

Supporting Information

Total Synthesis and Structural Revision of (±)-Nidemone

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Comparison of ^1H NMR data for natural nidemone, synthetic **1, and our previously synthetic nidemone**

Natural nidemone (300 MHz, CDCl_3) δ (ppm)	Synthetic 1 (500 MHz, CDCl_3) δ (ppm)	Our previously synthetic nidemone (500 MHz, CDCl_3) δ (ppm)
12.23 (s, 1H)	12.22 (s, 1H)	12.13 (s, 1H)
7.39 (dd, $J = 7.5, 7.5$ Hz, 1H)	7.39 (dd, $J = 8.5, 7.5$ Hz, 1H)	7.42 (dd, $J = 8.5, 7.5$ Hz, 1H)
6.80 (dd, $J = 7.5, 1.0$ Hz, 1H)	6.80 (d, $J = 8.5$ Hz, 1H)	6.84 (d, $J = 8.5$ Hz, 1H)
6.72 (brdd, $J = 7.5, 1.0$ Hz, 1H)	6.71 (d, $J = 7.5$ Hz, 1H)	6.72 (d, $J = 7.5$ Hz, 1H)
5.27 (t, $J = 1.2$ Hz, 1H)	5.26 (s, 1H)	5.49 (s, 1H)
3.91 (s, 3H)	3.91 (s, 3H)	3.88 (s, 3H)
3.48 (brddd, $J = 16.7, 9.4, 5.0$ Hz, 1H)	3.48 (ddd, $J = 17.0, 9.5, 5.0$ Hz, 1H)	3.06–3.02 (m, 2H)
3.29 (dd, $J = 17.5, 1.5$ Hz, 1H)	3.29 (d, $J = 17.5$ Hz, 1H)	
2.87 (ddd, $J = 16.8, 5.8, 5.8$ Hz, 1H)	2.86 (ddd, $J = 17.0, 5.5, 5.5$ Hz, 1H)	2.67 (d, $J = 17.5$ Hz, 1H)
2.48 (dd, $J = 17.5, 1.5$ Hz, 1H)	2.47 (d, $J = 17.5$ Hz, 1H)	2.66–2.60 (m, 1H)
2.47 (ddd, $J = 14.0, 7.4, 4.0$ Hz, 1H)	2.46 (ddd, $J = 14.0, 5.5, 5.5$ Hz, 1H)	2.57 (d, $J = 17.5$ Hz, 1H)
2.20 (ddd, $J = 13.8, 9.3, 4.8$ Hz, 1H)	2.20 (ddd, $J = 14.0, 9.5, 5.0$ Hz, 1H)	2.04 (ddd, $J = 13.5, 5.1, 5.1$ Hz, 1H)

Comparison of ^{13}C NMR data for natural nidemone, synthetic **1, and our previously synthetic nidemone**

Natural nidemone (75 MHz, CDCl_3) δ (ppm)	Synthetic 1 (125 MHz, CDCl_3) δ (ppm)	Our previously synthetic nidemone (125 MHz, CDCl_3) δ (ppm)
202.7 (C)	202.7 (C)	202.8 (C)
202.3 (C)	202.2 (C)	201.9 (C)
189.6 (C)	189.6 (C)	190.8 (C)
163.4 (C)	163.4 (C)	163.9 (C)
145.4 (C)	145.3 (C)	144.6 (C)
137.0 (CH)	137.0 (CH)	137.5 (CH)
118.9 (CH)	118.9 (CH)	119.2 (CH)
116.4 (C)	116.4 (C)	116.4 (CH)
115.8 (CH)	115.8(CH)	116.4 (C)
101.9 (CH)	101.9 (CH)	105.8 (CH)
59.2 (C)	59.2 (C)	59.6 (CH_3)
59.1 (CH_3)	59.1 (CH_3)	55.7 (C)
39.7 (CH_2)	39.7 (CH_2)	44.9 (CH_2)
32.1 (CH_2)	32.1 (CH_2)	32.1 (CH_2)
25.6 (CH_2)	25.6 (CH_2)	26.5 (CH_2)

Table 1. Crystal data and structure refinement for LCY14.

Identification code	lcy14
Empirical formula	C H O
Formula weight	29.02
Temperature	173(2) K
Wavelength	0.71073 Å
Crystal system, space group	Monoclinic , Pbc _a
Unit cell dimensions	a = 10.287(2) Å alpha = 90 deg. b = 11.291(2) Å beta = 90 deg. c = 20.866(4) Å gamma = 90 deg.
Volume	2423.6(9) Å ³
Z, Calculated density	71, 1.412 Mg/m ³
Absorption coefficient	0.131 mm ⁻¹
F(000)	1065
Crystal size	0.38 x 0.16 x 0.15 mm ³
Theta range for data collection	1.95 to 28.33 deg.
Limiting indices	-13<= <i>h</i> <=13, -15<= <i>k</i> <=14, -27<= <i>l</i> <=27
Reflections collected / unique	26046 / 2965 [R(int) = 0.0253]
Completeness to theta = 28.33	98.3 %
Refinement method	Full-matrix least-squares on F ²

Data / restraints / parameters	2965 / 0 / 228
Goodness-of-fit on F^2	1.098
Final R indices [$I > 2\sigma(I)$]	R1 = 0.0406, wR2 = 0.1169
R indices (all data)	R1 = 0.0506, wR2 = 0.1324
Largest diff. peak and hole	0.340 and -0.300 e.Å ⁻³

Table 2. Atomic coordinates ($\times 10^4$) and equivalent isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for LCY14.

U(eq) is defined as one third of the trace of the orthogonalized U_{ij} tensor.

	x	y	z	U(eq)
C(10)	1216(1)	1654(1)	4415(1)	22(1)
C(5)	926(1)	1646(1)	3729(1)	24(1)
C(13)	3043(1)	1760(1)	5702(1)	24(1)
C(12)	3781(1)	1104(1)	5300(1)	25(1)
C(9)	2373(1)	2370(1)	4658(1)	23(1)
C(14)	2216(1)	2658(1)	5373(1)	28(1)
C(11)	3530(1)	1474(1)	4650(1)	22(1)
C(6)	1603(1)	2388(1)	3295(1)	30(1)
C(8)	2595(1)	3472(1)	4242(1)	31(1)
C(4)	-58(1)	880(1)	3495(1)	30(1)
C(7)	2656(2)	3210(1)	3527(1)	36(1)
C(3)	-351(2)	861(2)	2843(1)	42(1)
C(1)	1285(2)	2341(2)	2650(1)	42(1)
C(2)	321(2)	1590(2)	2431(1)	46(1)
O(1)	2984(1)	1728(1)	6335(1)	32(1)
C(15)	3704(2)	787(2)	6637(1)	37(1)

O(3)	579(1)	1060(1)	4797(1)	30(1)
O(2)	4080(1)	1141(1)	4162(1)	30(1)
O(4)	-736(1)	142(1)	3877(1)	39(1)

Table 3. Bond lengths [Å] and angles [deg] for LCY14.

C(10)-O(3)	1.2307(15)
C(10)-C(5)	1.4637(17)
C(10)-C(9)	1.5243(16)
C(5)-C(6)	1.4159(18)
C(5)-C(4)	1.4180(18)
C(13)-O(1)	1.3227(15)
C(13)-C(12)	1.3516(17)
C(13)-C(14)	1.4915(18)
C(12)-C(11)	1.4439(17)
C(9)-C(8)	1.5331(17)
C(9)-C(14)	1.5354(17)
C(9)-C(11)	1.5623(16)
C(11)-O(2)	1.2232(15)
C(6)-C(1)	1.386(2)
C(6)-C(7)	1.506(2)
C(8)-C(7)	1.522(2)
C(4)-O(4)	1.3476(18)
C(4)-C(3)	1.394(2)
C(3)-C(2)	1.377(3)
C(1)-C(2)	1.383(3)
O(1)-C(15)	1.4403(18)
O(3)-C(10)-C(5)	121.46(11)
O(3)-C(10)-C(9)	119.34(11)
C(5)-C(10)-C(9)	119.14(10)
C(6)-C(5)-C(4)	119.52(12)
C(6)-C(5)-C(10)	121.41(11)
C(4)-C(5)-C(10)	119.08(11)
O(1)-C(13)-C(12)	129.05(12)

O(1)-C(13)-C(14)	116.91(11)
C(12)-C(13)-C(14)	114.02(11)
C(13)-C(12)-C(11)	108.91(11)
C(10)-C(9)-C(8)	111.07(10)
C(10)-C(9)-C(14)	110.64(10)
C(8)-C(9)-C(14)	113.14(10)
C(10)-C(9)-C(11)	104.36(9)
C(8)-C(9)-C(11)	113.92(10)
C(14)-C(9)-C(11)	103.13(9)
C(13)-C(14)-C(9)	104.13(10)
O(2)-C(11)-C(12)	127.63(11)
O(2)-C(11)-C(9)	124.05(11)
C(12)-C(11)-C(9)	108.29(10)
C(1)-C(6)-C(5)	118.75(14)
C(1)-C(6)-C(7)	120.39(13)
C(5)-C(6)-C(7)	120.86(12)
C(7)-C(8)-C(9)	113.72(11)
O(4)-C(4)-C(3)	117.14(13)
O(4)-C(4)-C(5)	122.89(12)
C(3)-C(4)-C(5)	119.96(14)
C(6)-C(7)-C(8)	113.91(11)
C(2)-C(3)-C(4)	119.50(15)
C(2)-C(1)-C(6)	120.99(15)
C(3)-C(2)-C(1)	121.28(14)
C(13)-O(1)-C(15)	115.73(10)

Symmetry transformations used to generate equivalent atoms:

Table 4. Anisotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for LCY14.

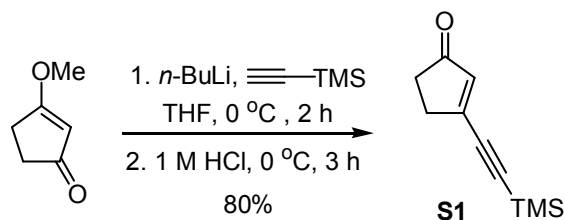
The anisotropic displacement factor exponent takes the form:

$$-2 \pi^2 [h^2 a^{*2} U_{11} + \dots + 2 h k a^* b^* U_{12}]$$

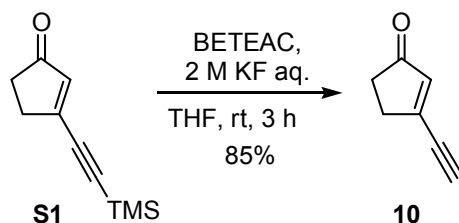
	U11	U22	U33	U23	U13	U12
C(10)	19(1)	19(1)	26(1)	0(1)	1(1)	2(1)
C(5)	23(1)	26(1)	25(1)	-1(1)	1(1)	4(1)
C(13)	20(1)	27(1)	26(1)	-5(1)	-2(1)	-3(1)
C(12)	20(1)	26(1)	29(1)	-2(1)	-1(1)	2(1)
C(9)	21(1)	20(1)	27(1)	-2(1)	-1(1)	1(1)
C(14)	26(1)	26(1)	30(1)	-9(1)	-2(1)	4(1)
C(11)	18(1)	19(1)	29(1)	-2(1)	1(1)	-2(1)
C(6)	33(1)	29(1)	28(1)	4(1)	5(1)	8(1)
C(8)	30(1)	21(1)	43(1)	3(1)	0(1)	-3(1)
C(4)	26(1)	32(1)	33(1)	-6(1)	-3(1)	5(1)
C(7)	36(1)	31(1)	40(1)	12(1)	7(1)	-2(1)
C(3)	42(1)	48(1)	36(1)	-14(1)	-11(1)	8(1)
C(1)	53(1)	44(1)	27(1)	6(1)	7(1)	14(1)
C(2)	58(1)	54(1)	26(1)	-7(1)	-7(1)	21(1)
O(1)	31(1)	38(1)	25(1)	-6(1)	-2(1)	2(1)
C(15)	34(1)	48(1)	28(1)	2(1)	-4(1)	2(1)
O(3)	28(1)	32(1)	29(1)	5(1)	2(1)	-7(1)
O(2)	28(1)	31(1)	30(1)	-2(1)	6(1)	4(1)
O(4)	29(1)	43(1)	43(1)	-4(1)	-5(1)	-11(1)

Table 5. Hydrogen coordinates ($\times 10^4$) and isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for LCY14.

	x	y	z	U(eq)
H(1)	3337(17)	39(17)	6507(9)	41(4)
H(2)	3585(19)	898(18)	7093(10)	49(5)
H(3)	4640(20)	808(18)	6538(10)	52(5)
H(4)	4404(15)	496(14)	5404(8)	30(4)
H(6)	1315(18)	2609(16)	5513(8)	41(5)
H(5)	2529(18)	3448(17)	5480(8)	40(5)
H(12)	2572(19)	3952(17)	3304(9)	46(5)
H(11)	3524(17)	2862(16)	3429(9)	40(4)
H(14)	1830(16)	4024(15)	4330(8)	36(4)
H(10)	1749(19)	2859(18)	2359(9)	49(5)
H(13)	3402(17)	3876(16)	4392(8)	38(4)
H(9)	130(20)	1580(20)	1991(11)	64(6)
H(7)	-410(20)	290(20)	4281(12)	65(6)
H(8)	-1030(20)	354(18)	2700(9)	51(5)



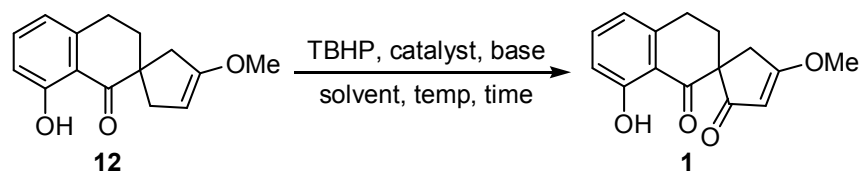
3-(Trimethylsilylethynyl)-2-cyclopenten-1-one (S1). To a stirred solution of trimethylsilylacetylene (336 mg, 3.42 mmol) in THF (3 mL) under Ar atmosphere at 0 °C was added *n*-BuLi (1.6 M in hexanes, 2.0 mL, 3.20 mmol) and stirred at 0 °C for 1 h. 3-Methoxy-2-cyclopenten-1-one (300 mg, 2.68 mmol) was added to the reaction mixture and this mixture was stirred at 0 °C for 2 h. The mixture was quenched with 1 M HCl at 0 °C and stirred at this temperature for 3 h. The aqueous solution was extracted with EtOAc, and the combined extracts were washed with brine, dried over MgSO₄, filtered and concentrated. The residue was purified by silica-gel chromatography (hexane/EtOAc = 6:1) to afford **S1** (382 mg, 80%) as a yellowish oil. IR (neat) ν 2961, 2150, 1709, 1582, 1267, 848 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.27 (t, *J* = 1.8 Hz, 1H), 2.74–2.70 (m, 2H), 2.42–2.39 (m, 2H), 0.23 (s, 9H); ¹³C NMR (100 MHz, CDCl₃): δ 209.6, 156.7, 136.8, 111.7, 99.7, 34.7, 32.5, –0.5; MS (EI) *m/z* (% base peak) 178 (M⁺, 22), 163 (100), 133 (1), 119 (1), 107 (8), 97 (2), 83 (4), 77 (3), 75 (3), 53 (2); HRMS (EI) calcd for C₁₀H₁₄OSi 178.0814, Found 178.0815.



