

**Inverse-electron-demand Diels–Alder reaction of
 α,β -unsaturated hydrazones with 3-methoxycarbonyl α -pyrone**

Supporting Information

Yoshimitsu Hashimoto, Ryo Abe, Nobuyoshi Morita and Osamu Tamura*

Showa Pharmaceutical University, 3-3165 Higashi-Tamagawagakuen, Machida, Tokyo 194-8543, Japan

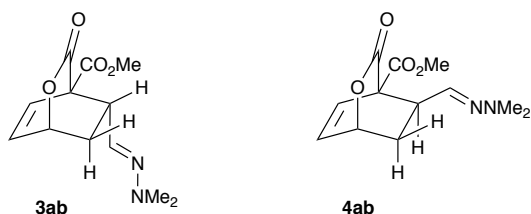
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General procedure

^1H and ^{13}C NMR spectra were recorded with JEOL JNM-AL300, JEOL JNM-ECZ400S, BRUKER AV300M or BRUKER AV600 spectrometer at room temperature, with tetramethylsilane ($\delta = 0$) as an internal standard (CDCl_3 solution). Chemical shifts were expressed in ppm, and coupling constants (J) in Hz. Infrared (IR) spectra were recorded with a Shimadzu FTIR-8200A spectrometer. Mass spectra were recorded on JEOL JMS-700 and JMS-T100LP spectrometers. Melting points were determined by using a Yanaco melting point apparatus MP-S3. Merck silica gel 60 (1.09385) and Merck silica gel 60 F254 were used for column chromatography and thin layer chromatography (TLC), respectively.

Synthesis and Data



endo-cycloadduct **3ab** and its *exo*-isomer **4ab**

A solution of α -pyrone **1a** (50.0 mg, 0.324 mmol), acrolein hydrazone **2b** (63.7 mg, 0.649 mmol) and $\text{Eu}(\text{hfc})_3$ (38.7 mg, 32.4 μmol) in 1,2-dichloroethane (1.6 mL) was stirred at room temperature for 26 hours. After the reaction mixture was concentrated under reduced pressure, the residue was purified by column chromatography on silica gel (CHCl_3 : acetone = 10 : 1) to afford *endo* adduct **3ab** (65.1 mg, 80%) as a pale yellow solid and *exo* adduct **4ab** (5.3 mg, 6%) as a brown amorphous.

3ab: mp 54.0–54.4 $^\circ\text{C}$; IR (KBr) 2953, 1751, 1277, 1105, 1070 cm^{-1} ; ^1H NMR (300 MHz, CDCl_3) δ 6.78 (1H, brd, $J = 7.8$ Hz), 6.50 (1H, dd, $J = 7.8, 5.1$ Hz), 6.21 (1H, d, $J = 3.9$ Hz), 5.29–5.21 (1H, m), 3.88 (3H, s), 3.48 (1H, dt, $J = 9.6, 3.9$ Hz), 2.68 (6H, s), 2.55 (1H, ddd, $J = 13.2, 9.6, 3.9$ Hz), 1.96 (1H, ddd, $J = 13.2, 3.9, 1.5$ Hz); ^{13}C NMR (75 MHz, CDCl_3) δ 170.3, 168.3, 131.8, 131.1, 130.4, 74.4, 58.3, 52.8, 42.8, 37.6, 31.0; HRMS (FAB) m/z calcd for $\text{C}_{12}\text{H}_{17}\text{N}_2\text{O}_4$ $[\text{M}+\text{H}]^+$ 253.1188, found 253.1193.

4ab: IR (KBr) 2955, 1746, 1275, 1076 cm^{-1} ; ^1H NMR (300 MHz, CDCl_3) δ 6.75 (1H, dd, $J = 7.8, 1.8$ Hz), 6.60 (1H, dd, $J = 7.8, 5.1$ Hz), 6.58 (1H, d, $J = 6.6$ Hz), 5.29–5.20 (1H, m), 3.85 (3H, s), 2.96 (1H, ddd, $J = 10.5, 6.6, 4.6$ Hz), 2.76 (6H, s), 2.42 (1H, dt, $J = 13.5, 4.6$ Hz), 2.06 (1H, ddd, $J = 13.5, 10.5, 1.5$ Hz); ^{13}C NMR (75 MHz, CDCl_3) δ 168.4, 168.2, 134.6, 132.8, 132.2, 74.1, 57.9, 52.7, 43.0, 39.9, 32.2; HRMS (FAB) m/z calcd for $\text{C}_{12}\text{H}_{17}\text{N}_2\text{O}_4$ $[\text{M}+\text{H}]^+$ 253.1188, found 253.1200.



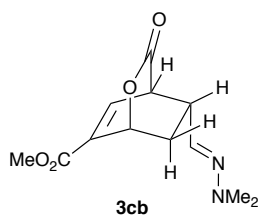
4-*p*-toluenesulfonyl-*endo*-cycloadduct **3bb and its *exo*-isomer **4bb****

A solution of 3-Ts- α -pyrone **1b** (50.0 mg, 0.200 mmol), acrolein hydrazone **2b** (44.9 mg, 0.400 mmol) and Eu(hfc)₃ (23.9 mg, 20.0 μ mol) in 1,2-dichloroethane (1.0 mL) was stirred at 50 °C for 10 hours. After the reaction mixture was concentrated under reduced pressure, the residue was purified by column chromatography on silica gel (hexane : AcOEt = 1 : 1 to 1 : 2) to afford a mixture of *endo* adduct **3bb** and *exo* adduct **4bb** (69.2 mg, 99%, **3bb** : **4bb** = 83 : 17) as pale yellow amorphous.

Analytical samples were obtained by separation of the mixture with PTLC (toluene : Et₂O = 1 : 5 ($\times 2$)) to afford **3bb** and **4bb** both as a white solid.

3bb: mp 129-130 °C (decomp.); IR (KBr) 2955, 2860, 1759, 1597, 1319, 1153, 1043 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 8.05 (2H, d, J = 8.1 Hz), 7.34 (2H, d, J = 8.1 Hz), 6.86 (1H, d, J = 7.8 Hz), 6.75 (1H, dd, J = 7.8, 4.8 Hz), 6.63 (1H, d, J = 6.0 Hz), 5.20–5.15 (1H, m), 3.34 (1H, ddd, J = 9.6, 6.0, 3.0 Hz), 2.83 (6H, s), 2.52 (1H, ddd, J = 13.2, 9.6, 3.6 Hz), 2.44 (3H, s), 2.10 (1H, ddd, J = 13.2, 3.0, 1.2 Hz); ¹³C NMR (150 MHz, CDCl₃) δ 166.6, 145.5, 134.0, 133.4, 132.4, 131.4, 129.1, 127.2, 73.3, 73.2, 42.9, 37.9, 32.5, 21.7; HRMS (ESI) m/z calcd for C₁₇H₂₀N₂O₄NaS [M+Na]⁺ 371.1042, found 371.1028.

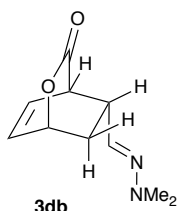
4bb: mp 137-139 °C (decomp.); IR (KBr) 2955, 2925, 2858, 1763, 1597, 1367, 1155, 1074, 1045 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 8.06 (2H, d, J = 8.4 Hz), 7.34 (2H, d, J = 8.4 Hz), 6.63 (1H, dd, J = 8.4, 1.8 Hz), 6.58 (1H, dd, J = 8.4, 5.4 Hz), 6.51 (1H, d, J = 6.3 Hz), 5.20–5.13 (1H, m), 3.36 (1H, ddd, J = 10.2, 6.3, 4.8 Hz), 2.77 (6H, s), 2.52 (1H, dt, J = 13.2, 4.8 Hz), 2.44 (3H, s), 2.10 (1H, ddd, J = 13.2, 10.2, 1.2 Hz); ¹³C NMR (150 MHz, CDCl₃) δ 165.7, 145.5, 134.3, 133.3, 131.34, 131.30, 130.4, 129.4, 73.6, 73.4, 42.6, 39.3, 33.5, 21.7; HRMS (ESI) m/z calcd for C₁₇H₂₀N₂O₄NaS [M+Na]⁺ 371.1042, found 371.1040.



2-methoxycarbonyl-*exo*-cycloadduct **4cb**

A solution of methyl coumalate **1c** (100 mg, 0.648 mmol), acrolein hydrazone **2b** (127 mg, 1.30 mmol) and Eu(hfc)₃ (77.4 mg, 64.8 μmol) in 1,2-dichloroethane (3.2 mL) was stirred at room temperature for 48 hours. After the reaction mixture was concentrated under reduced pressure, the residue was purified by column chromatography on silica gel (hexane : AcOEt = 1 : 1) and (CHCl₃ : acetone = 10 : 1) to afford *exo* adduct **4cb** (3.1 mg, 2%) as yellow oil.

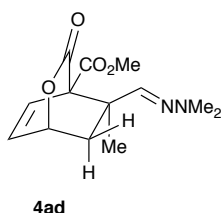
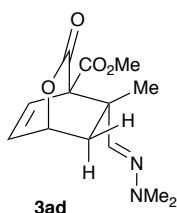
IR (NaCl) 2953, 2856, 1764, 1717, 1439, 1267, 1013 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.41 (1H, dd, *J* = 6.3, 2.1 Hz), 6.33 (1H, d, *J* = 6.3 Hz), 5.71 (1H, ddd, *J* = 3.9, 2.1, 1.8 Hz), 3.82 (3H, s), 3.65 (1H, dd, *J* = 6.3, 2.4 Hz), 2.80–2.71 (1H, m), 2.78 (6H, s), 2.31 (1H, ddd, *J* = 14.1, 5.4, 3.9 Hz), 2.01 (1H, ddd, *J* = 14.1, 10.5, 1.8 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 170.8, 162.6, 141.1, 136.6, 132.9, 73.7, 52.2, 47.5, 42.8, 36.8, 30.4; HRMS (FAB) *m/z* calcd for C₁₂H₁₇N₂O₄ [M+H]⁺ 253.1188, found 253.1182.



endo-cycloadduct **3db**

A solution of α-pyrone **1d** (50.0 mg, 0.520 mmol), acrolein hydrazone **2b** (102 mg, 1.04 mmol) and Eu(hfc)₃ (62.1 mg, 52.0 μmol) in 1,2-dichloroethane (2.6 mL) was stirred 80 °C for 40 hours. After the reaction mixture was concentrated under reduced pressure, the residue was purified by column chromatography on silica gel (hexane : AcOEt = 1 : 1) to afford *endo* adduct **3bd** (9.3 mg, 9%) as a brown oil. Unreacted **1d** was recovered (39.2 mg, 78%).

IR (NaCl) 2954, 2858, 1755, 1747, 1364, 1182, 1015 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 6.53 (1H, ddd, *J* = 7.8, 7.2, 1.5 Hz), 6.39 (1H, ddd, *J* = 7.8, 6.0, 1.5 Hz), 6.26 (1H, d, *J* = 4.8 Hz), 5.24 (1H, ddd, *J* = 7.2, 3.9, 1.5 Hz), 3.67 (1H, ddd, *J* = 6.0, 2.7, 1.5 Hz), 3.06–2.95 (1H, m), 2.73 (6H, s), 2.46 (1H, ddd, *J* = 13.5, 9.3, 3.9 Hz), 1.86 (1H, ddd, *J* = 13.5, 3.9, 1.5 Hz); ¹³C NMR (75 MHz, CDCl₃) δ 173.6, 134.7, 132.4, 130.1, 74.3, 45.4, 42.9, 34.8, 30.5; HRMS (FAB) *m/z* calcd for C₁₀H₁₅N₂O₂ [M+H]⁺ 195.1134, found 195.1140.

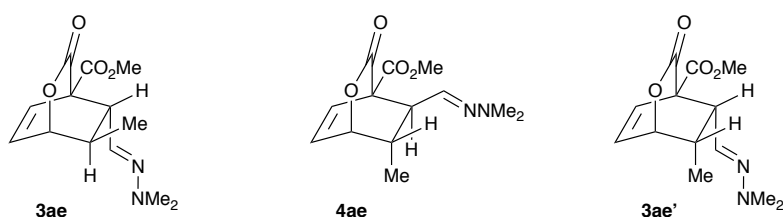


5-methyl-*endo*-cycloadduct **3ad** and its *exo*-isomer **4ad**

A solution of α-pyrone **1a** (50.0 mg, 0.324 mmol), methacrolein hydrazone **2d** (72.8 mg, 0.649 mmol) and Eu(hfc)₃ (38.7 mg, 32.4 μmol) in 1,2-dichloroethane (1.6 mL) was stirred at 0 °C for 4 hour and at room

temperature for 4.5 hours. After the reaction mixture was concentrated under reduced pressure, the residue was purified by column chromatography on silica gel (hexane : AcOEt = 1 : 1) to afford a mixture of *endo* adduct **3ad** and *exo* adduct **4ad** (57.7 mg, 67%, **3ad** : **4ad** = 52 : 48) as yellow oil.

IR (NaCl) 2955, 2856, 1766, 1758, 1744, 1732, 1445, 1067, 1020 cm^{-1} (described without distinction of the two isomers); ^1H NMR (300 MHz, CDCl_3 , signals from the *exo* isomer **4ad** are marked with an asterisk) δ 6.87* (1H, s), 6.76* (1H, dd, $J = 7.8, 1.8$ Hz), 6.69 (1H, dd, $J = 7.8, 1.8$ Hz), 6.59* (1H, dd, $J = 7.8, 5.1$ Hz), 6.47 (1H, dd, $J = 7.8, 5.1$ Hz), 6.33 (1H, s), 5.22–5.14* (1H, m), 5.21–5.13 (1H, m), 3.84* (3H, s), 3.84 (3H, s), 3.03* (1H, dd, $J = 13.5, 4.2$ Hz), 2.75* (6H, s), 2.67 (6H, s), 2.42 (1H, dd, $J = 13.5, 1.5$ Hz), 2.05 (1H, dd, $J = 13.5, 4.2$), 1.58 (3H, s), 1.46* (1H, dd, $J = 13.5, 1.5$ Hz), 1.19* (3H, s); ^{13}C NMR (75 MHz, CDCl_3 , signals that could be distinguished from the *exo* isomer **4ad** are marked with an asterisk) δ 169.8, 169.3*, 168.1, 167.7, 140.1*, 136.8, 132.4, 131.7*, 131.4*, 130.1, 74.5, 74.0*, 63.5*, 61.9, 52.5, 52.4, 43.0*, 42.9, 42.7, 42.1, 40.7, 39.5*, 26.7, 23.5*; HRMS (FAB) m/z calcd for $\text{C}_{13}\text{H}_{19}\text{N}_2\text{O}_4$ $[\text{M}+\text{H}]^+$ 267.1345, found 267.1357.



6-methyl-*endo*-cycloadduct **3ae**, its *exo*-isomer **4ae** and *cis*-alkene derived *endo*-adduct **3ae'**

A solution of α -pyrone **1a** (50.0 mg, 0.324 mmol), crotonaldehyde hydrazone **2e** (72.7 mg, 0.648 mmol) and $\text{Eu}(\text{hfc})_3$ (38.7 mg, 32.4 μmol) in 1,2-dichloroethane (1.6 mL) was stirred at room temperature for 16 hours. After the reaction mixture was concentrated under reduced pressure, the residue was purified by column chromatography on silica gel (hexane : AcOEt = 3 : 2 to 1 : 1) to afford *endo* adduct **3ae** (45.5 mg, 53%) as pale yellow oil and a mixture of **3ae**, *exo* adduct **4ae** and *cis*-alkene derived *endo*-adduct **3ae'** (19.9 mg, 23%, **3ae** : **4ae** : **3ae'** = 71 : 14 : 15) as orange oil.

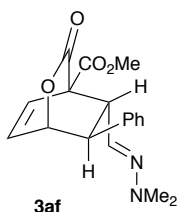
Analytical samples were obtained by separation of the mixture with PTLC (CH_2Cl_2 : AcOEt = 2 : 1) to afford *exo* adduct **4ae** and *cis*-alkene derived *endo*-adduct **3ae'** both as pale yellow oil.

3ae: IR (NaCl) 2968, 2851, 1755, 1724, 1435, 1304, 1279, 1119, 1099 cm^{-1} ; ^1H NMR (600 MHz, CDCl_3) δ 6.76 (1H, brd, $J = 7.8$ Hz), 6.53 (1H, dd, $J = 7.8, 5.4$ Hz), 6.23 (1H, d, $J = 4.2$ Hz), 4.90 (1H, dt, $J = 5.4, 1.2$ Hz), 3.85 (3H, s), 2.86 (1H, t, $J = 4.2$ Hz), 2.68 (6H, s), 2.13–2.06 (1H, m), 1.30 (3H, d, $J = 7.2$ Hz); ^{13}C NMR (150 MHz, CDCl_3) δ 170.2, 168.3, 131.4, 131.0, 130.3, 79.6, 58.3, 52.6, 46.0, 42.8, 38.0, 17.8; HRMS (FAB) m/z calcd for $\text{C}_{13}\text{H}_{19}\text{N}_2\text{O}_4$ $[\text{M}+\text{H}]^+$ 267.1345, found 267.1344.

4ae: IR (NaCl) 2957, 1749, 1734, 1286, 1096, 1034 cm^{-1} ; ^1H NMR (600 MHz, CDCl_3) δ 6.84 (1H, dd, $J =$

7.8, 1.8 Hz), 6.53 (1H, dd, $J = 7.8, 4.8$ Hz), 6.49 (1H, d, $J = 7.2$ Hz), 4.98 (1H, ddd, $J = 4.8, 3.6, 1.8$ Hz), 3.82 (3H, s), 2.76 (6H, s), 2.57–2.50 (1H, m), 2.34 (1H, dd, $J = 7.2, 5.4$ Hz), 1.01 (3H, d, $J = 6.6$ Hz); ^{13}C NMR (150 MHz, CDCl_3) δ 168.2, 167.8, 133.9, 133.3, 130.4, 78.3, 58.0, 52.6, 49.3, 43.0, 39.5, 17.8; HRMS (FAB) m/z calcd for $\text{C}_{13}\text{H}_{19}\text{N}_2\text{O}_4$ $[\text{M}+\text{H}]^+$ 267.1345, found 267.1340.

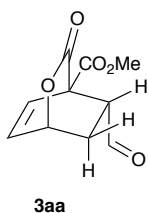
3ae': IR (NaCl) 2955, 1757, 1734, 1285, 1092, 1076 cm^{-1} ; ^1H NMR (600 MHz, CDCl_3) δ 6.98 (1H, brd, $J = 7.8$ Hz), 6.41 (1H, dd, $J = 7.8, 4.8$ Hz), 6.17 (1H, d, $J = 5.4$ Hz), 4.99 (1H, ddd, $J = 4.8, 3.6, 1.8$ Hz), 3.82 (3H, s), 3.55 (1H, dd, $J = 10.2, 5.4$ Hz), 2.88–2.78 (1H, m), 2.68 (6H, s), 0.91 (3H, d, $J = 7.8$ Hz); ^{13}C NMR (150 MHz, CDCl_3) δ 170.3, 168.3, 133.0, 130.6, 128.3, 78.8, 58.3, 52.6, 42.9, 42.8, 36.4, 15.1; HRMS (FAB) m/z calcd for $\text{C}_{13}\text{H}_{19}\text{N}_2\text{O}_4$ $[\text{M}+\text{H}]^+$ 267.1345, found 267.1335.



6-phenyl-*endo*-cycloadduct **3af**

A solution of α -pyrone **1a** (265 mg, 1.72 mmol), cinnamaldehyde hydrazone **2f** (30.0 mg, 0.172 mmol) and $\text{Eu}(\text{hfc})_3$ (20.5 mg, 17.2 μmol) in 1,2-dichloroethane (0.86 mL) was stirred at 80 $^\circ\text{C}$ for 24 hours. After the reaction mixture was concentrated under reduced pressure, the residue was purified by column chromatography on silica gel (hexane : AcOEt = 4 : 1 to 1 : 2) and (hexane : AcOEt = 2 : 1) to afford *endo* adduct **3af** (35.7 mg, 63%) with a trace amount of impurity as a pale yellow solid. Unreacted **1a** (206 mg, 78%) and **2f** (11.0 mg, 37%) were recovered.

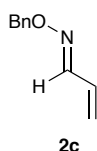
mp 96.2–97.3 $^\circ\text{C}$; IR (KBr) 1759, 1285, 1107, 1074 cm^{-1} ; ^1H NMR (300 MHz, CDCl_3) δ 7.41–7.25 (5H, m), 6.84 (1H, ddd, $J = 7.8, 1.8, 1.2$ Hz), 6.63 (1H, dd, $J = 7.8, 5.1$ Hz), 6.35 (1H, d, $J = 3.6$ Hz), 5.21 (1H, ddd, $J = 5.1, 1.8, 1.2$ Hz), 3.87 (3H, s), 3.52–3.46 (1H, m), 3.32 (1H, dd, $J = 5.1, 1.2$ Hz), 2.70 (6H, s); ^{13}C NMR (75 MHz, CDCl_3) δ 170.3, 168.1, 140.1, 130.9, 130.7, 130.5, 128.9, 127.9, 127.5, 79.0, 59.0, 52.7, 48.1, 45.5, 42.8; HRMS (FAB) m/z calcd for $\text{C}_{18}\text{H}_{21}\text{N}_2\text{O}_4$ $[\text{M}+\text{H}]^+$ 329.1501, found 329.1517.



aldehyde **3aa**

To a solution of hydrazone **3ab** (50.0 mg, 0.198 mmol) in THF (3.9 mL) was added 37% HCHO aq. (81 μ L) and conc. HCl (36%, 64 μ L). After stirring for 2 hours at room temperature, the reaction mixture was added water. The whole was extracted with CH₂Cl₂ ($\times$ 4), and combined organic layer was washed with brine ($\times$ 1). The resulting solution was dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (hexane : AcOEt = 1 : 2) to afford aldehyde **3aa** (41.6 mg, quant) as a white solid.

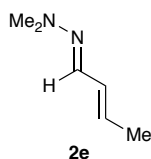
mp 80.0-80.8 °C; IR (KBr) 2926, 1757, 1738, 1711, 1288, 1099, 1076 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 9.49 (1H, s), 6.84 (1H, d, J = 7.8 Hz), 6.52 (1H, dd, J = 7.8, 4.8 Hz), 5.36–5.30 (1H, m), 3.93 (3H, s), 3.62 (1H, dd, J = 10.2, 3.6 Hz), 2.63 (1H, ddd, J = 13.8, 10.2, 3.6 Hz), 2.09 (1H, ddd, J = 13.8, 3.6, 1.2 Hz); ¹³C NMR (150 MHz, CDCl₃) δ 196.6, 169.0, 167.5, 131.8, 131.4, 74.0, 55.7, 53.4, 46.7, 27.9; HRMS (FAB) m/z calcd for C₁₀H₁₁O₅ [M+H]⁺ 211.0606, found 211.0613.



acrolein (*E*)-benzyloxime **2c**

To a solution of *O*-benzylhydroxylamine hydrochloride (5.00 g, 31.3 mmol) and sodium acetate (2.57 g, 31.3 mmol) in MeOH (100 mL) was added acrolein monomer (2.10 mL, 31.5 mmol). After stirring for an hour at room temperature, water was added to the reaction mixture. The whole was extracted with Et₂O (×2), and combined organic layer was washed with water (×1) and brine (×1). The resulting solution was dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (hexane : Et₂O = 15 : 1) to afford oxime **2c** (1.81 g, 36%) as colorless oil.

IR (NaCl) 3065, 2928, 1456, 1366, 1024, 939 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.78 (1H, d, *J* = 10.2 Hz), 7.40–7.26 (5H, m), 6.41 (1H, dt, *J* = 17.4, 10.2 Hz), 5.53 (1H, d, *J* = 10.2 Hz), 5.50 (1H, d, *J* = 17.4 Hz), 5.11 (2H, s); ¹³C NMR (75 MHz, CDCl₃) δ 151.0, 137.3, 130.7, 128.4, 128.2, 127.9, 123.9, 76.1; HRMS (EI) *m/z* calcd for C₁₀H₁₁NO 161.0841, found 161.0845.



crotonaldehyde dimethylhydrazone **2e**

To a solution of 1,1-dimethylhydrazine (3.30 mL, 40.0 mmol) and acetic acid (2.30 mL, 40.0 mmol) in CH₂Cl₂ (40 mL) was added crotonaldehyde (3.30 mL, 40.0 mmol). After stirring for 45 minutes at 0 °C, the reaction mixture was diluted with CH₂Cl₂. The whole was successively washed with water (×1), sat. NaHCO₃ aq. and brine (×1). The resulting solution was dried over MgSO₄, and concentrated under reduced pressure. The residue was distilled (2.0 kPa, 55–58 °C) to afford hydrazone **2e** (2.72 g, 61%, *E* : *Z* = 89 : 11) as colorless oil.

IR (NaCl) 2855, 1568, 1470, 1445, 1269, 1134, 1024, 968 cm⁻¹; ¹H NMR (600 MHz, CDCl₃, signals from the minor isomer are marked with an asterisk) δ 7.25* (1H, d, *J* = 9.6 Hz), 7.01 (1H, d, *J* = 9.0 Hz), 6.20 (1H, dd, *J* = 15.6, 9.0 Hz), 6.14* (1H, dd, *J* = 10.2, 9.6 Hz), 5.82 (1H, dq, *J* = 15.6, 7.2 Hz), 5.62* (1H, dq, *J* = 10.2, 7.2 Hz), 2.87* (6H, s), 2.81 (6H, s), 1.81 (3H+3H*, d, *J* = 7.2 Hz); ¹³C NMR (150 MHz, CDCl₃, weak signals from the minor isomer are marked with an asterisk) δ 137.0, 131.9*, 130.3, 130.2, 127.7*, 126.3*, 42.9, 42.8*, 18.2, 13.5*; HRMS (ESI) *m/z* calcd for C₆H₁₃N₂ [M+H]⁺ 113.1079, found 113.1076.

