# Supporting Information

# Synthesis of Chiral Seven-membered β-Substituted Lactams via Rh-

# catalyzed Asymmetric Hydrogenation

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# Contents

I. General remarks	2
II. General procedure for the preparation of substrates	3
III. General procedure for the Rh-catalyzed asymmetric hydrogenation	12
IV. Gram-scale asymmetric hydrogenation of model substrate	18
V. Transformation for hydrogenation product <b>2f</b>	19
VI. Reference	20
VII. NMR spectra	21
VIII. HPLC spectra	

## I. General remarks

All the reactions dealing with air- or moisture-sensitive compounds were carried out in a dry reaction vessel under a positive pressure of nitrogen or in the argon-filled glovebox. Unless otherwise noted, all reagents and solvents were purchased from commercial suppliers without further purification. The anhydrous CH<sub>2</sub>Cl<sub>2</sub> and MeOH (superdry solvent, 99.9%, water  $\leq$  30 ppm, with molecular sieves.) were purchased from J&K Chemical Technology company and transferred by syringe. <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were recorded on a Bruker ADVANCE III (400 MHz) spectrometer with CDCl<sub>3</sub> as the solvent and tetramethylsilane (TMS) as the internal standard. Chemical shifts are reported in parts per million (ppm,  $\delta$  scale) downfield from TMS at 0.00 ppm and referenced to the CDCl<sub>3</sub> at 7.26 ppm (for <sup>1</sup>H NMR) or 77.0 ppm (for <sup>13</sup>C NMR). Data are reported as: multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet), coupling constant in hertz (Hz) and signal area integration in natural numbers. <sup>13</sup>C NMR and <sup>1</sup>H NMR analyses were run with decoupling. Enantiomeric excess values were determined by Daicel chiral column on an Agilent 1260 Series HPLC instrument. Optical rotations  $[\alpha]_D$  were measured on a PERKIN ELMER polarimeter 343 instrument. High resolution mass spectra (HRMS) were recorded on DIONEX UltiMate 3000& Bruker Compact TOF mass spectrometer. The substrate 4phenyl-1,5,6,7-tetrahydro-2H-azepin-2-one 1a and its analogues were synthesized according to the literatures. <sup>1-6</sup> Compound **2f** was transformed into N-*p*-chlorobenzovl protected lactam 3 for the determination of absolute configuration. The absolute configuration of lactam 3 is (R), which was determined by X-ray analysis. <sup>7</sup> The absolute configurations of other hydrogenation products were assigned by analogy.

### **II.** General procedure for the preparation of substrates

 $\begin{array}{c} \overbrace{O}^{O} + EtOH \xrightarrow{TsOH H_2O (2 \text{ mol}\%)}_{\text{toluene, reflux, overnight}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O}^{O} + \frac{RMgX (2.0 \text{ eq.})}_{0 \text{ °C-rt}} & \overbrace{O$ 

General procedure for the synthesis of 1a and its analogues:

The 3-ethoxy-2-cyclohexen-1-one was synthesized according to the following procedures. <sup>1</sup> To a solution of 1,3-cyclohexanedione (100 mmol, 11.2 g, 1.0 equiv.) in toluene (50 mL) was added EtOH (760 mmol, 44 mL, 7.6 equiv.), *p*-TsOH·H<sub>2</sub>O (2 mmol, 0.02 equiv.) at ambient temperature. The flask was equipped with a reflux condenser and placed in oil bath maintained at 130 °C overnight. The reaction mixture was allowed to cool down to ambient temperature and the solvent was removed in *vacuo* to give orange oil. The crude product was dissolved in CH<sub>2</sub>Cl<sub>2</sub> and neutralized with aqueous NaOH (0.1 M). The organic phase and aqueous phase were separated, and the organic phases were combined, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and the solvent was removed under reduced pressure. The resulting crude product was purified by flash column chromatography using PE/EA (V: V = 10:1) as eluent to afford pure product as pale orange or colorless oil (11.7 g, 84% yield).

The corresponding cyclic unsaturated ketone was obtained according to the following procedure. <sup>1-2</sup> A flame-dried flask was charged with THF solution (30 mL) of corresponding Grignard reagent (2.0 equiv., 60 mmol) and cooled to 0 °C. 3-ethoxy-2-cyclohexen-1-one (1.0 equiv., 4.2 g, 30 mmol) was added to the solution dropwise. Once the addition was completed, the reaction mixture was warmed to room temperature and stirred for 2 h until the completion of the disappearance of starting material. The reaction mixture was placed in an ice bath and 0.2 M aqueous H<sub>2</sub>SO<sub>4</sub> was added to quench the reaction, and then extracted with CH<sub>2</sub>Cl<sub>2</sub>. The combined extracts were washed with brine, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and the solvent was removed under reduced pressure. The resulting crude product was purified by flash column

chromatography using PE/EA (10:1, V: V) as eluent to give desired product as yellow solid.

The oximes were obtained from corresponding ketone according to the following procedures. <sup>3</sup> The product corresponding ketone from last step (1.0 equiv., 20 mmol) was dissolved in MeOH (20 mL), and successive addition of hydroxylamine hydrochloride (1.2 equiv., 24 mmol, 1.7 g) and sodium acetate trihydrate (2.4 equiv., 48 mmol, 6.5 g). The flask was equipped with a reflux condenser and placed in oil bath maintained at 70 °C overnight or for 4 h. The reaction mixture was allowed to cool down to ambient temperature, washed with brine and extracted with CH<sub>2</sub>Cl<sub>2</sub>. The combined extracts were dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and the solvent was used for next step directly without further purification.

# Method A for the synthesis of seven-membered $\beta$ -substituted $\alpha$ , $\beta$ -unsaturated lactams.<sup>4</sup>

To PPA (polyphosphoric acid, 10 mL) at 120 °C was added the oxime (12.8 mmol) by portionwise over 5 min. The mixture was stirred at this temperature for 2 h resulting in a deep red-brown syrup. The syrup was poured slowly into ice water and then neutralized with 1 M aqueous NaHCO<sub>3</sub>, extracted with CH<sub>2</sub>Cl<sub>2</sub>. The combined organic phases were dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>. The resulting crude product was purified by flash column chromatography using PE/EA (V: V = 1: 5) as eluent to provide crude product, and the finally desired product was obtained after recrystallization from diethyl ether and CH<sub>2</sub>Cl<sub>2</sub>.

# Method B for the synthesis of seven-membered $\beta$ -substituted $\alpha$ , $\beta$ -unsaturated lactams.<sup>5</sup>

A solution of thionyl chloride (16 equiv.) in 1,4-dioxane was added dropwise into a stirred solution of oxime (1.0 equiv.) in 1,4-dioxane (0.5 M) in a 10 mL round-bottom flask at 0 °C. The resulting mixture was then placed at 70 °C and stirred for 6 h. The mixture was concentrated under *vacuo* and diluted with  $CH_2Cl_2$  and then the reaction was quenched with saturated aqueous sodium bicarbonate to adjust  $pH \approx 7$ . The organic phase was dried over anhydrous  $Na_2SO_4$  and concentrated under reduced pressure. The resulting crude product was purified by flash column chromatography using PE/EA (V: V = 1: 5) as eluent, and after which, recrystallized in  $CH_2Cl_2$  and petroleum ether to provide the desired product as brown solid.

Method C for the synthesis of N-*p*-Ts seven-membered  $\beta$ -phenyl substituted  $\alpha$ , $\beta$ unsaturated lactam 1j.



*n*-BuLi (2.4 M in hexane, 0.1 mL, 1.1 eq.) was added into the THF (1.0 mL) solution of  $\beta$ -phenyl substituted  $\alpha$ , $\beta$ -unsaturated lactam **1a** (0.21 mmol, 39.3 mg, 1.0 eq.) at -78 °C under N<sub>2</sub> atmosphere and stirred for 1 h, and 1.0 mL THF solution of *p*-TsCl (0.23 mmol, 44.0 mg, 1.1 eq.) was transferred into this reaction mixture at the same temperature under N<sub>2</sub> atmosphere. After stirring at -78 °C for another 5 min, the cooling bath was removed and warmed to room temperature and stirred for 1 h under N<sub>2</sub> atmosphere. The reaction mixture was subjected to flash column chromatography directly and the finally desired product N-Ts  $\beta$ -phenyl substituted  $\alpha$ , $\beta$ -unsaturated lactam **1j** was obtained as white solid (58.0 mg, 81% yield).