3-Ethynyl-2-cyclopenten-1-one (10). A mixture of **S1** (600 mg, 3.36 mmol) and benzyltriethylammonium chloride (BTEAC, 77 mg, 0.34 mmol) in THF (15 mL) at room temperature was added 2 M KF (3.5 mL). The reaction mixture was stirred at room temperature for 3 h and extracted with Et₂O. The combined extracts were washed with brine, dried over MgSO₄, filtered and concentrated.

The crude product was purified by column chromatography on silica gel (Et₂O/hexanes = 1:5) to afford **10** (304 mg, 85%) as a colorless solid. Mp 55–56 °C; IR (neat) ν 3194, 2926, 2854, 2095, 1701, 1664, 1578, 1435, 1285, 1177, 872, 729 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.35 (t, *J* = 1.6 Hz, 1H), 3.84 (s, 1H), 2.77–2.74 (m, 2H), 2.45–2.42 (m, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 209.2, 155.8, 137.8, 92.2, 79.0, 34.6, 32.3; MS (EI) *m/z* (% base peak) 106 (M⁺, 100), 105 (12), 78 (47), 77 (20), 52 (15), 51 (13); HRMS (ESI) calcd for C₇H₆O 106.0419, Found 106.0420.

Table S1. Attempts to the regioselective allylic oxidation of 12



entry	catalyst	base	solvent	temp	time	1 (yield)
1	Rh(cap) ₄	K ₂ CO ₃	CH ₂ Cl ₂	0 °C to rt	3 h	— ^a
2	Rh(cap) ₄	K ₂ CO ₃	CH ₂ Cl ₂	0 °C to rt	3 h	10% ^b
3	Pd(OH) ₂	K ₂ CO ₃	CH ₂ Cl ₂	0 °C	1 d	— ^a
4	Pd(OH) ₂	Cs ₂ CO ₃	CH ₂ Cl ₂	0 °C	1 d	— ^a
5	PhI(OAc) ₂	—	EtOAc	0 °C to rt	3 h	7% ^b
6	PhI(OAc) ₂	K ₂ CO ₃	EtOAc	0 °C to rt	2 h	18% ^b
7	PhI(OAc) ₂	Cs ₂ CO ₃	EtOAc	0 °C to rt	2 h	14% ^b
8	PhI(OAc) ₂	K ₂ CO ₃	EtOAc	0 °C	5 h	22% ^b
9	PhI(OAc) ₂	K ₂ CO ₃	EtOAc	−10 °C	1 d	— ^c
10	PhI(OAc) ₂	Na ₂ CO ₃	EtOAc	0 °C	6 h	27% ^b
11	Mn(OAc) ₃	—	EtOAc	rt	1 d	— ^a
12	Mn(OAc) ₃	—	EtOAc	rt	1 d	— ^{a,b}

^aComplicated mixture of products.

^bAdded 4A molecular sieve.

^cRecovery of starting material.

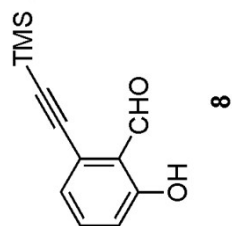
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 AQ 1.3664756 sec
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 DW 83.400 usec
 DE 6.50 usec
 TE 300.0 K
 D1 1.50000000 sec

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 LB 0.10 Hz
 GB 0
 PC 1.00

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 F1 5001.62 Hz
 F2P -0.500 ppm
 F2 -200.07 Hz
 PPMCM 0.60465 ppm/cm
 HZCM 241.93907 Hz/cm



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10.426

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Integral

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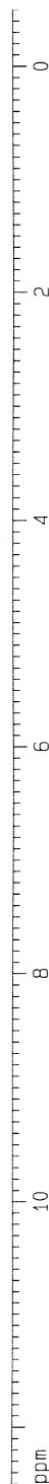
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1.0526

1.0236

9.9127



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119.53
124.71
127.15

136.36

162.20

196.85



ppm

Current Data Parameters
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PROCNO 1

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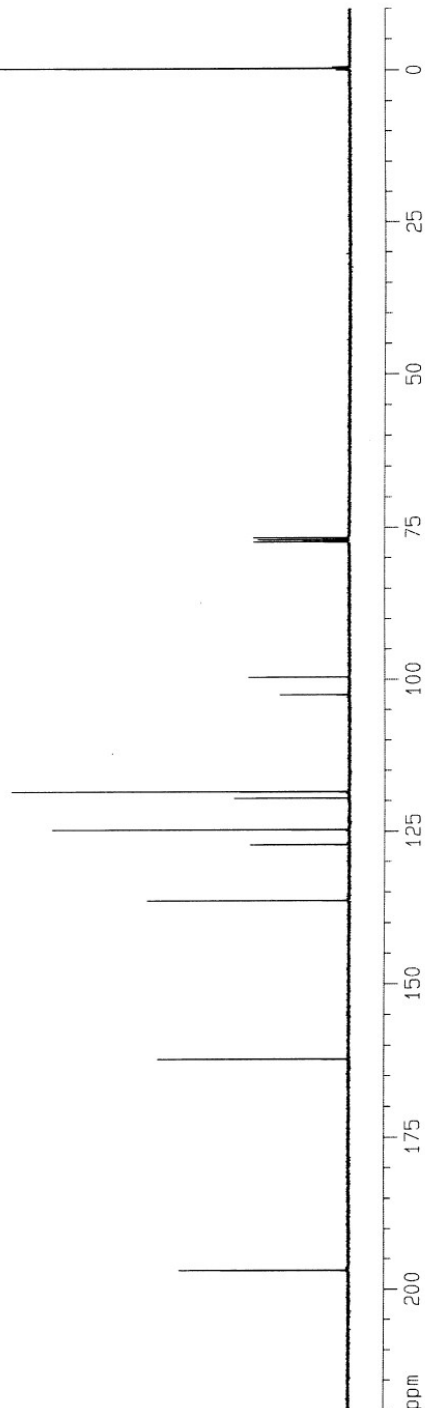
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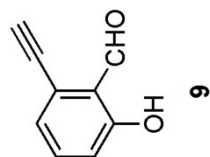
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1D NMR plot parameters

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F1 22134.81 Hz
F2P -10.000 ppm
F2 -1006.13 Hz
PPMCM 11.50000 ppm/cm
HZCM 1157.04688 Hz/cm





1.581

3.438

7.471
7.451
7.431
7.132
7.113
6.993
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10.451

11.664

ppm

Current Data Parameters
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PROCNO 1

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AQ 1.3664756 sec
RG 4597.6
DW 83.400 usec
DE 6.50 usec
TE 300.0 K
D1 1.5000000 sec

===== CHANNEL f1 =====

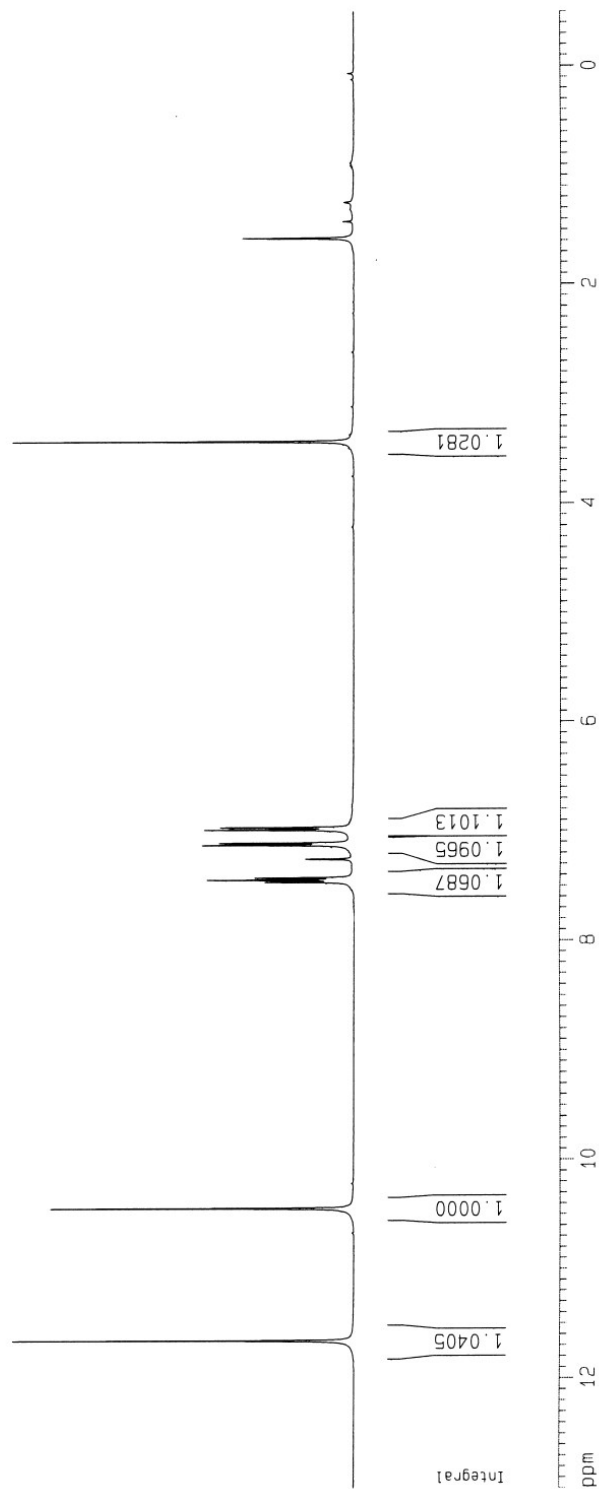
NUC1 1H
P1 11.90 usec
PL1 -3.00 dB
SF01 400.1326008 MHz

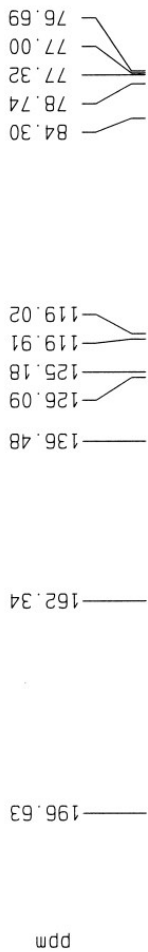
F2 - Processing parameters

SI 8192
SF 400.130099 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.00

1D NMR plot parameters

CX 21.50 cm
F1P 13.000 ppm
F1 5201.69 Hz
F2P -0.500 ppm
F2 -200.07 Hz
PPMCM 0.62791 ppm/cm
HZCM 251.24442 Hz/cm





Current Data Parameters
 NAME LCY-05
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters

Date_ 20151115
 Time 18.24
 INSTRUM spect
 PROBHD 5 mm GNP 1H
 PULPROG zgpg30
 TO 65536
 SOLVENT CDCl3
 NS 768
 DS 4
 SWH 25125.629 Hz
 FIDRES 0.383387 Hz
 AQ 1.3042164 sec
 RG 256
 DW 19.900 usec
 DE 6.50 usec
 TE 300.0 K
 D1 2.00000000 sec
 d11 0.03000000 sec
 d12 0.0002000 sec

