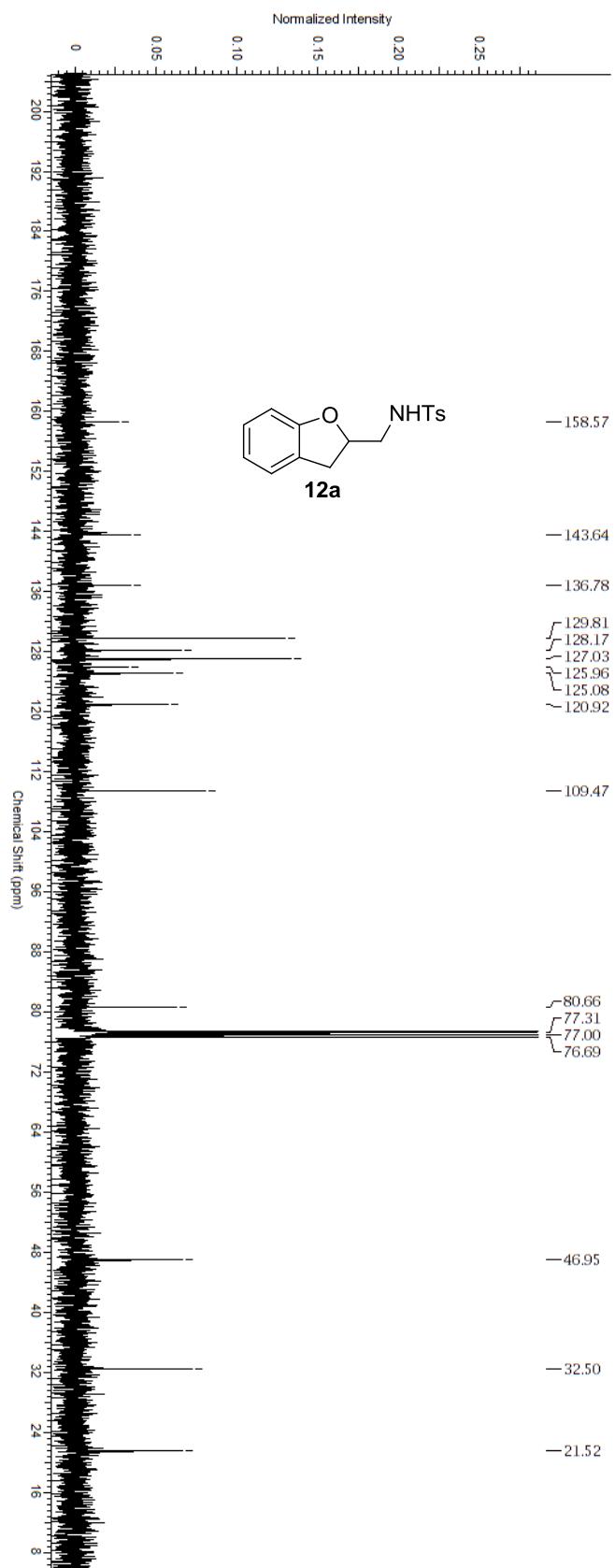
^1H NMR (400 MHz, CDCl_3) spectrum of **11h**



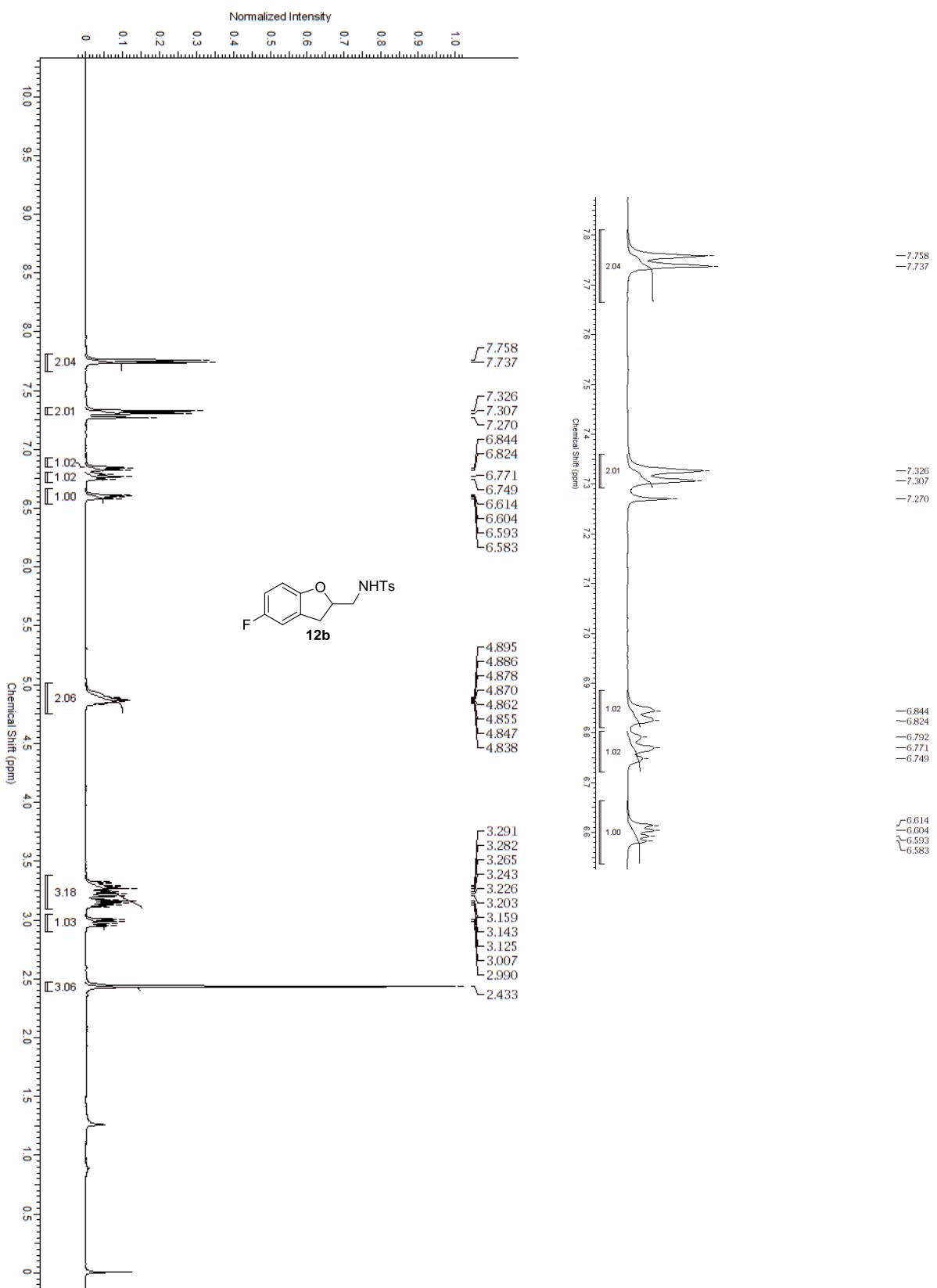
^{13}C NMR (100 MHz, CDCl_3) spectrum of **11h**



