

**Chiral isoxazolidine-mediated stereoselective umpolung
 α -phenylation of methyl ketones**

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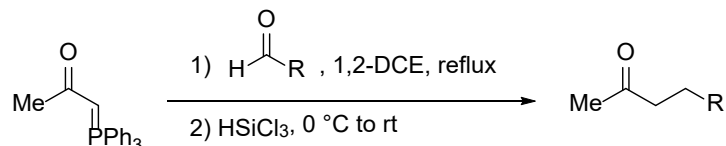
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I. General Information

All reactions were carried out under an argon atmosphere with dry solvents under anhydrous conditions, unless otherwise noted. Reagents were purchased at the highest commercial quality and used without further purification, unless otherwise stated. Flash column chromatography were performed using Silicycle silica gel (SiliaFlash® F60, 40-63 μm). Preparative thin-layer chromatography (preparative TLC) separations were carried out on 0.25 or 0.50 mm E. Merck silica gel plates (60 F₂₅₄). ¹H NMR and ¹³C NMR spectra were recorded on a Varian Mercury 300 MHz, a Varian VNS AS 500 MHz or a Varian VNS AS 600 MHz operating at 300 MHz/75 MHz, 500 MHz/125 MHz, or 600 MHz/150 MHz for ¹H and ¹³C acquisitions, respectively. Chemical shifts are reported in ppm with the solvent resonance or TMS as the internal standard. Multiplicities are indicated by s (singlet), d (doublet), t (triplet), q (quartet), qn (quintet), m (multiplet) and br (broad). Infrared (IR) spectra were recorded on a Perkin-Elmer SpectrumOne A spectrometer using NaCl plates. High-resolution mass spectra (HRMS) were obtained by ESI method on Thermo Fisher Scientific Exactive Instrument. Melting points (uncorrected) were determined on BÜCHI M-565 apparatus. HPLC analyses were carried out on a SHIMADZU LC-20AT pump and a SPD-20A UV/Vis detector or JASCO PU-4180 RHPLC pump and UV-4075 UV/Vis detector. Optical rotations were measured on a JASCO DIP-370 digital polarimeter. 4-Phenyl-2-butanone (**1a**), 4-(4-methoxyphenyl)-2-butanone (**1c**), 4-(1,3-benzodioxol-5-yl)-2-butanone (**1i**), 1-(4-methylphenyl)-2-propanone (**1o**), 5-hexen-2-one (**1p**), 5-oxo-hexanenitrile (**1q**), nabumethone, and pentoxifylline were purchased from Aldrich or TCI (Tokyo Chemical Industry Co., Ltd.). Triphenylaluminium (1.0 M in *n*-dibutyl ether) was purchased from Aldrich. (+)-Benzopyranoisoxazolidine **3** was prepared by the reported procedure.¹ The racemic materials **2a-2s** were prepared using isoxazolidine² or *rac*-benzopyranoisoxazolidine (*rac*-**3**) in the same procedure as their corresponding chiral compounds.

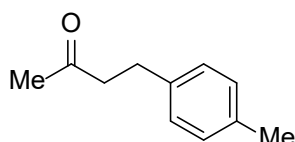
II. Experimental Section

General procedure for the preparation of 4-aryl-2-butanone (**1**)³



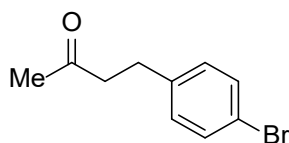
To a solution of (acetylmethylene)triphenylphosphorane (3.18 g, 10 mmol) in 1,2-dichloroethane (20 mL) was added aldehydes (10 mmol) at room temperature under an argon atmosphere. After being stirred at reflux for 3-7 h, the reaction mixture was cooled to $0\text{ }^\circ\text{C}$. Then trichlorosilane (2.0 mL, 20 mmol) was added at $0\text{ }^\circ\text{C}$ in one portion. After being stirred at room temperature for 4 h, the reaction mixture was concentrated under reduced pressure. The residue was purified by flash column chromatography (*n*hexane/AcOEt = 10/1 to 1/1) to give 4-aryl-2-butanone **1**.

4-(4-Methylphenyl)-2-butanone (**1b**)⁴



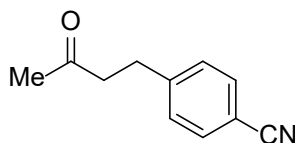
75% yield. a colorless oil; IR (neat) ν_{max} 1717 cm^{-1} ; ^1H NMR (300 MHz, CDCl_3) δ : 7.06 (m, 4H), 2.83 (br t, $J = 7.5\text{ Hz}$, 2H), 2.70 (br t, $J = 7.5\text{ Hz}$, 2H), 2.29 (s, 3H), 2.10 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ : 207.9, 137.8, 135.4, 129.0, 128.0, 45.2, 29.9, 29.2, 20.9; HRMS (ESI) calcd for $\text{C}_{11}\text{H}_{14}\text{ONa}$ [$\text{M}+\text{Na}^+$] 185.0937, found 185.0934.

4-(4-Bromophenyl)-2-butanone (**1d**)⁵



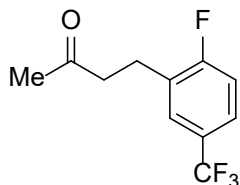
89% yield. white solid; mp $39\text{--}40\text{ }^\circ\text{C}$; IR (CHCl_3) ν_{max} 1716 cm^{-1} ; ^1H NMR (300 MHz, CDCl_3) δ : 7.33 (br d, $J = 8.0\text{ Hz}$, 2H), 7.00 (br d, $J = 8.0\text{ Hz}$, 2H), 2.78 (br t, $J = 7.5\text{ Hz}$, 2H), 2.68 (br t, $J = 7.5\text{ Hz}$, 2H), 2.07 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ : 207.3, 139.9, 131.4, 130.0, 119.7, 44.7, 30.0, 28.9; HRMS (ESI) calcd for $\text{C}_{10}\text{H}_{11}\text{O}^{78}\text{BrNa}$ [$\text{M}+\text{Na}^+$] 248.9885, found 248.9885.

4-(3-Oxobutyl)benzonitrile (**1e**)⁶



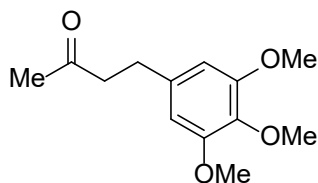
95% yield. white solid; mp 55-56 °C; IR (CHCl₃) ν_{\max} 2230, 1717 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ : 7.57 (br d, J = 8.5 Hz, 2H), 7.30 (br d, J = 8.5 Hz, 2H), 2.96 (br t, J = 7.5 Hz, 2H), 2.79 (br t, J = 7.5 Hz, 2H), 2.16 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ : 206.6, 146.6, 132.0, 129.0, 118.7, 109.6, 43.9, 29.8, 29.3; HRMS (ESI) calcd for C₁₁H₁₁ONNa [M+Na⁺] 196.0733, found 196.0734.

4-[2-Fluoro-(5-trifluoromethyl)phenyl]-2-butanone (1g)



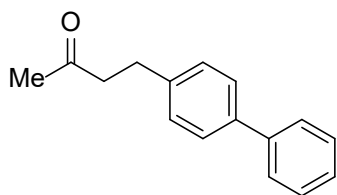
46% yield. a colorless oil; IR (neat) ν_{\max} 1721 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ : 7.52-7.44 (m, 2H), 7.11 (br t, J = 9.0 Hz, 1H), 2.96 (br t, J = 7.5 Hz, 2H), 2.80 (br t, J = 7.5 Hz, 2H), 2.17 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ : 206.8, 162.9 (d, J = 249.0 Hz), 128.9 (d, J = 21.0 Hz), 128.2 (qd, J = 7.0, 3.0 Hz), 126.7 (qd, J = 32.0, 3.5 Hz), 125.5 (qd, J = 7.0, 3.0 Hz), 123.8 (q, J = 270.0 Hz), 115.8 (d, J = 23.0 Hz), 43.1, 29.9, 23.2; HRMS (ESI) calcd for C₁₁H₁₀OF₄Na [M+Na⁺] 257.0560, found 257.0560.

4-(3,4,5-Trimethoxyphenyl)-2-butanone (1h)⁷



47% yield. a colorless oil; IR (neat) ν_{\max} 1717 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ : 6.41 (s, 2H), 3.85 (s, 6H), 3.82 (s, 3H), 2.84 (br td, J = 7.0, 3.0 Hz, 2H), 2.75 (br td, J = 7.0, 3.0 Hz, 2H), 2.16 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ : 207.7, 153.0, 136.7, 136.1, 105.1, 60.6, 55.9 (2C), 45.1, 29.9; HRMS (ESI) calcd for C₁₃H₁₈O₄Na [M+Na⁺] 261.1097, found 261.1096.

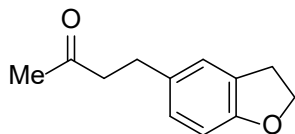
4-[1,1'-Biphenyl]-4-yl-2-butanone (1j)



80% yield. white solid; mp 77-78 °C; IR (CHCl₃) ν_{\max} 1716 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ : 7.57 (br d, J = 7.5 Hz, 2H), 7.51 (br d, J = 7.5 Hz, 2H), 7.43 (br t, J = 7.5 Hz, 2H), 7.32 (br t, J = 7.0 Hz, 1H), 7.26 (br d, J = 7.5 Hz, 2H), 2.94 (br t, J = 7.5 Hz, 2H), 2.80 (br t, J = 7.5 Hz, 2H), 2.16 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ : 207.8, 140.8, 140.0, 139.0, 128.7, 127.1, 127.0, 126.9, 45.0, 30.0, 29.2; HRMS (ESI) calcd for C₁₆H₁₆ONa [M+Na⁺] 247.1093, found 247.1093.

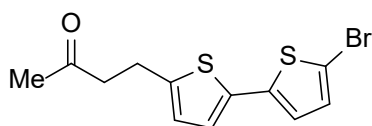
One of aromatic carbons overlapped with other aromatic carbons in ¹³C NMR spectrum.

4-(2,3-Dihydro-5-benzofuranyl)-2-butanone (1l)



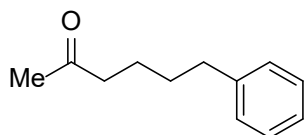
55% yield. white solid; mp 54-55 °C; IR (CHCl₃) ν_{\max} 1715 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ : 7.01 (br s, 1H), 6.90 (br d, J = 8.0 Hz, 1H), 6.69 (br d, J = 8.0 Hz, 1H), 4.53 (t, J = 8.5 Hz, 2H), 3.16 (t, J = 8.5 Hz, 2H), 2.82 (br t, J = 7.5 Hz, 2H), 2.71 (br t, J = 7.5 Hz, 2H), 2.13 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ : 208.1, 158.3, 132.8, 127.5, 127.0, 124.7, 108.9, 71.0, 45.6, 30.0, 29.6, 29.1; HRMS (ESI) calcd for C₁₂H₁₄O₂Na [M+Na⁺] 213.0886, found 213.0886.

4-[5'-Bromo-(2,2'-bithiophene)-5-yl]-2-butanone (1m)



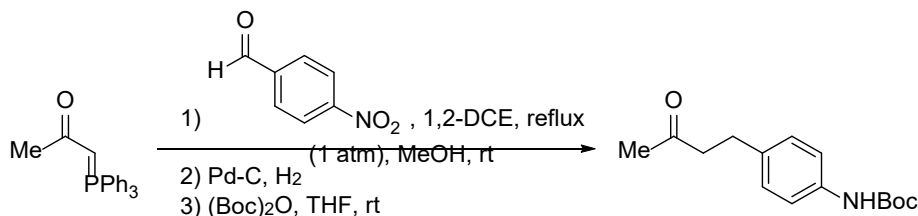
91% yield. white solid; mp 80-81 °C (decomposed); IR (CHCl₃) ν_{\max} 1719 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ : 6.93 (dd, J = 3.5, 1.0 Hz, 1H), 6.89 (d, J = 3.5 Hz, 1H), 6.82 (d, J = 3.5 Hz, 1H), 6.68 (br dd, J = 3.5, 1.0 Hz, 1H), 3.06 (br t, J = 7.5 Hz, 2H), 2.81 (br t, J = 7.5 Hz, 2H), 2.17 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ : 206.9, 143.5, 139.0, 134.3, 130.5, 125.4, 123.7, 123.2, 110.3, 44.8, 30.0, 23.9; HRMS (ESI) calcd for C₁₂H₁₁O⁷⁹BrS₂Na [M+Na⁺] 336.9327, found 336.9327.

6-Phenyl-2-hexanone (1n)⁸



94% yield. a colorless oil; IR (neat) ν_{\max} 1715 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ : 7.26-7.21 (m, 2H), 7.15-7.12 (m, 3H), 2.58 (br t, J = 7.5 Hz, 2H), 2.38 (br t, J = 7.5 Hz, 2H), 2.05 (s, 3H), 1.59-1.56 (m, 4H); ¹³C NMR (75 MHz, CDCl₃) δ : 208.7, 141.9, 128.13, 128.06, 125.5, 43.2, 35.5, 30.7, 29.6, 23.2; HRMS (APCI) calcd for C₁₂H₁₇O [M+H⁺] 177.1274, found 177.1274.

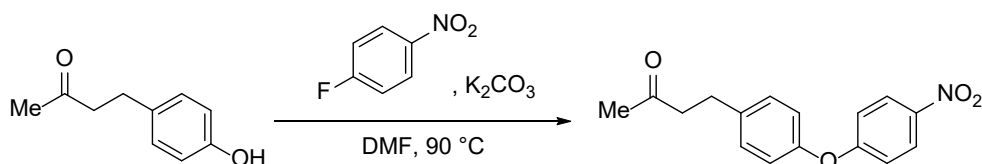
N-[4-(3-Oxobutyl)phenyl]carbamic acid 1,1-dimethylethyl ester (1f)



To a solution of (acetylmethylene)triphenylphosphorane (3.18 g, 10 mmol) in 1,2-dichloroethane (20 mL) was added 4-nitrobenzaldehyde (1.51 g, 10 mmol) at room temperature under an argon atmosphere. After being stirred at reflux for 7 h, the reaction mixture was concentrated under

reduced pressure. The crude product was dissolved in Et₂O (100 mL), then mixture was cooled to 0 °C. A pale yellow precipitate was removed by filtration. The filtrate was concentrated under reduced pressure. The residue was purified by flash column chromatography (*n*hexane/AcOEt = 5/1 to 1/1) to give α,β -unsaturated methyl ketone. To a suspension of α,β -unsaturated methyl ketone (1.74 g, 9.1 mmol) in MeOH (30 mL) and AcOEt (10 mL) was added Pd-C (10% wt, 100 mg) at room temperature under a hydrogen atmosphere (1 atm). After being stirred at the same temperature for overnight, the resulting mixture was filtrated through a silica gel pad and washed with AcOEt. The filtrate was concentrated under reduced pressure. The crude product was used to next reaction without further purification. To a solution of 4-(4-aminophenyl)-2-butanone (1.28 g, 7.3 mmol) in THF (4.5 mL) was added Boc₂O (2.0 g, 9.2 mmol) at room temperature. After being at the same temperature for 7 h, the reaction mixture was concentrated under reduced pressure. The residue was purified by flash column chromatography (*n*hexane/AcOEt = 10/1 to 5/1) to give 4-aryl-2-butanone **1f** (1.38 g, 48%, 3 steps) as white solid. mp 95-96 °C; IR (CHCl₃) ν_{\max} 3385, 1712 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ : 7.28-7.25 (m, 2H), 7.09 (d, *J* = 8.5 Hz, 2H), 6.49 (br s, 1H), 2.84 (t, *J* = 7.5 Hz, 2H), 2.72 (t, *J* = 7.5 Hz, 2H), 2.12 (s, 3H), 1.51 (s, 9H); ¹³C NMR (75 MHz, CDCl₃) δ : 208.1, 152.8, 136.4, 135.4, 128.6, 118.7, 80.1, 45.1, 30.0, 28.9, 28.2; HRMS (ESI) calcd for C₁₅H₂₁O₃NNa [M+Na⁺] 286.1414, found 286.1408.

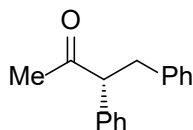
4-[4-(4-Nitrophenoxy)phenyl]-2-butanone (**1k**)



To a solution of 4-(4-hydroxyphenyl)-2-butanone (1.64 g, 10 mmol) in DMF (20 mL) was added K₂CO₃ (2.76 g, 20 mmol) and 1-fluoro-4-nitrobenzene (1.06 mL, 10 mmol) at room temperature under an argon atmosphere. After being stirred at 90 °C for 7 h, the reaction mixture was diluted with Et₂O. The resulting mixture was washed with H₂O (4 times) and saturated NaCl. The organic phase was dried over MgSO₄ and concentrated under reduced pressure. The residue was purified by flash column chromatography (*n*hexane/AcOEt = 5/1 to 1/1) to give 4-aryl-2-butanone **1k** (1.85 g, 65%) as yellow solid. mp 58-59 °C; IR (CHCl₃) ν_{\max} 1715, 1515, 1344 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ : 8.18 (br d, *J* = 8.5 Hz, 2H), 7.25 (br d, *J* = 8.5 Hz, 2H), 7.02-6.98 (m, 4H), 2.93 (t, *J* = 7.0 Hz, 2H), 2.80 (t, *J* = 7.0 Hz, 2H), 2.18 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ : 207.4, 163.4, 152.7, 142.3, 138.2, 130.0, 125.7, 120.4, 116.7, 44.8, 29.9, 28.8; HRMS (ESI) calcd for C₁₆H₁₅O₄NNa [M+Na⁺] 308.0893, found 308.0892.

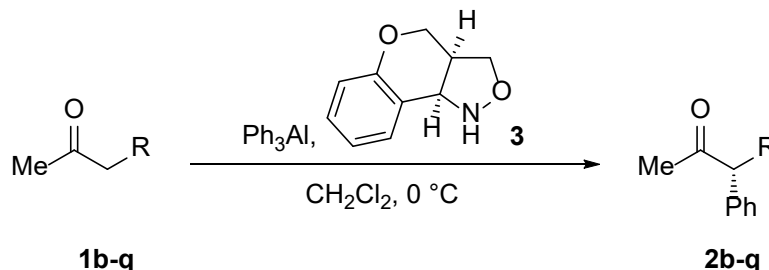
Asymmetric nucleophilic α -phenylation of ketone **1a** (Table 1, entry 2)

(3*R*)-3,4-Diphenyl-2-butanone (**2a**)⁹



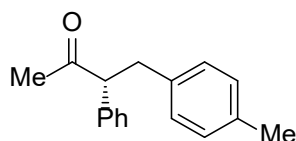
To a solution of (+)-benzopyranoisoxazolidine **3** (26.6 mg, 0.15 mmol) in CH_2Cl_2 (0.75 mL) were added 4-phenyl-2-butanone (**1a**) (11.1 mg, 0.075 mmol) and triphenylaluminium (1.0 M in *n*dibutyl ether, 0.23 mL, 0.23 mmol) dropwise at 0 °C under an argon atmosphere. After being stirred at the same temperature for 2 h, the reaction mixture was quenched with an aqueous Rochelle's salt (1.3 M). The resulting suspension was extracted with CHCl_3 . The organic phase was dried over MgSO_4 and concentrated under reduced pressure. The residue was purified by preparative TLC (*n*hexane/AcOEt = 5/1) to give α -phenylated ketone **2a** (11.1 mg, 66%, 94% ee) as a colorless oil. $[\alpha]_{\text{D}}^{26} -221.0$ (*c* 0.56, CHCl_3); The enantiomeric purity was determined by HPLC analysis [CHIRALPAK IC (0.46 cm x 25 cm, from Daicel Chemical Ind. Ltd.) *n*hexane/*i*PrOH = 99/1, flow rate = 0.5 mL/min, $\lambda = 215$ nm, temperature: 25 °C, retention times: 16.2 min (minor), 17.6 min (major)]. IR (neat) ν_{max} 1714 cm^{-1} ; ^1H NMR (300 MHz, CDCl_3) δ : 7.34-7.26 (m, 2H), 7.23-7.15 (m, 6H), 7.04 (br d, $J = 7.0$ Hz, 2H), 3.92 (br t, $J = 7.5$ Hz, 1H), 3.43 (dd, $J = 13.5, 7.5$ Hz, 1H), 2.90 (dd, $J = 13.5, 7.5$ Hz, 1H), 2.03 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ : 207.3, 139.5, 138.3, 128.8, 128.7, 128.2, 128.1, 127.2, 125.9, 61.5, 38.4, 29.5; HRMS (ESI) calcd for $\text{C}_{16}\text{H}_{16}\text{ONa}$ [$\text{M}+\text{Na}^+$] 247.1093, found 247.1095.

General procedure for asymmetric nucleophilic α -phenylation of ketones **1b-q** (Table 2)



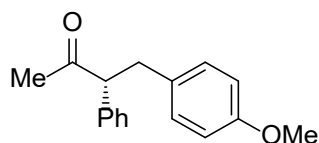
To a solution of (+)-benzopyranoisoxazolidine **3** (26.6 mg, 0.15 mmol) in CH_2Cl_2 (0.75 mL) were added ketone **1b-q** (0.075 mmol) and triphenylaluminium (1.0 M in *n*dibutyl ether, 0.23 mL, 0.23 mmol) dropwise at 0 °C under an argon atmosphere. After being stirred at the same temperature for 2-3 h, the reaction mixture was quenched with an aqueous Rochelle's salt (1.3 M). The resulting suspension was extracted with CHCl_3 . The organic phase was dried over MgSO_4 and concentrated under reduced pressure. The residue was purified by preparative TLC (*n*hexane/AcOEt = 5/1) to give α -phenylated ketone **2b-q**.

(3R)-4-(4-Methylphenyl)-3-phenyl-2-butanone (2b)



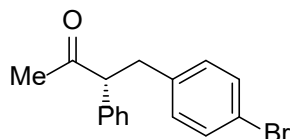
61% yield, 90% ee. a colorless oil. $[\alpha]_{\text{D}}^{25} -307$ (c 0.30, CHCl_3); The enantiomeric purity was determined by HPLC analysis [CHIRALPAK IC (0.46 cm x 25 cm, from Daicel Chemical Ind. Ltd.) n hexane/ i PrOH = 99/1, flow rate = 0.5 mL/min, λ = 215 nm, temperature: 25 °C, retention times: 26.1 min (minor), 29.9 min (major)]. IR (neat) ν_{max} 1715 cm^{-1} ; ^1H NMR (300 MHz, CDCl_3) δ : 7.33-7.25 (m, 3H), 7.20-7.17 (m, 2H), 7.00 (br d, J = 8.0 Hz, 2H), 6.93 (br d, J = 8.0 Hz, 2H), 3.90 (br t, J = 7.5 Hz, 1H), 3.38 (dd, J = 13.5, 7.5 Hz, 1H), 2.86 (dd, J = 13.5, 7.5 Hz, 1H), 2.27 (s, 3H), 2.01 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ : 207.5, 138.5, 136.5, 135.4, 128.83, 128.77, 128.72, 128.2, 127.2, 61.6, 38.0, 29.6, 21.1; HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{18}\text{ONa}$ $[\text{M}+\text{Na}^+]$ 261.1250, found 261.1250.

(3R)-4-(4-Methoxyphenyl)-3-phenyl-2-butanone (2c)



61% yield, 94% ee. white solid. mp 55-56 °C; $[\alpha]_{\text{D}}^{25} -262$ (c 0.39, CHCl_3); The enantiomeric purity was determined by HPLC analysis [CHIRALPAK IA (0.46 cm x 25 cm, from Daicel Chemical Ind. Ltd.) n hexane/ i PrOH = 99/1, flow rate = 1.0 mL/min, λ = 215 nm, temperature: 25 °C, retention times: 10.7 min (major), 12.2 min (minor)]. IR (CHCl_3) ν_{max} 1712 cm^{-1} ; ^1H NMR (300 MHz, CDCl_3) δ : 7.34-7.25 (m, 3H), 7.19-7.16 (m, 2H), 6.96 (br d, J = 8.5 Hz, 2H), 6.74 (br d, J = 8.5 Hz, 2H), 3.88 (br t, J = 7.5 Hz, 1H), 3.75 (s, 3H), 3.36 (dd, J = 13.5, 7.5 Hz, 1H), 2.84 (dd, J = 13.5, 7.5 Hz, 1H), 2.02 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ : 207.9, 157.9, 138.5, 131.7, 129.9, 128.9, 128.3, 127.3, 113.6, 61.8, 55.2, 37.5, 29.6; HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{18}\text{O}_2\text{Na}$ $[\text{M}+\text{Na}^+]$ 277.1199, found 277.1199.

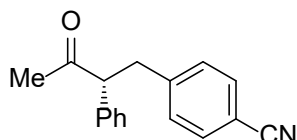
(3R)-4-(4-Bromophenyl)-3-phenyl-2-butanone (2d)



50% yield, 90% ee. white solid. mp 74-75 °C; $[\alpha]_{\text{D}}^{25} -218$ (c 0.33, CHCl_3); The enantiomeric purity was determined by HPLC analysis [CHIRALPAK IC (0.46 cm x 25 cm, from Daicel Chemical Ind. Ltd.) n hexane/ i PrOH = 99/1, flow rate = 0.5 mL/min, λ = 215 nm, temperature: 25 °C,

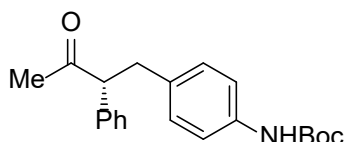
retention times: 18.2 min (minor), 19.2 min (major)]. IR (CHCl₃) ν_{\max} 1713 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ : 7.34-7.28 (m, 5H), 7.16-7.13 (m, 2H), 6.89 (br d, J = 8.5 Hz, 2H), 3.85 (br t, J = 7.5 Hz, 1H), 3.36 (dd, J = 13.5, 7.5 Hz, 1H), 2.84 (dd, J = 13.5, 7.5 Hz, 1H), 2.03 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ : 207.0, 138.5, 137.9, 131.2, 130.7, 128.9, 128.2, 127.4, 119.9, 61.4, 37.8, 29.5; HRMS (ESI) calcd for C₁₆H₁₅O⁷⁹BrNa [M+Na⁺] 325.0199, found 325.0196.

(2R)-4-(3-Oxo-2-phenylbutyl)benzonitrile (2e)



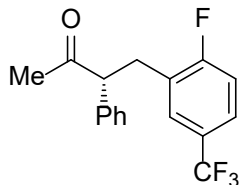
67% yield, 89% ee. white solid. mp 110-111 °C; [α]_D²⁵ -317 (c 0.55, CHCl₃); The enantiomeric purity was determined by HPLC analysis [CHIRALPAK IB (0.46 cm x 25 cm, from Daicel Chemical Ind. Ltd.) *n*hexane/*i*PrOH = 95/5, flow rate = 0.5 mL/min, λ = 215 nm, temperature: 25 °C, retention times: 21.5 min (minor), 24.7 min (major)]. IR (CHCl₃) ν_{\max} 2230, 1712 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ : 7.45 (br d, J = 8.0 Hz, 2H), 7.30-7.24 (m, 3H), 7.11-7.08 (m, 4H), 3.85 (br t, J = 7.5 Hz, 1H), 3.44 (dd, J = 13.5, 7.5 Hz, 1H), 2.94 (dd, J = 13.5, 7.5 Hz, 1H), 2.03 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ : 206.4, 145.3, 137.5, 131.9, 129.7, 129.0, 128.2, 127.6, 118.8, 110.0, 61.0, 38.4, 29.3; HRMS (ESI) calcd for C₁₇H₁₅ONNa [M+Na⁺] 272.1046, found 272.1046.

(2R)-N-[4-(3-Oxo-2-phenylbutyl)phenyl]carbamic acid 1,1-dimethylethyl ester (2f)



58% yield, 91% ee. white solid. mp 102-103 °C; [α]_D²⁵ -200 (c 0.74, CHCl₃); The enantiomeric purity was determined by HPLC analysis [CHIRALPAK IC (0.46 cm x 25 cm, from Daicel Chemical Ind. Ltd.) *n*hexane/*i*PrOH = 80/20, flow rate = 0.5 mL/min, λ = 215 nm, temperature: 25 °C, retention times: 15.3 min (minor), 16.8 min (major)]. IR (CHCl₃) ν_{\max} 3440, 1714 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ : 7.33-7.26 (m, 3H), 7.19-7.15 (m, 4H), 6.95 (d, J = 8.5 Hz, 2H), 6.38 (br s, 1H), 3.87 (br t, J = 7.5 Hz, 1H), 3.36 (dd, J = 13.5, 7.5 Hz, 1H), 2.84 (dd, J = 13.5, 7.5 Hz, 1H), 2.01 (s, 3H), 1.50 (s, 9H); ¹³C NMR (75 MHz, CDCl₃) δ : 207.8, 152.8, 138.3, 136.4, 134.2, 129.4, 128.8, 128.3, 127.3, 118.4, 80.3, 61.6, 37.6, 29.5, 28.3; HRMS (ESI) calcd for C₂₁H₂₅O₃NNa [M+Na⁺] 362.1727, found 362.1727.

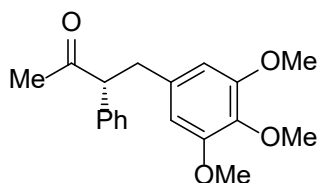
(3R)-4-[2-Fluoro-(5-trifluoromethyl)phenyl]-3-phenyl-2-butanone (2g)



61% yield, 95% ee. a colorless oil. $[\alpha]_D^{25} -222$ (c 0.45, CHCl_3); The enantiomeric purity was determined by HPLC analysis [CHIRALPAK IB (0.46 cm x 25 cm, from Daicel Chemical Ind. Ltd.) n hexane/ i PrOH = 100/1, flow rate = 0.5 mL/min, λ = 215 nm, temperature: 25 °C, retention times: 26.5 min (major), 29.9 min (minor)]. IR (CHCl_3) ν_{max} 1717 cm^{-1} ; ^1H NMR (300 MHz, CDCl_3) δ : 7.42-7.37 (m, 1H), 7.33-7.25 (m, 3H), 7.21-7.19 (m, 1H), 7.13-7.10 (m, 2H), 7.05 (t, J = 9.0 Hz, 1H), 3.94 (br t, J = 7.5 Hz, 1H), 3.44 (br dd, J = 13.5, 7.5 Hz, 1H), 2.97 (dd, J = 13.5, 7.5 Hz, 1H), 2.05 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3) δ : 206.8, 162.9 (d, J = 249.0 Hz), 137.6, 129.1 (m), 128.2, 127.7, 127.5 (d, J = 16.5 Hz), 126.3 (qd, J = 32.0, 3.5 Hz), 125.4 (qd, J = 7.0, 3.0 Hz), 123.7 (q, J = 270.0 Hz), 115.6 (d, J = 23.0 Hz), 59.4, 31.6, 29.2; HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{14}\text{OF}_4\text{Na}$ [$\text{M}+\text{Na}^+$] 333.0873, found 333.0870.

One of aromatic carbons overlapped with other aromatic carbons in ^{13}C NMR spectrum.

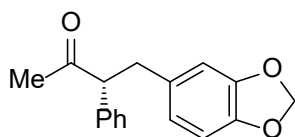
(3R)-4-(3,4,5-Trimethoxyphenyl)-3-phenyl-2-butanone (2h)



54% yield, 93% ee. white solid. mp 88-89 °C; $[\alpha]_D^{25} -182$ (c 0.62, CHCl_3); The enantiomeric purity was determined by HPLC analysis [CHIRALPAK IA (0.46 cm x 25 cm, from Daicel Chemical Ind. Ltd.) n hexane/ i PrOH = 95/5, flow rate = 0.5 mL/min, λ = 215 nm, temperature: 25 °C, retention times: 23.9 min (major), 26.9 min (minor)]. IR (CHCl_3) ν_{max} 1712 cm^{-1} ; ^1H NMR (300 MHz, CDCl_3) δ : 7.33-7.21 (m, 3H), 7.16-7.14 (m, 2H), 6.17 (s, 2H), 3.85 (br t, J = 7.5 Hz, 1H), 3.78 (s, 3H), 3.71 (s, 6H), 3.33 (dd, J = 13.5, 7.5 Hz, 1H), 2.84 (dd, J = 13.5, 7.5 Hz, 1H), 2.05 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ : 207.3, 152.7, 138.4, 135.2, 128.8, 128.3, 127.3, 106.0, 61.6, 60.8, 56.0, 38.7, 29.6; HRMS (ESI) calcd for $\text{C}_{19}\text{H}_{22}\text{O}_4\text{Na}$ [$\text{M}+\text{Na}^+$] 337.1410, found 337.1408.

One of aromatic carbons overlapped with other aromatic carbons in ^{13}C NMR spectrum.

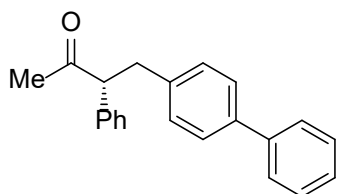
(3R)-4-(1,3-Benzodioxol-5-yl)-3-phenyl-2-butanone (2i)



58% yield, 93% ee. a colorless oil. $[\alpha]_D^{25} -225$ (c 0.44, CHCl_3); The enantiomeric purity was determined by HPLC analysis [CHIRALPAK IA (0.46 cm x 25 cm, from Daicel Chemical Ind. Ltd.)

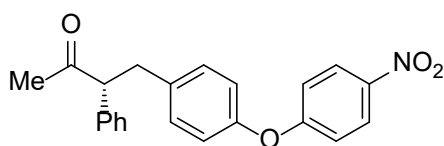
*n*hexane/*i*PrOH = 99/1, flow rate = 1.0 mL/min, λ = 215 nm, temperature: 25 °C, retention times: 12.5 min (minor), 14.2 min (major)]. IR (neat) ν_{\max} 1712 cm^{-1} ; ^1H NMR (300 MHz, CDCl_3) δ : 7.32-7.20 (m, 3H), 7.16-7.14 (m, 2H), 6.62 (d, J = 7.5 Hz, 1H), 6.52 (d, J = 1.5 Hz, 1H), 6.47 (dd, J = 7.5, 1.5 Hz, 1H), 5.86 (s, 2H), 3.85 (br t, J = 7.5 Hz, 1H), 3.33 (dd, J = 13.5, 7.5 Hz, 1H), 2.80 (dd, J = 13.5, 7.5 Hz, 1H), 2.02 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ : 207.7, 147.4, 145.8, 138.3, 133.4, 128.9, 128.3, 127.4, 121.9, 109.4, 108.0, 100.7, 61.7, 38.0, 29.6; HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{16}\text{O}_3\text{Na}$ [$\text{M}+\text{Na}^+$] 291.0992, found 291.0996.

(3R)-4-[1,1'-Biphenyl]-4-yl-3-phenyl-2-butanone (2j)



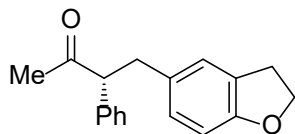
72% yield, 92% ee. white solid. mp 96-97 °C; $[\alpha]_{\text{D}}^{25}$ -259 (c 0.83, CHCl_3); The enantiomeric purity was determined by HPLC analysis [CHIRALPAK ID (0.46 cm x 25 cm, from Daicel Chemical Ind. Ltd.) *n*hexane/*i*PrOH = 99/1, flow rate = 0.5 mL/min, λ = 215 nm, temperature: 25 °C, retention times: 13.4 min (minor), 15.0 min (major)]. IR (CHCl_3) ν_{\max} 1712 cm^{-1} ; ^1H NMR (300 MHz, CDCl_3) δ : 7.55 (br d, J = 7.5 Hz, 2H), 7.46-7.19 (m, 10H), 7.12 (d, J = 8.0 Hz, 2H), 3.96 (br t, J = 7.5 Hz, 1H), 3.47 (dd, J = 13.5, 7.5 Hz, 1H), 2.94 (dd, J = 13.5, 7.5 Hz, 1H), 2.05 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ : 207.3, 140.7, 138.8, 138.7, 138.3, 129.3, 128.8, 128.6, 128.2, 127.3, 126.9, 126.81, 126.80, 61.5, 38.0, 29.6; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{20}\text{ONa}$ [$\text{M}+\text{Na}^+$] 323.1406, found 323.1406.

(3R)-4-[4-(4-Nitrophenoxy)phenyl]-3-phenyl-2-butanone (2k)



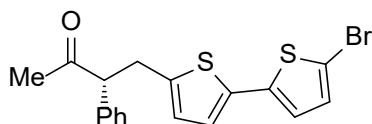
61% yield, 92% ee. pale yellow solid. mp 72-73 °C; $[\alpha]_{\text{D}}^{25}$ -226 (c 0.82, CHCl_3); The enantiomeric purity was determined by HPLC analysis [CHIRALPAK IB (0.46 cm x 25 cm, from Daicel Chemical Ind. Ltd.) *n*hexane/*i*PrOH = 95/5, flow rate = 0.5 mL/min, λ = 215 nm, temperature: 25 °C, retention times: 23.4 min (minor), 24.8 min (major)]. IR (CHCl_3) ν_{\max} 1713, 1516, 1344 cm^{-1} ; ^1H NMR (300 MHz, CDCl_3) δ : 8.15 (br d, J = 9.0 Hz, 2H), 7.33-7.24 (m, 3H), 7.15 (br d, J = 8.0 Hz, 2H), 7.07 (d, J = 8.0 Hz, 2H), 6.94-6.88 (m, 4H), 3.90 (br t, J = 7.5 Hz, 1H), 3.43 (dd, J = 13.5, 7.5 Hz, 1H), 2.92 (dd, J = 13.5, 7.5 Hz, 1H), 2.05 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ : 207.1, 163.3, 152.7, 142.4, 138.0, 136.9, 130.7, 128.9, 128.2, 127.4, 125.7, 120.2, 116.8, 61.5, 37.6, 29.5; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{19}\text{O}_4\text{NNa}$ [$\text{M}+\text{Na}^+$] 384.1206, found 384.1208.

(3R)-4-(2,3-Dihydro-5-benzofuranyl)-3-phenyl-2-butanone (2l)



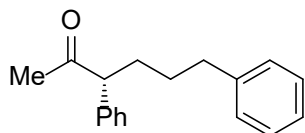
51% yield, 80% ee. a colorless oil. $[\alpha]_D^{25} -65$ (c 0.17, CHCl_3); The enantiomeric purity was determined by HPLC analysis [CHIRALPAK IC (0.46 cm x 25 cm, from Daicel Chemical Ind. Ltd.) *n*hexane/THF = 90/10, flow rate = 0.3 mL/min, λ = 215 nm, temperature: 25 °C, retention times: 21.4 min (minor), 22.4 min (major)]. IR (neat) ν_{max} 1712 cm^{-1} ; ^1H NMR (300 MHz, CDCl_3) δ : 7.34-7.26 (m, 3H), 7.19 (br d, J = 7.5 Hz, 2H), 6.89 (br s, 1H), 6.78 (br d, J = 8.0 Hz, 1H), 6.62 (br d, J = 8.0 Hz, 1H), 4.51 (t, J = 8.5 Hz, 2H), 3.88 (br t, J = 7.5 Hz, 1H), 3.35 (dd, J = 13.5, 7.5 Hz, 1H), 3.12 (t, J = 8.5 Hz, 2H), 2.81 (dd, J = 13.5, 7.5 Hz, 1H), 2.02 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ : 208.0, 158.4, 138.6, 131.6, 128.9, 128.5, 128.33, 128.31, 127.3, 125.5, 108.8, 71.1, 62.1, 62.0, 37.8, 29.7; HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{18}\text{O}_2\text{Na}$ $[\text{M}+\text{Na}^+]$ 289.1199, found 289.1202.

(3R)-4-[5'-Bromo-(2,2'-bithiophene)-5-yl]-3-phenyl-2-butanone (2m)



57% yield, 93% ee. white solid. mp 98-99 °C; $[\alpha]_D^{25} -202$ (c 0.50, CHCl_3); The enantiomeric purity was determined by HPLC analysis [CHIRALPAK IC (0.46 cm x 25 cm, from Daicel Chemical Ind. Ltd.) *n*hexane/*i*PrOH = 99/1, flow rate = 0.5 mL/min, λ = 215 nm, temperature: 25 °C, retention times: 24.1 min (minor), 29.1 min (major)]. IR (CHCl_3) ν_{max} 1713 cm^{-1} ; ^1H NMR (300 MHz, CDCl_3) δ : 7.36-7.20 (m, 5H), 6.92 (dd, J = 3.5, 1.0 Hz, 1H), 6.83 (br d, J = 3.5 Hz, 1H), 6.79 (br d, J = 3.5 Hz, 1H), 6.54 (br dd, J = 3.5, 1.0 Hz, 1H), 3.93 (br t, J = 7.5 Hz, 1H), 3.59 (dd, J = 13.5, 7.5 Hz, 1H), 3.07 (dd, J = 13.5, 7.5 Hz, 1H), 2.07 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ : 206.7, 141.8, 139.0, 137.6, 134.5, 130.4, 129.0, 128.2, 127.6, 126.3, 123.5, 123.1, 110.3, 61.4, 32.7, 29.4; HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{15}\text{O}^{79}\text{BrS}_2\text{Na}$ $[\text{M}+\text{Na}^+]$ 412.9640, found 412.9647.

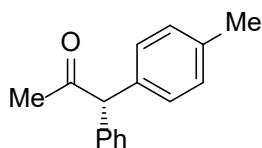
(3R)-3,6-diphenyl-2-hexanone (2n)



61% yield, 80% ee. a colorless oil. $[\alpha]_D^{25} -122$ (c 0.45, CHCl_3); The enantiomeric purity was determined by HPLC analysis [CHIRALPAK IC (0.46 cm x 25 cm, from Daicel Chemical Ind. Ltd.) *n*hexane/*i*PrOH = 99/1, flow rate = 0.5 mL/min, λ = 215 nm, temperature: 25 °C, retention times: 19.4 min (minor), 23.2 min (major)]. IR (neat) ν_{max} 1712 cm^{-1} ; ^1H NMR (300 MHz, CDCl_3) δ : 7.35-7.11 (m, 10H), 3.60 (br t, J = 7.5 Hz, 1H), 2.68-2.51 (m, 2H), 2.14-2.03 (m, 1H), 2.03, (s, 3H), 1.81-1.69 (m, 1H), 1.62-1.43 (m, 2H); ^{13}C NMR (75 MHz, CDCl_3) δ : 208.4, 142.1, 138.8, 128.9,

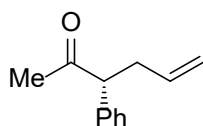
128.34, 128.26, 128.22, 127.2, 125.7, 59.6, 35.8, 31.4, 29.2, 29.0; HRMS (ESI) calcd for C₁₈H₂₀ONa [M+Na⁺] 275.1406, found 275.1405.

1-(4-Methylphenyl)-1-phenyl-2-propanone (2o)¹⁰



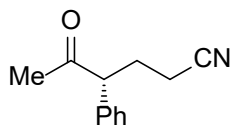
51% yield, 5% ee. a colorless oil. The enantiomeric purity was determined by HPLC analysis [CHIRALPAK IA (0.46 cm x 25 cm, from Daicel Chemical Ind. Ltd.) *n*hexane/*i*PrOH = 99/1, flow rate = 0.5 mL/min, λ = 215 nm, temperature: 25 °C, retention times: 20.1 min (major), 21.8 min (minor)]. IR (neat) ν_{\max} 1714 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ : 7.36-7.19 (m, 6H), 7.16-7.09 (m, 3H), 5.08 (s, 1H), 2.32 (s, 3H), 2.33 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ : 206.7, 138.5, 136.9, 135.2, 129.4, 128.9, 128.8, 128.7, 127.1, 64.7, 30.0, 21.0; HRMS (ESI) calcd for C₁₆H₁₆ONa [M+Na⁺] 247.1093, found 247.1092.

(3R)-3-Phenyl-5-hexen-2-one (2p)



54% yield, 85% ee. a colorless oil. [α]_D²⁵ -214 (*c* 0.35, CHCl₃); The enantiomeric purity was determined by HPLC analysis [CHIRALPAK IC (0.46 cm x 25 cm, from Daicel Chemical Ind. Ltd.) *n*hexane/*i*PrOH = 99/1, flow rate = 0.5 mL/min, λ = 215 nm, temperature: 25 °C, retention times: 13.0 min (minor), 14.0 min (major)]. IR (neat) ν_{\max} 1715 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ : 7.37-7.26 (m, 3H), 7.23-7.20 (m, 2H), 5.67 (ddt, *J* = 17.0, 10.0, 6.5 Hz, 1H), 4.99 (br d, *J* = 17.0 Hz, 1H), 4.95 (br d, *J* = 10.0 Hz, 1H), 3.70 (br t, *J* = 7.5 Hz, 1H), 2.85-2.75 (m, 1H), 2.48-2.38 (m, 1H), 2.07 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ : 207.5, 138.3, 135.7, 128.9, 128.3, 127.4, 116.6, 59.4, 36.1, 29.1; HRMS (ESI) calcd for C₁₂H₁₄ONa [M+Na⁺] 197.0937, found 197.0934.

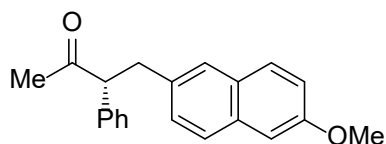
(4R)-5-Oxo-4-phenylhexanenitrile (2q)



61% yield, 83% ee. a colorless oil. [α]_D²⁵ -293 (*c* 0.43, CHCl₃); The enantiomeric purity was determined by HPLC analysis [CHIRALPAK IA (0.46 cm x 25 cm, from Daicel Chemical Ind. Ltd.) *n*hexane/*i*PrOH = 99/1, flow rate = 1.0 mL/min, λ = 215 nm, temperature: 25 °C, retention times: 16.2 min (minor), 18.4 min (major)]. IR (neat) ν_{\max} 2247, 1713 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ : 7.41-7.30 (m, 3H), 7.22-7.19 (m, 2H), 3.81 (br dd, *J* = 8.0, 6.5 Hz, 1H), 2.40-2.28 (m, 2H), 2.23-1.94 (m, 5H); ¹³C NMR (75 MHz, CDCl₃) δ : 206.6, 136.8, 129.4, 128.2, 128.1, 119.1, 57.5, 29.1, 27.2,

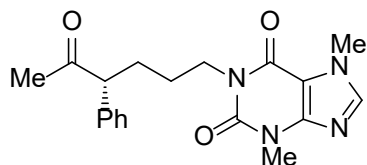
15.0; HRMS (ESI) calcd for C₁₂H₁₃ONNa [M+Na⁺] 210.0889, found 210.0890.

(3R)-4-(6-Methoxy-2-naphthalenyl)-3-phenyl-2-butanone (2r)



According to the general procedure for asymmetric nucleophilic α -phenylation of ketones **2b-q**, the reaction of nabumethone (17.1 mg, 0.075 mmol) with (+)-benzopyranoisoxazolidine **3** (26.6 mg, 0.15 mmol) and triphenylaluminium (1.0 M in *nd*ibutyl ether, 0.23 mL, 0.23 mmol) gave α -phenylated nabumetone **2r** (15.3 mg, 67%, 89% ee) as white solid. mp 85-86 °C; [α]_D²⁵ -195 (*c* 0.82, CHCl₃); The enantiomeric purity was determined by HPLC analysis [CHIRALPAK IC (0.46 cm x 25 cm, from Daicel Chemical Ind. Ltd.) *n*hexane/THF = 90/10, flow rate = 0.5 mL/min, λ = 215 nm, temperature: 25 °C, retention times: 12.1 min (minor), 12.3 min (major)]. IR (CHCl₃) ν_{\max} 1712 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ : 7.60 (br d, *J* = 7.0 Hz, 1H), 7.58 (br d, *J* = 7.0 Hz, 1H), 7.42 (br s, 1H), 7.32-7.10 (m, 7H), 7.07 (br s, 1H), 4.00 (br t, *J* = 7.5 Hz, 1H), 3.89 (s, 3H), 3.55 (dd, *J* = 13.5, 7.5 Hz, 1H), 3.01 (dd, *J* = 13.5, 7.5 Hz, 1H), 2.02 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ : 207.8, 157.2, 138.5, 134.8, 133.0, 129.0, 128.9 (2C), 128.3, 128.0, 127.4, 127.2, 126.6, 118.6, 105.5, 61.6, 55.2, 38.3, 29.6; HRMS (ESI) calcd for C₂₁H₂₀O₂Na [M+Na⁺] 327.1356, found 327.1357.