# Method D for the synthesis of N-Boc seven-membered $\beta$ -phenyl substituted $\alpha$ , $\beta$ unsaturated lactam 1k.<sup>6</sup>



The N-Boc substituted lactam was synthesized according to the following procedures: **1a** (2.0 mmol), DMAP (0.1 eq., 0.2 mmol), and NEt<sub>3</sub> (1.5 eq., 3.0 mmol) were dissolved in CH<sub>2</sub>Cl<sub>2</sub> (4.0 mL), followed by addition of  $(Boc)_2O$  (2.0 eq., 4.0

mmol). The reaction mixture was stirred at room temperature for another 5 h. The reaction mixture was subjected to flash column chromatography directly and the finally desired N-Boc product **1k** was obtained as white solid (487 mg, 85% yield).

# Method E for the synthesis of tetrasubstituted (3-methyl-4-phenyl-1,5,6,7-tetrahydro-2*H*-azepin-2one) lactam 1l



The 3-ethoxy-2-methyl-2-cyclohexen-1-one was synthesized according to the following procedures. To a solution of 2-methyl-1,3-cyclohexanedione (50 mmol, 6.3 g, 1.0 equiv.) in toluene (30 mL) was added EtOH (380 mmol, 22 mL, 7.6 equiv.), *p*-TsOH·H<sub>2</sub>O (1 mmol, 0.02 equiv.) at ambient temperature. The flask was equipped with a reflux condenser and placed in oil bath maintained at 130 °C overnight. The reaction mixture was allowed to cool down to ambient temperature and the solvent was removed in *vacuo* to give orange oil. The crude product was dissolved in CH<sub>2</sub>Cl<sub>2</sub> and neutralized with aqueous NaOH (0.1 M). The organic phase and aqueous phase were separated, and the organic phases were combined, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and the solvent was removed under reduced pressure. The resulting crude product was purified by flash column chromatography using PE/EA (V: V = 10:1) as eluent to afford pure product as yellow solid (2.6 g, 34% yield).

The cyclic unsaturated ketone 2-methyl-3-phenyl-2-cyclohexen-1-one was obtained according to the following procedure. A flame-dried flask was charged with THF solution (15 mL) of phenylmagnesium bromide (2.0 equiv., 34 mmol) and cooled to 0 °C. 3-ethoxy-2-methyl-2-cyclohexen-1-one (1.0 equiv., 2.6 g, 17 mmol) was added to the solution dropwise. Once the addition was completed, the reaction mixture was warmed to room temperature and stirred for 2 h until the completion of the

disappearance of starting material. The reaction mixture was placed in an ice bath and 0.2 M aqueous H<sub>2</sub>SO<sub>4</sub> was added to quench the reaction, and then extracted with CH<sub>2</sub>Cl<sub>2</sub>. The combined extracts were washed with brine, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and the solvent was removed under reduced pressure. The resulting crude product was purified by flash column chromatography using PE/EA (10:1, V: V) as eluent to give desired product as yellow viscous oil (1.6 g, 51% yield).

The oximes were obtained from corresponding ketone according to the following procedures. The product corresponding ketone from last step (1.0 equiv., 8.6 mmol, 1.6 g) was dissolved in MeOH (20 mL), and successive addition of hydroxylamine hydrochloride (1.2 equiv., 10.3 mmol, 716 mg) and sodium acetate trihydrate (2.4 equiv., 20.6 mmol, 2.8 g). The flask was equipped with a reflux condenser and placed in oil bath maintained at 70 °C overnight or for 4 h. The reaction mixture was allowed to cool down to ambient temperature, washed with brine and extracted with CH<sub>2</sub>Cl<sub>2</sub>. The combined extracts were dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and the solvent was removed from the mixture under reduced pressure. The resulting crude product was used for next step directly without further purification (1.65 g white solid, 95% yield).

To PPA (polyphosphoric acid, 10 mL) at 120 °C was added the oxime (8.2 mmol, 1.65 g) by portionwise over 5 min. The mixture was stirred at this temperature for 2 h resulting in a deep red-brown syrup. The syrup was poured slowly into ice water and then neutralized with 1 M aqueous NaHCO<sub>3</sub>, extracted with CH<sub>2</sub>Cl<sub>2</sub>. The combined organic phases were dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>. The resulting crude product was purified by flash column chromatography using PE/EA (V: V = 1: 5) as eluent to provide crude product, and the finally desired product was obtained as colorless crystal after recrystallization from diethyl ether and CH<sub>2</sub>Cl<sub>2</sub> (677 mg, 41% yield).

4-phenyl-1,5,6,7-tetrahydro-2*H*-azepin-2-one **1a** 



Brown solid; M. P. 133-134 °C; Total yield 38%; It was obtained according to

method A. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 7.47-7.44 (m, 2H), 7.40-7.32 (m, 3H), 6.19 (s, 1H), 3.36-3.32 (m, 2H), 2.80 (t, *J* = 8.0 Hz, 2H), 2.15-2.09 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 172.50, 150.42, 141.47, 128.50, 126.17, 122.75, 40.24, 30.81, 29.58. ESI-HRMS calculated for C<sub>12</sub>H<sub>13</sub>NONa<sup>+</sup> ([M+Na]<sup>+</sup>): 210.0889; Found: 210.0890.

4-(4-(trifluoromethyl)phenyl)-1,5,6,7-tetrahydro-2*H*-azepin-2-one **1b** 



Dark brown solid; M. P. 140-141 °C; Total yield 12%; It was obtained according to method B. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 7.64 (d, *J* = 8.0 Hz, 2H), 7.56 (d, *J* = 8.0 Hz, 2H), 7.28 (brs, 1H), 6.22 (s, 1H), 3.38-3.34 (m, 2H), 2.80 (t, *J* = 8.0 Hz, 2H), 2.18-2.12 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 171.74, 148.98, 145.14, 130.42 (q, *J* = 32.0 Hz), 126.59, 125.54 (q, *J* = 4.0 Hz), 124.47, 122.55, 40.22, 30.95, 29.42. ESI-HRMS calculated for C<sub>13</sub>H<sub>12</sub>F<sub>3</sub>NONa<sup>+</sup> ([M+Na]<sup>+</sup>): 278.0763; Found: 278.0766.

### 4-(4-fluorophenyl)-1,5,6,7-tetrahydro-2*H*-azepin-2-one 1c



Yellow solid; M. P. 150-152 °C; Total yield 45%. It was obtained according to method A. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 7.45-7.41 (m, 2H), 7.15 (brs, 1H), 7.08-7.04 (m, 2H), 6.14 (s, 1H), 3.36-3.32 (m, 2H), 2.78 (t, *J* = 4.0 Hz, 2H), 2.15-2.08 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 172.22, 162.87 (d, *J* = 247.0 Hz), 149.37, 137.52 (d, *J* = 3.0 Hz), 127.98 (d, *J* = 8.0 Hz), 122.74 (d, *J* = 1.0 Hz), 115.46 (d, *J* = 21.0 Hz), 40.26, 31.00, 29.42. ESI-HRMS calculated for C<sub>12</sub>H<sub>12</sub>FNONa<sup>+</sup> ([M+Na]<sup>+</sup>): 228.0795; Found: 228.0797.