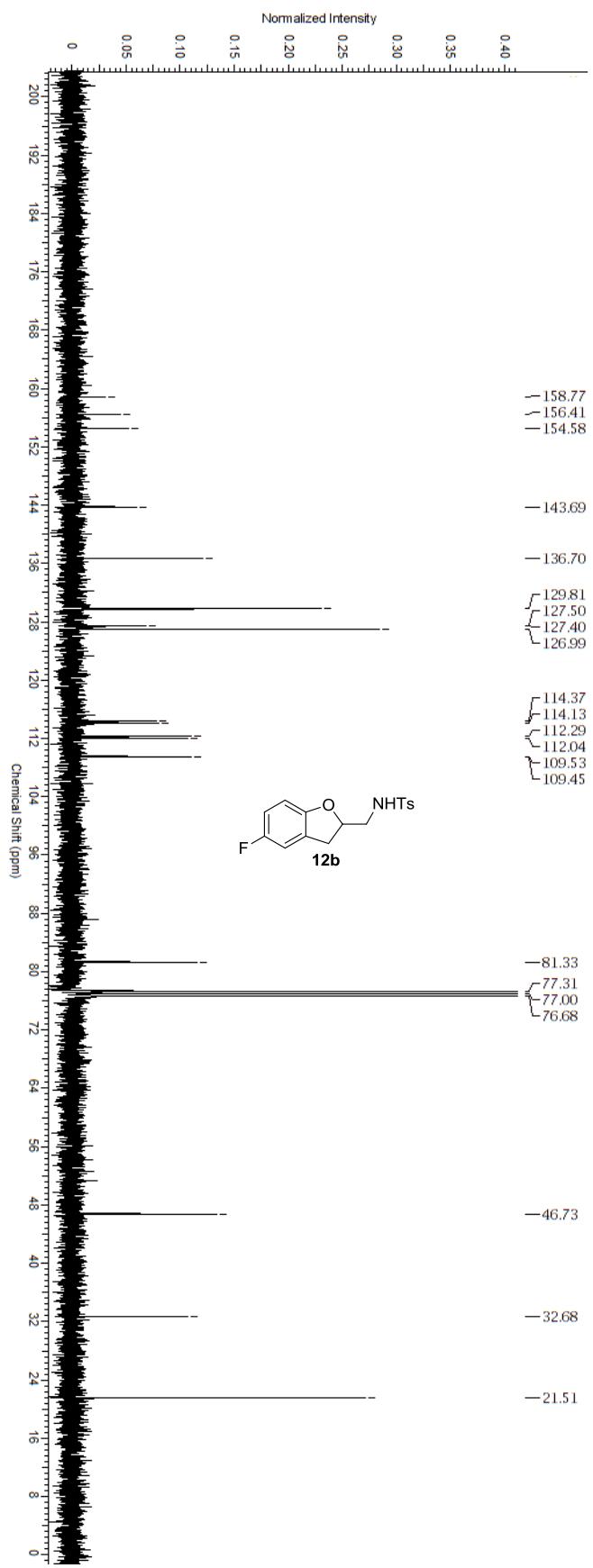
¹H NMR (400 MHz, CDCl₃) spectrum of 12a



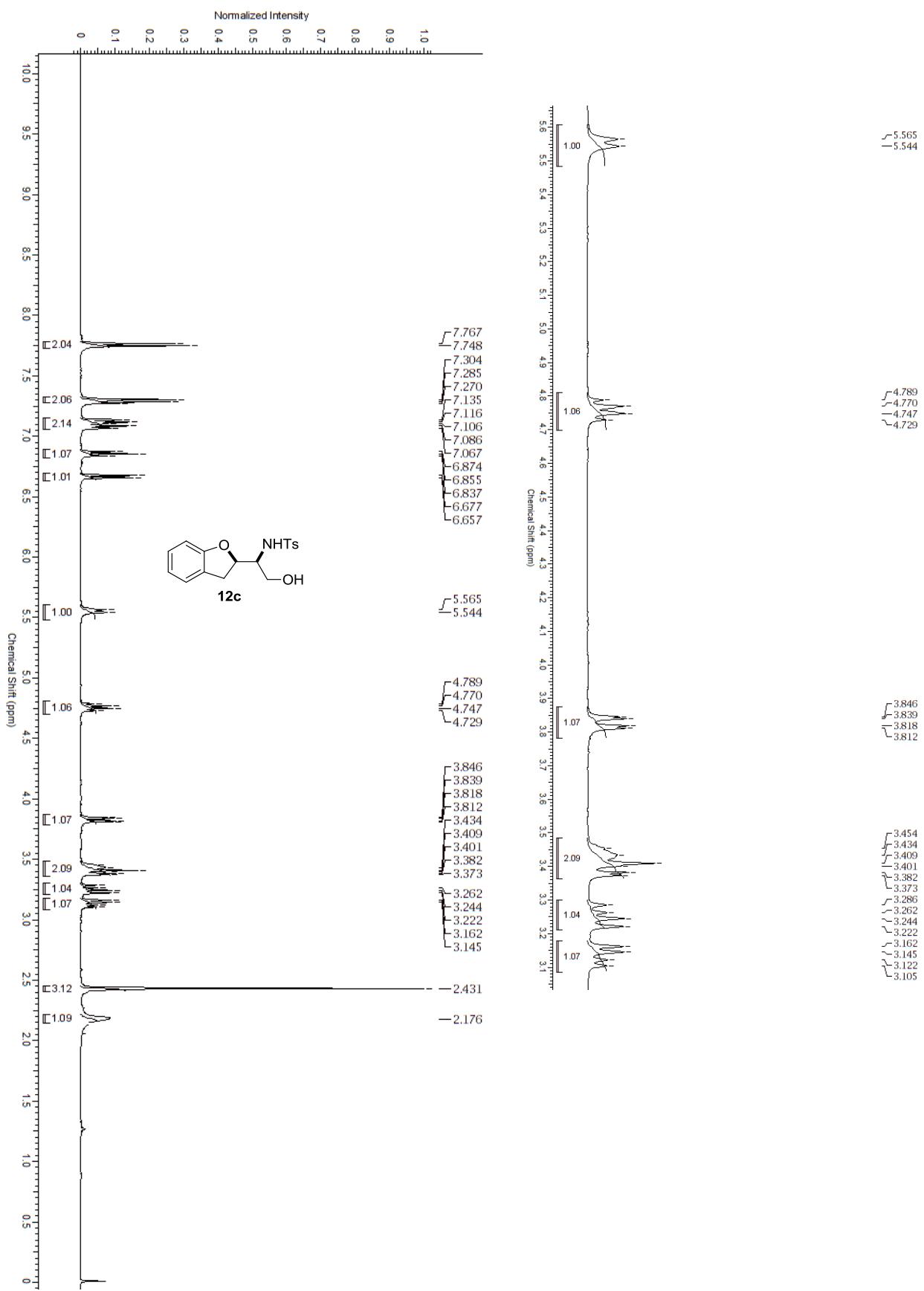
^{13}C NMR (100 MHz, CDCl_3) spectrum of 12a



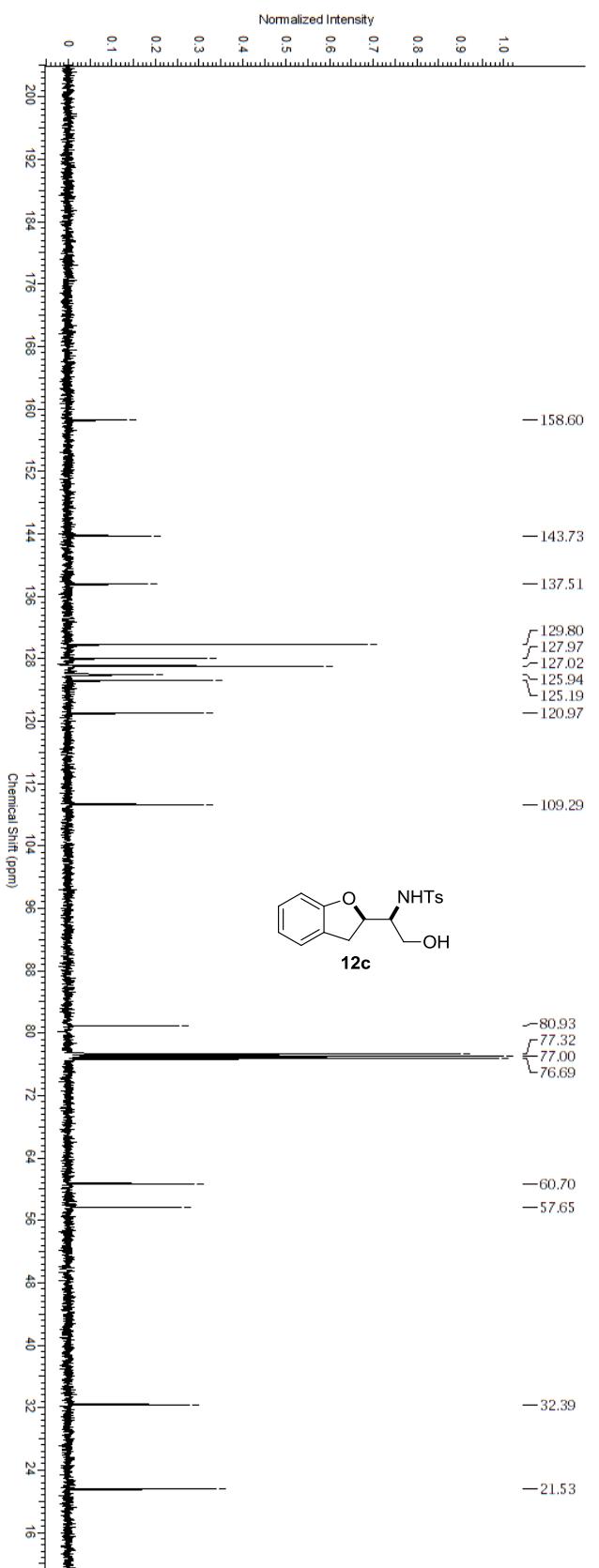
¹H NMR (400 MHz, CDCl₃) spectrum of **12b**