(R)-3,7-Dihydro-3,7-dimethyl-1-(5-oxo-4-phenylhexyl)-1H-purine-2,6-dione (2s)

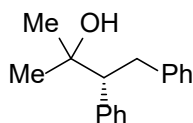


According to the general procedure for asymmetric nucleophilic α -phenylation of ketones **1b-q**, the reaction of pentoxifylline (20.9 mg, 0.075 mmol) with (+)-benzopyranoisoxazolidine **3** (26.6 mg, 0.15 mmol) and triphenylaluminium (1.0 M in *nd*ibutyl ether, 0.23 mL, 0.23 mmol) gave α -phenylated pentoxifylline **2s** (13.3 mg, 50%, 75% ee) as white solid. mp 168-169 °C; [α]_D²⁵ -147 (*c* 0.58, CHCl₃); The enantiomeric purity was determined by HPLC analysis [CHIRALPAK IC (0.46 cm x 25 cm, from Daicel Chemical Ind. Ltd.) *n*hexane/*i*PrOH = 50/50, flow rate = 2.0 mL/min, λ = 215 nm, temperature: 25 °C, retention times: 15.0 min (minor), 19.4 min (major)]. IR (CHCl₃) ν_{\max} 1706, 1658 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ : 7.50 (s, 1H), 7.35-7.20 (m, 5H), 4.00 (br t, *J* = 7.5 Hz, 2H), 3.97 (s, 3H), 3.71 (br t *J* = 7.5 Hz, 1H), 3.55 (s, 3H), 2.14-2.05 (m, 1H), 2.06 (s, 3H), 1.84-1.68 (m, 1H), 1.65-1.14 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ : 208.2, 155.2, 151.3, 148.7, 141.4, 138.7, 128.9, 128.3, 127.2, 107.5, 59.1, 40.8, 33.5, 29.6, 29.1, 28.8, 25.8; HRMS (ESI) calcd for C₁₉H₂₂O₃N₄Na [M+Na⁺] 377.1584, found 377.1585.

Asymmetric nucleophilic α -phenylation of ketone **1a** on 3.0 mmol scale (Scheme 4)

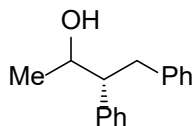
According to the experimental procedure for asymmetric α -phenylation of ketone **1a** (Table 1, entry 2), the reaction of 4-phenyl-2-butanone (**1a**) (444.6 mg, 3.0 mmol) and (+)-benzopyranoisoxazolidine **3** (1.06 g, 6.0 mmol) in CH_2Cl_2 (60 mL) with triphenylaluminium (1.0 M in *n*-dibutyl ether, 9.0 mL, 9.0 mmol) at 0 °C for 2 h gave α -phenylated ketone **2a** (376.8 mg, 56%, 94% ee) as a colorless oil.

(3*R*)-3,4-diphenyl-2-methyl-2-butanol (**4**)



To a solution of **2a** (11.2 mg, 0.050 mmol) in THF (0.20 mL) was added MeMgBr (3.0 M in Et_2O , 36.7 μL , 0.11 mmol) dropwise at 0 °C under an argon atmosphere. After being stirred at the same temperature for 1 h, the reaction mixture was quenched with saturated NH_4Cl . The resulting suspension was extracted with CHCl_3 . The organic phase was dried over MgSO_4 and concentrated under reduced pressure. The residue was purified by preparative TLC (*n*hexane/ AcOEt = 3/1) to give *tert*-alcohol **4** (8.8 mg, 73%, 94% ee) as white solid. mp 68-69 °C; $[\alpha]_{\text{D}}^{25}$ -96 (*c* 0.38, CHCl_3); The enantiomeric purity was determined by HPLC analysis [CHIRALPAK IA (0.46 cm x 25 cm, from Daicel Chemical Ind. Ltd.) *n*hexane/*i*PrOH = 99/1, flow rate = 0.5 mL/min, λ = 254 nm, temperature: 25 °C, retention times: 36.6 min (major), 44.8 min (minor)]. IR (CHCl_3) ν_{max} 3448 cm^{-1} ; ^1H NMR (300 MHz, CDCl_3) δ : 7.23-7.01 (m, 8H), 6.95 (br d, J = 7.5 Hz, 2H), 3.28 (dd, J = 13.0, 3.0 Hz, 1H), 3.02 (br t, J = 13.0 Hz, 1H), 2.90 (dd, J = 13.0, 3.0 Hz, 1H), 1.41 (br s, 1H), 1.27 (s, 3H), 1.25 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ : 141.0, 140.6, 129.6, 128.7, 127.9 (2C), 126.5, 125.4, 73.0, 59.3, 36.2, 28.4, 28.0; HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{20}\text{ONa}$ [$\text{M}+\text{Na}^+$] 263.1406, found 263.1407.

(3*R*)-3,4-Diphenyl-2-butanol (**5**)¹¹



To a solution of **2a** (10.8 mg, 0.044 mmol) in MeOH (0.50 mL) was added NaBH_4 (3.3 mg, 0.088 mmol) at 0 °C under an argon atmosphere. After being stirred at the same temperature for 30 min, the reaction mixture was quenched with saturated NH_4Cl . The resulting mixture was extracted with CHCl_3 . The organic phase was dried over MgSO_4 and concentrated under reduced pressure. The residue was purified by preparative TLC (*n*hexane/ Et_2O = 1/2) to give alcohol **5** (7.4 mg, 90%, 94% ee) as a colorless oil. The diastereomeric ratio (dr = 9:1) was calculated by ^1H NMR analysis of

crude product and the enantiomeric excess was calculated by HPLC analysis. The relative stereochemistry of the major diastereomer has not been established. The minor diastereomer could not be isolated.

(major diastereomer) $[\alpha]_D^{25} -61$ (*c* 0.33, CHCl₃); The enantiomeric purity was determined by HPLC analysis [CHIRALPAK IB (0.46 cm x 25 cm, from Daicel Chemical Ind. Ltd.) *n*hexane/*i*PrOH = 99/1, flow rate = 0.5 mL/min, λ = 254 nm, temperature: 25 °C, retention times: 22.4 min (major), 27.1 min (minor)]. IR (neat) ν_{\max} 3428 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ : 7.29-7.03 (m, 10H), 3.98 (m, 1H), 3.16 (dd, *J* = 13.0, 6.5 Hz, 1H), 2.90 (dd, *J* = 13.0, 8.5 Hz, 1H), 2.85-2.78 (m, 1H), 1.29 (br s, 1H), 1.17 (d, *J* = 6.0 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ : 140.4, 140.2, 129.00, 128.98, 128.2, 128.0, 126.7, 125.7, 69.5, 55.3, 38.6, 21.7; HRMS (ESI) calcd for C₁₆H₁₈ONa [M+Na⁺] 249.1250, found 249.1250.

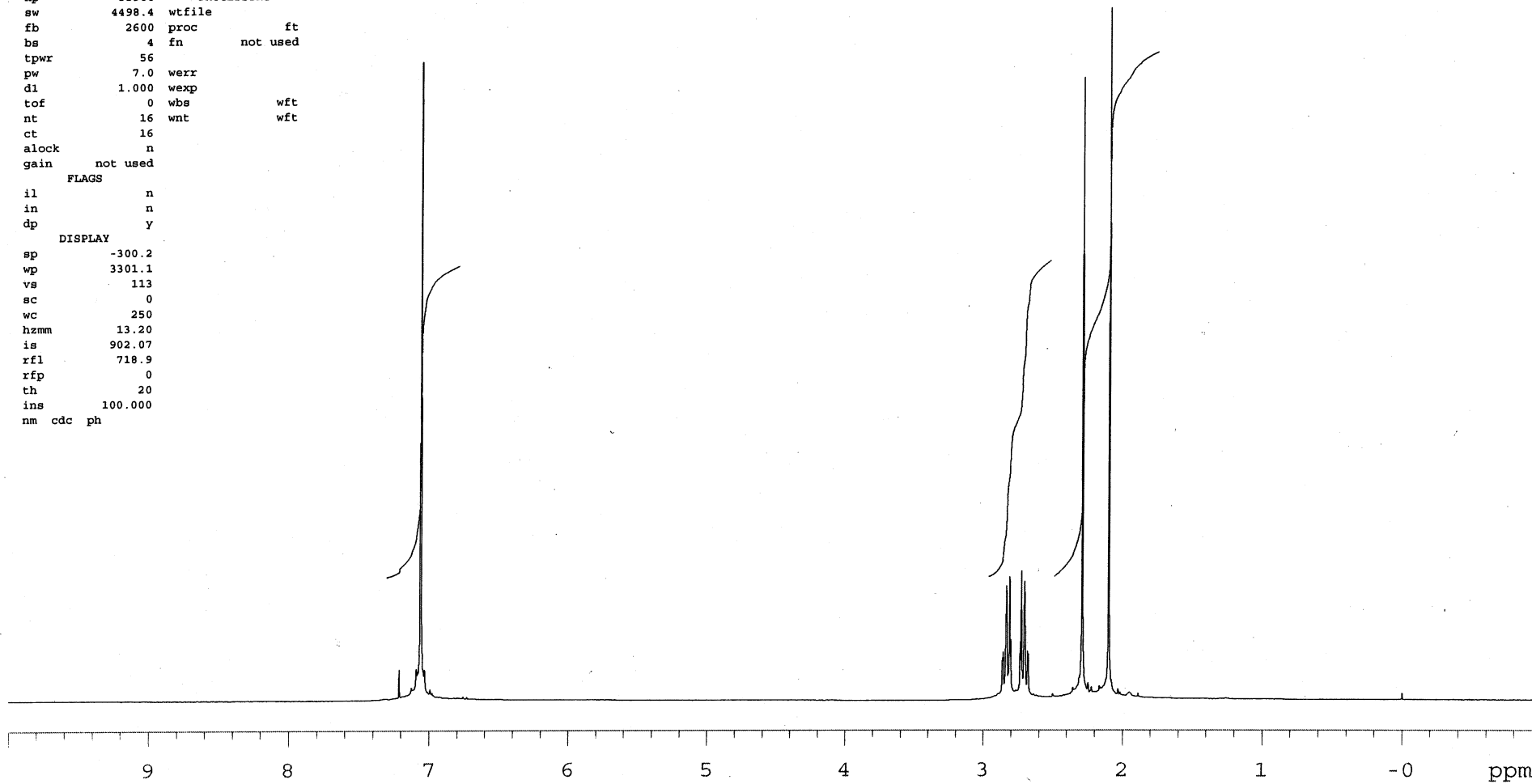
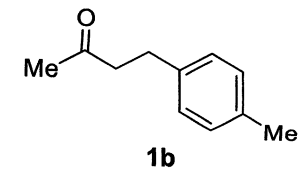
III. References

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STANDARD 1H OBSERVE

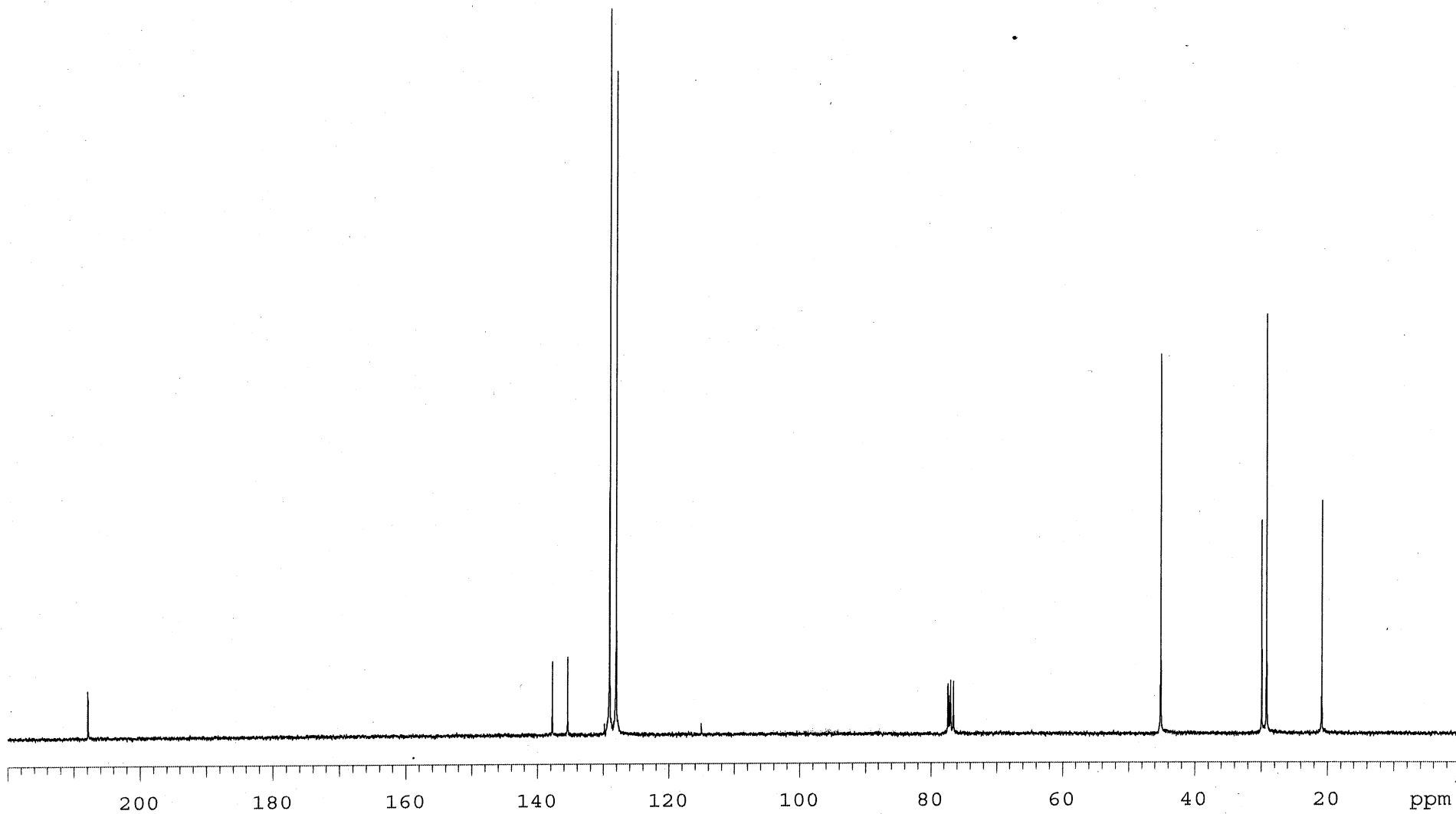
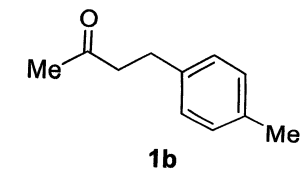
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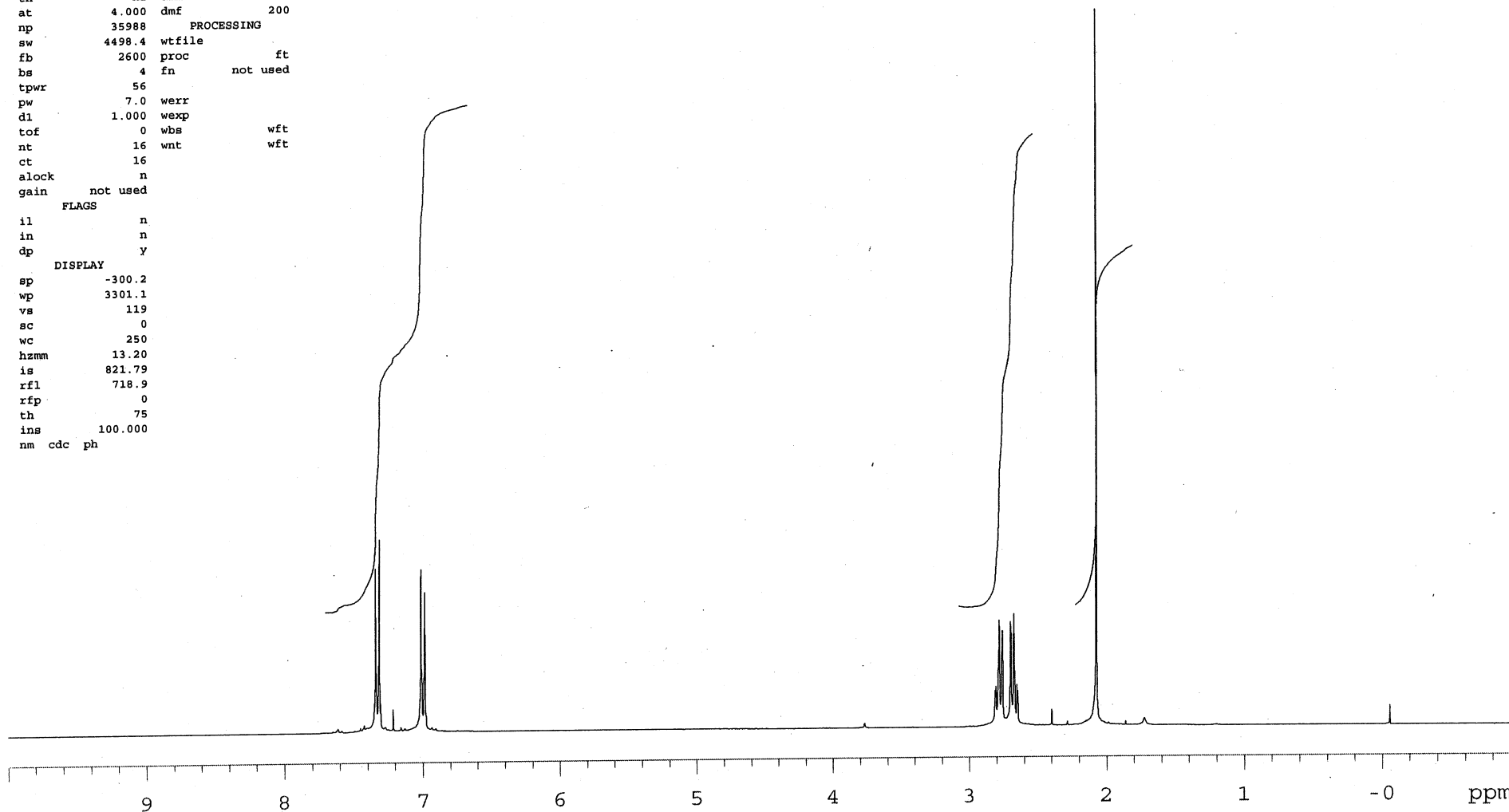
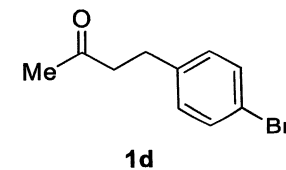
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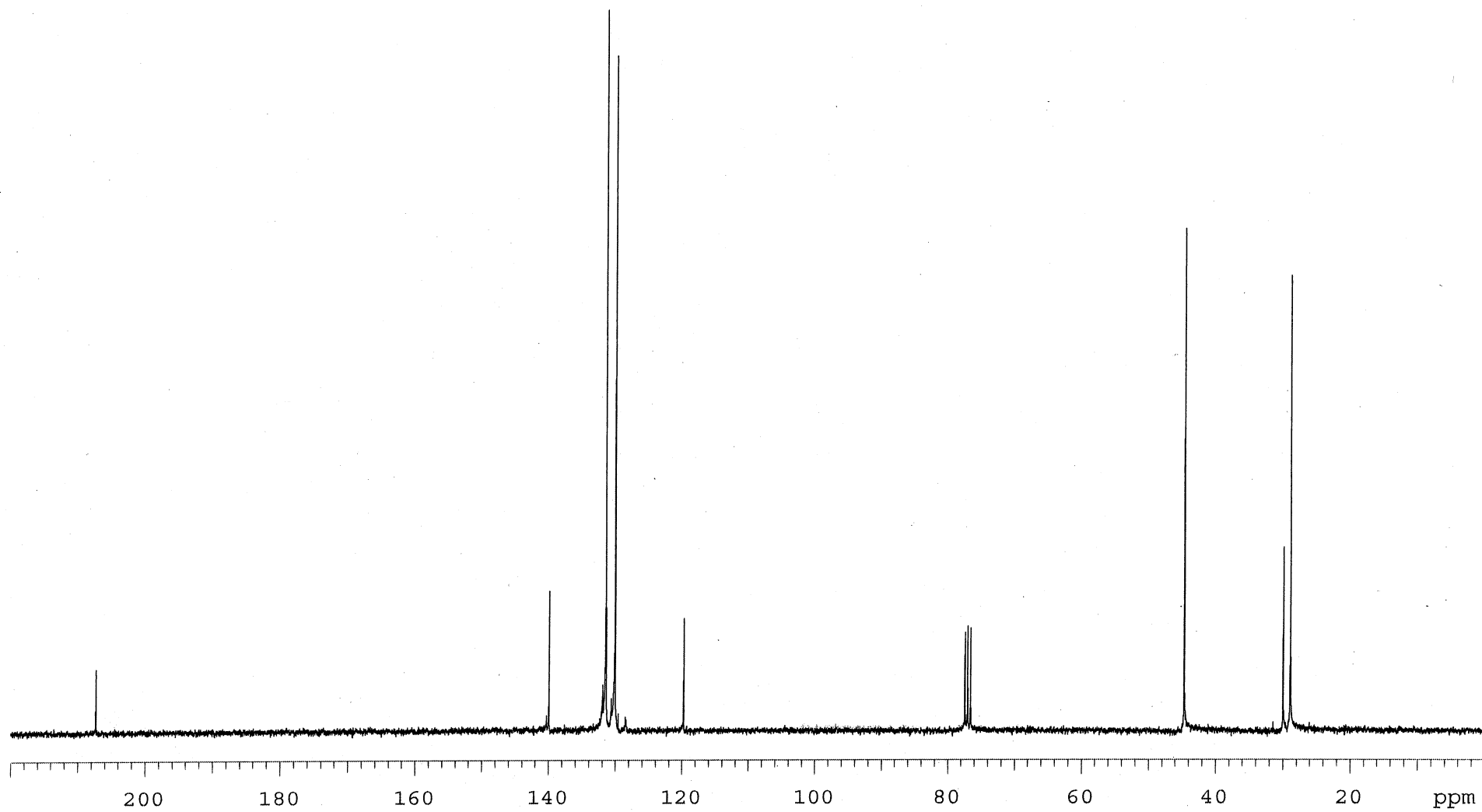
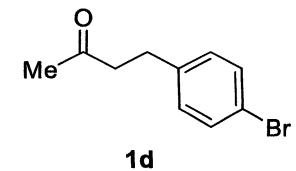
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KO A-17 FF2 13C

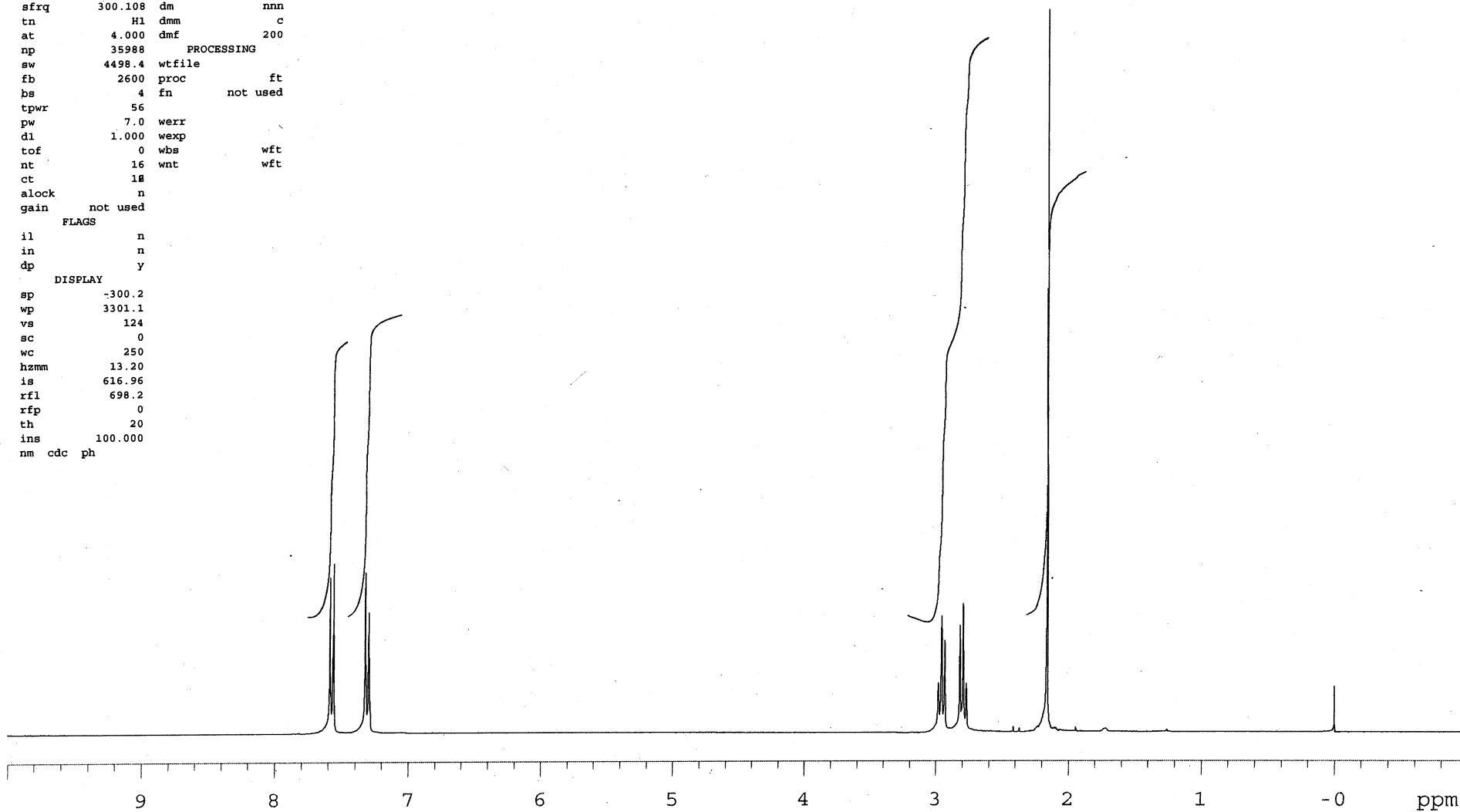
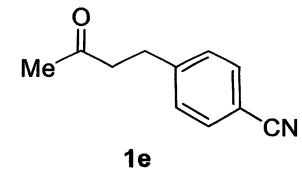
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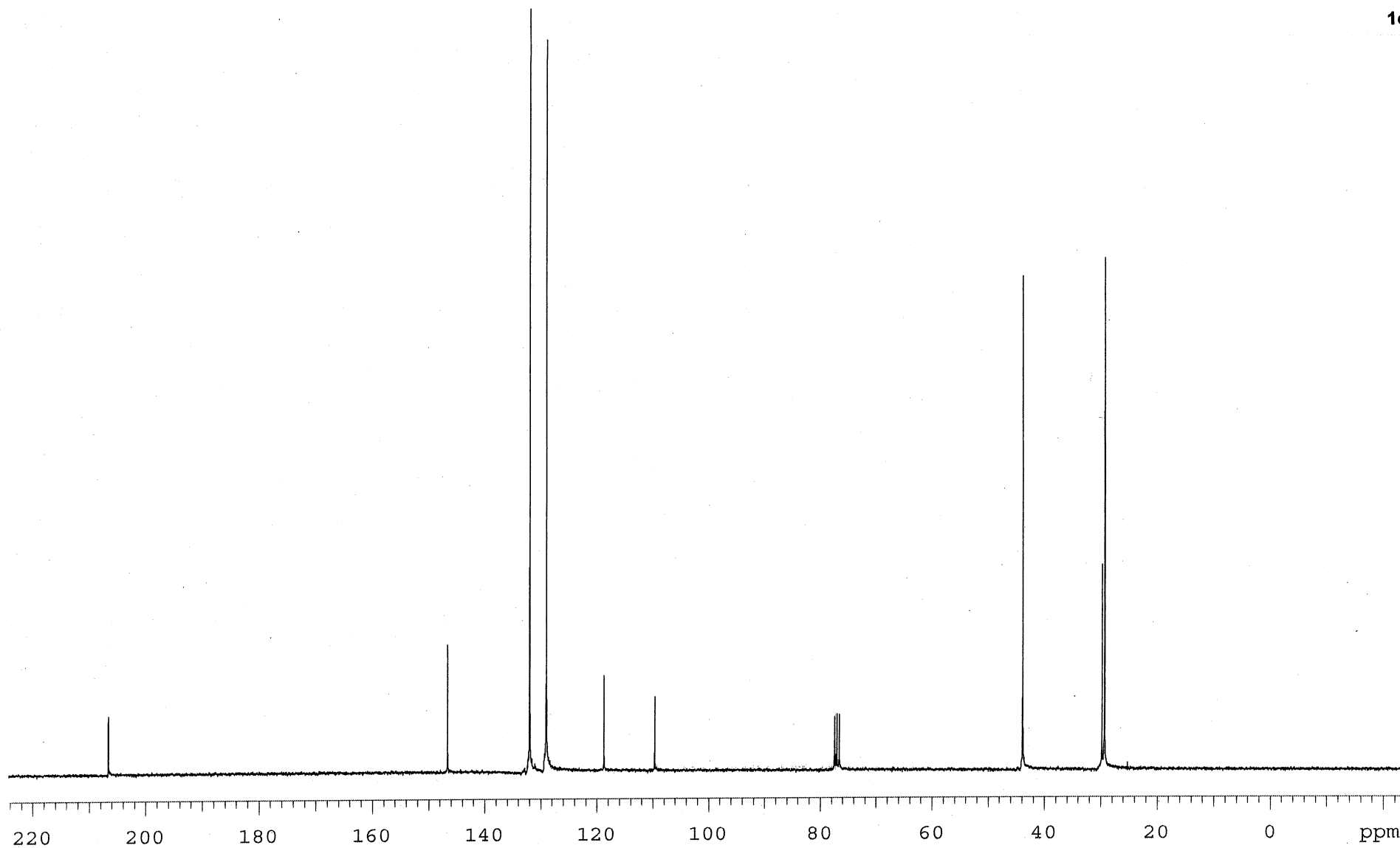
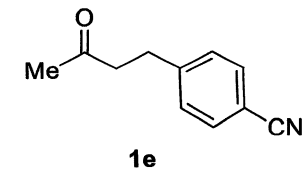
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MF D-52 FF1 13C

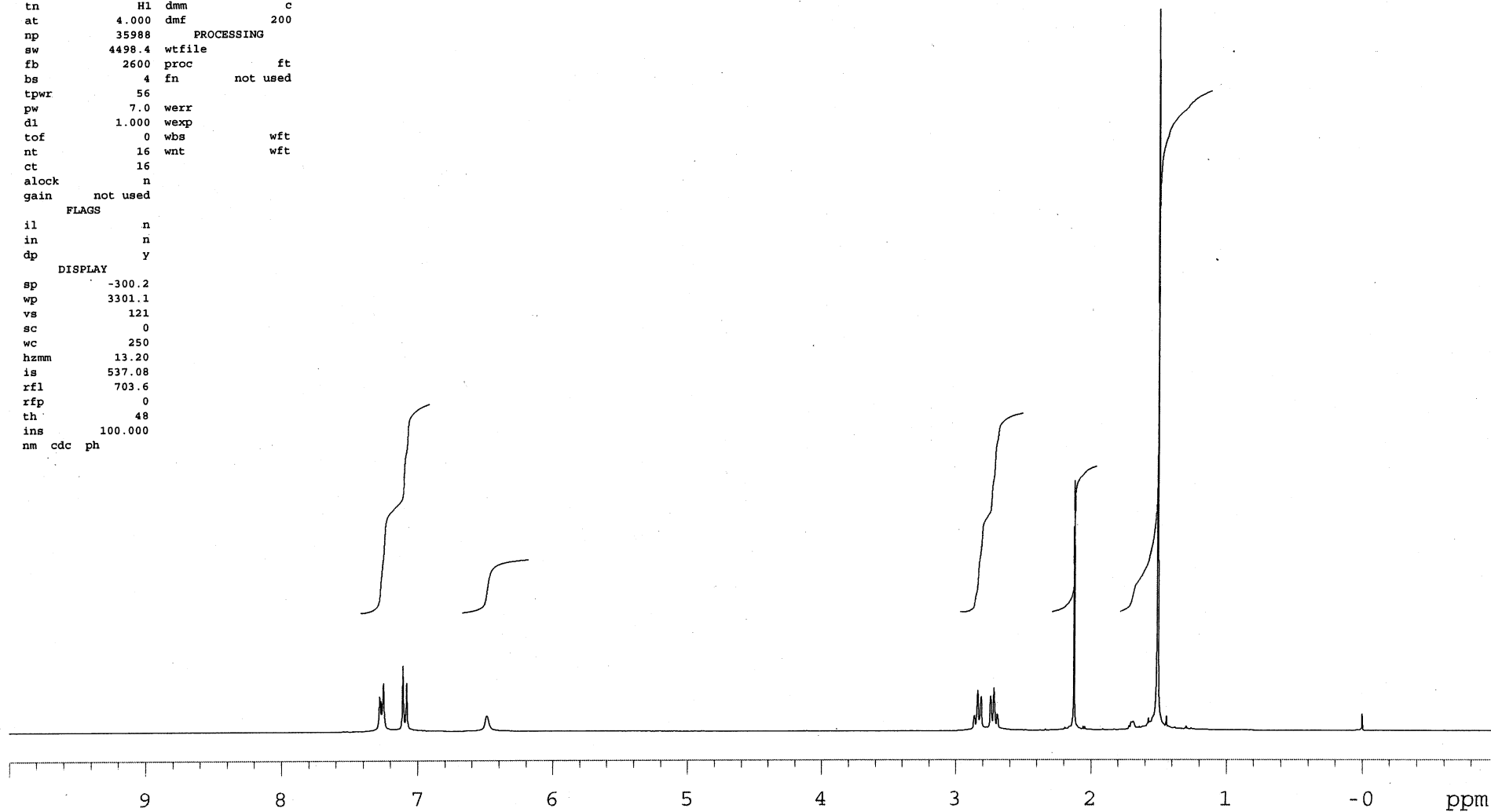
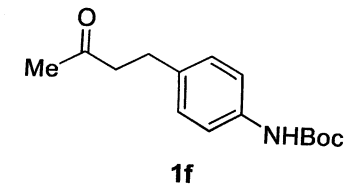
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MM A-33

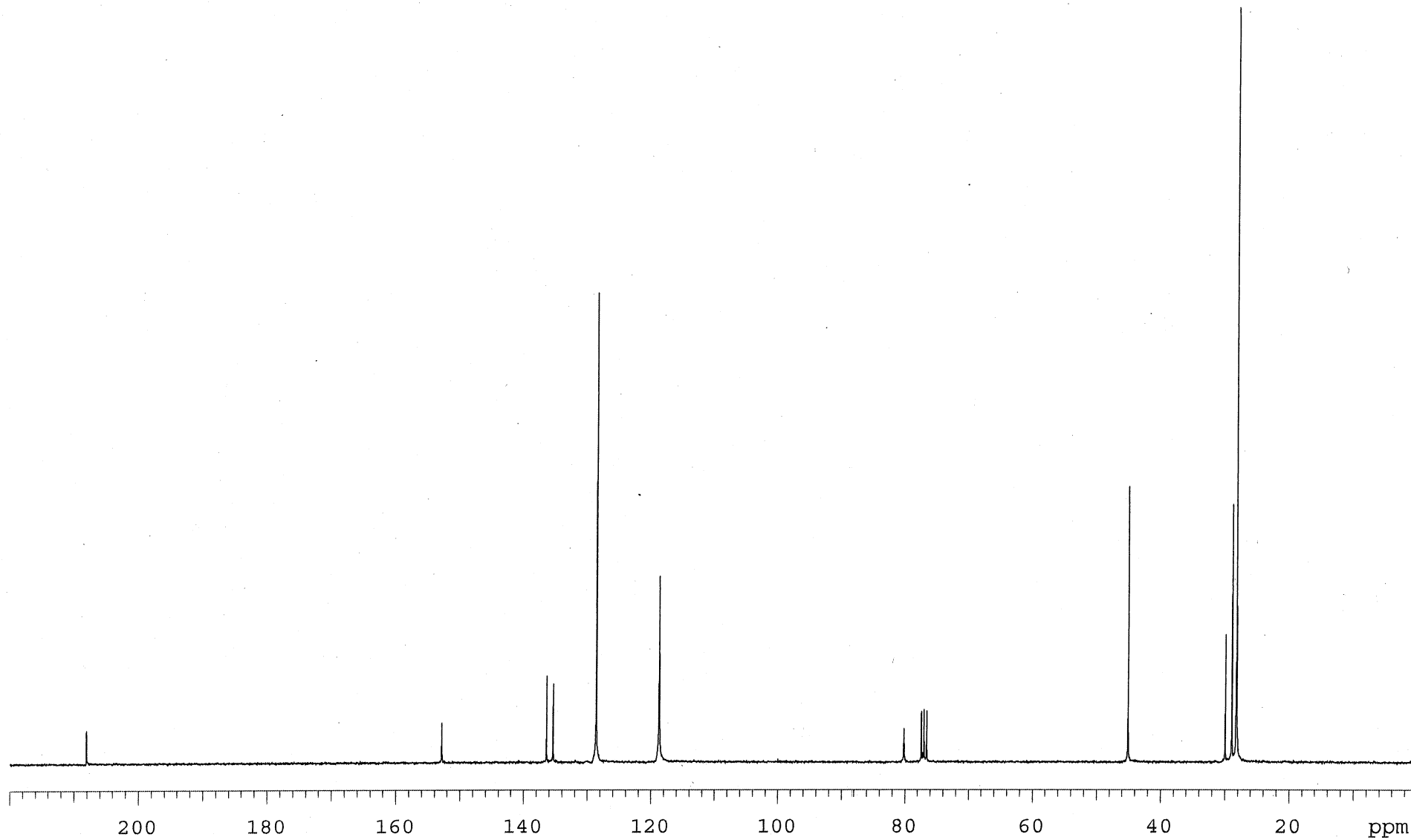
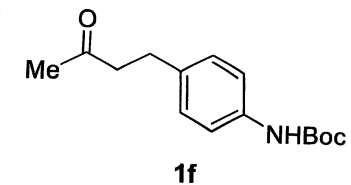
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MM A-33 13C

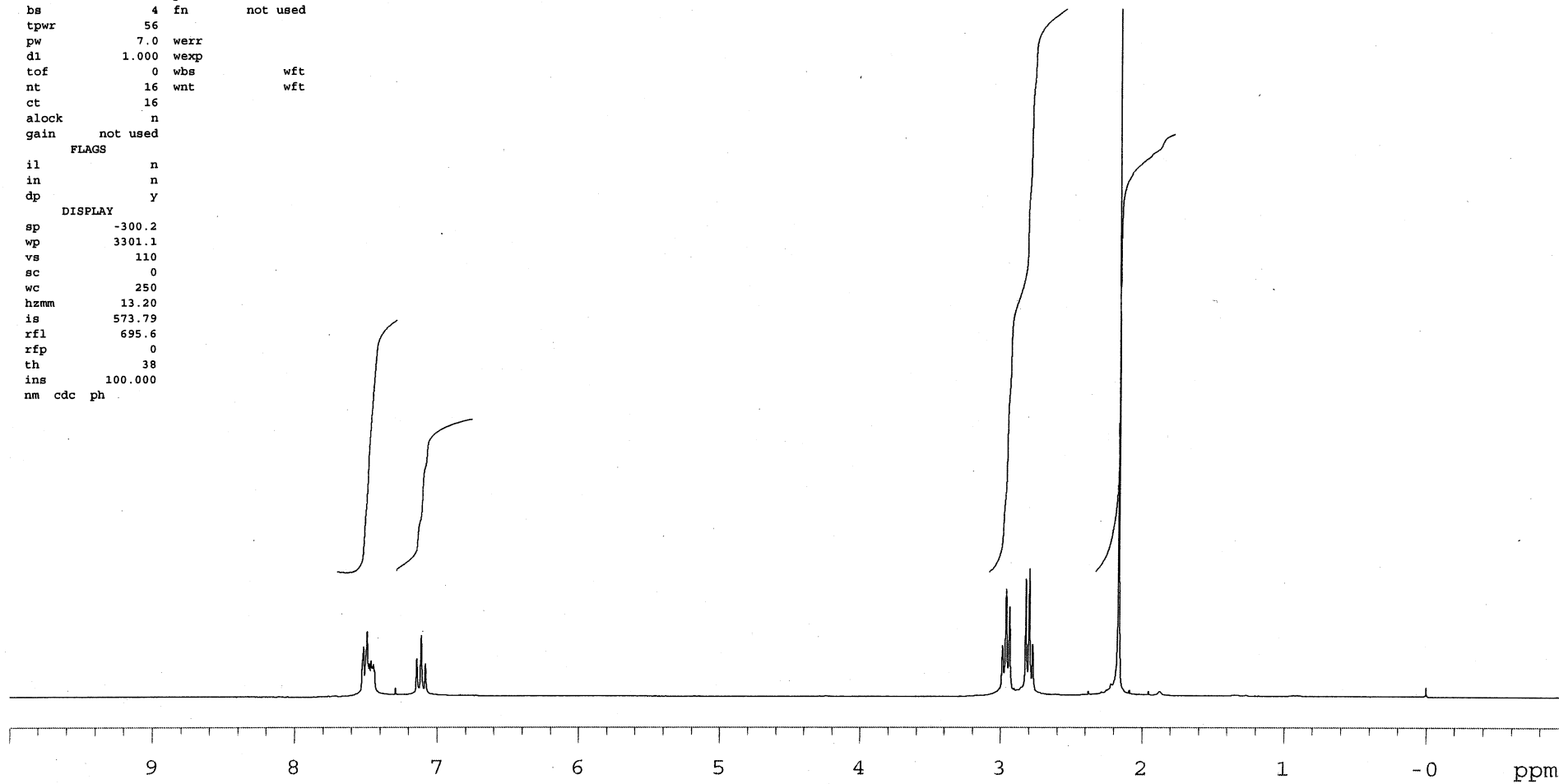
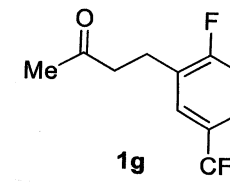
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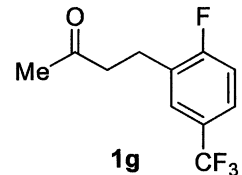
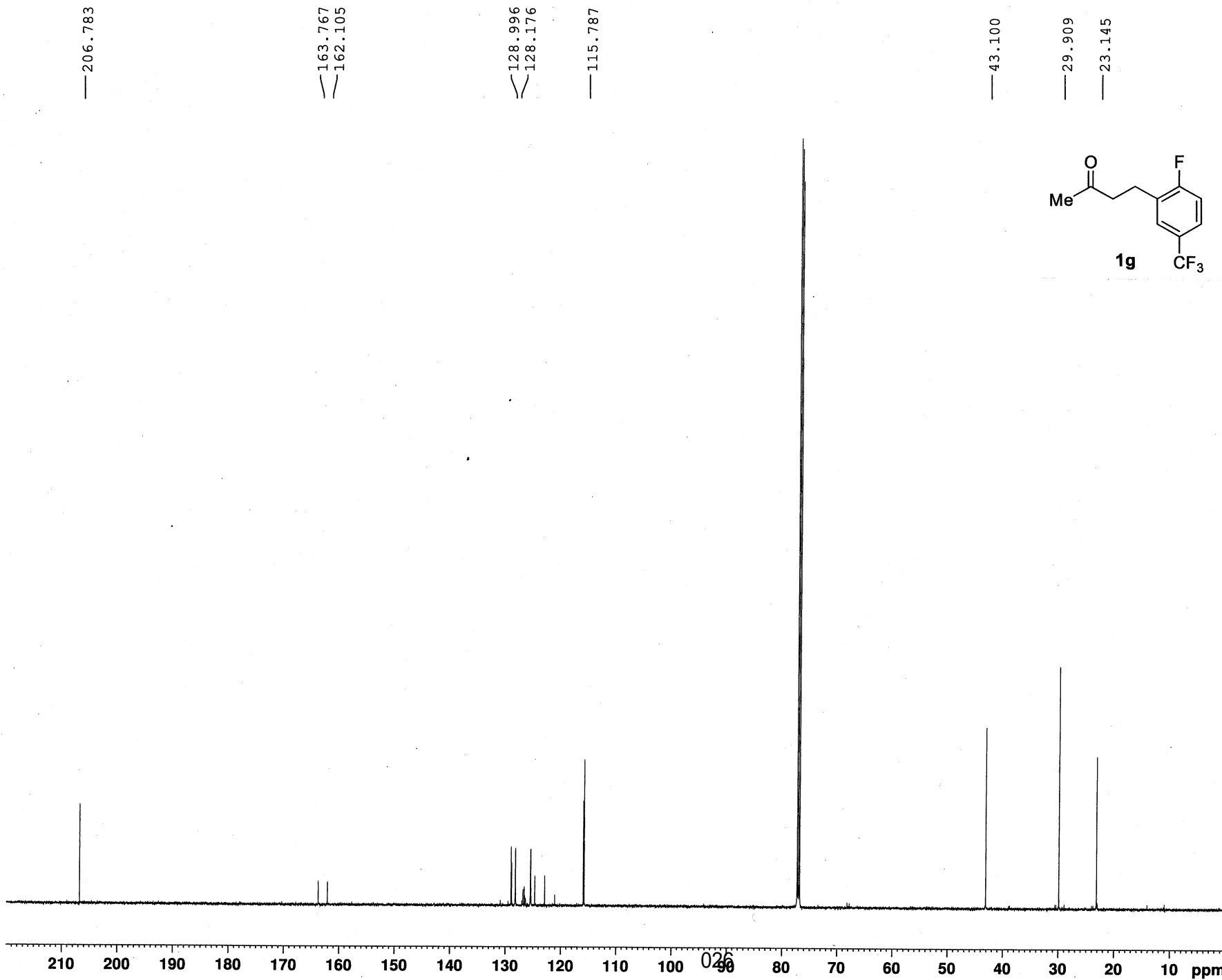