20170619 1



---- PROCESSING PARAMETERS ----

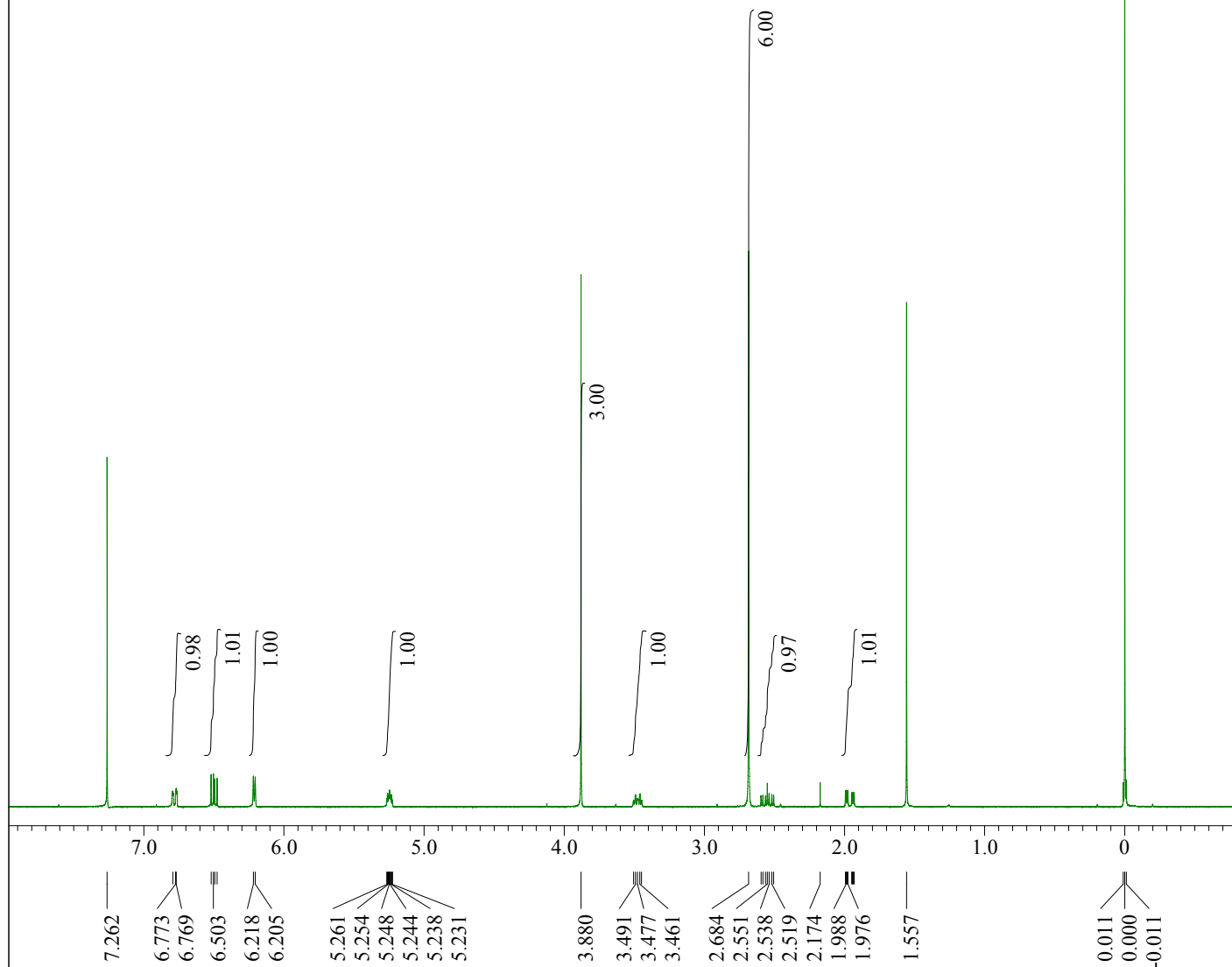
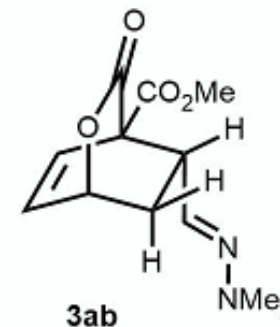
```
dc_balance( 0, FALSE )
sexp( 0.2, 0.0[s] )
trapezoid3( 0[%], 80[%], 100[%] )
zerofill( 1 )
fft( 1, TRUE, TRUE )
ppm
machinephase
```

Filename = Y-Abe_440 p23 9-12-1.jdf
Author = Administrator
Experiment = zg30
Sample_Id = 20170619 1
Solvent = CHLOROFORM-D
Creation_Time = 19-JUN-2017 16:15:28
Revision_Time = 19-JUN-2017 16:17:57
Current_Time = 7-SEP-2017 10:31:10

Comment = 20170619 1
Data_Format = 1D COMPLEX
Dim_Size = 32768
Dim_Title = 1H
Dim_Units = [ppm]
Dimensions = X
Spectrometer = BRUKER_DMX_NMR

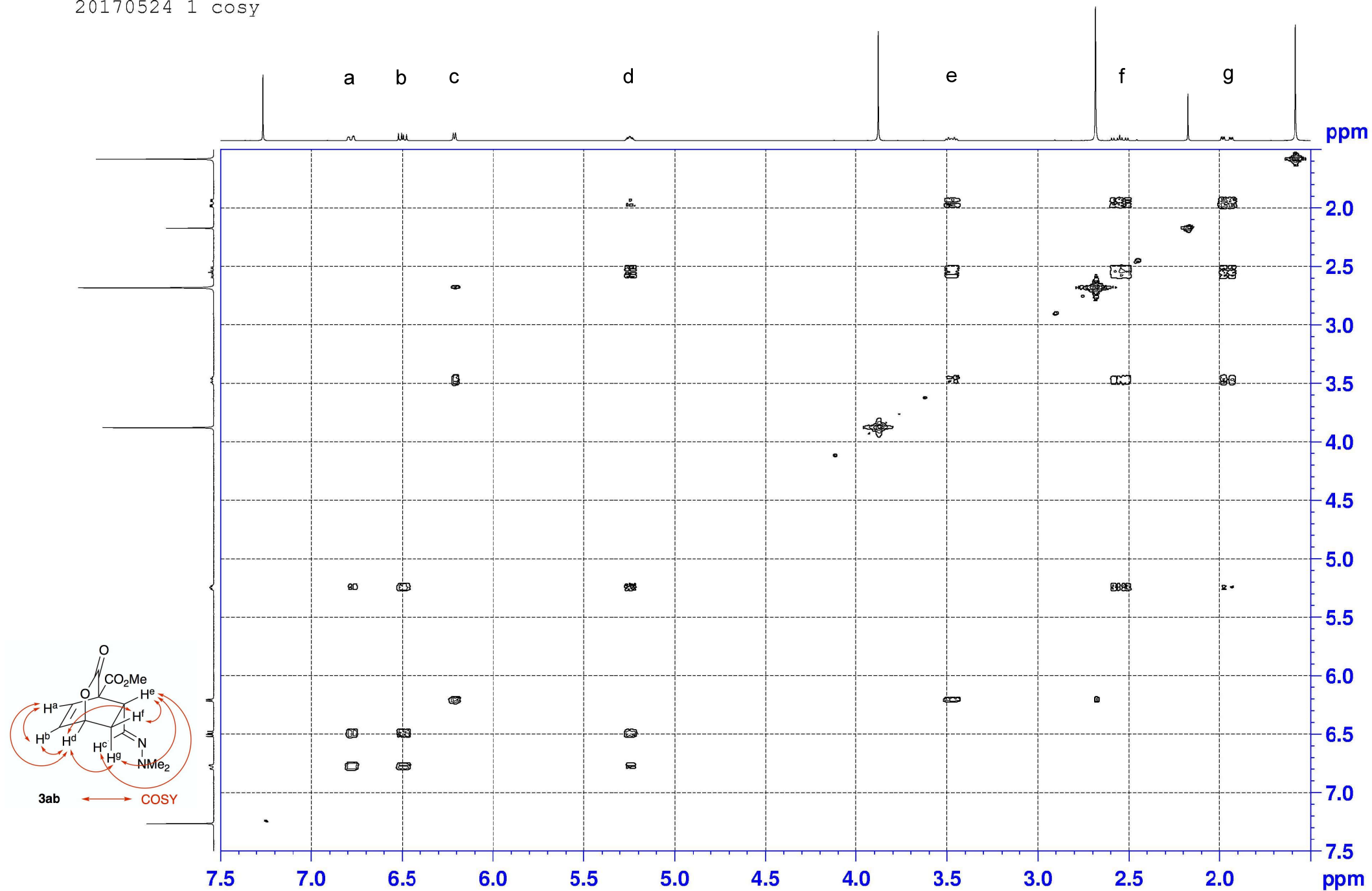
X_Freq = 300.13185343[MHz]
X_Offset = 1.85342561[kHz]
X_Sweep = 6.18811881[kHz]

Temp_Get = 295.66[K]
X_Points = 32768
X_Prescans = 2
Filter_Factor = 3232
Scans = 16



X : ppm : 1H

20170524 1 cosy



20170529 h1



---- PROCESSING PARAMETERS ----

```
dc_balance( 0, FALSE )
sexf( 2.0, 0.0[s] )
trapezoid3( 0[%], 80[%], 100[%] )
zerofill( 1 )
fft( 1, TRUE, TRUE )
ppm
machinephase
```

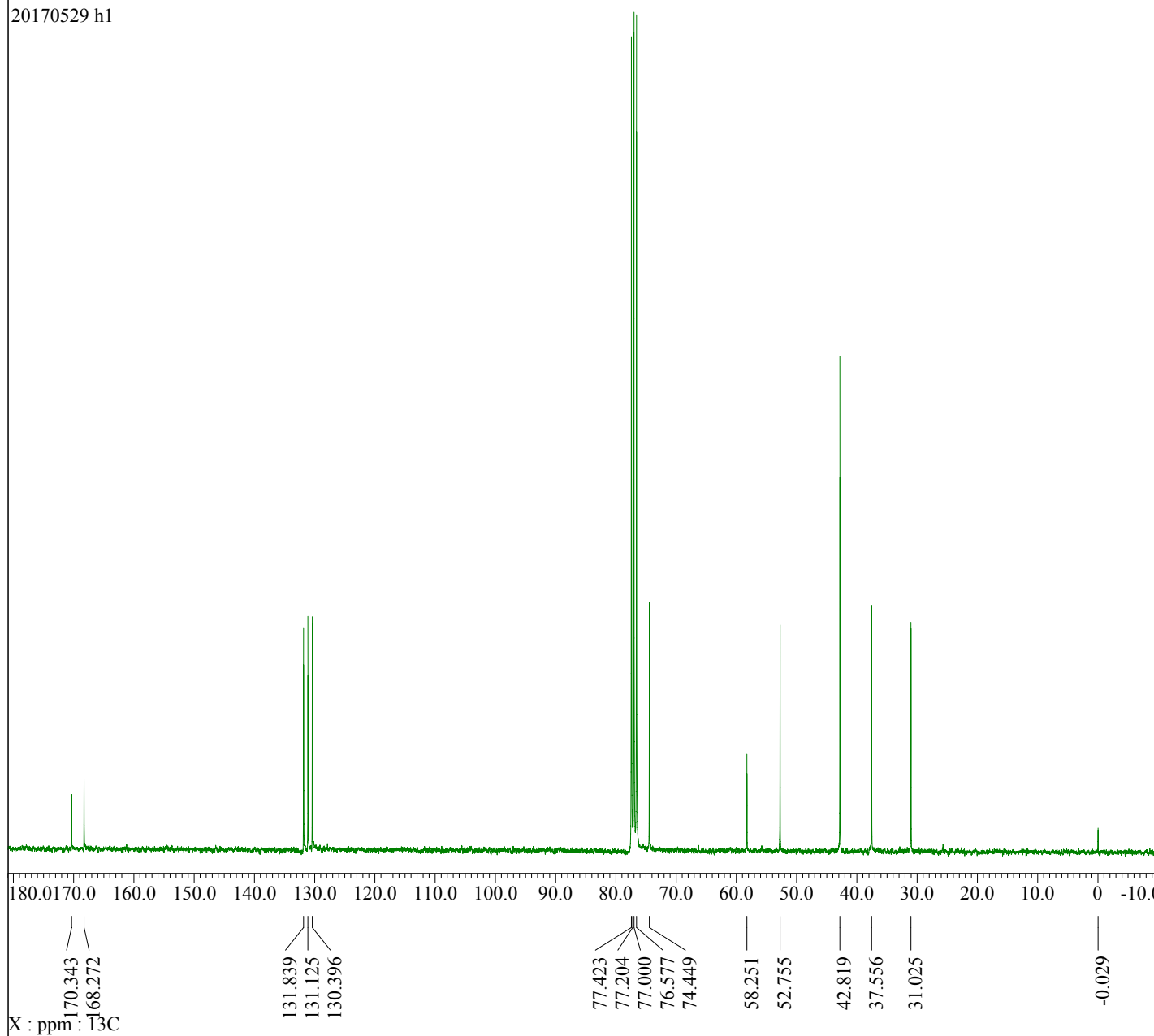
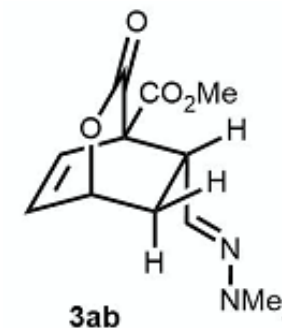
以下に由来: Y-Abe_222-12.jdf

```
Filename      = Y-Abe_222-16.jdf
Author       = Administrator
Experiment    = zgpg30
Sample_Id    = 20170529 h1
Solvent      = CHLOROFORM-D
Creation_Time = 7-SEP-2017 12:32:54
Revision_Time = 7-SEP-2017 12:40:48
Current_Time  = 7-SEP-2017 12:41:10
```

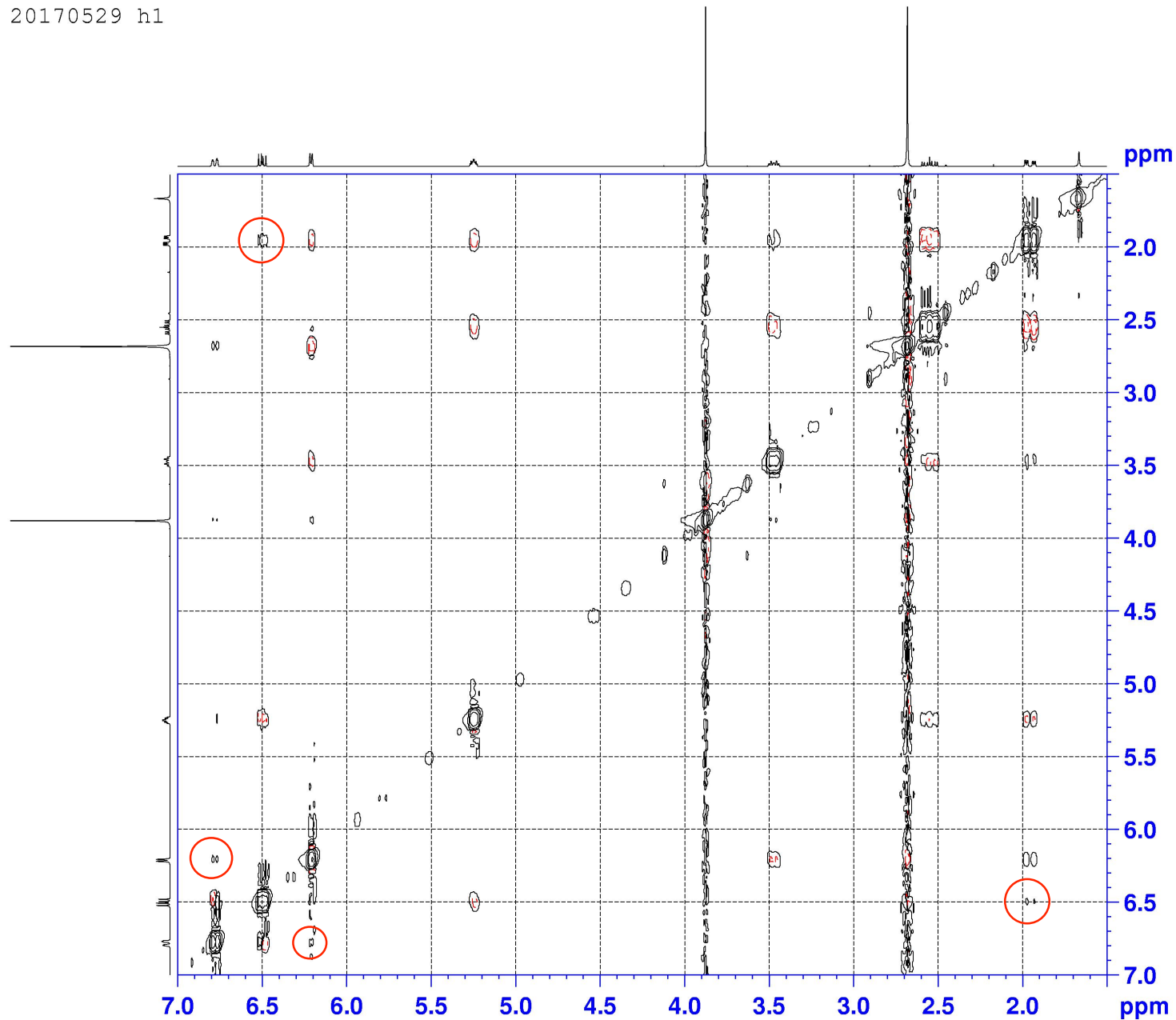
```
Comment      = 20170529 h1
Data_Format   = 1D COMPLEX
Dim_Size      = 32768
Dim_Title     = 13C
Dim_Units     = [ppm]
Dimensions    = X
Spectrometer  = BRUKER_DMX_NMR
```

```
X_Freq       = 75.4752953[MHz]
X_Offset     = 7.54630085[kHz]
X_Sweep      = 18.02884615[kHz]
```

```
Temp_Get     = 297.16[K]
X_Points     = 32768
X_Prescans   = 4
Filter_Factor = 1109
Scans        = 4096
```



20170529 h1

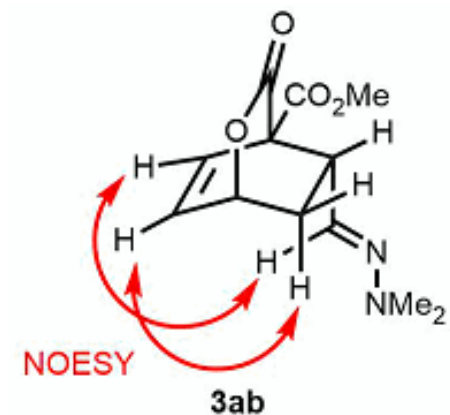


Current Data Parameters
NAME Y-Abe
EXPNO 221
PROCNO 1

F2 - Acquisition Parameters
Date_ 20170529
Time 21.09
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG noesyph
TD 2048
SOLVENT CDCl3
NS 16
DS 4
SWH 2525.252 Hz
FIDRES 1.233033 Hz
AQ 0.4055040 sec
RG 32
DW 198.000 usec
DE 6.50 usec
TE 295.5 K
D0 0.00017922 sec
D1 1.92872906 sec
D8 0.89999998 sec
IN0 0.00039600 sec

===== CHANNEL f1 =====
SFO1 300.1311040 MHz
NUC1 1H
P1 14.75 usec
PLW1 6.19999981 W

F1 - Acquisition parameters
TD 256
SFO1 300.1311 MHz
FIDRES 9.864267 Hz
SW 8.414 ppm
FnMODE States-TPPI



20170619 2



---- PROCESSING PARAMETERS ----

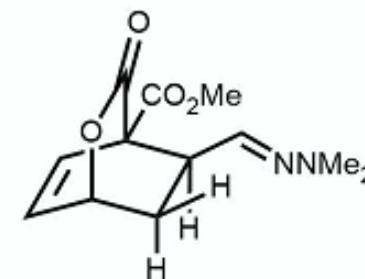
```
dc_balance( 0, FALSE )
sexp( 0.2, 0.0[s] )
trapezoid3( 0[%], 80[%], 100[%] )
zerofill( 1 )
fft( 1, TRUE, TRUE )
ppm
machinephase
```

Filename = Y-Abe_450 p23 14-17-2.jdf
Author = Administrator
Experiment = zg30
Sample_Id = 20170619 2
Solvent = CHLOROFORM-D
Creation_Time = 19-JUN-2017 16:20:25
Revision_Time = 7-SEP-2017 12:28:02
Current_Time = 7-SEP-2017 12:28:24

Comment = 20170619 2
Data_Format = 1D COMPLEX
Dim_Size = 32768
Dim_Title = 1H
Dim_Units = [ppm]
Dimensions = X
Spectrometer = BRUKER_DMX_NMR

X_Freq = 300.13185343 [MHz]
X_Offset = 1.85342561 [kHz]
X_Sweep = 6.18811881 [kHz]

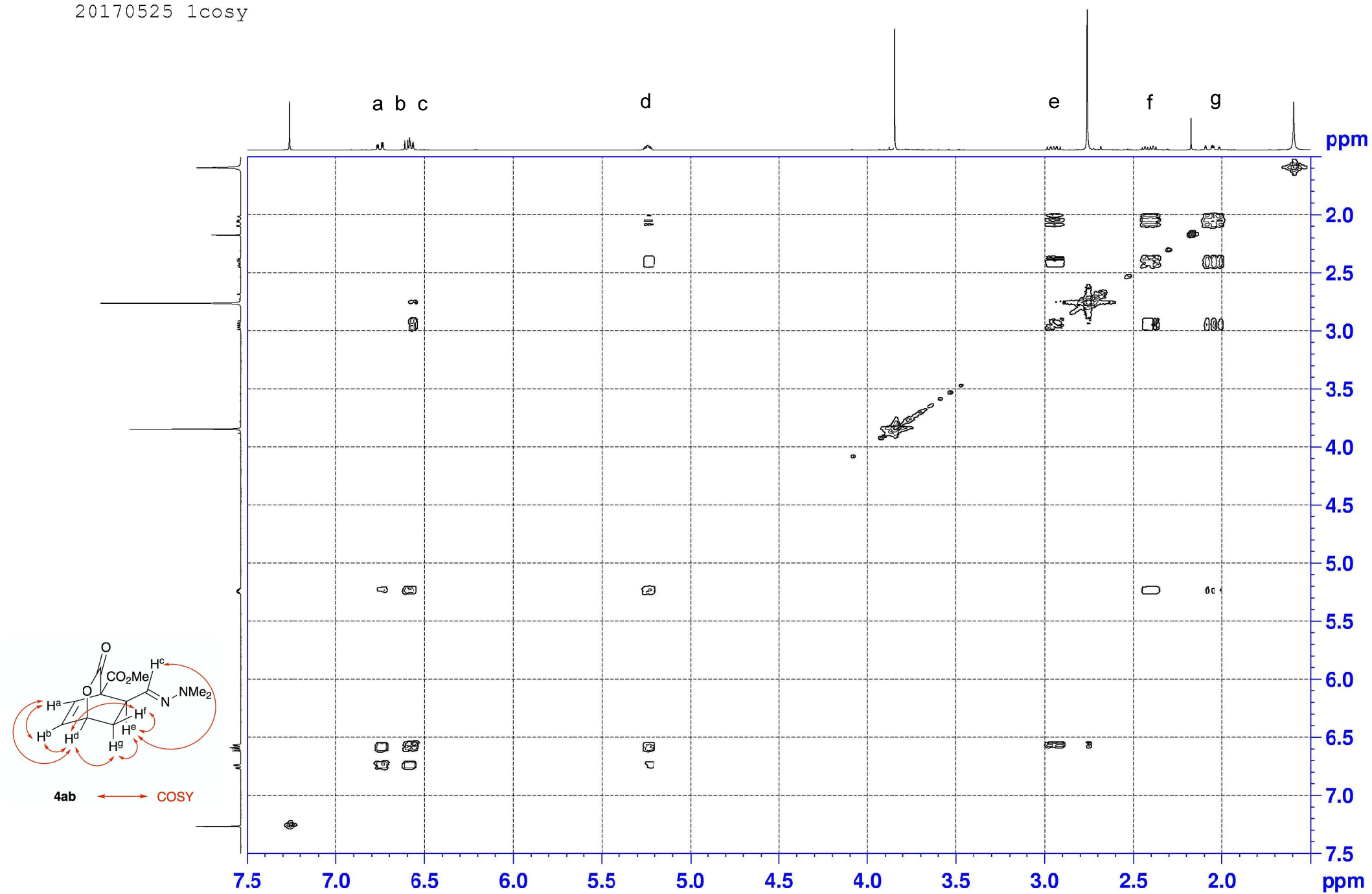
Temp_Get = 295.56 [K]
X_Points = 32768
X_Prescans = 2
Filter_Factor = 3232
Scans = 16



4ab

X : ppm : 1H

20170525 1cosy



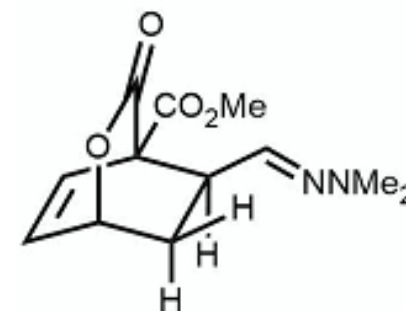
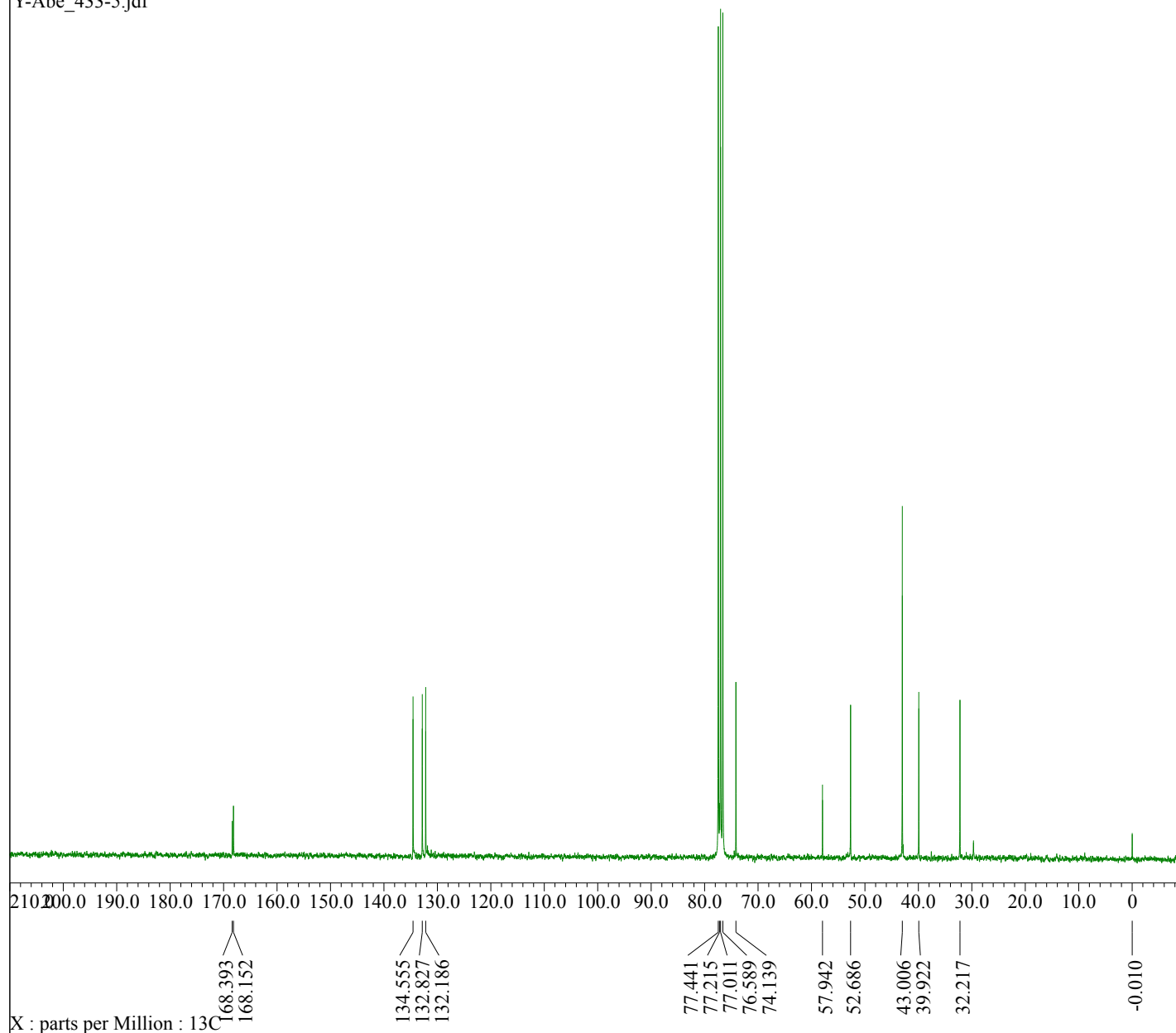
Y-Abe_433-5.jdf

Filename = Y-Abe_433-5.jdf
Author = Administrator
Experiment = zgpg30
Sample Id = Parameter file, TOPSPIN Vers
Solvent = CDCl3
Creation Time = 21-JUL-2018 14:11:55
Revision Time = 21-JUL-2018 14:12:58
Current Time = 21-JUL-2018 14:16:03

Comment = Parameter file, TOPSPIN Vers
Data Format = 1D COMPLEX
Dim Size = 32768
Dim Title = 13C
Dim Units = [ppm]
Dimensions = X
Spectrometer = BRUKER DMX_NMR