===== CHANNEL f1 =====

NUC1 13C
 P1 10.20 usec
 PL1 0.00 dB
 SF01 100.6237959 MHz

===== CHANNEL f2 =====

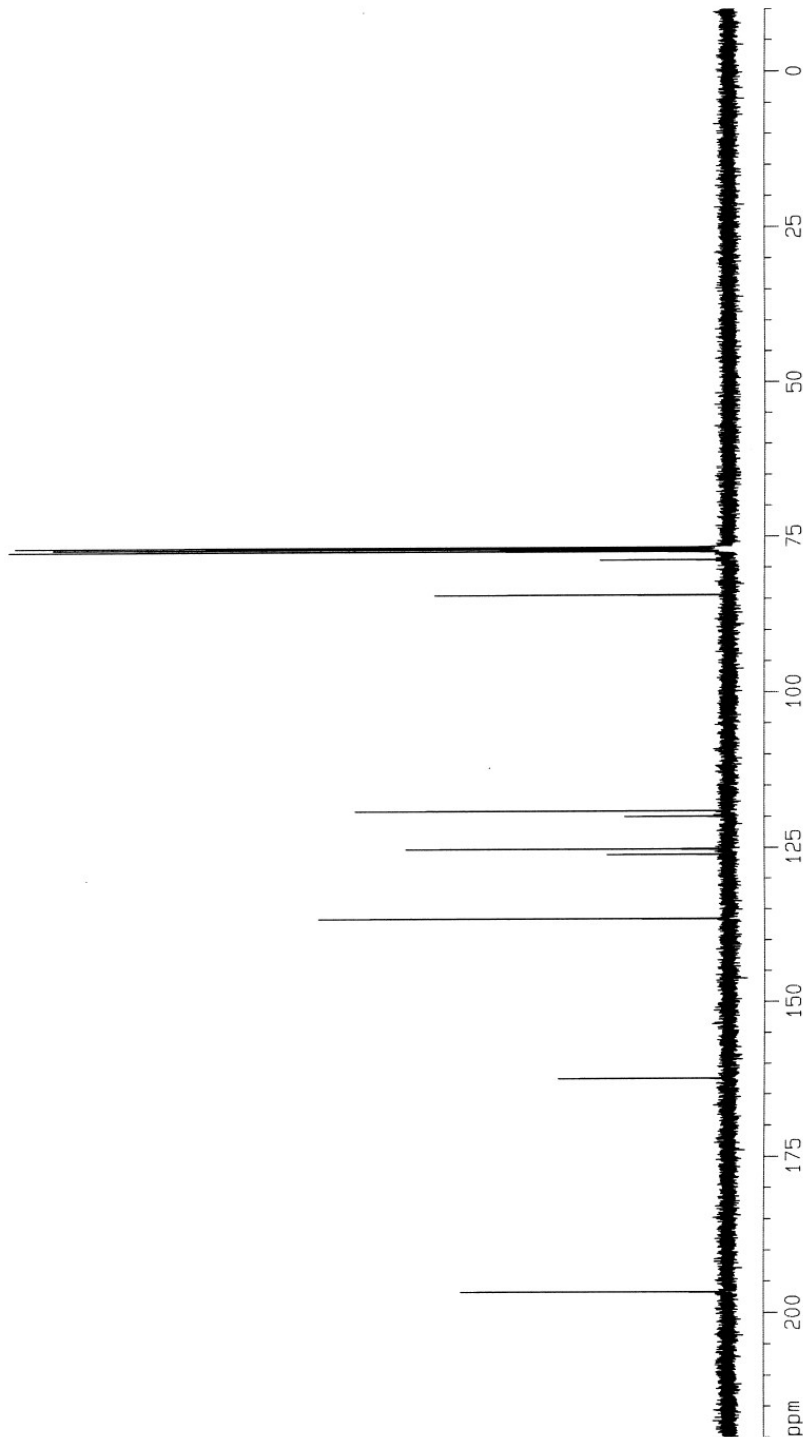
CPDPRG2 waltz16
 NUC2 1H
 PCPD2 90.00 usec
 PL2 -3.00 dB
 PL12 14.50 dB
 PL13 17.50 dB
 SF02 400.1326008 MHz

F2 - Processing parameters

SI 32768
 SF 100.6127700 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40

1D NMR plot parameters

CX 20.00 cm
 FIP 220.000 ppm
 F1 22134.81 Hz
 F2P -10.000 ppm
 F2 -1006.13 Hz
 PPMCM 11.50000 ppm/cm
 HZCM 1157.04688 Hz/cm



Current Data Parameters
NAME Icy-9
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters

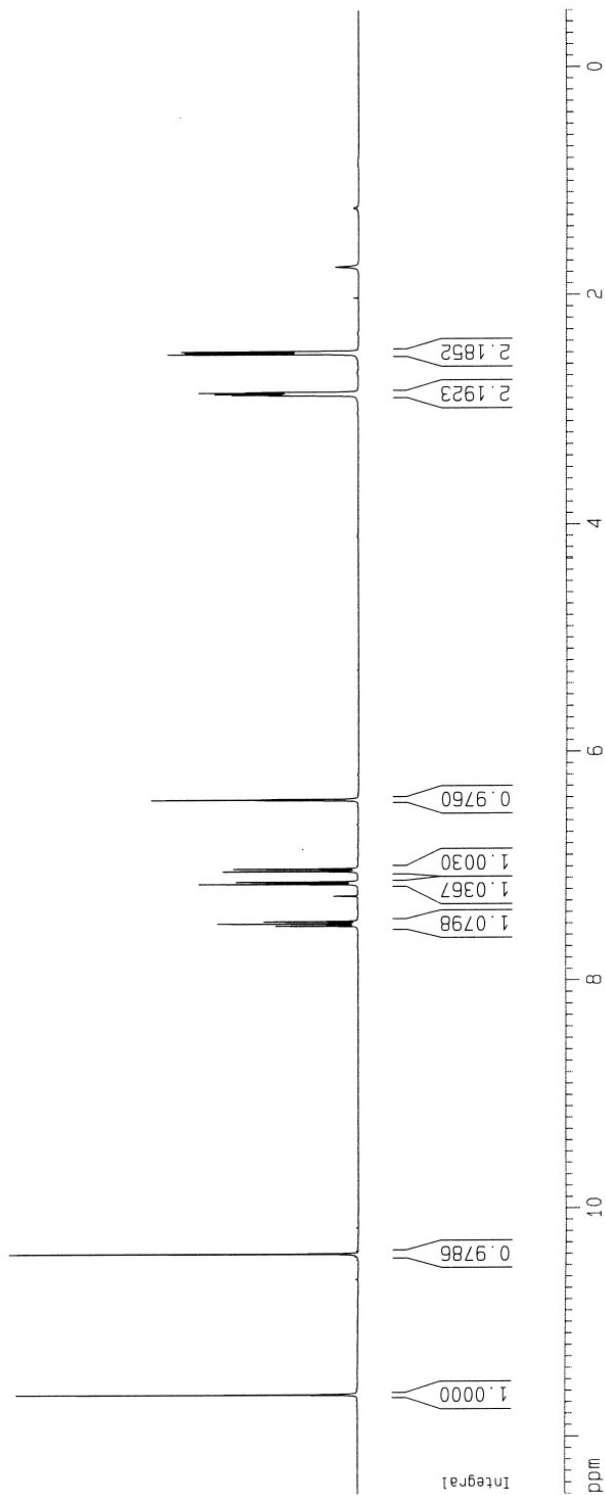
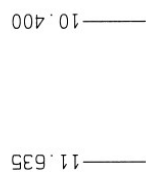
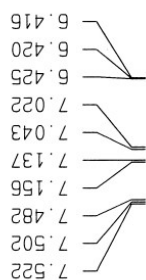
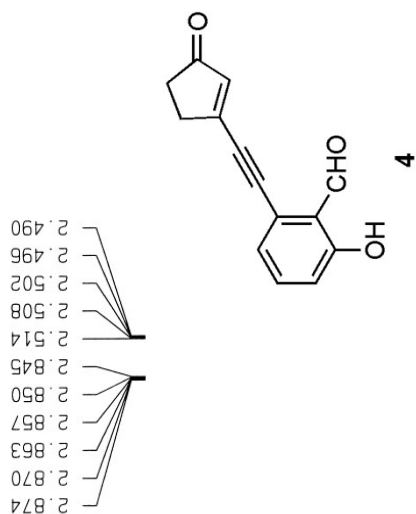
Date_ 20160305
Time 19.30
INSTRUM spect
PROBHD 5 mm QNP 1H
PULPROG zg30
TD 16384
SOLVENT CDCl3
NS 16
DS 0
SWH 5995.204 Hz
FIDRES 0.365918 Hz
AQ 1.3664756 sec
RG 2298.8
DM 83.400 usec
DE 6.50 usec
TE 300.0 K
D1 1.50000000 sec

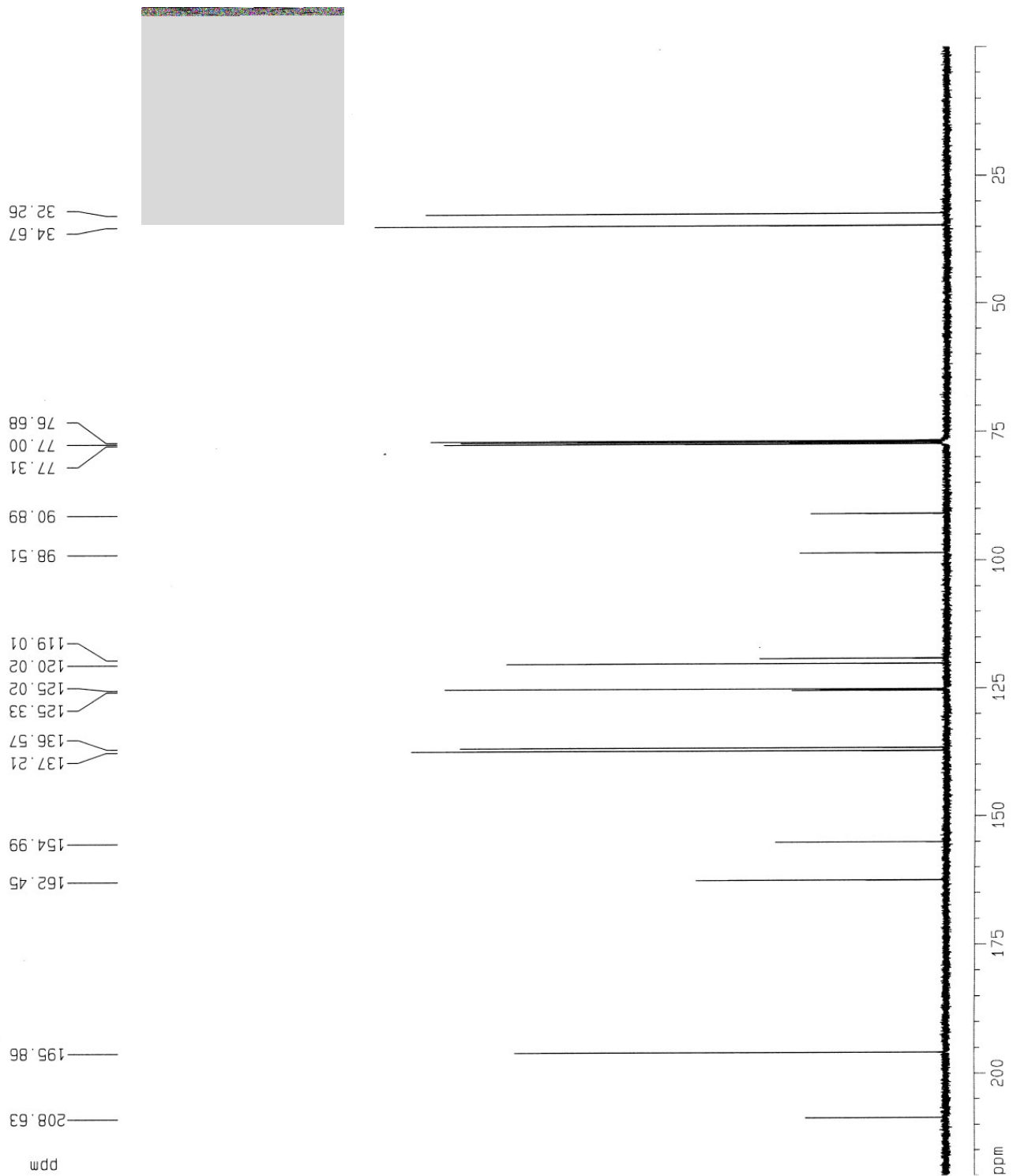
===== CHANNEL f1 =====

NUC1 1H
P1 11.90 usec
PL1 -3.00 dB
SF01 400.1326008 MHz

F2 - Processing parameters

SI 8192
SF 400.1300091 MHz
WDW EM
SSB 0
LB 0.10 Hz
GB 0
PC 1.00
1D NMR plot parameters
CX 21.50 cm
F1P 12.500 ppm
F1 5001.62 Hz
F2P -0.500 ppm
F2 -200.07 Hz
PPMCM 0.60465 ppm/cm
HZCM 241.93907 Hz/cm





```

Current Data Parameters
NAME          lcy-g
EXPNO         2
PROCNO        1

F2 - Acquisition Parameters
Date_         20160305
Time          20.27
INSTRUM       spect
PROBHD        5 mm GNP 1H
PULPROG       zgpg30
TD            65536
SOLVENT       CDCl3
NS            1024
DS            4
SWH           25125.629 Hz
FIDRES        0.363387 Hz
AQ            1.3042164 sec
RG            256
DE            19.900 usec
TE            300.0 K
D1            2.00000000 sec
d11           0.03000000 sec
d12           0.00002000 sec

===== CHANNEL f1 =====
NUC1          13C
P1            10.20 usec
PL1           0.00 dB
SF01          100.6237959 MHz

===== CHANNEL f2 =====
CPDPRG2       waltz16
NUC2          1H
PCPD2         90.00 usec
PL2           -3.00 dB
PL12          14.50 dB
PL13          17.50 dB
SF02          400.1326008 MHz

F2 - Processing parameters
SI            32768
SF            100.6127754 MHz
WDW           EM
SSB           0
LB            0.30 Hz
GB            0
PC            1.40

1D NMR plot parameters
CX            20.00 cm
F1P           220.000 ppm
F1            22134.81 Hz
F2P           0.000 ppm
F2            0.00 Hz
PPMCM         11.00000 ppm/cm
HZCM          1106.74048 Hz/cm
  
```

Current Data Parameters
 NAME LYC-10
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters