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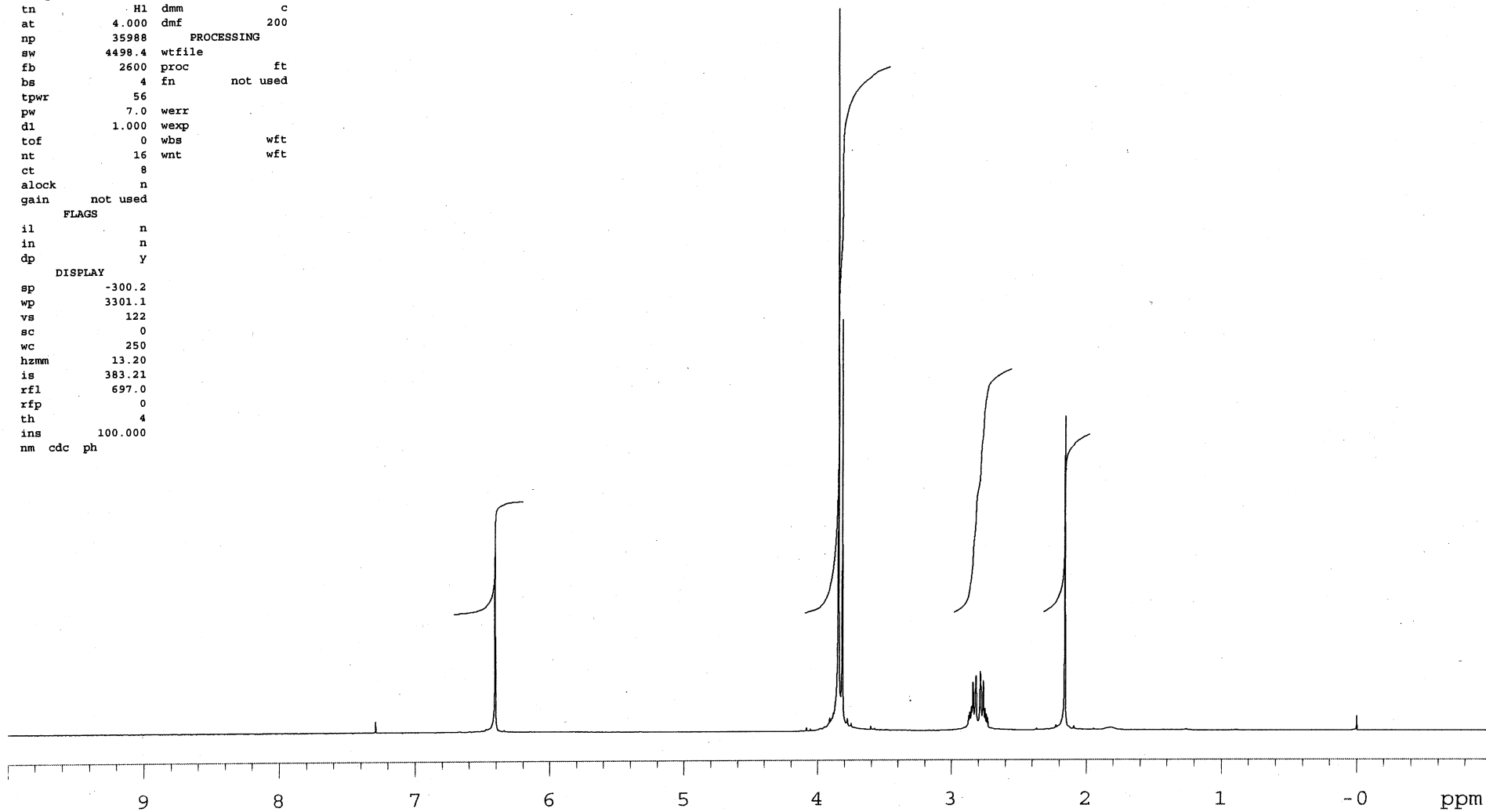
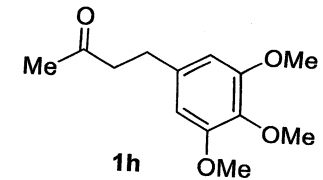
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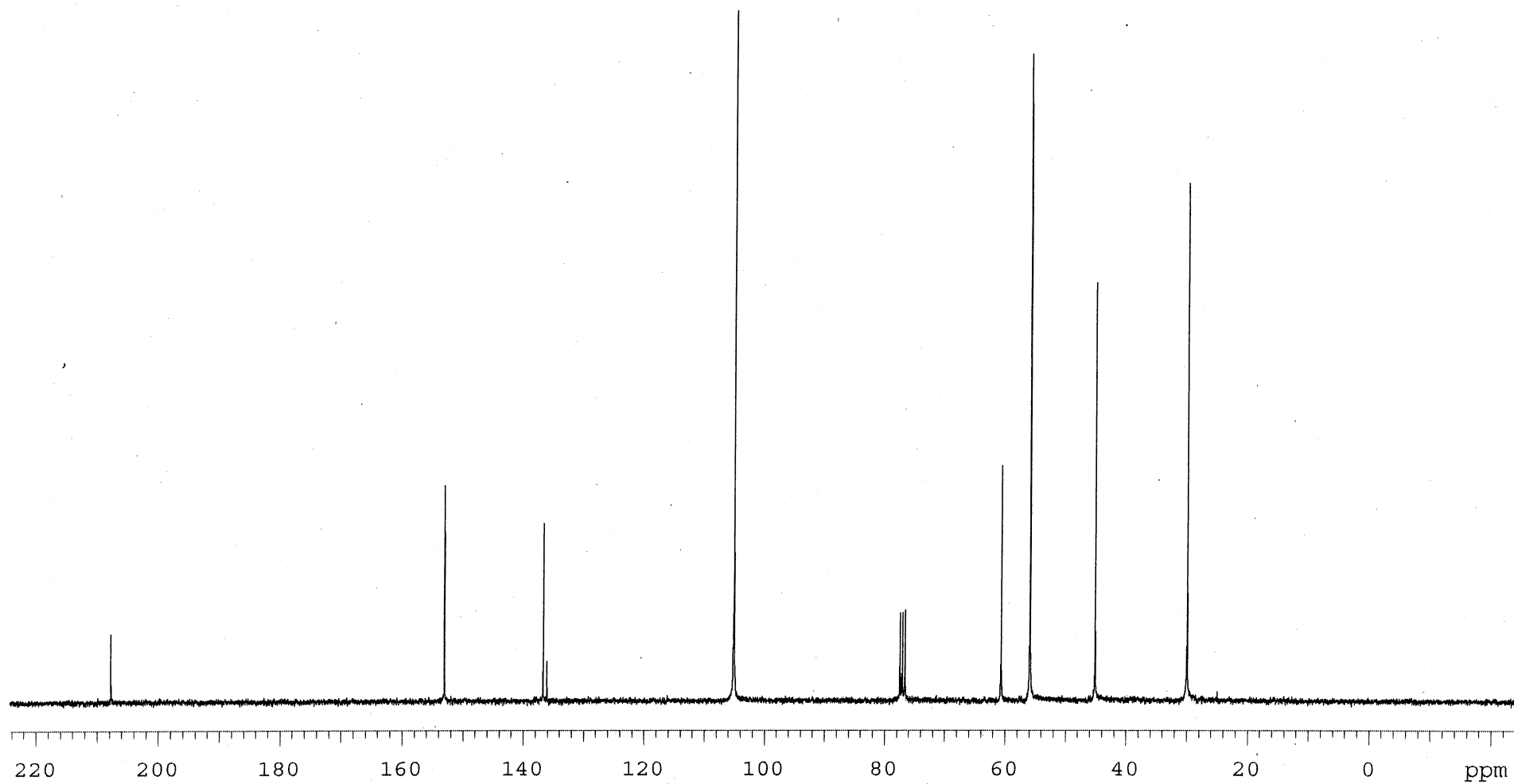
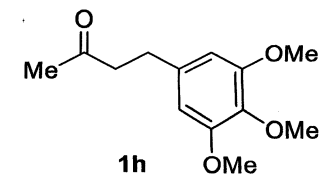
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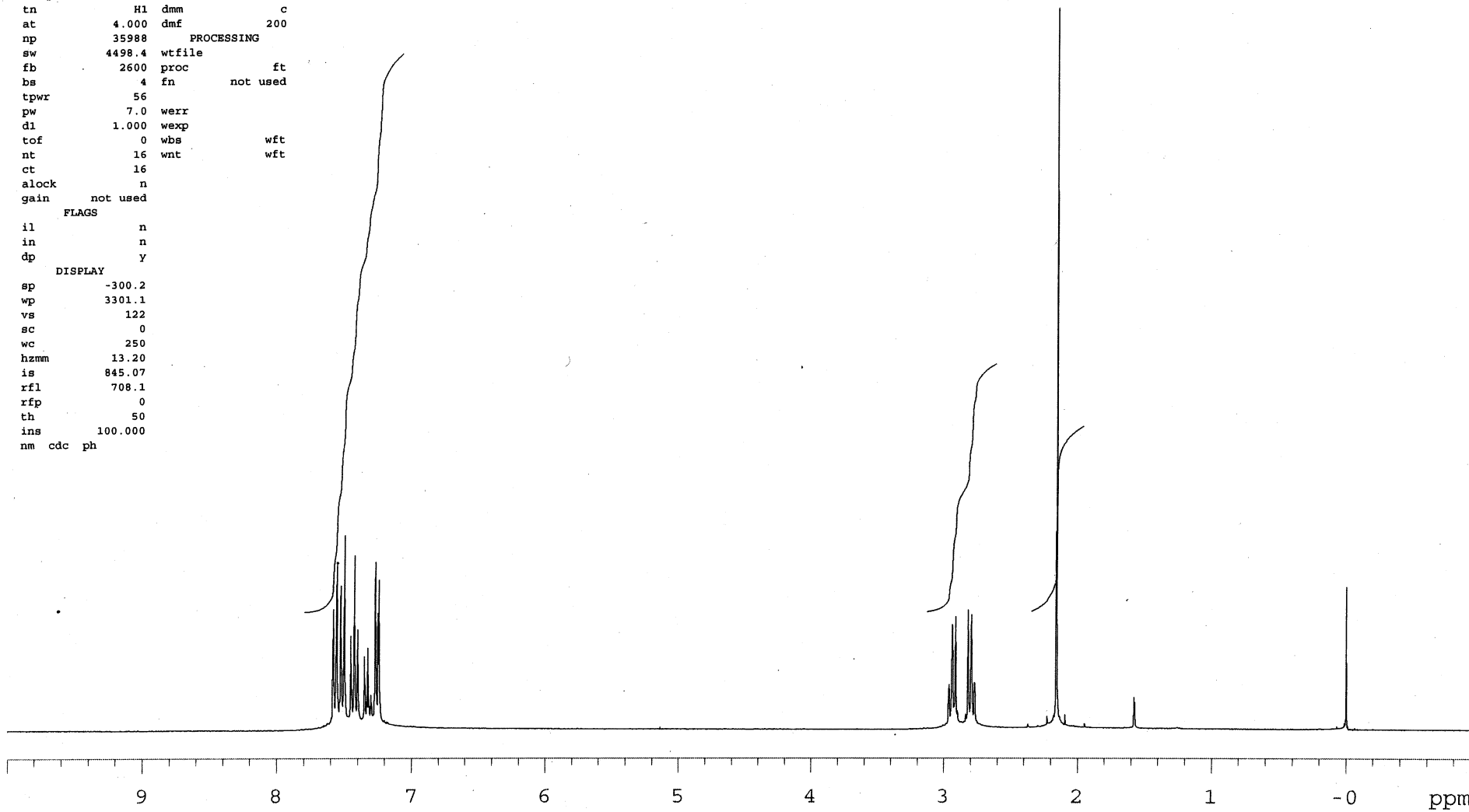
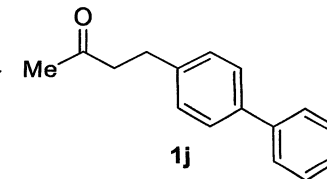
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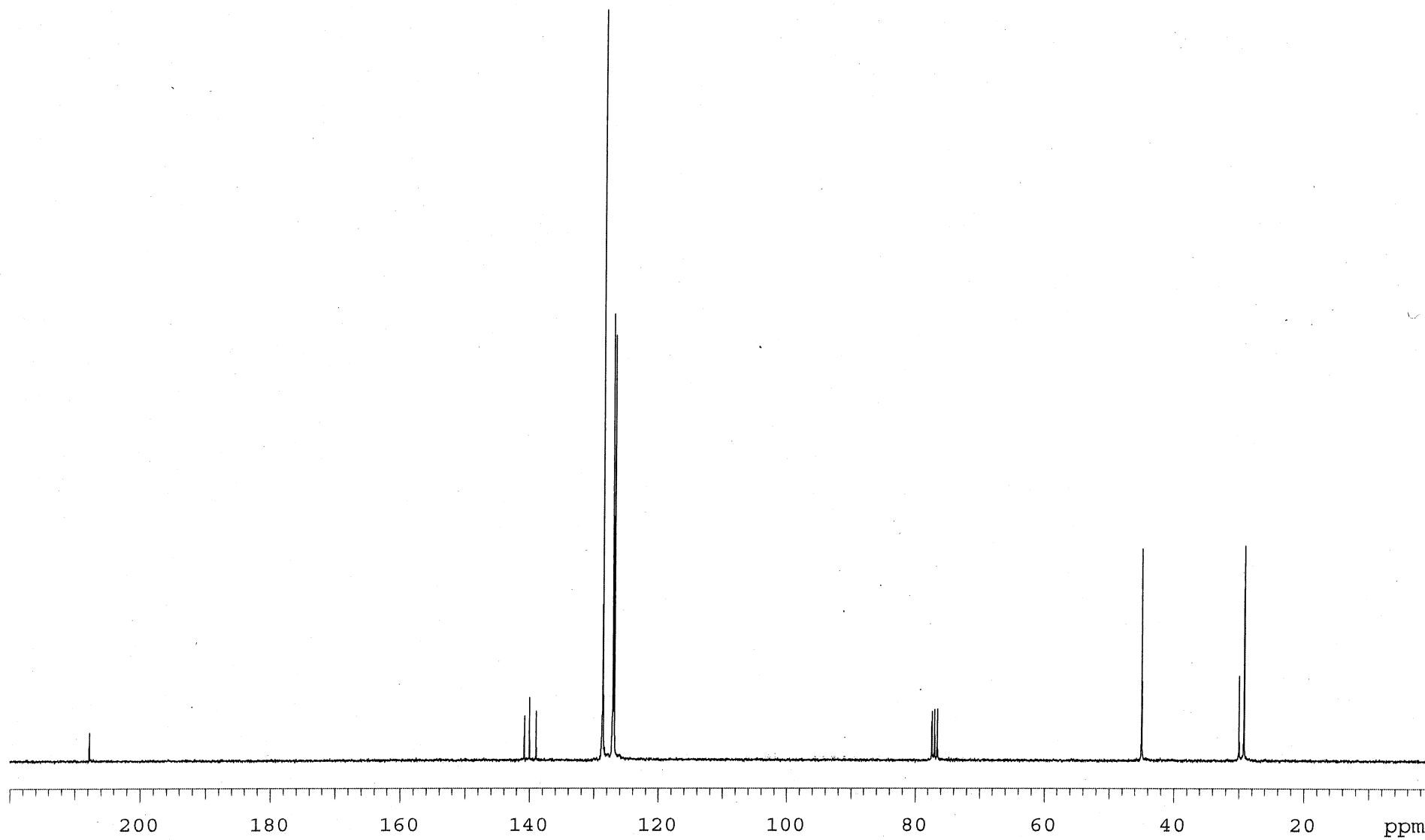
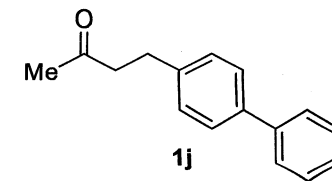
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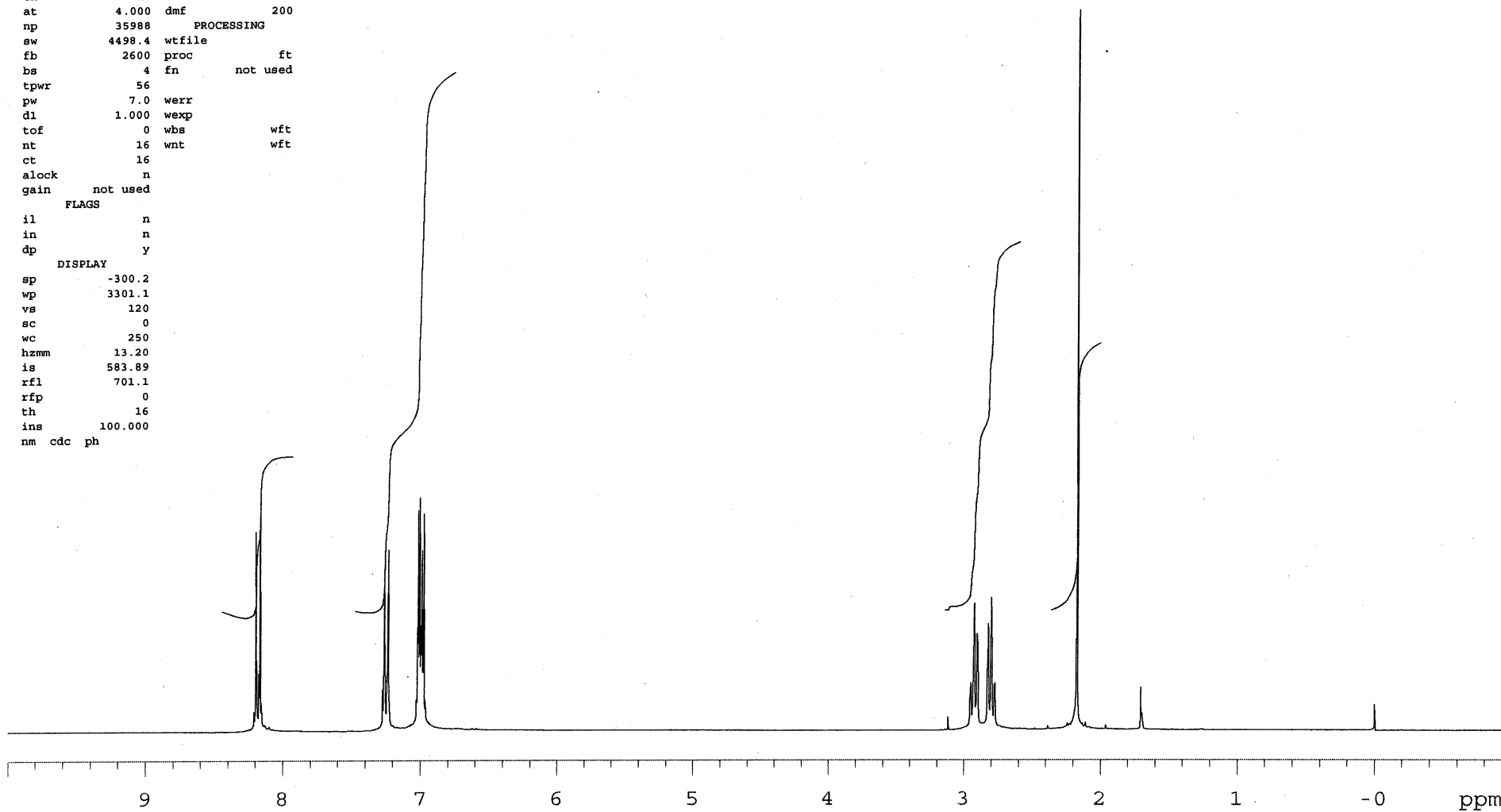
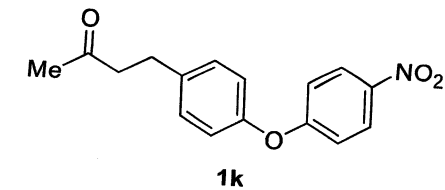
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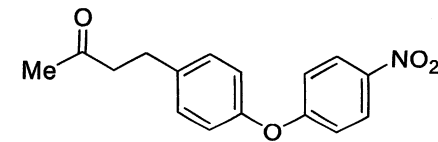
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```
SAMPLE          DEC. & VT
date  May 16 2017  dfrq    300.108
solvent  CDCl3    dn       H1
file     exp      dpwr    30
ACQUISITION  dof      0
sfrq     300.108  dm      nnn
tn       H1      dmm     c
at       4.000   dmf    200
np       35988   PROCESSING
sw       4498.4  wtfile
fb       2600   proc      ft
bs       4      fn      not used
tpwr     56
pw       7.0   werr
dl       1.000 wexp
tof      0     wbs      wft
nt       16   wnt      wft
ct       16
alock    n
gain     not used
FLAGS
il       n
in       n
dp       y
DISPLAY
sp      -300.2
wp      3301.1
vs      120
sc      0
wc      250
hzmm    13.20
is      583.89
rfl     701.1
rfp     0
th      16
ins     100.000
nm cdc ph
```

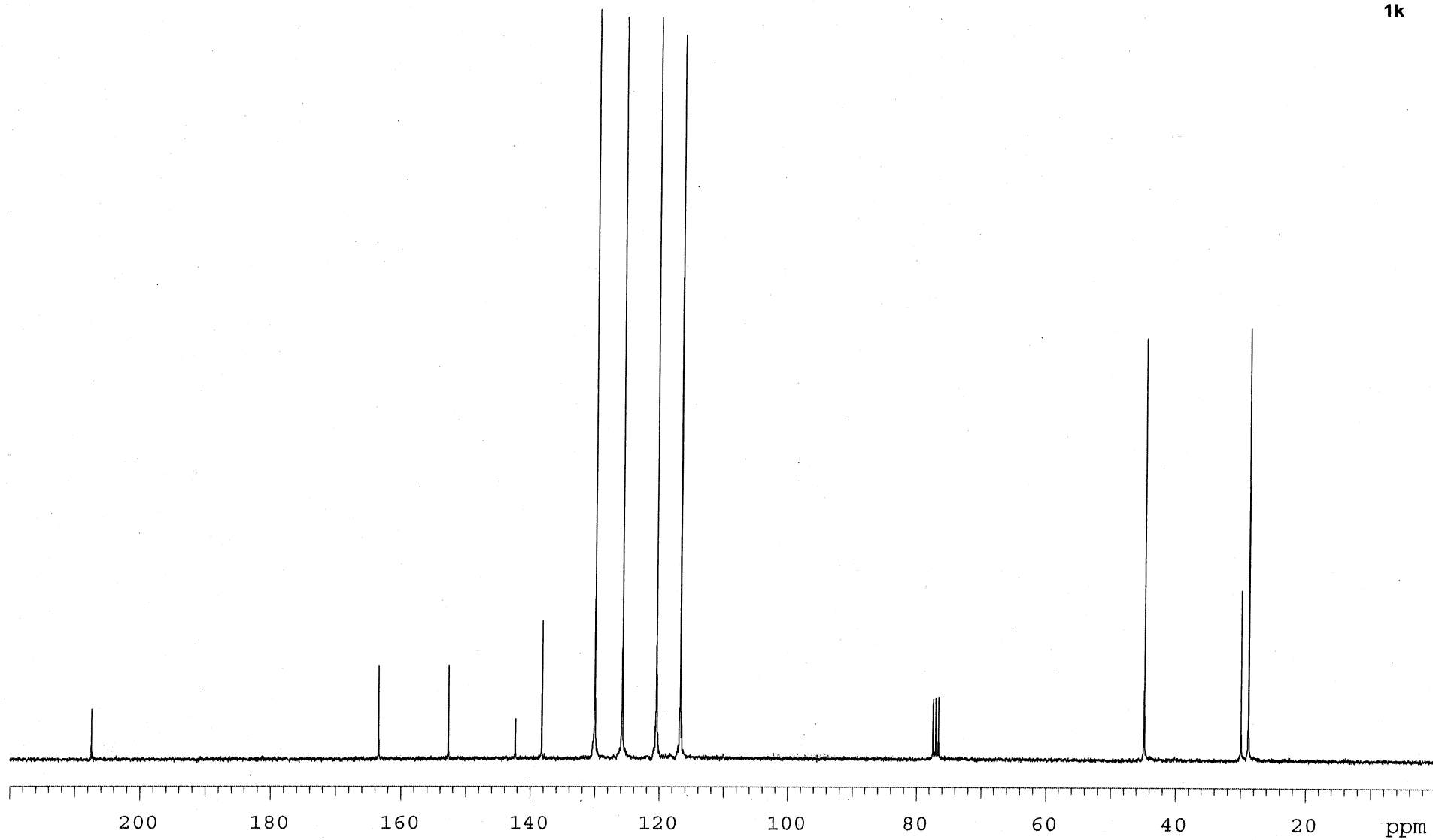


MF D-47 FF1 13C

Pulse Sequence: s2pul



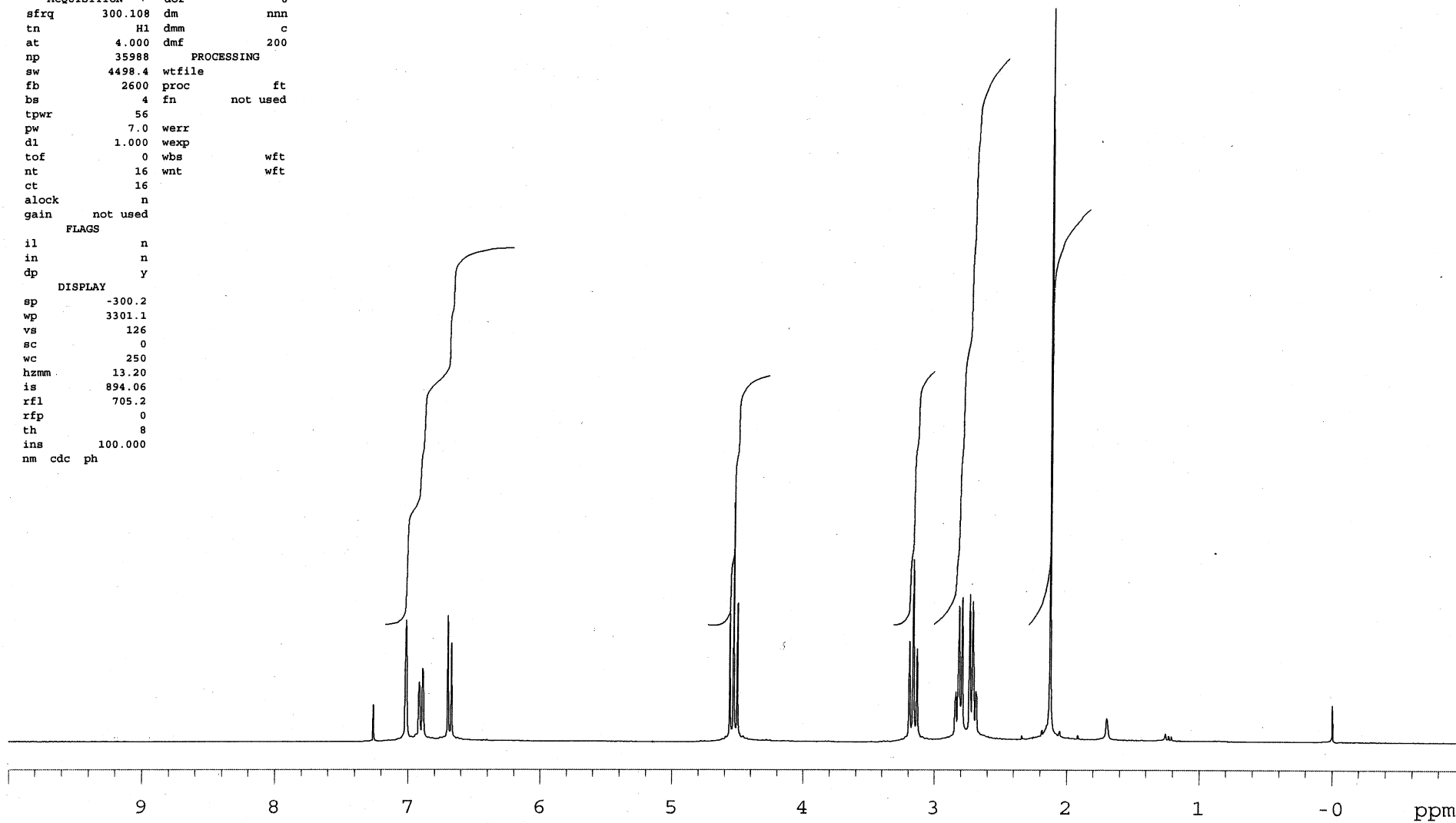
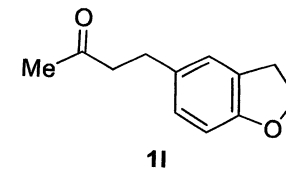
1k



TK A-14

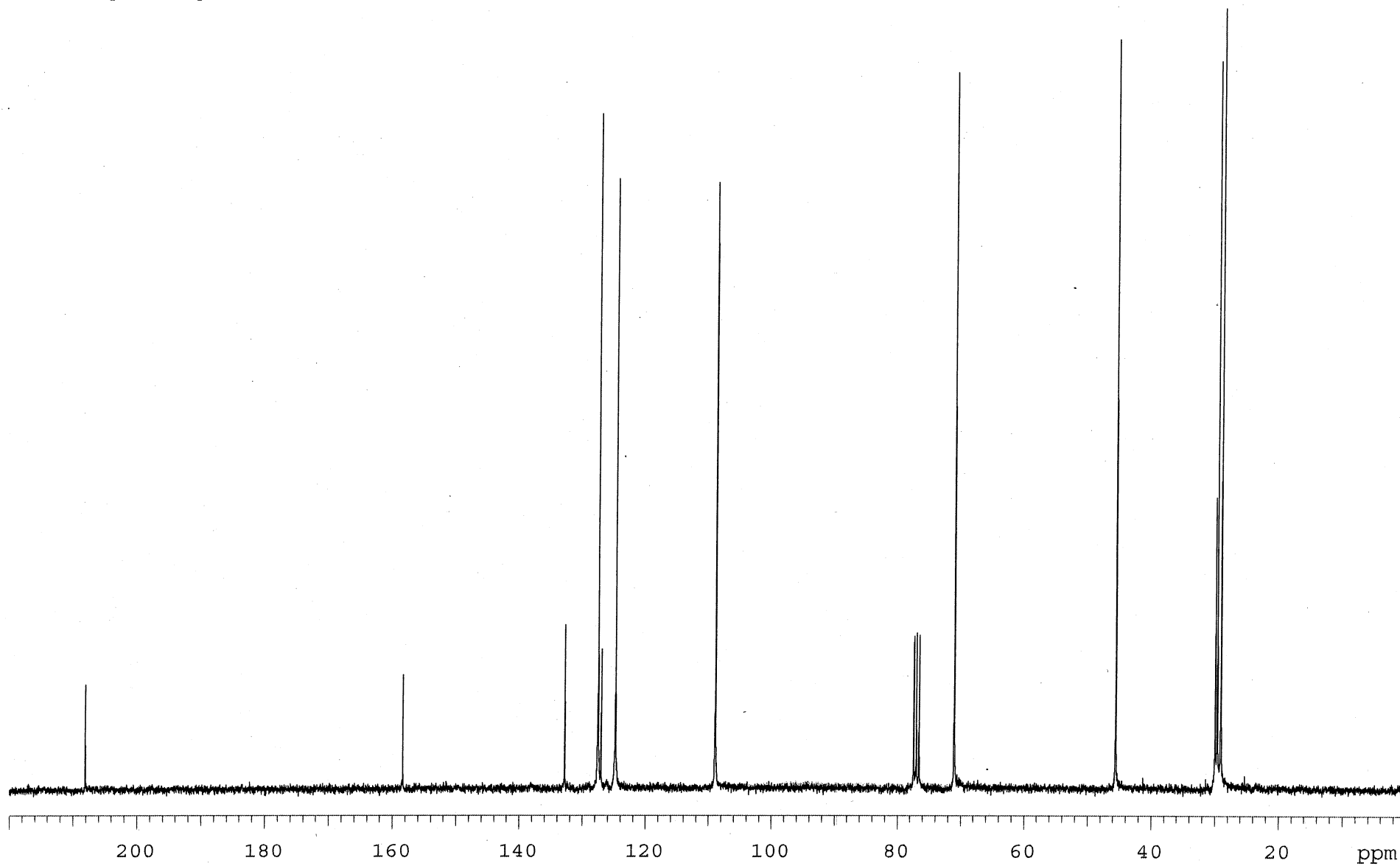
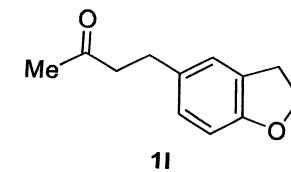
exp2 stdlh

```
SAMPLE          DEC. & VT
date  May 16 2017  dfrq    300.108
solvent CDCl3      dn      H1
file   exp        dpwr     30
ACQUISITION      dof      0
sfrq   300.108   dm       nnn
tn     H1        dmm      c
at     4.000     dmf      200
np     35988     PROCESSING
sw     4498.4    wtfile
fb     2600     proc      ft
bs     4         fn       not used
tpwr   56
pw     7.0      werr
d1     1.000    wexp
tof    0        wbs      wft
nt     16      wnt      wft
ct     16
alock  n
gain   not used
FLAGS
il     n
in     n
dp     Y
DISPLAY
sp     -300.2
wp     3301.1
vs     126
sc     0
wc     250
hzmm   13.20
is     894.06
rfl    705.2
rfp    0
th     8
ins    100.000
nm cdc ph
```



TK A-14 13C

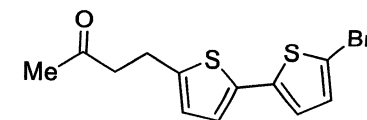
Pulse Sequence: s2pul



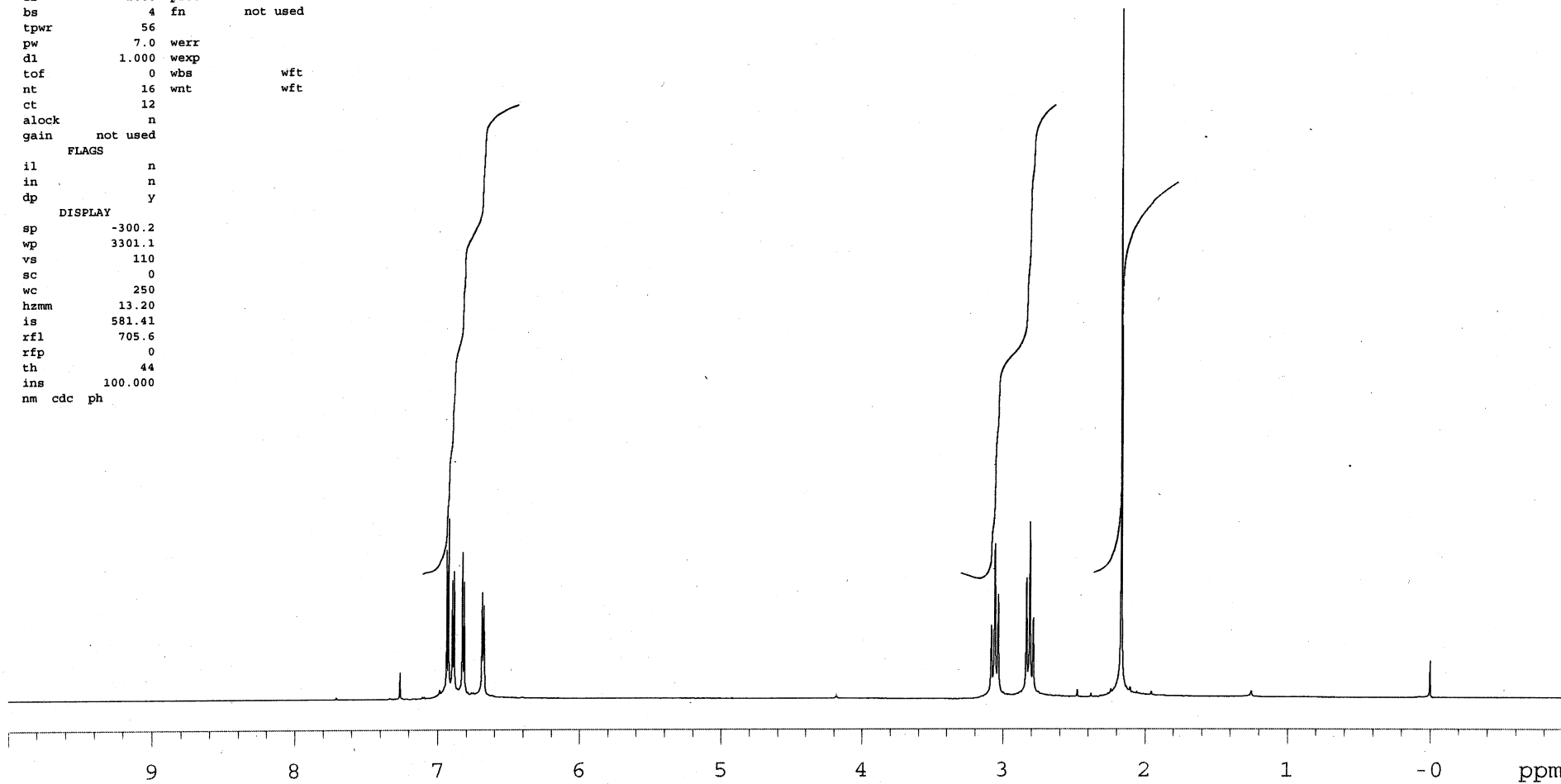
MF E-16 PTLCl

expl stdlh

SAMPLE		DEC. & VT	
date	May 29 2017	dfrq	300.108
solvent	CDCl3	dn	H1
file	exp	dpwr	30
ACQUISITION		PROCESSING	
sfrq	300.108	dm	nnn
tn	H1	dmm	c
at	4.000	dmf	200
np	35988		
sw	4498.4	wtfile	
fb	2600	proc	ft
bs	4	fn	not used
tpwr	56		
pw	7.0	werr	
d1	1.000	wexp	
tof	0	wbs	wft
nt	16	wnt	wft
ct	12		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	-300.2		
wp	3301.1		
vs	110		
sc	0		
wc	250		
hzmm	13.20		
is	581.41		
rfl	705.6		
rfp	0		
th	44		
ins	100.000		
nm	cdc ph		

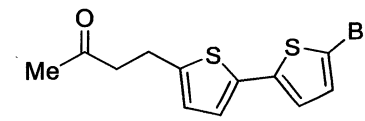


1m

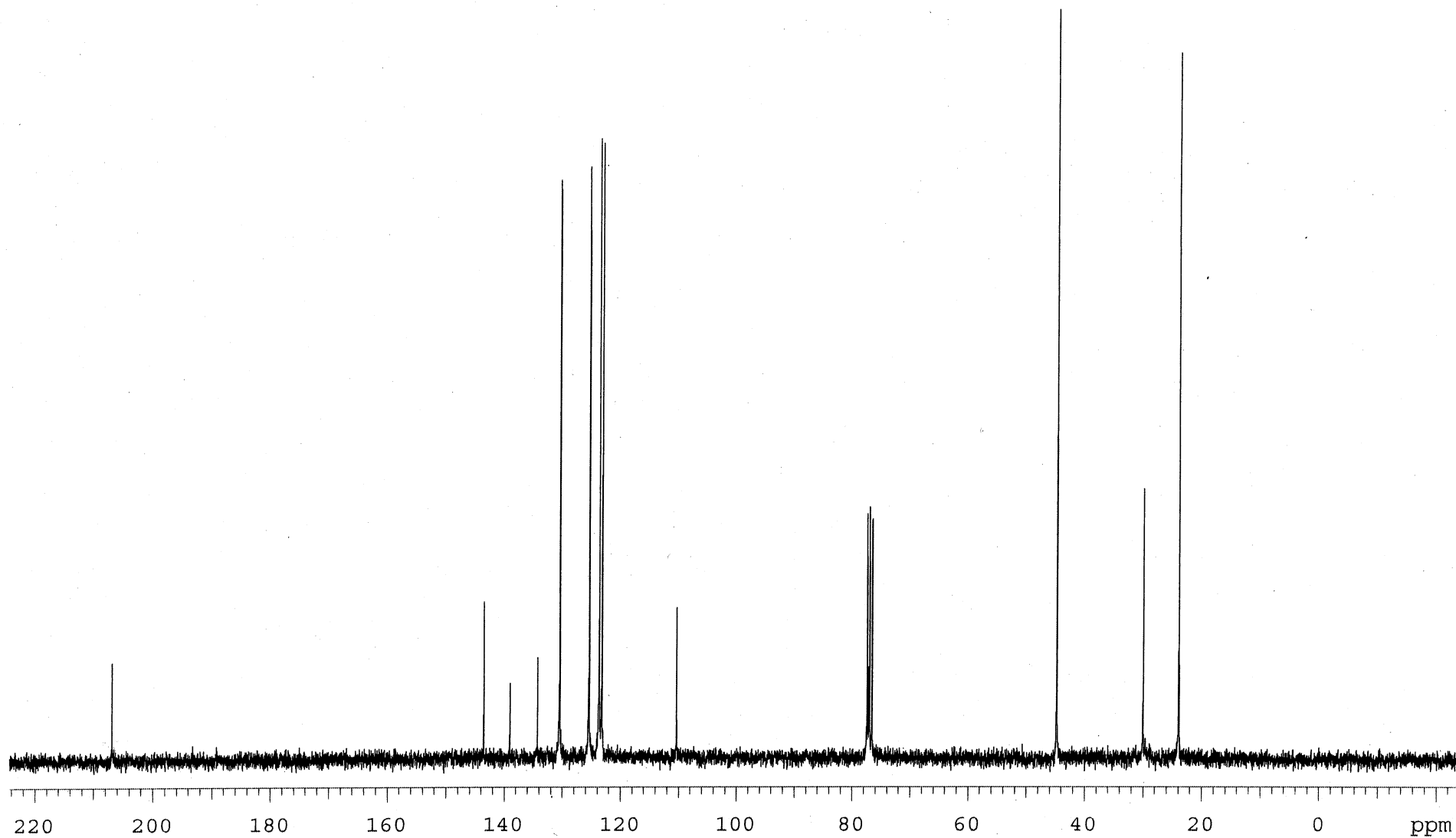


MF F-16 PTLCl 13C

Pulse Sequence: s2pul



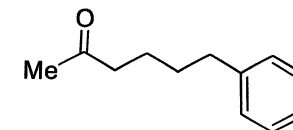
1m



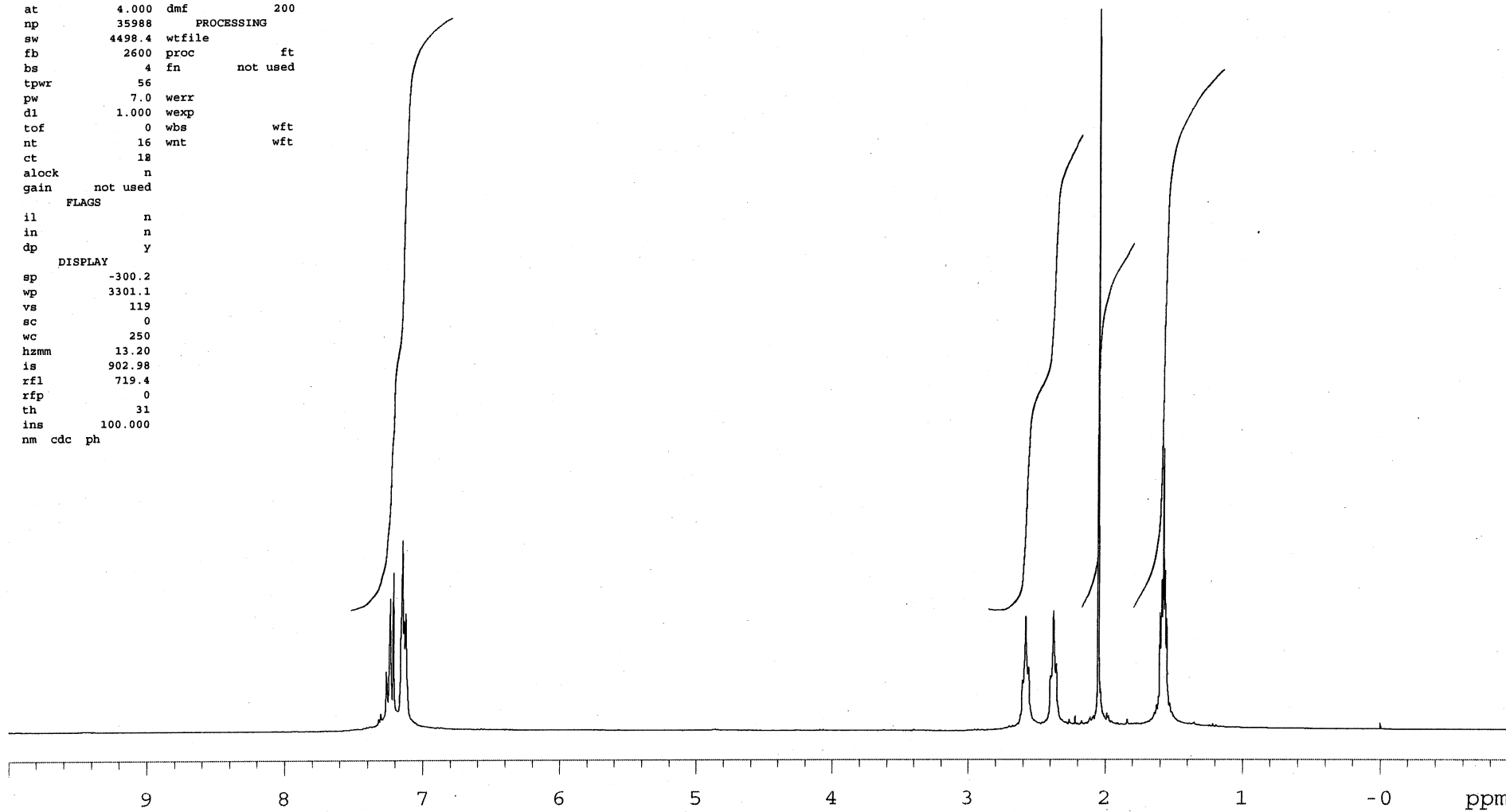
MF A-49

exp2 std1h