^{13}C NMR (100 MHz, CDCl_3) spectrum of **12b**



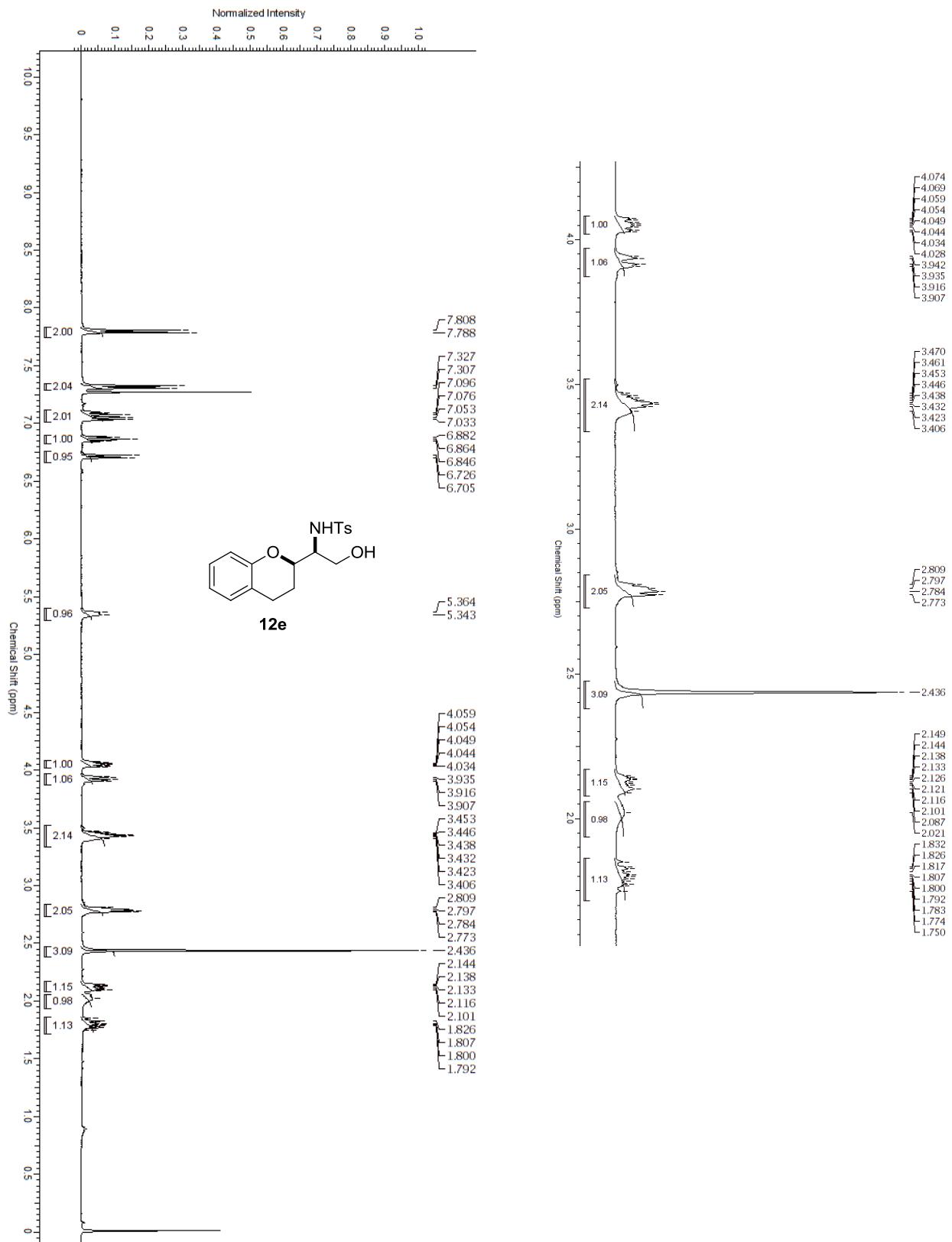
¹H NMR (400 MHz, CDCl₃) spectrum of 12c



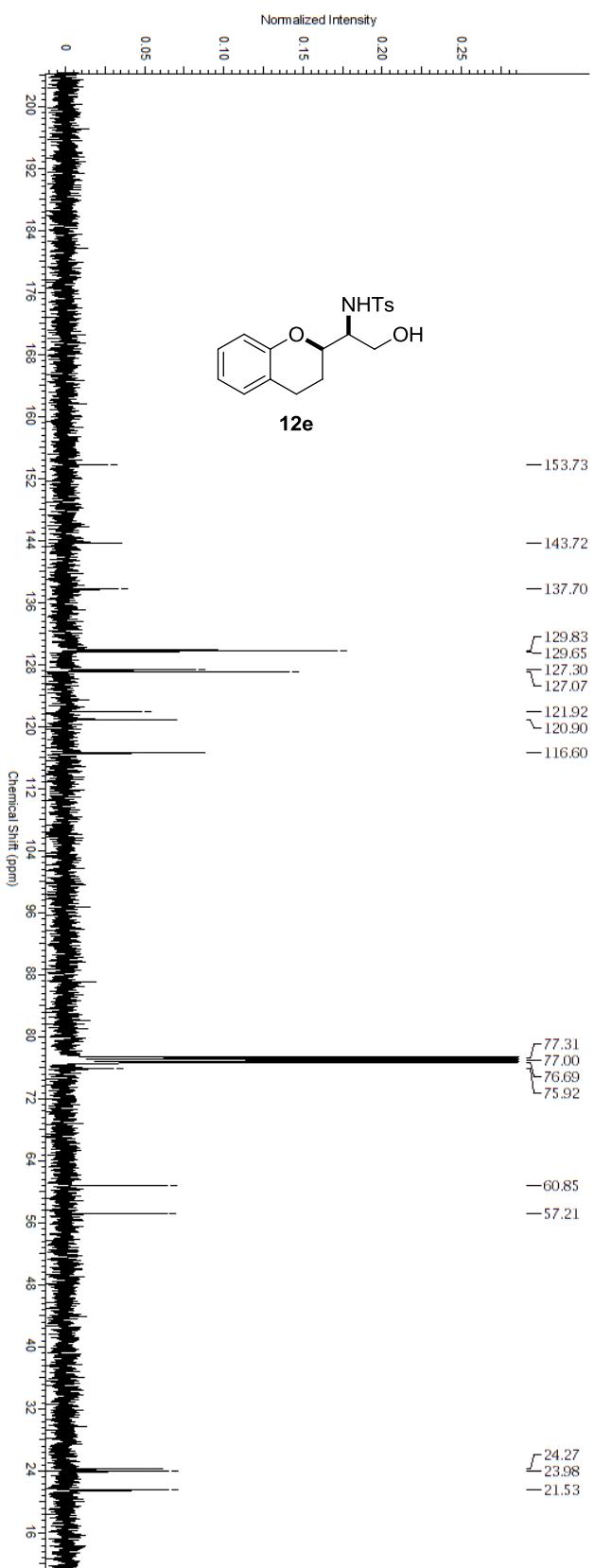
^{13}C NMR (100 MHz, CDCl_3) spectrum of 12c