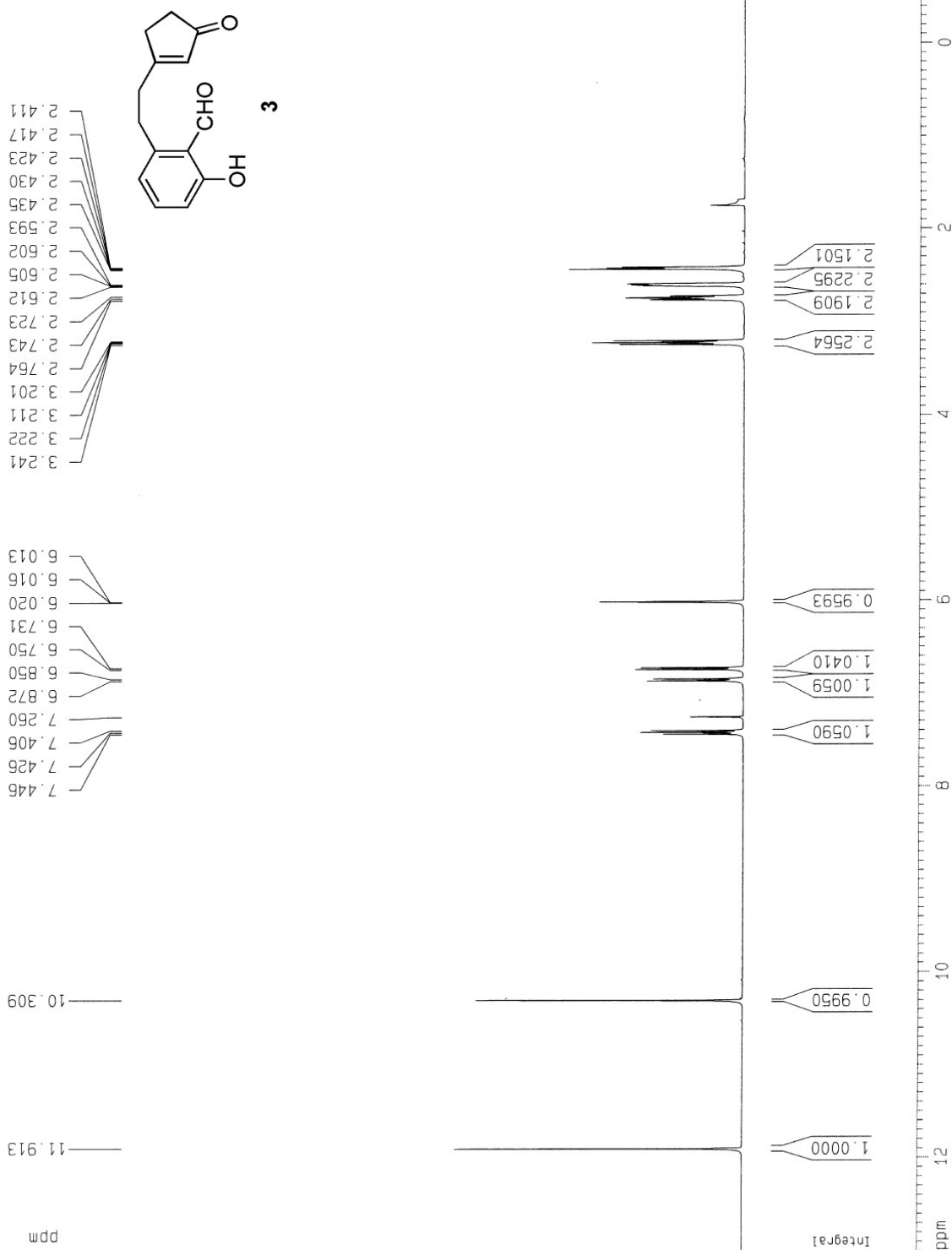
Date_ 20160705
 Time 21.32
 INSTRUM spect
 PROBD 5 mm QNP 1H
 PULPROG zg30
 TD 16384
 SOLVENT CDCl3
 NS 16
 DS 0
 SWH 5995.204 Hz
 FIDRES 0.365918 Hz
 AQ 1.3664756 sec
 RG 228.1
 DW 83.400 usec
 DE 6.50 usec
 TE 300.0 K
 D1 1.50000000 sec

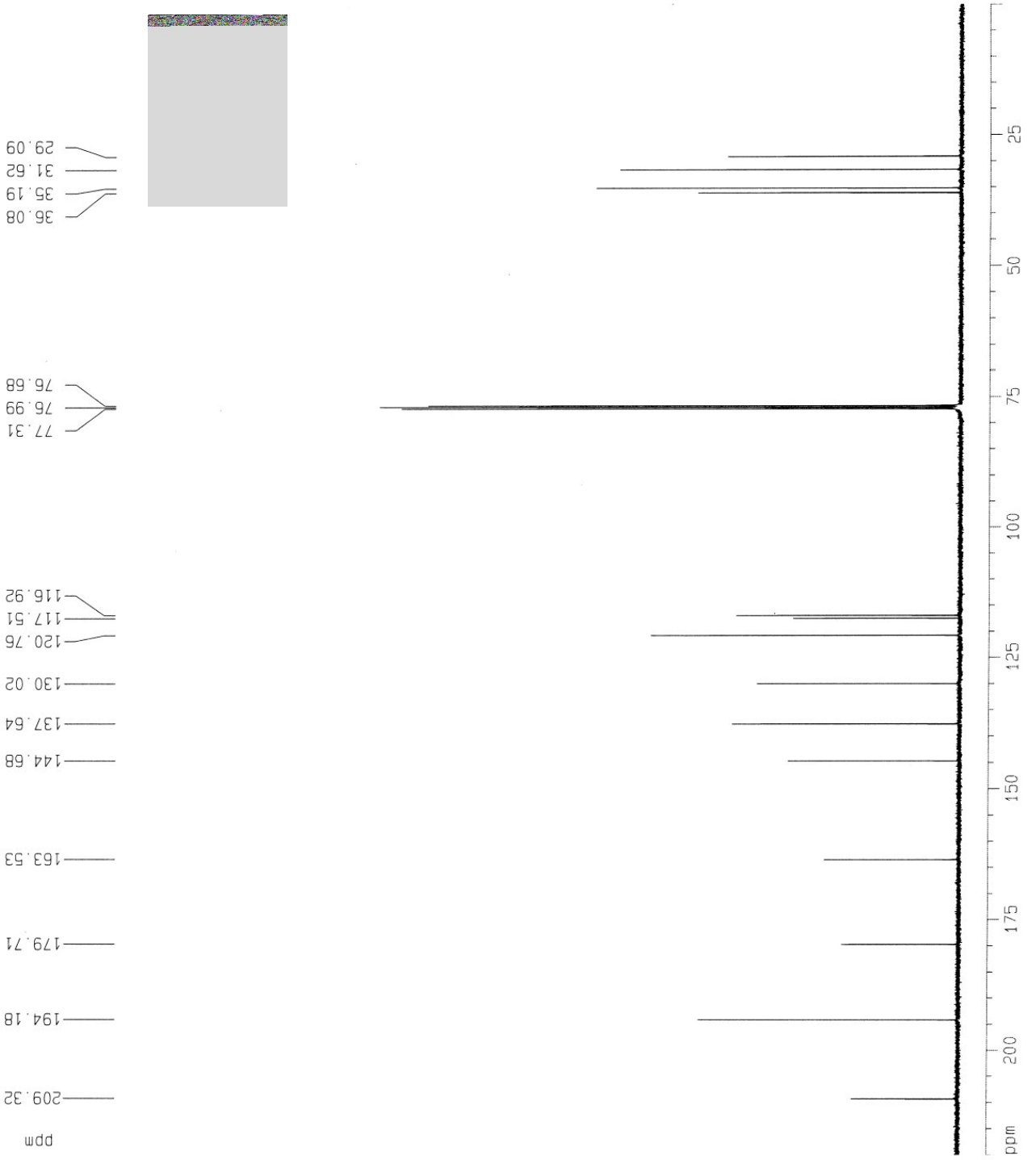
===== CHANNEL f1 =====

NUC1 1H
 P1 14.30 usec
 PL1 -0.30 dB
 SF01 400.1326008 MHz

F2 - Processing parameters

SI 8192
 SF 400.1300091 MHz
 WDW EM
 SSB 0
 LB 0.10 Hz
 GB 0
 PC 1.00
 ID NMR plot parameters
 CX 21.50 cm
 F1P 13.000 ppm
 F1 5201.69 Hz
 F2P -0.500 ppm
 F2 -200.07 Hz
 PPMCM 0.62791 ppm/cm
 HZCM 251.24442 Hz/cm





```

Current Data Parameters
NAME      LYC-10
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20160706
Time      1.43
INSTRUM   spect
PROBHD    5 mm QNP 1H
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         4505
DS         4
SWH        25125.629 Hz
FIDRES     0.383387 Hz
AQ         1.3042164 sec
RG         256
DM         19.900 usec
DE         6.50 usec
TE         300.0 K
D1         2.00000000 sec
d11        0.03000000 sec
d12        0.00002000 sec

===== CHANNEL f1 =====
NUC1       13C
P1         10.20 usec
PL1        0.00 dB
SF01       100.6237959 MHz

===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2       1H
PCPD2      90.00 usec
PL2        -3.00 dB
PL12       14.50 dB
PL13       17.50 dB
SF02       400.1326008 MHz

F2 - Processing parameters
S1         32768
SF         100.6127745 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.40

1D NMR plot parameters
CX         20.00 cm
F1P        220.000 ppm
F1         22134.81 Hz
F2P        0.000 ppm
F2         0.00 Hz
PPMCM      11.00000 ppm/cm
HZCM       1106.74048 Hz/cm

```

Current Data Parameters
 NAME LCY
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters

Date_ 20160308
 Time 23:51
 INSTRUM spect
 PROBHD 5 mm QNP 1H
 PULPROG zg30
 TD 16384
 SOLVENT CDCl3
 NS 16
 DS 0
 SWH 5995.204 Hz
 FIDRES 0.365918 Hz
 AQ 1.3664756 sec
 RG 3649.1
 DW 83.400 usec
 DE 6.50 usec
 TE 300.0 K
 D1 1.5000000 sec

===== CHANNEL f1 =====

NUC1 1H
 P1 11.90 usec
 PL1 -3.00 dB
 SF01 400.1326008 MHz

F2 - Processing parameters

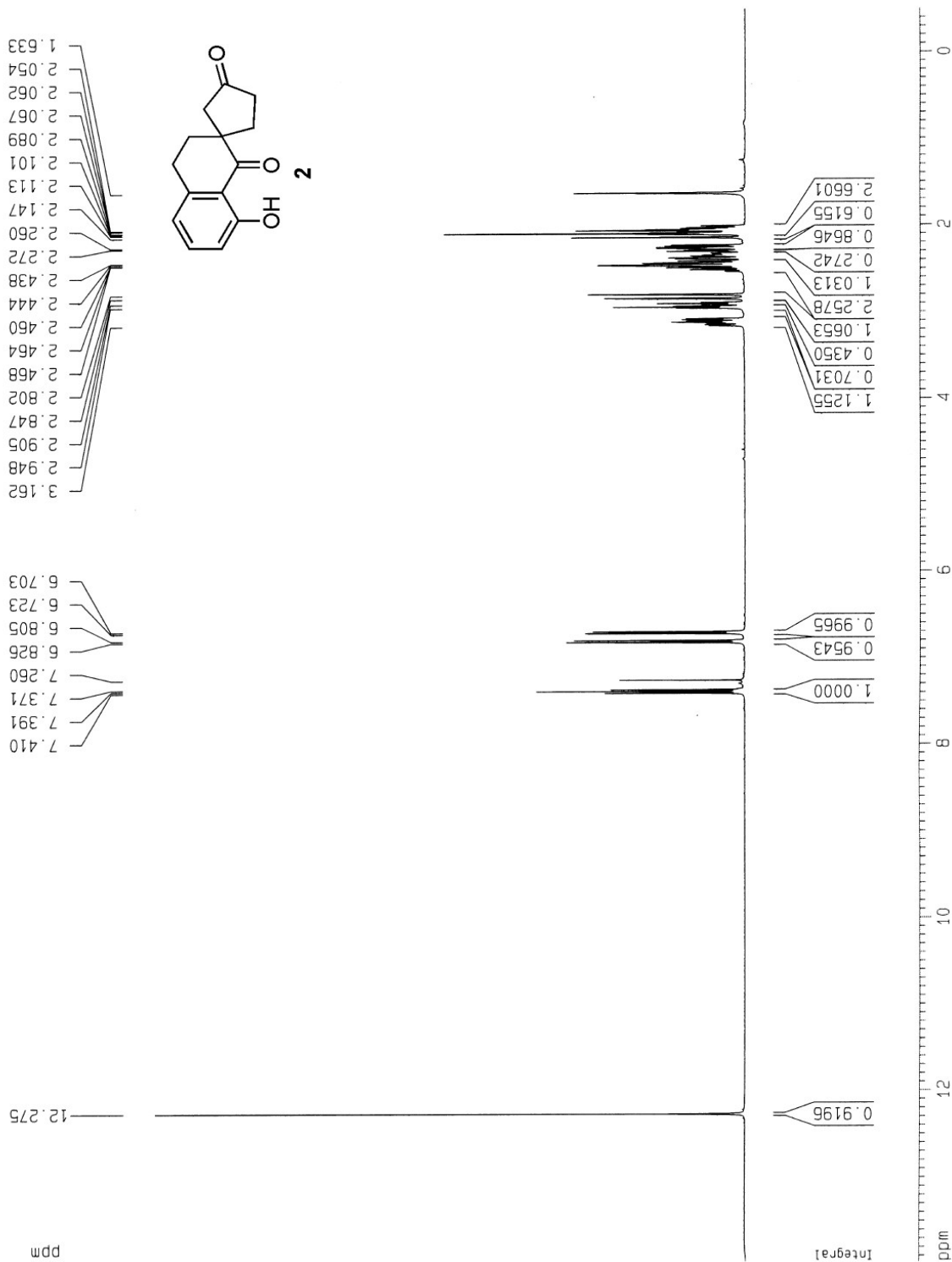
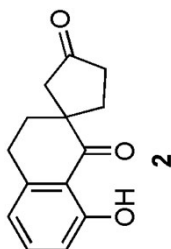
SI 8192
 SF 400.1300091 MHz
 WDW EM
 SSB 0
 LB 0.50 Hz
 GB 0
 PC 1.00