```
SAMPLE          DEC. & VT
date May 16 2017 dfrq      300.108
solvent CDCl3  dn         H1
file      exp  dpwr       30
ACQUISITION    dof        0
sfrq      300.108 dm       nnn
tn         H1  dmm        c
at         4.000 dmf      200
np         35988
sw         4498.4 wtfile
fb         2600  proc      ft
bs         4     fn       not used
tpwr       56
pw         7.0  werr
dl         1.000 wexp
tof        0    wbs      wft
nt         16  wnt      wft
ct         18
alock      n
gain      not used
FLAGS
il         n
in         n
dp         y
DISPLAY
sp        -300.2
wp        3301.1
vs        119
sc         0
wc         250
hzmm      13.20
is        902.98
rfl       719.4
rfp        0
th         31
ins       100.000
nm cdc ph
```

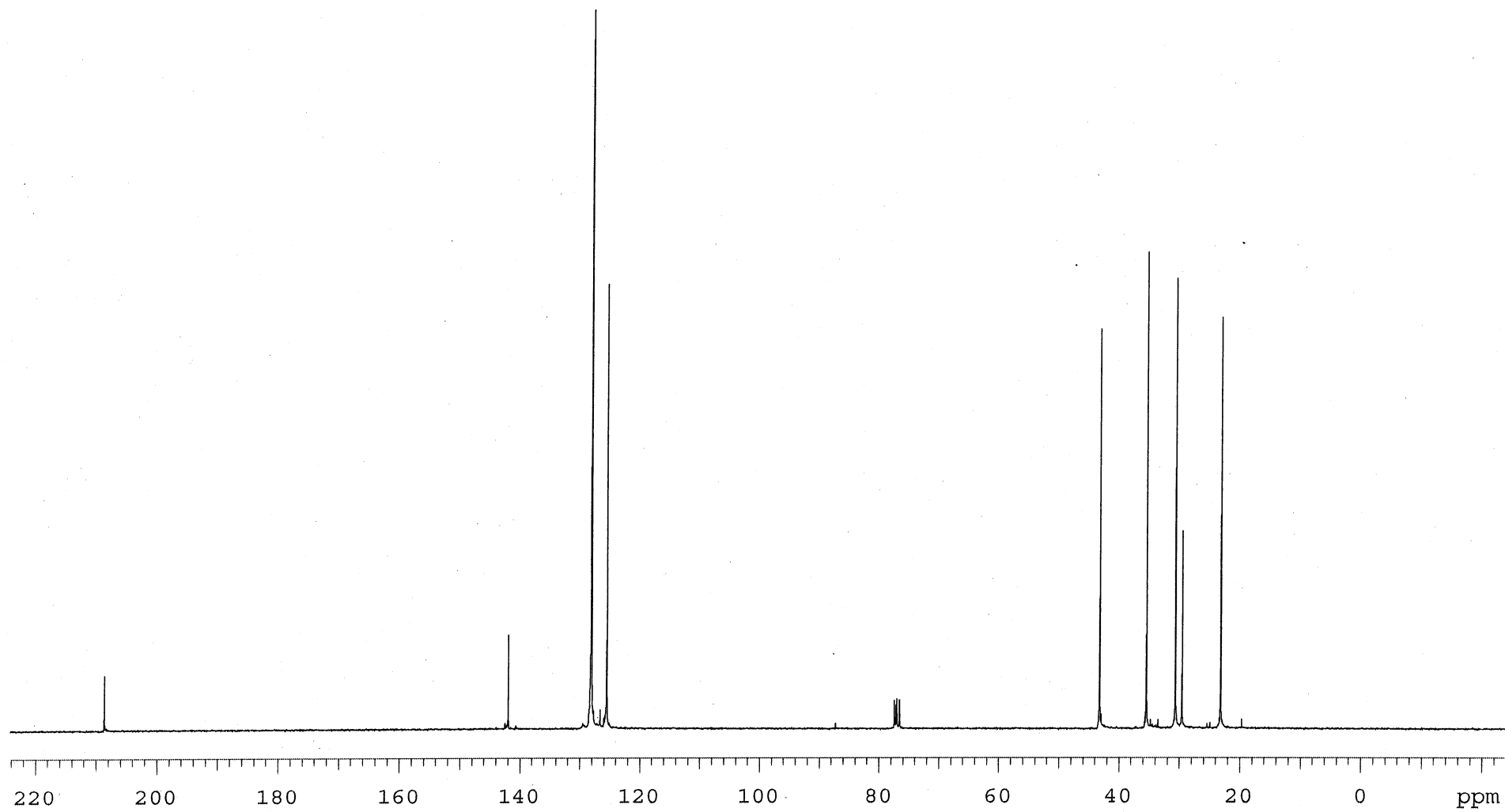
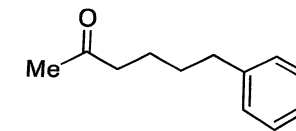


1n



MF A-49 13C

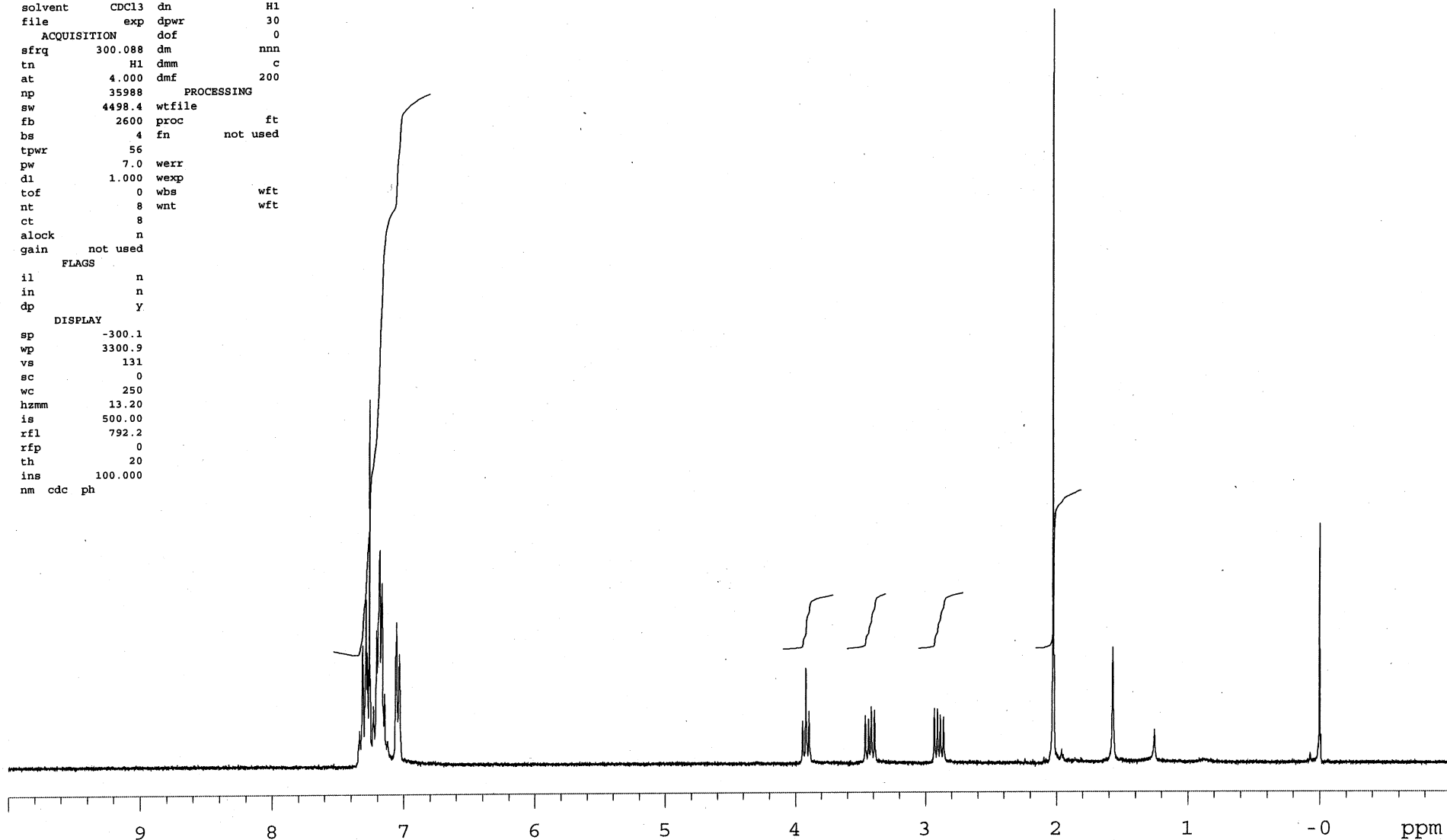
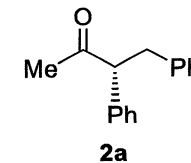
Pulse Sequence: s2pul



MF A-17

expl stdlh

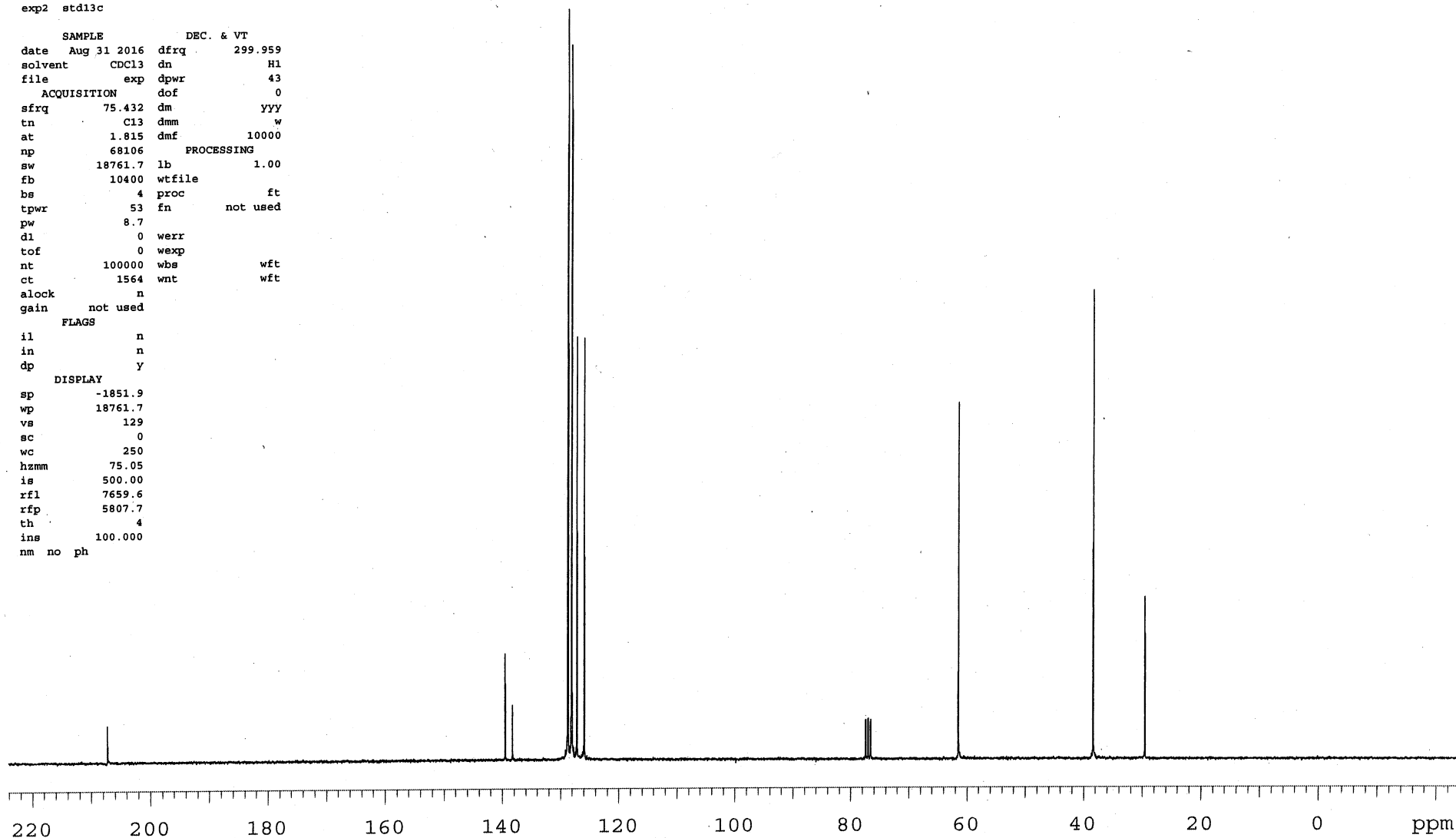
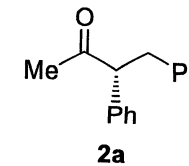
SAMPLE		DEC. & VT	
date	Mar 2 2016	dfrq	300.088
solvent	CDCl3	dn	H1
file	expl	dpwr	30
ACQUISITION		dof	0
sfrq	300.088	dm	nnn
tn	H1	dmm	c
at	4.000	dmf	200
np	35988	PROCESSING	
sw	4498.4	wtfile	
fb	2600	proc	ft
bs	4	fn	not used
tpwr	56		
pw	7.0	werr	
d1	1.000	wexp	
tof	0	wbs	wft
nt	8	wnt	wft
ct	8		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	Y		
DISPLAY			
sp	-300.1		
wp	3300.9		
vs	131		
sc	0		
wc	250		
hzmm	13.20		
is	500.00		
rfl	792.2		
rfp	0		
th	20		
ins	100.000		
nm	cdc ph		



MF C-21 FF1 13C

exp2 std13c

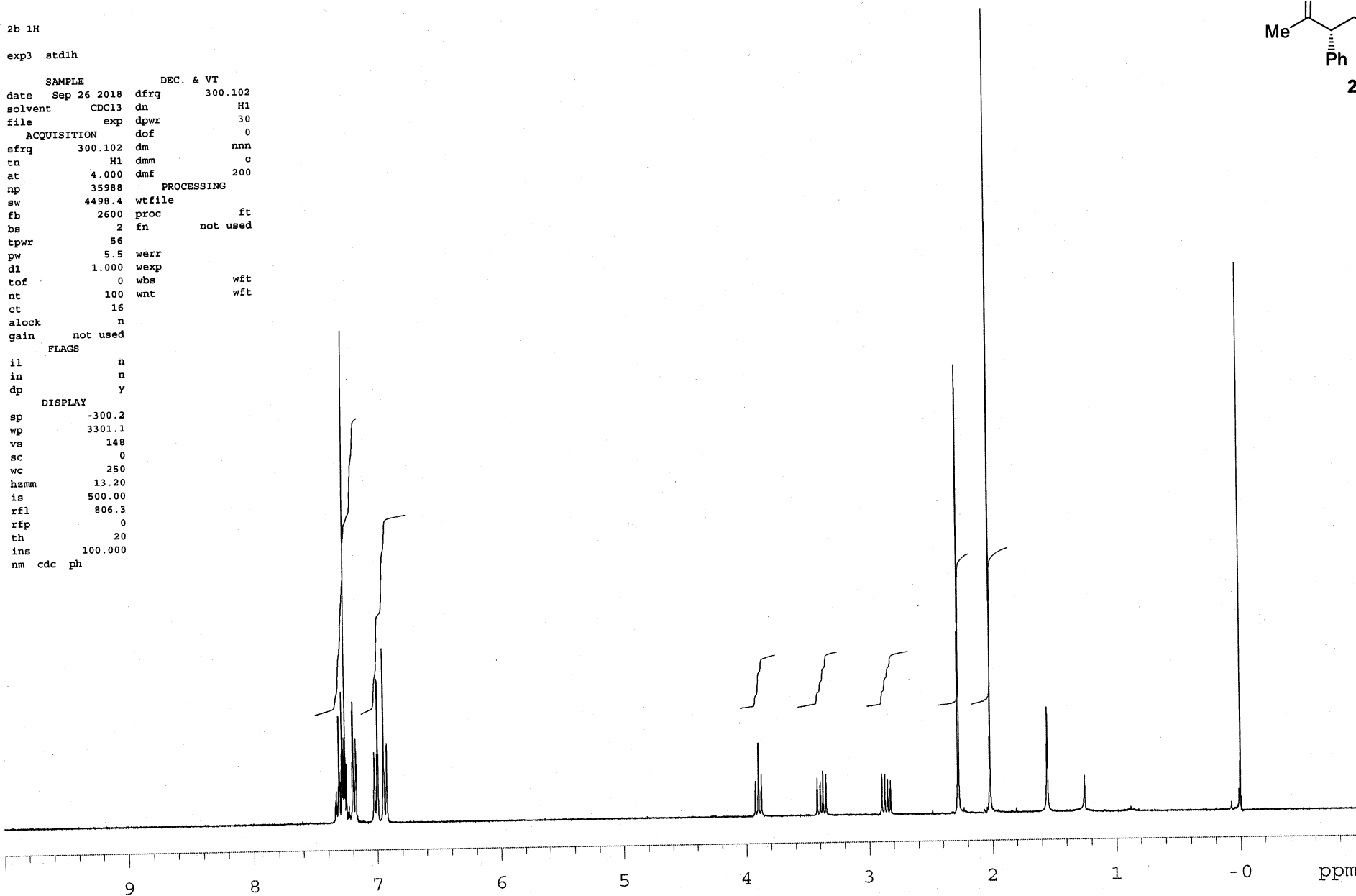
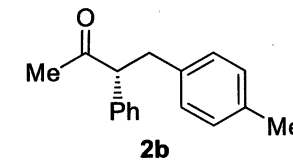
SAMPLE		DEC. & VT	
date	Aug 31 2016	dfrq	299.959
solvent	CDCl3	dn	H1
file	exp	dpwr	43
ACQUISITION		dof	
sfrq	75.432	dm	YYY
tn	C13	dmm	w
at	1.815	dmf	10000
np	68106	PROCESSING	
sw	18761.7	lb	1.00
fb	10400	wtfile	
bs	4	proc	ft
tpwr	53	fn	not used
pw	8.7		
d1	0	werr	
tof	0	wexp	
nt	100000	wbs	wft
ct	1564	wnt	wft
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	Y		
DISPLAY			
sp	-1851.9		
wp	18761.7		
vs	129		
sc	0		
wc	250		
hzmm	75.05		
is	500.00		
rfl	7659.6		
rfp	5807.7		
th	4		
ins	100.000		
nm	no ph		



2b 1H

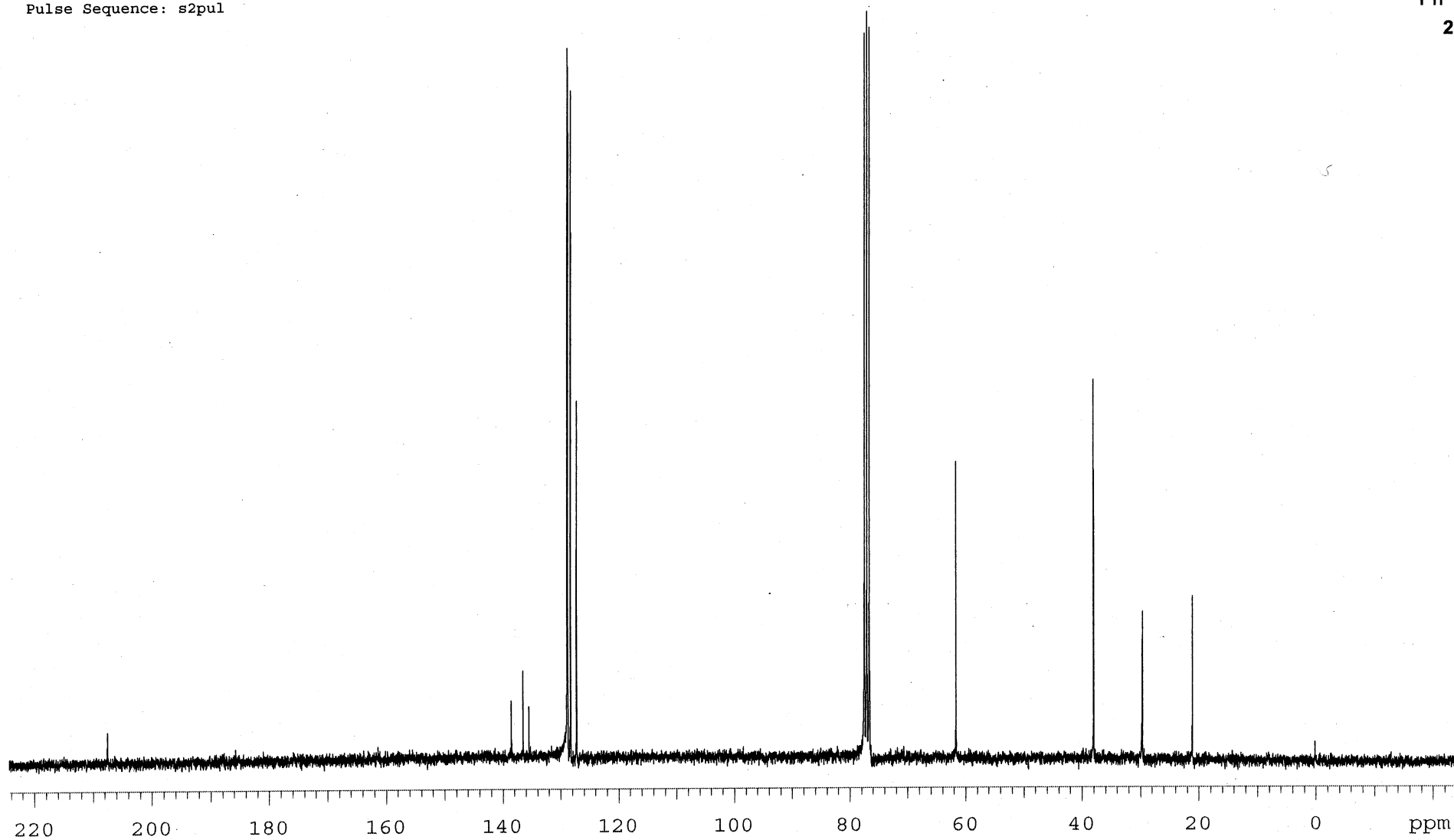
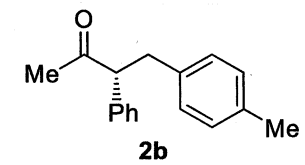
exp3 stdlh

SAMPLE		DEC. & VT	
date	Sep 26 2018	dfrq	300.102
solvent	CDCl3	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	
sfrq	300.102	dm	nnn
tn	H1	dmm	c
at	4.000	dmf	200
np	35988	PROCESSING	
sw	4498.4	wtfile	
fb	2600	proc	ft
bs	2	fn	not used
tpwr	56		
pw	5.5	werr	
d1	1.000	wexp	
tof	0	wbs	wft
nt	100	wnt	wft
ct	16		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	Y		
DISPLAY			
sp	-300.2		
wp	3301.1		
vs	148		
sc	0		
wc	250		
hzmm	13.20		
is	500.00		
rfl	806.3		
rfp	0		
th	20		
ins	100.000		
nm	cdc ph		



TN A-40 13C

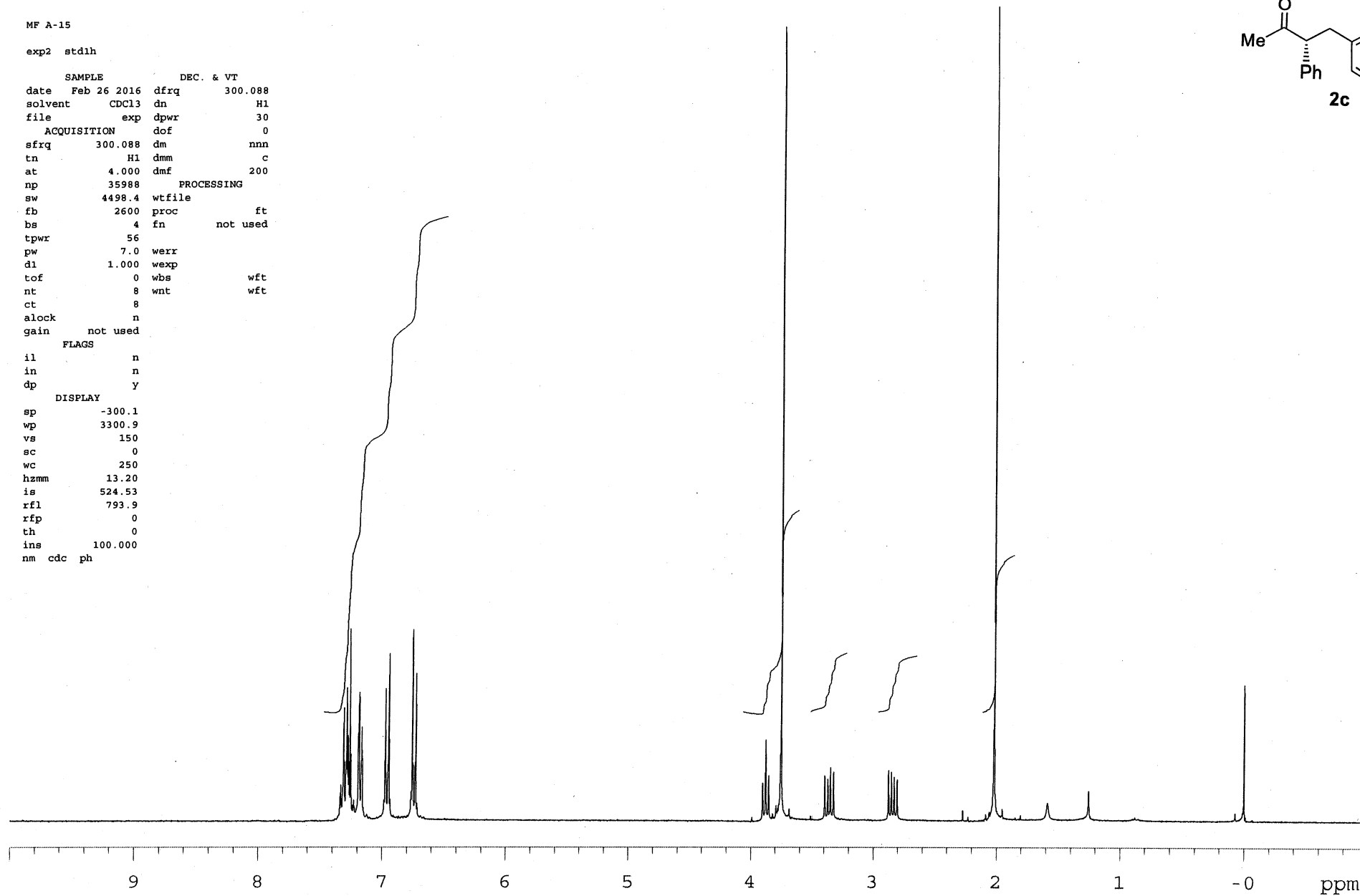
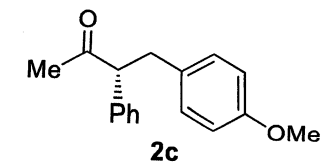
Pulse Sequence: s2pul



MF A-15

exp2 std1h

SAMPLE		DEC. & VT	
date	Feb 26 2016	dfrq	300.088
solvent	CDCl3	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	0
sfrq	300.088	dm	nnn
tn	H1	dmm	c
at	4.000	dmf	200
np	35988	PROCESSING	
sw	4498.4	wtfile	
fb	2600	proc	ft
bs	4	fn	not used
tpwr	56		
pw	7.0	werr	
d1	1.000	wexp	
tof	0	wbs	wft
nt	8	wnt	wft
ct	8		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	-300.1		
wp	3300.9		
vs	150		
sc	0		
wc	250		
hzmm	13.20		
is	524.53		
rfl	793.9		
rfp	0		
th	0		
ins	100.000		
nm	cdc ph		



MF A-15 C13

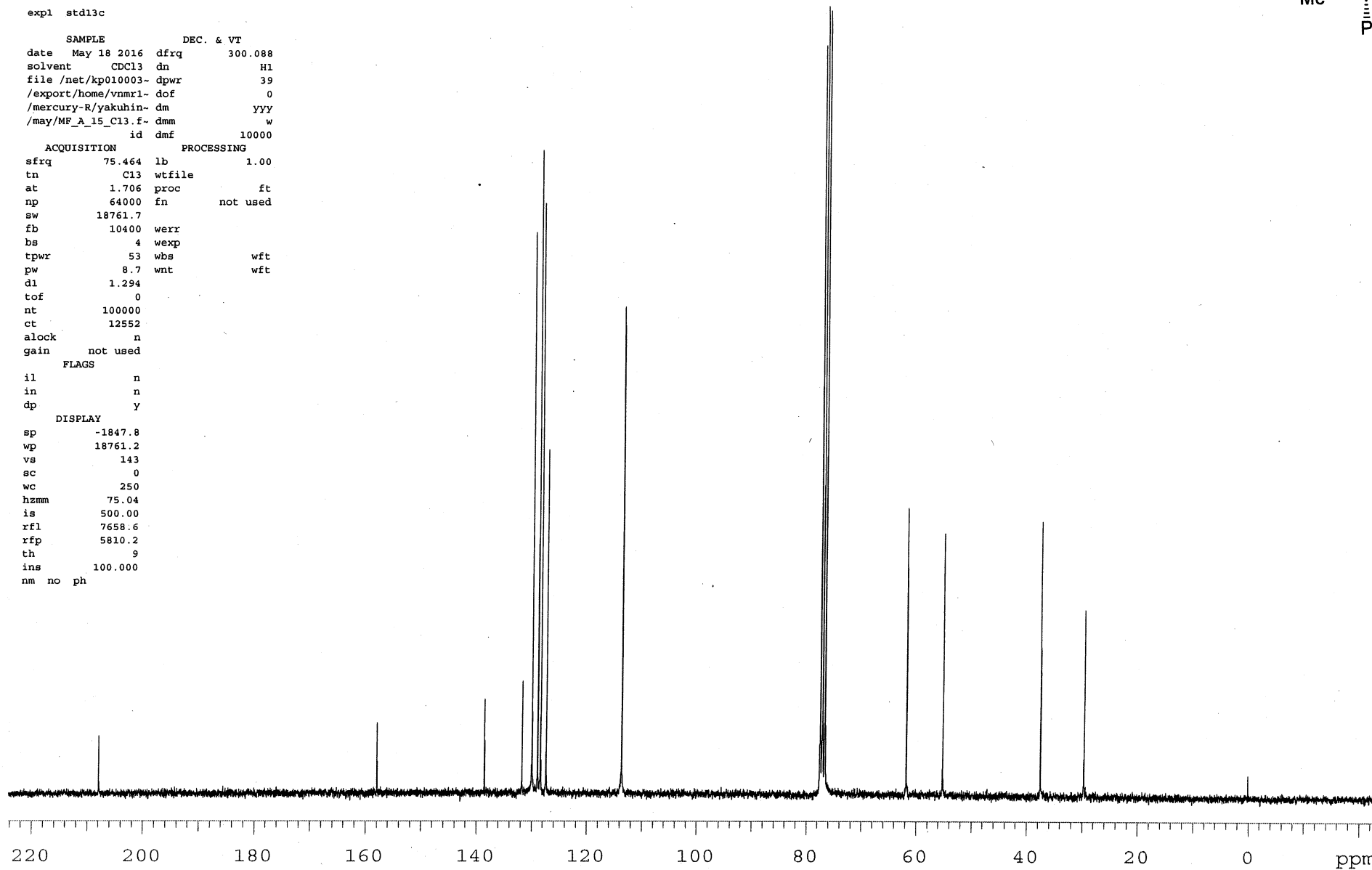
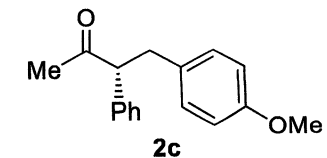
expl std13c

SAMPLE DEC. & VT
date May 18 2016 dfrq 300.088
solvent CDCl3 dn H1
file /net/kp010003- dpwr 39
/export/home/vnmr1- dof 0
/mercury-R/yakuhin- dm yyy
/may/MF_A_15_C13.f- dmm w
id dmf 10000

ACQUISITION PROCESSING
sfrq 75.464 lb 1.00
tn C13 wfile
at 1.706 proc ft
np 64000 fn not used
sw 18761.7
fb 10400 werr
bs 4 wexp
tpwr 53 wbs wft
pw 8.7 wnt wft
dl 1.294
tof 0
nt 100000
ct 12552
alock n
gain not used

FLAGS
il n
in n
dp Y

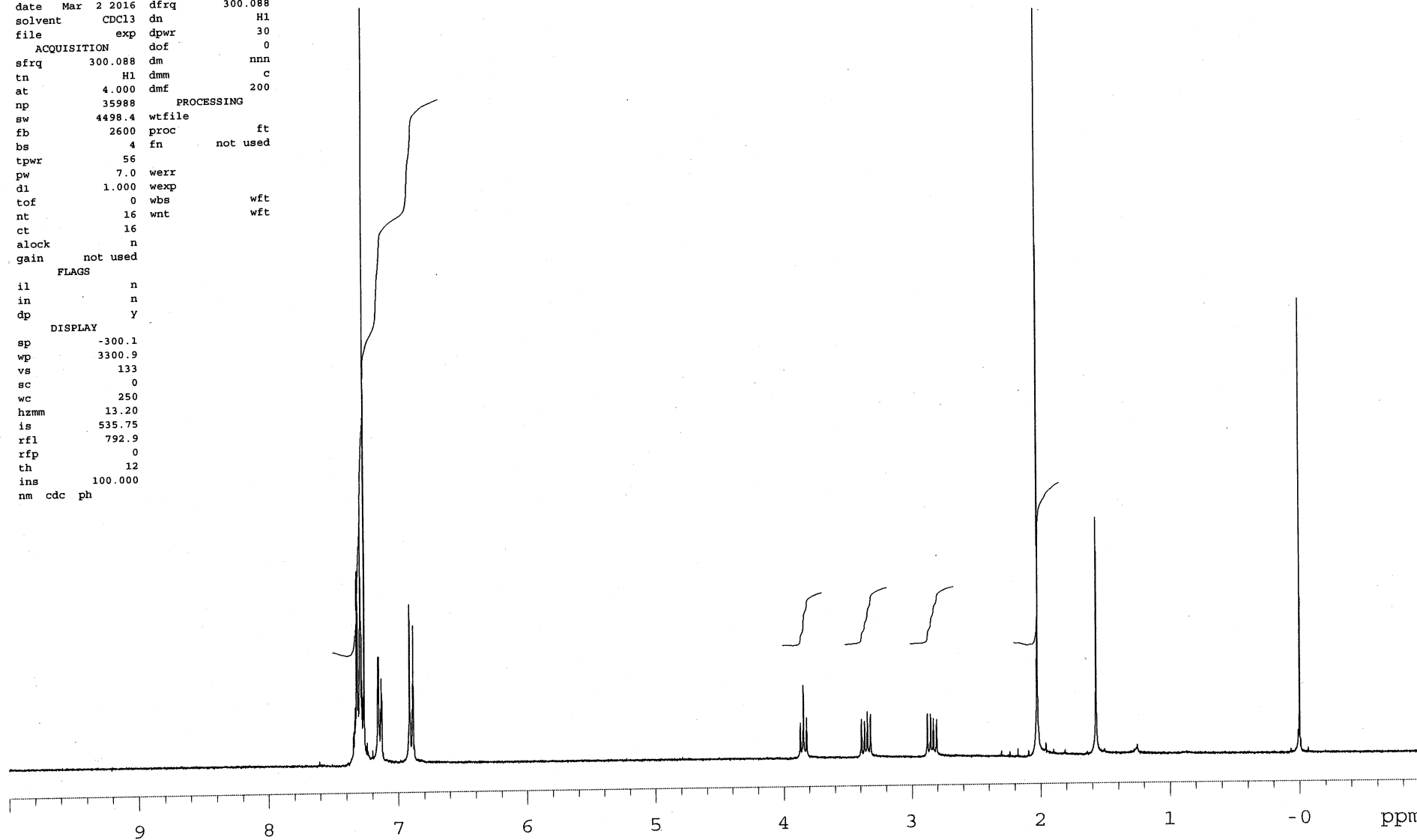
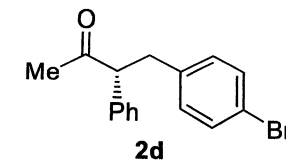
DISPLAY
sp -1847.8
wp 18761.2
vs 143
sc 0
wc 250
hzmm 75.04
is 500.00
rfl 7658.6
rfp 5810.2
th 9
ins 100.000
nm no ph



KO A-18 PTLCL1

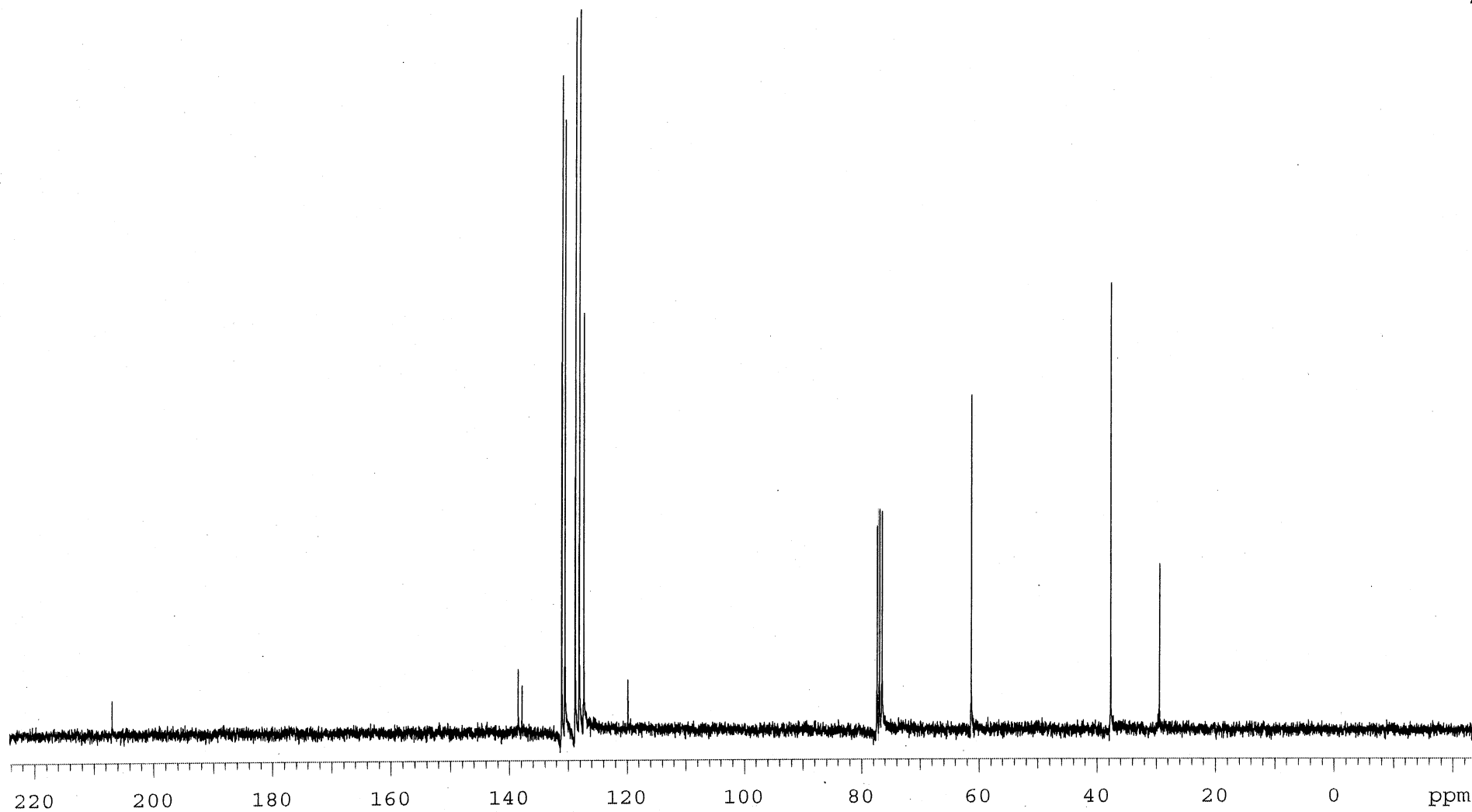
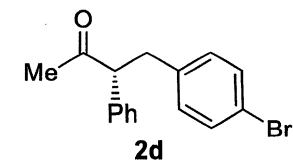
expl stdlh

SAMPLE		DEC. & VT	
date	Mar 2 2016	dfrq	300.088
solvent	CDCl3	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	
sfrq	300.088	dm	nnn
tn	H1	dmm	c
at	4.000	dmf	200
np	35988	PROCESSING	
sw	4498.4	wtfile	
fb	2600	proc	ft
bs	4	fn	not used
tpwr	56		
pw	7.0	werr	
d1	1.000	wexp	
tof	0	wbs	wft
nt	16	wnt	wft
ct	16		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	-300.1		
wp	3300.9		
vs	133		
sc	0		
wc	250		
hzmm	13.20		
is	535.75		
rfl	792.9		
rfp	0		
th	12		
ins	100.000		
nm	cdc ph		



KO-A-18 PTLCl

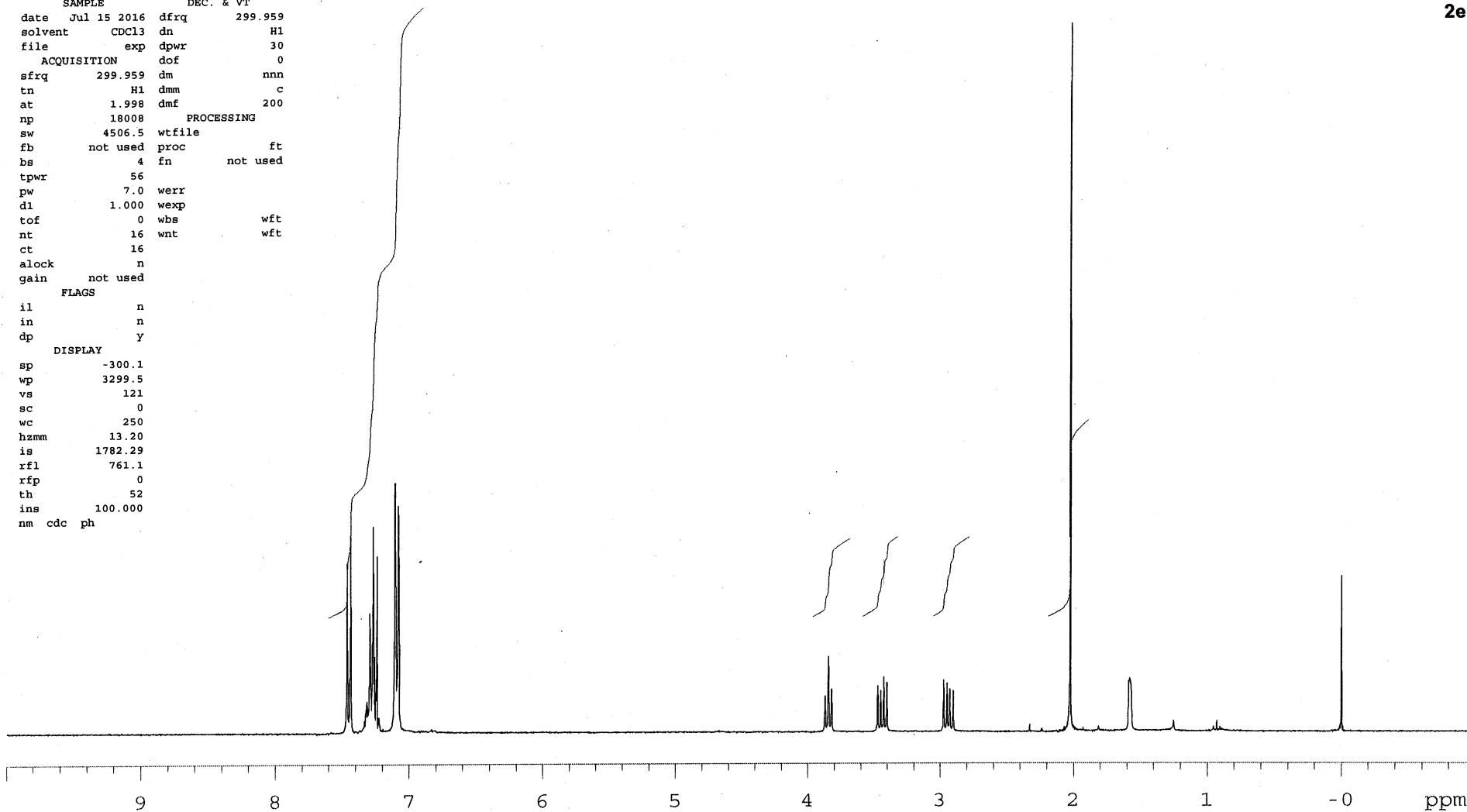
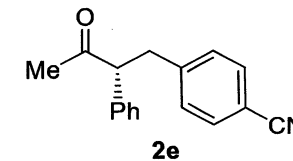
Pulse Sequence: s2pul



MF D-59 PTLC2

expl stdlh

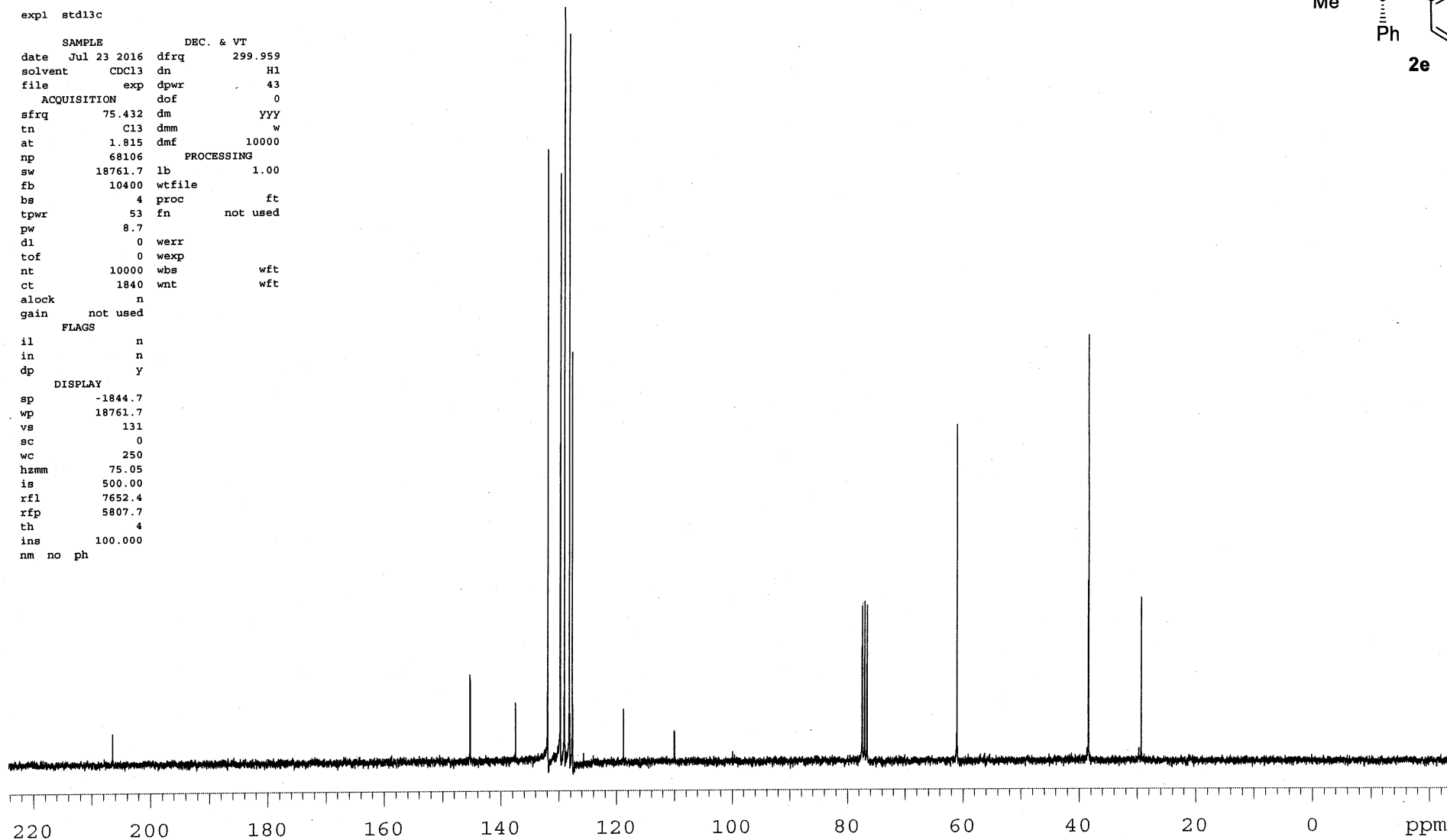
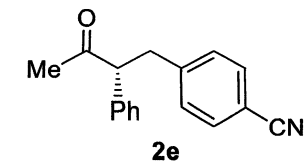
SAMPLE		DEC. & VT	
date	Jul 15 2016	dfrq	299.959
solvent	CDCl3	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	
sfrq	299.959	dm	nnn
tn	H1	dmm	c
at	1.998	dmf	200
np	18008	PROCESSING	
sw	4506.5	wtfile	
fb	not used	proc	ft
bs	4	fn	not used
tpwr	56		
pw	7.0	werr	
dl	1.000	wexp	
tof	0	wbs	wft
nt	16	wnt	wft
ct	16		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	Y		
DISPLAY			
sp	-300.1		
wp	3299.5		
vs	121		
sc	0		
wc	250		
hzmm	13.20		
is	1782.29		
rfl	761.1		
rfp	0		
th	52		
ins	100.000		
nm	cdc ph		



MF-D-59 PTLC2 13C

expl std13c

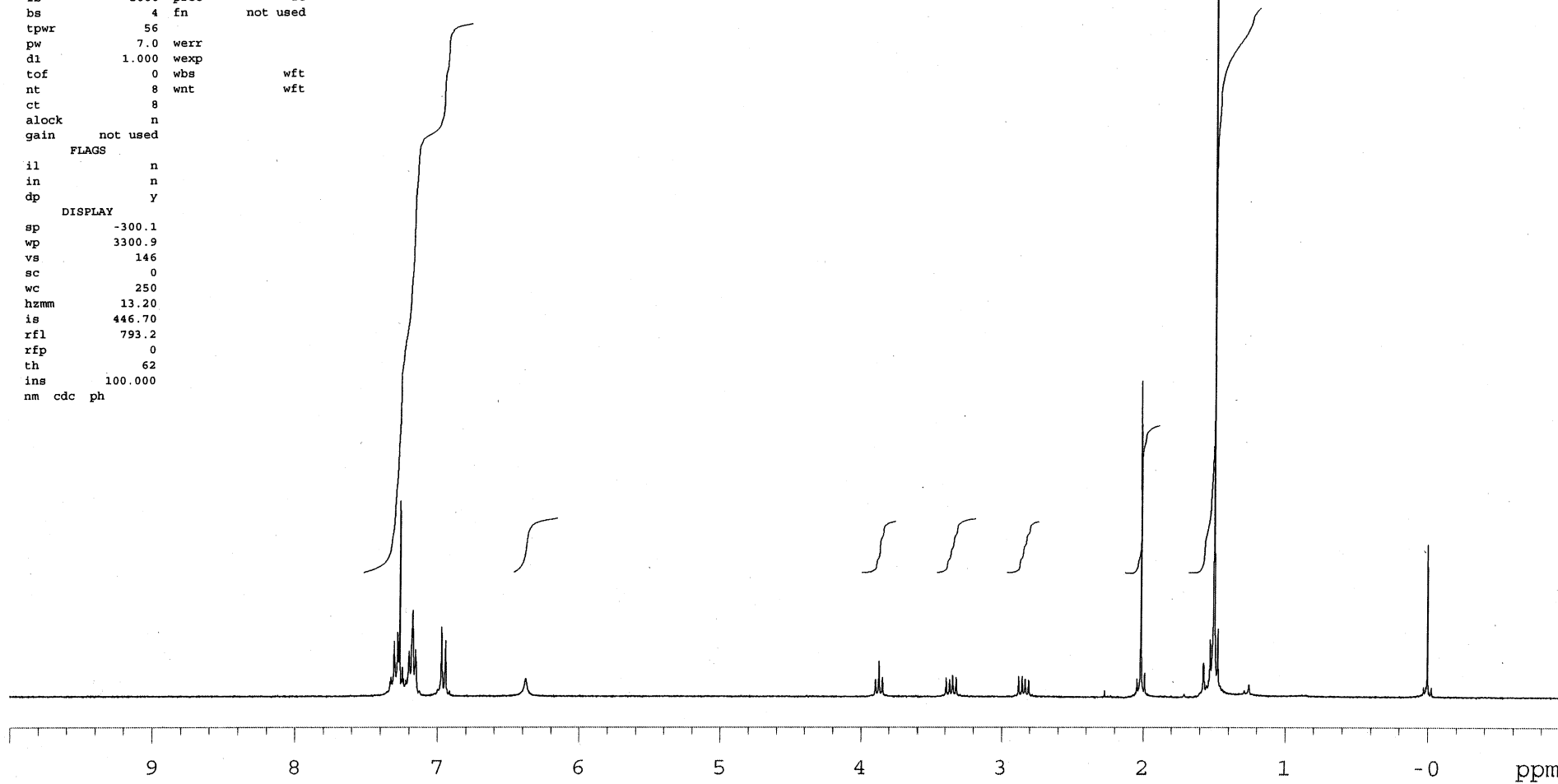
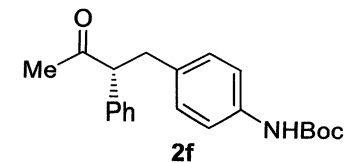
```
SAMPLE          DEC. & VT
date Jul 23 2016 dfrq      299.959
solvent CDCl3  dn         H1
file      exp      dpwr      43
ACQUISITION    dof         0
sfrq      75.432  dm         YY
tn         C13      dmm        w
at         1.815  dmf        10000
np         68106  PROCESSING
sw      18761.7  lb         1.00
fb      10400   wtfile
bs         4     proc         ft
tpwr      53   fn         not used
pw         8.7
d1         0     werr
tof         0     wexp
nt      10000   wbs         wft
ct      1840   wnt         wft
alock      n
gain      not used
FLAGS
il         n
in         n
dp         Y
DISPLAY
sp      -1844.7
wp      18761.7
vs      131
sc         0
wc      250
hzmm     75.05
is      500.00
rfl     7652.4
rfp     5807.7
th         4
ins     100.000
nm no ph
```



MM A-37

exp2 std1h

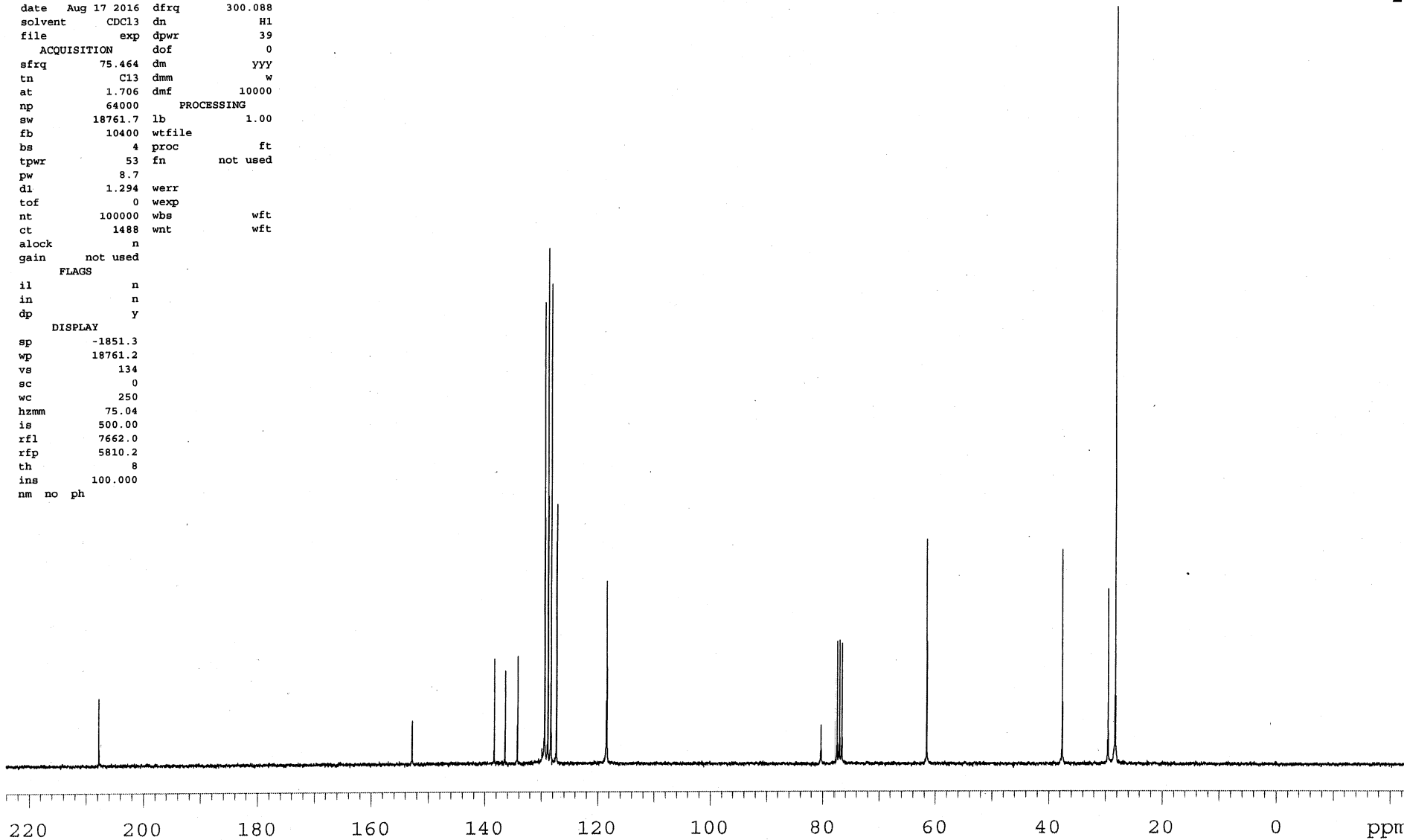
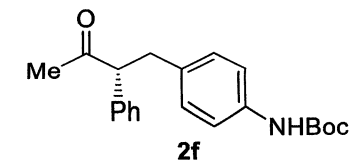
```
SAMPLE          DEC. & VT
date Feb 26 2016 dfrq      300.088
solvent CDCl3  dn         H1
file      exp  dpwr       30
ACQUISITION     dof       0
sfrq  300.088  dm         nnn
tn     H1  dmm          c
at     4.000  dmf       200
np     35988  PROCESSING
sw     4498.4  wfile
fb     2600  proc         ft
bs     4      fn         not used
tpwr   56
pw     7.0  werr
dl     1.000  wexp
tof     0    wbs         wft
nt     8     wnt         wft
ct     8
alock   n
gain   not used
FLAGS
il     n
in     n
dp     Y
DISPLAY
sp     -300.1
wp     3300.9
vs     146
sc     0
wc     250
hzmm   13.20
is     446.70
rfl    793.2
rfp    0
th     62
ins    100.000
nm  cdc ph
```



MM A-37 13C

exp2 std13c

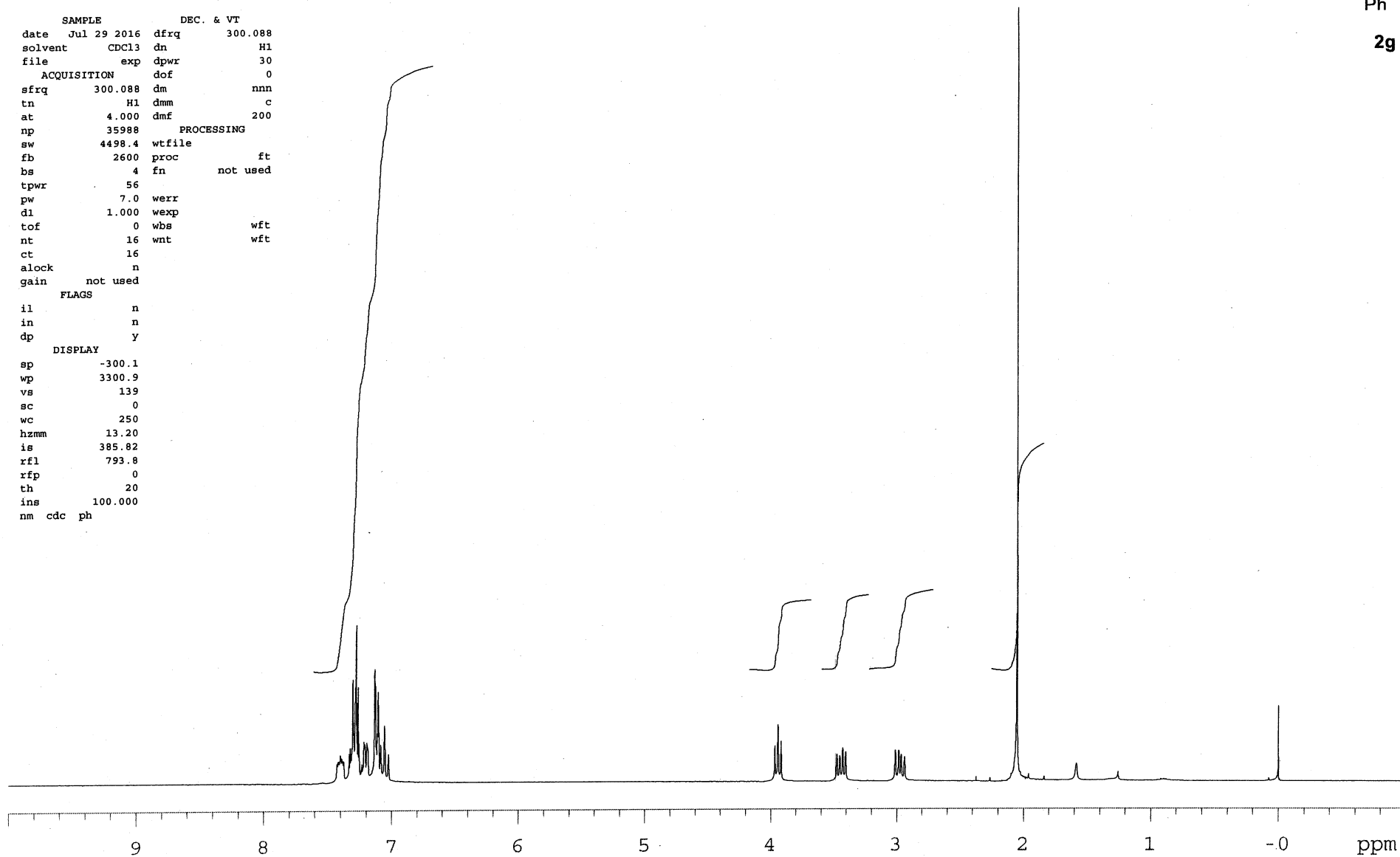
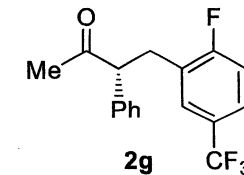
```
SAMPLE          DEC. & VT
date Aug 17 2016 dfrq          300.088
solvent CDCl3  dn              H1
file      exp  dpwr           39
ACQUISITION    dof            0
sfrq       75.464 dm          YYY
tn         C13 dmm            w
at         1.706 dmf          10000
np         64000 PROCESSING
sw         18761.7 lb          1.00
fb         10400 wtfile
bs         4   proc           ft
tpwr      53   fn            not used
pw         8.7
dl         1.294 werr
tof        0   wexp
nt         100000 wbs          wft
ct         1488 wnt           wft
alock      n
gain       not used
FLAGS
il         n
in         n
dp         y
DISPLAY
sp         -1851.3
wp         18761.2
vs         134
sc         0
wc         250
hzmm      75.04
is         500.00
rfl       7662.0
rfp       5810.2
th         8
ins       100.000
nm no ph
```

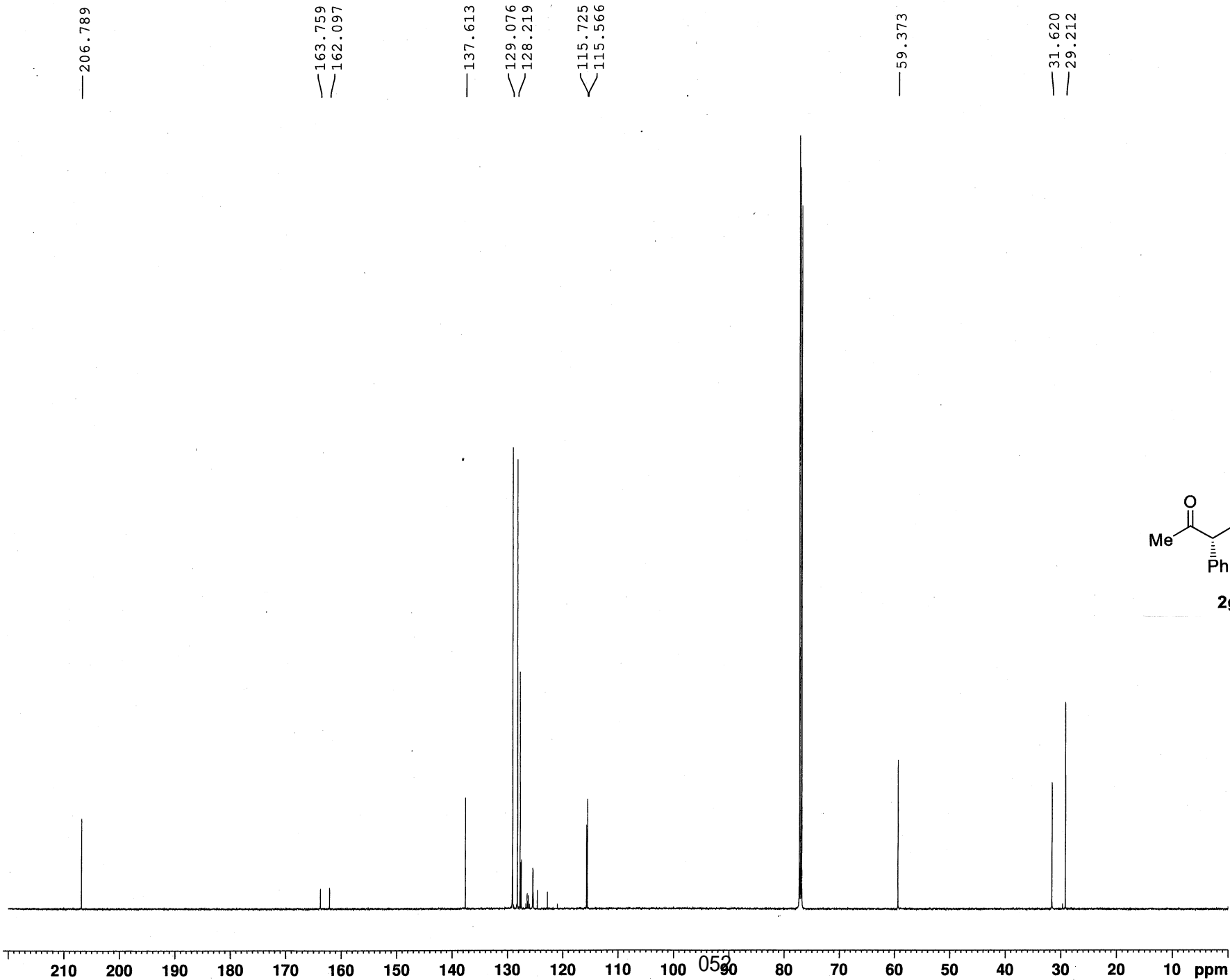


MF E-18 PTLCL

exp2 stdlh

SAMPLE		DEC. & VT	
date	Jul 29 2016	dfrq	300.088
solvent	CDCl3	dn	H1
file	exp	dpwr	30
ACQUISITION		PROCESSING	
sfrq	300.088	dof	0
tn	H1	dm	nnn
at	4.000	dmm	c
np	35988	dmf	200
sw	4498.4	wtfile	
fb	2600	proc	ft
bs	4	fn	not used
tpwr	56		
pw	7.0	werr	
d1	1.000	wexp	
tof	0	wbs	wft
nt	16	wnt	wft
ct	16		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	-300.1		
wp	3300.9		
vs	139		
sc	0		
wc	250		
hzmm	13.20		
is	385.82		
rfl	793.8		
rfp	0		
th	20		
ins	100.000		
nm	cdc ph		

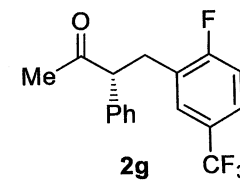




Current Data Parameters
 NAME KPNN-5613
 EXPNO 20
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20170803
 Time 14.22 h
 INSTRUM spect
 PROBHD Z114607_0202 (
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 3208
 DS 4
 SWH 36231.883 Hz
 FIDRES 1.105709 Hz
 AQ 0.9043968 sec
 RG 194.13
 DW 13.800 usec
 DE 6.50 usec
 TE 296.4 K
 D1 2.0000000 sec
 D11 0.0300000 sec
 TD0 300
 SFO1 151.0184889 MHz
 NUC1 13C
 P1 12.00 usec
 PLW1 80.0000000 W
 SFO2 600.5336032 MHz
 NUC2 1H
 CPDPRG[2] waltz65
 PCPD2 80.00 usec
 PLW2 24.00600052 W
 PLW12 0.37509000 W
 PLW13 0.16671000 W

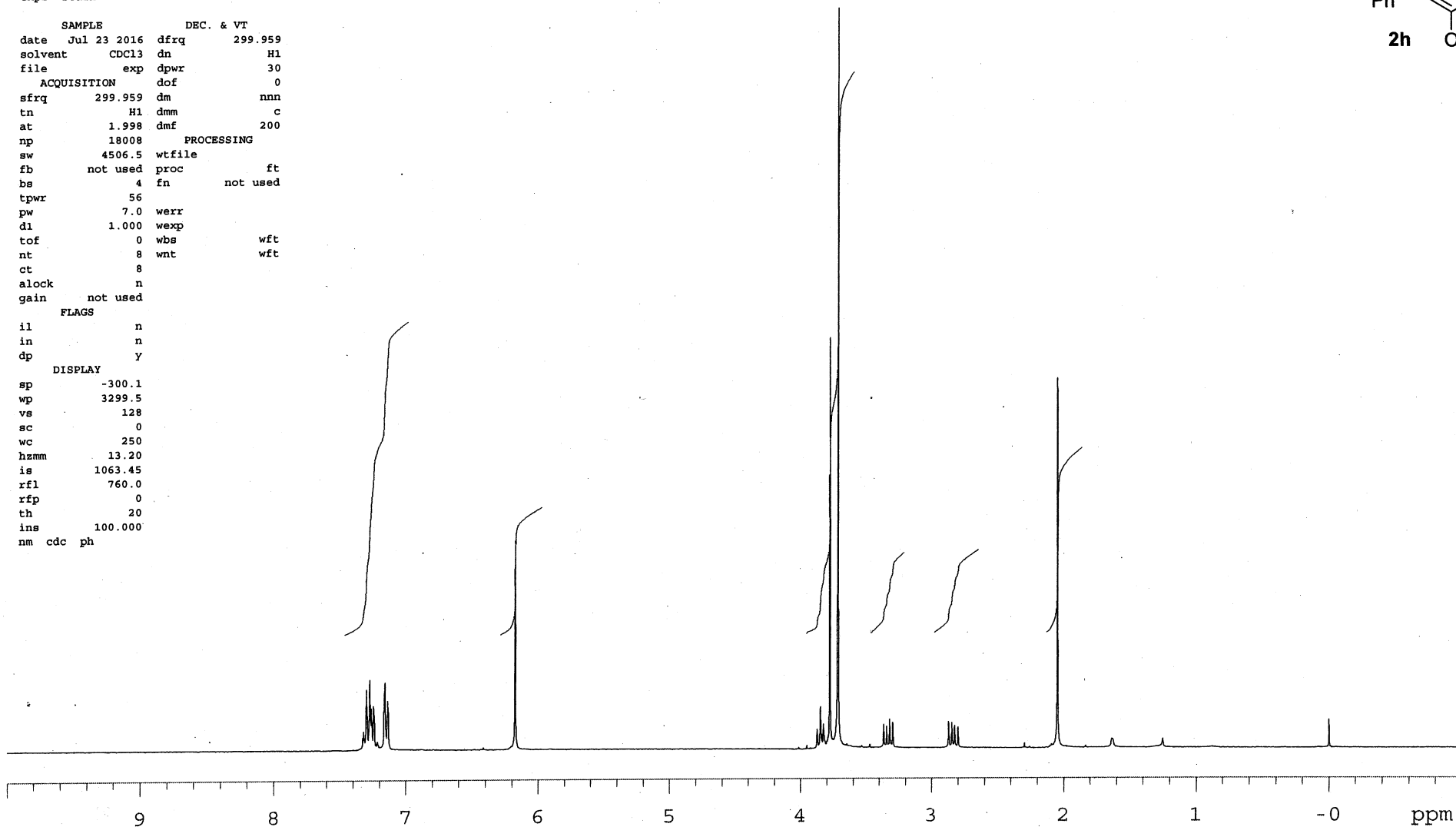
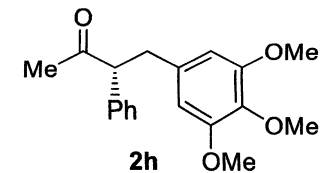
F2 - Processing parameters
 SI 32768
 SF 151.0033883 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



MF E-11 PTLCl

exp2 std1h

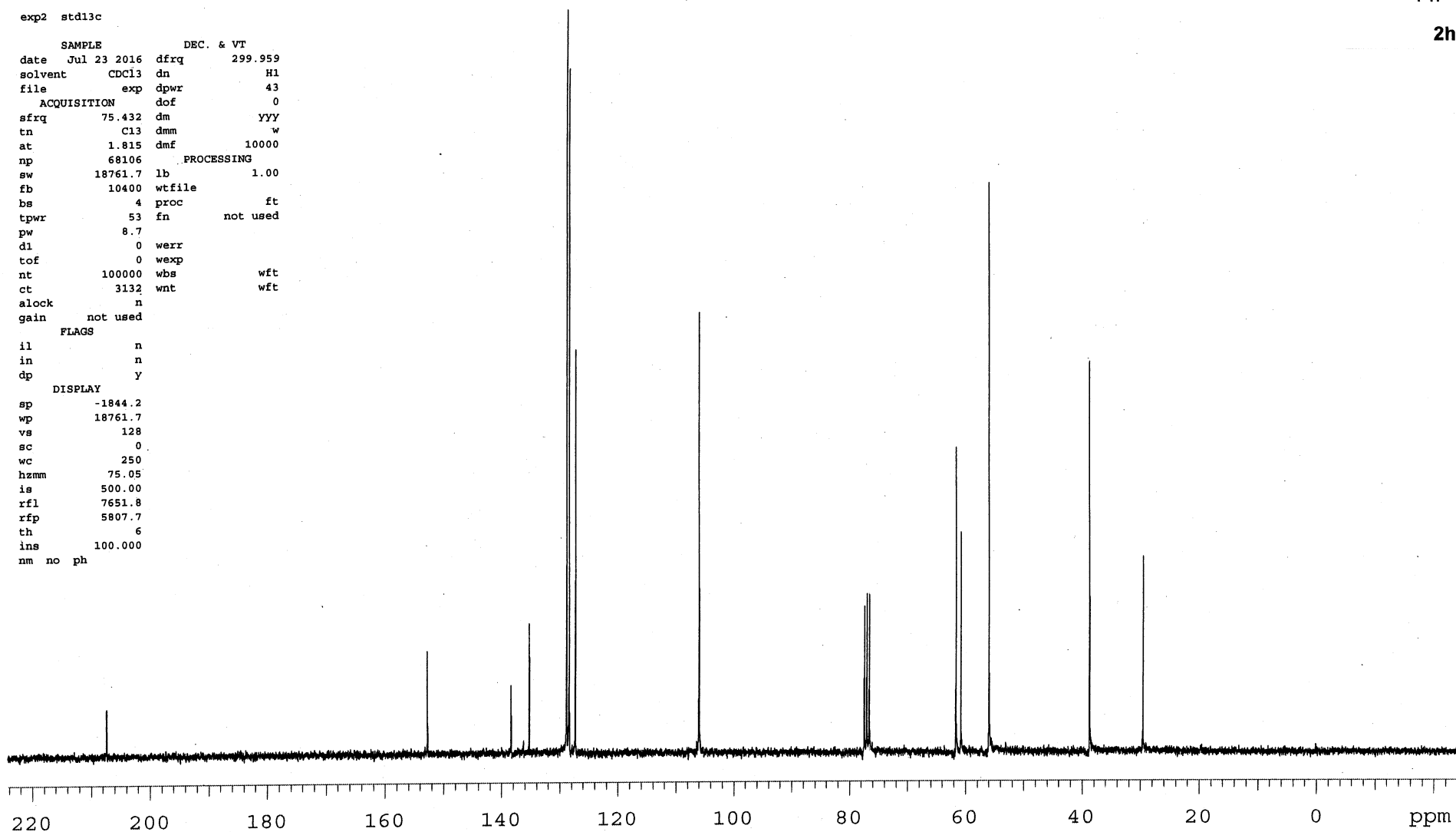
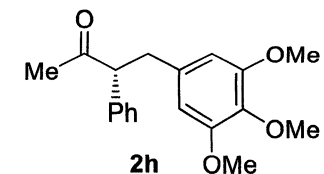
SAMPLE		DEC. & VT	
date	Jul 23 2016	dfrq	299.959
solvent	CDCl3	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	
sfrq	299.959	dm	nnn
tn	H1	dmm	c
at	1.998	dmf	200
np	18008	PROCESSING	
sw	4506.5	wtfile	
fb	not used	proc	ft
bs	4	fn	not used
tpwr	56		
pw	7.0	werr	
dl	1.000	wexp	
tof	0	wbs	wft
nt	8	wnt	wft
ct	8		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	-300.1		
wp	3299.5		
vs	128		
sc	0		
wc	250		
hzmm	13.20		
is	1063.45		
rfl	760.0		
rfp	0		
th	20		
ins	100.000		
nm	cdc ph		



13C OBSERVE

exp2 std13c

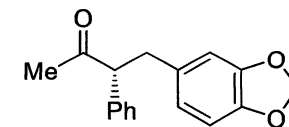
```
SAMPLE          DEC. & VT
date Jul 23 2016 dfrq      299.959
solvent CDCl3  dn         H1
file      exp  dpwr       43
ACQUISITION    dof       0
sfrq      75.432 dm       YYY
tn         C13 dmm        w
at         1.815 dmf      10000
np         68106          PROCESSING
sw      18761.7 lb        1.00
fb      10400 wtfile
bs         4 proc        ft
tpwr      53 fn         not used
pw         8.7
dl         0 werr
tof         0 wexp
nt      100000 wbs        wft
ct         3132 wnt        wft
alock      n
gain      not used
FLAGS
il         n
in         n
dp         y
DISPLAY
sp      -1844.2
wp      18761.7
vs       128
sc        0
wc       250
hzmm     75.05
is       500.00
rfl     7651.8
rfp     5807.7
th        6
ins     100.000
nm no ph
```



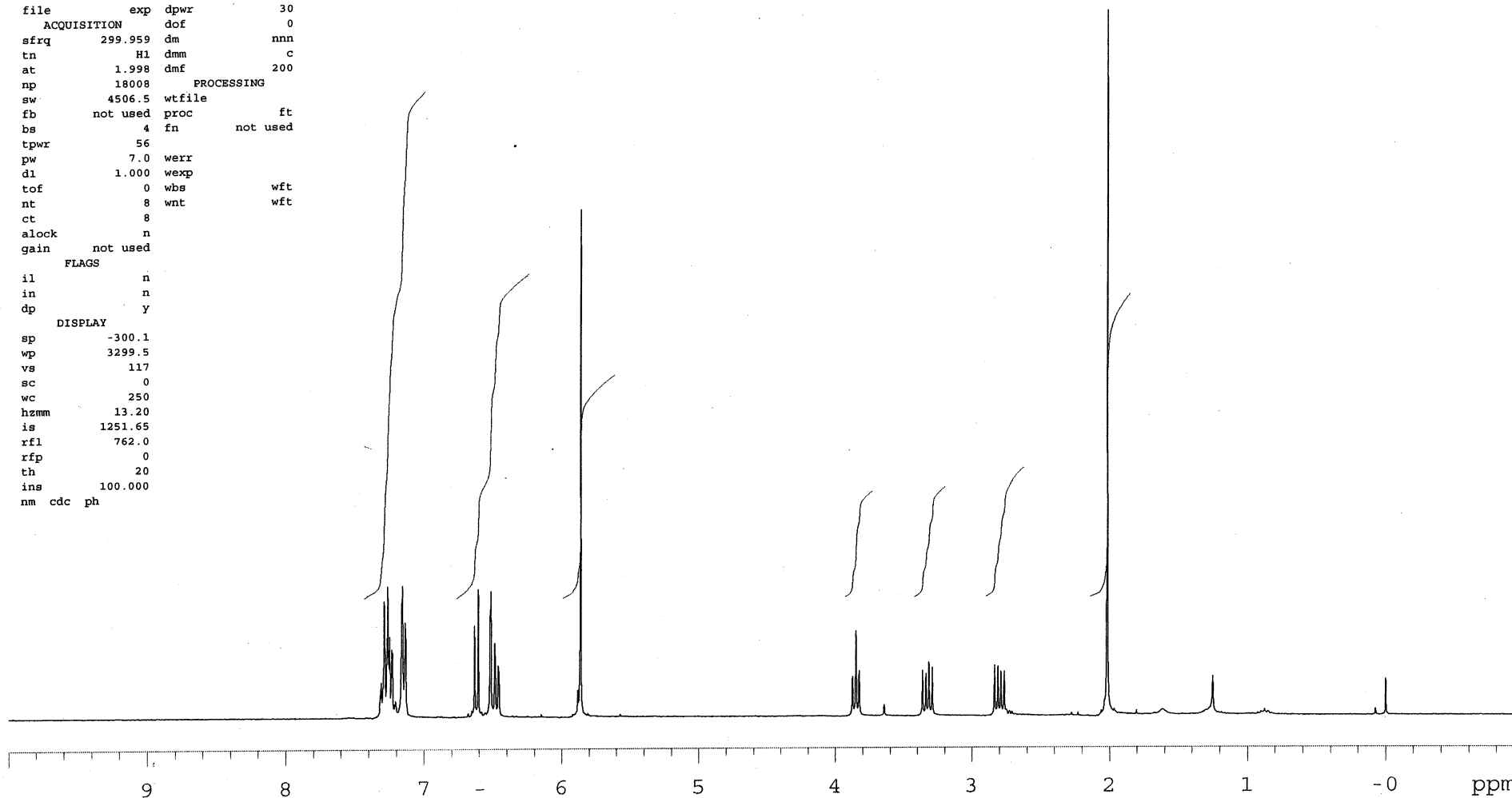
MF E-58 PTLC1

exp2 std1h