4-(3-methoxyphenyl)-1,5,6,7-tetrahydro-2H-azepin-2-one 1d



Dark brown solid; M. P. 101-102 °C; Total yield 14%. It was obtained according to method B. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 7.49 (brs, 1H), 7.31-7.27 (m, 1H), 7.04 (d, *J* = 8.0 Hz, 1H), 6.98 (s, 1H), 6.91-6.88 (m, 1H), 6.19 (s, 1H), 3.83 (s, 3H), 3.35-3.31 (m, 2H), 2.78 (t, *J* = 8.0 Hz, 2H), 2.14-2.08 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 172.48, 159.52, 150.29, 142.96, 129.48, 122.81, 118.60, 113.92, 111.83, 55.21, 40.20, 30.86, 29.57. ESI-HRMS calculated for C<sub>13</sub>H<sub>15</sub>NO<sub>2</sub>Na<sup>+</sup> ([M+Na]<sup>+</sup>): 240.0995; Found: 240.0998.

4-(p-tolyl)-1,5,6,7-tetrahydro-2H-azepin-2-one 1e



Dark brown solid; M. P. 147-148 °C; Total yield 13%. It was obtained according to method B. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 7.36 (d, J = 8.0 Hz, 2H), 7.19-7.17 (m, 3H), 6.17 (s, 1H), 3.34-3.30 (m, 2H), 2.78 (t, J = 8.0 Hz, 2H), 2.37 (s, 3H), 2.14-2.07 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 172.63, 150.36, 138.62, 138.48, 129.21, 126.10, 121.97, 40.31, 30.71, 29.57, 21.10. ESI-HRMS calculated for C<sub>13</sub>H<sub>15</sub>NONa<sup>+</sup> ([M+Na]<sup>+</sup>): 224.1046; Found: 224.1049.

4-(4-methoxyphenyl)-1,5,6,7-tetrahydro-2H-azepin-2-one 1f



Pale brown solid; M. P. 122-123 °C; Total yield 27%. It was obtained according to method A. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 7.42 (d, *J* = 8.0 Hz, 2H), 6.90 (d, *J* = 8.0 Hz, 2H), 6.78 (brs, 1H), 6.15 (d, *J* = 4.0 Hz, 1H), 3.83 (s, 3H), 3.35-3.30 (m, 2H), 2.78 (t, *J* = 8.0 Hz, 2H), 2.13-2.07 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 172.60, 160.01, 149.91, 133.58, 127.53, 121.13, 113.88, 55.31, 40.37, 30.61, 29.52. ESI-HRMS calculated for C<sub>13</sub>H<sub>15</sub>NO<sub>2</sub>Na<sup>+</sup> ([M+Na]<sup>+</sup>): 240.0995; Found: 240.0999.

4-(2-naphthenyl)-1,5,6,7-tetrahydro-2H-azepin-2-one 1g



White solid; M. P. 170-172 °C; Total yield 45%. It was obtained according to method A. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 7.92 (s, 1H), 7.86-7.82 (m, 3H), 7.60-7.57 (m, 2H), 7.52-7.49 (m, 2H), 6.34 (s, 1H), 3.40-3.35 (m, 2H), 2.91 (t, *J* = 7.2 Hz, 2H), 2.20-2.14 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 172.38, 150.23, 138.65, 133.15, 133.08, 128.28, 128.25, 127.54, 126.52, 126.49, 125.49, 124.08, 123.19, 40.35, 30.79, 29.62. ESI-HRMS calculated for C<sub>16</sub>H<sub>15</sub>NONa<sup>+</sup> ([M+Na]<sup>+</sup>): 260.1046; Found: 260.1050.

4-(3,5-dimethylphenyl)-1,5,6,7-tetrahydro-2*H*-azepin-2-one **1h** 



White solid; M. P. 206-208 °C; Total yield 8%. It was obtained according to method A. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 7.07 (s, 2H), 6.99 (s, 1H), 6.70 (brs, 1H), 6.16 (d, *J* = 4.0 Hz, 1H), 3.35-3.30 (m, 2H), 2.78 (t, *J* = 7.2 Hz, 2H), 2.33 (s, 6H), 2.14-2.07 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 172.35, 150.77, 141.59, 138.08, 130.20, 124.14, 122.45, 40.40, 30.95, 29.59, 21.29. ESI-HRMS calculated for C<sub>14</sub>H<sub>17</sub>NONa<sup>+</sup> ([M+Na]<sup>+</sup>): 238.1202; Found: 238.1205.

4-benzyl-1,5,6,7-tetrahydro-2*H*-azepin-2-one 1i



Brown solid; M. P. 120-121 °C; Total yield 2%. It was obtained according to method A. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 7.32-7.28 (m, 2H), 7.25-7.19 (m, 3H), 6.57 (brs, 1H), 5.84 (s, 1H), 3.45 (s, 2H), 3.21-3.17 (m, 2H), 2.31 (t, *J* = 8.0 Hz, 2H),

1.84-1.78 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 171.41, 152.76, 138.10, 128.98, 128.57, 126.71, 122.87, 46.28, 40.86, 31.92, 28.55. ESI-HRMS calculated for C<sub>13</sub>H<sub>15</sub>NONa<sup>+</sup> ([M+Na]<sup>+</sup>): 224.1046; Found: 224.1046.

4-phenyl-1-tosyl-1,5,6,7-tetrahydro-2H-azepin-2-one 1j



White solid; M. P. 94-97 °C; Total yield 31%. It was obtained according to method A and C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 7.98 (d, *J* = 8.0 Hz, 2H), 7.40-7.33 (m, 7H), 6.09 (s, 1H), 4.00 (t, *J* = 8.0 Hz, 2H), 2.84 (t, *J* = 8.0 Hz, 2H), 2.44 (s, 3H), 2.29-2.22 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 169.11, 151.96, 144.68, 140.04, 136.24, 129.39, 129.33, 128.77, 128.55, 126.26, 122.01, 45.00, 28.89, 28.33, 21.65. ESI-HRMS calculated for C<sub>19</sub>H<sub>19</sub>NO<sub>3</sub>SNa<sup>+</sup> ([M+Na]<sup>+</sup>): 364.0978; Found: 364.0984.

tert-butyl 7-oxo-5-phenyl-2,3,4,7-tetrahydro-1*H*-azepine-1-carboxylate 1k



White solid; M. P. 52-54 °C; Total yield 32%. It was obtained according to method A and D. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 7.47-7.44 (m, 2H), 7.42-7.35 (m, 3H), 6.24 (s, 1H), 3.85 (t, *J* = 6.4 Hz, 2H), 2.79 (t, *J* = 7.2 Hz, 2H), 2.13-2.07 (m, 2H), 1.56 (s, 9H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 170.27, 151.99, 150.46, 140.55, 129.05, 128.70, 126.30, 123.72, 82.77, 43.46, 28.19, 28.06, 27.00. ESI-HRMS calculated for C<sub>17</sub>H<sub>21</sub>NO<sub>3</sub>Na<sup>+</sup> ([M+Na]<sup>+</sup>): 310.1414; Found: 310.1410.

2-methyl-3-phenyl-1,5,6,7-tatrahydro-2*H*-azepin-2-one 11



White crystal; M. P. 130-132 °C; Total yield 7%; It was obtained according to method E. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 7.75 (brs, 1H), 7.35 (t, *J* = 7.2 Hz,

2H), 7.27-7.20 (m, 3H), 2.55 (q, J = 7.2 Hz, 4H), 2.25-2.18 (m, 2H), 1.82 (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 176.31, 141.61, 130.60, 129.07, 128.41, 128.20, 126.59, 34.11, 32.33, 29.78, 18.71. ESI-HRMS calculated for C<sub>13</sub>H<sub>15</sub>NONa<sup>+</sup> ([M+Na]<sup>+</sup>): 224.1051; Found: 224.1045.

### III. General procedure for the Rh-catalyzed asymmetric hydrogenation

In an argon-filled glove-box, a solution of ZhaoPhos L1 (6.6 mol%) and Rh(NBD)<sub>2</sub>BF<sub>4</sub> (6.0 mol%) in the total volume of 0.5 mL of anhydrous CH<sub>2</sub>Cl<sub>2</sub> and MeOH (V/V = 1:1) was stirred at room temperature for 2.0 h. 0.2 mmol substrate **1** was transferred into the resulting catalytic complex to make the concentration of substrate 0.4 M. And then stirred for another 5 min. The vials were transferred into autoclave, which was then charged with 50 atm H<sub>2</sub> and stirred at room temperature for 48 h. The hydrogen gas was released carefully and the solution was concentrated and purified through a short celite column on silica gel to remove the metal complex. The product was analyzed by <sup>1</sup>H NMR spectra for conversion-determination and purification by flash column chromatography using PE/EA (V: V = 1: 5) to provide desired hydrogenation product, which was further analyzed by chiral HPLC for ee values determination.