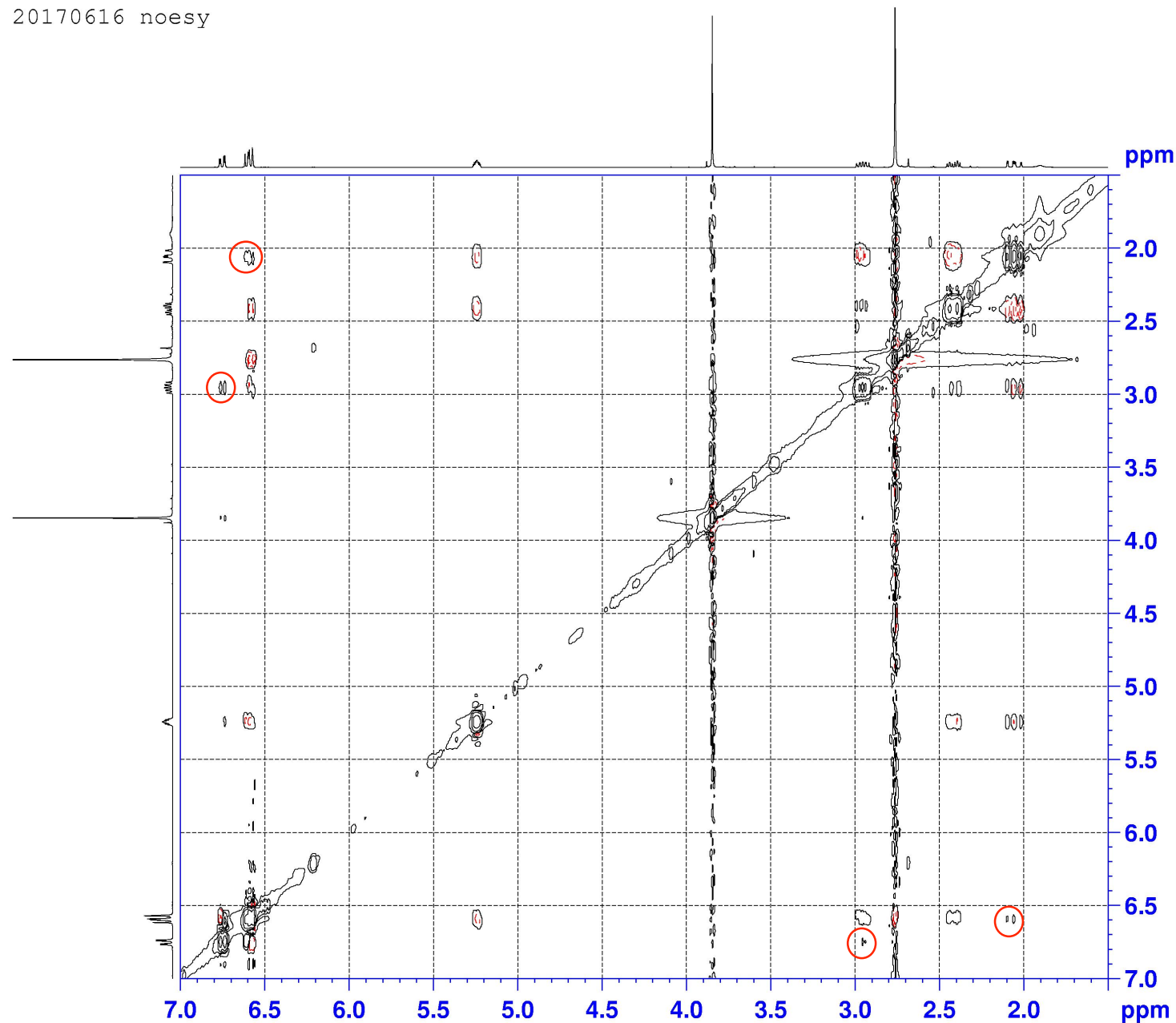
X Freq = 75.4752953 [MHz]
X Offset = 7.54630085 [kHz]
X Sweep = 18.02884615 [kHz]

Temp Get = 297.06 [K]
X Points = 32768
X Prescans = 4
Filter Factor = 1109
Scans = 4096



4ab

20170616 noesy

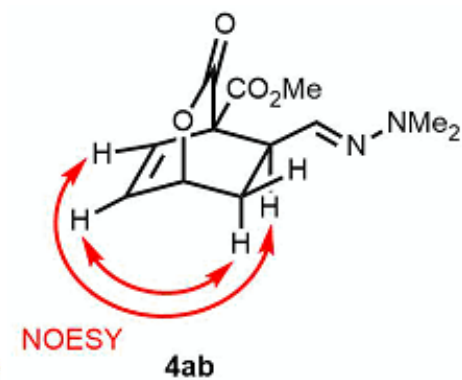


Current Data Parameters
NAME Y-Abe
EXPNO 432
PROCNO 1

F2 - Acquisition Parameters
Date_ 20170616
Time 21.20
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG noesyph
TD 2048
SOLVENT CDC13
NS 16
DS 4
SWH 2577.320 Hz
FIDRES 1.258457 Hz
AQ 0.3973120 sec
RG 32
DW 194.000 usec
DE 6.50 usec
TE 295.5 K
D0 0.00017522 sec
D1 1.93692100 sec
D8 0.89999998 sec
IN0 0.00038800 sec

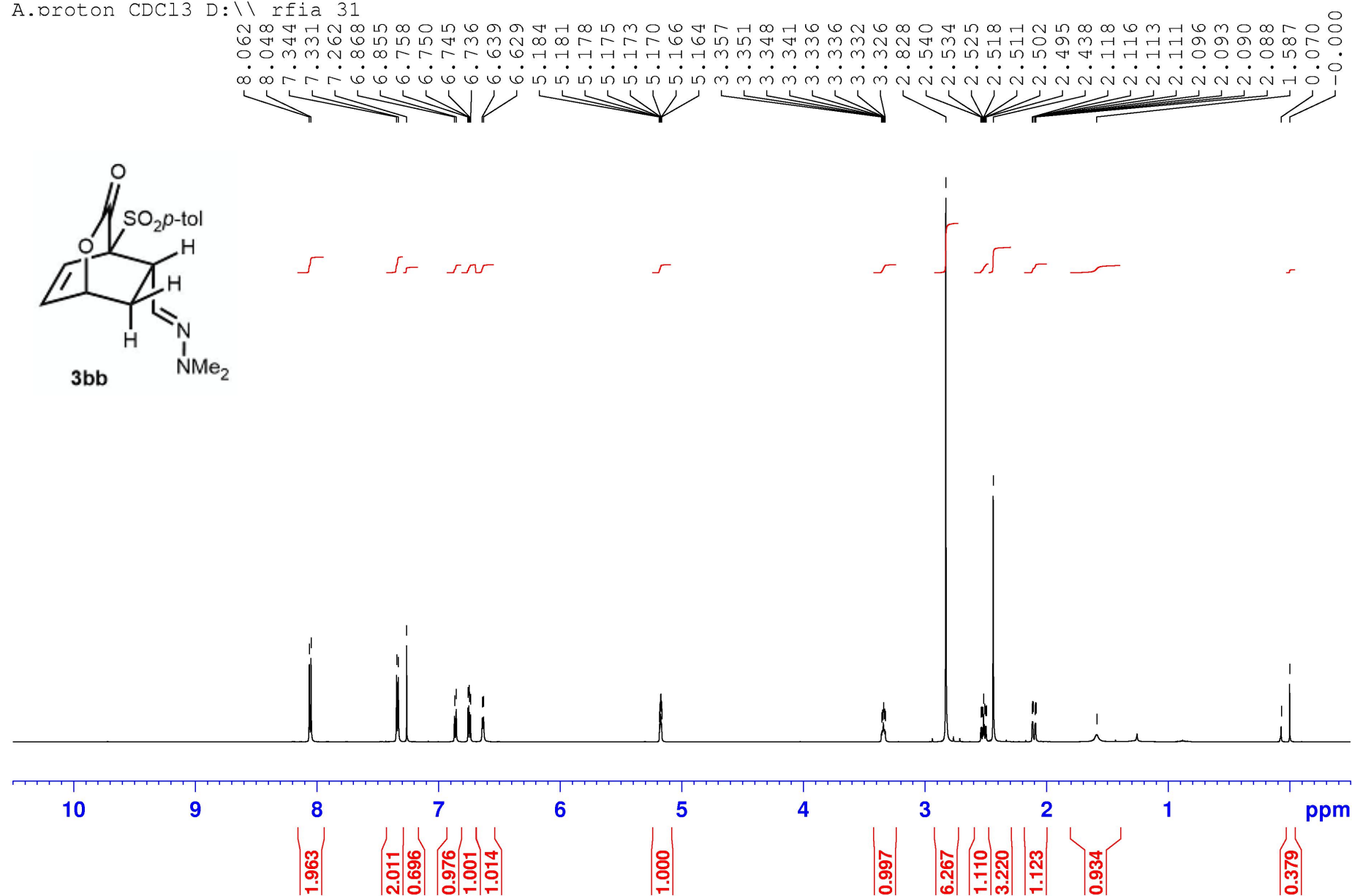
===== CHANNEL f1 =====
SFO1 300.1310959 MHz
NUC1 1H
P1 14.75 usec
PLW1 6.19999981 W

F1 - Acquisition parameters
TD 256
SFO1 300.1311 MHz
FIDRES 10.067655 Hz
SW 8.587 ppm
FnMODE States-TPPI

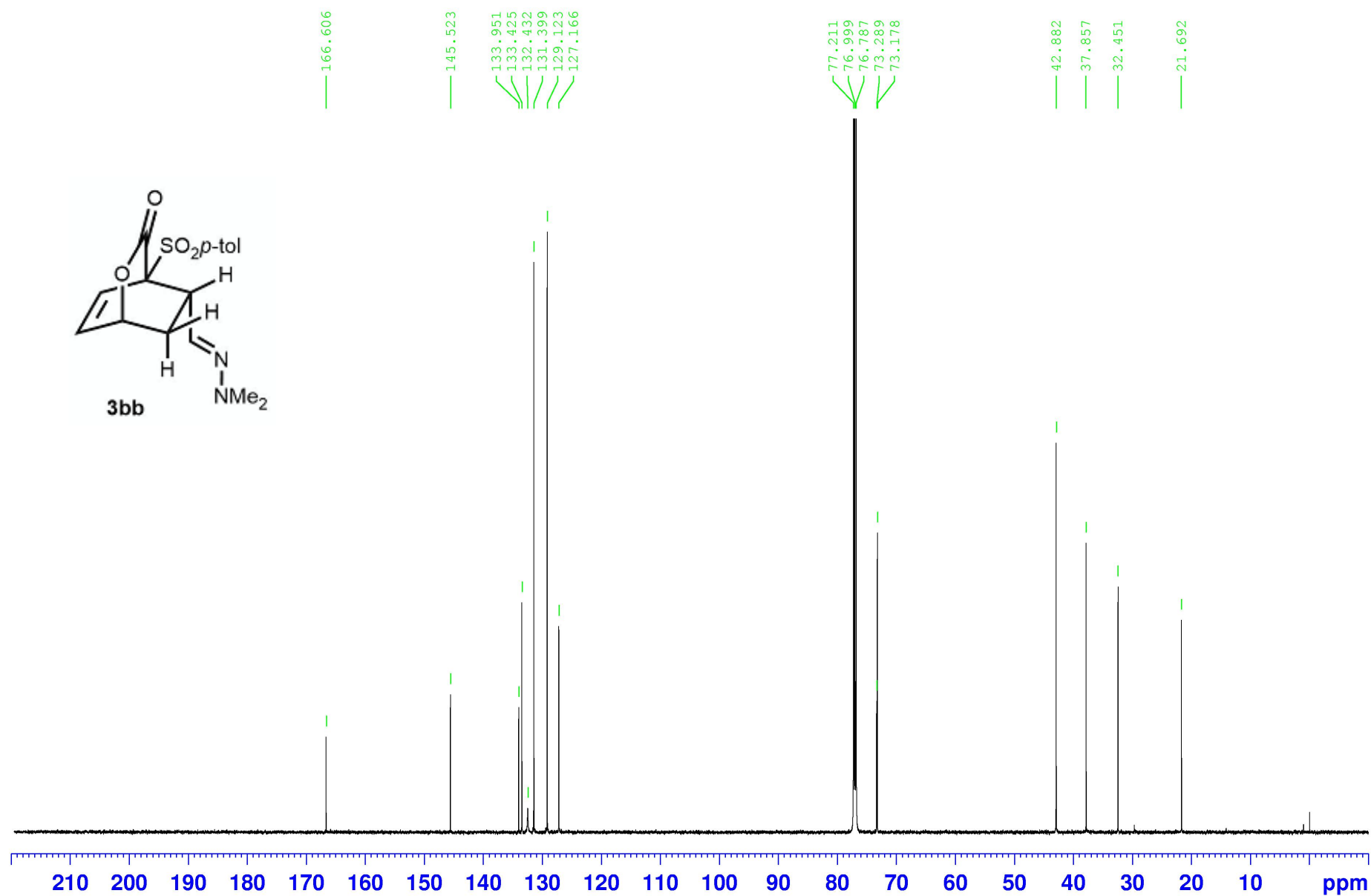
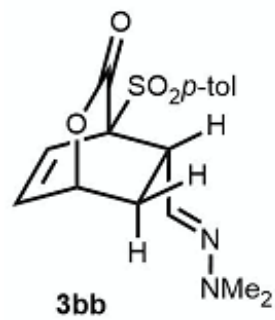


YH_0329_1and6

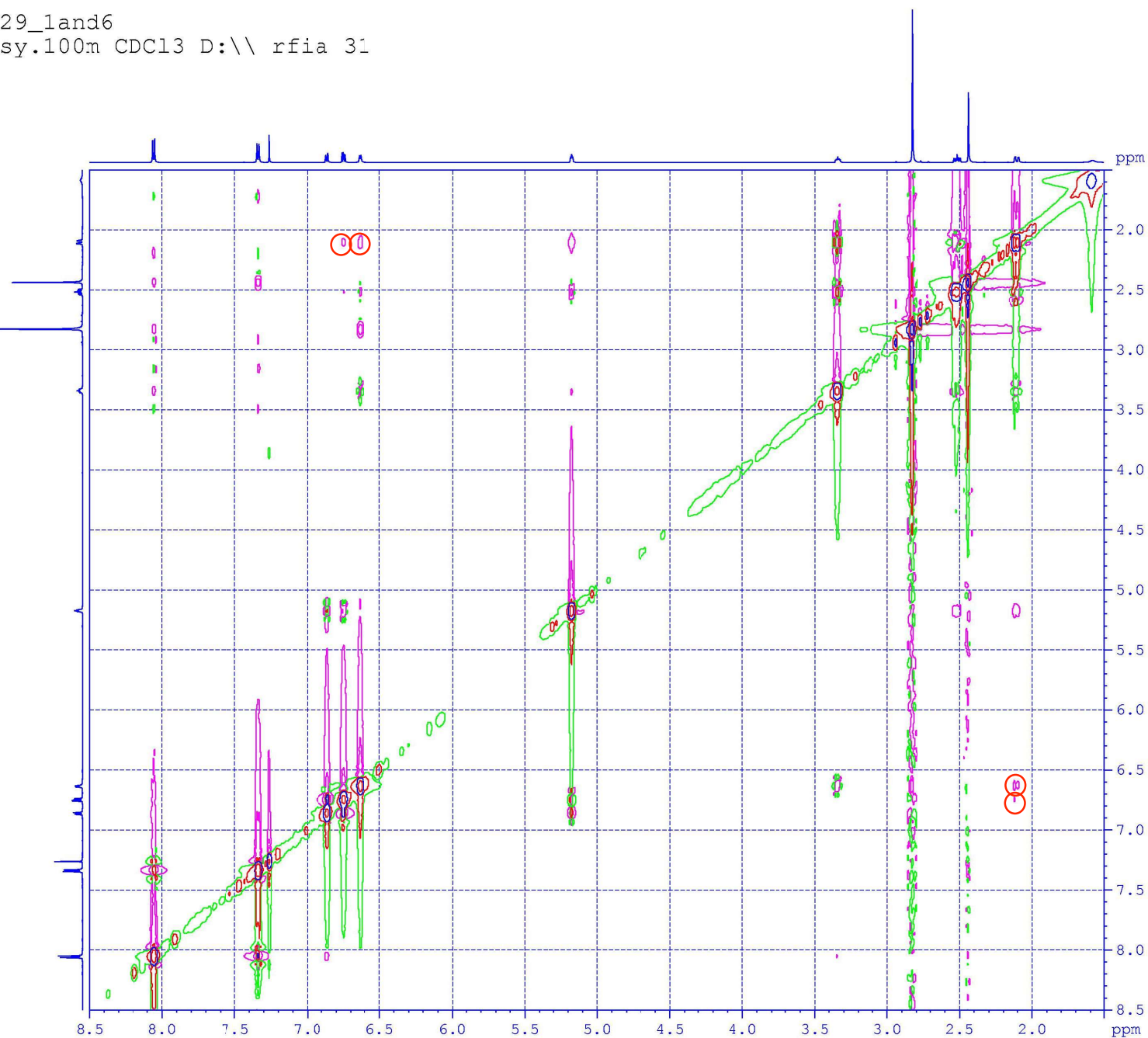
A.proton CDC13 D:\\ rfia 31



YH_0329_1and6
A.carbon CDCl3 D:\\ rfia 31



YH_0329_1and6
B.noesy.100m CDC13 D:\ rfia 31



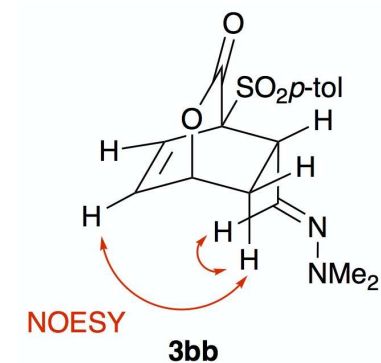
Current Data Parameters
NAME Hashimoto
EXPNO 1532
PROCNO 1

F2 - Acquisition Parameters
Date_ 20180214
Time 6.40
INSTRUM spect
PROBHD 5 mm CPTCI 1H-
PULPROG noesygpph
TD 2048
SOLVENT CDC13
NS 32
DS 16
SWH 5681.818 Hz
FIDRES 2.774325 Hz
AQ 0.1802240 sec
RG 20.2
DW 88.000 usec
DE 10.00 usec
TE 298.0 K
D0 0.00007807 sec
D1 1.98689306 sec
D8 0.10000000 sec
D11 0.03000000 sec
D12 0.00002000 sec
D16 0.00020000 sec
IN0 0.00017600 sec

===== CHANNEL f1 =====
SFO1 600.1324381 MHz
NUC1 1H
P1 7.80 usec
P2 15.60 usec
P17 2500.00 usec
PLW1 8.80000019 W
PLW10 0.79200000 W

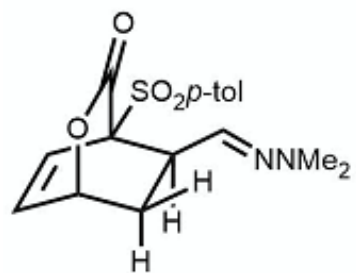
===== GRADIENT CHANNEL =====
GPNAM[1] SINE.100
GPZ1 40.00 %
P16 1000.00 usec

F1 - Acquisition parameters
TD 256
SFO1 600.1324 MHz
FIDRES 22.194603 Hz
SW 9.468 ppm
FnMODE States-TPP1

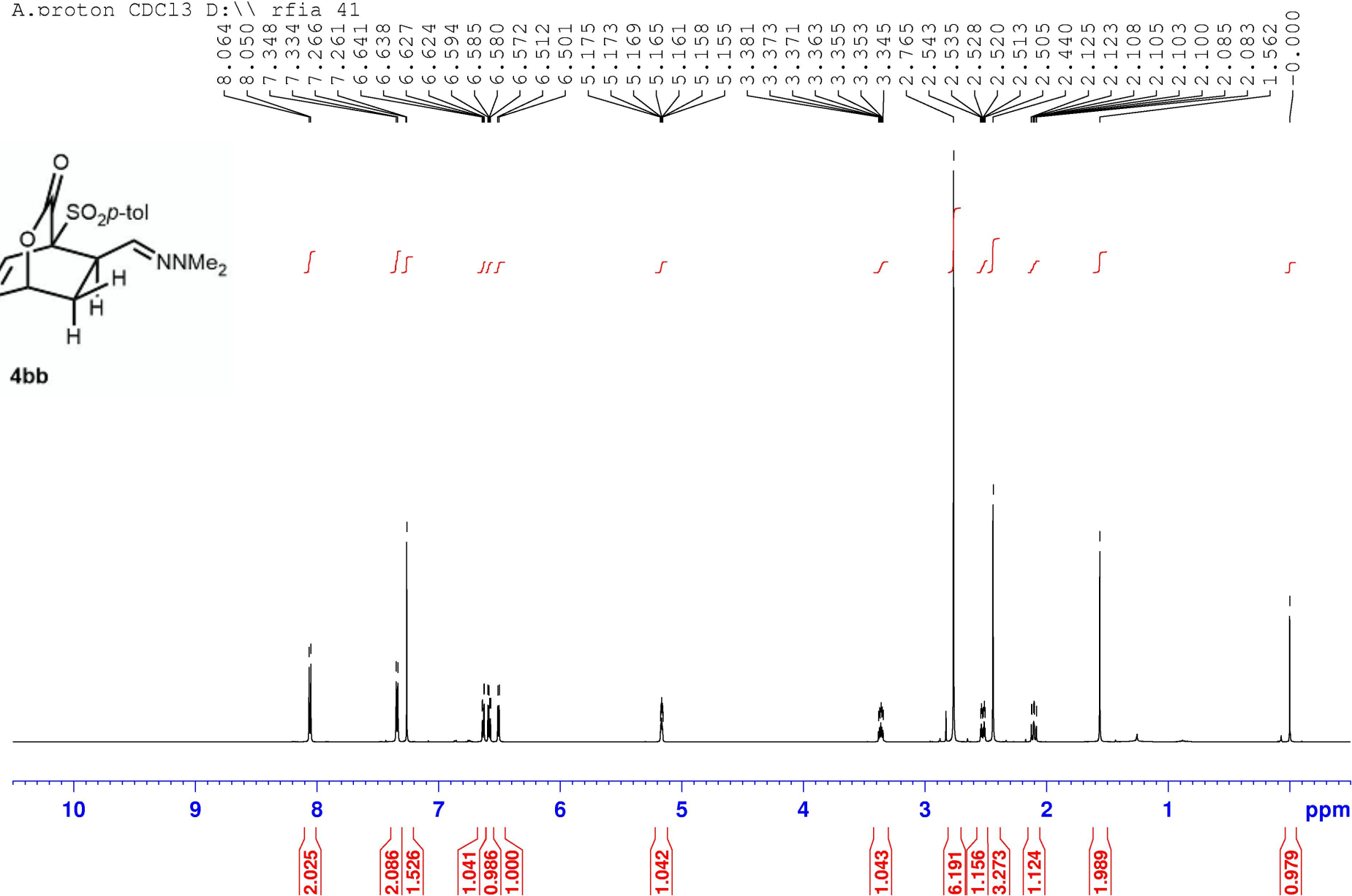


YH_0329_7

A.proton CDC13 D:\ rfia 41

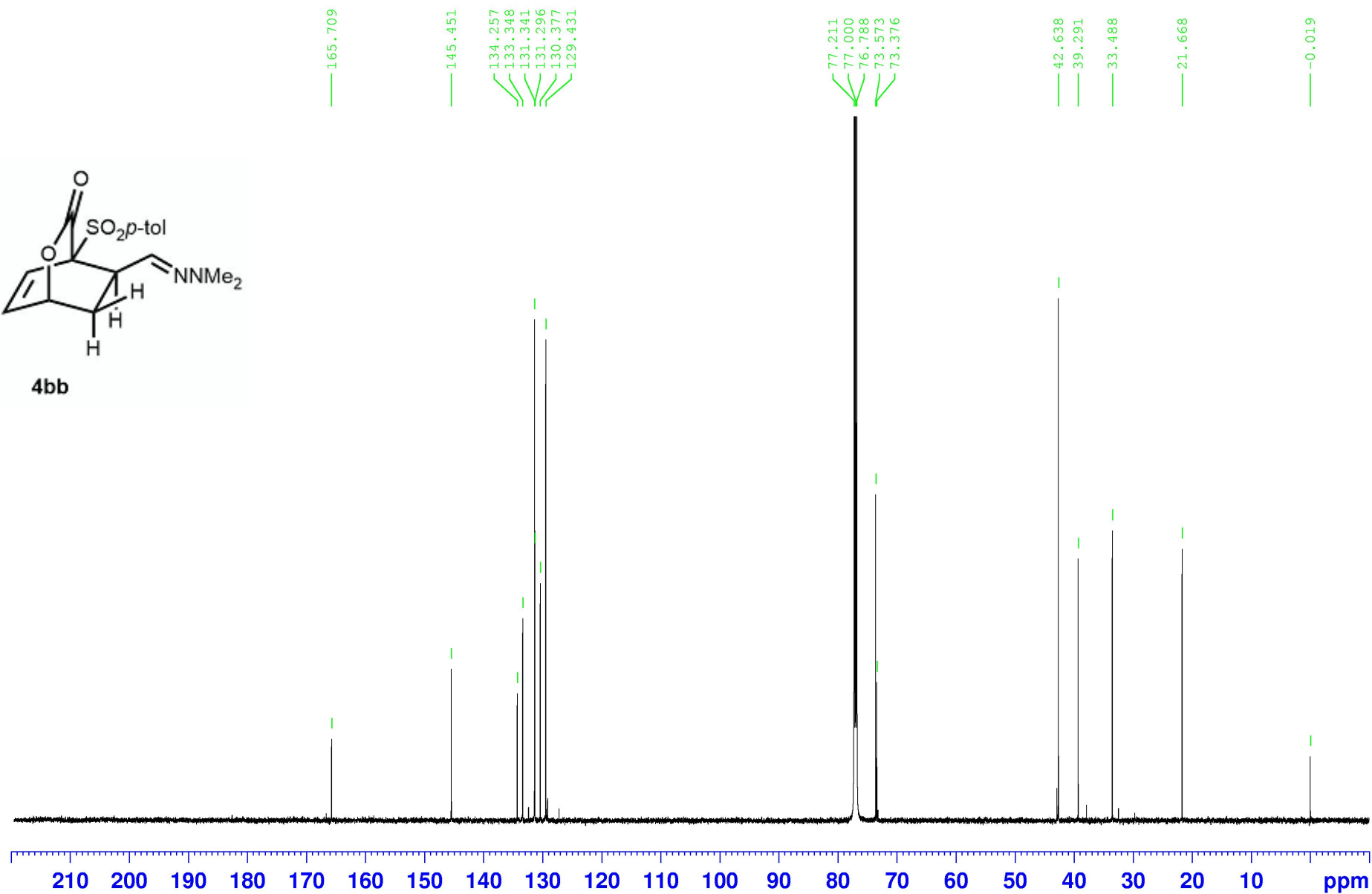
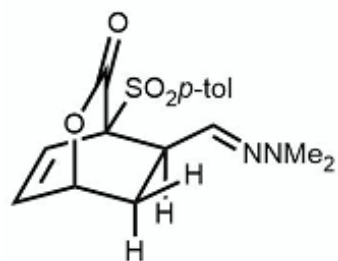