```
SAMPLE          DEC. & VT
date Dec 14 2016 dfrq      299.959
solvent CDCl3  dn         H1
file      exp  dpwr       30
ACQUISITION    dof        0
sfrq  299.959 dm         nnn
tn      H1  dmm         c
at      1.998 dmf       200
np      18008 PROCESSING
sw      4506.5 wtfile
fb      not used proc      ft
bs      4      fn      not used
tpwr    56
pw      7.0  werr
d1      1.000 wexp
tof     0    wbs
nt      8    wnt      wft
ct      8
alock   n
gain    not used
FLAGS
il      n
in      n
dp      Y
DISPLAY
sp      -300.1
wp      3299.5
vs      117
sc      0
wc      250
hzmm    13.20
is      1251.65
rfl     762.0
rfp     0
th      20
ins     100.000
nm cdc ph
```

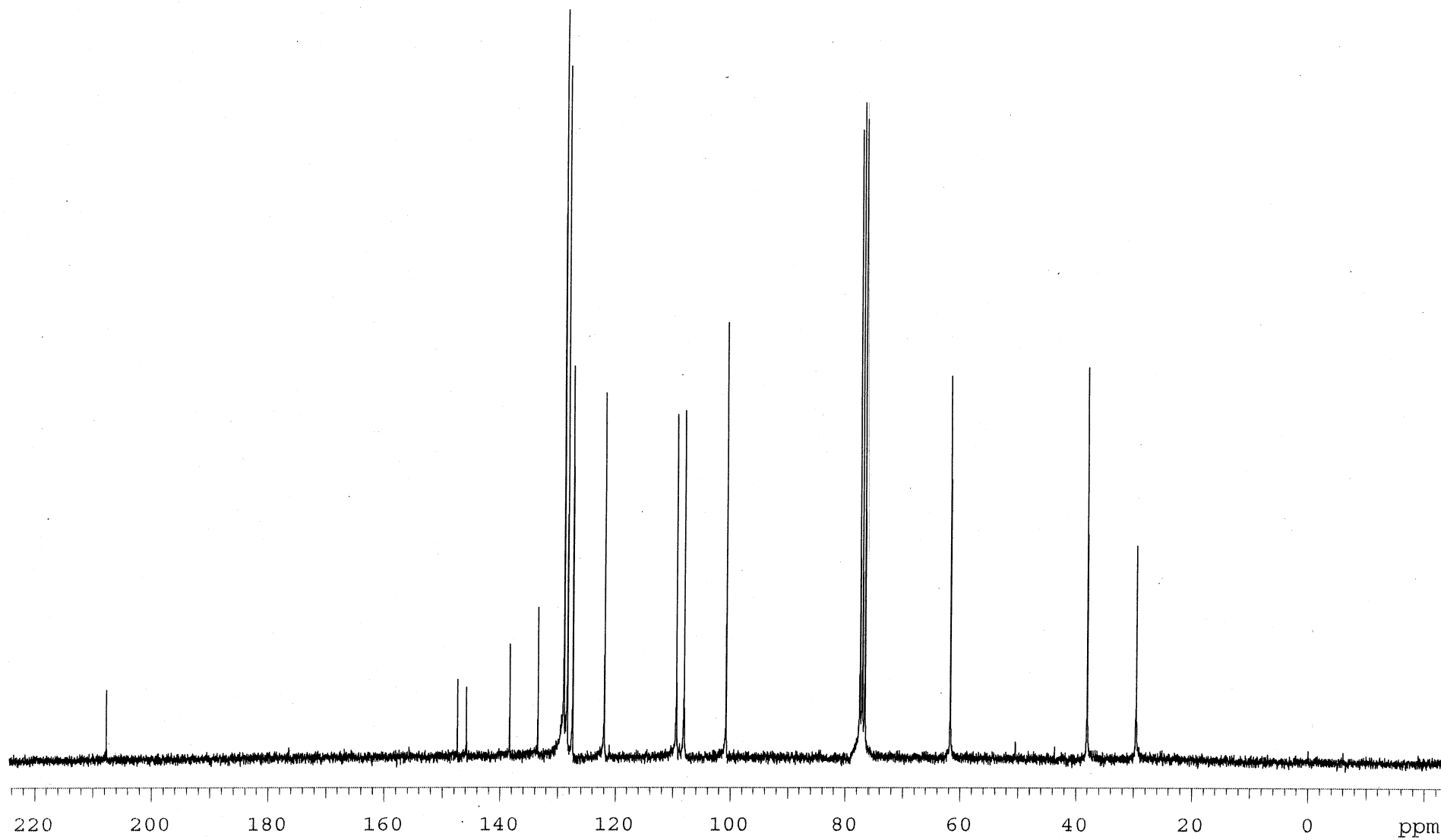
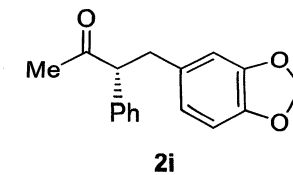


2i



MF E-58 PTLCl

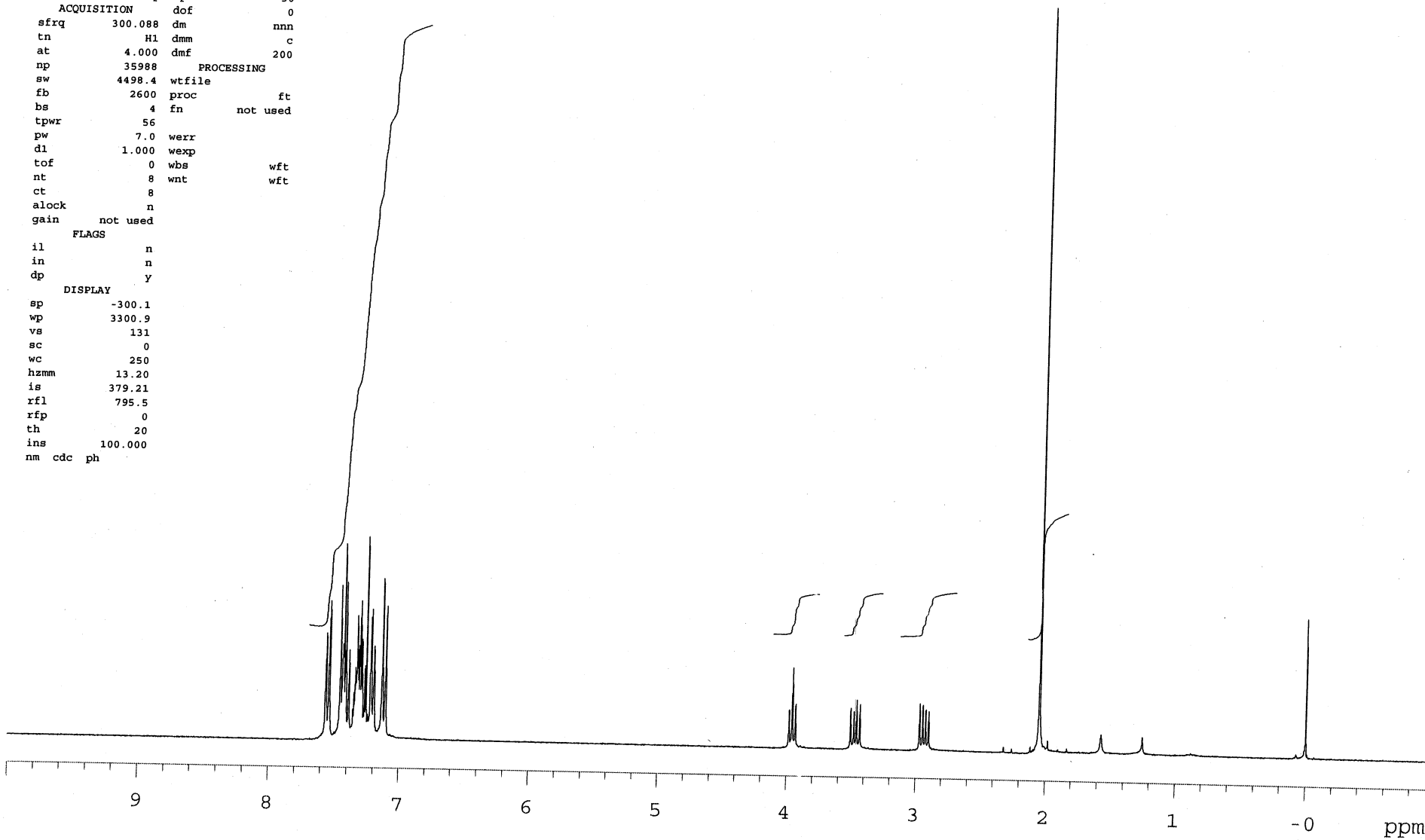
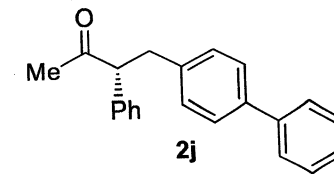
Pulse Sequence: s2pul



MF A-36 PTLC1

exp2 stdlh

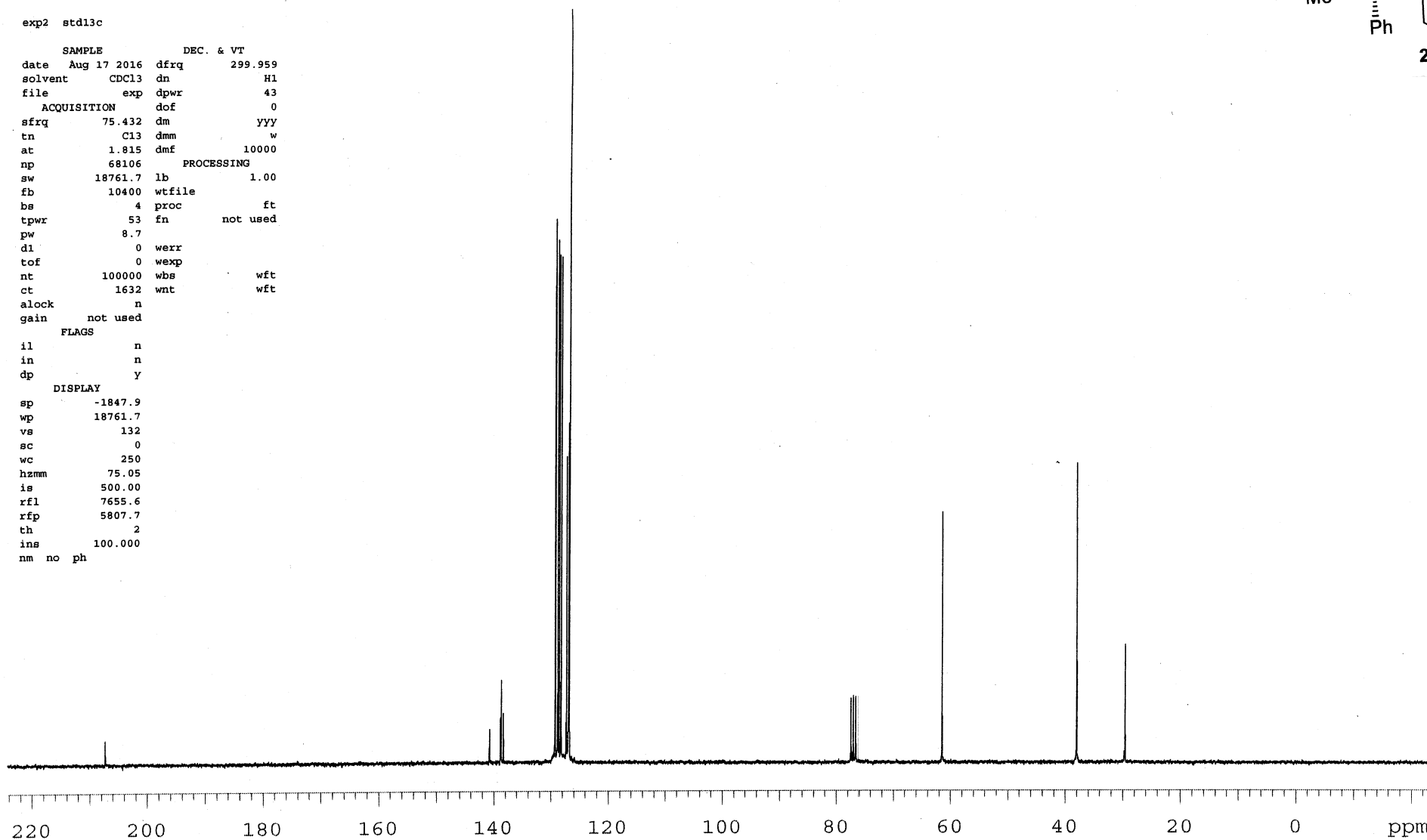
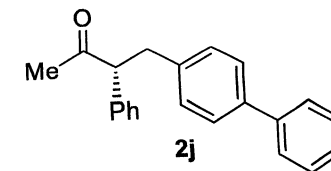
```
SAMPLE          DEC. & VT
date Feb 26 2016 dfrq      300.088
solvent CDCl3  dn         H1
file      exp  dpwr       30
ACQUISITION    dof       0
sfrq      300.088 dm      nnn
tn         H1  dmm        c
at         4.000 dmf      200
np      35988  PROCESSING
sw      4498.4 wtfile
fb      2600  proc       ft
bs         4  fn      not used
tpwr      56
pw         7.0 wexr
dl         1.000 wexp
tof         0  wbs      wft
nt         8  wnt      wft
ct         8
alock      n
gain      not used
FLAGS
il         n
in         n
dp         y
DISPLAY
sp      -300.1
wp      3300.9
vs      131
sc       0
wc      250
hzmm    13.20
is      379.21
rfl     795.5
rfp      0
th       20
ins     100.000
nm cdc ph
```



MF A-36 13C

exp2 std13c

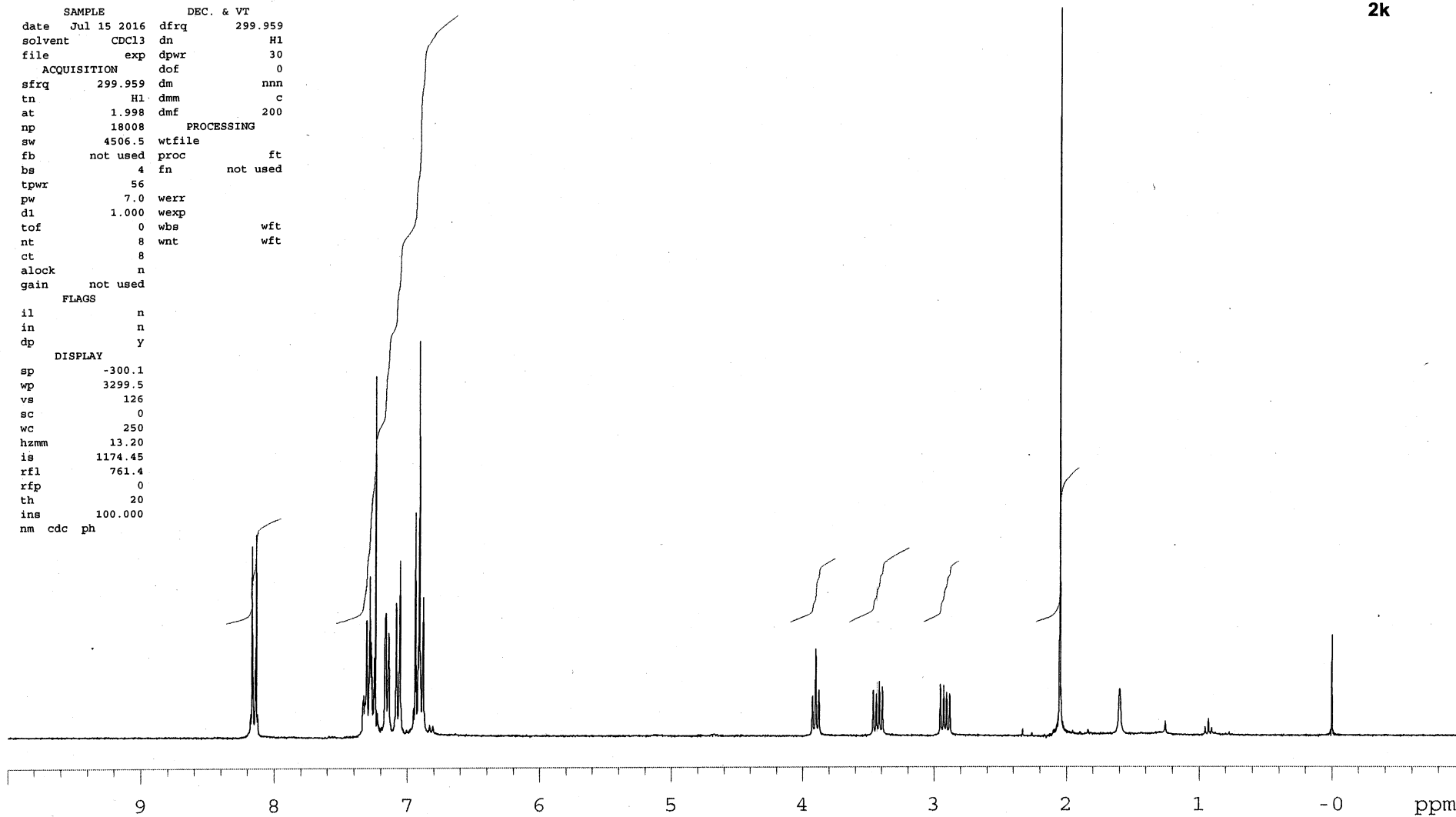
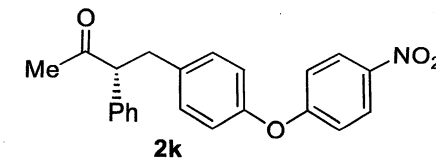
```
SAMPLE          DEC. & VT
date Aug 17 2016 dfrq      299.959
solvent CDCl3  dn          H1
file      exp  dpwr        43
ACQUISITION  dof          0
sfrq      75.432 dm         YYY
tn         C13 dmm         w
at         1.815 dmf       10000
np         68106 PROCESSING
sw         18761.7 lb        1.00
fb         10400 wtfile
bs         4   proc         ft
tpwr       53   fn         not used
pw         8.7
dl         0   weyr
tof        0   wexp
nt         100000 wbs       wft
ct         1632 wnt       wft
alock      n
gain       not used
FLAGS
il         n
in         n
dp         Y
DISPLAY
sp        -1847.9
wp        18761.7
vs         132
sc         0
wc         250
hzmm      75.05
is        500.00
rfl       7655.6
rfp       5807.7
th         2
ins       100.000
nm no ph
```



MF E-9 PTLCL1

exp2 std1h

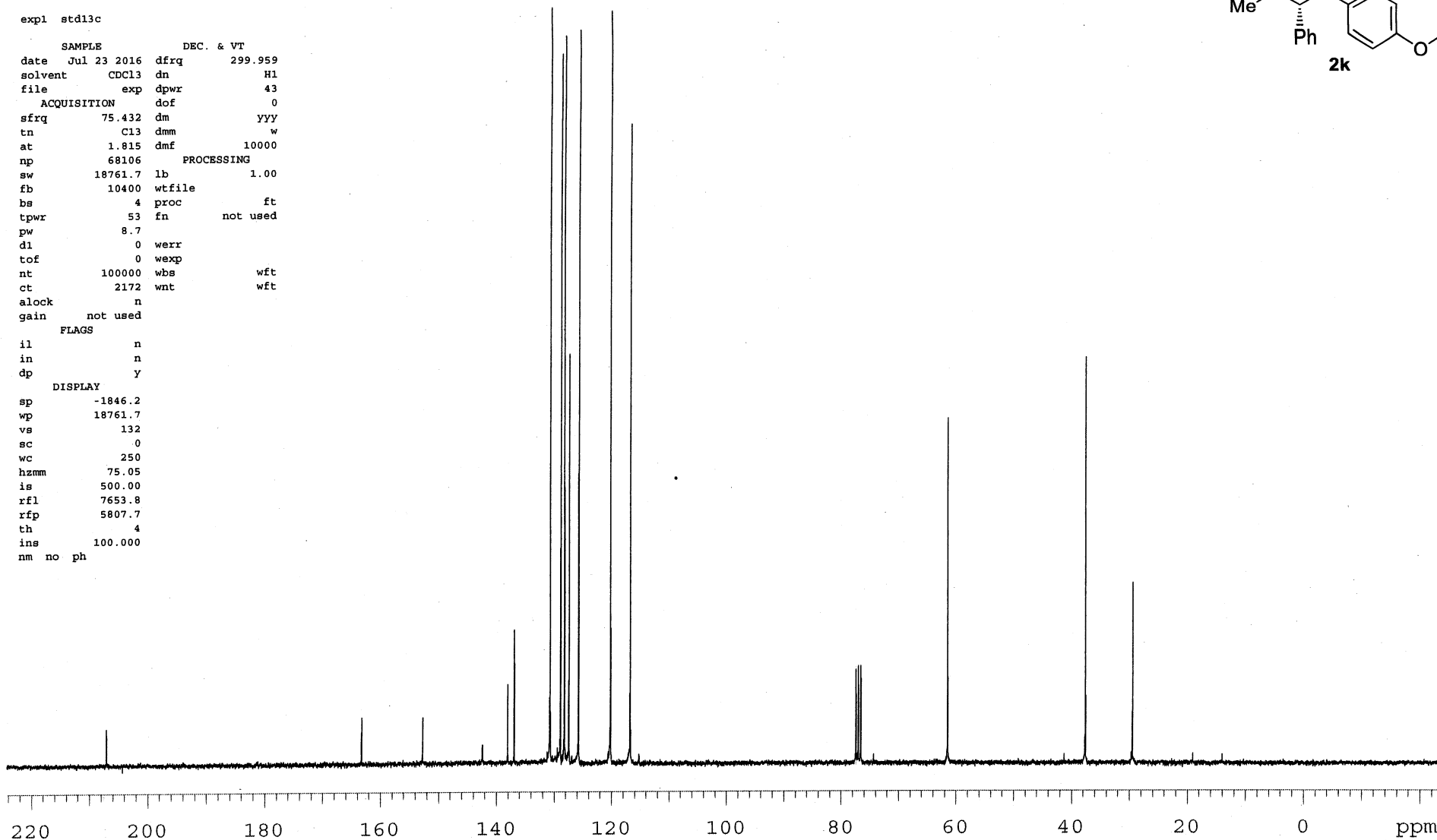
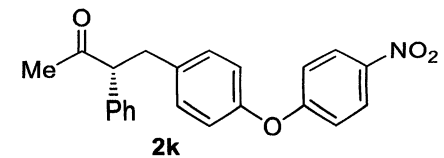
SAMPLE		DEC. & VT	
date	Jul 15 2016	dfrq	299.959
solvent	CDCl3	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	
sfrq	299.959	dm	nnn
tn	H1	dmm	c
at	1.998	dmf	200
np	18008	PROCESSING	
sw	4506.5	wtfile	
fb	not used	proc	ft
bs	4	fn	not used
tpwr	56		
pw	7.0	werr	
dl	1.000	wexp	
tof	0	wbs	wft
nt	8	wnt	wft
ct	8		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	-300.1		
wp	3299.5		
vs	126		
sc	0		
wc	250		
hzmm	13.20		
is	1174.45		
rfl	761.4		
rfp	0		
th	20		
ins	100.000		
nm	cdc ph		



MF E-9 PTLCL1 13C

expl std13c

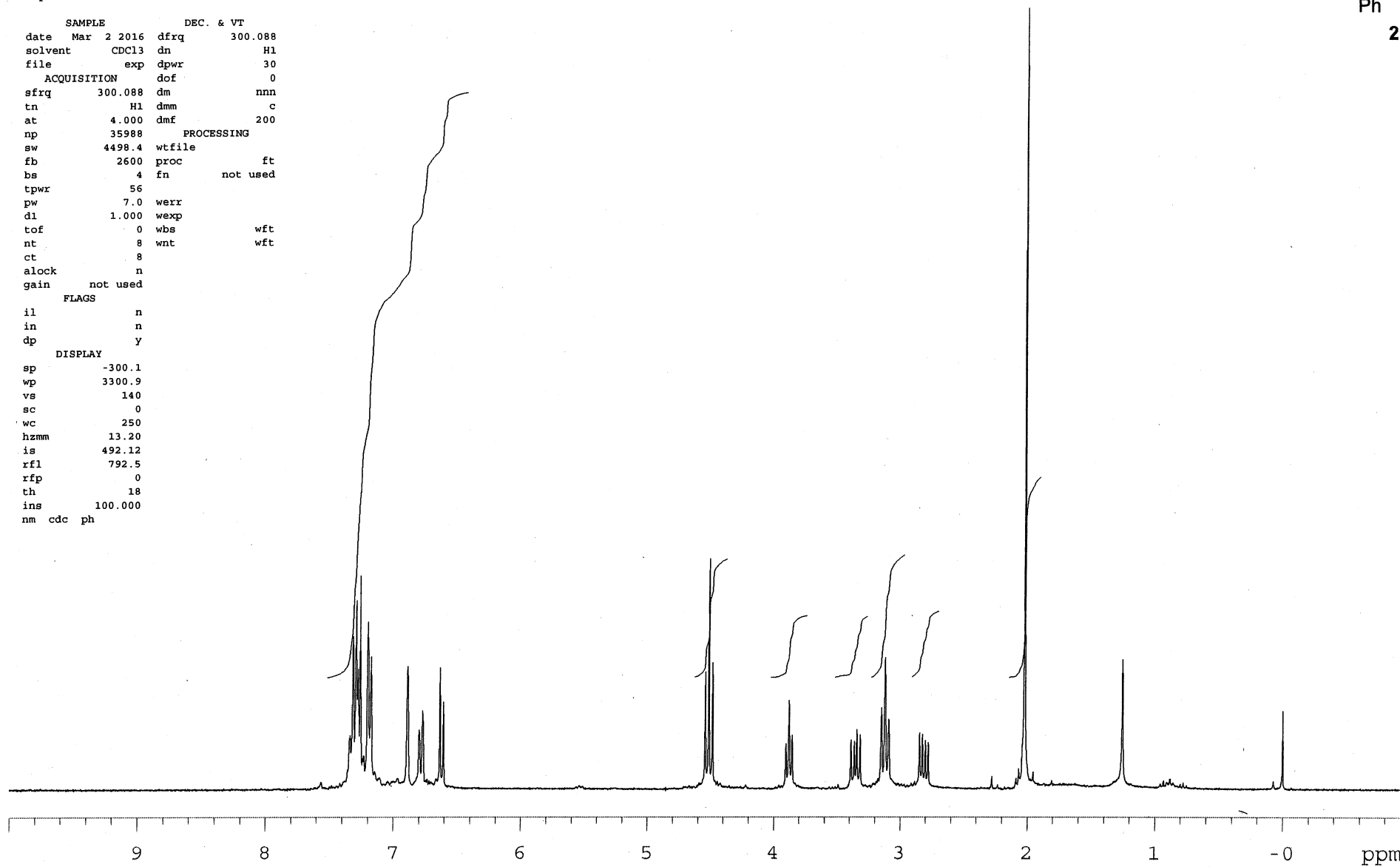
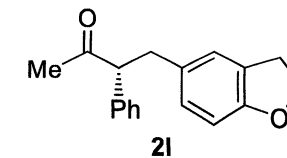
SAMPLE		DEC. & VT	
date	Jul 23 2016	dfrq	299.959
solvent	CDC13	dn	H1
file	exp	dpwr	43
ACQUISITION		dof	0
sfrq	75.432	dm	YYY
tn	C13	dmm	w
at	1.815	dmf	10000
np	68106	PROCESSING	
sw	18761.7	lb	1.00
fb	10400	wtfile	
bs	4	proc	ft
tpwr	53	fn	not used
pw	8.7		
d1	0	werr	
tof	0	wexp	
nt	100000	wbs	wft
ct	2172	wnt	wft
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	Y		
DISPLAY			
sp	-1846.2		
wp	18761.7		
vs	132		
sc	0		
wc	250		
hzmm	75.05		
is	500.00		
rfl	7653.8		
rfp	5807.7		
th	4		
ins	100.000		
nm	no ph		



TK A-16 PTLCL1

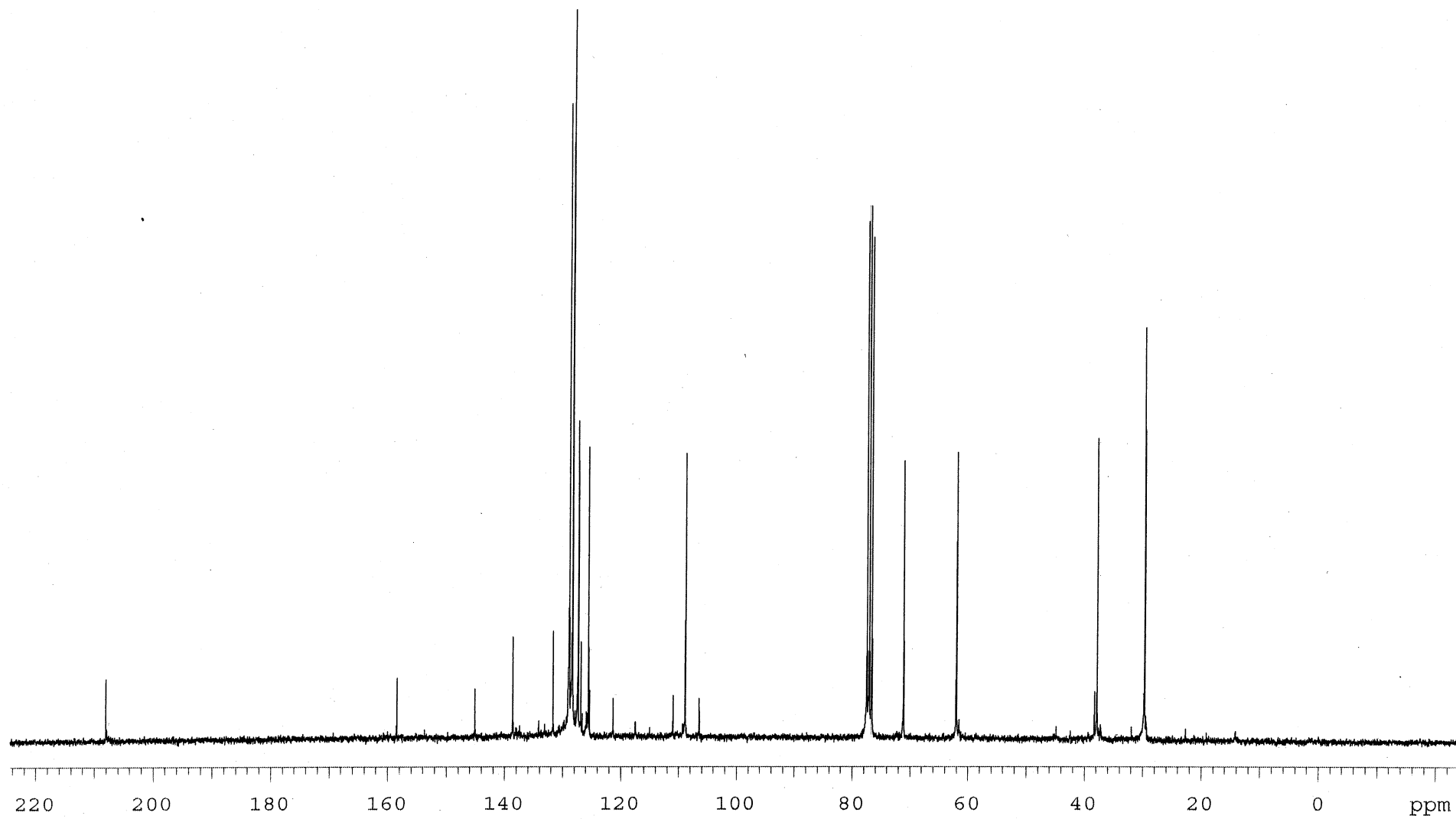
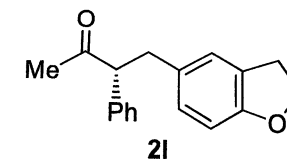
expl stdlh

SAMPLE		DEC. & VT	
date	Mar 2 2016	dfrq	300.088
solvent	CDCl3	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	0
sfrq	300.088	dm	nnn
tn	H1	dmm	c
at	4.000	dmf	200
np	35988	PROCESSING	
sw	4498.4	wtfile	
fb	2600	proc	ft
bs	4	fn	not used
tpwr	56		
pw	7.0	werr	
dl	1.000	wexp	
tof	0	wbs	wft
nt	8	wnt	wft
ct	8		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	Y		
DISPLAY			
sp	-300.1		
wp	3300.9		
vs	140		
sc	0		
wc	250		
hzmm	13.20		
is	492.12		
rfl	792.5		
rfp	0		
th	18		
ins	100.000		
nm	cdc ph		



TK A-16 PTLCl 13C

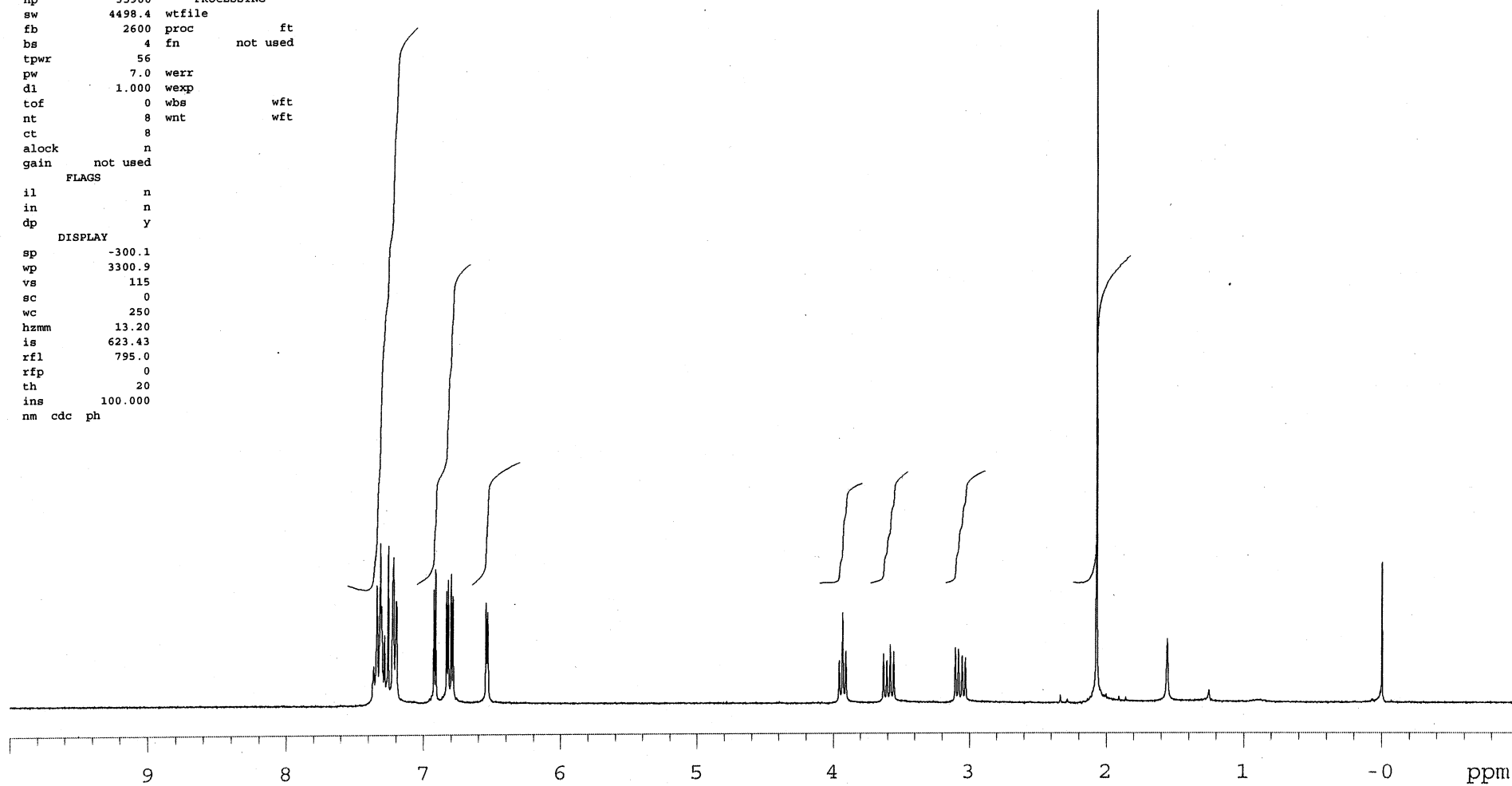
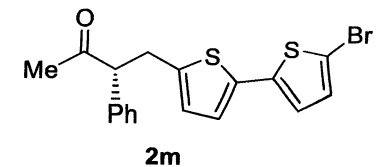
Pulse Sequence: s2pul



MF-E-21 PTLCl

expl stdlh

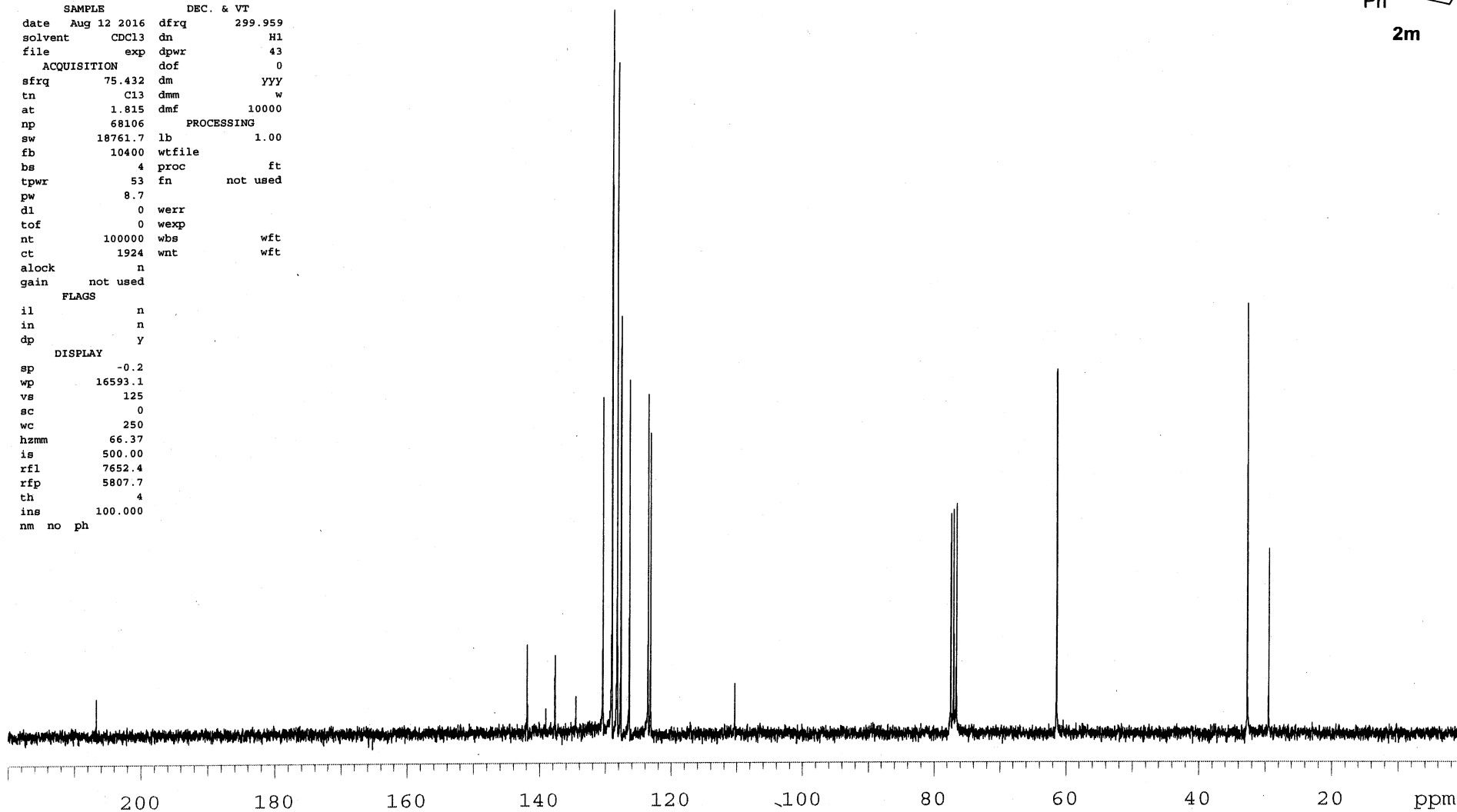
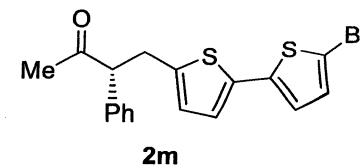
SAMPLE		DEC. & VT	
date	Jul 29 2016	dfrq	300.088
solvent	CDCl3	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	
sfrq	300.088	dm	nnn
tn	H1	dmm	c
at	4.000	dmf	200
np	35988	PROCESSING	
sw	4498.4	wtfile	
fb	2600	proc	ft
bs	4	fn	not used
tpwr	56		
pw	7.0	werr	
di	1.000	wexp	
tof	0	wbs	wft
nt	8	wnt	wft
ct	8		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	-300.1		
wp	3300.9		
vs	115		
sc	0		
wc	250		
hzmm	13.20		
is	623.43		
rfl	795.0		
rfp	0		
th	20		
ins	100.000		
nm	cdc ph		



ME E-21 PTLCL1 13C

expl std13c

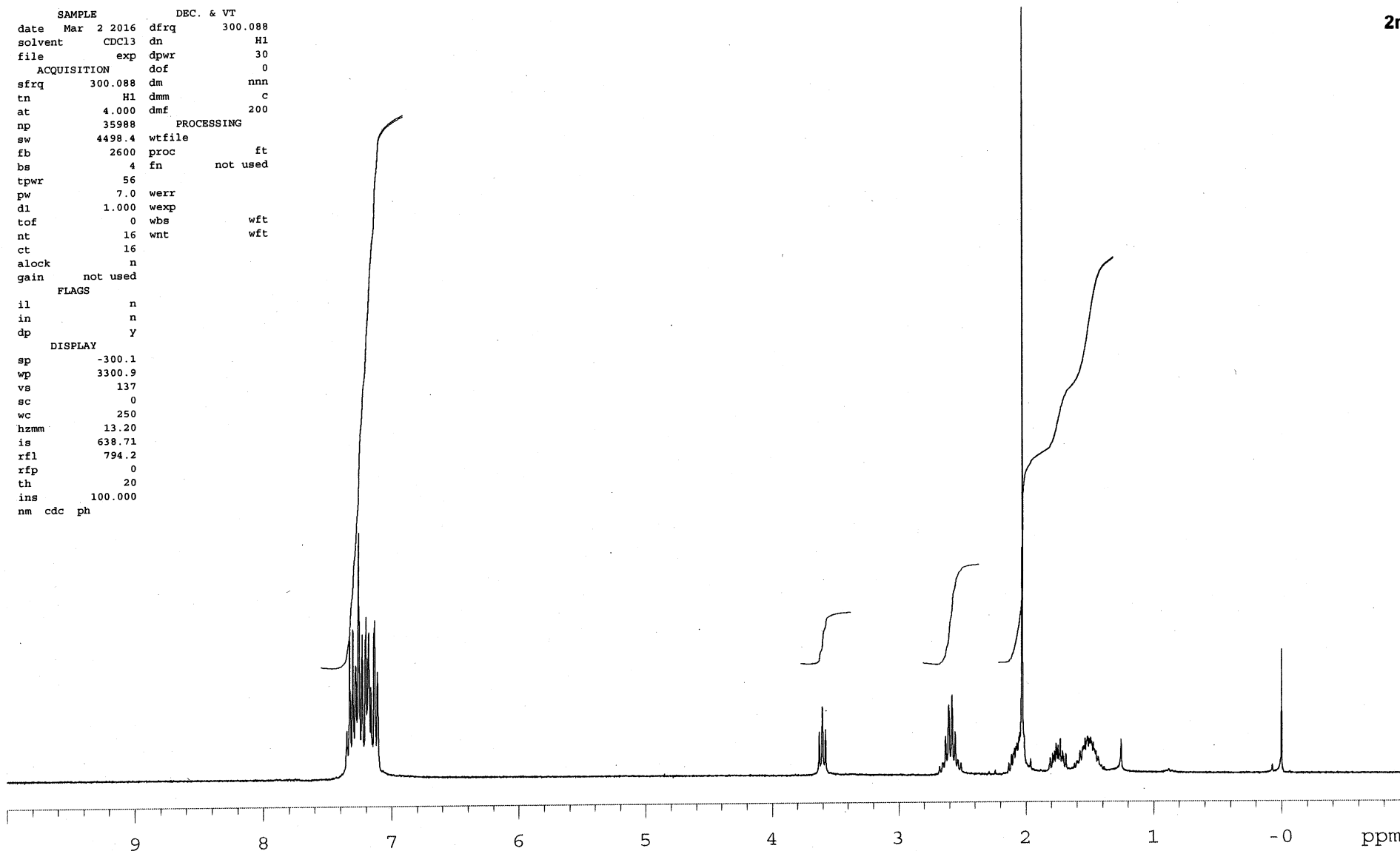
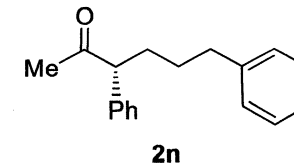
SAMPLE		DEC. & VT	
date	Aug 12 2016	dfrq	299.959
solvent	CDCl3	dn	H1
file	expl	dpwr	43
ACQUISITION		dof	0
sfrq	75.432	dm	YYY
tn	C13	dmm	w
at	1.815	dmf	10000
np	68106	PROCESSING	
sw	18761.7	lb	1.00
fb	10400	wtfile	
bs	4	proc	ft
tpwr	53	fn	not used
pw	8.7		
d1	0	werr	
tof	0	wexp	
nt	100000	wbs	wft
ct	1924	wnt	wft