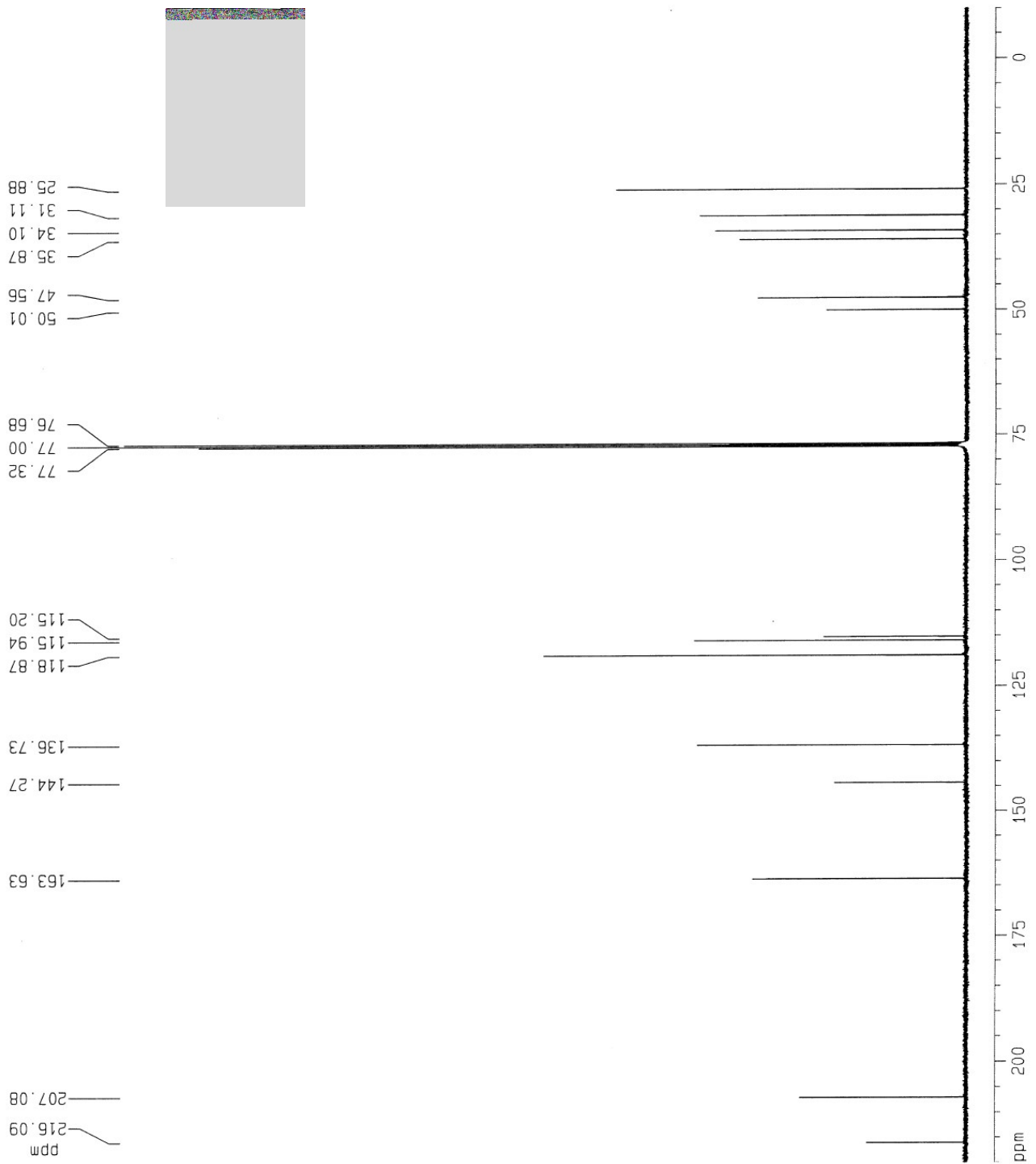
1D NMR plot parameters

CX 21.50 cm
 F1P 14.000 ppm
 F1 5601.82 Hz
 F2P -0.500 ppm
 F2 -200.06 Hz
 PPMCM 0.67442 ppm/cm
 HZCM 269.85510 Hz/cm

3.162
2.948
2.905
2.847
2.802
2.468
2.464
2.460
2.444
2.438
2.272
2.260
2.147
2.113
2.101
2.089
2.067
2.062
2.054
1.633

7.410
7.391
7.371
7.260
6.826
6.805
6.723
6.703





```

Current Data Parameters
NAME          LCY
EXPNO         2
PROCNO        1

F2 - Acquisition Parameters
Date_         20160309
Time          6.34
INSTRUM       spect
PROBHD        5 mm QNP 1H
PULPROG       zgpg30
TD            65536
SOLVENT       CDCl3
NS            7168
DS            4
SWH           25125.629 Hz
FIDRES        0.383387 Hz
AQ            1.3042164 sec
RG            256
DW            19.900 usec
DE            6.50 usec
TE            300.0 K
D1            2.00000000 sec
d11           0.03000000 sec
d12           0.00002000 sec

===== CHANNEL f1 =====
NUC1          13C
P1            10.20 usec
PL1           0.00 dB
SF01          100.6237959 MHz

===== CHANNEL f2 =====
CPDPRG2       waltz16
NUC2          1H
PCPD2         90.00 usec
PL2           -3.00 dB
PL12          14.50 dB
PL13          17.50 dB
SF02          400.1326008 MHz

F2 - Processing parameters
SI            32768
SF            100.6127723 MHz
WDW           EM
SSB           0
LB            0.30 Hz
GB            0
PC            1.40

1D NMR plot parameters
CX            20.00 cm
F1P           220.000 ppm
F1            22134.81 Hz
F2P           -10.000 ppm
F2            -1006.13 Hz
PPMCM         11.50000 ppm/cm
HZCM          1157.04688 Hz/cm

```

Current Data Parameters
 NAME WYC-test
 EXPNO 70
 PROCNO 1

F2 - Acquisition Parameters

Date_ 20160301
 Time 22.34
 INSTRUM spect
 PROBD 5 mm QNP 1H
 PULPROG zg30
 TD 16384
 SOLVENT CDCl3
 NS 16
 DS 0
 SWH 5995.204 Hz
 FIDRES 0.365918 Hz
 AQ 1.3664756 sec
 RG 912.3
 DW 83.400 usec
 DE 6.50 usec
 TE 300.0 K
 D1 1.50000000 sec

===== CHANNEL f1 =====

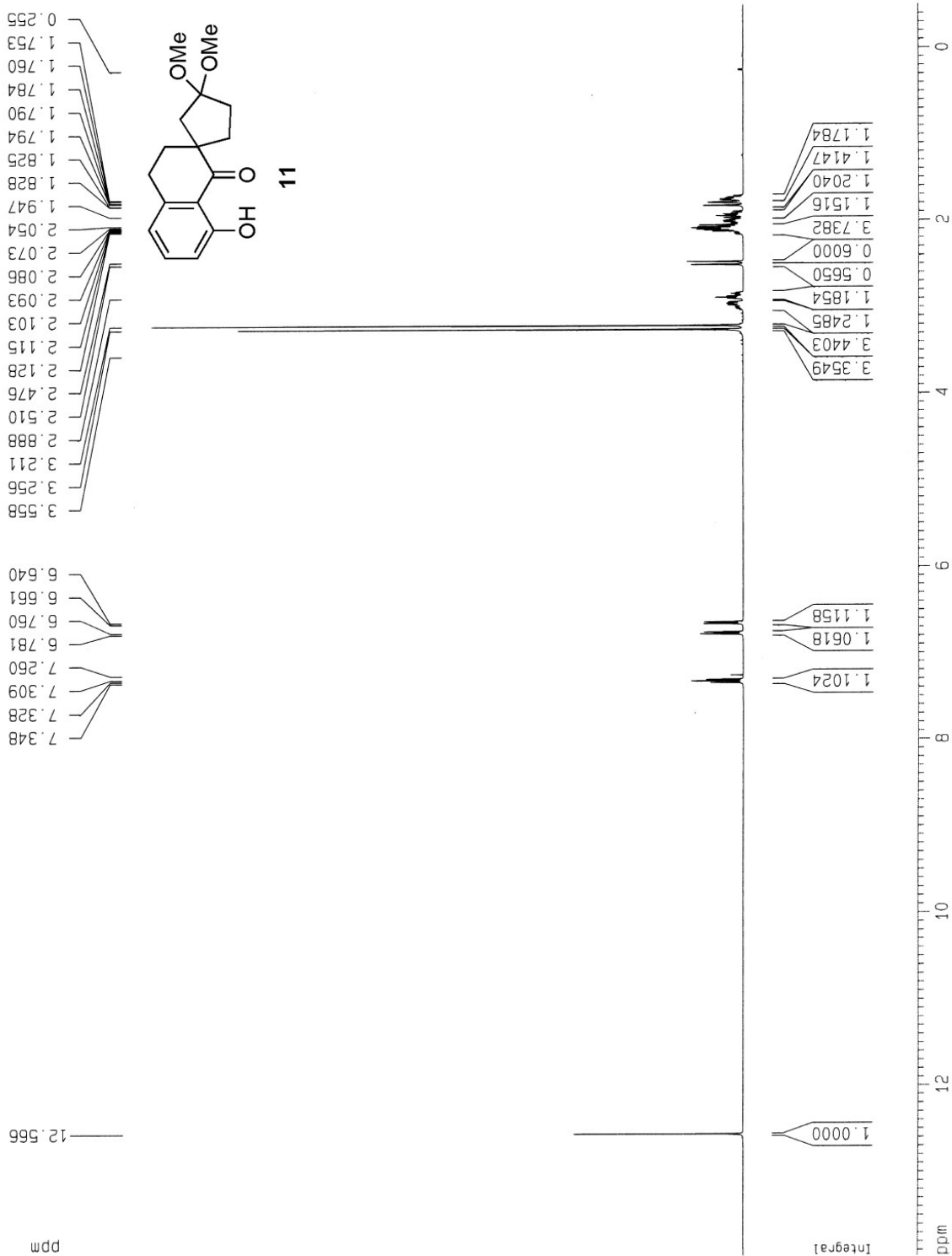
NUC1 1H
 P1 11.90 usec
 PL1 -3.00 dB
 SF01 400.1326008 MHz

F2 - Processing parameters

SI 8192
 SF 400.1300091 MHz
 WDW EM
 SSB 0
 LB 0.10 Hz
 GB 0
 PC 1.00

1D NMR plot parameters

CX 21.50 cm
 F1P 14.000 ppm
 F1 5601.82 Hz
 F2P -0.500 ppm
 F2 -200.06 Hz
 PPMCM 0.67442 ppm/cm
 HZCM 269.85510 Hz/cm



ppm

208.01

163.46

144.65

136.05

118.60

115.54

115.50

111.01

77.31

77.00

76.68

50.70

50.00

48.72

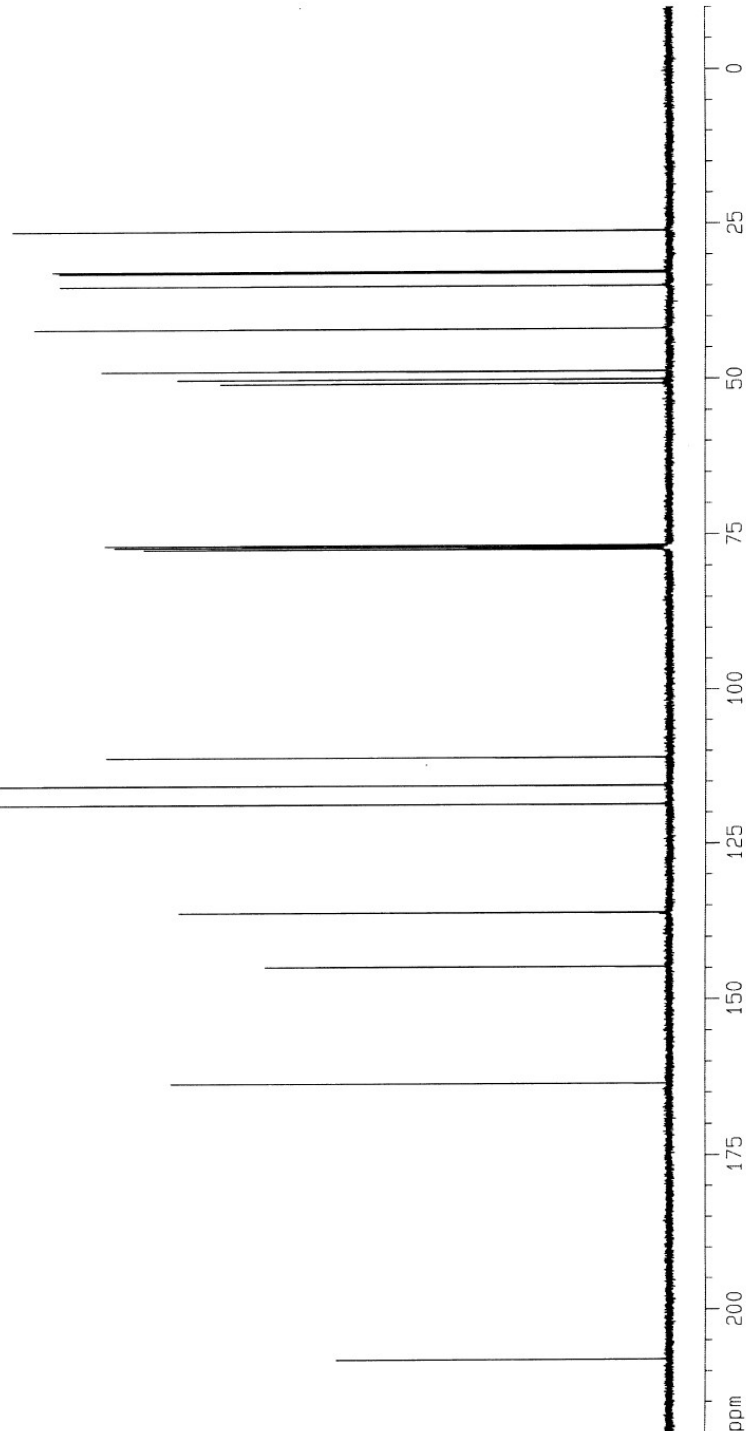
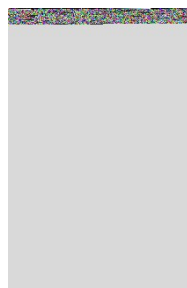
41.90