4bb

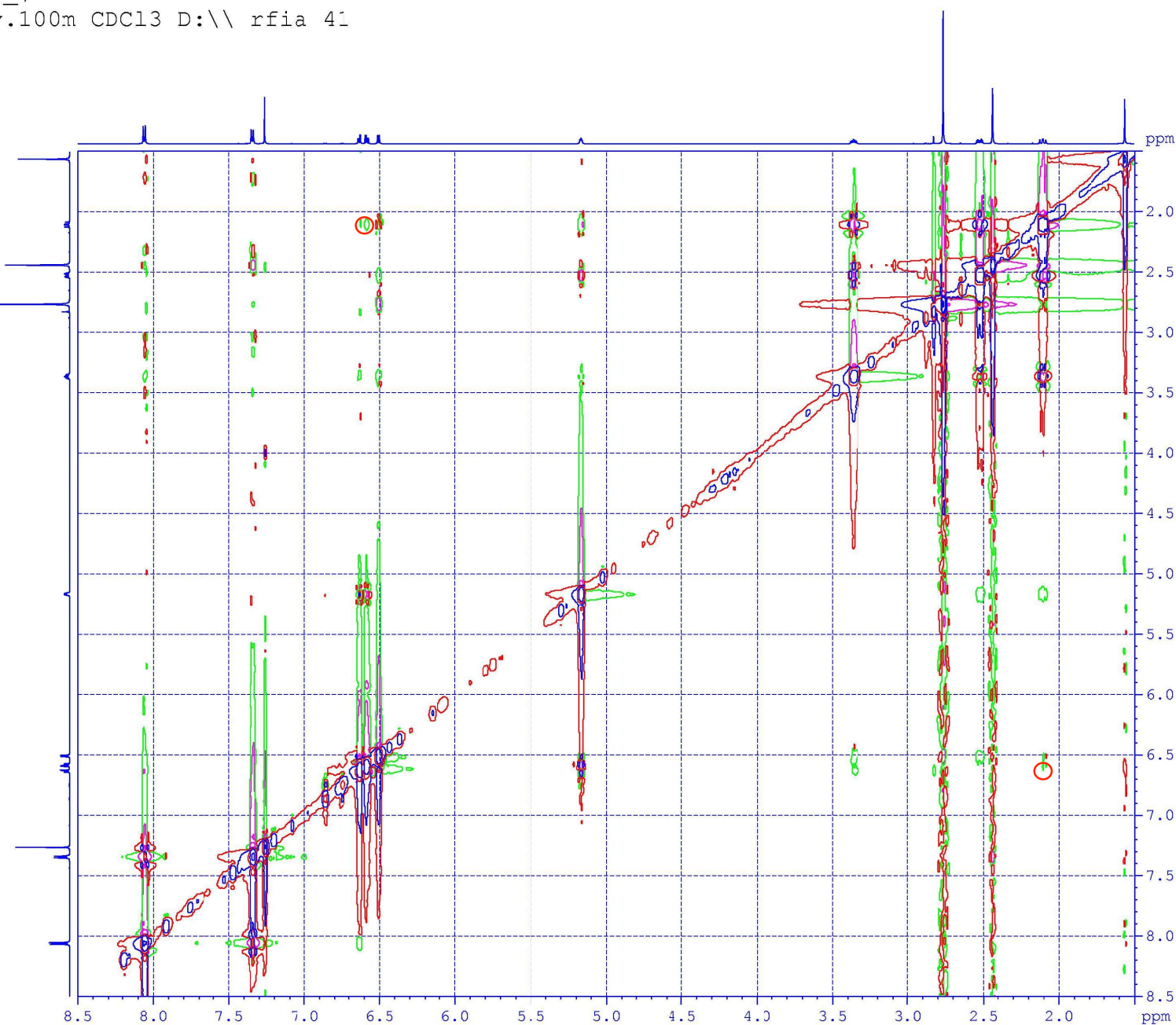


YH_0329_7

A.carbon CDCl₃ D:\\ rfia 41



YH_0329_7
B.noesy.100m CDC13 D:\\ rfia 41



Current Data Parameters
NAME Hashimoto
EXPNO 1522
PROCNO 1

F2 - Acquisition Parameters
Date_ 20180213
Time 15.31
INSTRUM spect
PROBHD 5 mm CPTCI 1H-
PULPROG noesygpph
TE 2048
SOLVENT CDC13
NS 32
DS 16
SWH 5681.818 Hz
FIDRES 2.774325 Hz
AQ 0.1802240 sec
RG 40.3
DW 88.000 usec
DE 10.00 usec
TE 298.0 K
DC 0.00007807 sec
D1 1.98689305 sec
D8 0.10000000 sec
D11 0.03000000 sec
D12 0.00002000 sec
D16 0.00020000 sec
IN0 0.00017600 sec

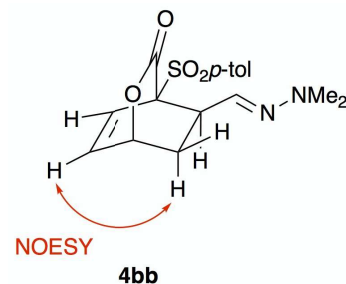
===== CHANNEL f1 =====
SF01 600.1324588 MHz
NUC1 1H
P1 7.80 usec
P2 15.60 usec
P17 2500.00 usec
PLW1 8.80000019 W
PLW10 0.79200000 W

===== GRADIENT CHANNEL =====
GENAM[1] SINE.100
GEZ1 40.00 %
P16 1000.00 usec

F1 - Acquisition parameters
TE 256
SF01 600.1325 MHz
FIDRES 22.194603 Hz
SW 9.468 ppm
PRMODE States-TPPI

F2 - Processing parameters
SI 1024
SF 600.1300099 MHz
WDW QSINE
SSB 2
LB 0 Hz
GB 0
PC 1.00

F1 - Processing parameters
SI 1024
MC2 States-TPPI
SF 600.1300099 MHz
WDW QSINE
SSB 2
LB 0 Hz
GB 0



20170804 4



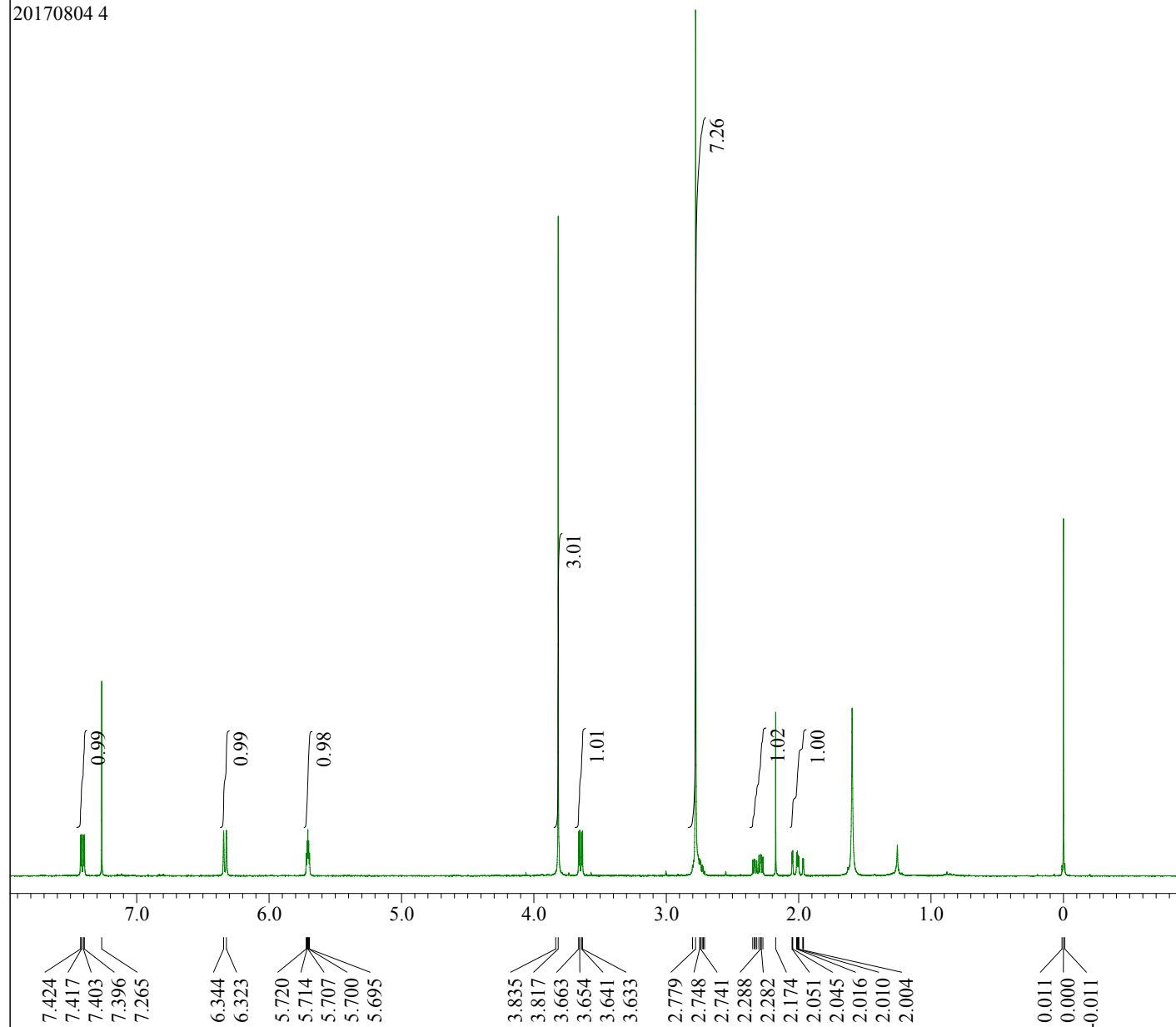
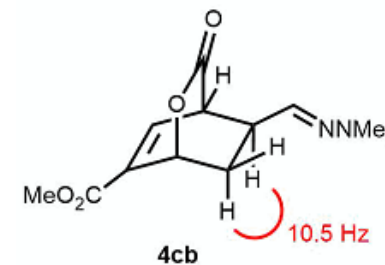
---- PROCESSING PARAMETERS ----
dc_balance(0, FALSE)
sexp(0.2, 0.0[s])
trapezoid3(0[%], 80[%], 100[%])
zerofill(1)
fft(1, TRUE, TRUE)
ppm
machinephase

Filename = Y-Abe_1180 honmonohonmono-2.
Author = Administrator
Experiment = zg30
Sample_Id = 20170804 4
Solvent = CHLOROFORM-D
Creation_Time = 5-AUG-2017 11:35:21
Revision_Time = 7-SEP-2017 14:39:45
Current_Time = 7-SEP-2017 14:40:26

Comment = 20170804 4
Data_Format = 1D COMPLEX
Dim_Size = 32768
Dim_Title = 1H
Dim_Units = [ppm]
Dimensions = X
Spectrometer = BRUKER_DMX_NMR

X_Freq = 300.13185343[MHz]
X_Offset = 1.85342561[kHz]
X_Sweep = 6.18811881[kHz]

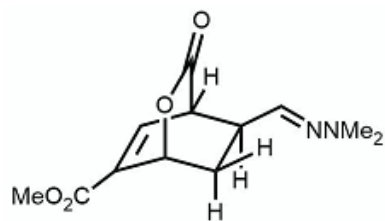
Temp_Get = 296.16[K]
X_Points = 32768
X_Prescans = 2
Filter_Factor = 3232
Scans = 16



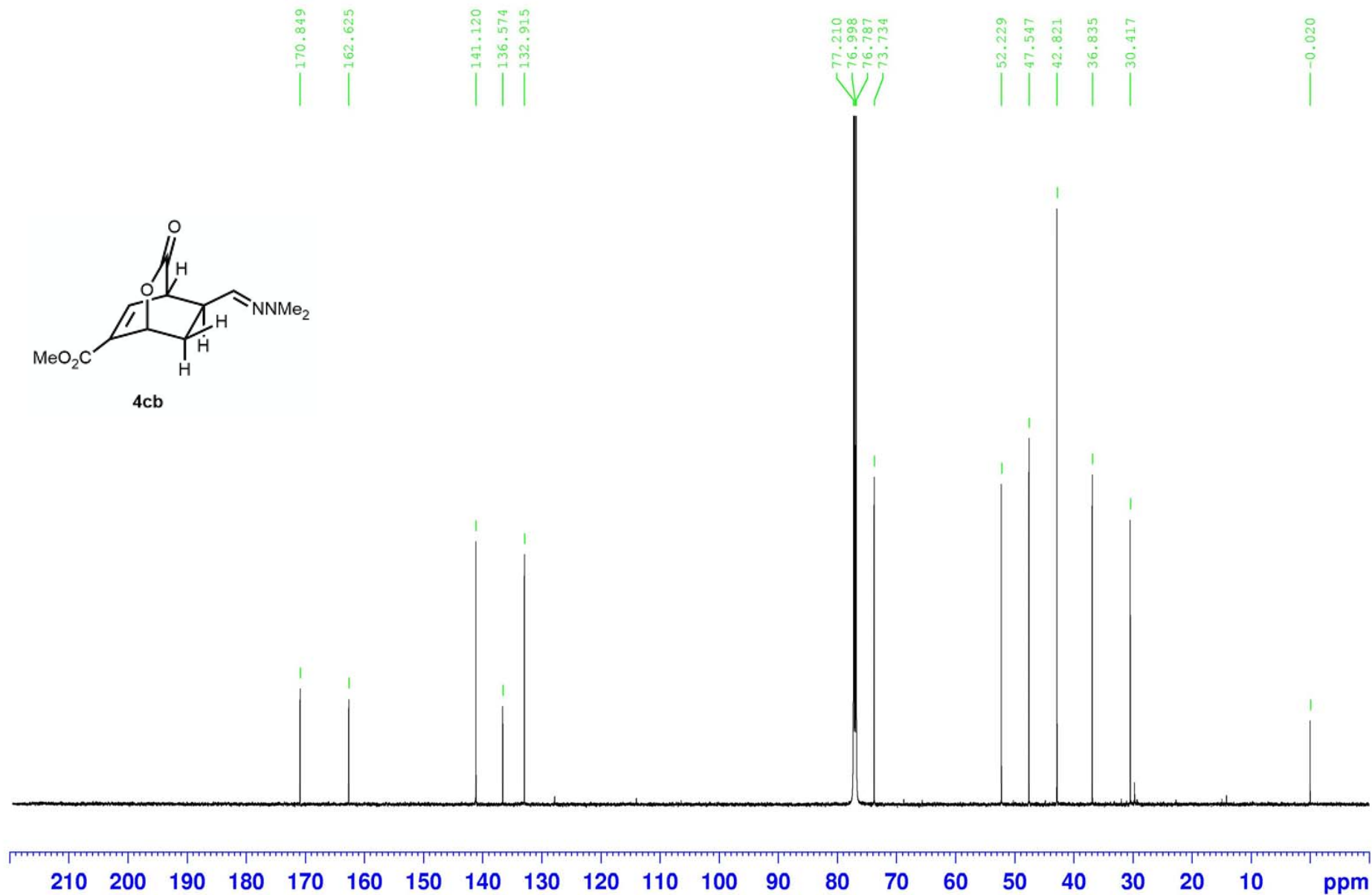
X : ppm : 1H

abe_no3_p43_3_12_15

A.carbon CDCl3 D:\ rfia 41



4cb



20170803 1



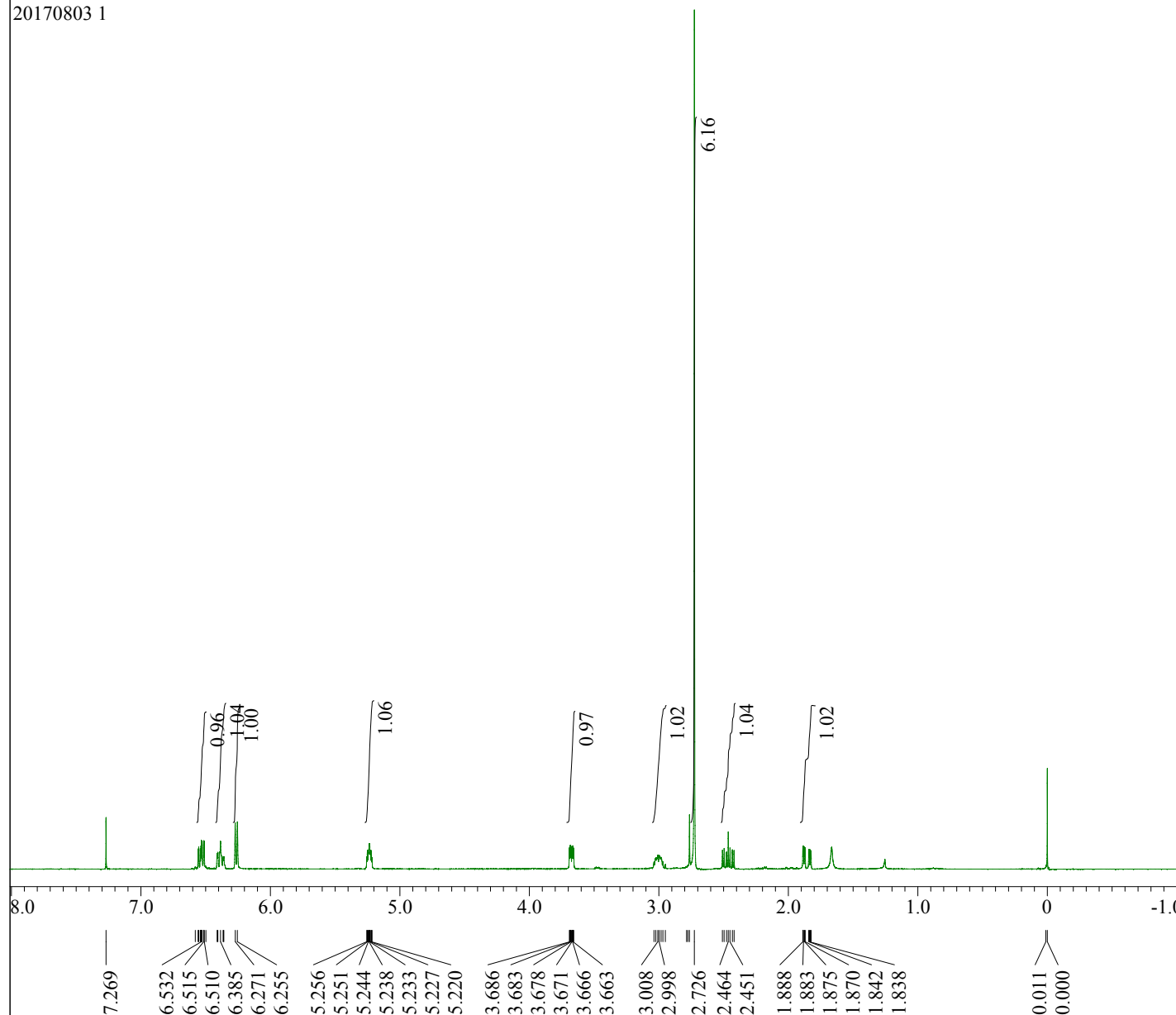
---- PROCESSING PARAMETERS ----
dc_balance(0, FALSE)
sexf(0.2, 0.0[s])
trapezoid3(0[%], 80[%], 100[%])
zerofill(1)
fft(1, TRUE, TRUE)
ppm
machinephase

Filename = Y-Abe_1110-2.jdf
Author = Administrator
Experiment = zg30
Sample_Id = 20170803 1
Solvent = CHLOROFORM-D
Creation_Time = 3-AUG-2017 13:45:51
Revision_Time = 7-SEP-2017 14:44:06
Current_Time = 7-SEP-2017 14:44:17

Comment = 20170803 1
Data_Format = 1D COMPLEX
Dim_Size = 32768
Dim_Title = 1H
Dim_Units = [ppm]
Dimensions = X
Spectrometer = BRUKER DMX NMR

X_Freq = 300.13185343[MHz]
X_Offset = 1.85342561[kHz]
X_Sweep = 6.18811881[kHz]

Temp_Get = 295.86[K]
X_Points = 32768
X_Prescans = 2
Filter_Factor = 3232
Scans = 16

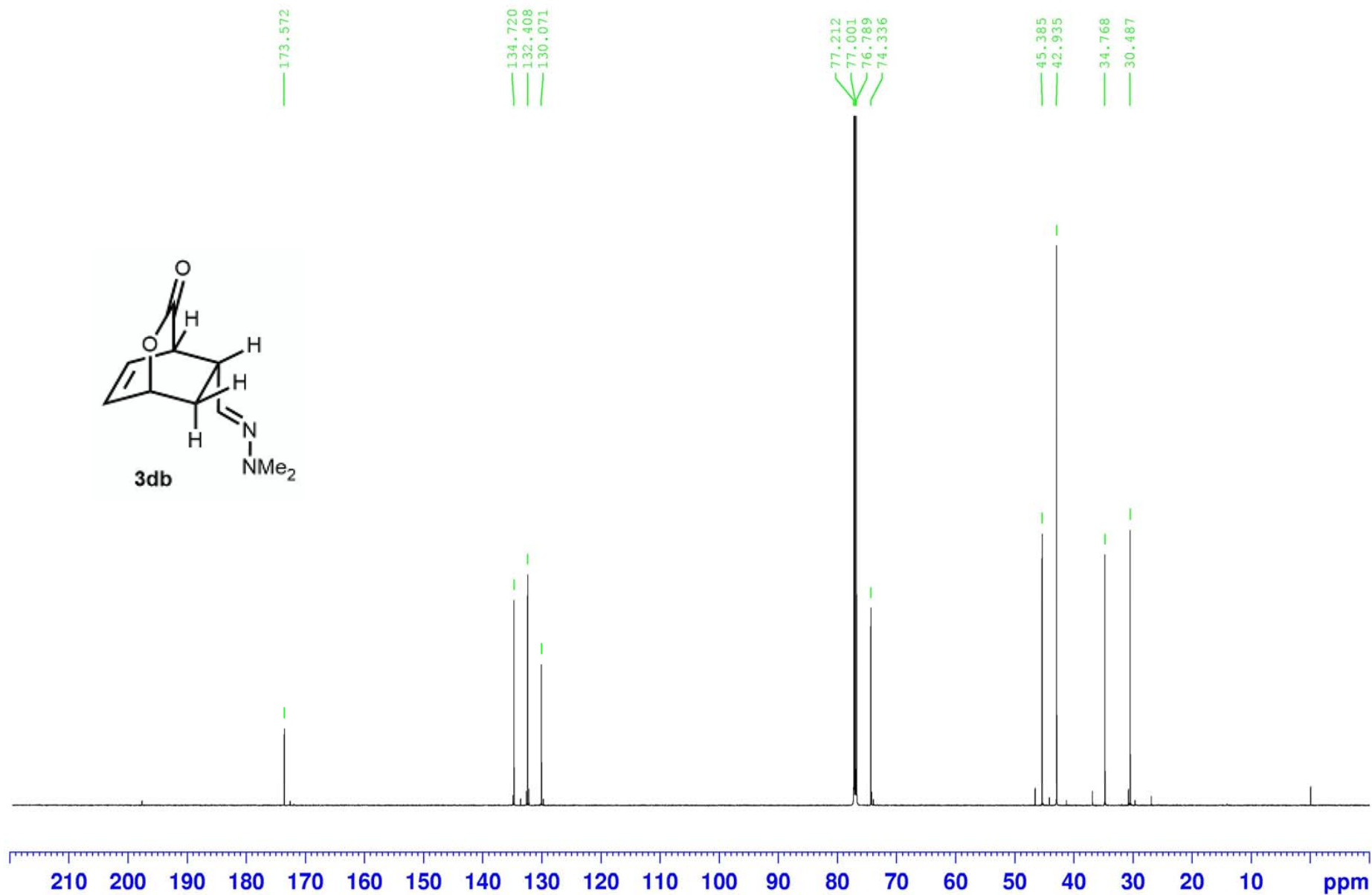
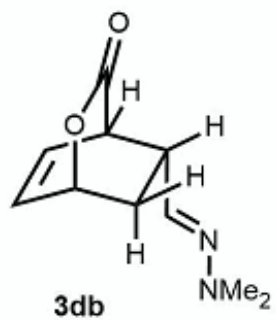


X : ppm : 1H

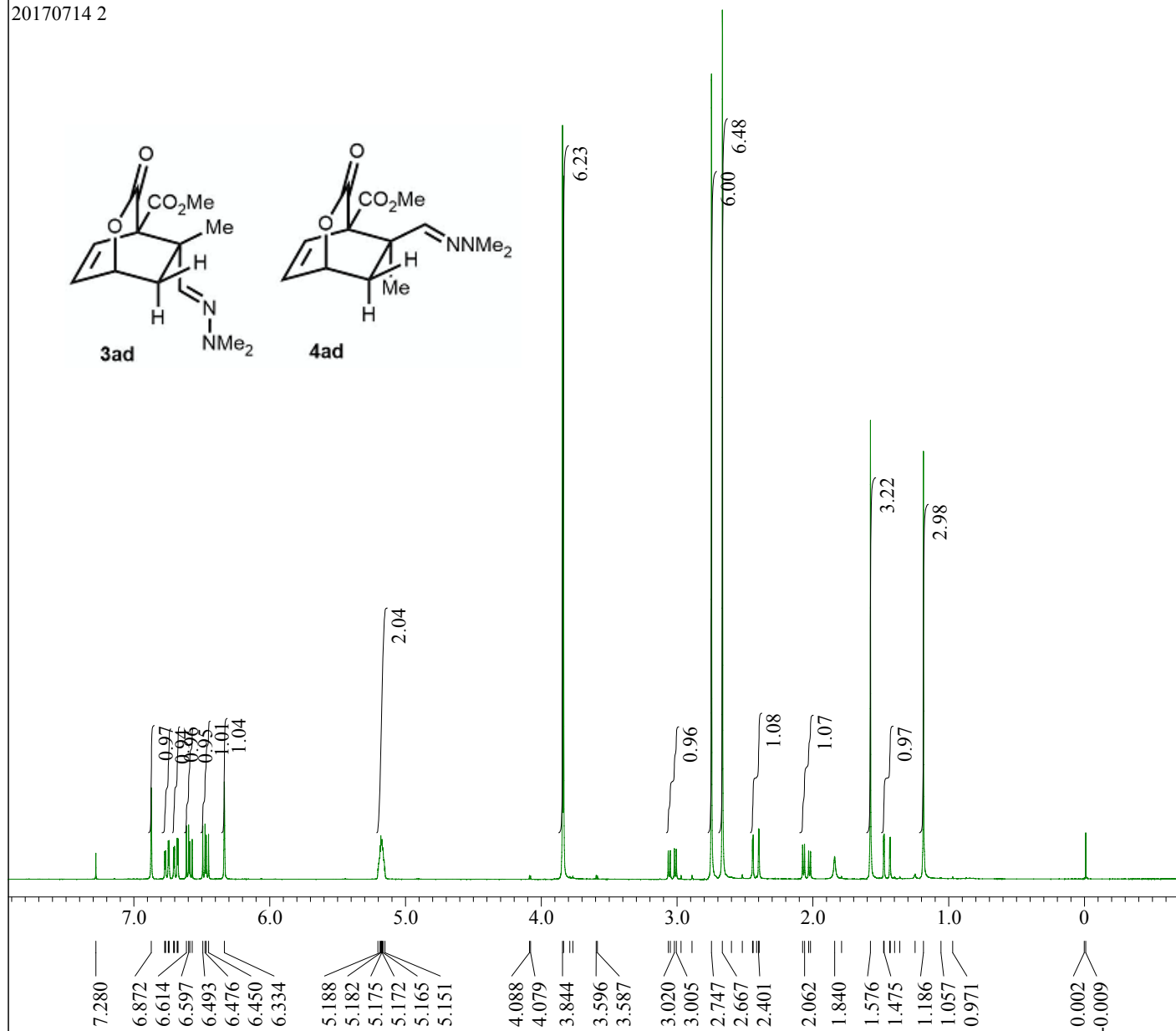
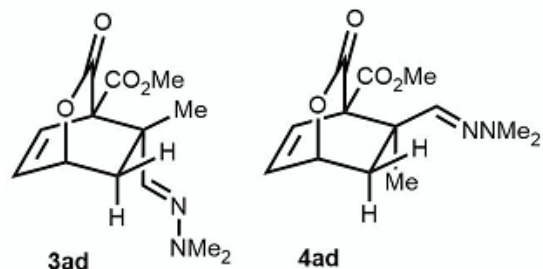


abe_no3_p45_32_44

A.carbon CDCl₃ D:\ rfia 22



20170714 2



---- PROCESSING PARAMETERS ----
 dc_balance(0, FALSE)
 secp(0.2, 0.0[s])
 trapezoid3(0[%], 80[%], 100[%])
 zerofill(1)
 fft(1, TRUE, TRUE)
 ppm
 machinephase
 phase(-25, -140, 50[%])

Filename = Y-Abe_900 2-1.jdf
 Author = Administrator
 Experiment = zg30
 Sample_Id = 20170714 2
 Solvent = CHLOROFORM-D
 Creation_Time = 19-JUL-2017 18:06:35
 Revision_Time = 19-JUL-2017 18:10:14
 Current_Time = 7-SEP-2017 14:24:59

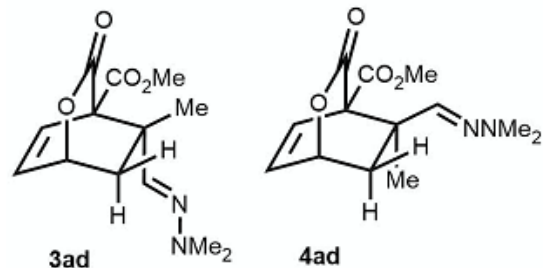
Comment = 20170714 2
 Data_Format = 1D COMPLEX
 Dim_Size = 32768
 Dim_Title = 1H
 Dim_Units = [ppm]
 Dimensions = X
 Spectrometer = BRUKER_DMX_NMR