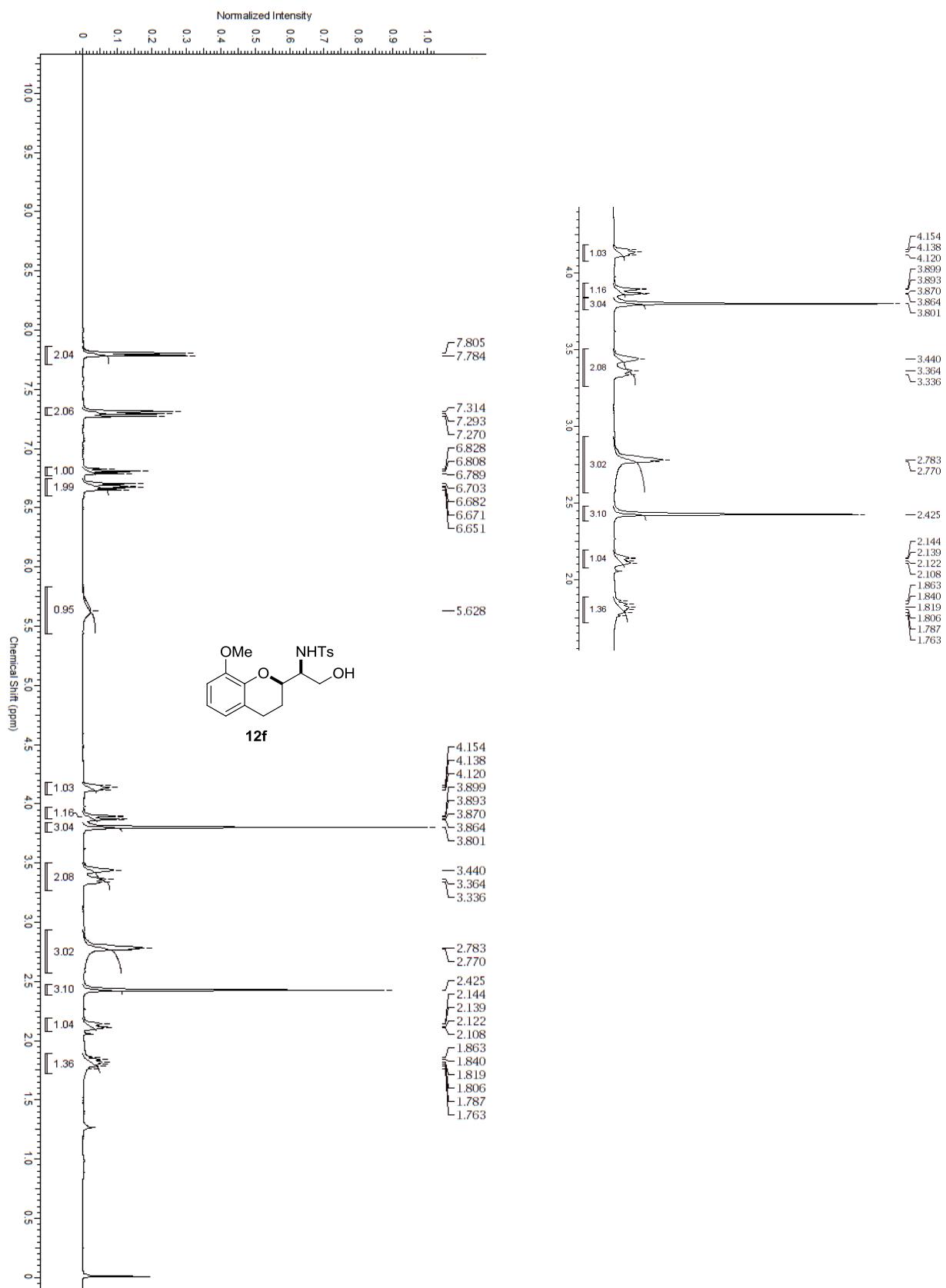
¹H NMR (400 MHz, CDCl₃) spectrum of 12d



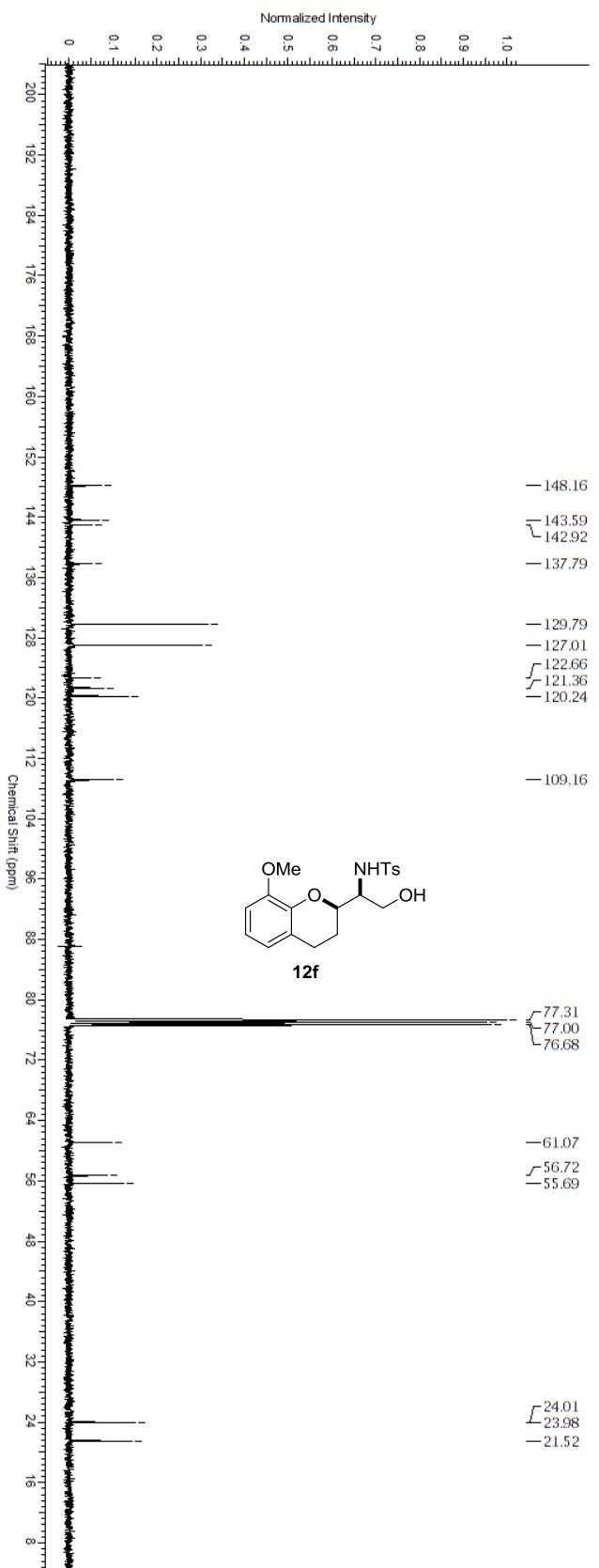
¹H NMR (400 MHz, CDCl₃) spectrum of 12e



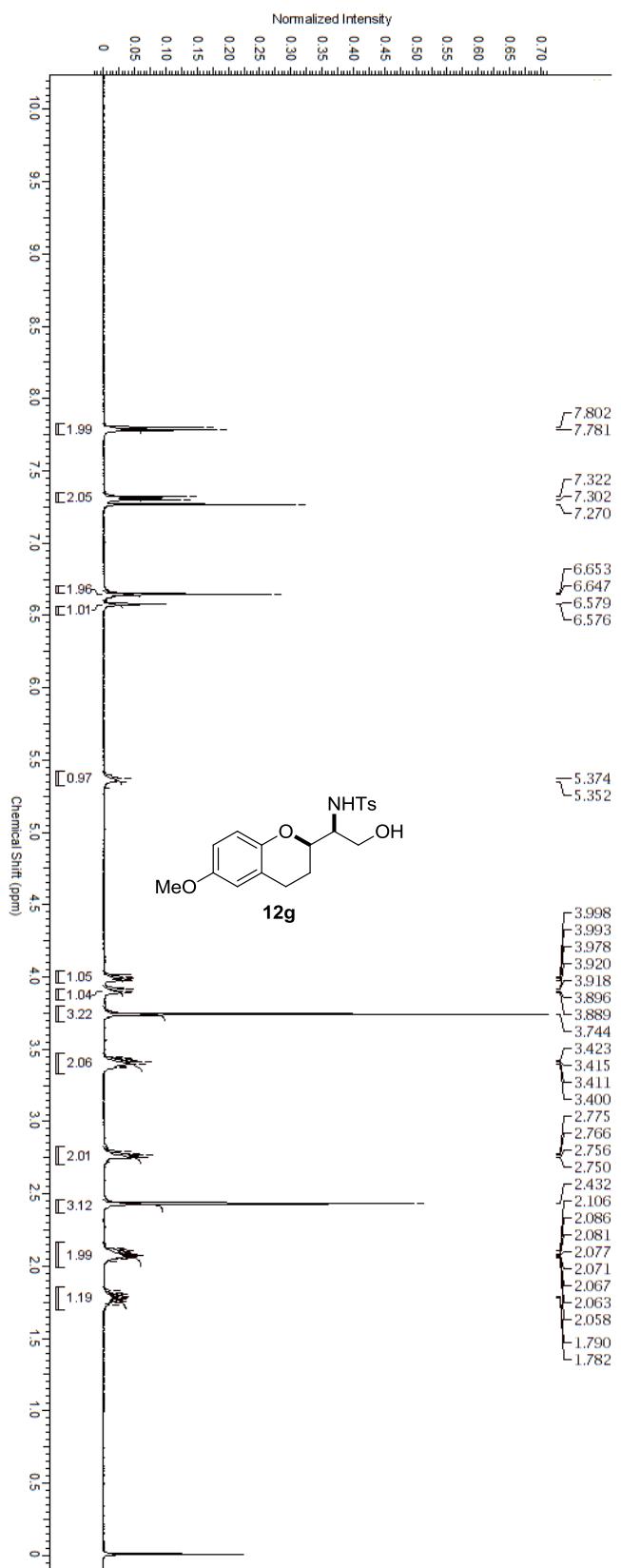
^{13}C NMR (100 MHz, CDCl_3) spectrum of 12e