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	-0.2		
wp	16593.1		
vs	125		
sc	0		
wc	250		
hzmm	66.37		
is	500.00		
rfl	7652.4		
rfp	5807.7		
th	4		
ins	100.000		
nm	no ph		



MF B-33 FTLC1

expl std1h

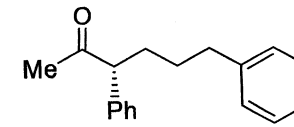
SAMPLE		DEC. & VT	
date	Mar 2 2016	dfrq	300.088
solvent	CDCl3	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	0
sfrq	300.088	dm	nnn
tn	H1	dmm	c
at	4.000	dmf	200
np	35988	PROCESSING	
sw	4498.4	wtfile	
fb	2600	proc	ft
bs	4	fn	not used
tpwr	56		
pw	7.0	werr	
d1	1.000	wexp	
tof	0	wbs	wft
nt	16	wnt	wft
ct	16		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	Y		
DISPLAY			
sp	-300.1		
wp	3300.9		
vs	137		
sc	0		
wc	250		
hzmm	13.20		
is	638.71		
rfl	794.2		
rpf	0		
th	20		
ins	100.000		
nm	cdc ph		



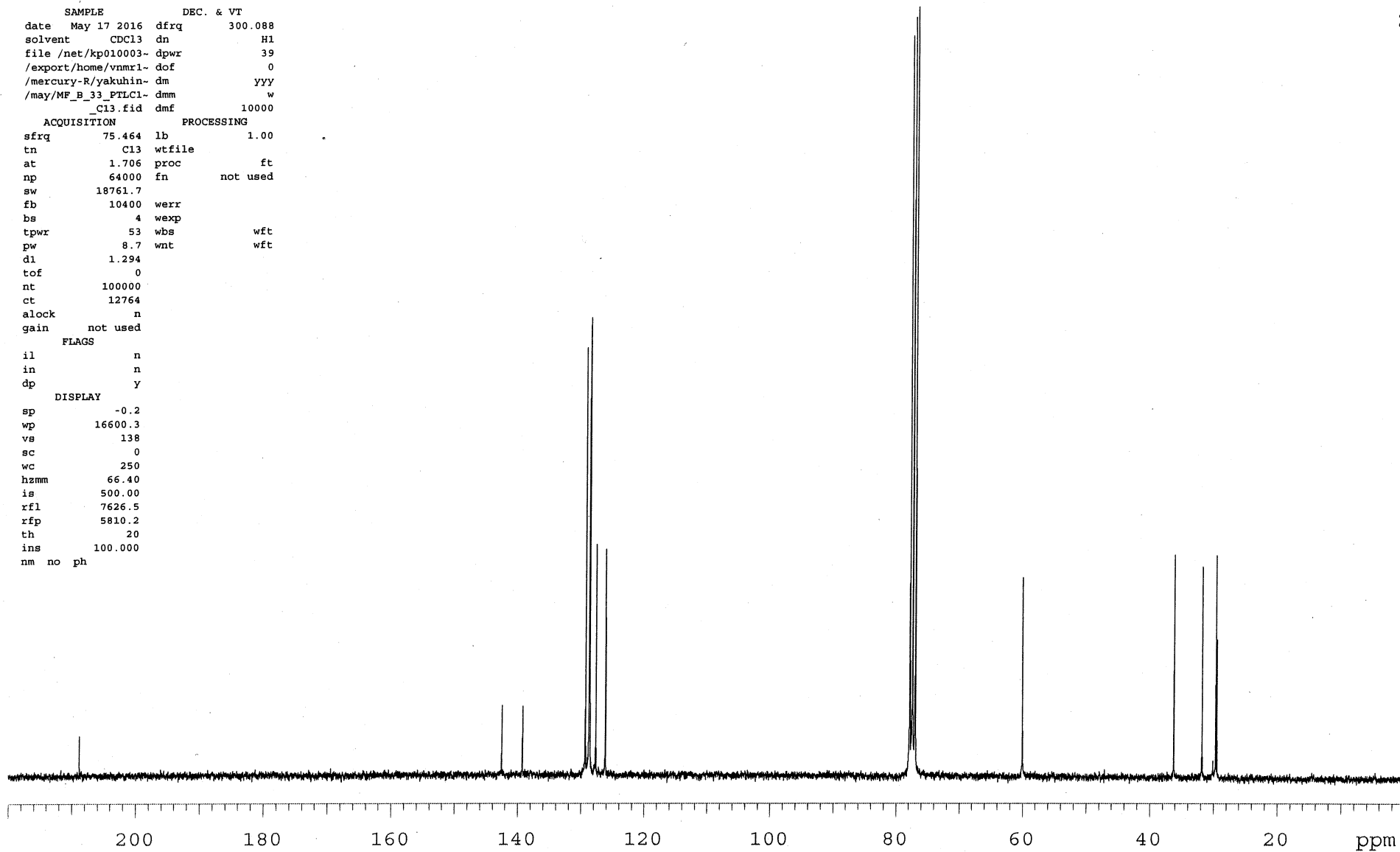
MF B-33 PTLCl C13

expl std13c

SAMPLE		DEC. & VT	
date	May 17 2016	dfrq	300.088
solvent	CDCl3	dn	H1
file	/net/kp010003-	dpwr	39
/export/home/vnmr1-	dof		0
/mercury-R/yakuhin-	dm	YYY	
/may/MF_B_33_PTLCl-	dmm	w	
_C13.fid	dmf	10000	
ACQUISITION		PROCESSING	
sfrq	75.464	lb	1.00
tn	C13	wtfile	
at	1.706	proc	ft
np	64000	fn	not used
sw	18761.7		
fb	10400	werr	
bs	4	wexp	
tpwr	53	wbs	wft
pw	8.7	wnt	wft
d1	1.294		
tof	0		
nt	100000		
ct	12764		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	-0.2		
wp	16600.3		
vs	138		
sc	0		
wc	250		
hzmm	66.40		
is	500.00		
rfl	7626.5		
rfp	5810.2		
th	20		
ins	100.000		
nm	no	ph	



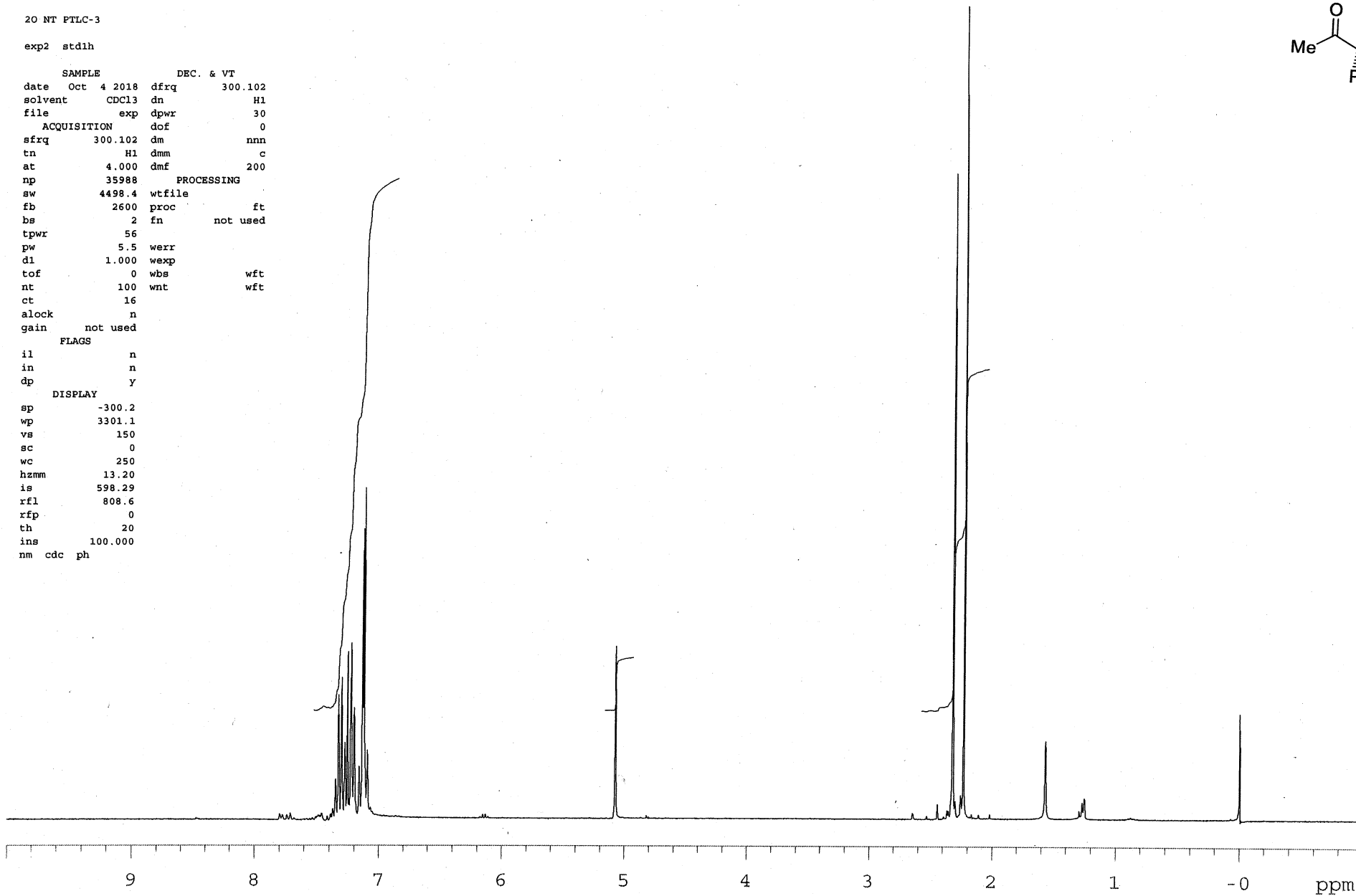
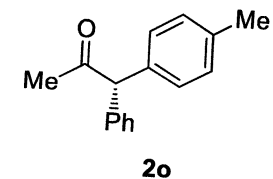
2n



20 NT FTLC-3

exp2 stdih

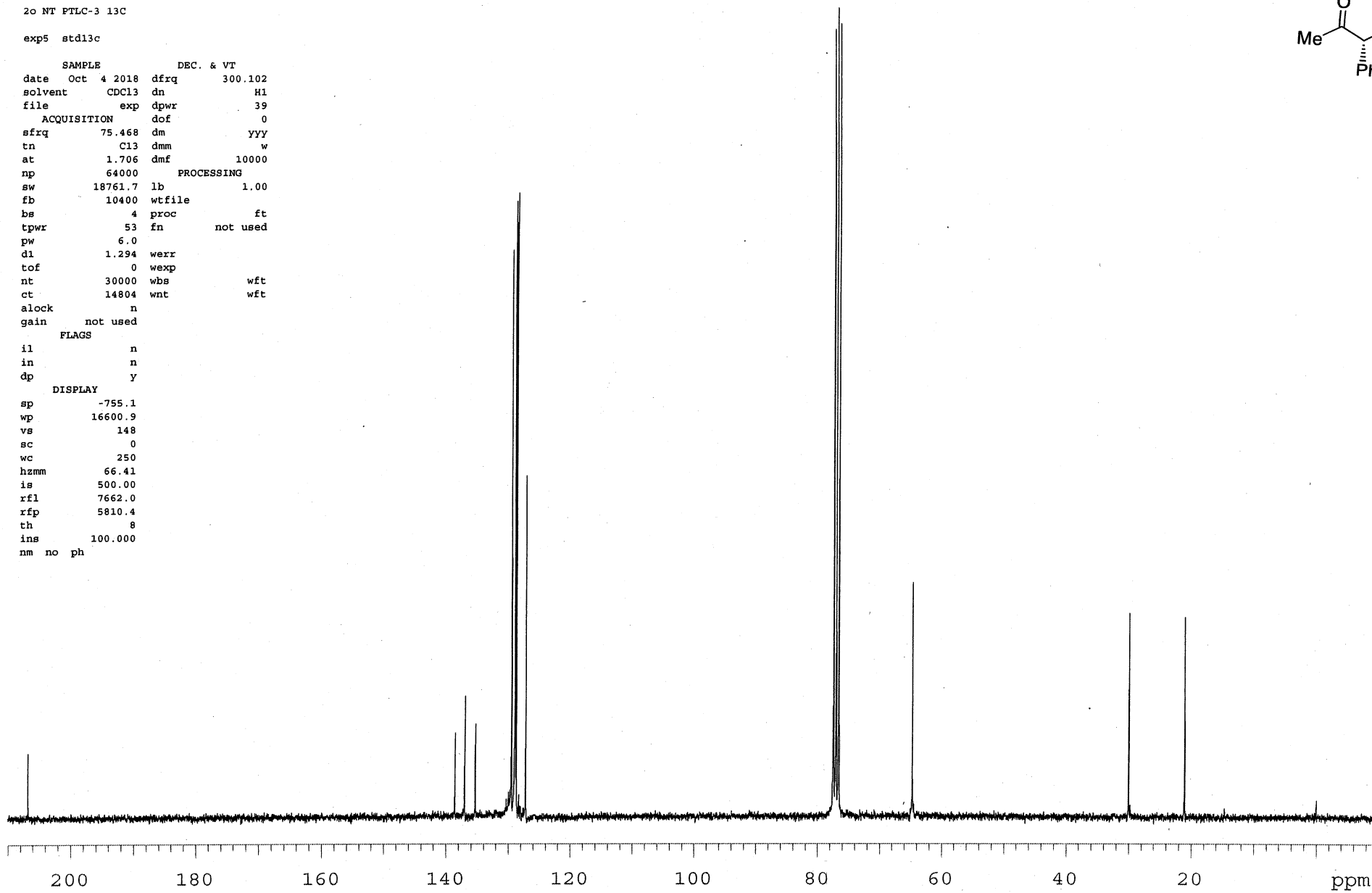
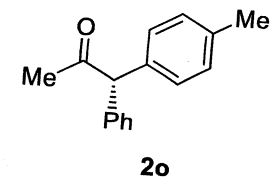
```
SAMPLE          DEC. & VT
date Oct 4 2018 dfrq      300.102
solvent CDCl3  dn        H1
file      exp dpwr       30
ACQUISITION    dof       0
sfrq      300.102 dm      nnn
tn         H1 dmm        c
at         4.000 dmf     200
np         35988  PROCESSING
sw         4498.4 wtfile
fb         2600  proc      ft
bs         2    fn        not used
tpwr       56
pw         5.5  werr
dl         1.000 wexp
tof        0    wbs      wft
nt         100  wnt      wft
ct         16
alock      n
gain       not used
FLAGS
il         n
in         n
dp         Y
DISPLAY
sp        -300.2
wp        3301.1
vs        150
sc         0
wc        250
hzmm     13.20
is        598.29
rfl       808.6
xrp       0
th        20
ins      100.000
nm cdc ph
```



2o NT FTLC-3 13C

exp5 std13c

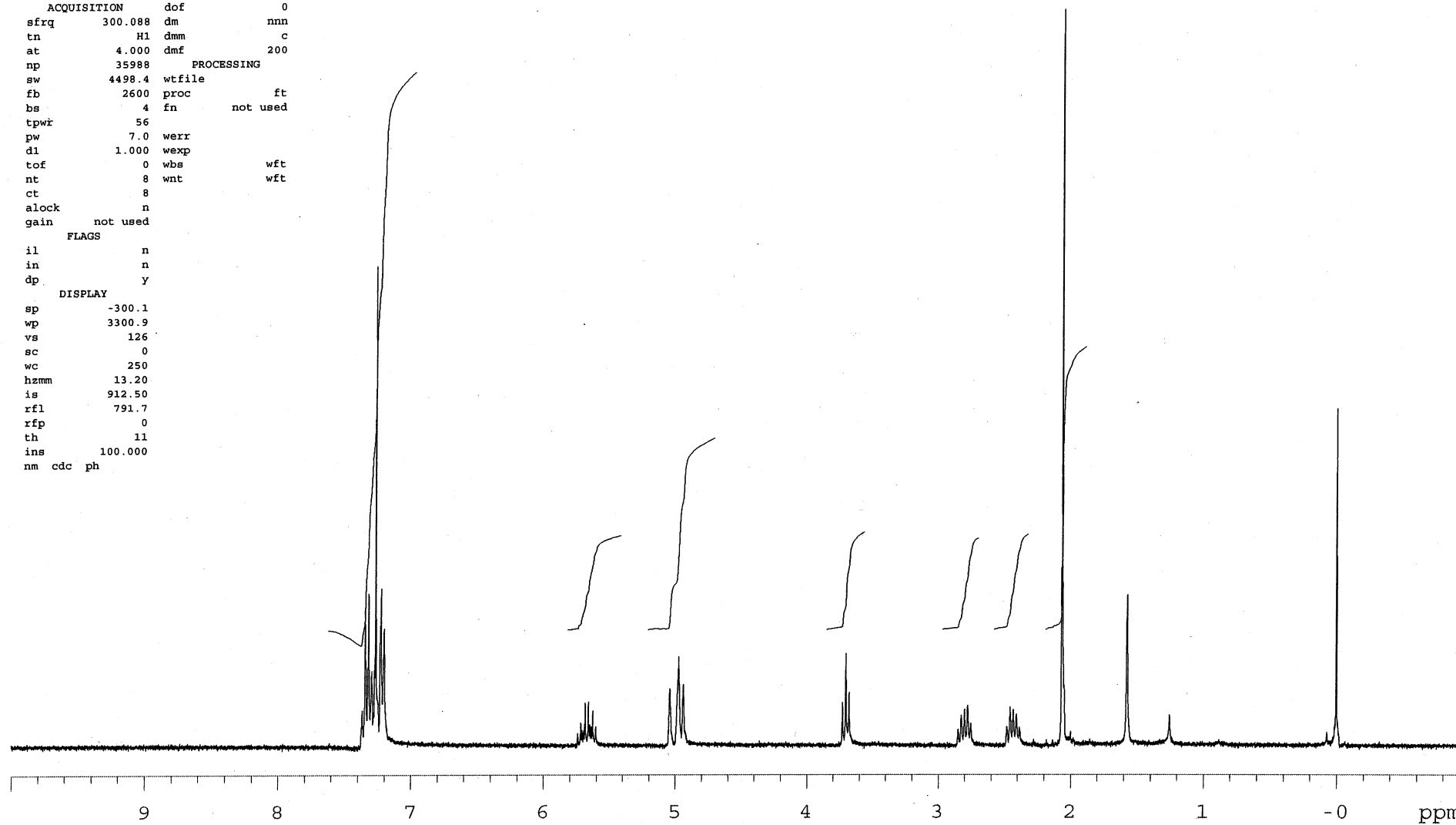
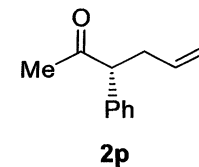
```
SAMPLE          DEC. & VT
date   Oct  4 2018  dfrq      300.102
solvent CDCl3      dn         H1
file    exp      dpwr         39
ACQUISITION      dof         0
sfrq     75.468  dm          yyy
tn        C13     dmm         w
at        1.706  dmf        10000
np        64000  PROCESSING
sw     18761.7  lb          1.00
fb        10400  wtfile
bs         4     proc         ft
tpwr       53   fn         not used
pw         6.0
d1        1.294  werr
tof         0    wexp
nt        30000  wbs          wft
ct        14804  wnt          wft
alock      n
gain      not used
FLAGS
il         n
in         n
dp         y
DISPLAY
sp        -755.1
wp       16600.9
vs         148
sc         0
wc         250
hzmm      66.41
is         500.00
rfl       7662.0
rfp       5810.4
th         8
ins       100.000
nm no ph
```



MF A-33

exp2 std1h

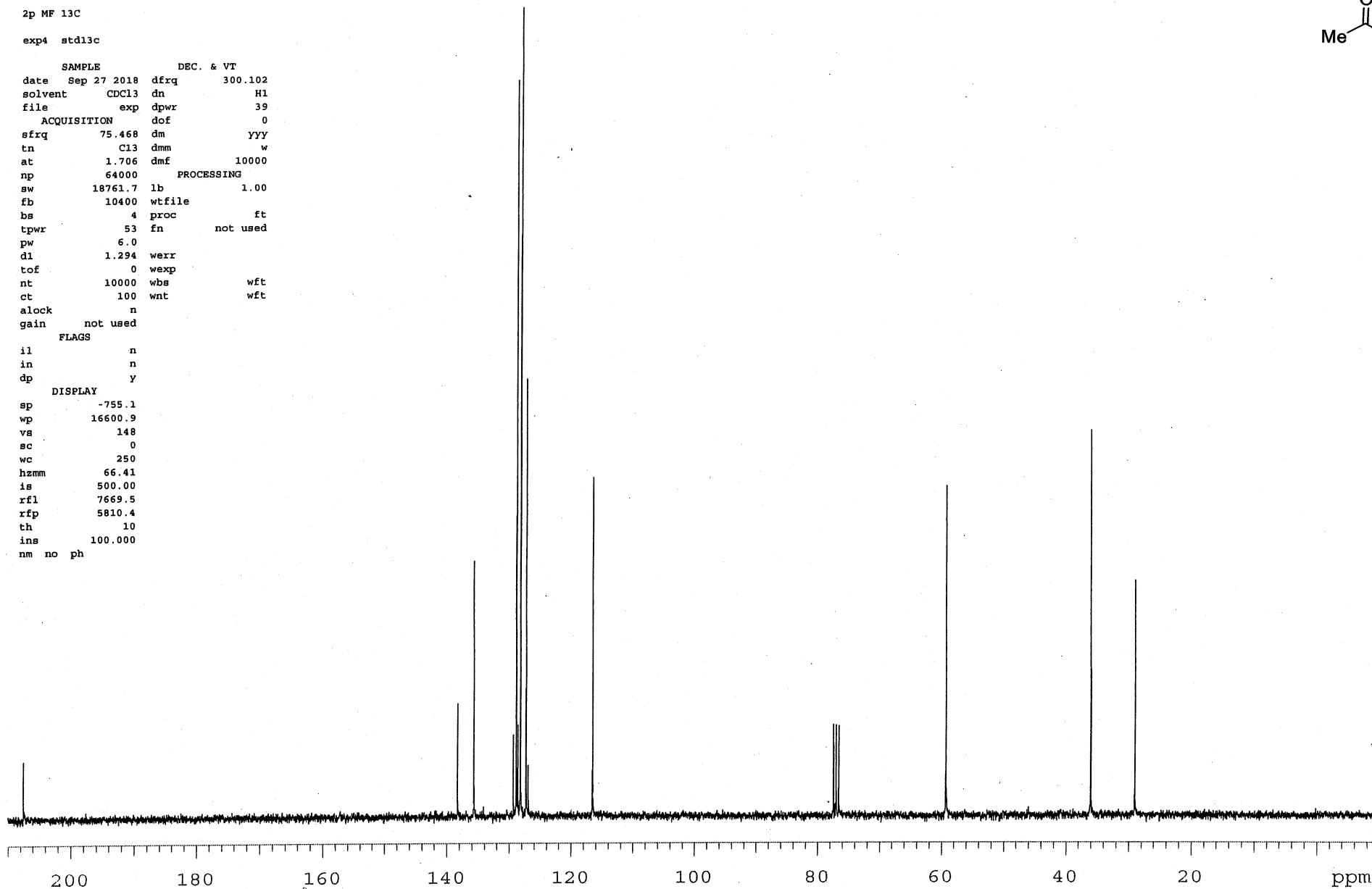
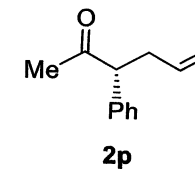
SAMPLE		DEC. & VT	
date	Mar 1 2016	dfrq	300.088
solvent	CDCl3	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	0
sfrq	300.088	dm	nmn
tn	H1	dmm	c
at	4.000	dmf	200
np	35988	PROCESSING	
sw	4498.4	wtfile	
fb	2600	proc	ft
bs	4	fn	not used
tpwr	56		
pw	7.0	werr	
d1	1.000	wexp	
tof	0	wbs	wft
nt	8	wnt	wft
ct	8		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	-300.1		
wp	3300.9		
vs	126		
sc	0		
wc	250		
hzmm	13.20		
is	912.50		
rfl	791.7		
rfl	0		
th	11		
ins	100.000		
nm	cdc ph		



2p MF 13C

exp4 std13c

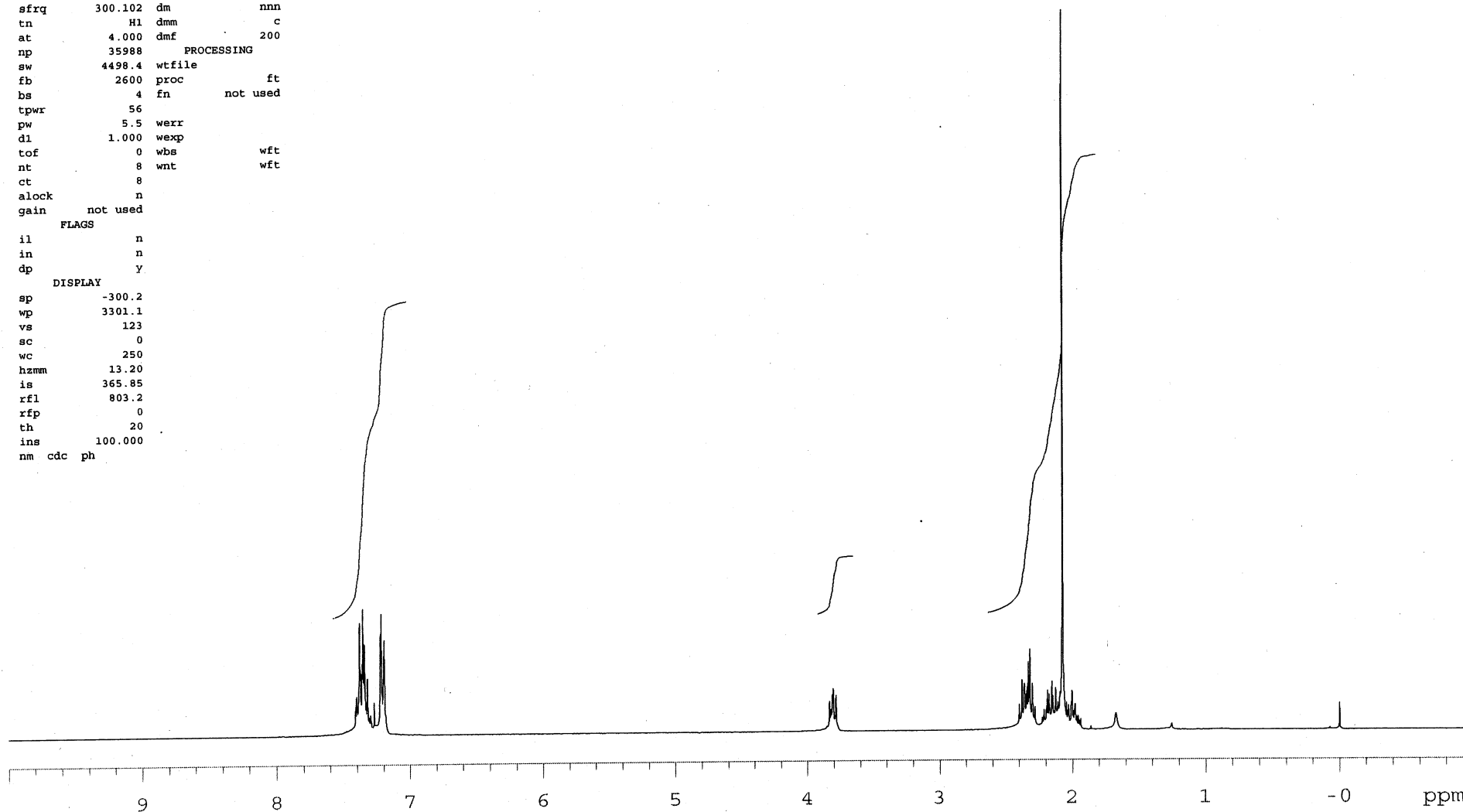
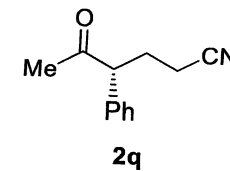
```
SAMPLE          DEC. & VT
date Sep 27 2018 dfrq      300.102
solvent CDC13      dn       H1
file      exp      dpwr     39
ACQUISITION      dof       0
sfrq      75.468  dm       yyy
tn         C13     dmm      w
at         1.706  dmf     10000
np         64000  PROCESSING
sw         18761.7 lb       1.00
fb         10400  wtfile
bs         4      proc     ft
tpwr      53     fn       not used
pw         6.0
dl         1.294  werr
tof        0     wexp
nt         10000  wbs     wft
ct         100   wnt     wft
alock      n
gain      not used
FLAGS
il         n
in         n
dp         Y
DISPLAY
sp         -755.1
wp         16600.9
va         148
sc         0
wc         250
hzmm      66.41
is         500.00
rfl       7669.5
rfp       5810.4
th         10
ins       100.000
nm no ph
```



MF F-10 PTL2

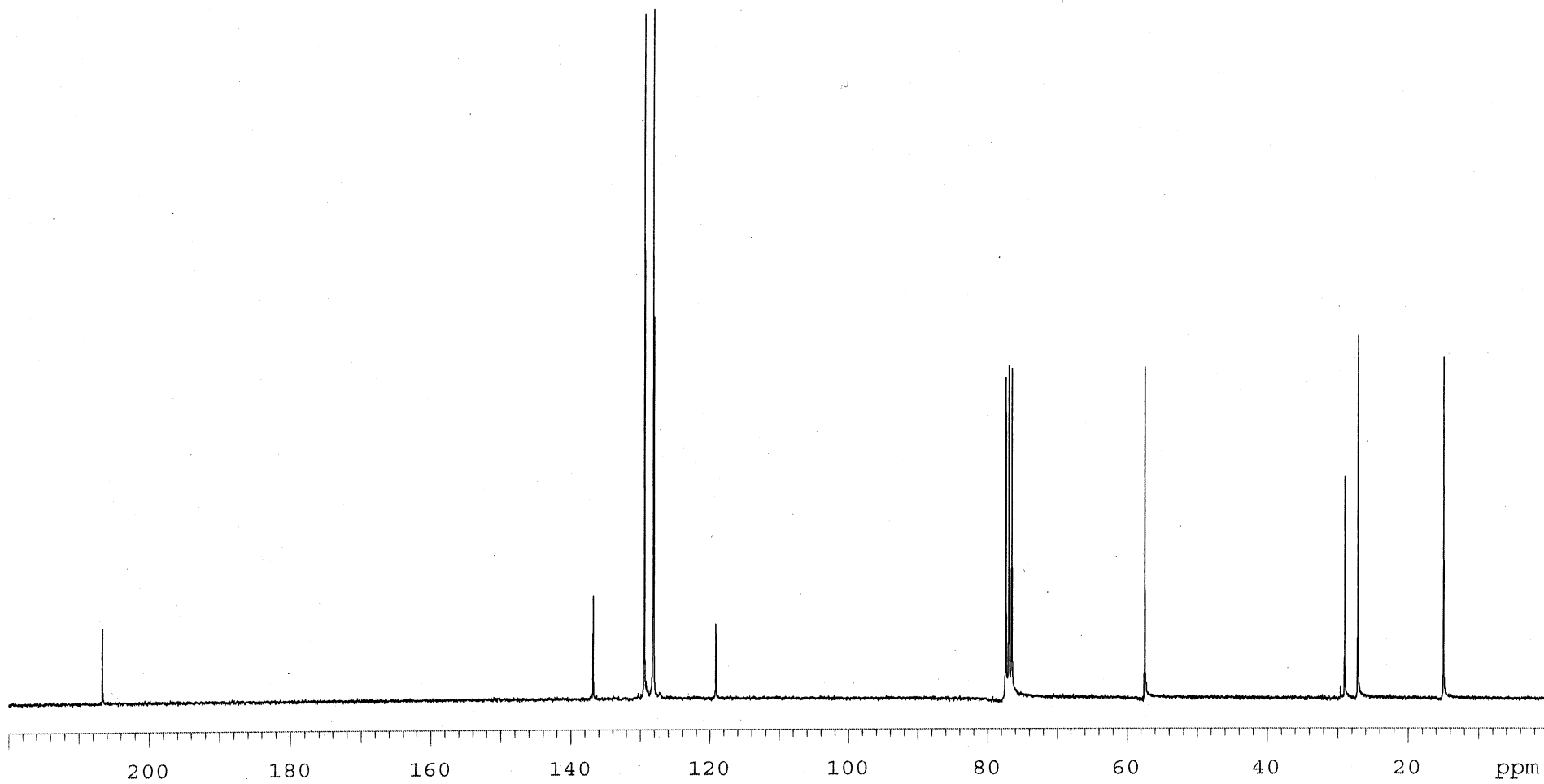
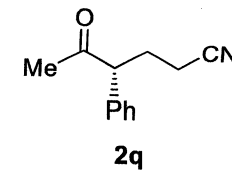
exp2 stdlh

SAMPLE		DEC. & VT	
date	Jul 24 2017	dfxq	300.102
solvent	CDCl3	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	0
sfrq	300.102	dm	nnn
tn	H1	dmm	c
at	4.000	dmf	200
np	35988	PROCESSING	
sw	4498.4	wtfile	
fb	2600	proc	ft
bs	4	fn	not used
tpwr	56		
pw	5.5	werr	
d1	1.000	wexp	
tof	0	wbs	wft
nt	8	wnt	wft
ct	8		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	Y		
DISPLAY			
sp	-300.2		
wp	3301.1		
vs	123		
sc	0		
wc	250		
hzmm	13.20		
is	365.85		
rfl	803.2		
rfp	0		
th	20		
ins	100.000		
nm	cdc ph		



MF F-10 PTLC2

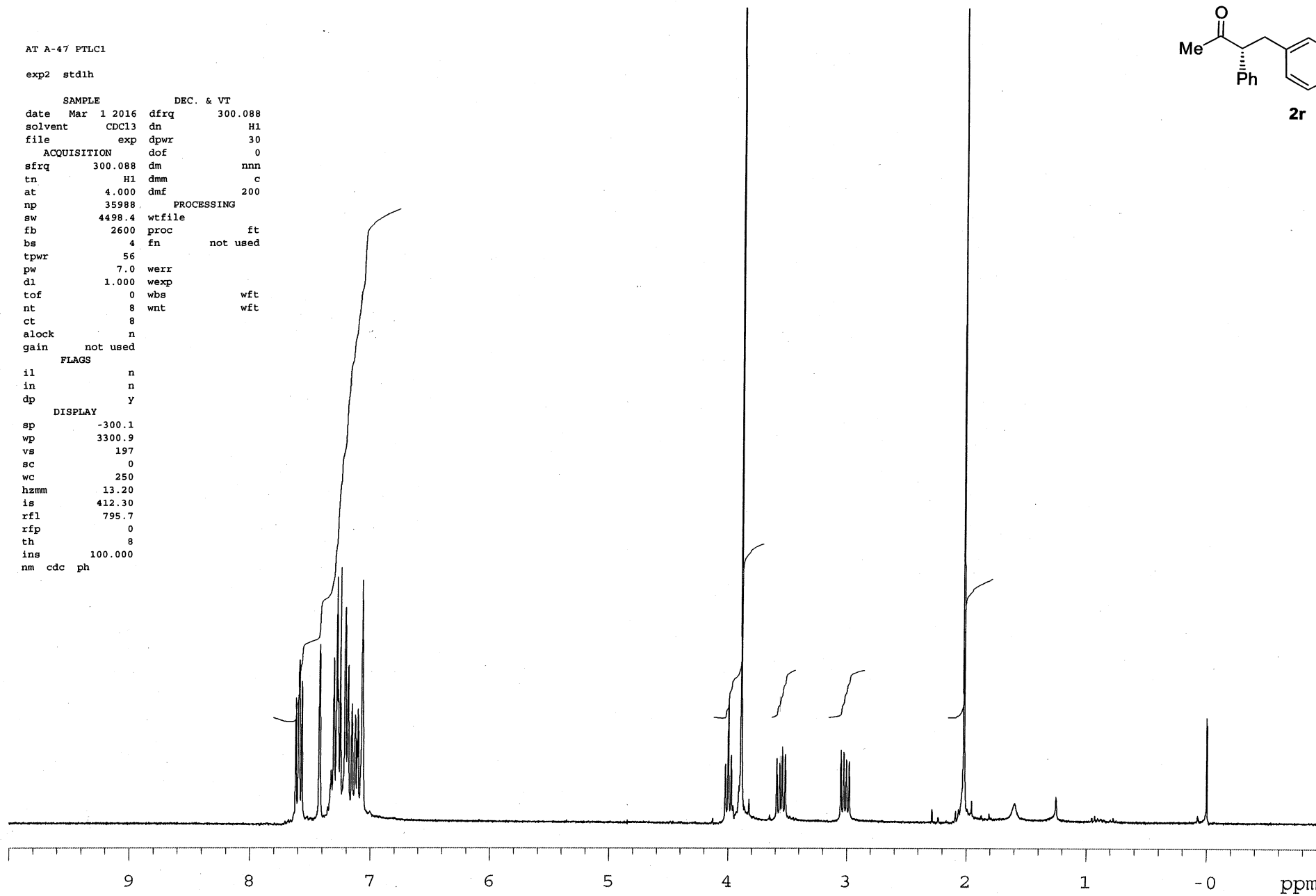
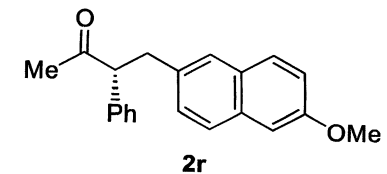
Pulse Sequence: s2pul



AT A-47 PTLCl

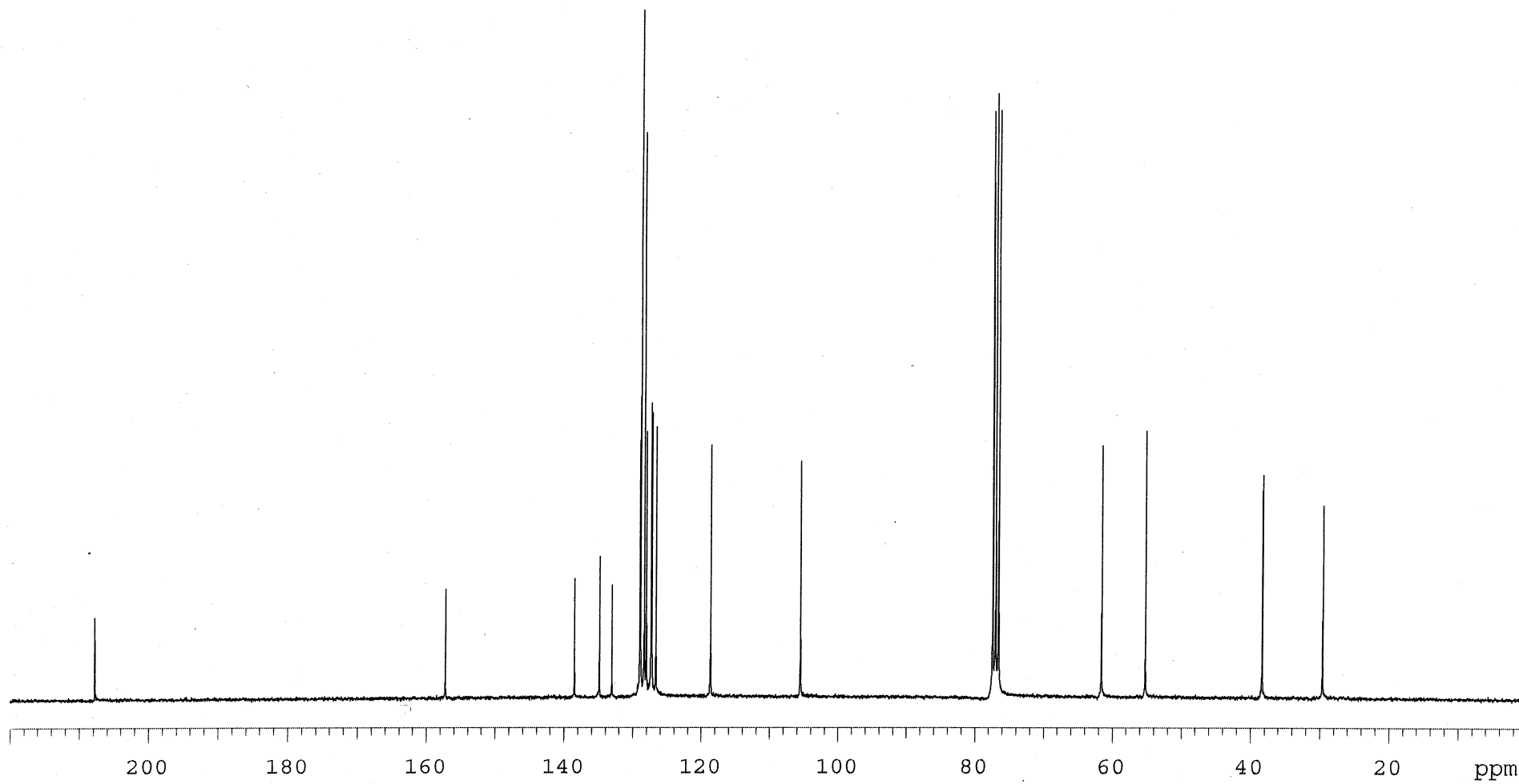
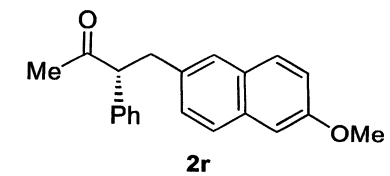
exp2 stdlh

