# **Electronic Supplementary Information**

## Regio-specific prenylation of pterocarpans by a membrane-bound

## prenyltransferase from Psoralea corylifolia

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### Experimental

#### Species identification of Psoralea corylifolia by DNA barcoding

Dried seeds of Psoralea corylifolia L. were collected from Yunnan province in China and were identified by DNA barcoding analysis of the chloroplast *psbA-trnH* spacer. Genomic DNA was extracted using the plant genomic DNA Kit (Tiangen Biotech, China). The primers and the reaction conditions for the *psbA-trnH* barcode used for PCR amplification were as follows. Primers for trnH-psbA sequence were psbA-F and trnH-R (Table S1). PCR amplification was performed in 25 µL reaction mixtures containing approximately 50 ng of genomic DNA template (0.5 µL), 1×TransStart KD Plus Buffer, 0.4 µM of each primer, 200 µM dNTPs, and 1.0 unit *TransStart*<sup>®</sup> KD Plus DNA Polymerase (TransGen Biotech, Beijing, China). The PCR conditions were initial denaturation at 94 °C for 3 min, followed by 35 cycles at 94 °C for 30 s, 55 °C for 30 s, and 68 °C for 45 s, and a final extension at 68 °C for 10 min. The products were then checked on 1.5% agarose gel electrophoresis, and purified using TIANquick Midi Purification Kit (Tiangen Biotech). Samples were sequenced using a 3730XL sequencer (Applied Biosystems, USA), and sequence alignment and analysis were performed with DNAMAN (version 8.0, lynnon biosoft, USA) software. The sample was finally identified as *P. corylifolia* based on the sequence alignment (GenBank accession number GU396699, GU396701, GU396700 and GQ434957). The psbA-trnH spacer sequence was given below.

#### psbA-trnH sequence

#### ATAATTTCCCTCTAGATCTAGCTGCGATCGACGCTCCATCTATTAATGGATAAAATTTTTATC

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## ESI-MS, <sup>1</sup>H and <sup>13</sup>C NMR data for prenylated product 1a

4-dimethylallylmaackiain (**1a**) (1.52 mg; amorphous powder, white):  $[α]^{25}_{D}$  –120.0 (*c* 0.05, MeOH). <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 400 MHz): δ 9.49 (1H, s, 3-OH), 7.09 (1H, d, *J* = 8.4 Hz, H-1), 6.97 (1H, s, H-7), 6.52 (1H, d, *J* = 8.4 Hz, H-2), 6.51 (1H, s, H-10), 5.94 (2H, two d, *J* = 0.9 Hz, -O-CH<sub>2</sub>-O-), 5.51 (1H, d, *J* = 6.8 Hz, H-11a), 4.27 (1H, dd, *J* = 9.0, 6.3 Hz, H-6α), 3.62–3.48 (2H, m, H-6β, H-6a), 5.12 (1H, t, *J* = 7.2 Hz, H-2'), 3.19 (2H, d, *J* = 6.8 Hz, H-1'), 1.69 (3H, s, H-4'), 1.60 (3H, s, H-5'). <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 100 MHz): δ 155.8 (C-3), 154.0 (C-4a), 153.7 (C-10a), 147.4 (C-9), 140.9 (C-8), 128.5 (C-1), 118.5 (C-7a), 115.1 (C-4), 111.4 (C-1a), 108.9 (C-2), 105.4 (C-7), 100.9 (-O-CH<sub>2</sub>-O-), 93.07 (C-10), 78.6 (C-11a), 66.2 (C-6), 40.1 (C-6a), 130.0 (C-3'), 122.9 (C-2'), 25.4 (C-5'), 21.8 (C-1'), 17.6 (C-4'). ESI-MS *m/z*: 353.0 [M+H]<sup>+</sup>. All of these data are in good agreement with the previously reported data (Figure S4, S5 and S9).<sup>1</sup>