34.97

32.90

32.64

26.10



Current Data Parameters

NAME icy-10

EXPNO 1

PROCNO 1

F2 - Acquisition Parameters

Date_ 20160302

Time 11.10

INSTRUM spect

PROBHD 5 mm QNP 1H

PULPROG zgpg30

TD 65536

SOLVENT CDCl3

NS 1024

DS 4

SWH 25125.629 Hz

FIDRES 0.383387 Hz

AQ 1.3042164 sec

RG 256

DW 19.900 usec

DE 6.50 usec

TE 300.0 K

D1 2.00000000 sec

d11 0.03000000 sec

d12 0.00002000 sec

===== CHANNEL f1 =====

NUC1 13C

P1 10.20 usec

PL1 0.00 dB

SF01 100.6237959 MHz

===== CHANNEL f2 =====

CPDPRG2 waltz16

NUC2 1H

PCPD2 90.00 usec

PL2 -3.00 dB

PL12 14.50 dB

PL13 17.50 dB

SF02 400.1326008 MHz

F2 - Processing parameters

SI 32768

SF 100.6127754 MHz

WDW EM

SSB 0

LB 0.30 Hz

GB 0

PC 1.40

1D NMR plot parameters

CX 20.00 cm

F1P 220.000 ppm

F1 22134.81 Hz

F2P -10.000 ppm

F2 -1006.13 Hz

PPMCM 11.50000 ppm/cm

HZCM 1157.04688 Hz/cm

LCY-13

Sample Name
Date collected

LCY-13
2016-05-13

Pulse sequence
Solvent

PROTON
cdcl3

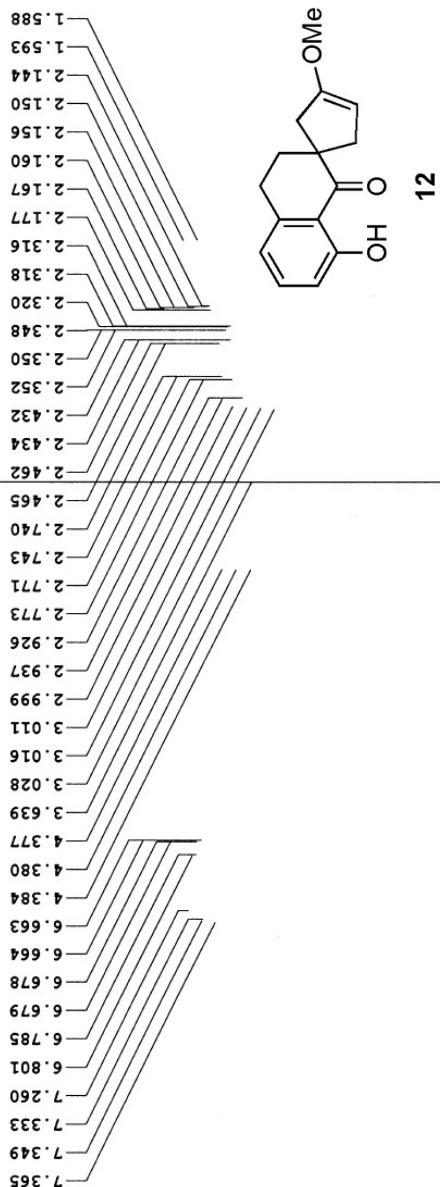
Temperature
Spectrometer

25
Agilent-NMR-inova500

Study Owner
Operator

vnmr2
vnmr2

12.587



LCY-13

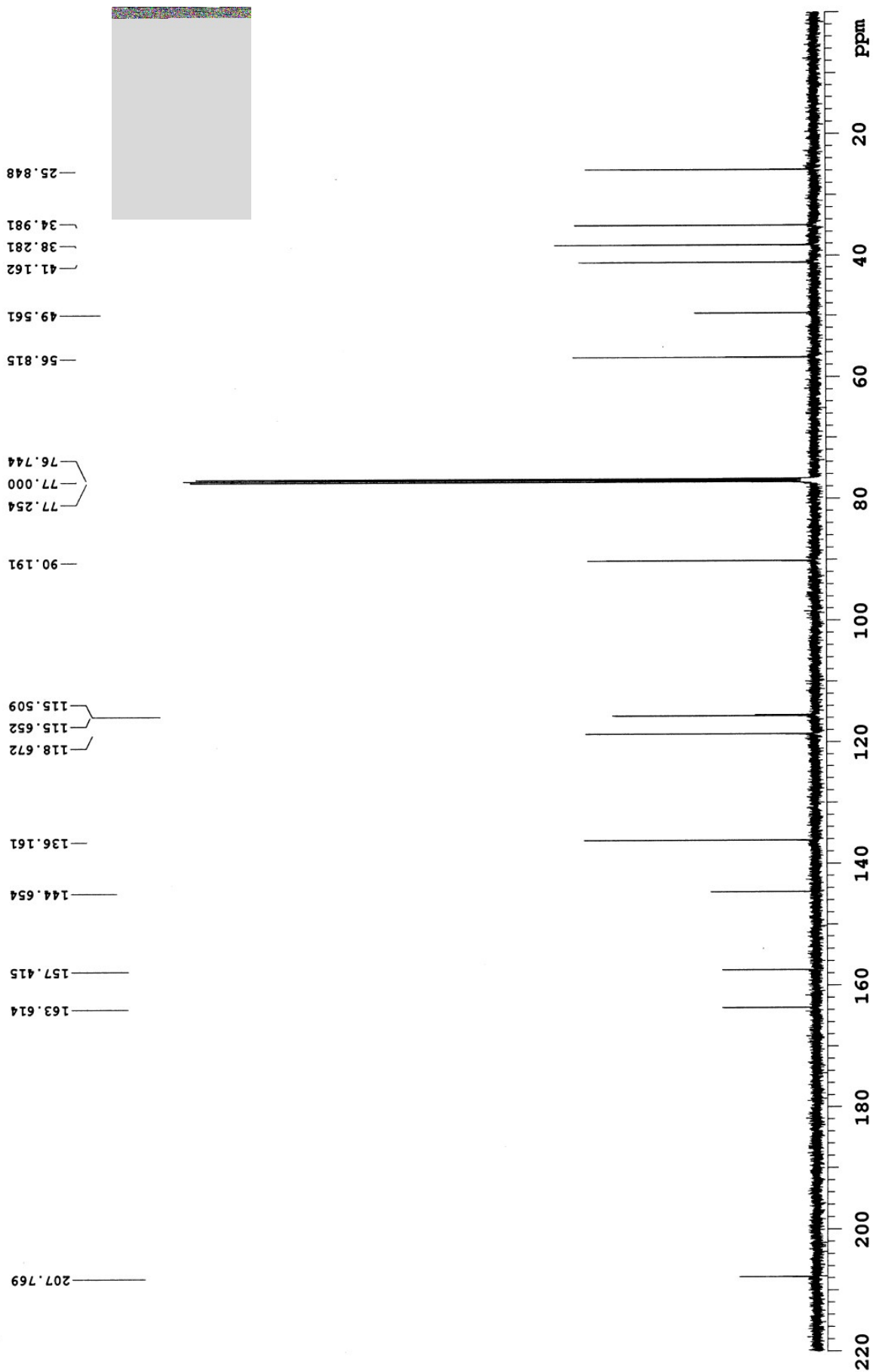
Sample Name **LCY-13**
Date collected **2016-05-13**

Pulse sequence
Solvent **cdcl3**

CARBON

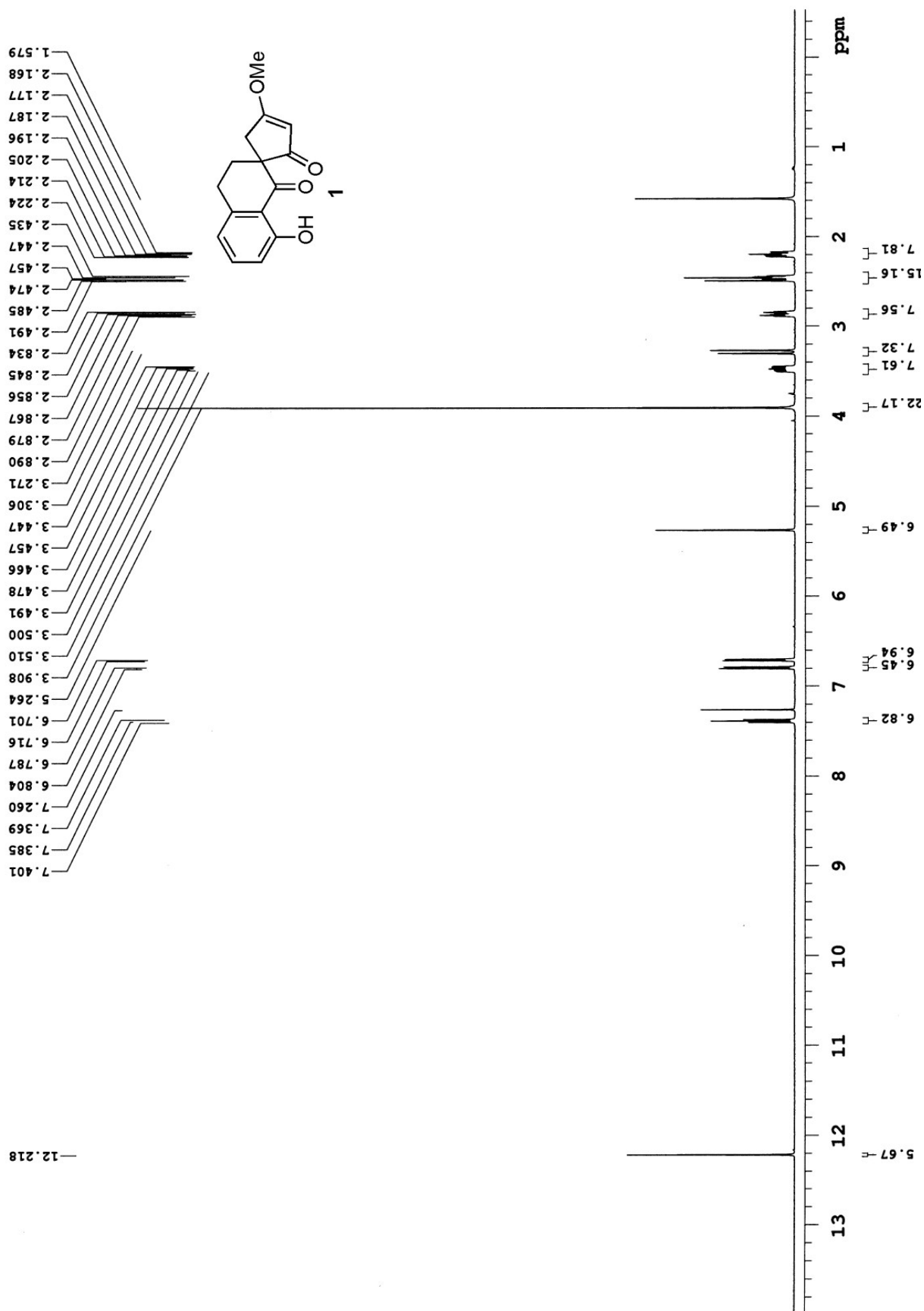
Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**



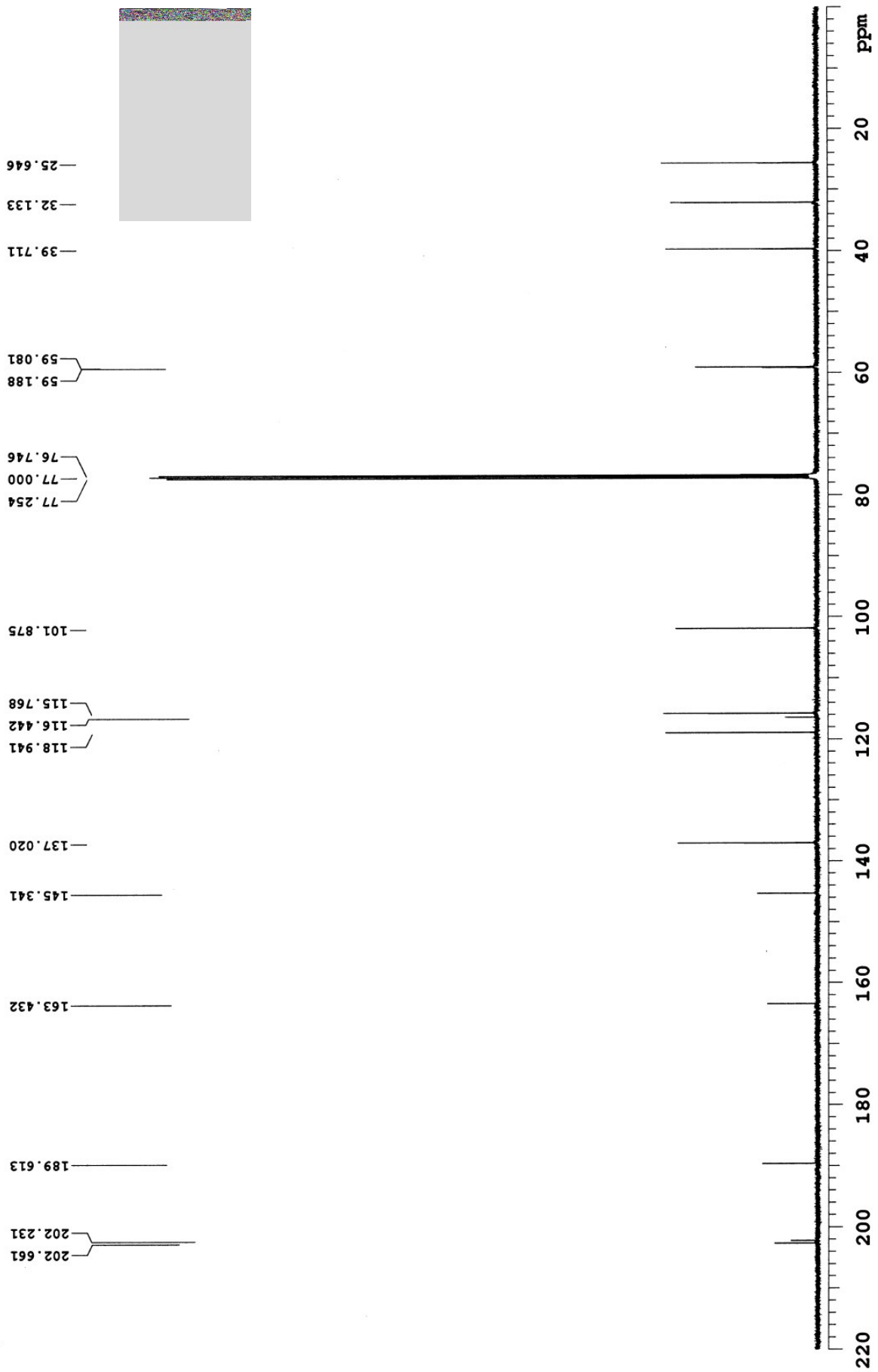
LCY-14

Sample Name **LCY-14** Date collected **2016-12-12** Pulse sequence **PROTON** Solvent **cdcl3** Temperature **25** Spectrometer **Agilent-NMR-inova500** Study owner **vnmr2** Operator **vnmr2**