```
SAMPLE          DEC. & VT
date  Mar  1 2016  dfrq      300.088
solvent CDCl3      dn        H1
file          exp  dpwr       30
ACQUISITION    dof         0
sfrq      300.088  dm        nnn
tn          H1    dmm        c
at          4.000  dmf       200
np      35988    PROCESSING
sw      4498.4  wtfile
fb      2600    proc        ft
bs          4    fn        not used
tpwr       56
pw          7.0  weirr
d1          1.000 wexp
tof          0   wbs        wft
nt           8   wnt        wft
ct           8
alock       n
gain    not used
FLAGS
il          n
in          n
dp          y
DISPLAY
sp      -300.1
wp      3300.9
vs       197
sc         0
wc       250
hzmm     13.20
is       412.30
rfl      795.7
rfp       0
th         8
ins     100.000
nm cdc ph
```



AT A-47 PTLCl

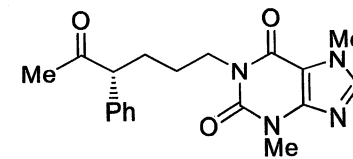
Pulse Sequence: s2pul



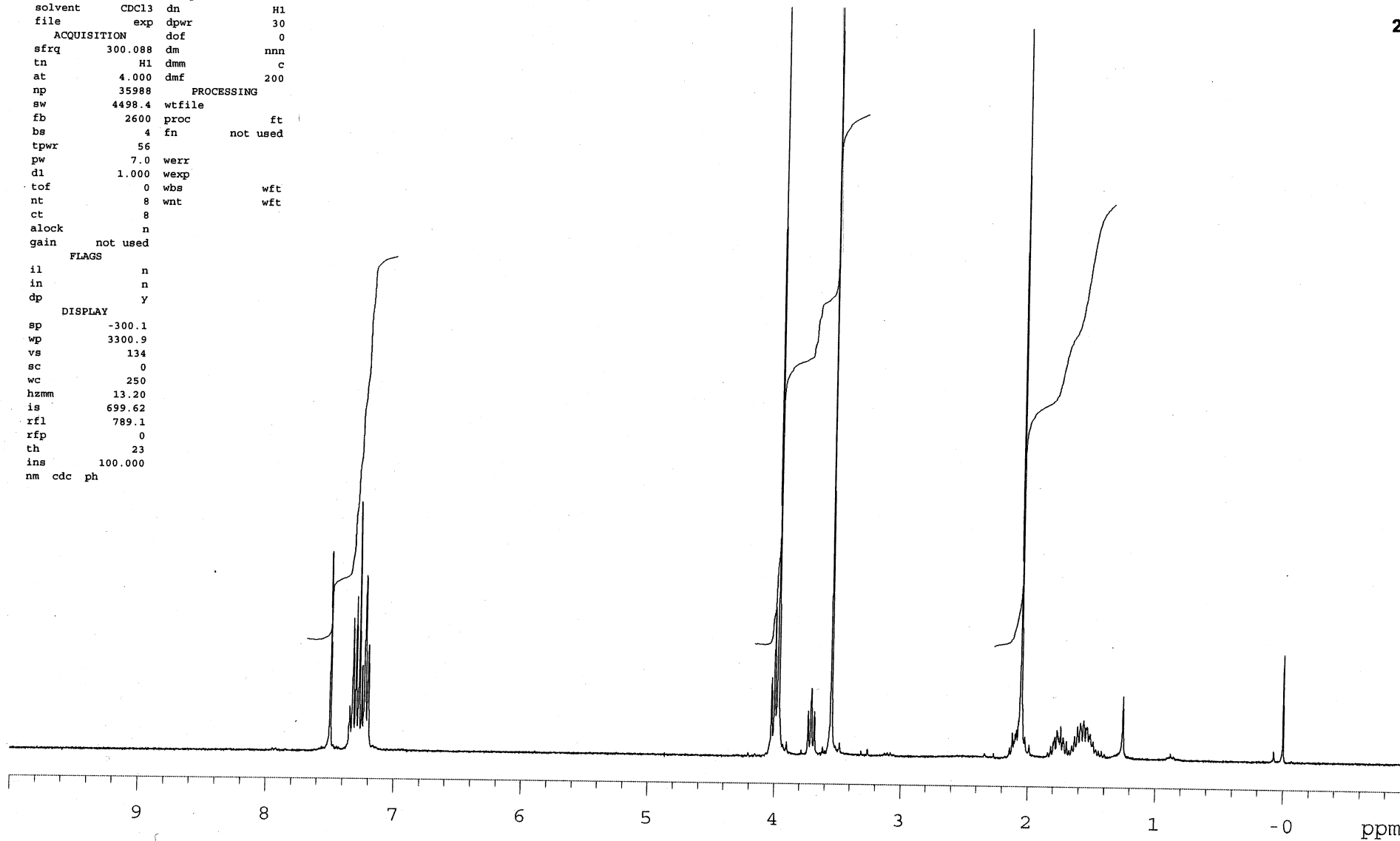
TK A-12 PTL2

expl stdih

SAMPLE		DEC. & VT	
date	Mar 2 2016	dfrq	300.088
solvent	CDCl3	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	0
sfrq	300.088	dm	nnn
tn	H1	dmm	c
at	4.000	dmf	200
np	35988	PROCESSING	
sw	4498.4	wtfile	
fb	2600	proc	ft
bs	4	fn	not used
tpwr	56		
pw	7.0	werr	
dl	1.000	wexp	
tof	0	wbs	wft
nt	8	wnt	wft
ct	8		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	-300.1		
wp	3300.9		
vs	134		
sc	0		
wc	250		
hzmm	13.20		
is	699.62		
rfl	789.1		
rfp	0		
th	23		
ins	100.000		
nm	cdc	ph	



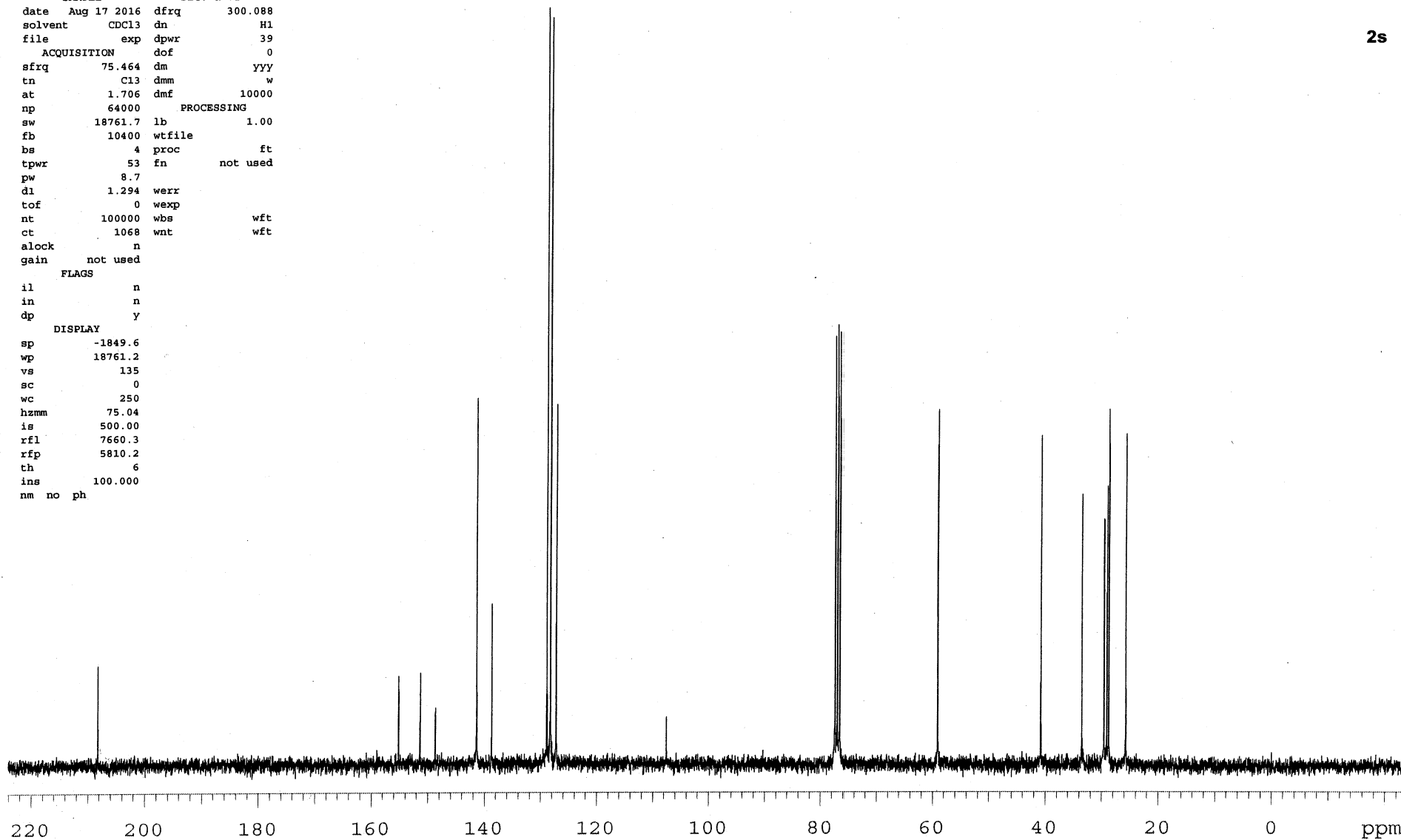
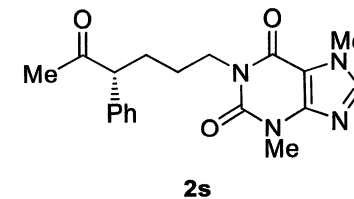
2s



TK-A-12 FTLC2 13C

expl std13c

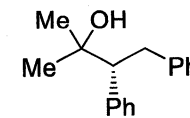
SAMPLE		DEC. & VT	
date	Aug 17 2016	dfrq	300.088
solvent	CDCl3	dn	H1
file	exp	dpwr	39
ACQUISITION		dof	0
sfrq	75.464	dm	YYY
tn	C13	dmm	w
at	1.706	dmf	10000
np	64000	PROCESSING	
sw	18761.7	lb	1.00
fb	10400	wtfile	
bs	4	proc	ft
tpwr	53	fn	not used
pw	8.7		
dl	1.294	werr	
tof	0	wexp	
nt	100000	wbs	wft
ct	1068	wnt	wft
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
DISPLAY			
sp	-1849.6		
wp	18761.2		
vs	135		
sc	0		
wc	250		
hzmm	75.04		
is	500.00		
rfl	7660.3		
rfp	5810.2		
th	6		
ins	100.000		
nm	no ph		



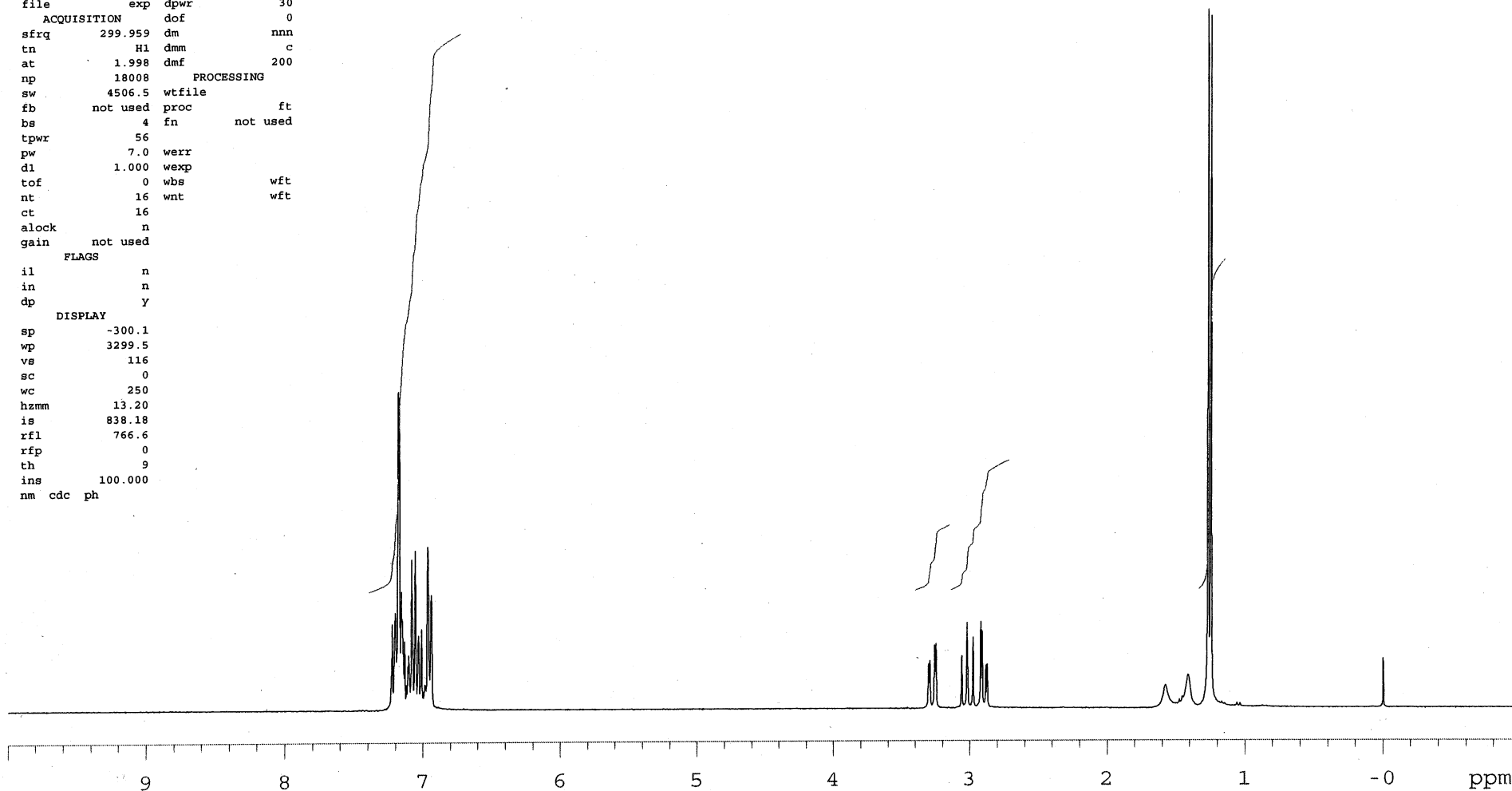
MF D-14 PTLCl

exp2 stdlh

SAMPLE		DEC. & VT	
date	Jul 15 2016	dfrq	299.959
solvent	CDCl3	dn	H1
file	exp	dpwr	30
ACQUISITION		dof	
sfrq	299.959	dm	nnn
tn	H1	dmm	c
at	1.998	dmf	200
np	18008	PROCESSING	
sw	4506.5	wtfile	
fb	not used	proc	ft
bs	4	fn	not used
tpwr	56		
pw	7.0	werr	
d1	1.000	wexp	
tof	0	wbs	wft
nt	16	wnt	wft
ct	16		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	Y		
DISPLAY			
sp	-300.1		
wp	3299.5		
vs	116		
sc	0		
wc	250		
hzmm	13.20		
is	838.18		
rfl	766.6		
rfp	0		
th	9		
ins	100.000		
nm	cdc ph		



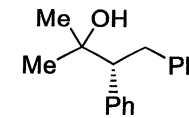
4



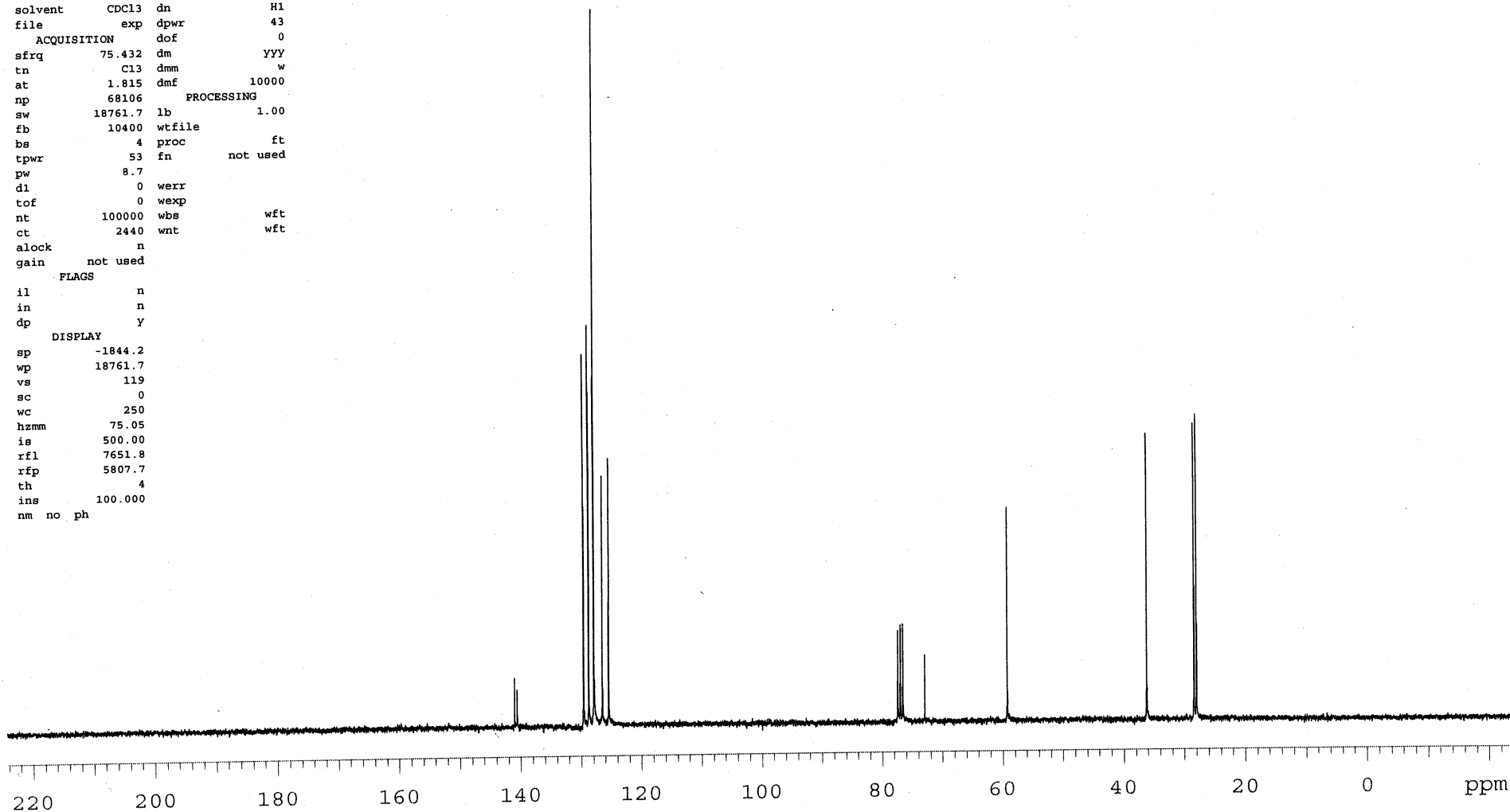
MF D-14 PTLCl 13C

exp2 std13c