Primer name	Sequences (5'–3')
psbA-F	GTTATGCATGAACGTAATGCTC
<i>trnH-</i> R	CGCGCATGGTGGATTCACAATCC
PCPT-F	ΑΑΑΤWGACAAGATAAAC
PcPT-R	GTCAGGTATATCCTTGYAC
PcPT-5'GSP	CAATCATGCACATTGCCGCAACCAATGG
PcPT-3'GSP	ATGGAAGAGATACCCATTGGTTGCGGCA
PcM4DT-BamH I-F*	GGAGAAAAAACCCCG <b>GATCC</b> ATGGATTGGGGATTAGCAATAT
PcM4DT- <i>Xho</i> I-R*	CGGTACCAAGCTTA <b>CTCGAG</b> TTATCTGACAAAAGCCATGAGC
ΔTP <sub>41</sub> -PcM4DT- <i>Bam</i> H I-F*	GGAGAAAAAACCCCG <b>GATCC</b> ATGGCTTCACAACACAAAAAGA

 Table S1. PCR primers used in this experiment.

\* Bold text represents restriction sites for subcloning.

# **Table S2.** Accession numbers of plant PTs used for phylogenetic analysis.

Name	Accession number	Definition	Organism
PcM4DT	MH626730	pterocarpan prenyltransferase	Psoralea corylifolia
AhR4DT-1	AQM74172.1	resveratrol-4-dimethylallyltransferase	Arachis hypogaea
AhR3'DT-1	AQM74173.1	resveratrol-3'-dimethylallyltransferase	Arachis hypogaea
AhR3'DT-2	AQM74174.1	resveratrol-3'-dimethylallyltransferase	Arachis hypogaea
AhR3'DT-3	AQM74175.1	resveratrol-3'-dimethylallyltransferase	Arachis hypogaea
AhR3'DT-4	AQM74176.1	resveratrol-3'-dimethylallyltransferase	Arachis hypogaea
AaVTE2-1	ABB70124.1	homogentisate phytyltransferase VTE2-1	Allium ampeloprasum
AtHPT1	AAM10489.1	homogentisate phytylprenyltransferase	Arabidopsis thaliana
AtPPT1	BAB20818.2	p-hydroxybenzoate polyprenyltransferase	Arabidopsis thaliana
AtVTE2-1	AAM10489.1	homogentisate phytylprenyltransferase	Arabidopsis thaliana
AtVTE2-2	ABB70127.1	homogentisate phytyltransferase VTE2-2	Arabidopsis thaliana
CIPT1a	BAP27988.1	umbelliferone 8-geranyltransferase	Citrus limon
CpVTE2-1	ABB70125.1	homogentisate phytyltransferase VTE2-1	Cuphea pulcherrima
CtIDT	AJD80983.1	isoliquiritigenin 3'-dimethylallyltransferase	Cudrania tricuspidata
GmG2DT	BAW32578.1	pterocarpan 2-dimethylallyltransferase	Glycine max
GmG4DT	BAH22520.1	pterocarpan 4-dimethylallyltransferase	Glycine max
GmVTE2-1	ABB70126.1	homogentisate phytyltransferase VTE2-1	Glycine max
GmVTE2-2	ABB70128.1	homogentisate phytyltransferase VTE2-2	Glycine max
GuA6DT	AIT11912.1	flavone prenyltransferase	Glycyrrhiza uralensis
HIPT-1	BAJ61049.1	aromatic prenyltransferase	Humulus lupulus
HIPT-2	AJD80255.1	aromatic prenyltransferase HIPT	Humulus lupulus
HvHGGT	AAP43911.1	homogentisic acid geranylgeranyl transferase	Hordeum vulgare
LaPT1	AER35706.1	genistein 3'-dimethylallyltransferase	Lupinus albus
LePGT-1	BAB84122.1	4-hydroxybenzoate geranyltransferase	Lithospermum erythrorhizon
LePGT-2	BAB84123.1	4-hydroxybenzoate geranyltransferase	Lithospermum erythrorhizon
HcPT	ALD84371.1	xanthone-specific prenyltransferase	Hypericum calycinum
MaIDT	AJD80982.1	isoliquiritigenin 3'-dimethylallyltransferase	Morus alba
OsHGGT	AAP43913.1	homogentisic acid geranylgeranyl transferase	Oryza sativa
OsPPT1	BAE96574.1	p-hydroxybenzoate polyprenyltransferase	Oryza sativa
PcPT	BAO31627.1	umbelliferone 6-dimethylallyltransferase	Petroselinum crispum
PsPT1	AJW31563.1	umbelliferone dimethylallyl transferase	Pastinaca sativa
PsPT2	AJW31564.1	umbelliferone dimethylallyl transferase	Pastinaca sativa
SfFPT	AHA36633.1	flavonoid prenyltransferase	Sophora flavescens
SfG6DT	BAK52291.1	genistein 6-dimethylallyltransferase	Sophora flavescens
SfiLDT	BAK52290.1	isoliquiritigenin dimethylallyltransferase	Sophora flavescens
SfN8DT-1	BAG12671.1	naringenin 8-dimethylallyltransferase	Sophora flavescens
SfN8DT-2	BAG12673.1	naringenin 8-dimethylallyltransferase	Sophora flavescens
SfN8DT-3	BAK52289.1	naringenin 8-dimethylallyltransferase	Sophora flavescens
TaHGGT	AAP43912.1	homogentisic acid geranylgeranyl transferase	Triticum aestivum
TaVTE2-1	ABB70123.1	homogentisate phytyltransferase VTE2-1	Triticum aestivum
ZmVTE2-1	ABB70122.1	homogentisate phytyltransferase VTE2-1	Zea mays
AgGPPS2	AAN01134.1	geranyl diphosphate synthase	Abies grandis
AtGPPS	CAC16849.1	geranyl diphosphate synthase	Arabidopsis thaliana