X_Freq = 300.13185343[MHz]
 X_Offset = 1.85342561[kHz]
 X_Sweep = 6.18811881[kHz]

Temp_Get = 295.56[K]
 X_Points = 32768
 X_Prescans = 2
 Filter_Factor = 3232
 Scans = 16

X : ppm : 1H

20170714 c



---- PROCESSING PARAMETERS ----

```
dc_balance( 0, FALSE )
sexp( 2.0, 0.0[s] )
trapezoid3( 0[%], 80[%], 100[%] )
zerofill( 1 )
fft( 1, TRUE, TRUE )
ppm
machinephase
```

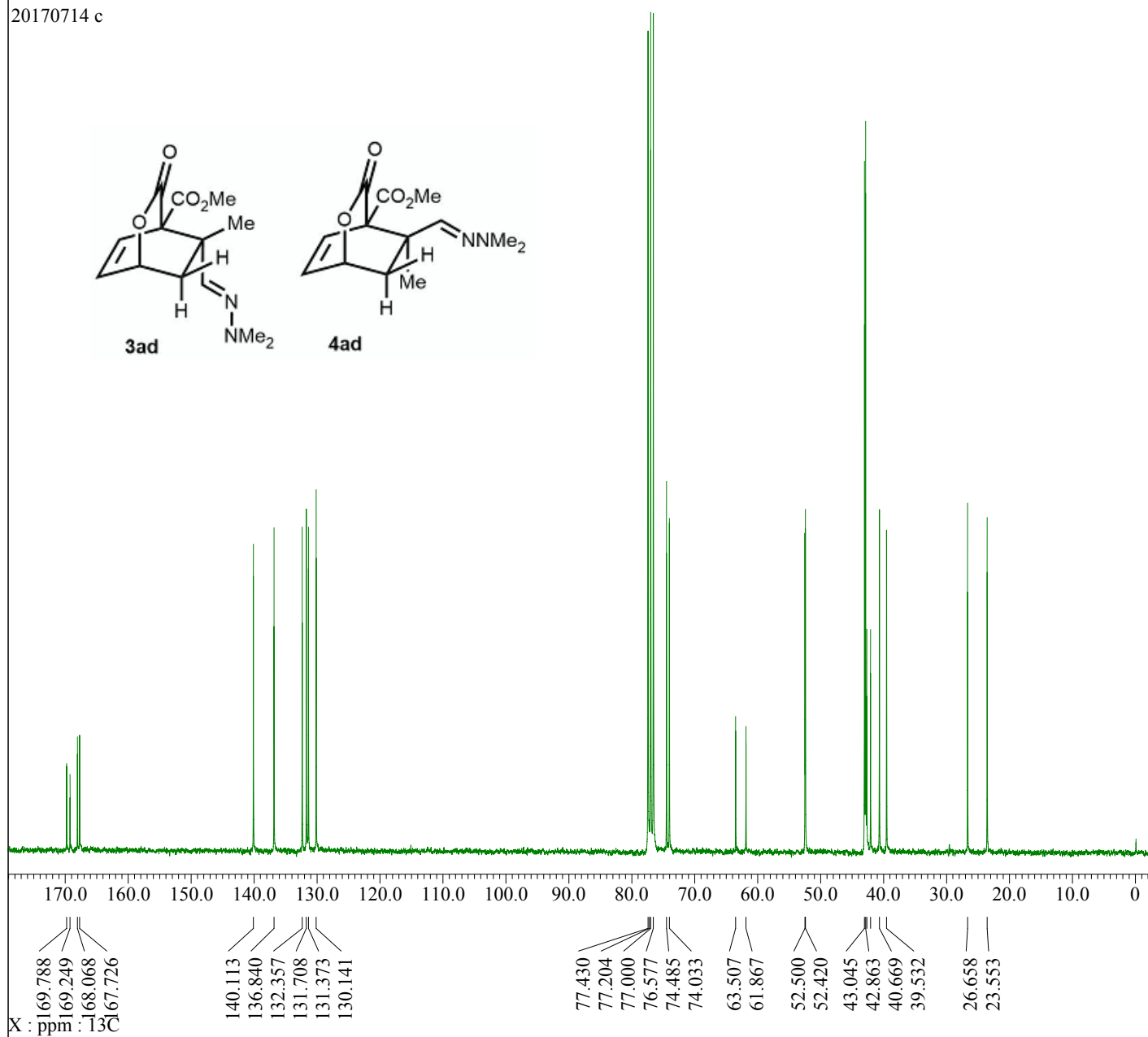
以下に由来: Y-Abe_882-1.jdf

```
Filename      = Y-Abe_882-5.jdf
Author       = Administrator
Experiment    = zgpg30
Sample_Id    = 20170714 c
Solvent      = CHLOROFORM-D
Creation_Time = 7-SEP-2017 14:26:36
Revision_Time = 7-SEP-2017 14:26:54
Current_Time  = 7-SEP-2017 14:29:06
```

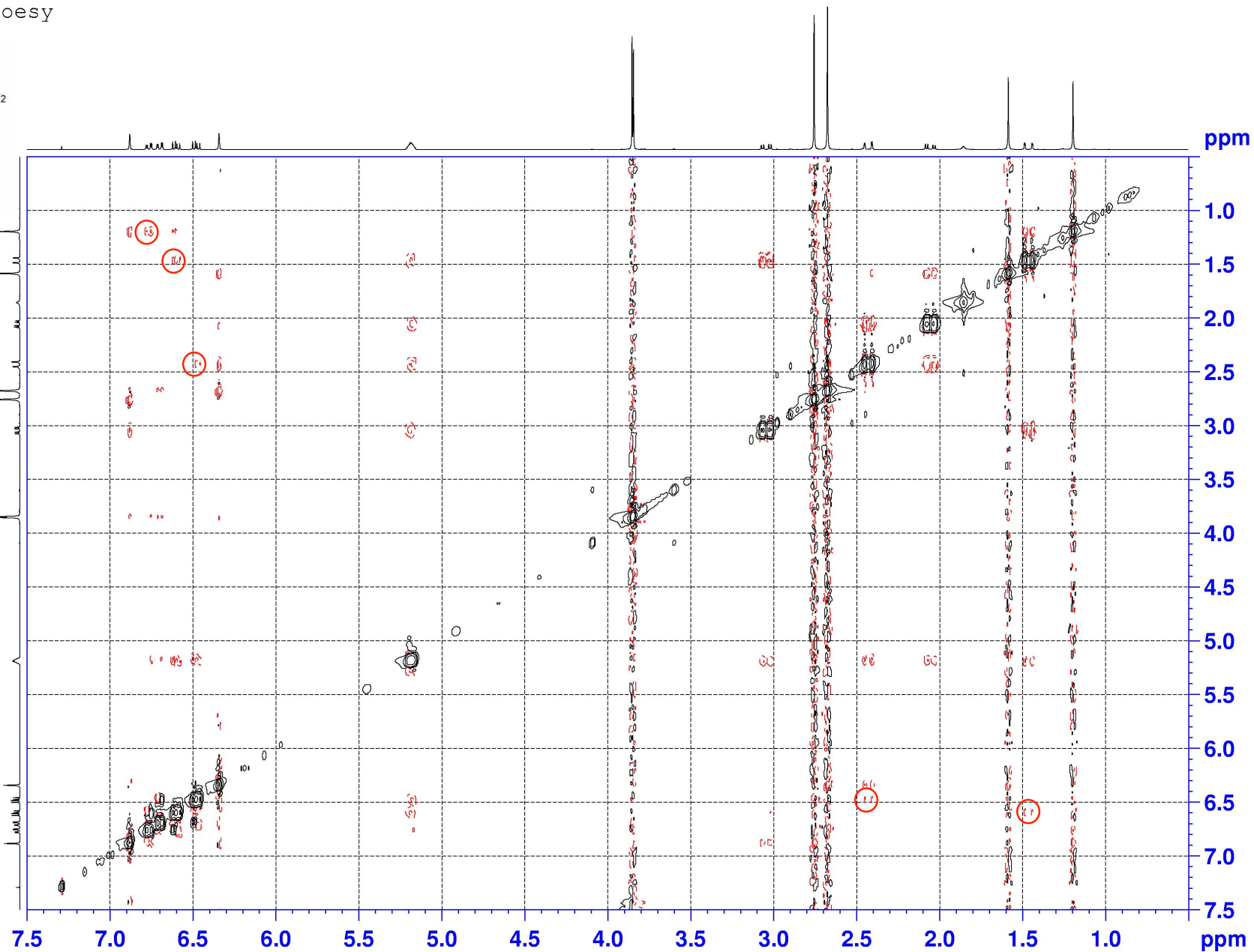
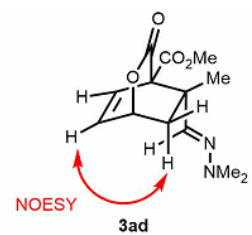
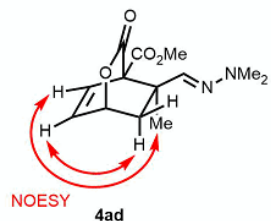
```
Comment      = 20170714 c
Data_Format   = 1D COMPLEX
Dim_Size      = 32768
Dim_Title     = 13C
Dim_Units     = [ppm]
Dimensions    = X
Spectrometer  = BRUKER_DMX_NMR
```

```
X_Freq       = 75.4752953[MHz]
X_Offset     = 7.54630085[kHz]
X_Sweep      = 18.02884615[kHz]
```

```
Temp_Get     = 297.26[K]
X_Points     = 32768
X_Prescans   = 4
Filter_Factor = 1109
Scans        = 4096
```

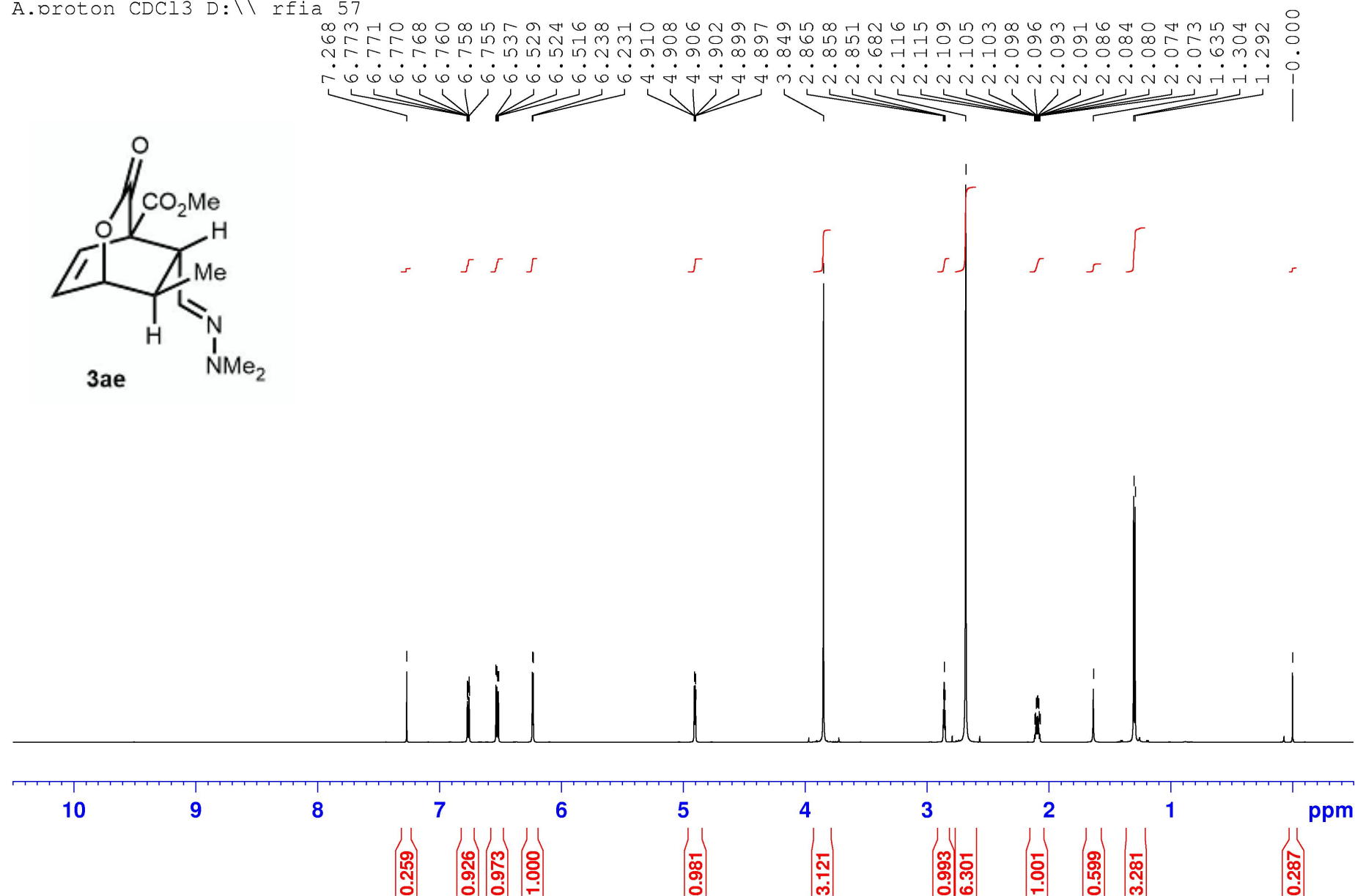
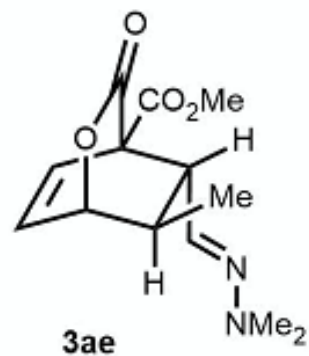


20170714 noesy



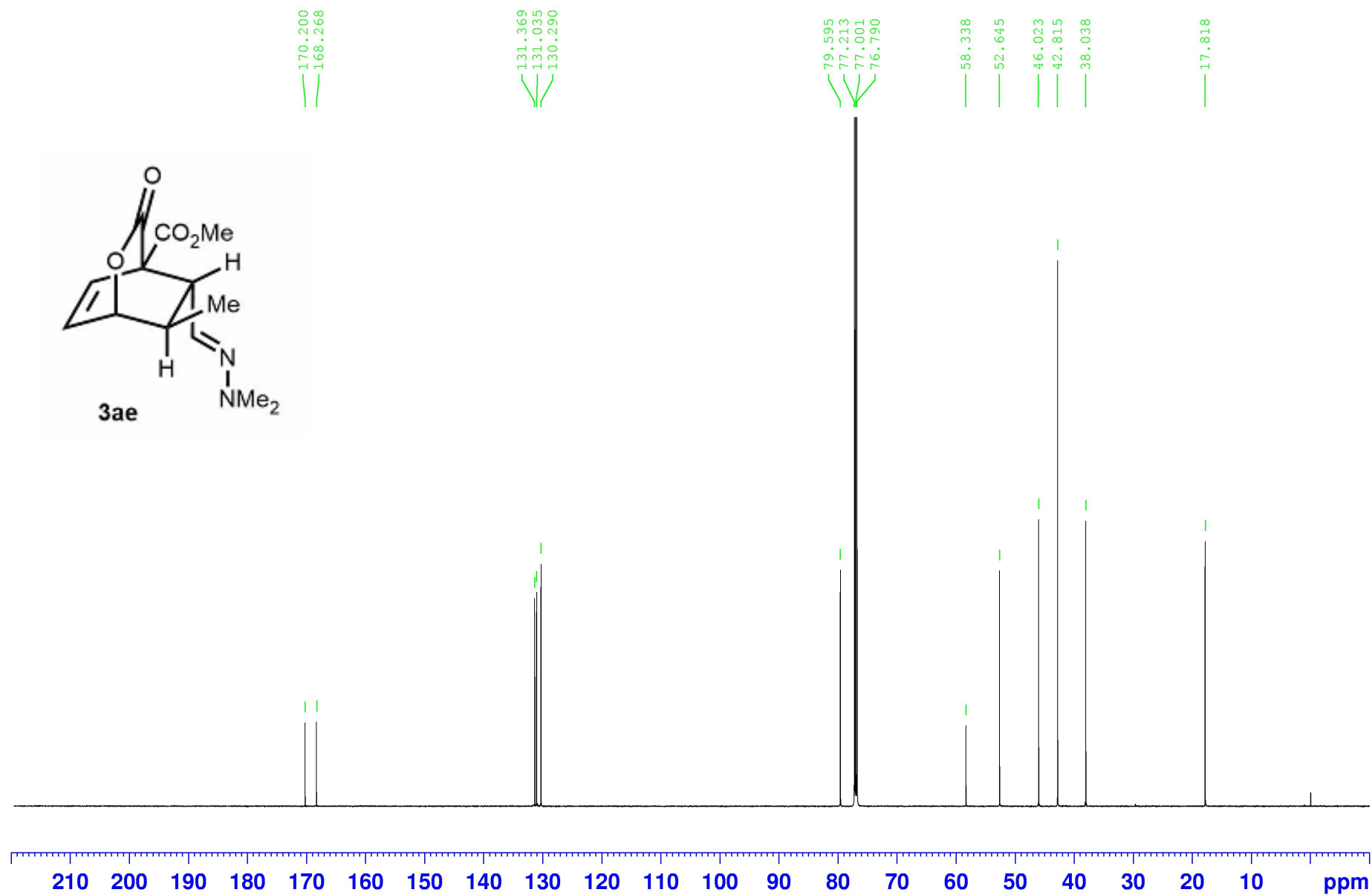
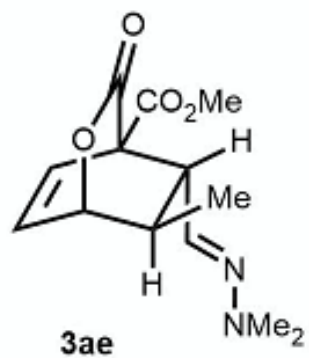
YH_0316_2

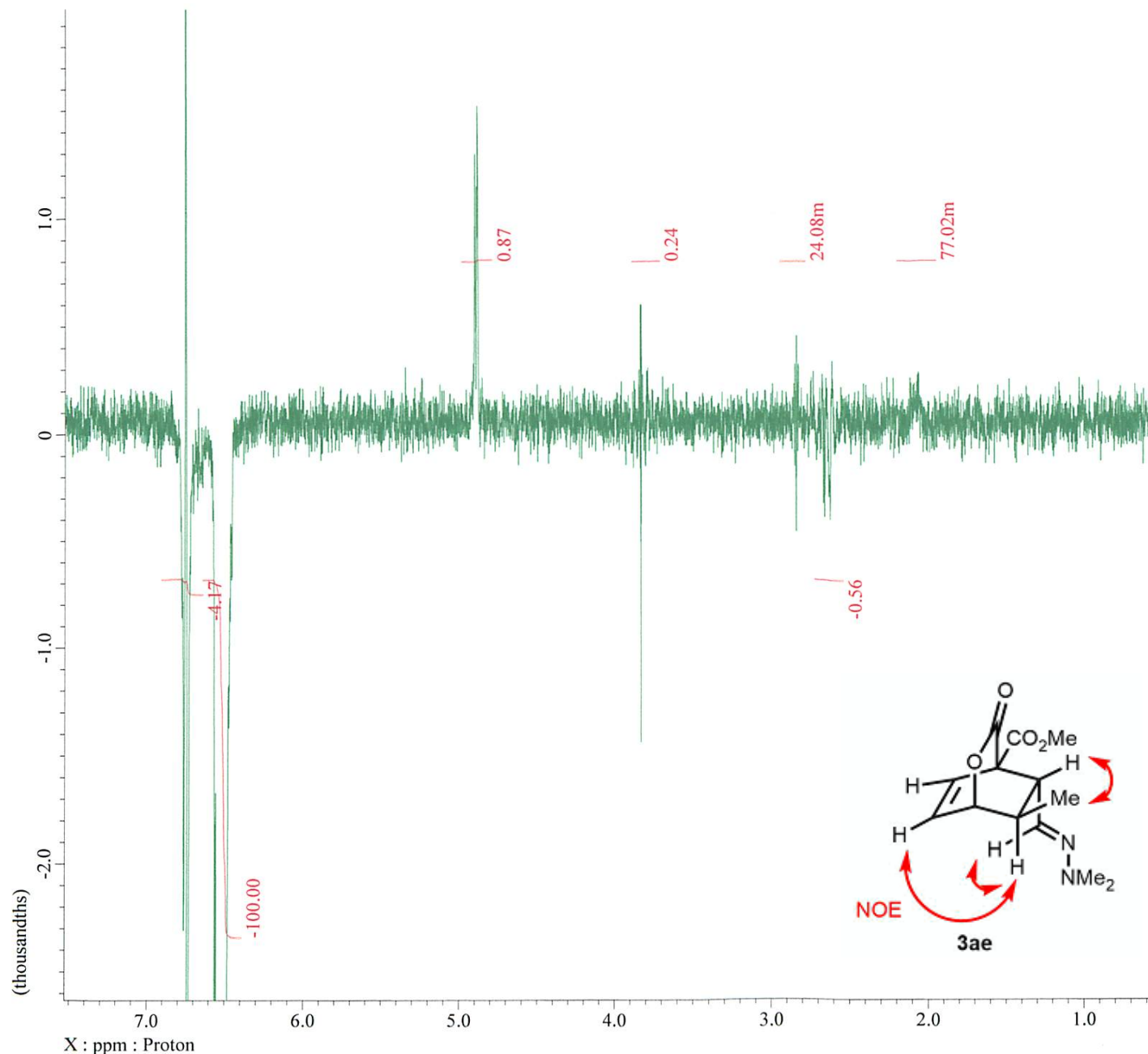
A.proton CDC13 D:\\ rfia 57



YH_0316_2

A.carbon CDC13 D:\\ rfia 57





---- PROCESSING PARAMETERS ----
sexf(0.2[Hz], 0.0[s])
trapezoid(0[%], 0[%], 80[%], 100[%])
zerofill(1)
fft(1, TRUE, TRUE)
ppm

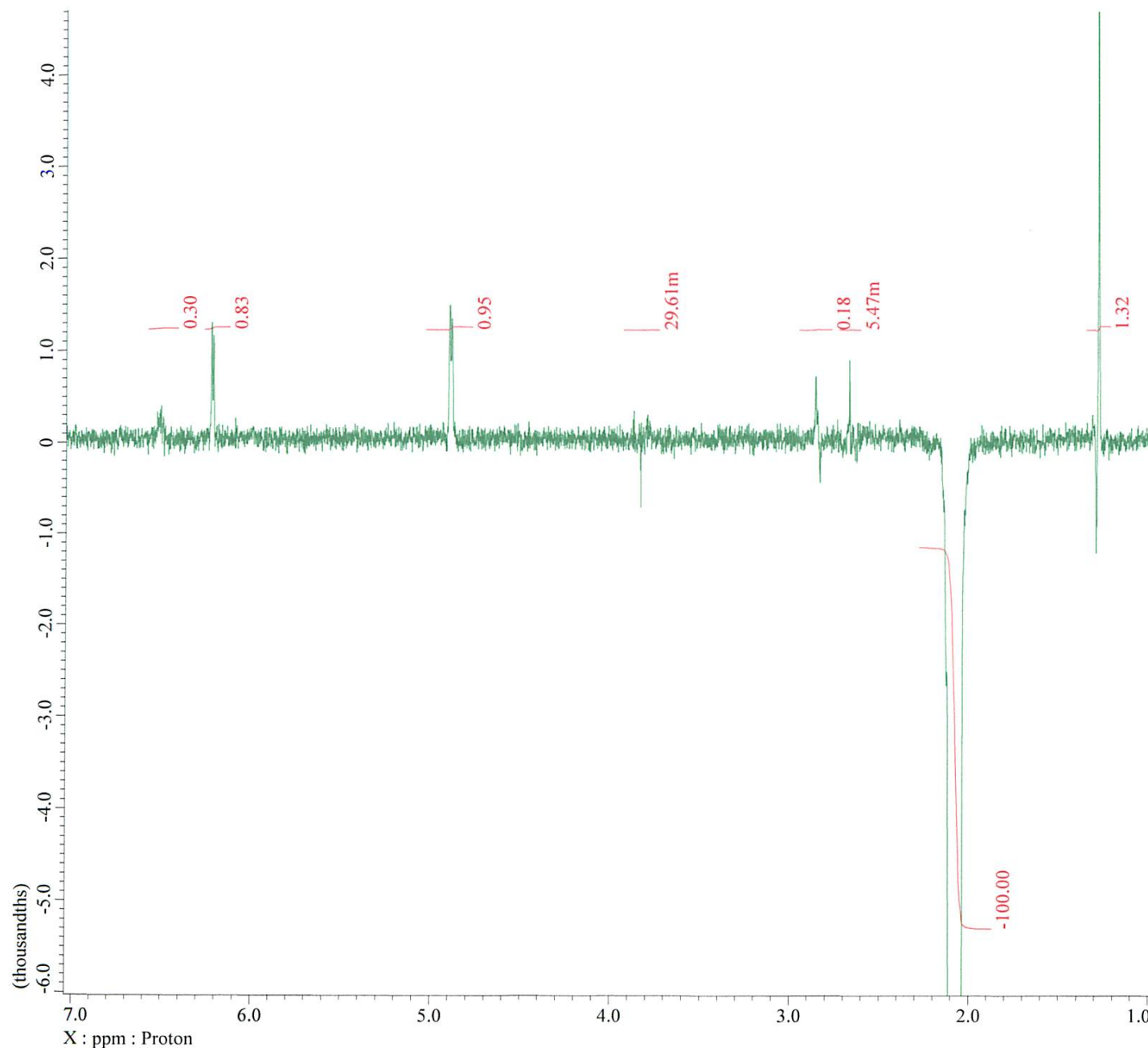
以下に由来: YH_0319_1_noe_1d-2-1.jdf

Filename = YH_0319_1_noe_1d-2-7.
Author = delta
Experiment = noe_1d.jxp
Sample_Id = YH_0319_1
Solvent = CHLOROFORM-D
Actual_Start_Time = 20-OCT-2017 11:31:23
Revision_Time = 20-OCT-2017 12:53:10

Comment = DPGSE NOE 1D
Data_Format = 1D_COMPLEX
Dim_Size = 13107
Dim_Title = Proton
Dim_Units = [ppm]
Dimensions = X
Spectrometer = JNM-ECZ400S/L1

Field_Strength = 9.389766[T] (400[MHz])
X_Acq_Duration = 2.18628096[s]
X_Domain = 1H
X_Freq = 399.78219838 [MHz]
X_Offset = 5[ppm]
X_Points = 16384
X_Prescans = 2
X_Resolution = 0.45739775 [Hz]
X_Sweep = 7.4940048 [kHz]
X_Sweep_Clipped = 5.99520384 [kHz]
Irr_Domain = Proton
Irr_Freq = 399.78219838 [MHz]
Irr_Offset = 5[ppm]
Tri_Domain = Proton
Tri_Freq = 399.78219838 [MHz]
Tri_Offset = 5[ppm]
Blanking = 2[us]
Clipped = FALSE
Scans = 16
Total_Scans = 16

Relaxation_Delay = 7[s]
Recvr_Gain = 50
Temp_Get = 21.8[dC]
Mix_Time = 0.5[s]
X_Acq_Time = 2.18628096[s]
X_Atn = 3[dB]
X_Pulse = 6.1[us]
Irr_Mode = Off
Obs_Sel_180 = 60[ms]
Obs_Sel_Atn = 69.125[dB]
Obs_Sel_Offset = 6.52819633[ppm]



---- PROCESSING PARAMETERS ----
sexp(0.2[Hz], 0.0[s])
trapezoid(0[%], 0[%], 80[%], 100[%])
zerofill(1)
fft(1, TRUE, TRUE)
ppm

以下に由来: YH_0319_1_noe_1d-6-1.jdf

Filename = YH_0319_1_noe_1d-6-6.
Author = delta
Experiment = noe_1d.jxp
Sample_Id = YH_0319_1
Solvent = CHLOROFORM-D
Actual_Start_Time = 20-OCT-2017 11:43:35
Revision_Time = 20-OCT-2017 13:20:15