LCY-14

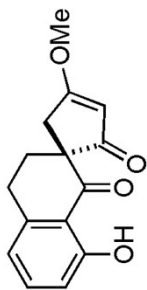
Sample Name **LCY-14** Date collected **2016-12-11** Pulse sequence **CARBON** Solvent **cdcl3** Temperature **25** Spectrometer **Agilent-NMR-inova500** Study owner **vnmr2** Operator **vnmr2**



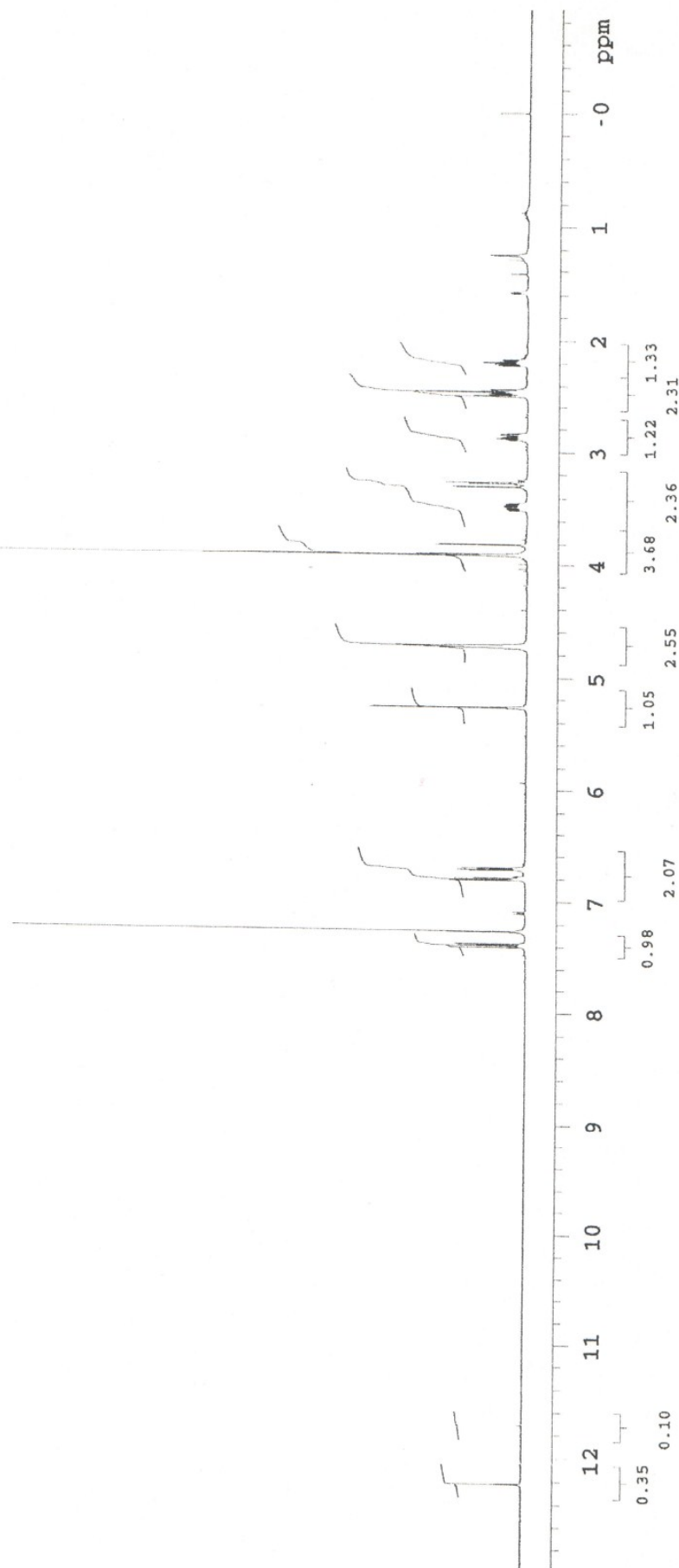
Instituto de Química UNAM ICH
 Dra. R. Mata
 Yanet Hernandez
 clave:RM63_EB-P14
 unity unity plus500 (7)
 disolvente:CDCl3
 hidrogeno-1+D2O
 28-01-03

Pulse Sequence: s2pul

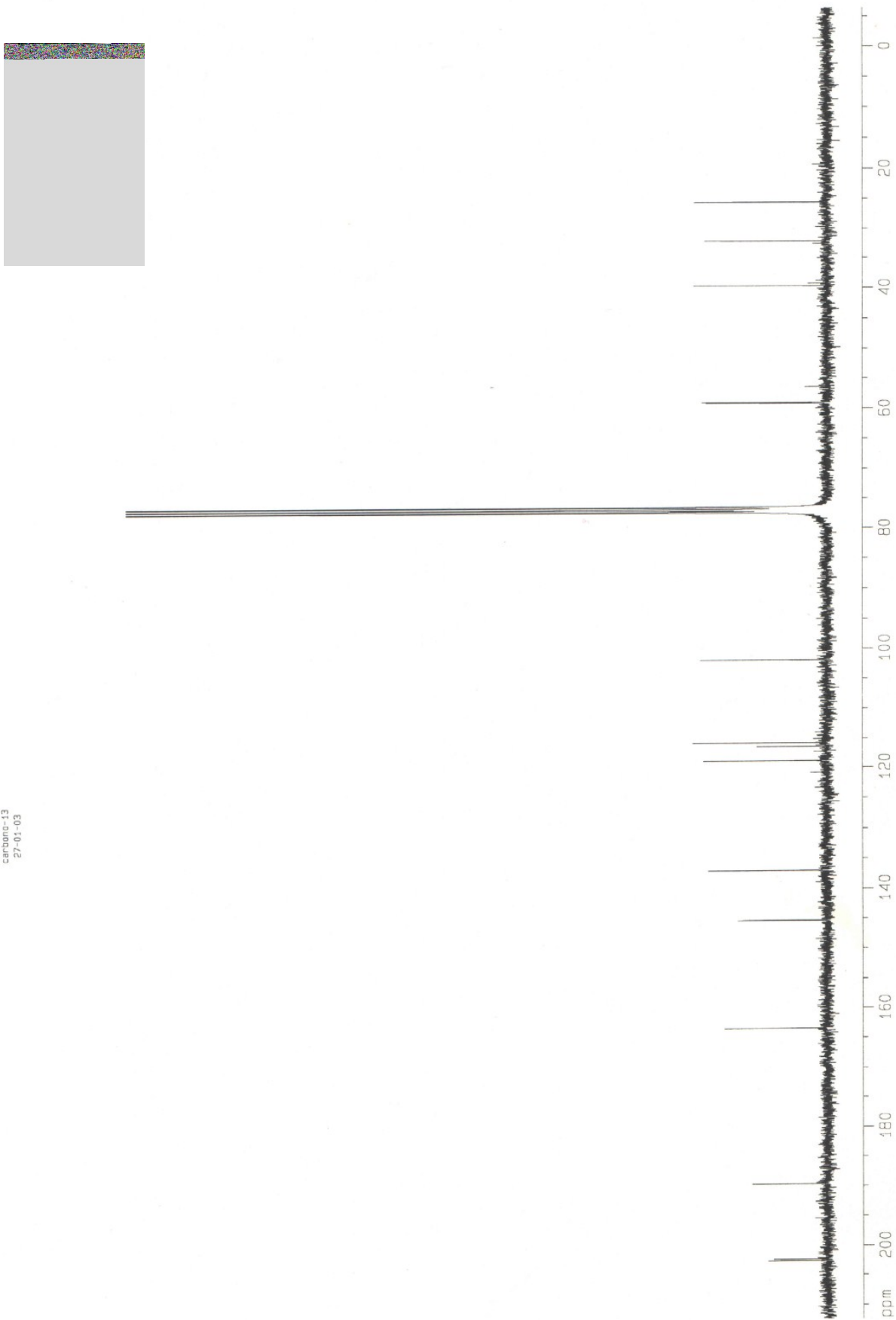
C15 H14 O4



natural (-)-nidemone



Instituto de Quimica UNAM ICH
Dra. R. Mota //Yeniet Hernandez
clave: RME3-EB014
disolvente: CDCl3
bruker avance300 75 MHz (6)
carburo-13
27-01-03



Current Data Parameters
 NAME LSP-test
 EXPNO 13
 PROCNO 1

F2 - Acquisition Parameters

Date_ 20150322
 Time 17.29
 INSTRUM spect
 PROBHD 5 mm QNP 1H
 PULPROG zg30
 TD 16384
 SOLVENT CDCl3
 NS 16
 DS 0
 SWH 5995.204 Hz
 FIDRES 0.365918 Hz
 AQ 1.3664756 sec
 RG 1024
 DW 83.400 usec
 DE 6.50 usec
 TE 300.0 K
 D1 1.50000000 sec

===== CHANNEL f1 =====

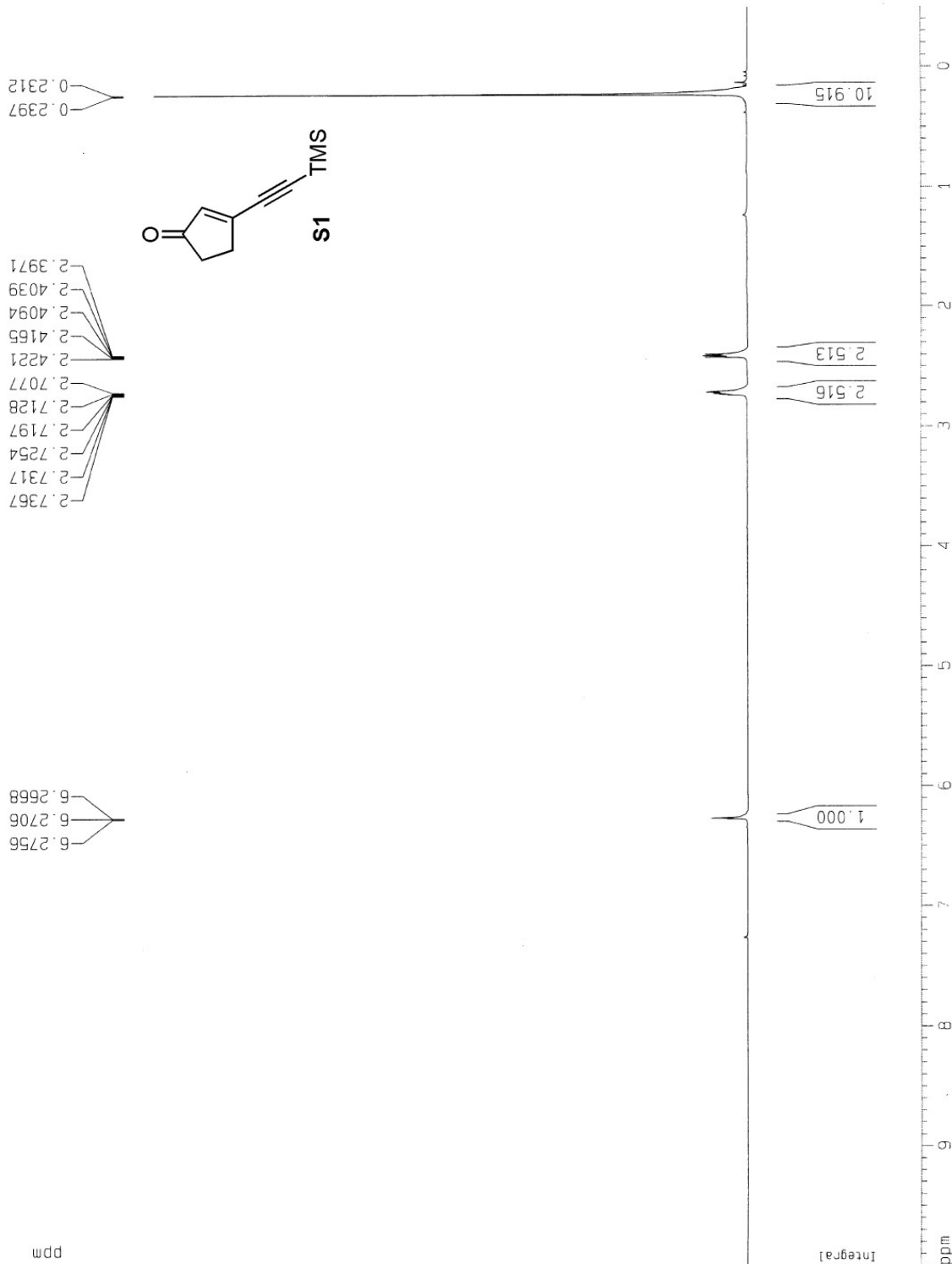
NUC1 1H
 P1 11.90 usec
 PL1 -3.00 dB
 SF01 400.1326008 MHz