The absolute configuration of product 2f was determined by the X-ray analysis of its N-*p*-chlorobenzoyl derivative **3**, <sup>7</sup> and the others were determined by analogy.

## (*R*)-4-phenylazepan-2-one **2a**



The title compound was purified by flash column chromatography using PE/EA (V: V = 1: 5) to provide desired hydrogenation product as brown solid; M. P. 174-177 °C; >99% conv.; 37.1 mg, 98% yield; 93% ee;  $[\alpha]_D^{20} = -49.40$  (c = 0.5, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on chiralpak AD-H column, hexane: isopropanol = 90:10; flow rate = 1.0 mL/min; UV detection at 210 nm; t<sub>R</sub> = 11.3 min

(minor),  $t_R = 12.6 \text{ min}$  (major). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 7.32-7.29 (m, 2H), 7.23-7.18 (m, 3H), 6.34 (brs, 1H), 3.38-3.25 (m, 2H), 2.97-2.85 (m, 2H), 2.61 (d, J = 12.0 Hz, 1H), 2.15-2.11 (m, 1H), 1.99-1.92 (m, 1H), 1.83-1.65 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 177.13, 146.86, 128.59, 126.41, 126.32, 43.61, 42.65, 40.66, 39.31, 29.73. ESI-HRMS calculated for C<sub>12</sub>H<sub>15</sub>NONa<sup>+</sup> ([M+Na]<sup>+</sup>): 212.1046; Found: 212.1048.

(*R*)-4-(4-(trifluoromethyl)phenyl)azepan-2-one **2b** 



The title compound was purified by flash column chromatography using PE/EA (V: V = 1: 5) as eluent to give desired hydrogenation product as orange solid; M. P. 142-144 °C; >99% conv.; 50.4 mg, 98% yield; 93% ee;  $[\alpha]_D^{20}$  = -38.70 (c = 1.2, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on chiralpak AD-H column, hexane: isopropanol = 90:10; flow rate = 1.0 mL/min; UV detection at 210 nm; t<sub>R</sub> = 13.8 min (minor), t<sub>R</sub> = 12.1 min (major). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 7.57 (d, *J* = 8.0 Hz, 2H), 7.31 (d, *J* = 8.0 Hz, 2H), 6.66 (brs, 1H), 3.40-3.26 (m, 2H), 2.99-2.92 (m, 2H), 2.62-2.57 (m, 1H), 2.14-2.10 (m, 1H), 2.00-1.96 (m, 1H), 1.86-1.63 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 176.70, 150.67, 128.75 (q, *J* = 33.0 Hz), 126.72, 125.60 (q, *J* = 4.0 Hz), 124.11 (d, *J* = 270.0 Hz), 43.25, 42.50, 40.51, 39.04, 29.56. ESI-HRMS calculated for C<sub>13</sub>H<sub>14</sub>F<sub>3</sub>NONa<sup>+</sup> ([M+Na]<sup>+</sup>): 280.0920; Found: 280.0919.

(*R*)-4-(4-fluorophenyl)azepan-2-one 2c

The title compound was purified by flash column chromatography using PE/EA (V: V = 1:5) as eluent to give desired hydrogenation product as orange solid; M. P. 189-190 °C; >99% conv.; 40.6 mg, 98% yield; 91% ee;  $[\alpha]_D^{20} = -37.14$  (c = 0.7, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on chiralpak AD-H column, hexane:

isopropanol = 90:10; flow rate = 1.0 mL/min; UV detection at 210 nm;  $t_R$  = 13.3 min (minor),  $t_R$  = 14.4 min (major). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 7.17-7.13 (m, 2H), 7.02-6.97 (m, 2H), 6.50 (brs, 1H), 3.35-3.27 (m, 2H), 2.94-2.87 (m, 2H), 2.58-2.55 (m, 1H), 2.12-2.09 (m, 1H), 1.98-1.93 (m, 1H), 1.76-1.64 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 176.93, 161.32 (d, *J* = 243.0 Hz), 142.55 (d, *J* = 4.0 Hz), 127.72 (d, *J* = 8.0 Hz), 115.33 (d, *J* = 21.0 Hz), 43.78, 42.58, 39.95, 39.40, 29.64. ESI-HRMS calculated for C<sub>12</sub>H<sub>14</sub>FNONa<sup>+</sup> ([M+Na]<sup>+</sup>): 230.0952; Found: 230.0954.

(*R*)-4-(3-methoxyphenyl)azepan-2-one 2d



The title compound was purified by flash column chromatography using PE/EA (V: V = 1: 5) as eluent to give desired hydrogenation product as orange solid; M. P. 108-110 °C; >99% conv.; 42.5 mg, 97% yield; 86% ee;  $[\alpha]_D^{20}$  = -12.73 (c = 1.1, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on chiralcel OD-H column, hexane: isopropanol = 90:10; flow rate = 1.0 mL/min; UV detection at 210 nm; t<sub>R</sub> = 21.8 min (minor), t<sub>R</sub> = 28.7 min (major). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 7.22 (t, *J* = 8.0 Hz, 1H), 6.79-6.73 (m, 3H), 6.45 (brs, 1H), 3.80 (s, 3H), 3.38-3.25 (m, 2H), 2.95-2.82 (m, 2H), 2.62-2.59 (m, 1H), 2.15-2.12 (m, 1H), 1.98-1.93 (m, 1H), 1.83-1.60 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 177.27, 159.66, 148.48, 129.60, 118.69, 112.34, 111.45, 55.16, 43.55, 42.65, 40.69, 39.16, 29.66. ESI-HRMS calculated for C<sub>13</sub>H<sub>17</sub>NO<sub>2</sub>Na<sup>+</sup> ([M+Na]<sup>+</sup>): 242.1151; Found: 242.1153.

(*R*)-4-(*p*-tolyl)azepan-2-one **2e** 

The title compound was purified by flash column chromatography using PE/EA (V: V = 1: 5) as eluent to give desired hydrogenation product as orange solid; M. P. 139-140 °C; >99% conv.; 39.8 mg, 98% yield; 87% ee;  $[\alpha]_D^{20} = -31.81$  (c = 0.72, CHCl<sub>3</sub>);

The enantiomeric excess was determined by HPLC on chiralcel OD-H column, hexane: isopropanol = 90:10; flow rate = 1.0 mL/min; UV detection at 210 nm;  $t_R$  = 13.0 min (minor),  $t_R$  = 13.7 min (major). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 7.13-7.07 (m, 4H), 6.47 (brs, 1H), 3.39-3.23 (m, 2H), 2.95-2.82 (m, 2H), 2.60-2.57 (m, 1H), 2.32 (s, 3H), 2.12-2.09 (m, 1H), 1.95-1.92 (m, 1H), 1.81-1.60 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 177.29, 143.93, 135.89, 129.23, 126.17, 43.75, 42.64, 40.22, 39.38, 29.72, 20.96. ESI-HRMS calculated for C<sub>13</sub>H<sub>17</sub>NONa<sup>+</sup> ([M+Na]<sup>+</sup>): 226.1202; Found: 226.1204.