Comment = DPGSE NOE 1D
Data Format = 1D COMPLEX
Dim Size = 13107
Dim Title = Proton
Dim Units = [ppm]
Dimensions = X
Spectrometer = JNM-ECZ400S/L1

Field Strength = 9.389766[T] (400[MHz])
X_Acq_Duration = 2.18628096[s]
X_Domain = 1H
X_Freq = 399.78219838[MHz]
X_Offset = 5[ppm]
X_Points = 16384
X_Prescans = 2
X_Resolution = 0.45739775[Hz]
X_Sweep = 7.4940048[kHz]
X_Sweep_Clipped = 5.99520384[kHz]
Irr_Domain = Proton
Irr_Freq = 399.78219838[MHz]
Irr_Offset = 5[ppm]
Tri_Domain = Proton
Tri_Freq = 399.78219838[MHz]
Tri_Offset = 5[ppm]
Blanking = 2[us]
Clipped = FALSE
Scans = 16
Total_Scans = 16

Relaxation_Delay = 7[s]
Recvr_Gain = 50
Temp_Get = 21.8[dC]
Mix_Time = 0.5[s]
X_Acq_Time = 2.18628096[s]
X_Atn = 3[dB]
X_Pulse = 6.1[us]
Irr_Mode = Off
Obs_Sel_180 = 60[ms]
Obs_Sel_Atn = 69.125[dB]
Obs_Sel_Offset = 2.0926187[ppm]



---- PROCESSING PARAMETERS ----
sexp(0.2[Hz], 0.0[s])
trapezoid(0[%], 0[%], 80[%], 100[%])
zerofill(1)
fft(1, TRUE, TRUE)
ppm

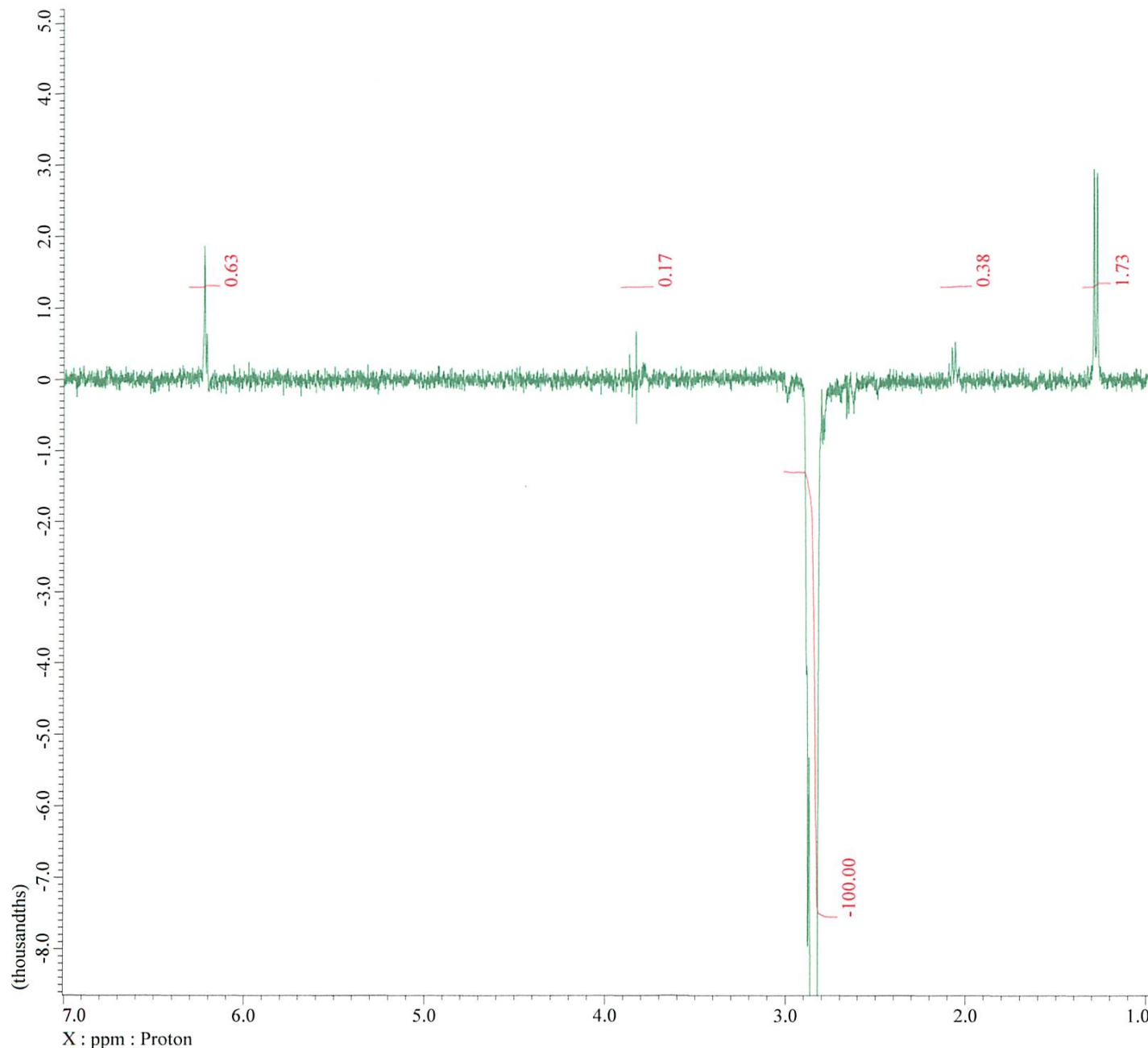
以下に由来: YH_0319_1_noe_1d-5-1.jdf

Filename = YH_0319_1_noe_1d-5-6.
Author = delta
Experiment = noe_1d.jxp
Sample_Id = YH_0319_1
Solvent = CHLOROFORM-D
Actual_Start_Time = 20-OCT-2017 11:40:32
Revision_Time = 20-OCT-2017 13:17:49

Comment = DPGSE NOE 1D
Data_Format = 1D COMPLEX
Dim_Size = 13107
Dim_Title = Proton
Dim_Units = [ppm]
Dimensions = X
Spectrometer = JNM-ECZ400S/L1

Field Strength = 9.389766[T] (400[MHz])
X_Acq_Duration = 2.18628096[s]
X_Domain = 1H
X_Freq = 399.78219838[MHz]
X_Offset = 5[ppm]
X_Points = 16384
X_Prescans = 2
X_Resolution = 0.45739775[Hz]
X_Sweep = 7.4940048[kHz]
X_Sweep_Clipped = 5.99520384[kHz]
Irr_Domain = Proton
Irr_Freq = 399.78219838[MHz]
Irr_Offset = 5[ppm]
Tri_Domain = Proton
Tri_Freq = 399.78219838[MHz]
Tri_Offset = 5[ppm]
Blanking = 2[us]
Clipped = FALSE
Scans = 16
Total_Scans = 16

Relaxation_Delay = 7[s]
Recvr_Gain = 50
Temp_Get = 21.8[dC]
Mix_Time = 0.5[s]
X_Acq_Time = 2.18628096[s]
X_Atn = 3[dB]
X_Pulse = 6.1[us]
Irr_Mode = Off
Obs_Sel_180 = 60[ms]
Obs_Sel_Atn = 69.125[dB]
Obs_Sel_Offset = 2.86090207[ppm]





----- PROCESSING PARAMETERS -----
sexp(0.2[Hz], 0.0[s])
trapezoid(0[%], 0[%], 80[%], 100[%])
zerofill(1)
fft(1, TRUE, TRUE)
ppm

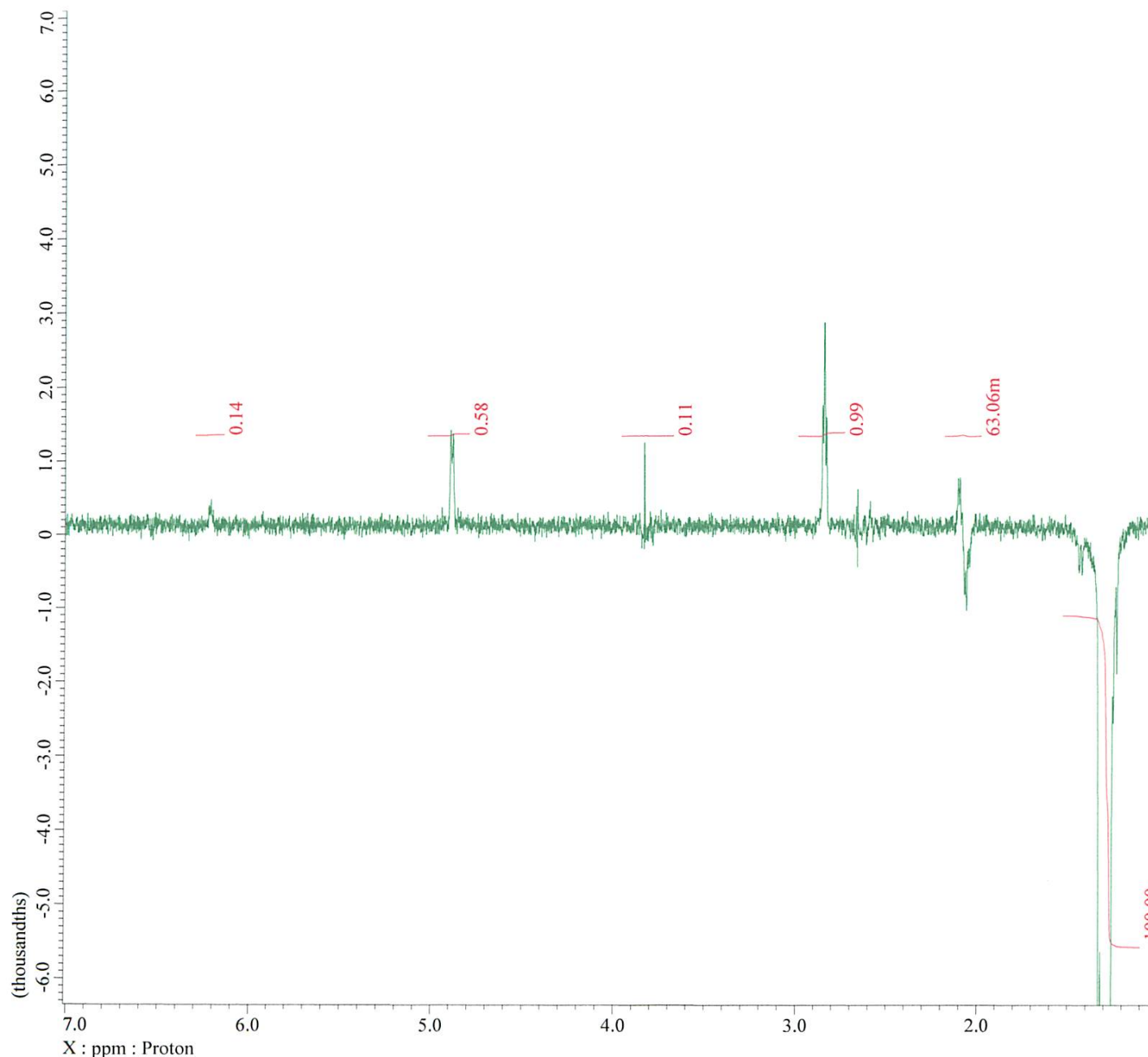
以下に由来: YH_0319_1_noe_1d-7-1.jdf

Filename = YH_0319_1_noe_1d-7-7.
Author = delta
Experiment = noe_1d.jxp
Sample_Id = YH_0319_1
Solvent = CHLOROFORM-D
Actual_Start_Time = 20-OCT-2017 11:46:38
Revision_Time = 20-OCT-2017 13:23:20

Comment = DPGSE NOE 1D
Data_Format = 1D COMPLEX
Dim_Size = 13107
Dim_Title = Proton
Dim_Units = [ppm]
Dimensions = X
Spectrometer = JNM-ECZ400S/L1

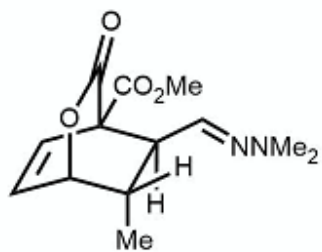
Field_Strength = 9.389766[T] (400[MHz])
X_Acq_Duration = 2.18628096[s]
X_Domain = 1H
X_Freq = 399.78219838[MHz]
X_Offset = 5[ppm]
X_Points = 16384
X_Prescans = 2
X_Resolution = 0.45739775[Hz]
X_Sweep = 7.4940048[kHz]
X_Sweep_Clipped = 5.99520384[kHz]
Irr_Domain = Proton
Irr_Freq = 399.78219838[MHz]
Irr_Offset = 5[ppm]
Tri_Domain = Proton
Tri_Freq = 399.78219838[MHz]
Tri_Offset = 5[ppm]
Blanking = 2[us]
Clipped = FALSE
Scans = 16
Total_Scans = 16

Relaxation_Delay = 7[s]
Recvr_Gain = 50
Temp_Get = 21.8[dC]
Mix_Time = 0.5[s]
X_Acq_Time = 2.18628096[s]
X_Atn = 3[dB]
X_Pulse = 6.1[us]
Irr_Mode = Off
Obs_Sel_180 = 60[ms]
Obs_Sel_Atn = 69.125[dB]
Obs_Sel_Offset = 1.29994524[ppm]

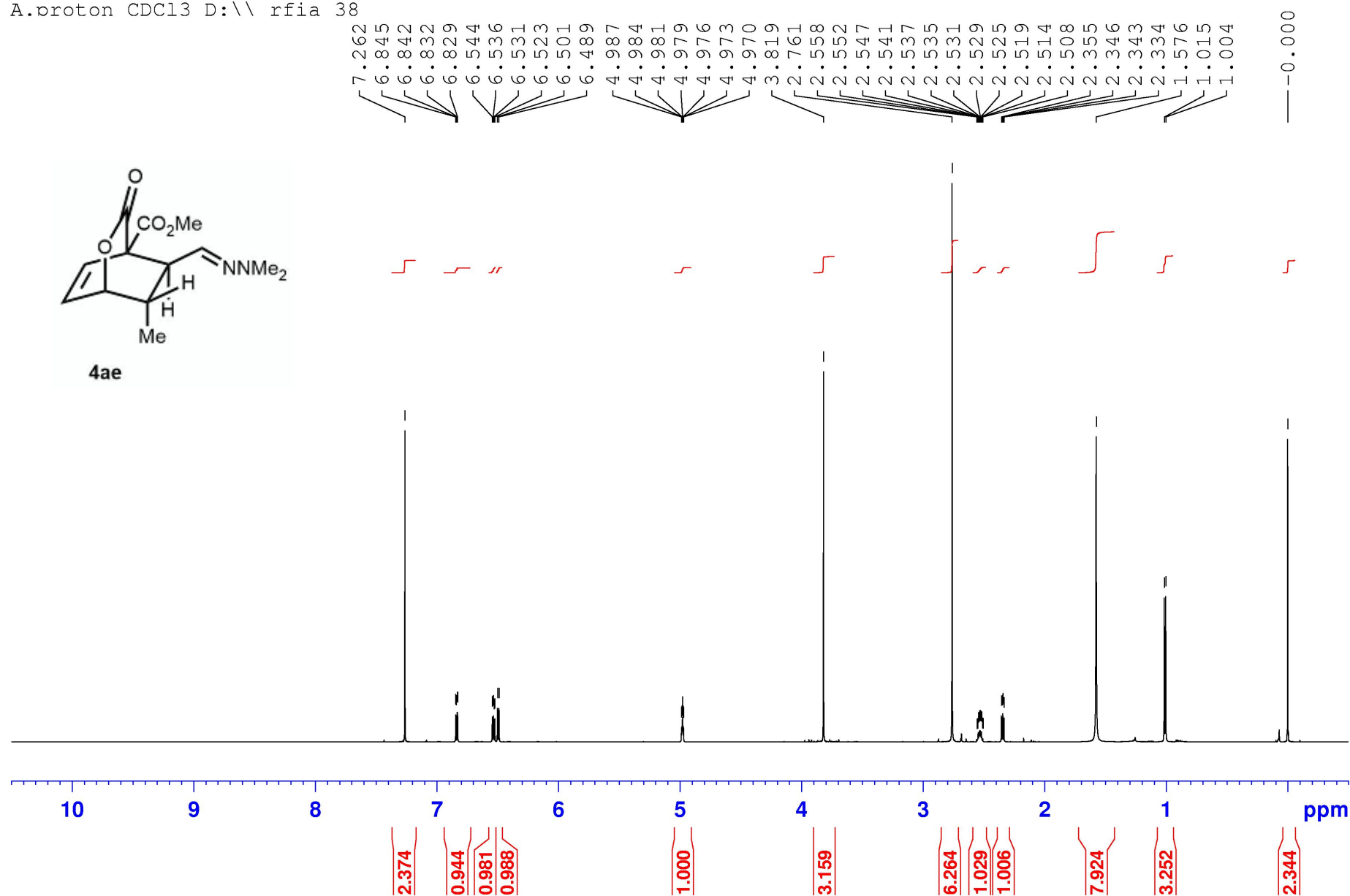


YH_0319_8

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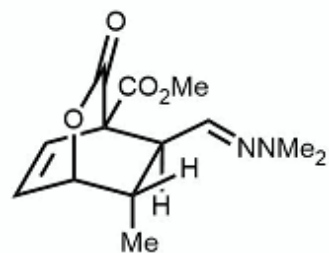


4ae

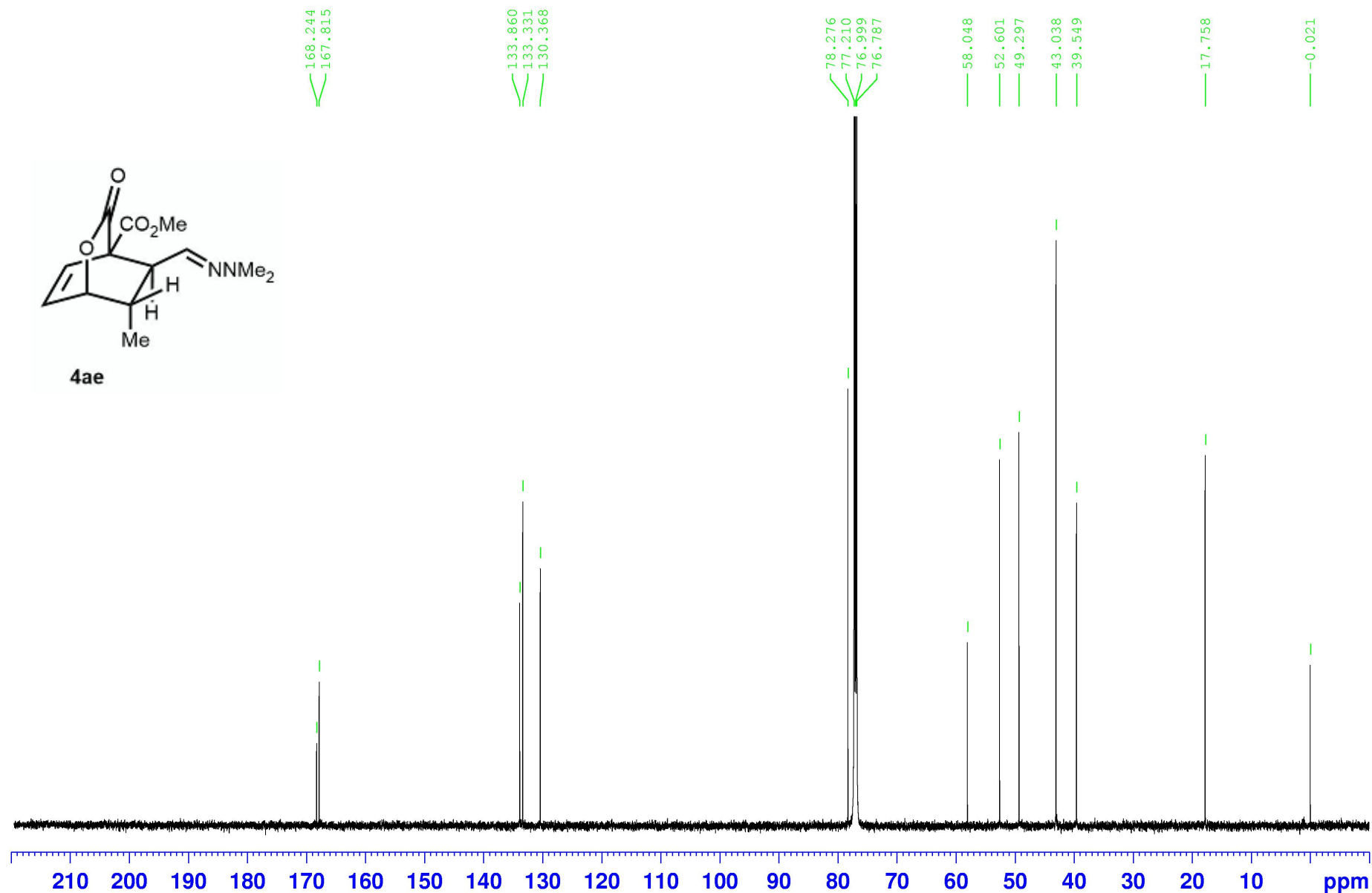


YH_0319_8

A.carbon CDCl₃ D:\\ rfia 38



4ae





----- PROCESSING PARAMETERS -----
 sexp(0.2[Hz], 0.0[s])
 trapezoid(0[%], 0[%], 80[%], 100[%])
 zerofill(1)
 fft(1, TRUE, TRUE)
 ppm

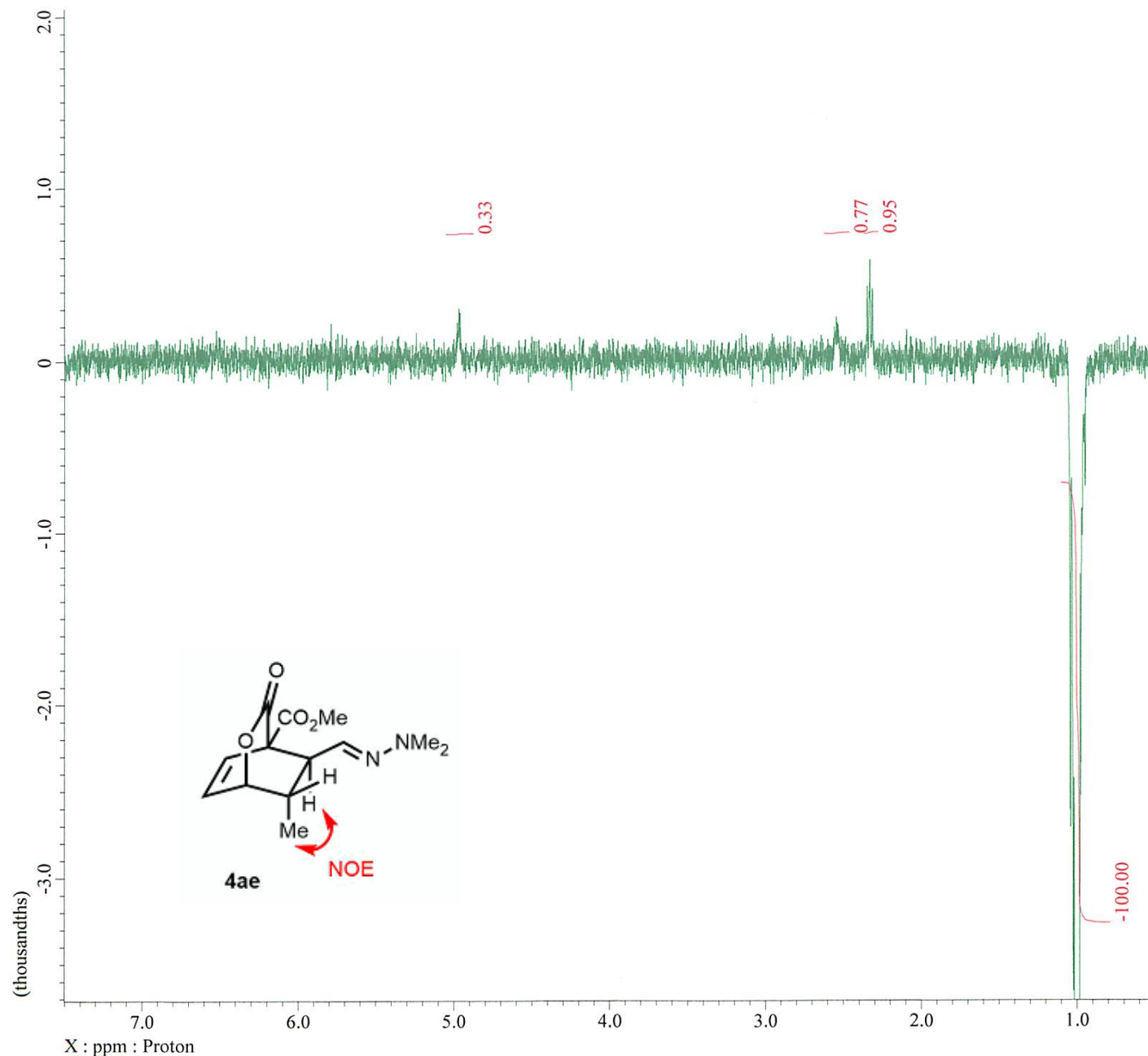
以下に由来: YH_0319_6_8_noe_1d-8-1.jdf

Filename = YH_0319_6_8_noe_1d-8-
 Author = delta
 Experiment = noe_1d.jxp
 Sample_Id = YH_0319_6_8
 Solvent = CHLOROFORM-D
 Actual_Start_Time = 20-OCT-2017 13:44:54
 Revision_Time = 20-OCT-2017 13:54:20

Comment = DPGSE NOE 1D
 Data_Format = 1D COMPLEX
 Dim_Size = 13107
 Dim_Title = Proton
 Dim_Units = [ppm]
 Dimensions = X
 Spectrometer = JNM-ECZ400S/L1

Field_Strength = 9.389766[T] (400[MHz])
 X_Acq_Duration = 2.18628096[s]
 X_Domain = 1H
 X_Freq = 399.78219838[MHz]
 X_Offset = 5[ppm]
 X_Points = 16384
 X_Prescans = 2
 X_Resolution = 0.45739775[Hz]
 X_Sweep = 7.4940048[kHz]
 X_Sweep_Clipped = 5.99520384[kHz]
 Irr_Domain = Proton
 Irr_Freq = 399.78219838[MHz]
 Irr_Offset = 5[ppm]
 Tri_Domain = Proton
 Tri_Freq = 399.78219838[MHz]
 Tri_Offset = 5[ppm]
 Blanking = 2[us]
 Clipped = FALSE
 Scans = 32
 Total_Scans = 32

Relaxation_Delay = 7[s]
 Recvr_Gain = 50
 Temp_Get = 21.9[dC]
 Mix_Time = 0.5[s]
 X_Acq_Time = 2.18628096[s]
 X_Atn = 3[dB]
 X_Pulse = 6.1[us]
 Irr_Mode = Off
 Obs_Sel_180 = 60[ms]
 Obs_Sel_Atn = 69.125[dB]
 Obs_Sel_Offset = 1.01213503[ppm]





---- PROCESSING PARAMETERS ----
sexp(0.2[Hz], 0.0[s])
trapezoid(0[%], 0[%], 80[%], 100[%])
zerofill(1)
fft(1, TRUE, TRUE)
ppm