PgFPS	AAY87903.1	farnesyl diphosphate synthase	Panax ginseng	
AtFPS	AAB07248.1	farnesyl diphosphate synthase	Arabidopsis thaliana	
GbGGPPS	AAQ72786.1	geranylgeranyl diphosphate synthase	Ginkgo biloba	
SmGGPPS	ACR19637.1	geranylgeranyl diphosphate synthase	Salvia miltiorrhiza	
CsPT1	Page J, Boubakir Z. 2010. Aromatic prenyltransferase from cannabis. Patent WO 2011017798 A1.			



**Figure S1**. Four-week-old seedlings of *Psoralea corylifolia* (A) and agarose gel electrophoresis of RT-PCR products (B). M: DNA marker; 1: *PcM4DT*; 2: Δ*TP41-PcM4DT*.



**Figure S2**. A multiple alignment of the amino acid sequences of PcM4DT and other plant pterocarpans prenyltransferases. GmG4DT: pterocarpan 4-dimethylallyltransferase form *Glycine max* (Genebank accession number BAH22520); GmG2DT: pterocarpan 2-dimethylallyltransferase form *Glycine max* (Genebank accession number BAW32578). Identical amino acid are shown in white on a black background. The conserved aspartate-rich motifs NQxxDxxxD and KD(I/L)xDx(E/D)GD are highlighted with red box.



**Figure S3**. Putative transmembrane domains of PcM4DT were predicted by TMHMM Server program version 2.0. TP: Transit peptide; TM: transmembrane α-helix; L2: loop number 2; L5: loop number 5.



Figure S4. <sup>1</sup>H NMR spectrum of **1a** in DMSO- $d_6$  (400 MHz).



Figure S5. <sup>13</sup>C NMR spectrum of **1a** in DMSO- $d_6$  (100 MHz).



**Figure S6**. Determination of kinetic parameters for recombinant PcM4DT. The apparent  $K_m$  values for maackiain (**1**) were determined with varying concentrations (5–400  $\mu$ M) using 400  $\mu$ M DMAPP as prenyl donor (A), while that for DMAPP were determined with various concentrations (10–800  $\mu$ M) using 400  $\mu$ M **1** as prenyl acceptor (B). The values are averages of three parallel experiments.



**Figure S7**. Enzymatic characterization of recombinant PcM4DT. (A) HPLC analysis of the enzymatic reaction mixture from microsomal fractions containing 3-hydroxy-9-methoxy-pterocarpan (**2**), DMAPP, and Mg<sup>2+</sup>. (B) MS analysis of prenylated product **2a** from  $\Delta$ TP<sub>41</sub>-PcM4DT-expressing microsomes.



Figure S8. Aromatic compounds that are not prenylated by PcM4DT.



Figure S9. The CD spectra of compound 1 (red) and 1a (blue).



Figure S10. IC  $_{\rm 50}$  curves of compounds 1a and 2a.

## References

1 G. L. DaSilva, F. J. D. Matos and E. R. Silveira, *Phytochemistry*, 1997, 46, 1059-1062.