(*R*)-4-(4-methoxyphenyl)azepan-2-one 2f



The title compound was purified by flash column chromatography using PE/EA (V: V = 1: 5) as eluent to give desired hydrogenation product as brown solid; M. P. 138-139 °C; >99% conv.; 43.4 mg, 99% yield; 88% ee;  $[\alpha]_D^{20} = -44.22$  (c = 1.31, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on chiralcel OD-H column, hexane: isopropanol = 90:10; flow rate = 0.5 mL/min; UV detection at 210 nm; t<sub>R</sub> = 40.9 min (minor), t<sub>R</sub> = 43.6 min (major). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 7.13-7.09 (m, 2H), 6.86-6.82 (m, 2H), 6.63 (brs, 1H), 3.79 (s, 3H), 3.35-3.24 (m, 2H), 2.93-2.81 (m, 2H), 2.57 (d, *J* = 12.0 Hz, 1H), 2.12-2.09 (m, 1H), 1.95-1.91 (m, 1H), 1.80-1.63 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 177.40, 157.94, 139.11, 127.22, 113.85, 55.19, 43.95, 42.59, 39.79, 39.44, 29.67. ESI-HRMS calculated for C<sub>13</sub>H<sub>17</sub>NO<sub>2</sub>Na<sup>+</sup> ([M+Na]<sup>+</sup>): 242.1151; Found: 242.1156.

(*R*)-4-(2-naphthenyl)azepan-2-one **2g** 



The title compound was purified by flash column chromatography using PE/EA (V: V = 1:5) as eluent to give desired hydrogenation product as white solid; M. P. 176-178

<sup>o</sup>C; >99% conv.; 47.8 mg, 99% yield; 93% ee;  $[α]_D^{20} = -55.02$  (c = 1.43, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on chiralcel OD-H column, hexane: isopropanol = 90: 10; flow rate = 0.5 mL/min; UV detection at 220 nm; t<sub>R</sub> = 43.4 min (minor), t<sub>R</sub> = 45.8 min (major). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>), δ (ppm) 7.82-7.78 (m, 3H), 7.62 (s, 1H), 7.48-7.41 (m, 2H), 7.35-7.33 (m, 1H), 6.52 (brs, 1H), 3.39-3.29 (m, 2H), 3.06-2.99 (m, 2H), 2.70 (d, *J* = 12.0 Hz, 1H), 2.20 (d, *J* = 12.0 Hz, 1H), 2.00-1.69 (m, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>), δ (ppm) 177.15, 144.26, 133.50, 132.20, 128.29, 127.61, 127.56, 126.07, 125.46, 125.26, 124.38, 43.70, 42.66, 40.75, 39.15, 29.78. ESI-HRMS calculated for C<sub>16</sub>H<sub>17</sub>NONa<sup>+</sup> ([M+Na]<sup>+</sup>): 262.1202; Found: 262.1204.

(R)-4-(3,5-dimethylphenyl)azepan-2-one 2h



The title compound was purified by flash column chromatography using PE/EA (V: V = 1: 5) as eluent to give desired hydrogenation product as white solid; M. P. 156-157 °C. >99% conv.; 43.4 mg, 99% yield; 96% ee;  $[\alpha]_D^{20} = -28.64$  (c = 0.43, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on chiralcel OD-H column, hexane: isopropanol = 90: 10; flow rate = 0.5 mL/min; UV detection at 220 nm; t<sub>R</sub> = 24.9 min (minor), t<sub>R</sub> = 31.0 min (major). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 6.85 (s, 1H), 6.80 (s, 2H), 6.77 (brs, 1H), 3.37-3.23 (m, 2H), 2.93-2.77 (m, 2H), 2.56 (d, *J* = 16.0 Hz, 1H), 2.29 (s, 6H), 2.13-2.09 (m, 1H), 1.95-1.89 (m, 1H), 1.82-1.59 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 177.45, 146.89, 137.99, 127.95, 124.13, 43.86, 42.58, 40.53, 39.10, 29.73, 21.26. ESI-HRMS calculated for C<sub>14</sub>H<sub>19</sub>NONa<sup>+</sup> ([M+Na]<sup>+</sup>): 240.1359; Found: 240.1360.

(+)-4-benzylazepan-2-one 2i



The title compound was purified by flash column chromatography using PE/EA (V: V = 1: 5) as eluent to give desired hydrogenation product as brown solid; M. P. 110-111 <sup>o</sup>C; 86% conv.; 33.7 mg; 83% yield; 84% ee;  $[\alpha]_D^{20} = 1.91$  (c = 0.21, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on chiralcel OD-H column, hexane: isopropanol = 90: 10; flow rate = 1.0 mL/min; UV detection at 210 nm;  $t_R = 17.2$  min (minor),  $t_R = 15.5$  min (major). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 7.30-7.26 (m, 3H), 7.22-7.18 (m, 1H), 7.16-7.14 (m, 1H), 6.15 (brs, 1H), 3.22-3.19 (m, 2H), 2.75-2.70 (m, 1H), 2.57-2.41 (m, 3H), 2.01-1.97 (m, 1H), 1.88-1.78 (m, 2H), 1.53-1.33 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 177.35, 139.76, 129.18, 128.35, 126.17, 43.16, 42.76, 42.39, 35.86, 35.66, 28.60. ESI-HRMS calculated for C<sub>13</sub>H<sub>17</sub>NONa<sup>+</sup> ([M+Na]<sup>+</sup>): 226.1202; Found: 226.1202.

(R)-4-phenyl-1-tosylazepan-2-one 2j

The title compound was purified by flash column chromatography using PE/EA (V: V = 5: 1) as eluent to give the desired hydrogenation product as white solid; M. P. 113-115 °C; >99% conv.; 68.3 mg; 99% yield; >99% ee;  $[\alpha]_D^{20} = -1.47$  (c = 1.77, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on chiralcel AD-H column, hexane: isopropanol = 95: 5; flow rate = 1.0 mL/min; UV detection at 210 nm; t<sub>R</sub> = 70.5 min (minor), t<sub>R</sub> = 84.6 min (major). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 7.91 (d, *J* = 8.0 Hz, 2H), 7.33 (d, *J* = 8.0 Hz, 2H), 7.28 (t, *J* = 8.0 Hz, 2H), 7.22-7.18 (m, 1H), 7.10 (d, *J* = 8.0 Hz, 2H), 4.69-4.63 (m, 1H), 3.65-3.59 (m, 1H), 3.03-2.97 (m, 1H), 2.86-2.84 (m, 1H), 2.68 (d, *J* = 12.0 Hz, 1H), 2.44 (s, 3H), 2.19-2.05 (m, 2H), 1.79-1.73 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 173.16, 145.79, 144.63, 136.41, 129.31, 128.72, 128.55, 126.70, 126.21, 46.08, 45.62, 40.50, 38.03, 29.23, 21.66. ESI-HRMS calculated

for C<sub>19</sub>H<sub>21</sub>NO<sub>3</sub>SNa<sup>+</sup> ([M+Na]<sup>+</sup>): 366.1134; Found: 366.1143.

tert-butyl (R)-2-oxo-4-phenylazepane-1-carboxylate 2k



The title compound was purified by flash column chromatography using PE/EA (V: V = 5: 1) as eluent to give the desired hydrogenation product as white solid; M. P. 71-72 °C; >99% conv; 17.4 mg; 30% yield (70% Boc group was removed, determined by <sup>1</sup>H crude NMR); 99% ee;  $[\alpha]_D{}^{20}$  = -39.70 (c = 0.68, CHCl<sub>3</sub>); The enantiomeric excess was determined by HPLC on chiralcel OD-H column, hexane: isopropanol = 98: 2; flow rate = 1.0 mL/min; UV detection at 220 nm; t<sub>R</sub> = 16.2 min (minor), t<sub>R</sub> = 19.6 min (major). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 7.32-7.28 (m, 2H), 7.23-7.16 (m, 3H), 4.33 (dd, *J* = 6.4 Hz, *J* = 15.2 Hz, 1H), 3.43 (dd, *J* = 10.0 Hz, *J* = 15.2 Hz, 1H), 3.13-3.07 (m, 1H), 2.95 (t, *J* = 11.6 Hz, 1H), 2.78 (d, *J* = 13.6 Hz, 1H), 2.09-2.04 (m, 2H), 1.81-1.63 (m, 2H), 1.54 (s, 9H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 173.96, 152.80, 146.34, 128.62, 126.50, 126.28, 82.97, 46.33, 45.70, 41.06, 37.97, 28.46, 27.98. ESI-HRMS calculated for C<sub>17</sub>H<sub>23</sub>NO<sub>3</sub>Na<sup>+</sup> ([M+Na]<sup>+</sup>): 312.1570; Found: 312.1567.