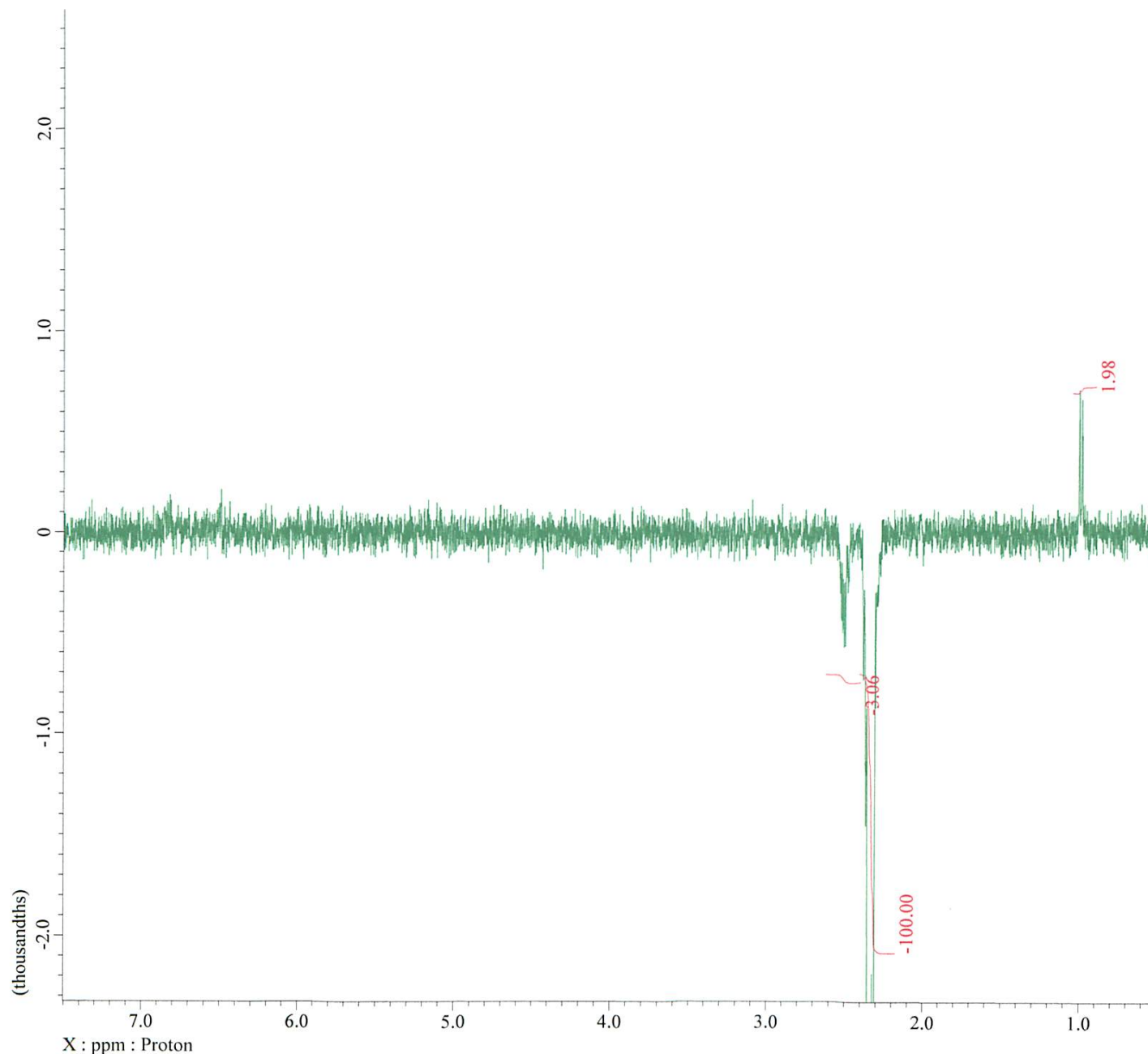
以下に由来: YH_0319_6_8_noe_1d-7-1.jdf

Filename = YH_0319_6_8_noe_1d-7-
Author = delta
Experiment = noe_1d.jxp
Sample Id = YH_0319_6_8
Solvent = CHLOROFORM-D
Actual_Start_Time = 20-OCT-2017 13:39:12
Revision_Time = 20-OCT-2017 13:52:00

Comment = DPGSE NOE 1D
Data Format = 1D COMPLEX
Dim_Size = 13107
Dim_Title = Proton
Dim_Units = [ppm]
Dimensions = X
Spectrometer = JNM-ECZ400S/L1

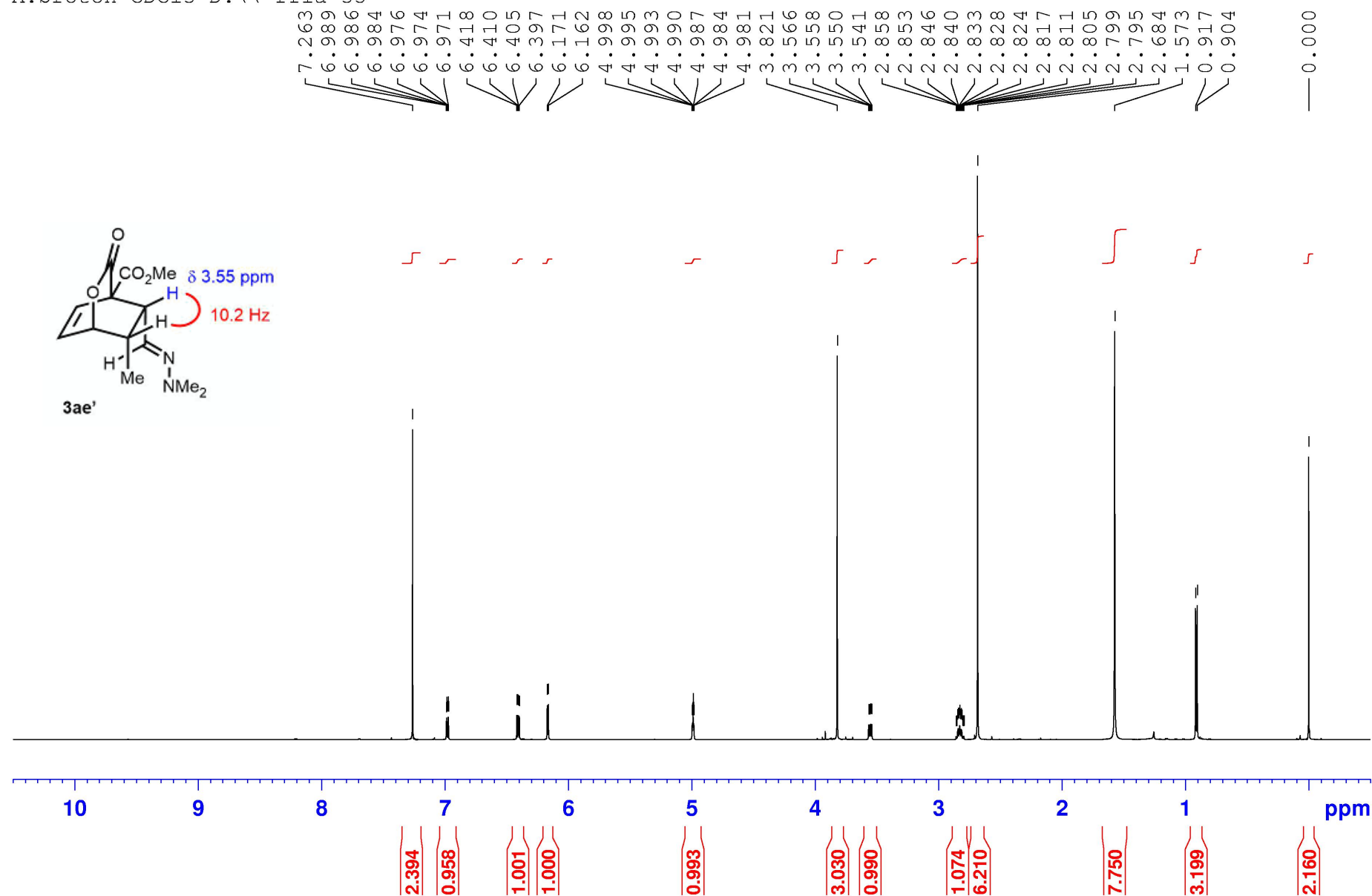
Field_Strength = 9.389766[T] (400[MHz])
X_Acq_Duration = 2.18628096[s]
X_Domain = 1H
X_Freq = 399.78219838[MHz]
X_Offset = 5[ppm]
X_Points = 16384
X_Prescans = 2
X_Resolution = 0.45739775[Hz]
X_Sweep = 7.4940048[kHz]
X_Sweep_Clipped = 5.99520384[kHz]
Irr_Domain = Proton
Irr_Freq = 399.78219838[MHz]
Irr_Offset = 5[ppm]
Tri_Domain = Proton
Tri_Freq = 399.78219838[MHz]
Tri_Offset = 5[ppm]
Blanking = 2[us]
Clipped = FALSE
Scans = 32
Total_Scans = 32

Relaxation_Delay = 7[s]
Recvr_Gain = 50
Temp_Get = 21.9[$^{\circ}$ C]
Mix_Time = 0.5[s]
X_Acq_Time = 2.18628096[s]
X_Atn = 3[dB]
X_Pulse = 6.1[us]
Irr_Mode = Off
Obs_Sel_180 = 60[ms]
Obs_Sel_Atn = 69.125[dB]
Obs_Sel_Offset = 2.34604478[ppm]



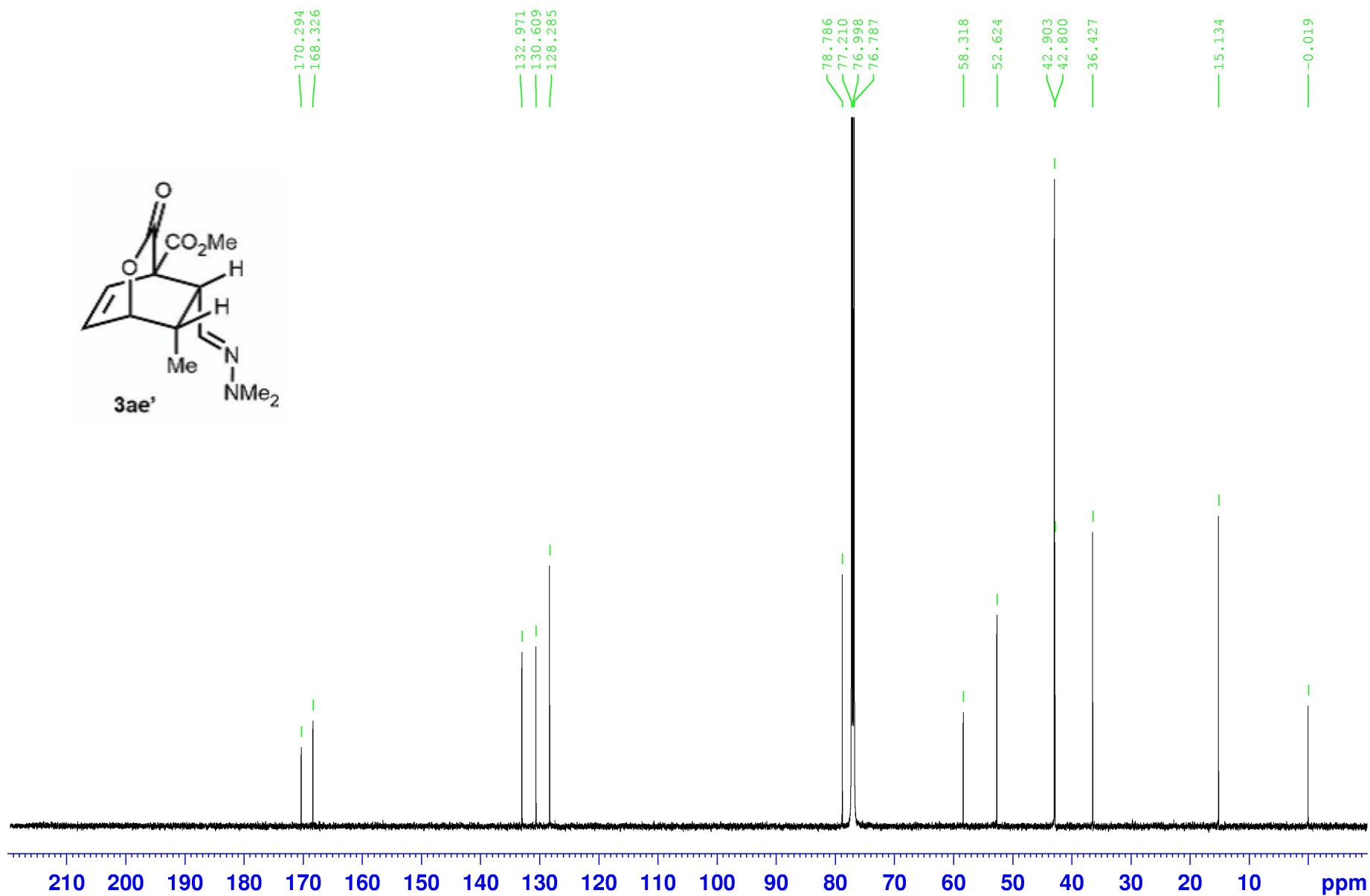
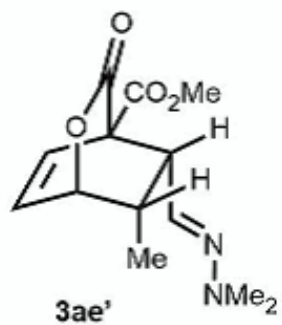
YH_0319_5

A.proton CDC13 D:\\ rfia 35



YH_0319_5

A.carbon CDC13 D:\\ rfia 35



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---- PROCESSING PARAMETERS ----

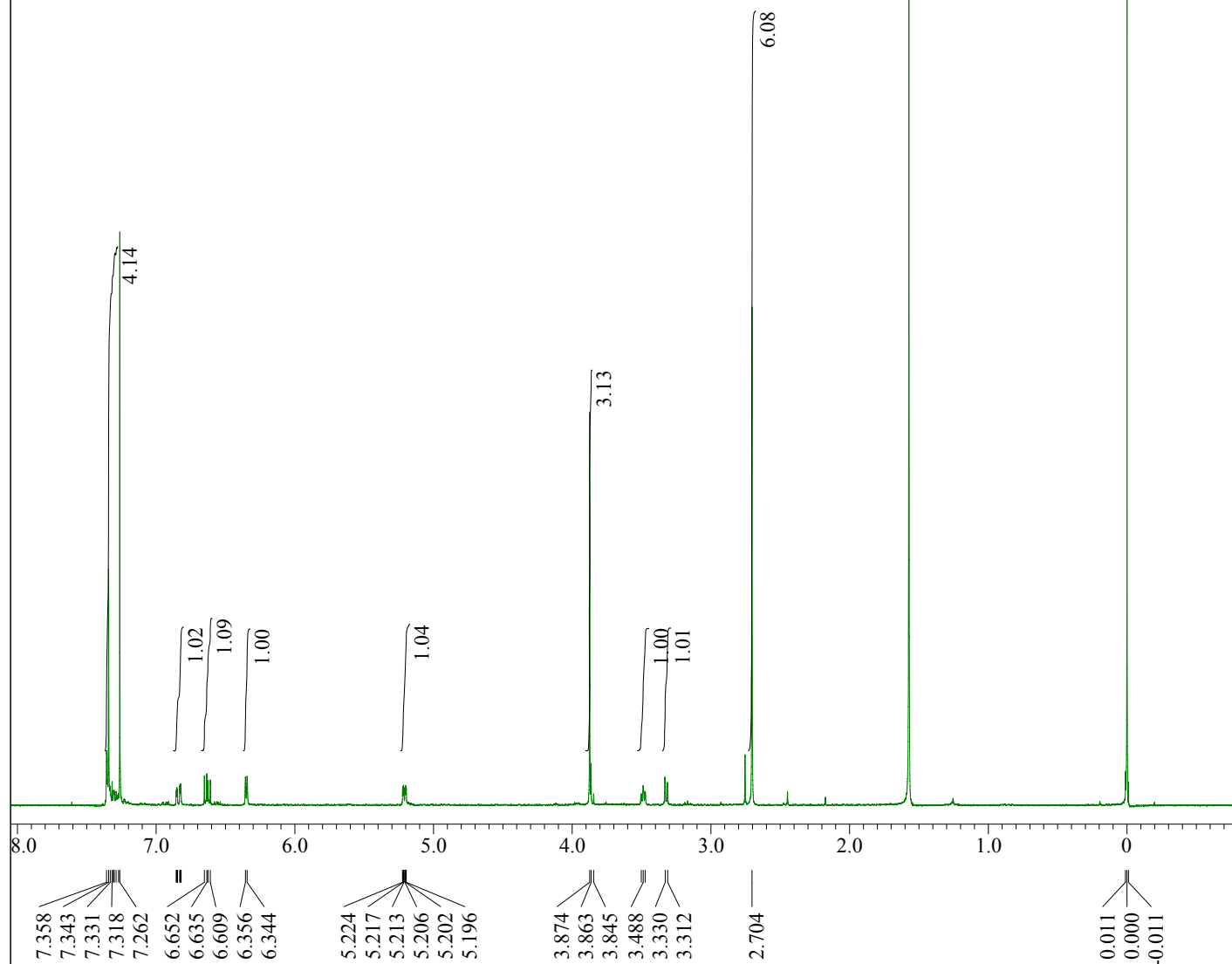
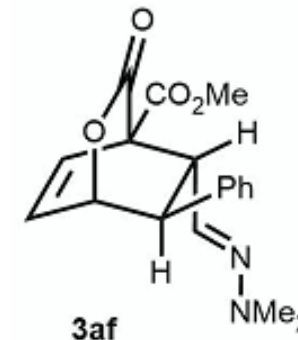
```
dc_balance( 0, FALSE )
sexp( 0.2, 0.0[s] )
trapezoid3( 0[%], 80[%], 100[%] )
zerofill( 1 )
fft( 1, TRUE, TRUE )
ppm
machinephase
```

Filename = Y-Abe_990-2.jdf
Author = Administrator
Experiment = zg30
Sample_Id = 20170726 3
Solvent = CHLOROFORM-D
Creation_Time = 26-JUL-2017 18:52:52
Revision_Time = 7-SEP-2017 14:33:32
Current_Time = 7-SEP-2017 14:33:48

Comment = 20170726 3
Data_Format = 1D COMPLEX
Dim_Size = 32768
Dim_Title = 1H
Dim_Units = [ppm]
Dimensions = X
Spectrometer = BRUKER_DMX_NMR

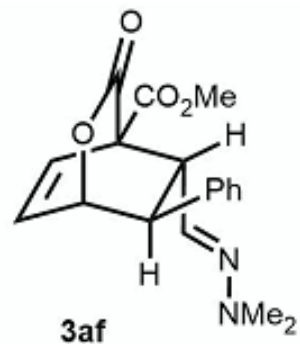
X_Freq = 300.13185343[MHz]
X_Offset = 1.85342561[kHz]
X_Sweep = 6.18811881[kHz]

Temp_Get = 295.46[K]
X_Points = 32768
X_Prescans = 2
Filter_Factor = 3232
Scans = 16



X : ppm : 1H

20170801 c



170.34
168.06

140.12
130.97
130.68
130.54
128.88
127.91
127.47

79.04
77.42
77.00
76.58

59.03
52.73
48.14
45.51
42.83

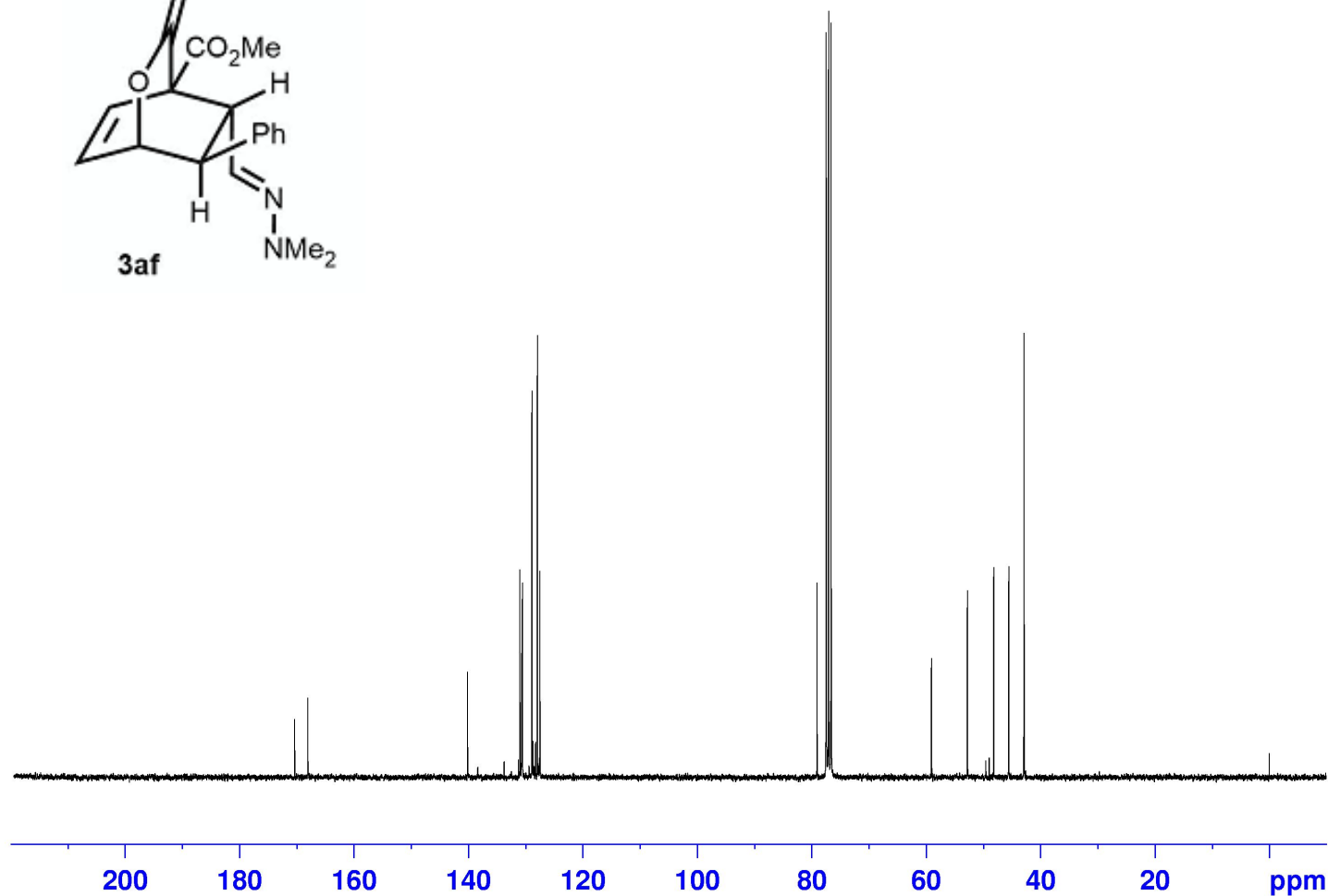
Current Data Parameters
NAME Y-Abe
EXPNO 1042
PROCNO 1

F2 - Acquisition Parameters
Date_ 20170802
Time 5.40
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDC13
NS 4096
DS 4
SWH 18028.846 Hz
FIDRES 0.275098 Hz
AQ 1.8175317 sec
RG 2050
DW 27.733 usec
DE 6.50 usec
TE 297.4 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

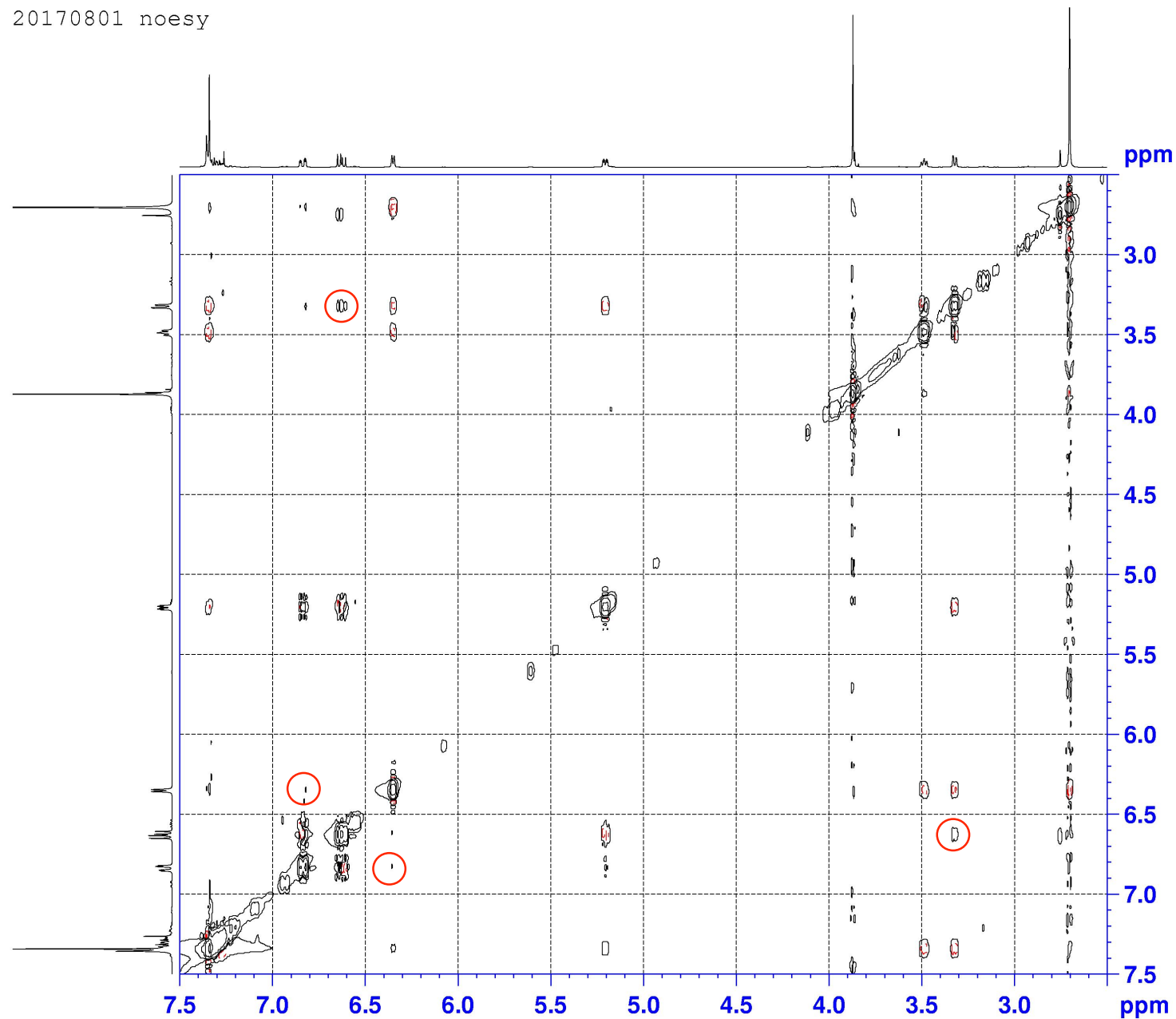
===== CHANNEL f1 =====
SFO1 75.4752953 MHz
NUC1 13C
P1 10.00 usec
PLW1 34.00000000 W

===== CHANNEL f2 =====
SFO2 300.1312005 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 80.00 usec
PLW2 6.19999981 W
PLW12 0.21797000 W
PLW13 0.13950001 W

F2 - Processing parameters
SI 32768
SF 75.4677509 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