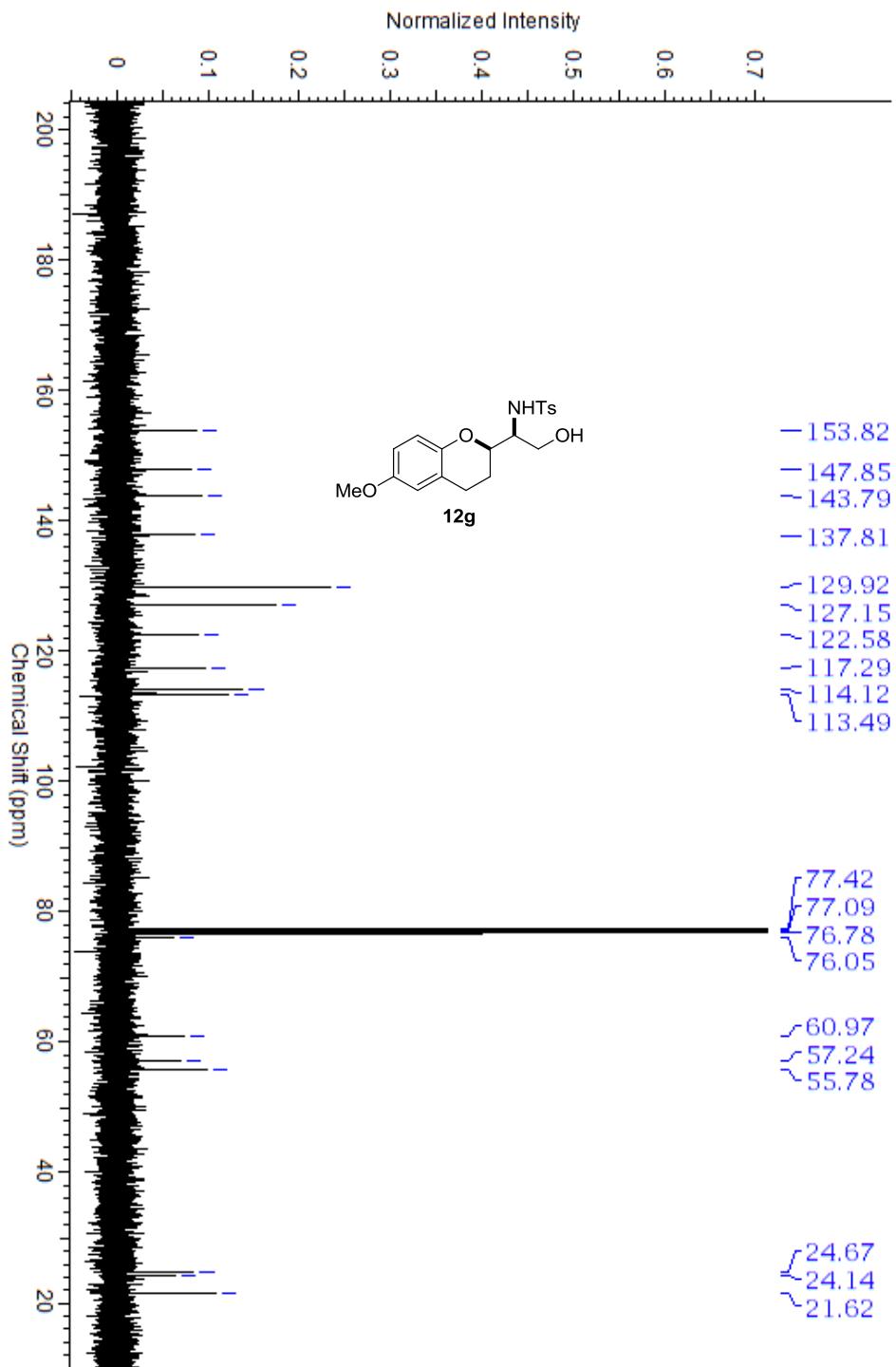
¹H NMR (400 MHz, CDCl₃) spectrum of 12f



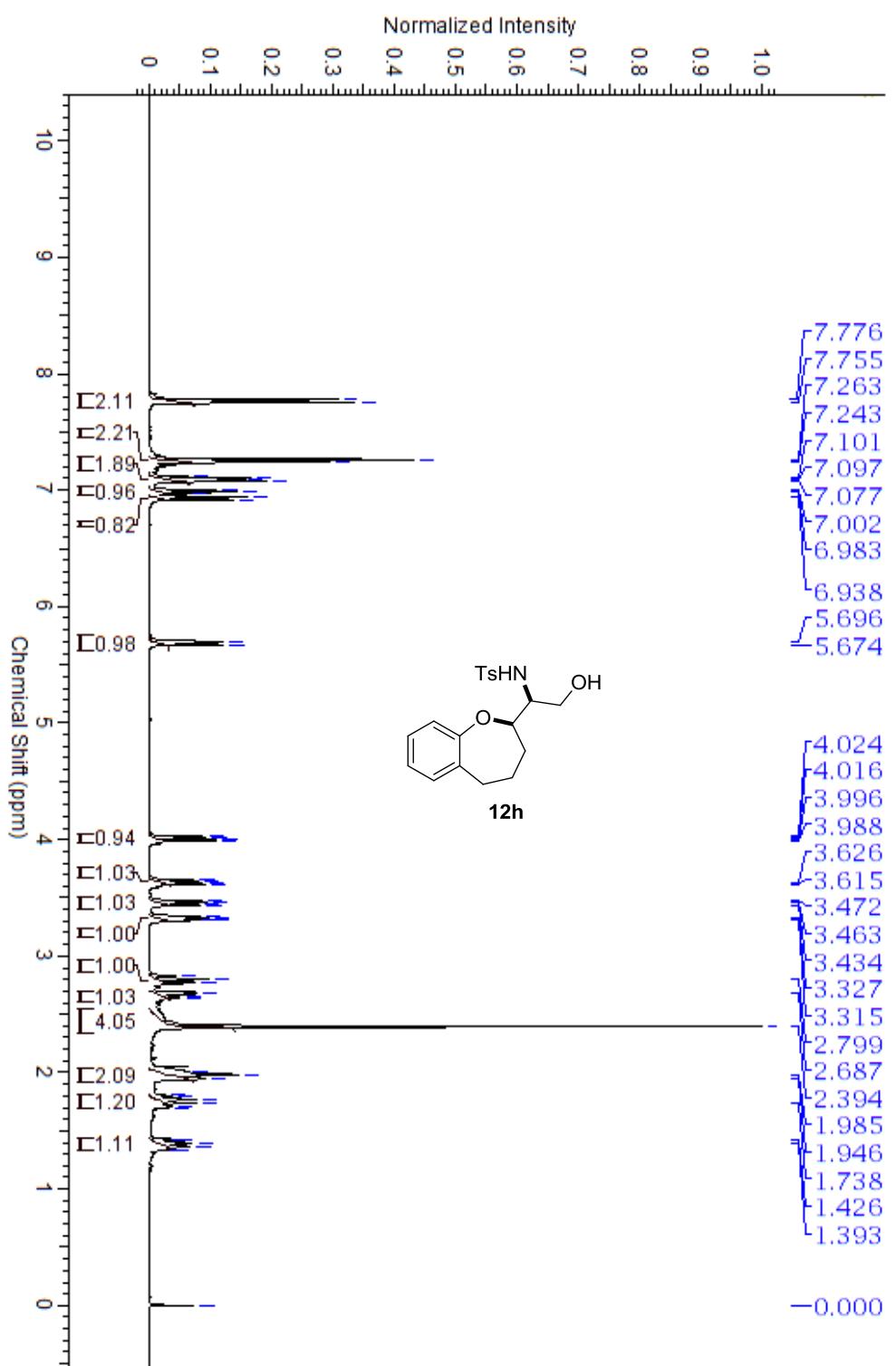
^{13}C NMR (100 MHz, CDCl_3) spectrum of 12f



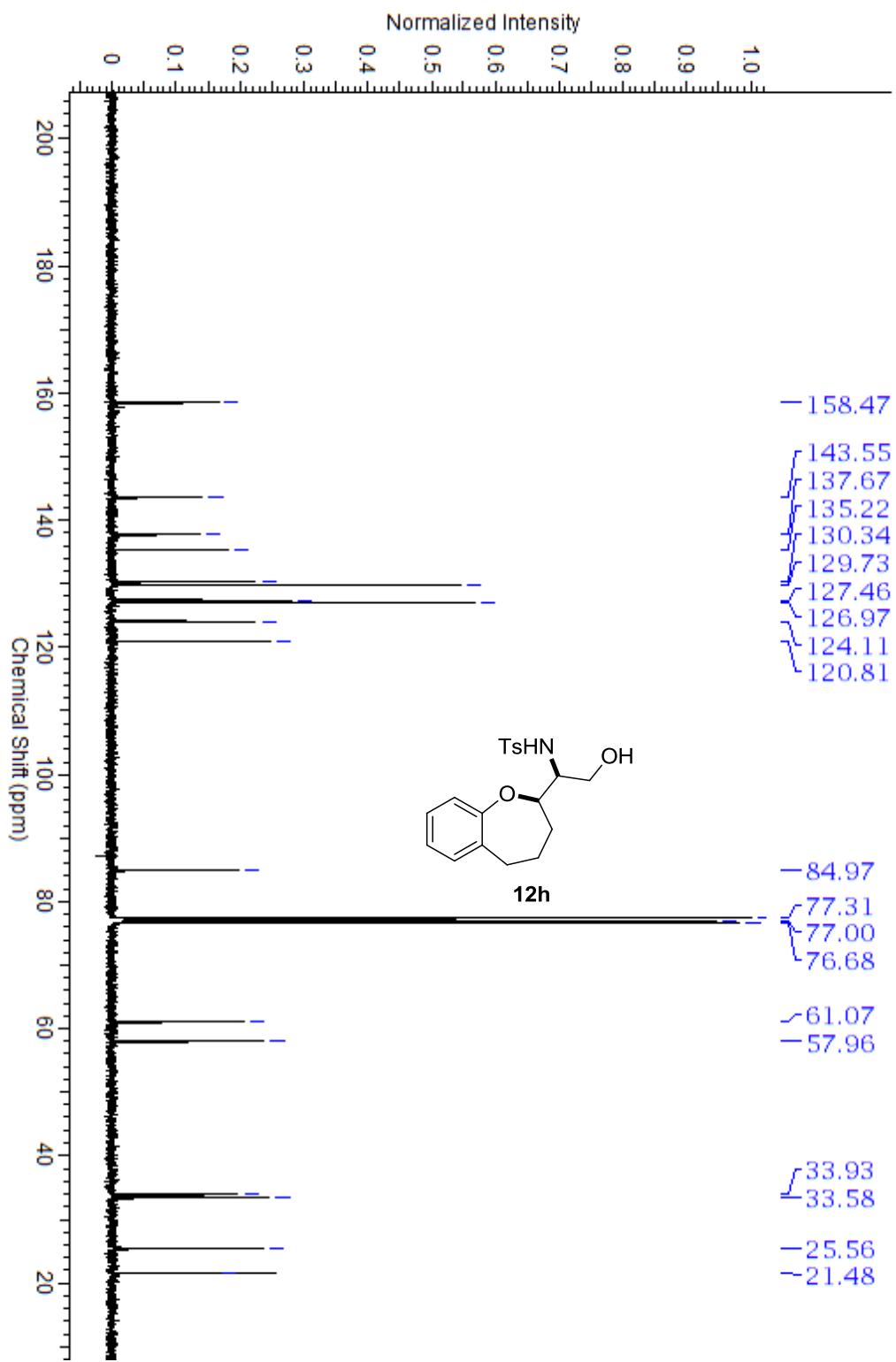
^1H NMR (400 MHz, CDCl_3) spectrum of **12g**