### IV. Gram-scale asymmetric hydrogenation of model substrate

In an argon-filled glove box, a solution of Rh(NBD)<sub>2</sub>BF<sub>4</sub> (6.0 mol%) and ZhaoPhos L1 (6.6 mol%) in anhydrous CH<sub>2</sub>Cl<sub>2</sub> and MeOH (1: 1, V: V) was stirred at room temperature for 2 h at least. 0.51g model substrate **1a** (2.72 mmol) was transferred into the resulting complex to make the concentration 0.4 M, and stirred at the same temperature for another 5 min, and then was transferred into autoclave. 50 atm of H<sub>2</sub> was charged and stirred at room temperature for 48 h. The hydrogen gas was released carefully and the solution was concentrated and purified through a short celite column on silica gel to remove the metal complex. The product was analyzed by <sup>1</sup>H NMR spectra for conversion-determination and purification by flash column chromatography using PE/EA (V: V = 1: 5) to provide desired hydrogenation product **2a** (97%)

conversion, 95% yield), which was further analyzed by chiral HPLC for ee values determination (92% ee).

### V. Transformation for hydrogenation product 2f

(R)-1-(4-chlorobenzoyl)-4-(4-methoxyphenyl)azepan-2-one **3** 



The title compound was synthesized via following procedures: n-BuLi (1.1 equiv., 2.4 M in hexane, 0.11 mL, 0.275 mmol) was added dropwise to the THF (1.0 mL) solution of desired product 2f (1.0 equiv., 54.8 mg, 0.25 mmol) at -78 °C over 5 min. The resulting mixture was stirred at this temperature for 1 h, after which, the corresponding 4-chlorobenzoyl chloride (1.1 equiv., 0.275 mmol, 48.1 mg) was added in one portion. And the resulting reaction mixture was warmed to ambient temperature and stirred for overnight. After concentrating in *vacuo*, the resulting precipitate was directly subjected to flash column chromatography on silica gel using petroleum/ethyl acetate (V: V = 5: 1) to provide the desired corresponding product 3. The enantiomeric excess was determined by chiral HPLC, and absolute configuration was determined as (*R*) by X-ray crystallography; 70% isolated yield; 63.0 mg; 86% ee;  $\lceil \alpha \rceil_D^{20} = -2.54$  (c = 1.18, CHCl<sub>3</sub>); M. P. 123-125 °C. The enantiomeric excess was determined by chiral HPLC on chiralpak AD-H column, hexane/isopropanol = 90:10; flow rate = 1.0 ml/min; UV detection at 254 nm;  $t_R = 26.9 \text{ min}$  (minor),  $t_R = 31.4 \text{ min}$  (major). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) 7.45 (d, J = 8.0 Hz, 2H), 7.35 (d, J = 8.0 Hz, 2H), 7.13 (d, J =8.0 Hz, 2H), 6.87 (d, J = 8.0 Hz, 2H), 4.52 (dd, J = 4.0, 12.0 Hz, 1H), 3.80 (s, 3H), 3.58 (dd, J = 8.0, 12.0 Hz, 1H), 3.21-3.15 (m, 1H), 3.02-2.96 (m, 1H), 2.78 (d, J = 16.0 Hz, 1H), 2.22-2.12 (m, 2H), 1.92-1.74 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>), δ (ppm) 175.89, 173.08, 158.24, 137.74, 137.59, 134.80, 129.24, 128.43, 127.30, 114.07, 55.25, 45.75, 45.00, 40.29, 38.07, 28.69. ESI-HRMS calculated for  $C_{20}H_{20}CINO_3Na^+$  ([M+Na]<sup>+</sup>): 380.1024; Found: 380.1025.

### **VI. Reference**

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# VII. NMR spectra



























































#### **VIII. HPLC spectra**

Data File E:\DATA\HYI\HYI-2-25-RAC\HYI-2-25-RAC 2017-06-10 16-39-21\064-0201.D Sample Name: HYI-2-25-RAC-AD

\_\_\_\_\_ Acq. Operator : SYSTEM Seq. Line : 2 Acq. Instrument : 1260HPLC-VWD Location : Vial 64 Inj : 1 Inj Volume : 10.000 μl Injection Date : 6/10/2017 6:04:36 PM : E:\DATA\HYI\HYI-2-25-RAC\HYI-2-25-RAC 2017-06-10 16-39-21\VWD-ADH(1-2)-Acg. Method 90-10-1.0-210NM-60MIN.M : 6/10/2017 6:55:22 PM by SYSTEM Last changed (modified after loading) Analysis Method : E:\DATA\HYI\HYI-2-25-RAC\HYI-2-25-RAC 2017-06-10 16-39-21\VWD-ADH(1-2)-90-10-1.0-210NM-60MIN.M (Sequence Method) Last changed : 12/22/2017 12:00:25 PM by SYSTEM (modified after loading) Additional Info : Peak(s) manually integrated VWD1A, Wavelength=210 nm (E:\DATAWY\\HY1-2:25 RAC\HY1-2:25 RAC 2017-06-10 16:39-21\0640201.D) mAU 11.088 2000 12.468 HN 1750 (racemic)-1500 2a 1250 1000 750 500 250 o 10 18 16 14 min 12 Area Percent Report Sorted By Signal : Multiplier : 1.0000 Dilution 1.0000 : Do not use Multiplier & Dilution Factor with ISTDs Signal 1: VWD1 A, Wavelength=210 nm Peak RetTime Type Width Area Height Area [min] [mAU\*s] [mAU] ÷ [min] # ----|-----|-1 11.099 VB 0.3308 3.91619e4 1836.01221 49.4965 2 12.458 BB 0.3567 3.99587e4 1728.76416 50.5035 7.91206e4 3564.77637 Totals : \_\_\_\_\_

1260HPLC-DAD 12/22/2017 12:00:31 PM SYSTEM

Data File E:\DATA\HYI\HYI-2-62\LWD-2-164 2017-08-17 16-20-24\017-0401.D Sample Name: HYI-2-62-3



Data File E:\DATA\HYI\HYI-3-30\HYI-3-30 2018-01-29 09-31-17\031-0201.D Sample Name: HYI-3-30-4



Data File E:\DATA\HYI\HYI-3-30\HYI-3-30 2018-01-29 09-31-17\032-0301.D Sample Name: HYI-3-30-5



Data File E:\DATA\HYI\HYI-2-68\HYI-2-68 2017-08-25 21-13-09\031-0201.D Sample Name: HYI-2-68-2 Acq. Operator : SYSTEM Acq. Instrument : 1260HPLC-VWD Seg. Line : 2 Location : Vial 31 Injection Date : 8/25/2017 10:14:55 PM Inj: 1 Inj Volume : 10.000 µl : E:\DATA\HYI\HYI-2-68\HYI-2-68 2017-08-25 21-13-09\VWD-AD(1-2)-90-10-1ML-Acg. Method 210NM-20MIN-10UL.M Last changed : 8/25/2017 9:13:09 PM by SYSTEM Analysis Method : E:\DATA\HYI\HYI-2-68\HYI-2-68 2017-08-25 21-13-09\VWD-AD(1-2)-90-10-1ML-210NM-20MIN-10UL.M (Sequence Method) Last changed : 12/22/2017 12:11:53 PM by SYSTEM (modified after loading) Additional Info : Peak(s) manually integrated VWD1A,Wavelength=210 nm (E:\DATAWHYNHY1-268XHY1-2682017-08-2521-13-09'031-0201.D) mAU 140 13.279 14.479 120 -HN (racemic)-100 2c 80 -60 · 40 20 0 -20 14 12 16 18 min Area Percent Report Sorted Bv : Signal Multiplier 1.0000 : 1.0000 Dilution : Do not use Multiplier & Dilution Factor with ISTDs Signal 1: VWD1 A, Wavelength=210 nm Peak RetTime Type Width Height Area Area # [min] -- [min] [mAU\*s] [mAU] % 1 13.279 BV 0.3460 2708.44067 118.36437 50.1114 2 14.479 VB 0.3710 2696.39868 109.61091 49.8886 Totals : 5404.83936 227.97528 -----\*\*\* End of Report \*\*\*