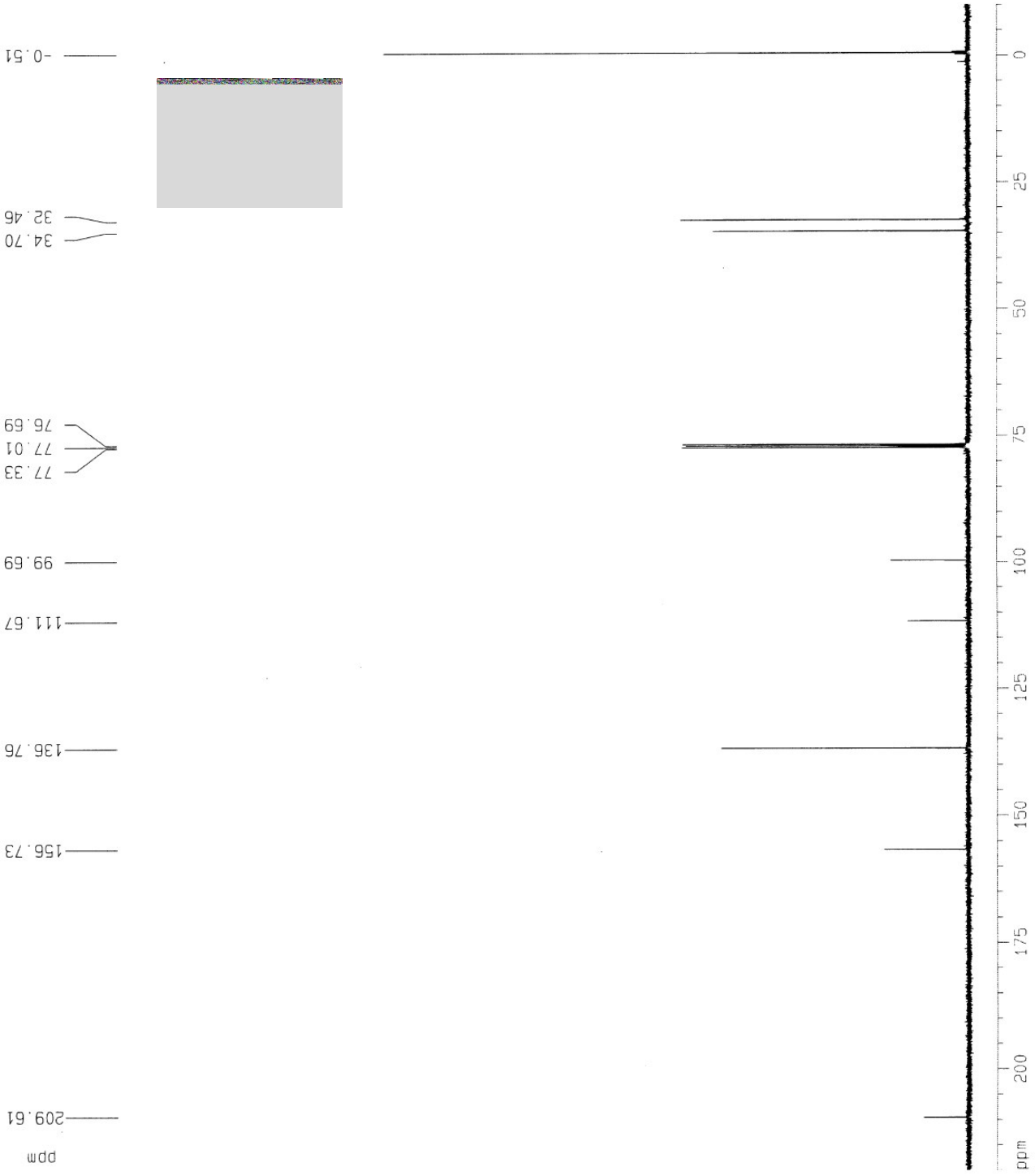
F2 - Processing parameters

SI 8192
 SF 400.1300099 MHz
 WDM EM
 SSB 0
 LB 0.10 Hz
 GB 0
 PC 1.00

1D NMR plot parameters

CX 21.50 cm
 F1P 10.000 ppm
 F1 4001.30 Hz
 F2P -0.500 ppm
 F2 -200.06 Hz
 PPMCM 0.48837 ppm/cm
 HZCM 195.41232 Hz/cm





Current Data Parameters
 NAME LSP-test
 EXPNO 14
 PROCNO 1

F2 - Acquisition Parameters

Date_ 20150322
 Time 18.27
 INSTRUM spect
 PROBHD 5 mm QNP 1H
 PULPROG zgpg30
 TO 65536
 SOLVENT DMSO
 NS 1024
 DS 4
 SWH 25125.629 Hz
 FIDRES 0.383387 Hz
 AQ 1.3042164 sec
 RG 256
 DW 19.900 usec
 DE 6.50 usec
 TE 300.0 K
 D1 2.00000000 sec
 d11 0.03000000 sec
 d12 0.00002000 sec

===== CHANNEL f1 =====

NUC1 13C
 P1 10.20 usec
 PL1 0.00 dB
 SF01 100.6237959 MHz

===== CHANNEL f2 =====

CPDPRG2 waltz16
 NUC2 1H
 PCPD2 90.00 usec
 PL2 -3.00 dB
 PL12 14.50 dB
 PL13 17.50 dB
 SF02 400.1326008 MHz

F2 - Processing parameters

SI 32768
 SF 100.6127731 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40

10 NMR plot parameters

CX 20.00 cm
 F1P 220.000 ppm
 F1 22134.81 Hz
 F2P -10.000 ppm
 F2 -1006.13 Hz
 PPMCM 11.50000 ppm/cm
 HZCM 1157.04688 Hz/cm

Current Data Parameters
 NAME LSP-test
 EXPNO 11
 PROCNO 1

F2 - Acquisition Parameters

Date_ 20150322
 Time 16.41
 INSTRUM spect
 PROBHD 5 mm QNP 1H
 PULPROG zg30
 TD 16384
 SOLVENT CDCl3
 NS 16
 DS 0
 SWH 5995.204 Hz
 FIDRES 0.365918 Hz
 AQ 1.3664756 sec
 RG 2896.3
 DW 83.400 usec
 DE 6.50 usec
 TE 300.0 K
 D1 1.50000000 sec

===== CHANNEL f1 =====

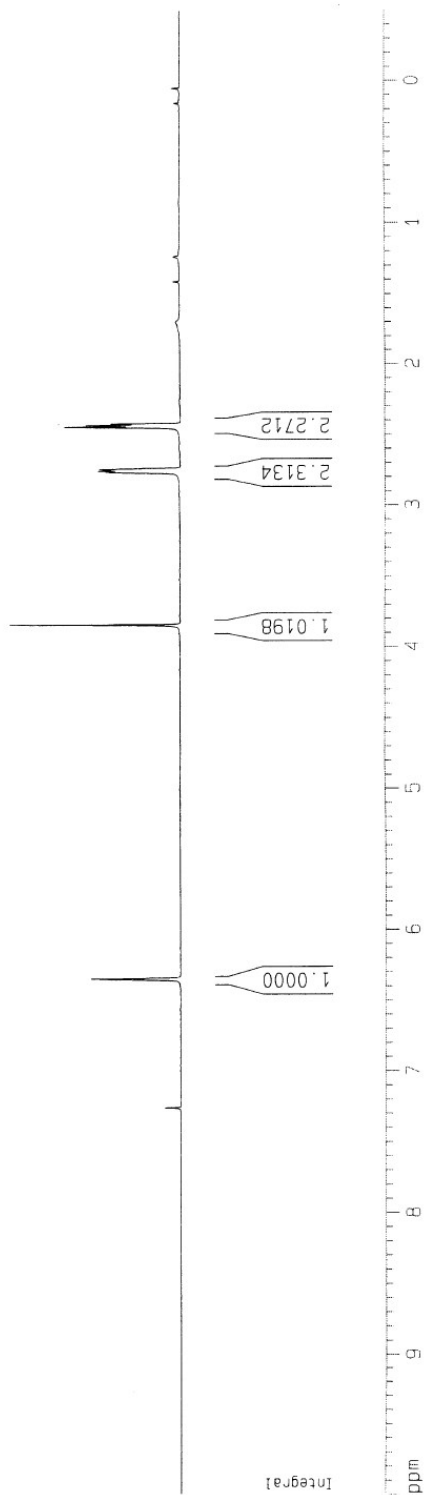
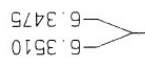
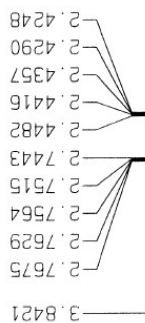
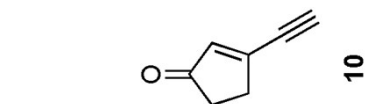
NUC1 1H
 P1 11.90 usec
 PL1 -3.00 dB
 SF01 400.1326008 MHz

F2 - Processing parameters

SI 8192
 SF 400.1300099 MHz
 WDW EM
 SSB 0
 LB 0.10 Hz
 GB 0
 PC 1.00

1D NMR plot parameters

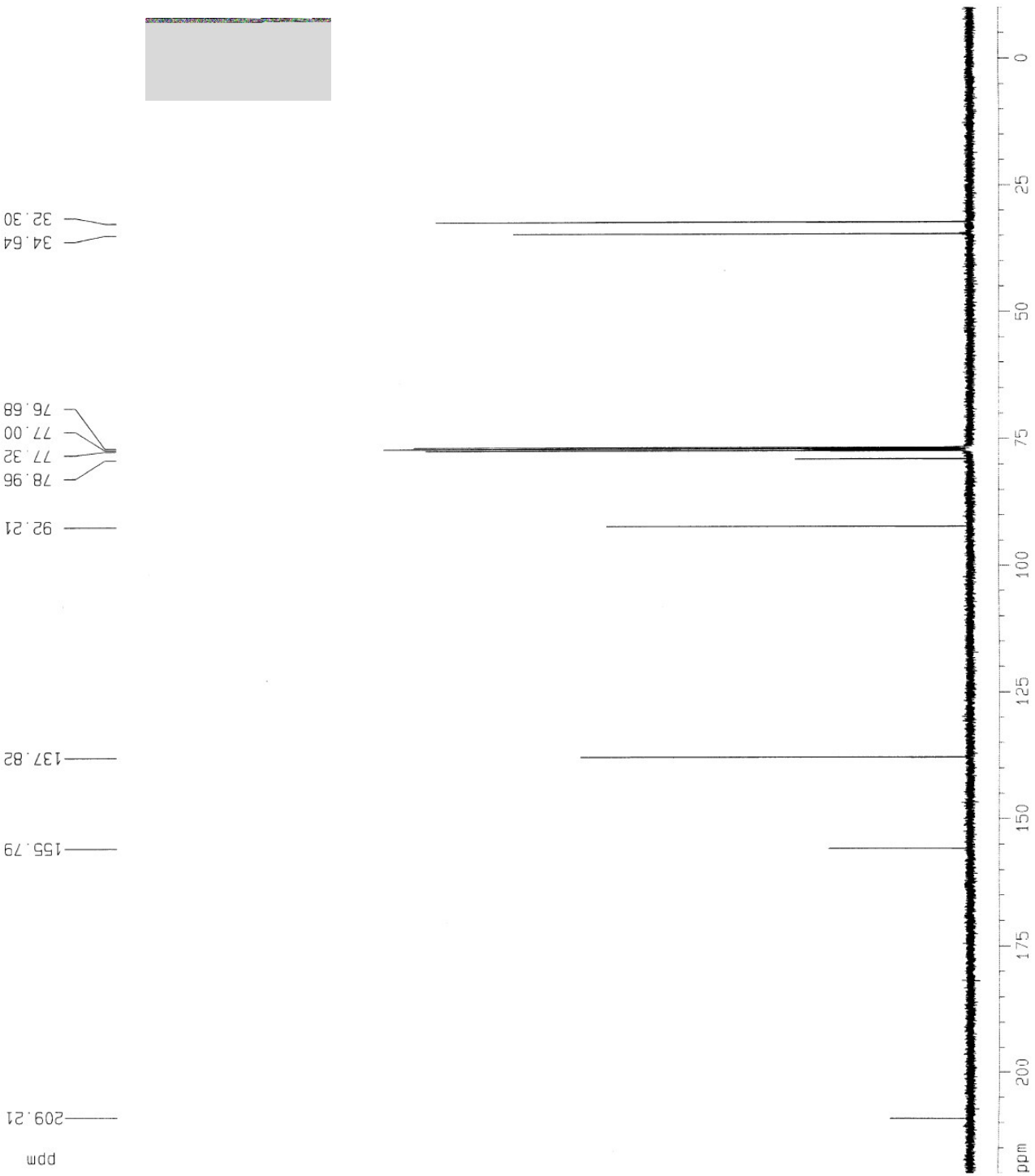
CX 21.50 cm
 F1P 10.000 ppm
 F1 4001.30 Hz
 F2P -0.500 ppm
 F2 -200.06 Hz
 PPMCM 0.48837 ppm/cm
 HZCM 195.41232 Hz/cm



ppm

Integral

ppm



Current Data Parameters
 NAME LSP-test
 EXPNO 12
 PROCNO 1

F2 - Acquisition Parameters

Date_ 20150322
 Time 17.24
 INSTRUM spect
 PROBHD 5 mm QNP 1H
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 757
 DS 4
 SWH 25125.629 Hz
 FIDRES 0.383387 Hz
 AQ 1.3042164 sec
 RG 256
 DW 19.900 usec
 DE 6.50 usec
 TE 300.0 K
 D1 2.00000000 sec
 d11 0.03000000 sec
 d12 0.00002000 sec

===== CHANNEL f1 =====

NUC1 13C
 P1 10.20 usec
 PL1 0.00 dB
 SF01 100.6237959 MHz

===== CHANNEL f2 =====

CPDPRG2 waltz16
 NUC2 1H
 PCPD2 90.00 usec
 PL2 -3.00 dB
 PL12 14.50 dB
 PL13 17.50 dB
 SF02 400.1326008 MHz

F2 - Processing parameters

SI 32768
 SF 100.6127738 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40

1D NMR plot parameters

CX 20.00 cm
 F1P 220.000 ppm
 F1 22134.81 Hz
 F2P -10.000 ppm
 F2 -1006.13 Hz
 PPMCM 11.50000 ppm/cm
 HZCM 1157.04688 Hz/cm