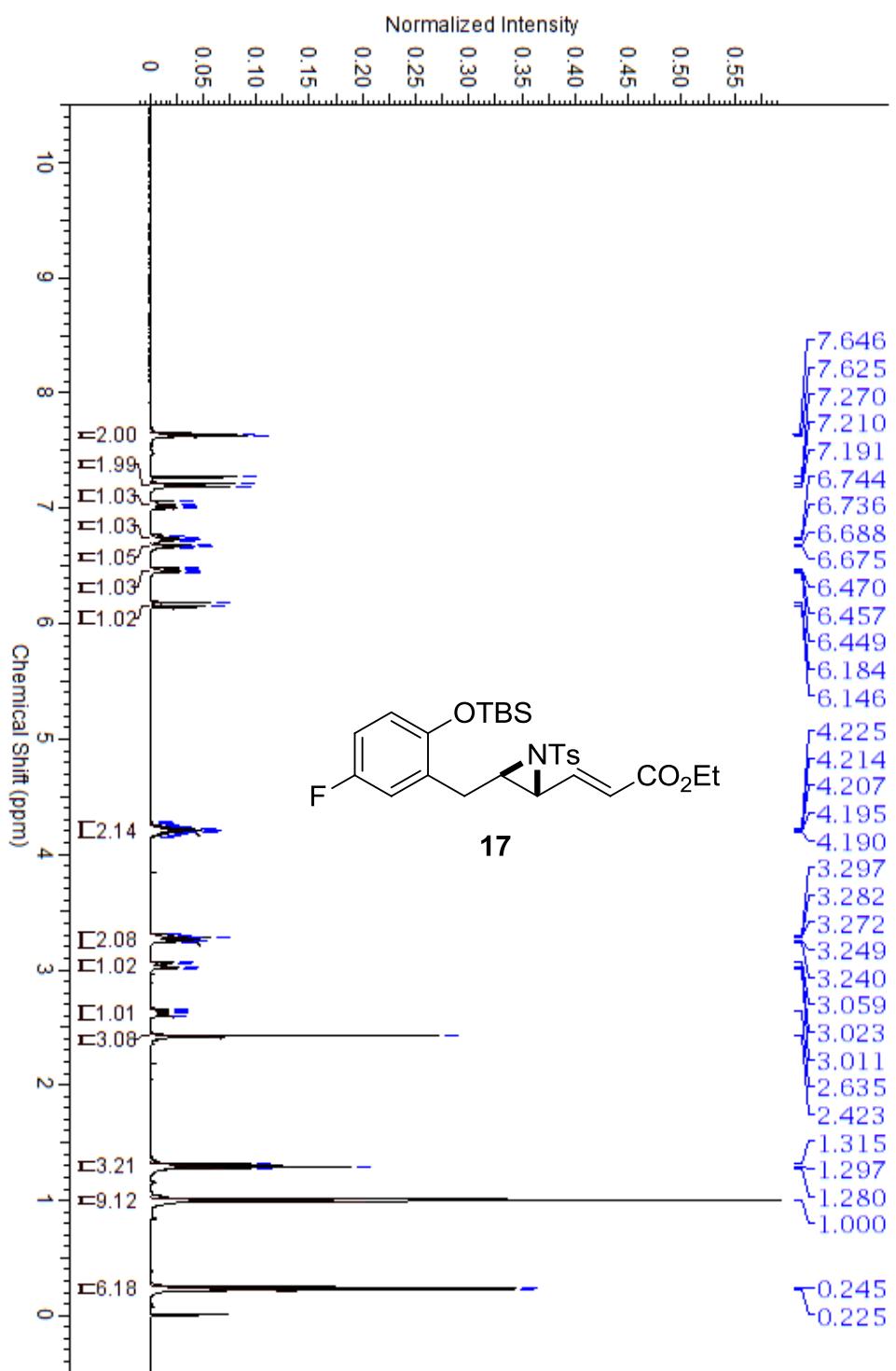
^{13}C NMR (100 MHz, CDCl_3) spectrum of **12g**