1260HPLC-DAD 12/22/2017 12:12:41 PM SYSTEM

Data File E:\DATA\HYI\HYI-2-68\HYI-2-68 2017-08-25 21-13-09\033-0301.D Sample Name: HYI-2-68-1



Data File E:\DATA\YCC\YCC-2018-01-17\YCC-210-9 2018-02-05 10-19-21\021-0601.D Sample Name: HYI-m-0Me-20180205



Data File E:\DATA\LYH\LYH-2-357\LYH-2-357 2017-10-31 15-19-16\003-0401.D Sample Name: HYI-2-121-3-0D-CHIRAL

![](_page_51_Figure_1.jpeg)

1260HPLC-DAD 1/26/2018 4:32:01 PM SYSTEM

Data File E:\DATA\HYI\HYI-2-121\HYI-2-121 2017-10-30 19-38-13\004-0601.D Sample Name: HYI-2-121-5-RAC

Acq. Operator :	SYSTEM Seq. Line : 6	
Acq. Instrument :	1260HPLC-DAD Location : Vial 4	
injection Date :	10/30/2017 10:54:10 PM Inj : 1 Tri Volume : 10.000 r	1
Acq. Method :	E:\DATA\HYI\HYI-2-121\HYI-2-121 2017-10-30 19-38-13	3\DAD-OD(1-2)-90-10-1.
	OML-10UL-ALL-40MIN.M	
Last changed :	10/30/2017 7:38:13 PM by SYSTEM	
Analysis Method :	E:\DATA\HYI\HYI-2-121\HYI-2-121 2017-10-30 19-38-13 OML-10UL-ALL-40MIN.M (Sequence Method)	3\DAD-OD(1-2)-90-10-1.
Last changed :	1/26/2018 4:51:37 PM by SYSTEM	
Additional Info :	Peak(s) manually integrated	
DAD1 C, Sig=21	0,4 Ref≕off (E:\DATA\HYI\HYI-2-121\HYI-2-121 2017-10-30 19-38-13\004-0601.D)	
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Sorted By	: Signal	
Multiplier	: 1.0000	
Dilution	: 1.0000	
Do not use Multip)	ier & Dilution Factor with ISTDs	
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angunar r. PRPI 6,	weg droys wer-old	
Peak RetTime Type	Width Area Height Area	
# [min] 	(min) (mao's) (mao) % 	
1 13.124 BV	0.4556 2010.67249 67.09521 49.2823	
2 14.225 VB	0.6017 2069.23608 51.39780 50.7177	
Totals :	4079.90857 118.49301	
		-
	*** End of Report ***	
HPLC-DAD 1/26/2018	3 4:51:43 PM SYSTEM	Page 1 of 1

Data File E:\DATA\ZX\ZX-2-25\ZX-2-25 2017-11-25 10-12-47\064-2701.D Sample Name: HYI-2-142-2

![](_page_53_Figure_1.jpeg)

1260HPLC-DAD 1/26/2018 4:58:01 PM SYSTEM

Data File E:\DATA\YC\YC-5-188-2-0D\YC-5-188-2-0D 2017-10-28 16-55-16\016-0401.D Sample Name: HYI-2-100-4-p-0ME

![](_page_54_Figure_1.jpeg)

1260HPLC-VWD 1/31/2018 10:10:36 PM SYSTEM

Data File E:\DATA\HYI\HYI-2-121\HYI-2-121 2017-10-30 19-38-13\001-0201.D Sample Name: HYI-2-121-1

![](_page_55_Figure_1.jpeg)

Data File D:\DATA\HY\HY-4-17-6 20180828\HY-4-17-6 20180828 2018-08-28 11-47-51\042-0201.D Sample Name: HY-4-17-6

Acq. Operator :	Seq. Line : 2
Acq. Instrument :	Instrument 2 Location : Vial 42
injection date :	8/28/2018 11:59:56 AM INJ: 1
Aca. Method :	D:\DATA\HY\HY-4-17-6 20180828\HY-4-17-6 20180828 2018-08-28 11-47-51\DAD-0D
Acq. nethod .	(1-2)-90-10-0.5ML-3UL-ALL-60MIN.M
Last changed :	8/28/2018 11:44:59 AM
Analysis Method :	D:\METHOD\LWD\DAD-OJ(1-6)-85-15-1ML-3UL-ALL-50MIN.M
Last changed :	9/1/2018 12:24:09 PM
	(modified after loading)
Additional Info :	Peak(s) manually integrated
mAll	220,4 Rel=011 (D.:DATAIRT17-0 20180828)rt14-17-6 20180828 2018-08-28 11-47-51/042-0201.D)
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Dilution	: 1.0000
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Peak RetTime Type	Width Area Height Area
# [min]	[min] [mAU] %
1 43.047 BV	1.2825 6.58445e4 764.77026 49.0214
2 46.652 VB	1.5342 6.84735e4 656.86407 50.9786
Totals :	1.34318e5 1421.63434

Instrument 2 9/1/2018 12:24:16 PM

Data File D:\DATA\HY\HY-4-17-6 20180828\HY-4-17-6 20180828 2018-08-28 11-47-51\043-1201.D Sample Name: HY-4-17-5

![](_page_57_Figure_1.jpeg)

Instrument 2 9/1/2018 12:26:08 PM

Data File D:\DATA\LYH\LYH-3-523\LYH-3-523 2018-08-30 16-08-52\052-0601.D Sample Name: HY-4-21-6

Acq. Operator :	Seq. Line : 6
Acq. Instrument : Inst	trument 2 Location : Vial 52
Injection Date : 8/3	0/2018 8:19:50 PM Inj: 1
	Inj Volume : 3.000 µl
Acq. Method : D:	DATA\LYH\LYH-3-523\LYH-3-523 2018-08-30 16-08-52\DAD-OD(1-2)-90-10-0.5ML
- 31	L-ALL-60MIN.M
Last changed : 8/2	8/2018 11:44:59 AM
Analysis Method : D:	METHOD\LWD\DAD-OJ(1-6)-85-15-1ML-3UL-ALL-50MIN.M
Last changed : 9/2	/2018 12:16:19 PM
(mo	dified after loading)
Additional Info : Pea	k(s) manually integrated
DAD1 B, Sig=220,4	Ref=off (D:\DATA\LYH\LYH-3-523\LYH-3-523 2018-08-30 16-08-52\052-0601.D)
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Sorted By	: Signal
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Use Multiplier & Dili	tion Factor with ISIDS
Signal 1: DAD1 B, Sig	=220,4 Ret=ott
Peak RetTime Type W	dth Area Height Area
# [min] [r	in] [mAU*s] [mAU] %
1 24.445 BB 0	6744 1.02854e4 234.17816 50.2120
2 30.792 BB 0	9169 1.01985e4 170.24193 49.7880
Totals :	2.04839e4 404.42009

Instrument 2 9/1/2018 12:16:28 PM

Data File D:\DATA\HY\HY-4-21\HY-4-21 2018-08-31 09-48-53\003-0301.D Sample Name: HY-4-21-5

![](_page_59_Figure_1.jpeg)

Instrument 2 9/1/2018 12:21:39 PM

Data File E:\DATA\HYI\HYI-2-153-154\HYI-2-153-154 2017-12-09 22-08-04\052-0401.D Sample Name: HYI-2-153-5