20170801 noesy

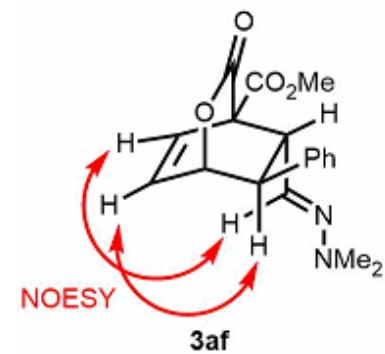


Current Data Parameters
 NAME Y-Abe
 EXPNO 1041
 PROCNO 1

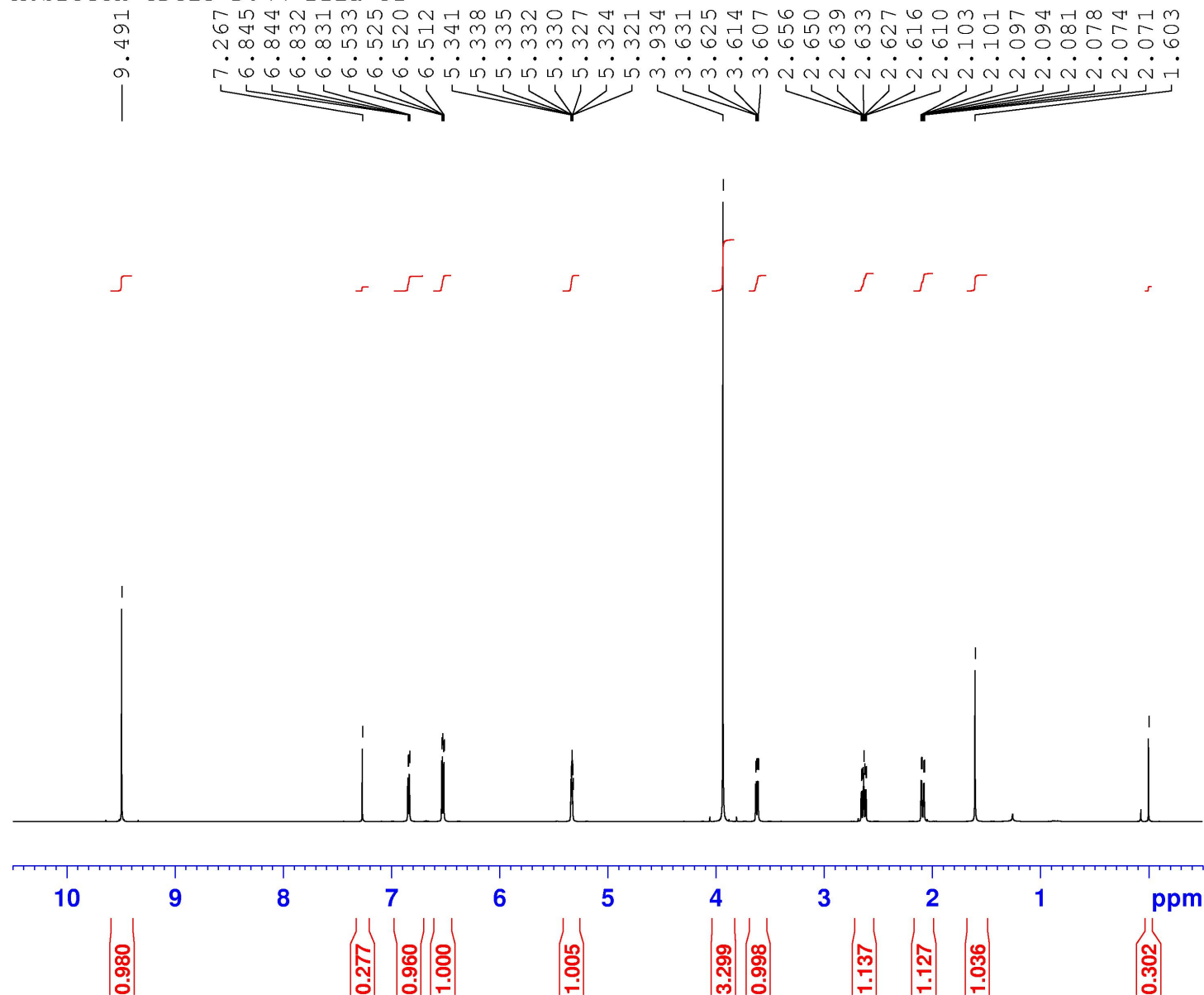
F2 - Acquisition Parameters
 Date_ 20170801
 Time 21.14
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG noesyph
 TD 2048
 SOLVENT CDC13
 NS 16
 DS 4
 SWH 2415.459 Hz
 FIDRES 1.179423 Hz
 AQ 0.4239360 sec
 RG 32
 DW 207.000 usec
 DE 6.50 usec
 TE 295.7 K
 D0 0.00018822 sec
 D1 1.91029704 sec
 D8 0.89999998 sec
 IN0 0.00041400 sec

===== CHANNEL f1 =====
 SFO1 300.1313051 MHz
 NUC1 1H
 P1 14.75 usec
 PLW1 6.19999981 W

F1 - Acquisition parameters
 TD 256
 SFO1 300.1313 MHz
 FIDRES 9.435387 Hz
 SW 8.048 ppm
 FnMODE States-TPPI



YH_0348_1_2nd
A.proton CDC13 D:\ rfia 51

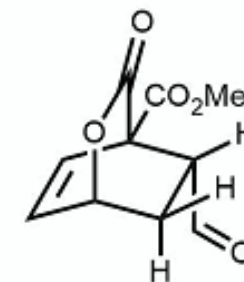


Current Data Parameters
NAME Hashimoto
EXPNO 1630
PROCNO 1

F2 - Acquisition Parameters
Date_ 20180801
Time 15.21
INSTRUM spect
PROBHD 5 mm CPTCI 1H-
PULPROG zg30
TD 32768
SOLVENT CDC13
NS 16
DS 2
SWH 12335.526 Hz
FIDRES 0.376450 Hz
AQ 1.3281963 sec
RG 20.2
DW 40.533 usec
DE 10.00 usec
TE 298.0 K
D1 1.00000000 sec
TD0 1

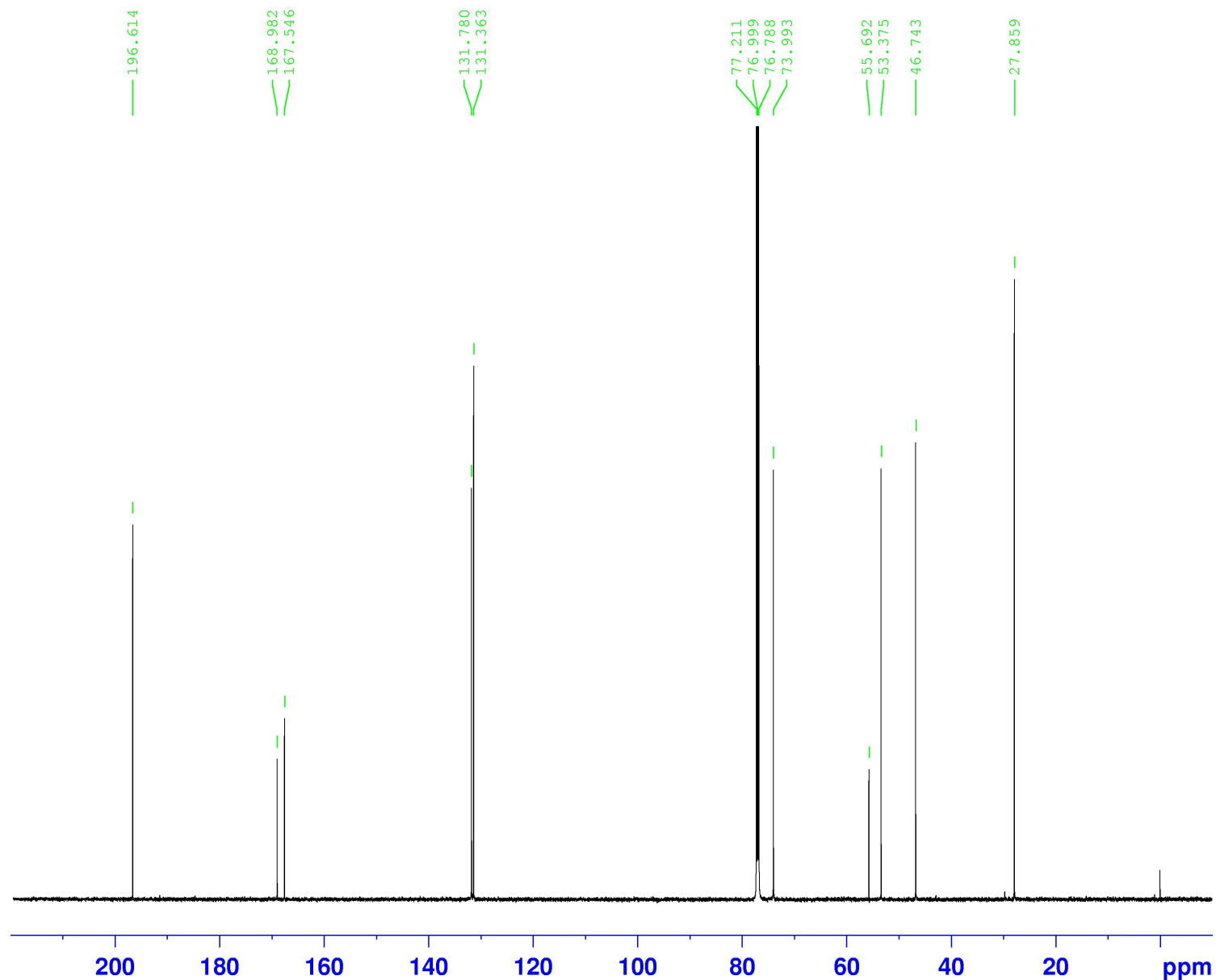
===== CHANNEL f1 =====
SFO1 600.1337060 MHz
NUC1 1H
P1 7.80 usec
PLW1 8.80000019 W

F2 - Processing parameters
SI 32768
SF 600.1300057 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



3aa

YH_0348_1_2nd
A.carbon CDCl3 D:\\ rfia 51



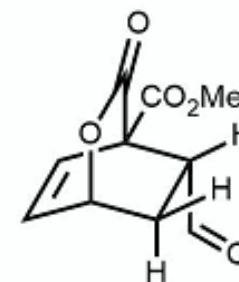
Current Data Parameters
NAME Hashimoto
EXPNO 1632
PROCNO 1

F2 - Acquisition Parameters
Date_ 20180801
Time 17.10
INSTRUM spect
PROBHD 5 mm CPTCI 1H-
PULPROG zgpg30
TD 65356
SOLVENT CDCl3
NS 2048
DS 4
SWH 36057.691 Hz
FIDRES 0.551712 Hz
AQ 0.9062698 sec
RG 2050
DW 13.867 usec
DE 6.50 usec
TE 298.0 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 150.9178988 MHz
NUC1 13C
P1 15.00 usec
PLW1 107.00000000 W

===== CHANNEL f2 =====
SFO2 600.1324005 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 70.00 usec
PLW2 8.80000019 W
PLW12 0.10926000 W
PLW13 0.05353900 W

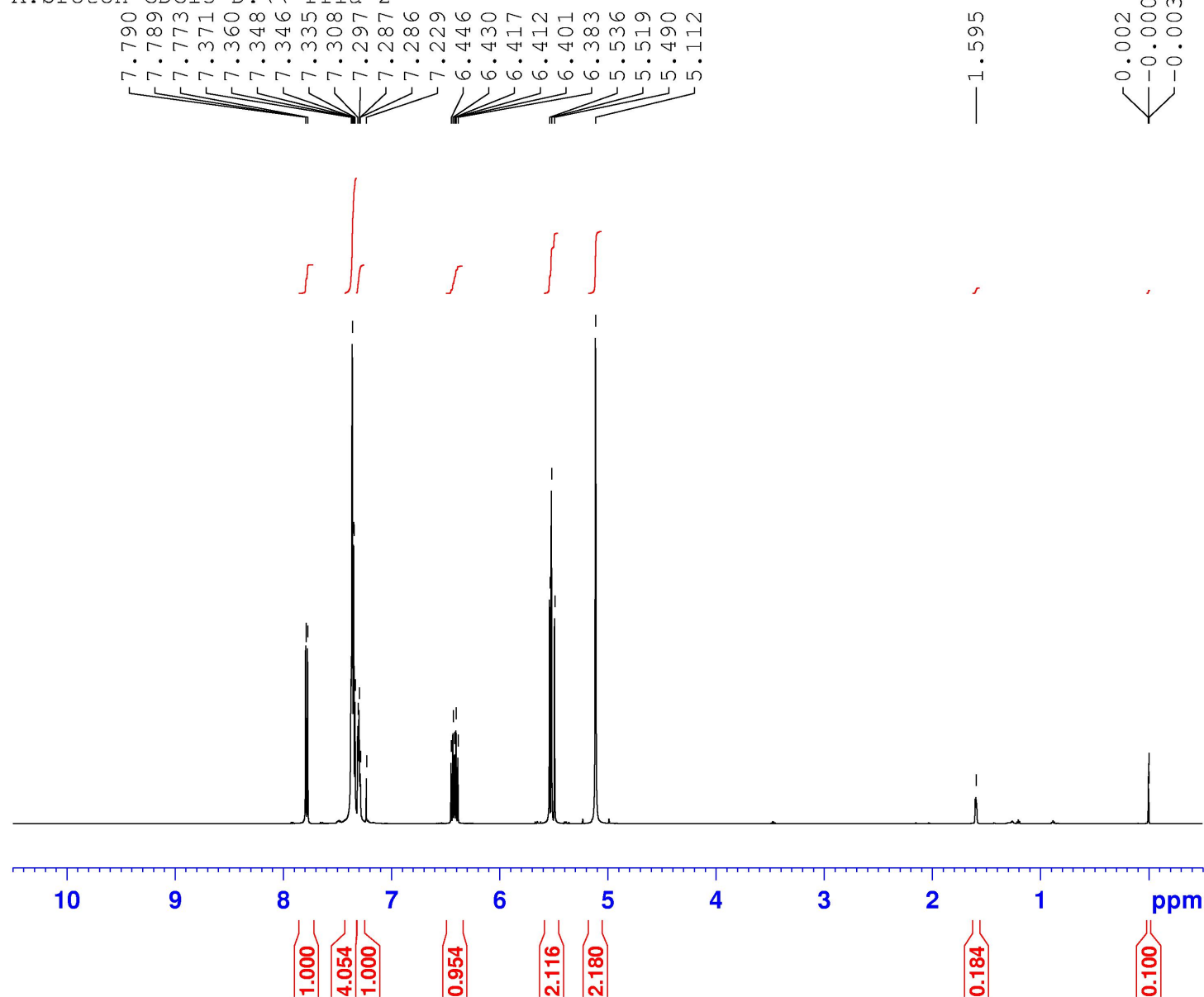
F2 - Processing parameters
SI 32768
SF 150.9028125 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



3aa

20170711_ikeda_55_1

A.proton CDCl3 D:\rfia 2

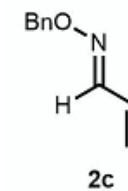


Current Data Parameters
NAME Hashimoto
EXPNO 790
PROCNO 1

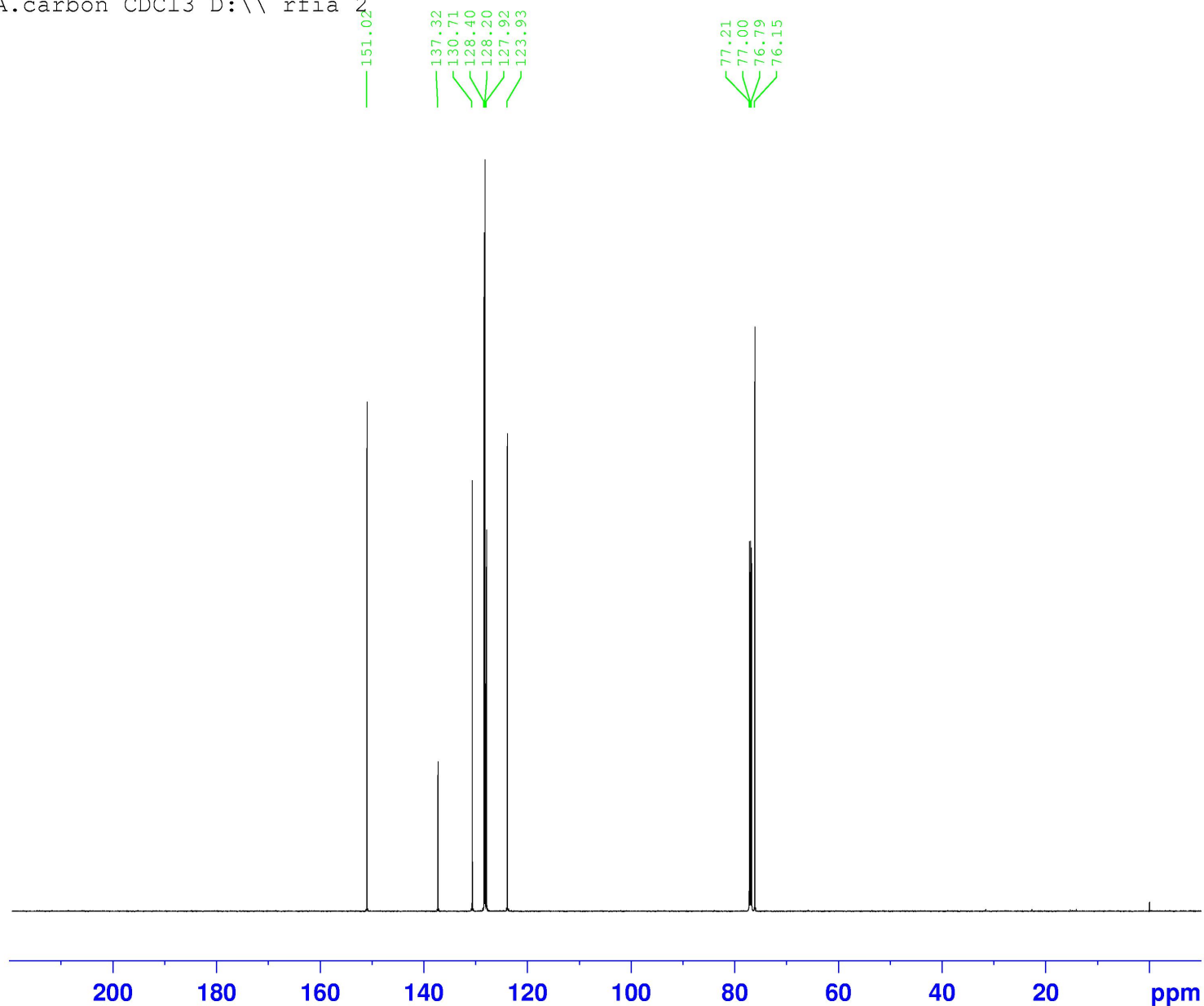
F2 - Acquisition Parameters
Date_ 20170711
Time 23.03
INSTRUM spect
PROBHD 5 mm CPTCI 1H-
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 16
DS 2
SWH 12335.526 Hz
FIDRES 0.376450 Hz
AQ 1.3281963 sec
RG 11.3
DW 40.533 usec
DE 10.00 usec
TE 298.0 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 600.1337060 MHz
NUC1 1H
P1 7.40 usec
PLW1 8.80000019 W

F2 - Processing parameters
SI 32768
SF 600.1300348 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



20170711_ikeda_55_1
A.carbon CDCl3 D:\ rfia 2



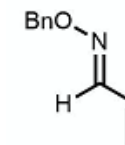
Current Data Parameters
NAME Hashimoto
EXPNO 791
PROCNO 1

F2 - Acquisition Parameters
Date_ 20170712
Time 0.45
INSTRUM spect
PROBHD 5 mm CPTCI 1H-
PULPROG zgpg30
TD 65356
SOLVENT CDCl3
NS 2048
DS 4
SWH 36057.691 Hz
FIDRES 0.551712 Hz
AQ 0.9062698 sec
RG 2050
DW 13.867 usec
DE 6.50 usec
TE 298.0 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 150.9178988 MHz
NUC1 13C
P1 15.00 usec
PLW1 102.00000000 W

===== CHANNEL f2 =====
SFO2 600.1324005 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 70.00 usec
PLW2 8.80000019 W
PLW12 0.09834400 W
PLW13 0.04818900 W

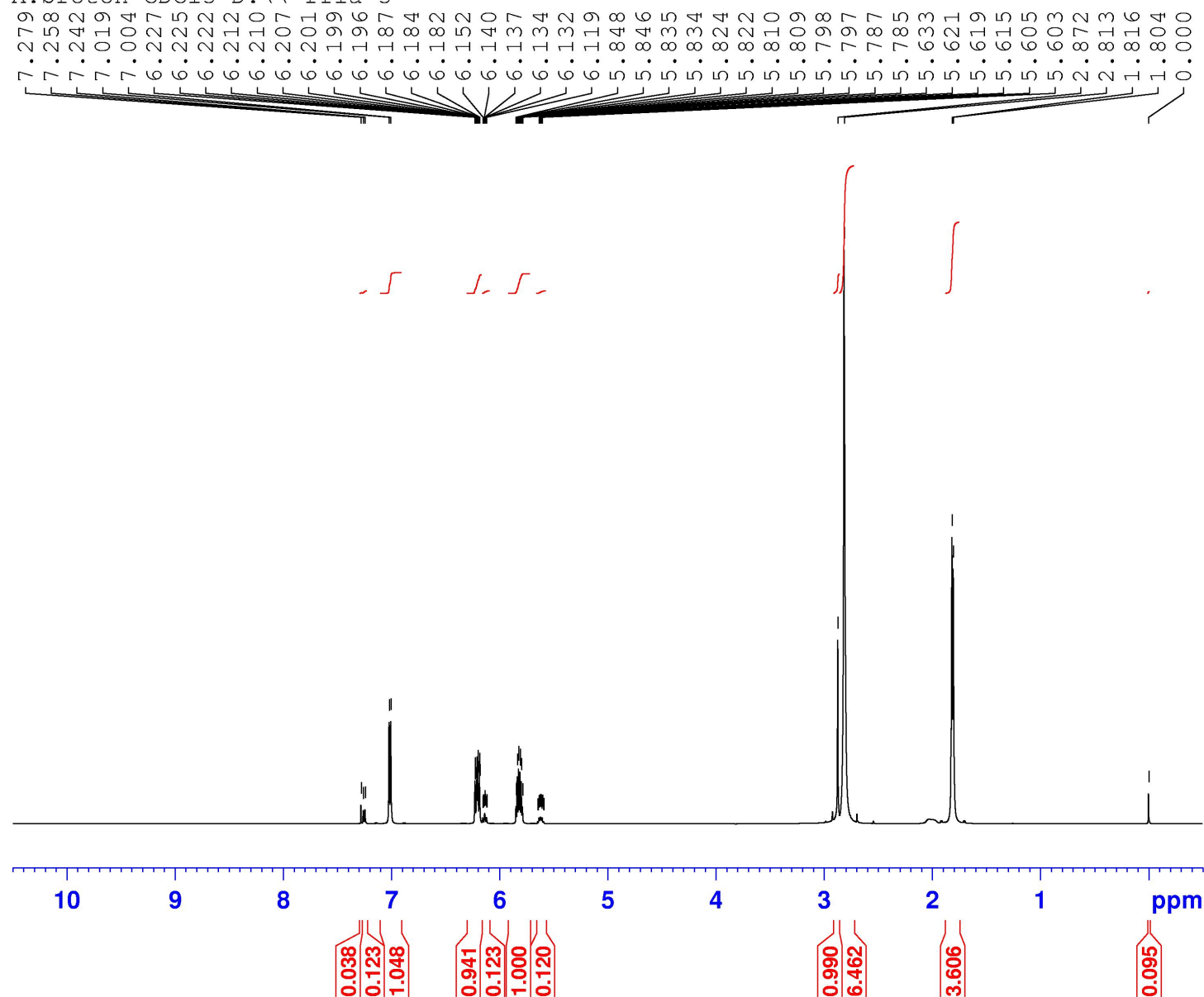
F2 - Processing parameters
SI 32768
SF 150.9028202 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



2c

20170711_iked_a_56_1

A.proton CDC13 D:\ rfia 3

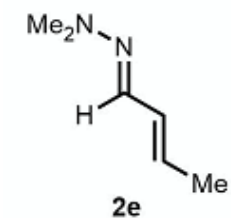


Current Data Parameters
NAME Hashimoto
EXPNO 800
PROCNO 1

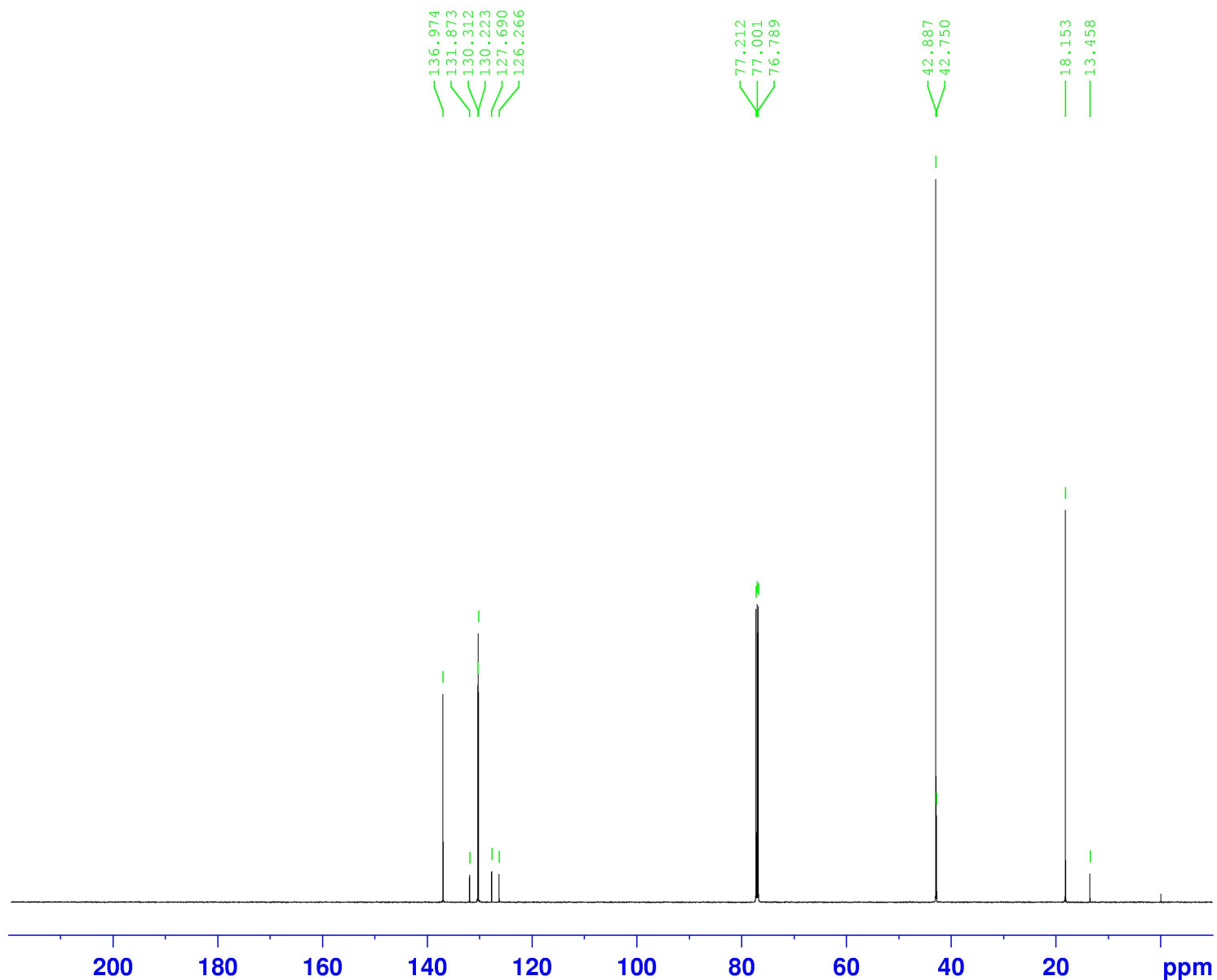
F2 - Acquisition Parameters
Date_ 20170712
Time 0.50
INSTRUM spect
PROBHD 5 mm CPTCI 1H-
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 16
DS 2
SWH 12335.526 Hz
FIDRES 0.376450 Hz
AQ 1.3281963 sec
RG 10
DW 40.533 usec
DE 10.00 usec
TE 298.0 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 600.1337060 MHz
NUC1 1H
P1 7.40 usec
PLW1 8.80000019 W

F2 - Processing parameters
SI 32768
SF 600.1300054 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



20170711_ikeda_56_1
A.carbon CDCl3 D:\\ rfia 3



Current Data Parameters
NAME Hashimoto
EXPNO 801
PROCNO 1

F2 - Acquisition Parameters
Date_ 20170712
Time 2.32
INSTRUM spect
PROBHD 5 mm CPTCI 1H-
PULPROG zgpg30
TD 65356
SOLVENT CDCl3
NS 2048
DS 4
SWH 36057.691 Hz
FIDRES 0.551712 Hz
AQ 0.9062698 sec
RG 2050
DW 13.867 usec
DE 6.50 usec
TE 298.0 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 150.9178988 MHz
NUC1 13C
P1 15.00 usec
PLW1 102.00000000 W

===== CHANNEL f2 =====
SFO2 600.1324005 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 70.00 usec
PLW2 8.80000019 W
PLW12 0.09834400 W
PLW13 0.04818900 W

F2 - Processing parameters
SI 32768
SF 150.9028180 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