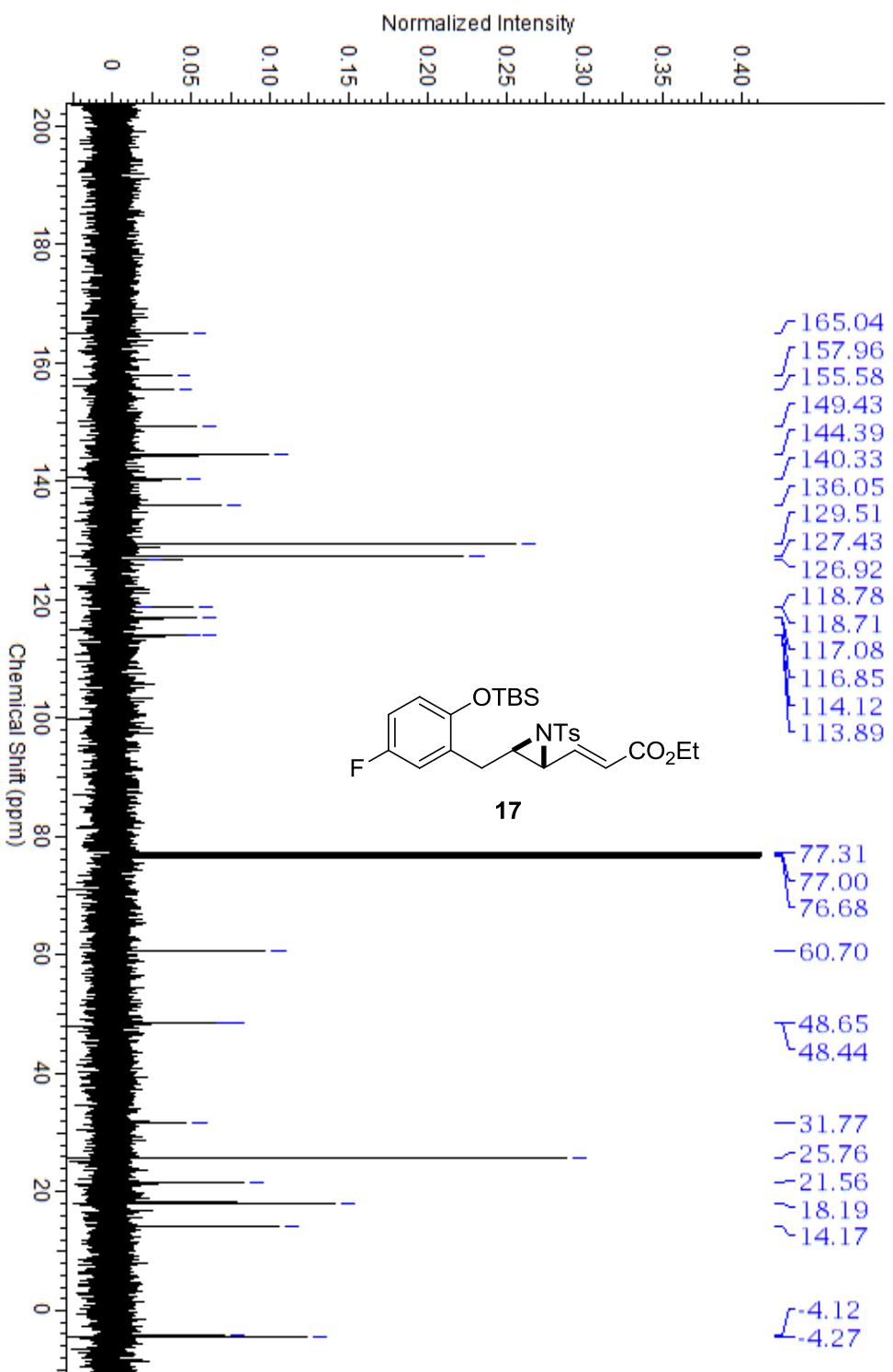
¹H NMR (400 MHz, CDCl₃) of 12h



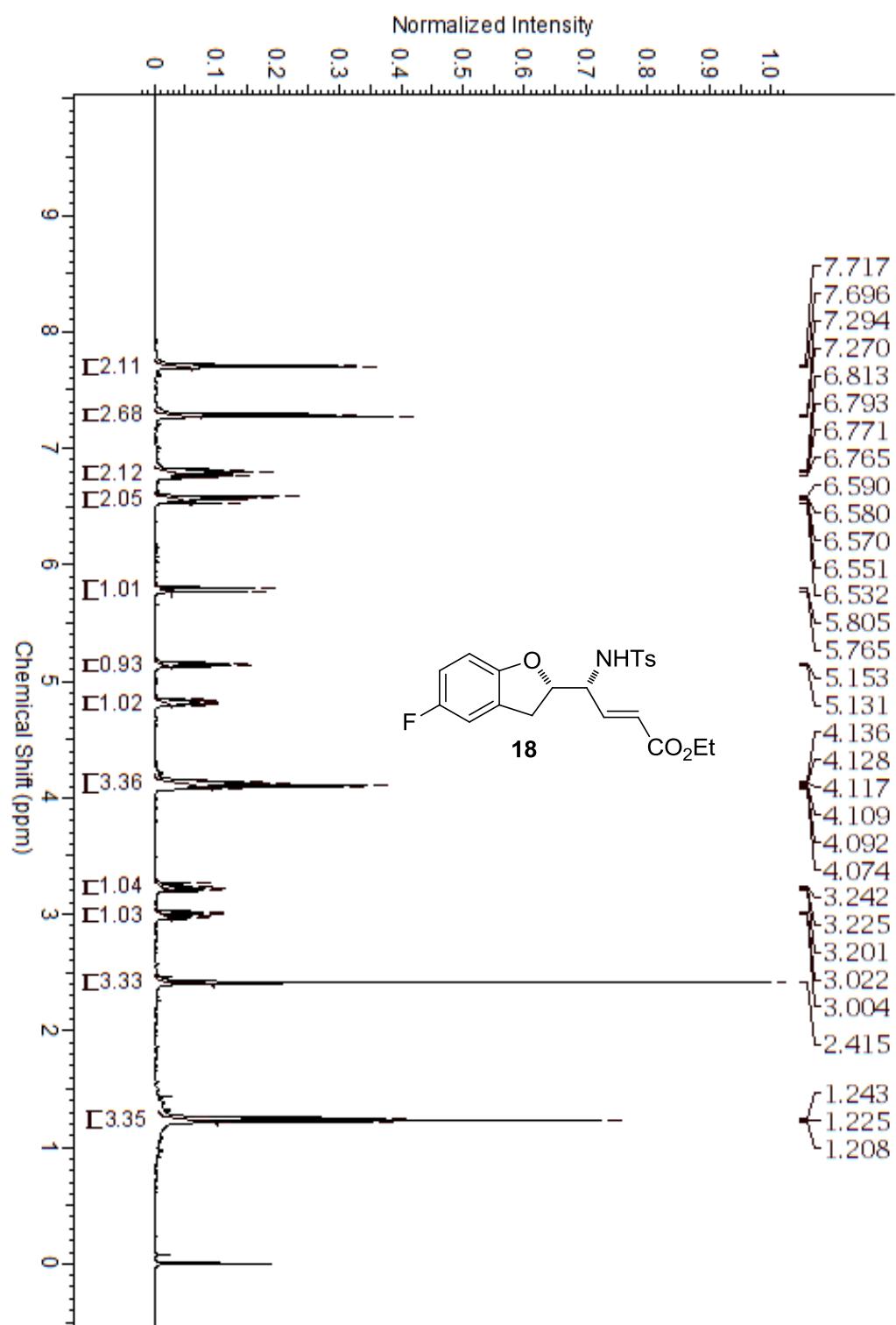
^{13}C NMR (100 MHz, CDCl_3) of 12h



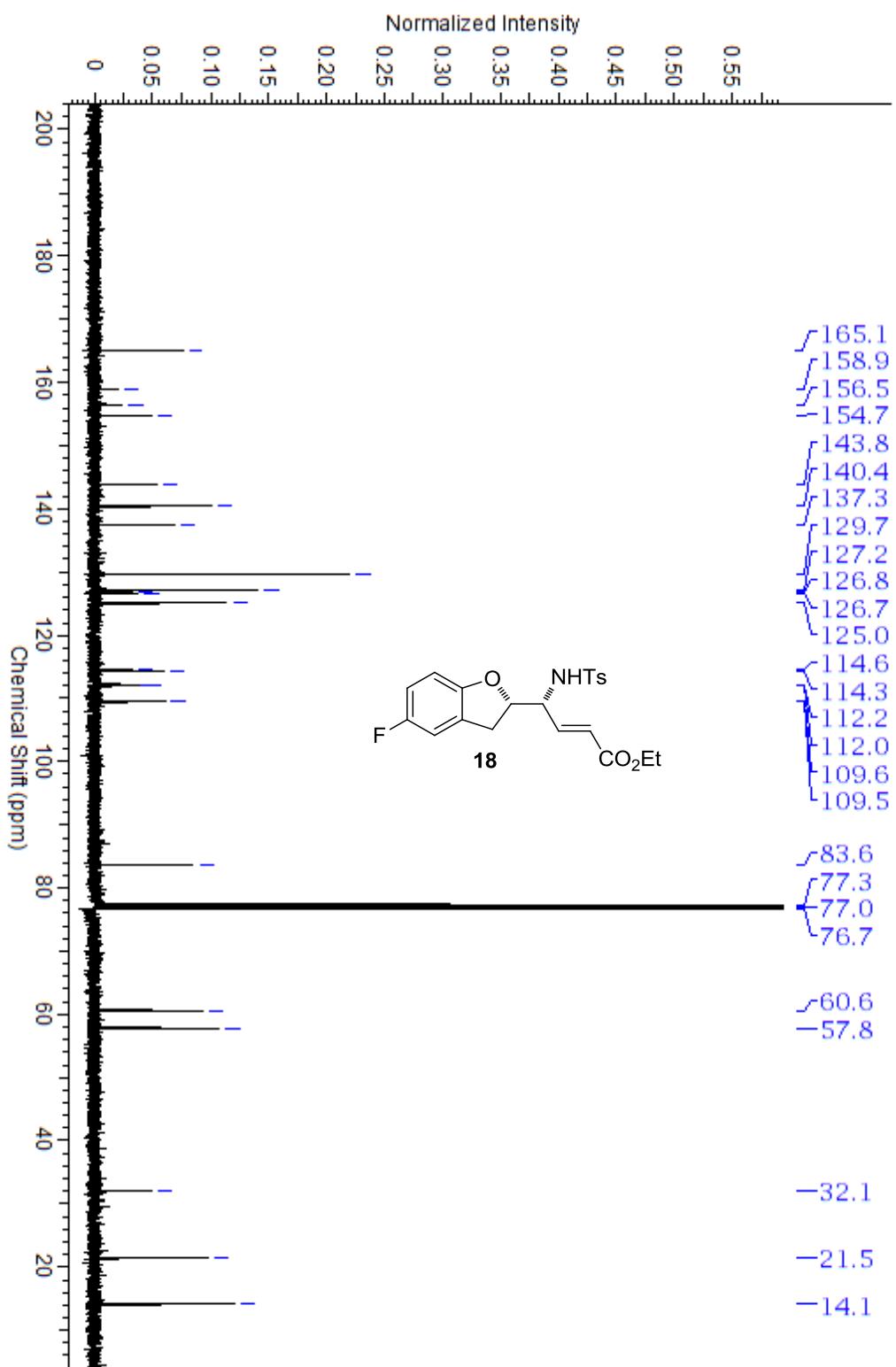
¹H NMR (400 MHz, CDCl₃) of 17



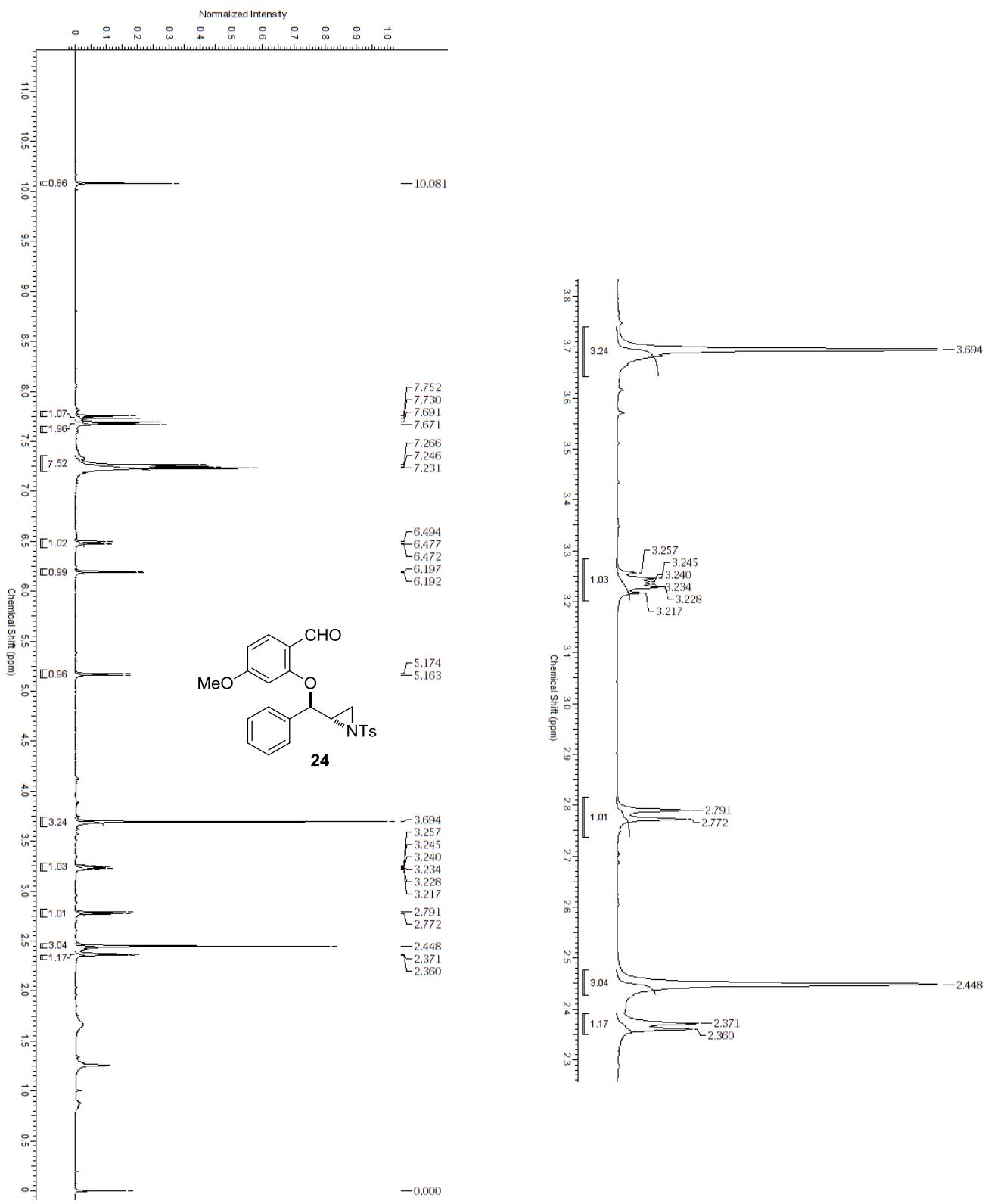
¹³C NMR (100 MHz, CDCl₃) of 17



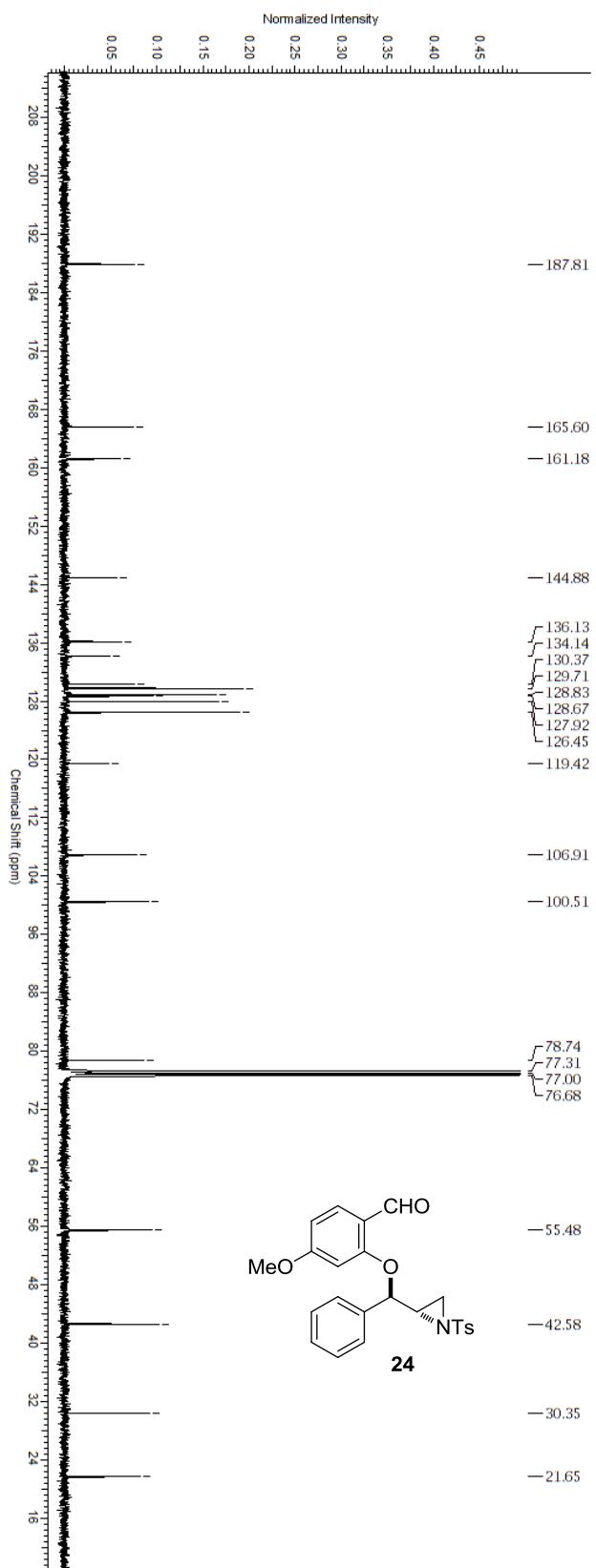