Acq. Operator : SYSTEM	Seq. Line: 4
Acq. Instrument : 1260HPLC-DAD	Location : Vial 52
Injection Date : 12/10/2017 12:15:17 AM	Inj: 1
	Inj Volume : 2.000 µl
Acq. Method : E:\DATA\HYI\HYI-2-153-154	HYI-2-153-154 2017-12-09 22-08-04\DAD-OD(1-2)-90-
10-1ML-2UL-ALL-60MIN.M	
Last changed : 12/9/2017 10:08:04 PM by S	SYSTEM
Analysis Method : E:\DATA\HYI\HYI-2-153-154	HYI-2-153-154 2017-12-09 22-08-04\DAD-OD(1-2)-90-
10-1ML-2UL-ALL-60MIN.M (Se	equence Method)
Last changed : 9/5/2018 5:40:07 PM by SYS	STEM
(modified after loading)	
Additional Info : Peak(s) manually integrate	ed
DAD1 C, Sig=210,4 Ref=360,100 (E:\DATA\HYI\HYI-2-153-15	4\HYI-2-153-154 2017-12-09 22-08-04\052-0401.D)
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20 0 10 11 12 13 14 Area Percent Report Sorted By : Signal	15 16 17 18 19 min
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20 0 10 11 12 13 14 Area Percent Report Sorted By : Signal Multiplier : 1.0000 Dilution : 1.0000	15 16 17 18 19 min
20 0 10 11 12 13 14 Area Percent Report Sorted By : Signal Multiplier : 1.0000 Dilution : 1.0000 Do not use Multiplier & Dilution Factor with	15 16 17 18 19 min
20 0 10 11 12 13 14 Area Percent Report Area Percent Report Sorted By : Signal Multiplier : 1.0000 Dilution : 1.0000 Do not use Multiplier & Dilution Factor with	15 16 17 18 19 min
20 0 10 11 12 13 14 Area Percent Report  Sorted By : Signal Multiplier : 1.0000 Dilution : 1.0000 Do not use Multiplier & Dilution Factor with	15 16 17 18 19 min
20 	15 16 17 18 19 min
20 	15 16 17 18 19 min
20 0 10 11 12 13 14 Area Percent Report Area Percent Report Sorted By : Signal Multiplier : 1.0000 Dilution : 1.0000 Do not use Multiplier & Dilution Factor with Signal 1: DAD1 C, Sig=210,4 Ref=360,100 Peak RetTime Type Width Area Heigh	15 16 17 18 19 min
20 0 10 11 12 13 14 Area Percent Report Area Percent Report Sorted By : Signal Multiplier : 1.0000 Dilution : 1.0000 Do not use Multiplier & Dilution Factor with Signal 1: DAD1 C, Sig=210,4 Ref=360,100 Peak RetTime Type Width Area Heigh # [min] [min] [mAU*s] [mAU]	n ISTDs
20 0 10 11 12 13 4 Area Percent Report Area Percent Report Sorted By : Signal Multiplier : 1.0000 Dilution : 1.0000 Do not use Multiplier & Dilution Factor with Signal 1: DAD1 C, Sig=210,4 Ref=360,100 Peak RetTime Type Width Area Heigh # [min] [min] [mAU*s] [mAU] 	n ISTDs
20 0 10 11 12 13 4 	n ISTDs
20 0 10 11 12 13 4 	n ISTDs
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20         0         10       11       12       13       14         Area Percent Report         Area Percent Report         Sorted By : Signal         Multiplier       : 1.0000         Dilution       : 1.0000         Do not use Multiplier & Dilution Factor with         Signal 1: DAD1 C, Sig=210,4 Ref=360,100         Peak RetTime Type Width Area Heigh         # [min]       [min] [mAU*s]	nt Area % 

1260HPLC-DAD 9/5/2018 5:40:17 PM SYSTEM

Data File D:\DATA\LG\20180905\NI-SUBSTRATE-DATA 2018-09-05 09-21-25\023-1901.D Sample Name: HY-4-24-2

![](_page_61_Figure_1.jpeg)

Instrument 2 9/5/2018 5:37:09 PM

Data File D:\DATA\HY\HY-4-58\LWD-4-145-1 2018-10-12 08-51-15\014-0201.D Sample Name: Ts-20181012-RAC

![](_page_62_Figure_1.jpeg)

Data File D:\DATA\HY\HY-4-58\LWD-4-145-1 2018-10-12 01-31-31\012-0801.D Sample Name: HY-4-58-1

![](_page_63_Figure_1.jpeg)

Instrument 1 10/12/2018 11:48:19 AM

Data File D:\DATA\HY\HY-4-38-20181002\HY-4-38-20181002 2018-10-02 20-42-55\023-0201.D Sample Name: HY-4-38-1

![](_page_64_Figure_1.jpeg)

Instrument 2 10/18/2018 8:20:47 PM

Data File D:\DATA\HY\HY-4-38-20181002\HY-4-38-20181002 2018-10-02 20-42-55\024-0301.D Sample Name: HY-4-38-2

![](_page_65_Figure_1.jpeg)

Instrument 2 10/18/2018 8:26:04 PM

Data File D:\DATA\LWD\LWD-4-95-2\LWD-4-95-2 2018-07-12 20-29-34\033-2301.D Sample Name: HY-3-170-RAC

Acq. Operator : Seq. Line : 23 Location : Vial 33 Acq. Instrument : Instrument 1 Injection Date : 7/13/2018 2:31:37 PM Inj: 1 Inj Volume : 2.000 μl : D:\DATA\LWD\LWD-4-95-2\LWD-4-95-2 2018-07-12 20-29-34\VWD-AD(1-2)-90-10-1ML Acq. Method -2UL-254NM-60MIN.M Last changed : 7/13/2018 10:23:58 AM Analysis Method : D:\METHOD\CZY\VWD-AD(1-2)-97-3-1ML-3UL-210NM-10MIN.M Last changed : 7/18/2018 11:13:35 AM (modified after loading) Additional Info : Peak(s) manually integrated VWD1 A, Wavelength=254 nm (D:\DATA\LWD\LWD-4-95-2\LWD-4-95-2 2018-07-12 20-29-34\033-2301.D) mAU 26.399 (racemic)-OMe 200 31.070 CI. 3 150 100 50 0 15 20 25 30 35 40 mi Area Percent Report -----Sorted By : Signal Multiplier 1.0000 : Dilution : 1.0000 Use Multiplier & Dilution Factor with ISTDs Signal 1: VWD1 A, Wavelength=254 nm Peak RetTime Type Width Height Area Area # [min] [min] [mAU\*s] [mAU] % 1 26.399 BB 1 26.399 BB 0.6608 8693.81152 200.52046 50.0285 2 31.070 BB 0.7882 8683.89063 167.96379 49.9715 Totals : 1.73777e4 368.48425

Instrument 1 7/18/2018 11:16:23 AM

Data File D:\DATA\GUAN YUQING\LJ-3-164\LJ-3-164 2018-07-13 17-50-28\052-0701.D Sample Name: HY-3-157-CHIRAL

Acq. Operator : Seq. Line : 7 Acq. Instrument : Instrument 1 Location : Vial 52 Injection Date : 7/14/2018 12:01:38 AM Inj: 1 Inj Volume : 2.000 μl : D:\DATA\GUAN YUQING\LJ-3-164\LJ-3-164 2018-07-13 17-50-28\VWD-AD(1-2)-90-10 Acq. Method -1ML-2UL-254NM-60MIN.M Last changed : 7/13/2018 10:23:58 AM Analysis Method : D:\METHOD\CZY\VWD-AD(1-2)-97-3-1ML-3UL-210NM-10MIN.M Last changed : 7/18/2018 11:29:30 AM (modified after loading) Additional Info : Peak(s) manually integrated VWD1 A, Wavelength=254 nm (D:\DATA\GUAN YUQING\LJ-3-164\LJ-3-164 2018-07-13 17-50-28\052-0701.D) mAU Ο  $\cap$ 140 OMe (chiral)-120 31.411 CI 3 100 80 60 40 26.860 20 0 15 20 25 30 35 40 mir Area Percent Report ------Sorted By : Signal Multiplier 1.0000 : : 1.0000 Dilution Use Multiplier & Dilution Factor with ISTDs Signal 1: VWD1 A, Wavelength=254 nm Height Peak RetTime Type Width Area Area # [min] [min] [mAU\*s] [mAU] % 1 26.860 BB 0.6374 439.71262 10.41835 7.0320 2 31.411 BB 0.8134 5813.31201 108.62247 92.9680 Totals : 6253.02463 119.04082

Instrument 1 7/18/2018 11:29:40 AM