¹H NMR (400 MHz, CDCl₃) of 18



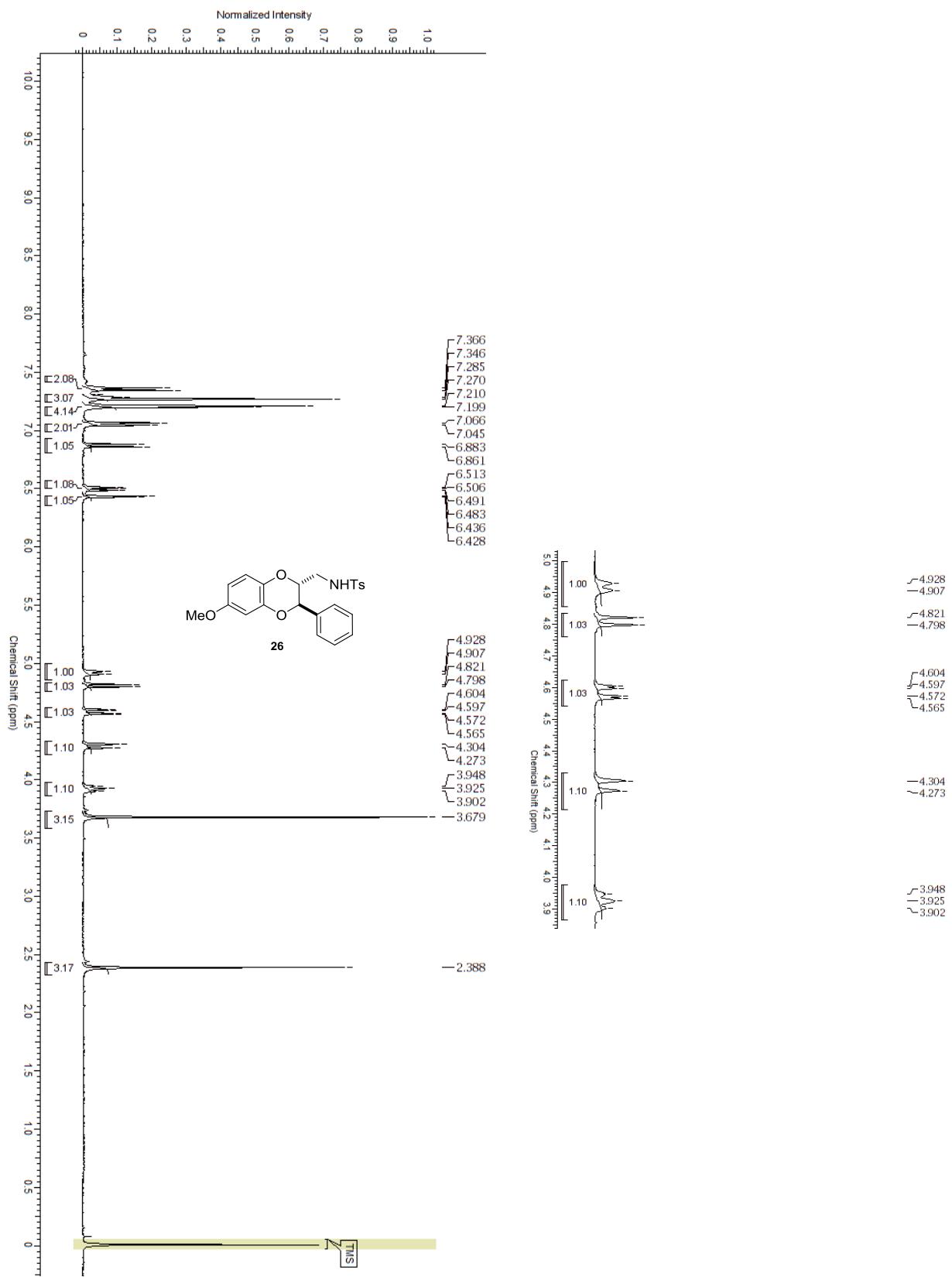
¹³C NMR (100 MHz, CDCl₃) of **18**



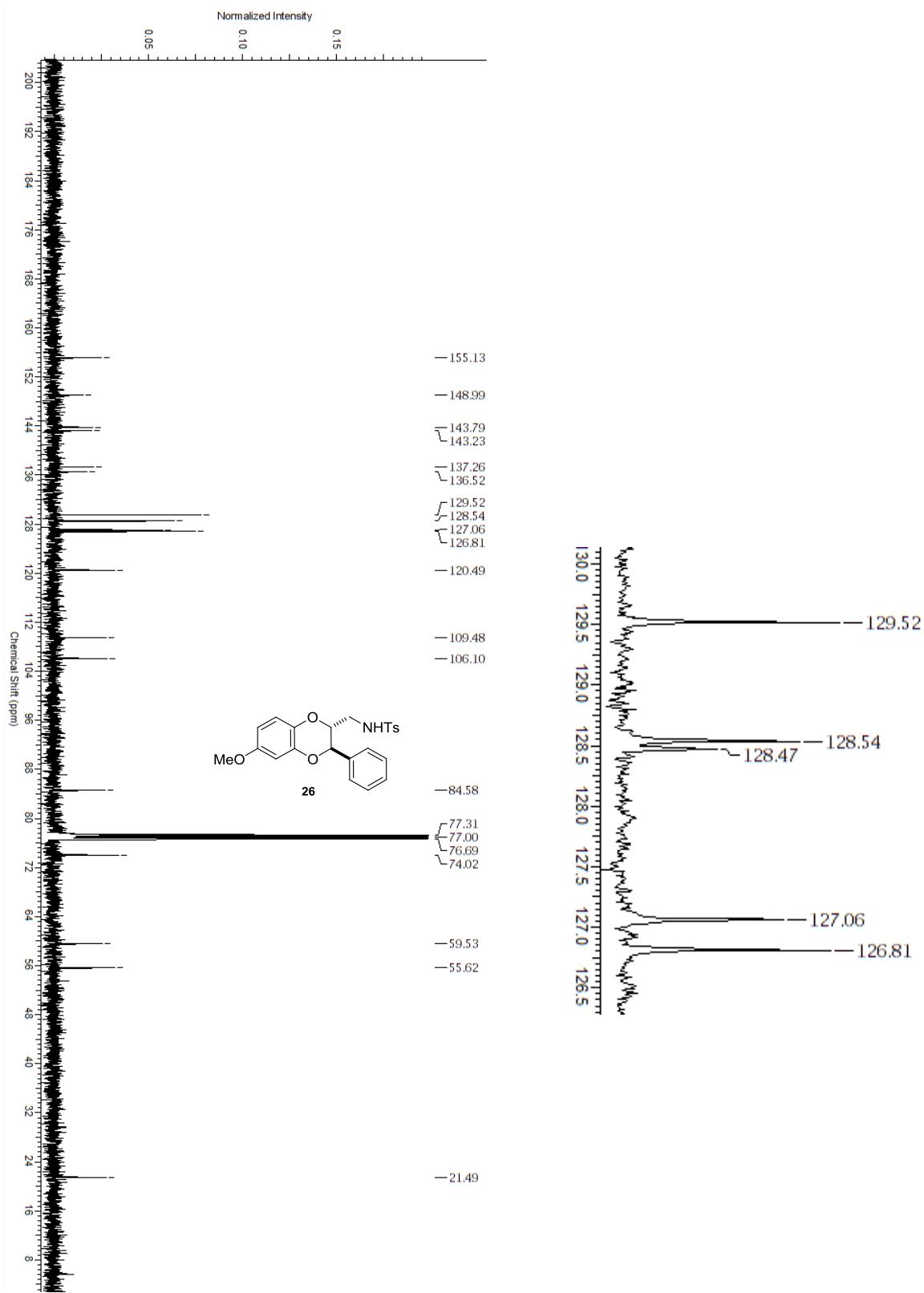
¹H NMR (400 MHz, CDCl₃) spectrum of 24



^{13}C NMR (100 MHz, CDCl_3) spectrum of 24



¹H NMR (400 MHz, CDCl₃) spectrum of 26



¹³C NMR (100 MHz, CDCl₃) spectrum of 26