```
SAMPLE          DEC. & VT
date   Jul 25 2016  dfrq   299.959
solvent  CDCl3     dn      H1
file     exp       dpwr    43
ACQUISITION    dof      0
sfrq     75.432   dm      YYY
tn        c13     dmm      w
at        1.815   dmf     10000
np        68106   PROCESSING
sw       18761.7  lb      1.00
fb        10400  wtfile
bs         4     proc     ft
tpwr      53     fn      not used
pw         8.7
dl         0     werr
tof        0     wexp
nt       100000  wbs      wft
ct        2440  wnt      wft
alock     n
gain     not used
FLAGS
il        n
in        n
dp        Y
DISPLAY
sp       -1844.2
wp       18761.7
vs       119
sc        0
wc       250
hzmm     75.05
is       500.00
rfl      7651.8
rfp      5807.7
th        4
ins     100.000
nm no ph
```



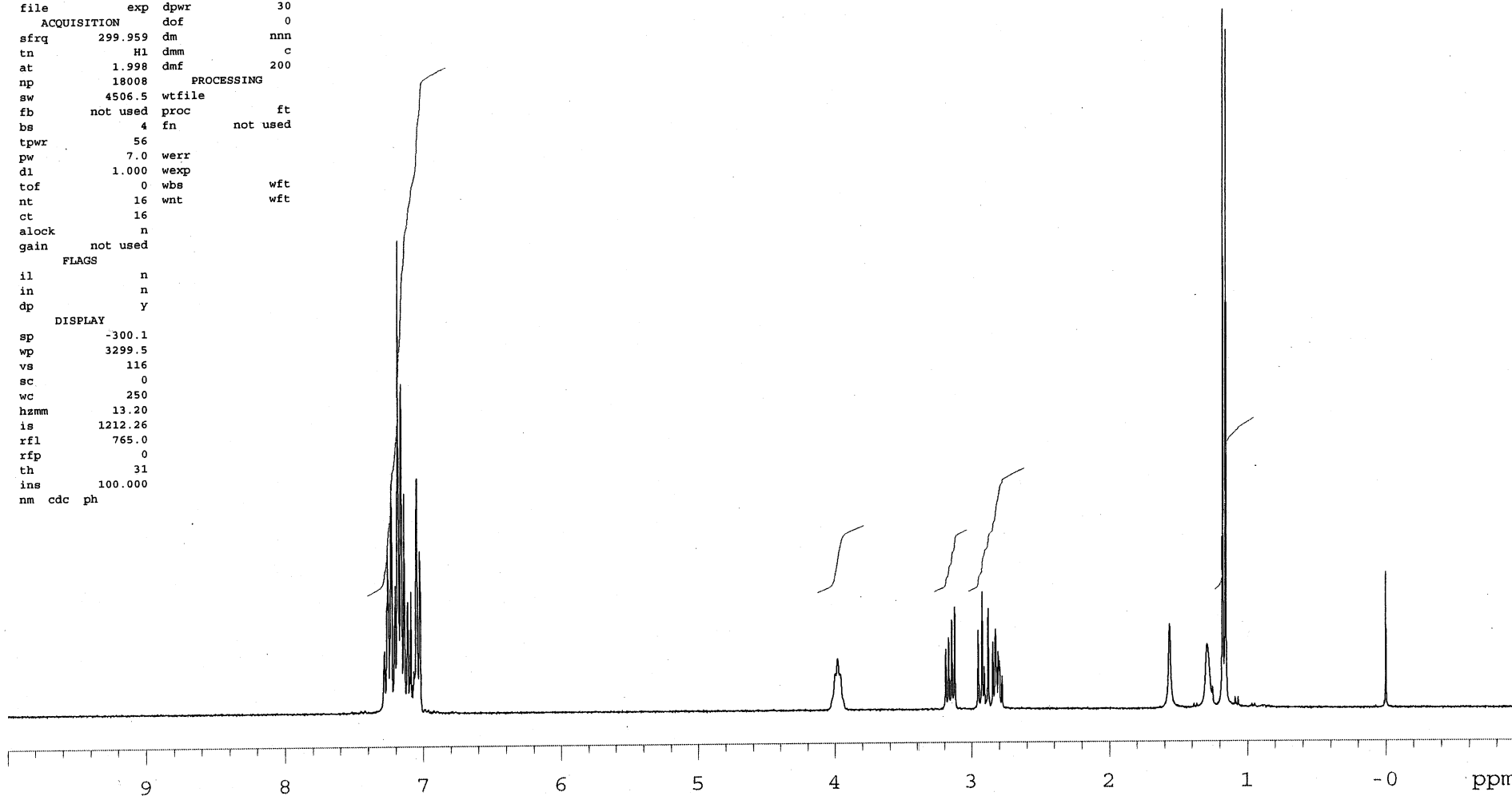
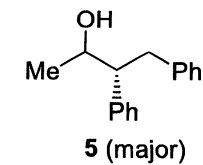
4



MF D-8 PTLCl

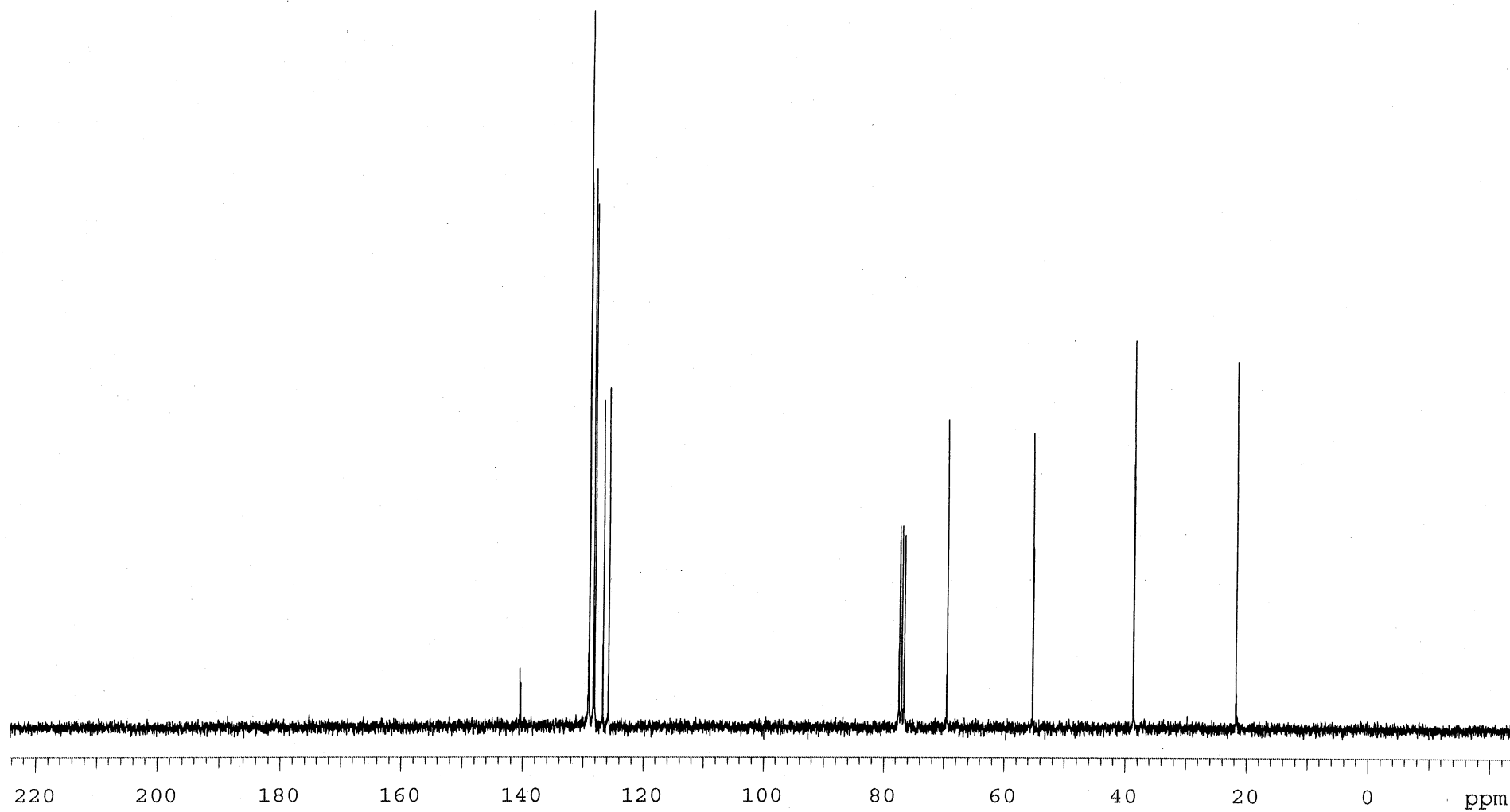
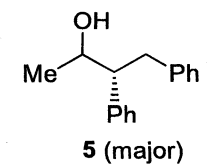
exp2 stdih

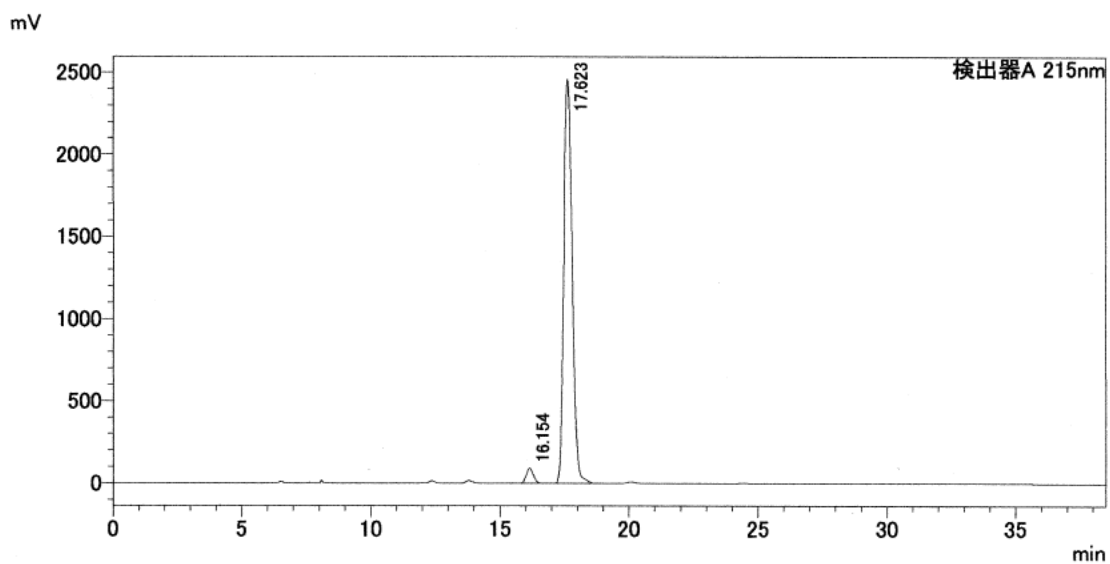
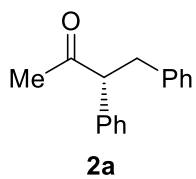
```
SAMPLE          DEC. & VT
date Jul 15 2016 dfrq      299.959
solvent CDCl3  dn         H1
file      exp  dpwr       30
ACQUISITION    dof        0
sfrq      299.959 dm       nnn
tn         H1  dmm        c
at         1.998 dmf      200
np         18008
sw         4506.5 wtfile
fb not used proc         ft
bs         4   fn       not used
tpwr       56
pw         7.0 werr
d1         1.000 wexp
tof        0   wbs       wft
nt         16  wnt       wft
ct         16
alock      n
gain not used
FLAGS
il         n
in         n
dp         y
DISPLAY
sp        -300.1
wp        3299.5
vs        116
sc         0
wc        250
hzmm     13.20
is        1212.26
rfl       765.0
rfp        0
th         31
ins       100.000
nm cdc ph
```



MF D-8 PTLC1 major 13C

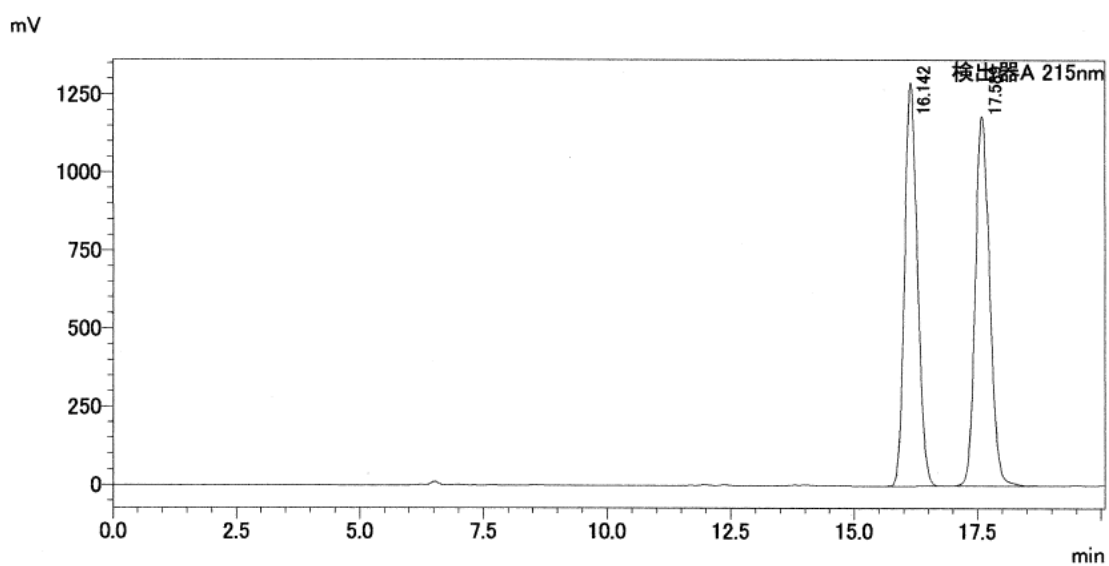
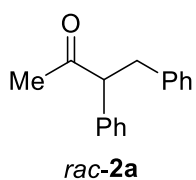
Pulse Sequence: s2pul

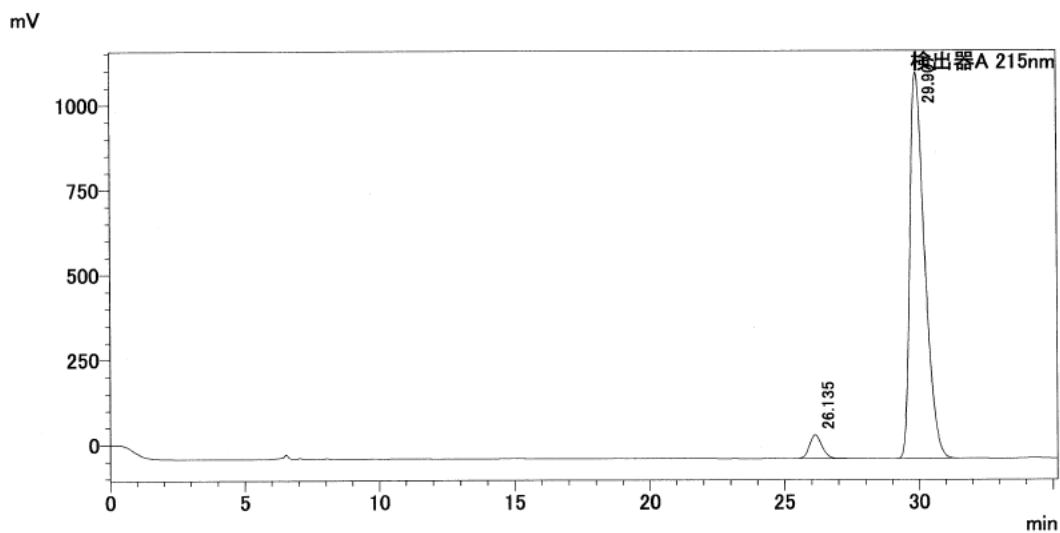
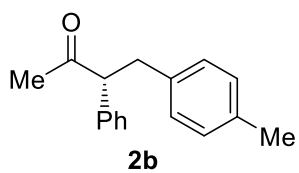




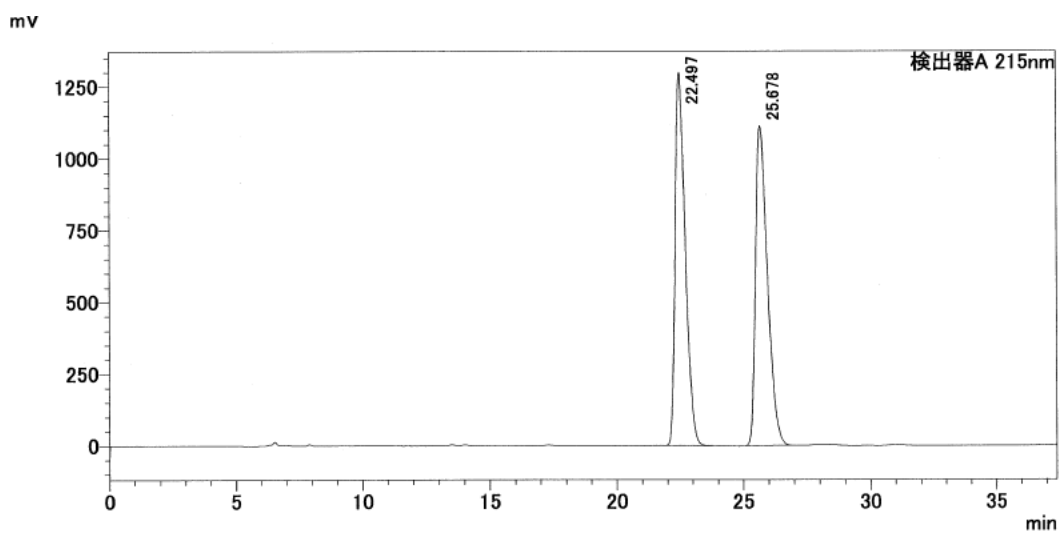
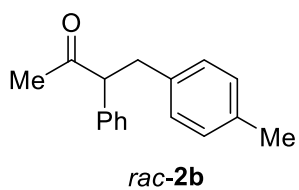
検出器A 215nm

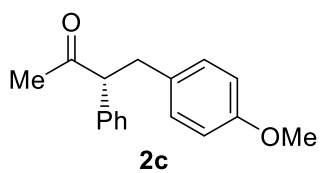
ピーク#	保持時間	面積	高さ	濃度	単位	マーク	化合物名
1	16.154	1780230	92318	3.110			
2	17.623	55469793	2458425	96.890			
合計		57250023	2550743				



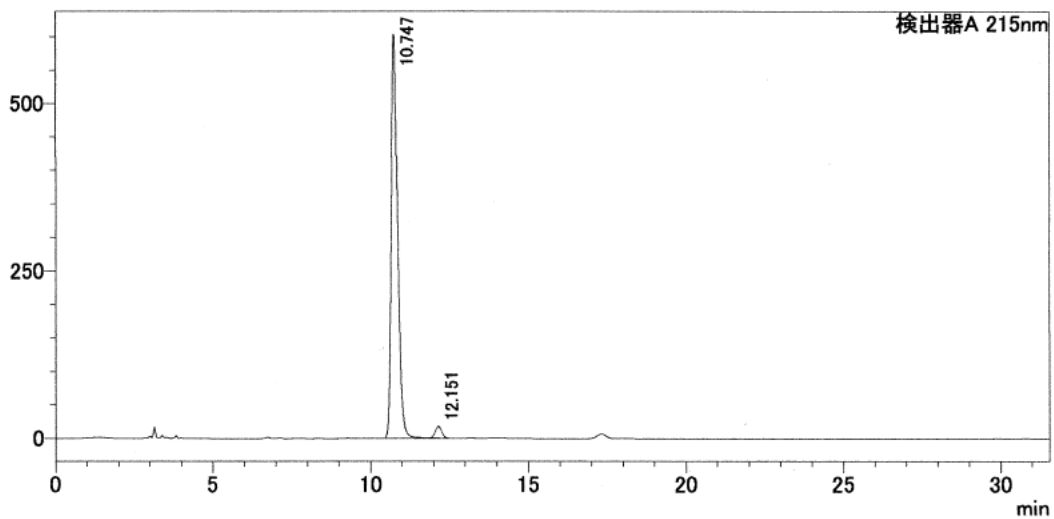


ピーク#	保持時間	面積	高さ	濃度	単位	マーク	化合物名
1	26.135	2202748	69045	4.789			
2	29.907	43795973	1136605	95.211			
合計		45998721	1205650				



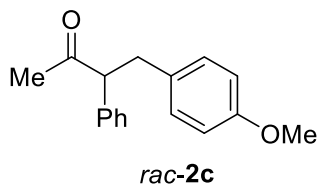


mV

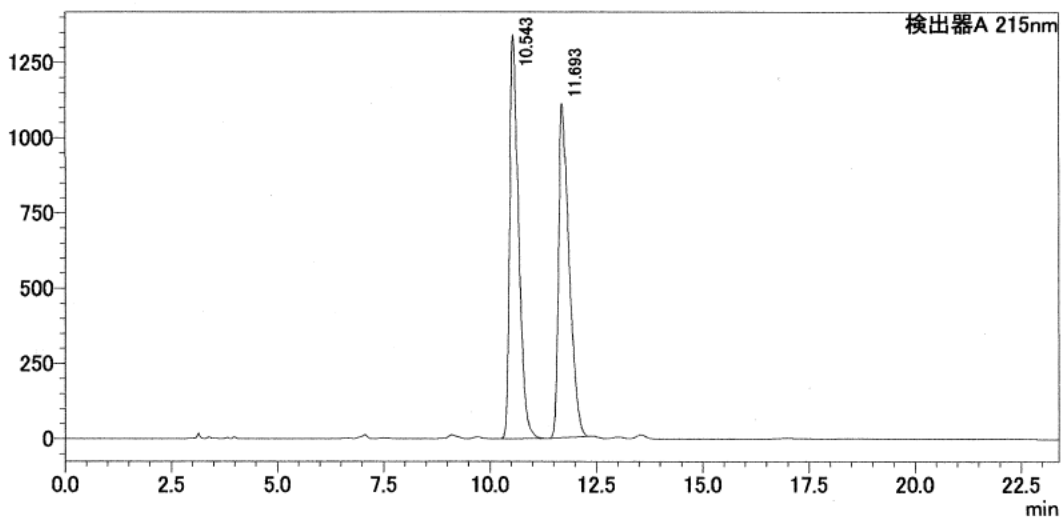


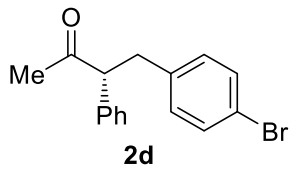
検出器A 215nm

ピーク#	保持時間	面積	高さ	濃度	単位	マーク	化合物名
1	10.747	8498327	603610	97.174		S	
2	12.151	247170	17345	2.826		T	
合計		8745497	620955				

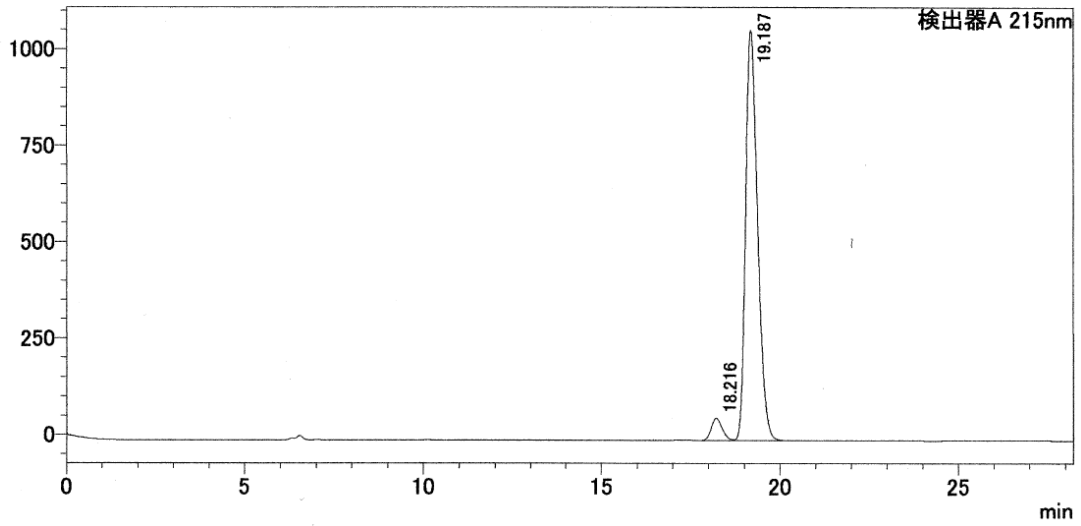


mV



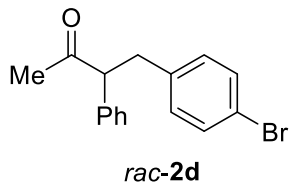


mV

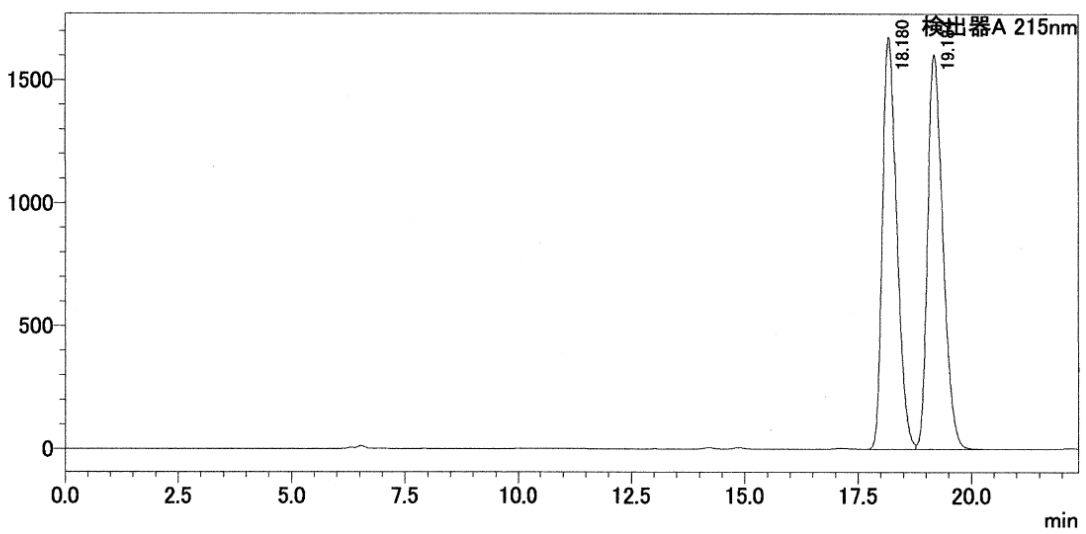


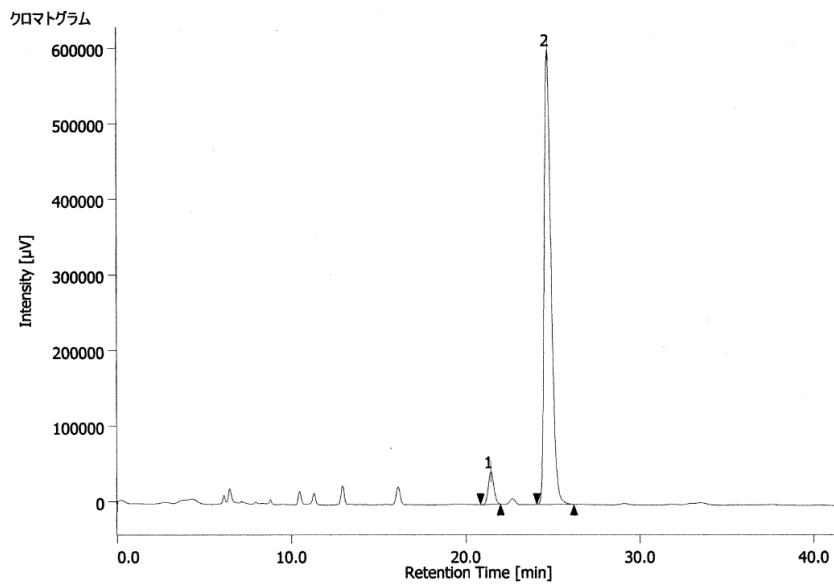
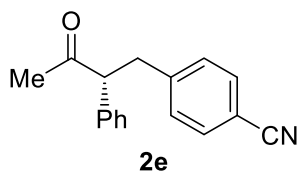
検出器A 215nm

ピーク#	保持時間	面積	高さ	濃度	単位	マーク	化合物名
1	18.216	1248264	57927	4.786			
2	19.187	24834901	1063180	95.214		V	
合計		26083166	1121107				



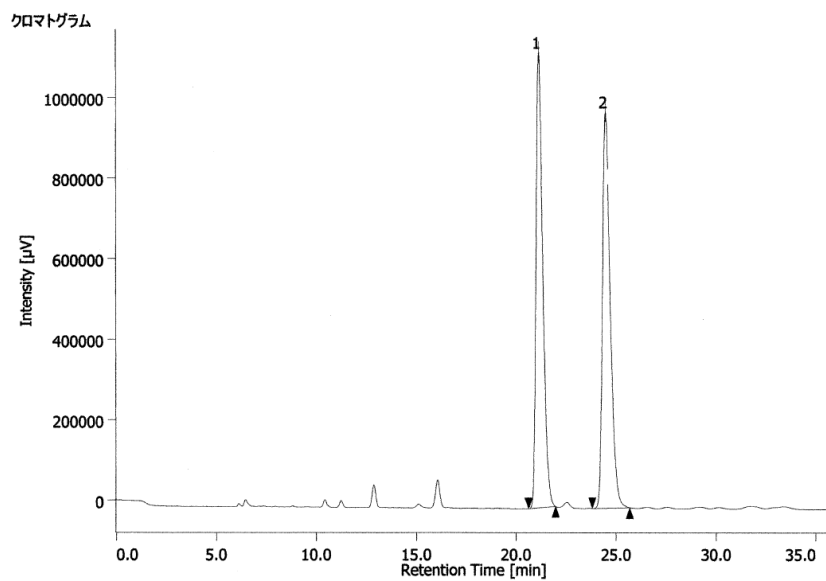
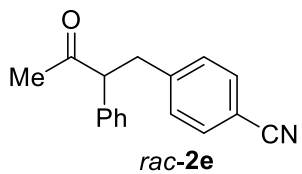
mV

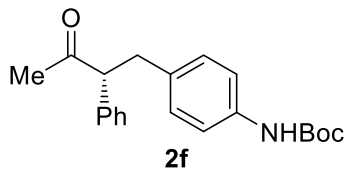




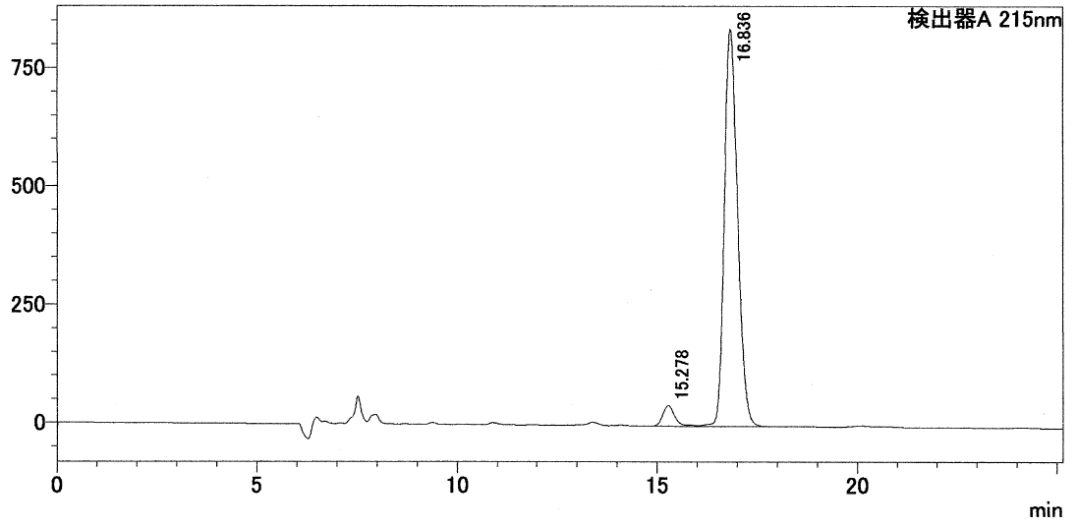
ピーク情報

ピークNo.	CH	tR [min]	面積 [$\mu\text{V}\cdot\text{sec}$]	高さ [μV]	面積%	高さ%
1	1	21.458	958253	43450	5.595	6.735
2	1	24.700	16169003	601680	94.405	93.265



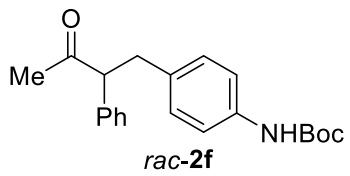


mV

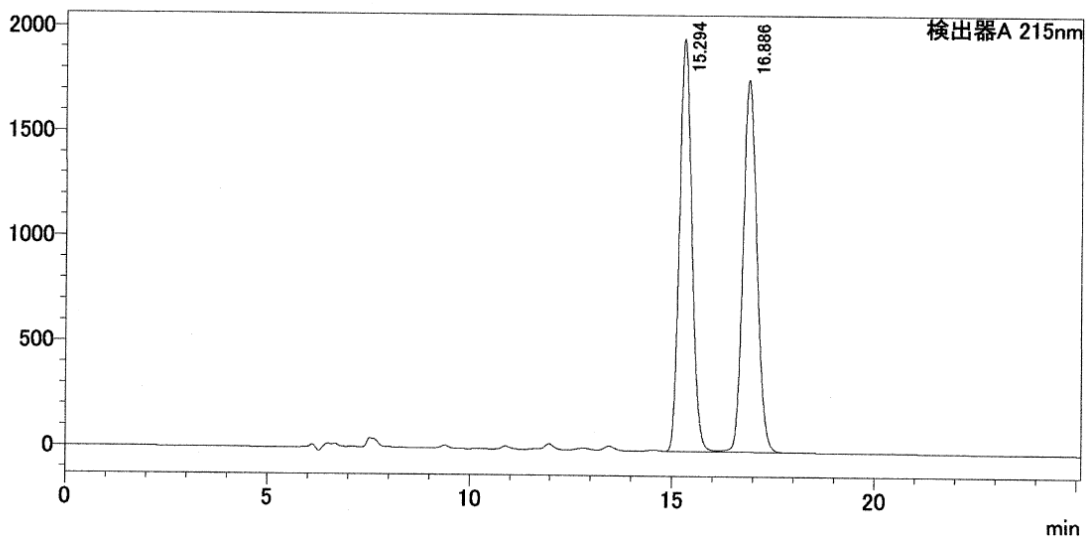


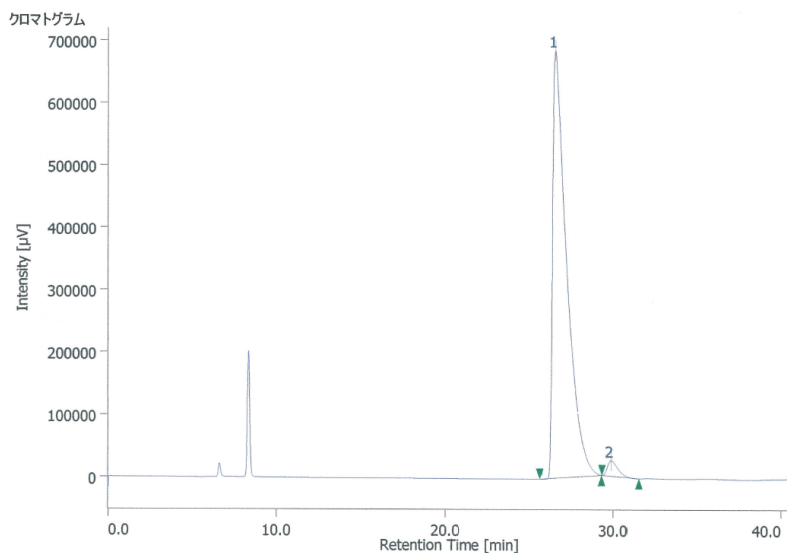
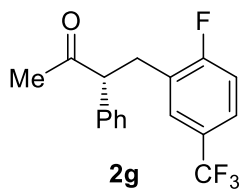
検出器A 215nm

ピーク#	保持時間	面積	高さ	濃度	単位	マーク	化合物名
1	15.278	919960	43460	4.541			
2	16.836	19340456	839970	95.459		V	
合計		20260415	883430				



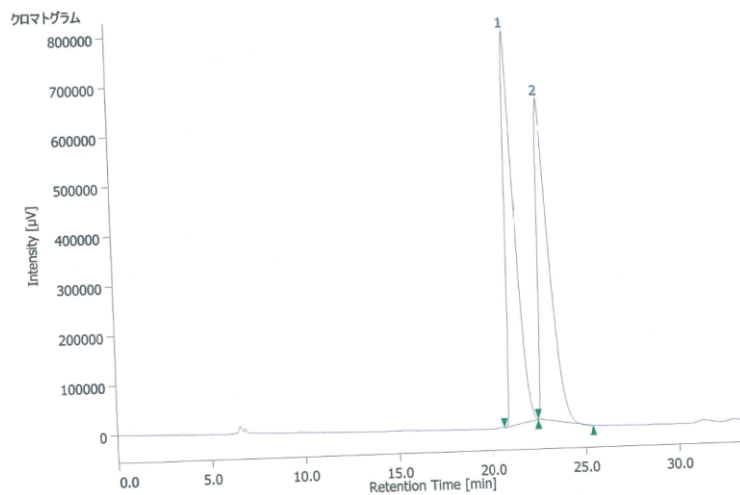
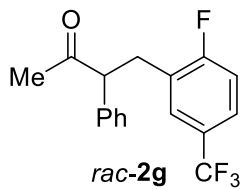
mV

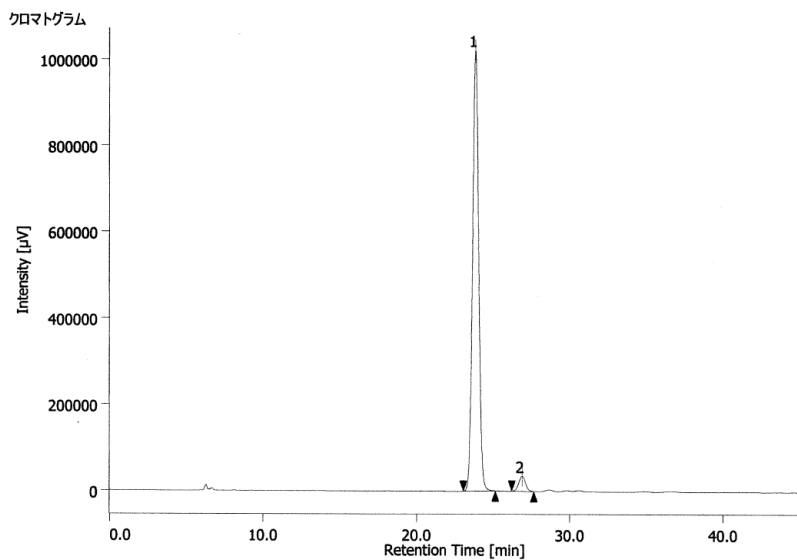
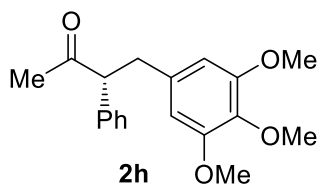




ピーク情報

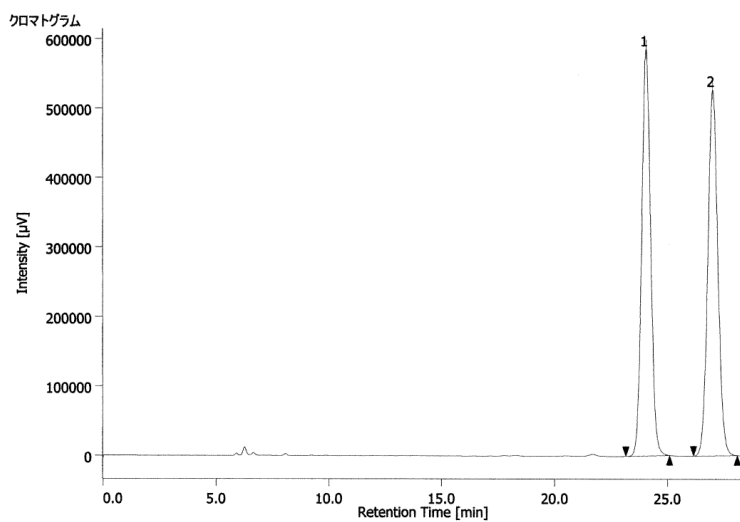
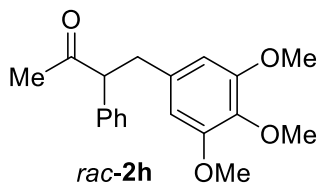
ピークNo.	CH	tR [min]	面積 [μ V \cdot sec]	高さ [μ V]	面積%	高さ%
1	1	26.533	39317243	685294	97.417	96.512
2	1	29.867	1042561	24764	2.583	3.488

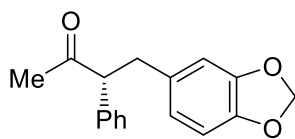




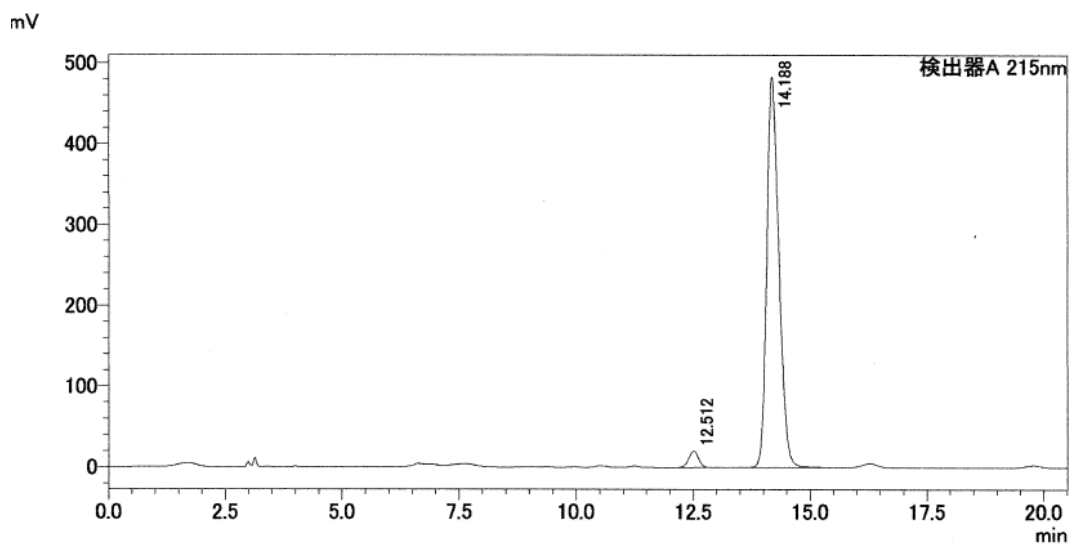
ピーク情報

ピークNo.	CH	tR [min]	面積 [$\mu\text{V}\cdot\text{sec}$]	高さ [μV]	面積%	高さ%
1	1	23.858	28004449	1022506	96.301	96.683
2	1	26.892	1075787	35077	3.699	3.317



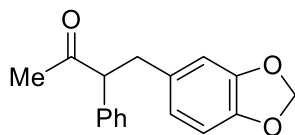


2i

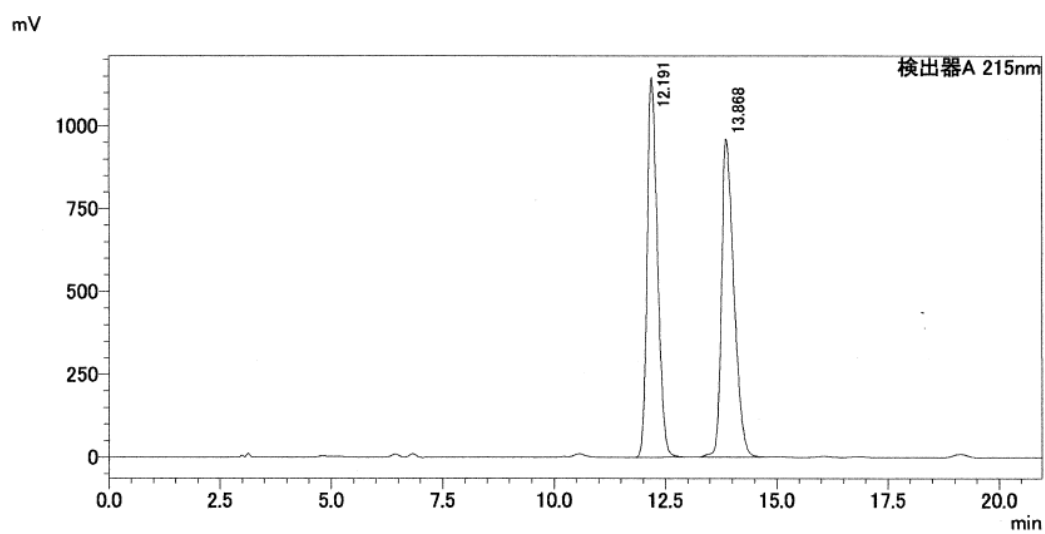


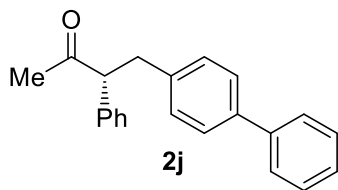
検出器A 215nm

ピーク#	保持時間	面積	高さ	濃度	単位	マーク	化合物名
1	12.512	303433	20343	3.382			
2	14.188	8667260	482779	96.618			
合計		8970693	503122				

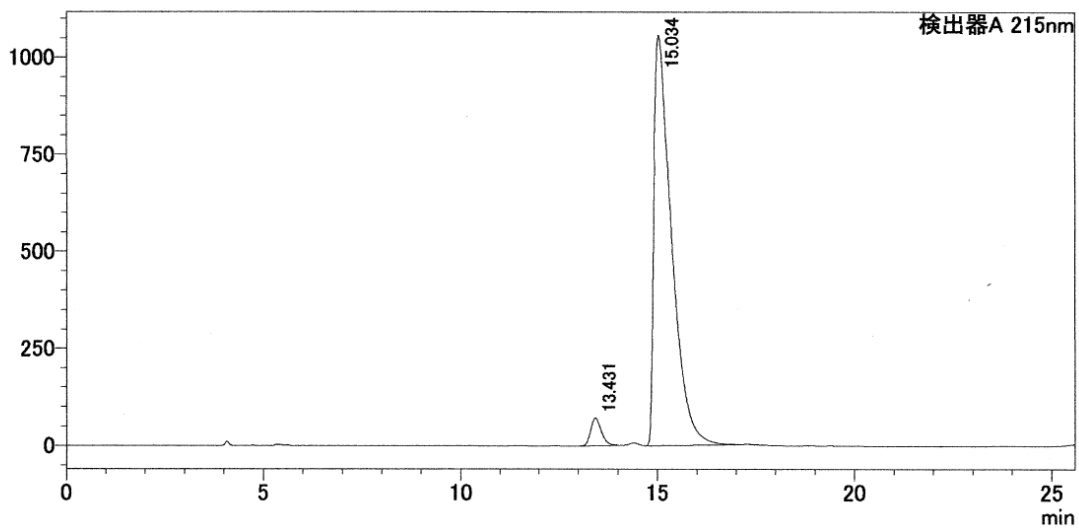


rac-2i



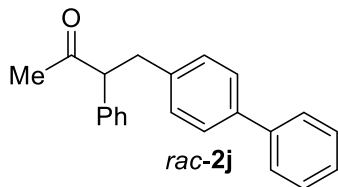


mV

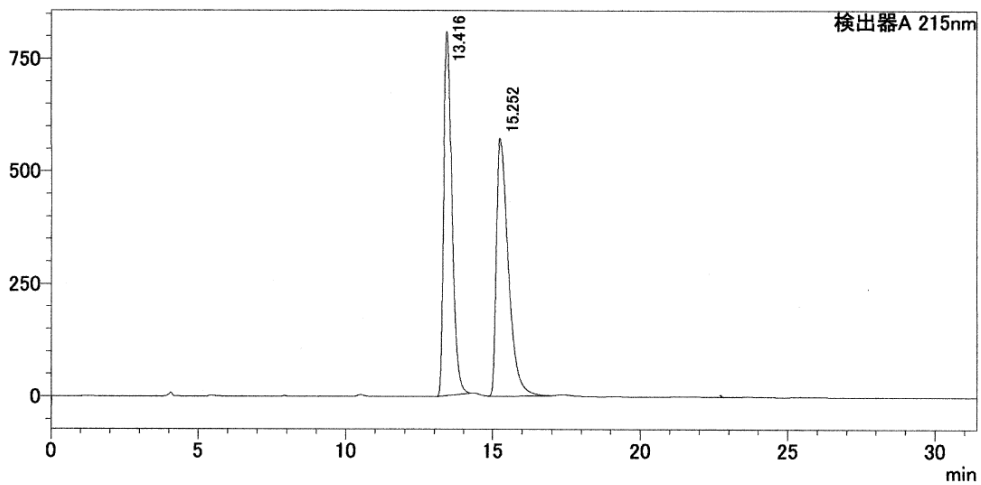


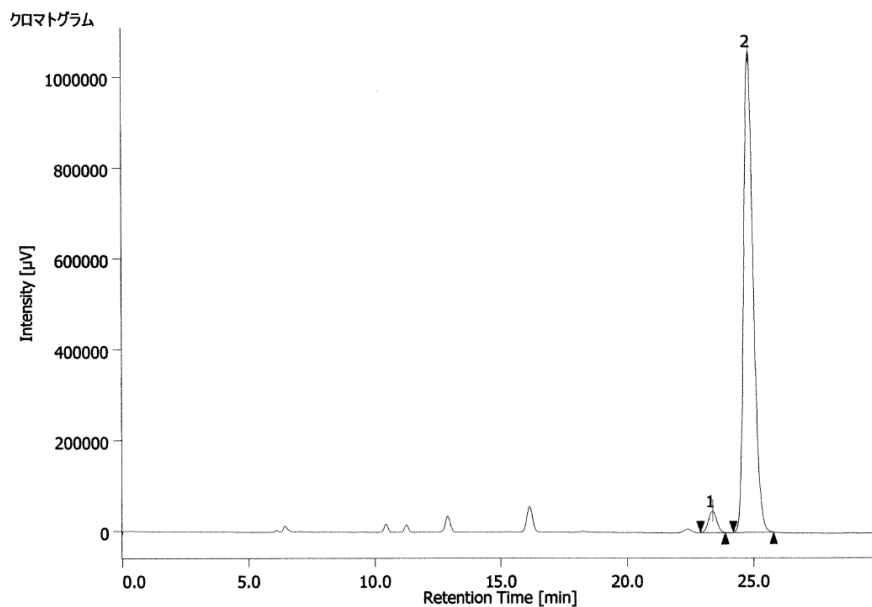
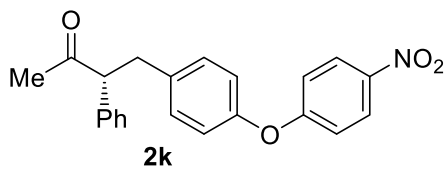
検出器A 215nm

ピーク#	保持時間	面積	高さ	濃度	単位	マーク	化合物名
1	13.431	1312220	70644	3.837			
2	15.034	32884389	1056371	96.163			
合計		34196609	1127015				



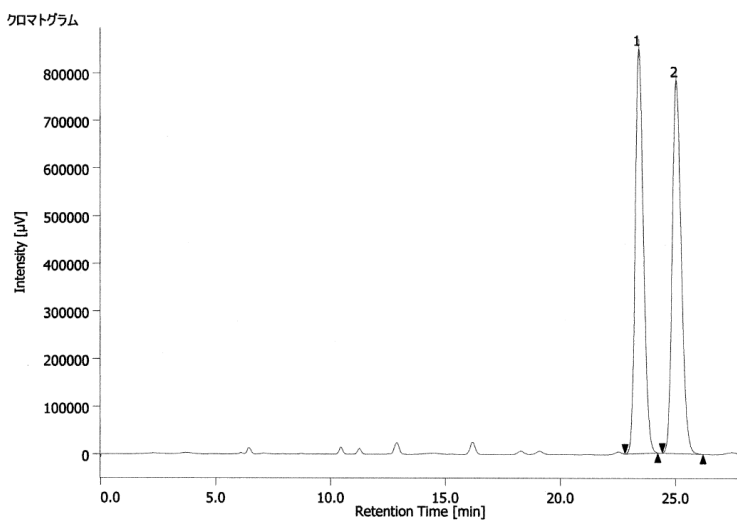
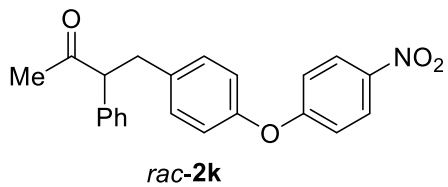
mV

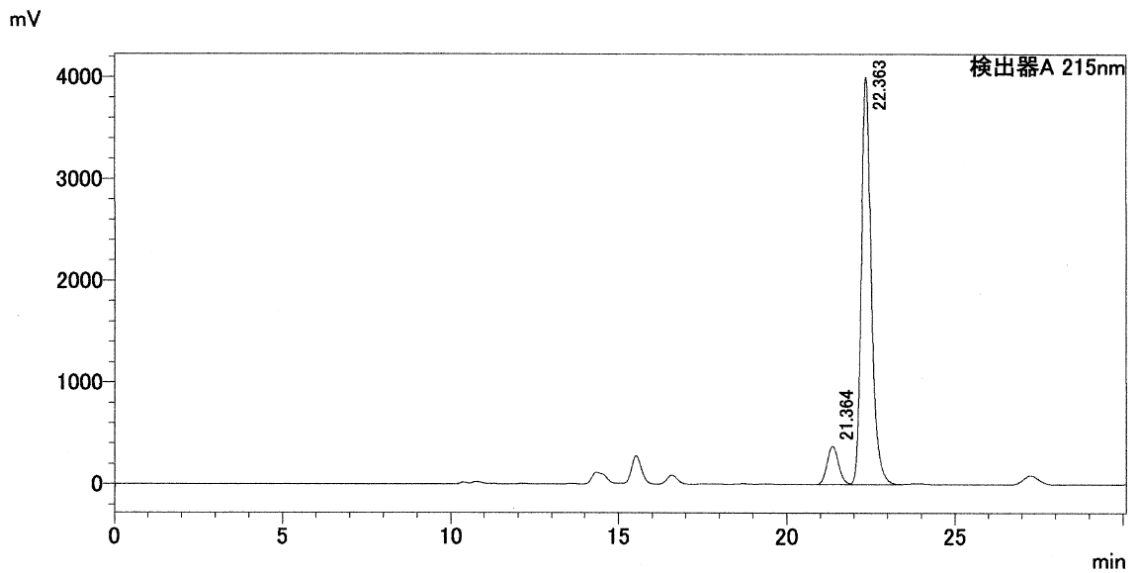
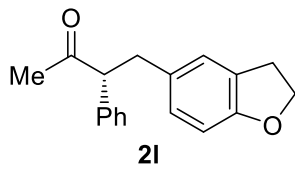




ピーク情報

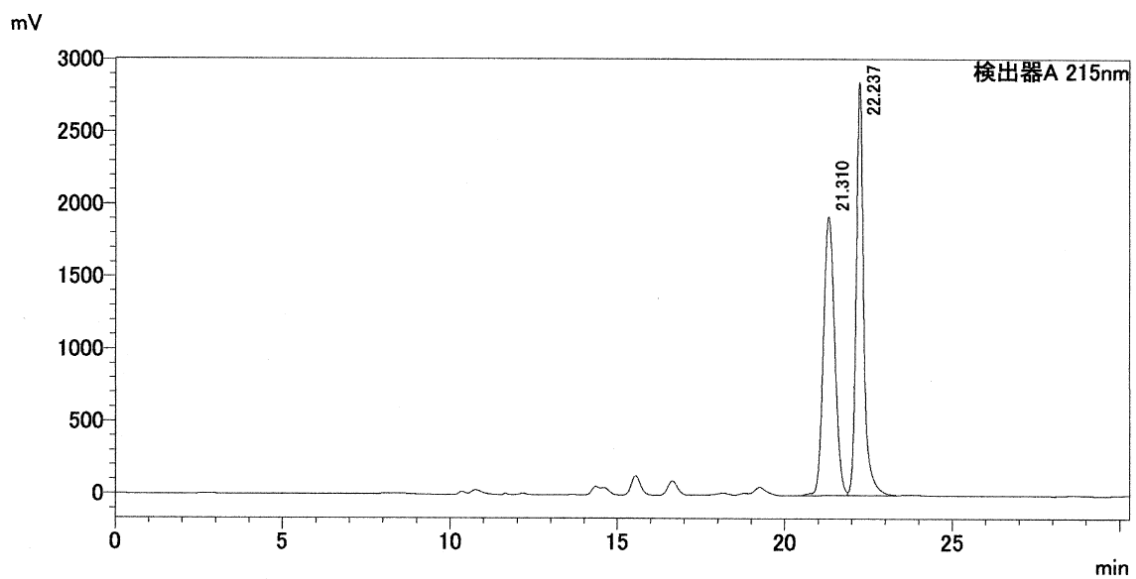
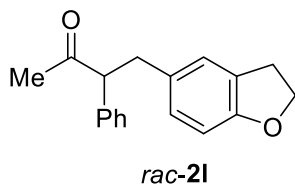
ピークNo.	CH	tR [min]	面積 [$\mu\text{V}\cdot\text{sec}$]	高さ [μV]	面積%	高さ%
1	1	23.358	1097756	47774	3.797	4.314
2	1	24.792	27814514	1059776	96.203	95.686

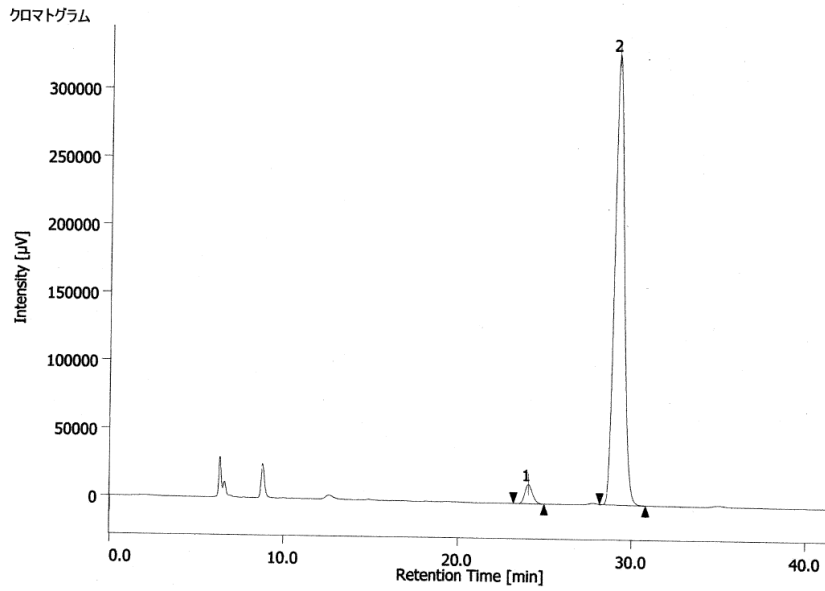
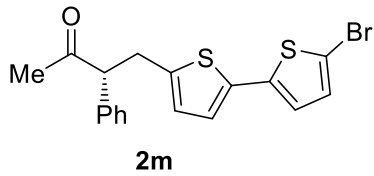




検出器A 215nm

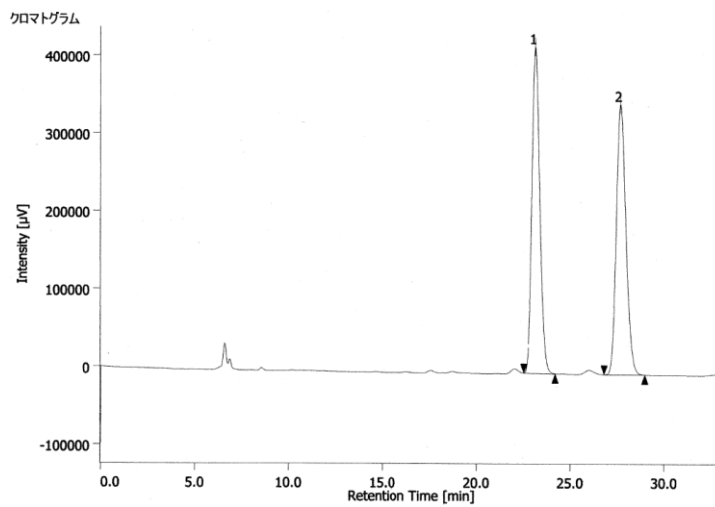
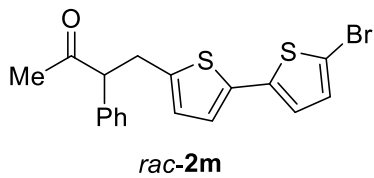
ピーク#	保持時間	面積	高さ	濃度	単位	マーク	化合物名
1	21.364	9022241	376133	9.935			
2	22.363	81789261	3997662	90.065		V	
合計		90811502	4373795				

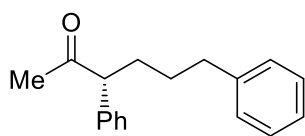




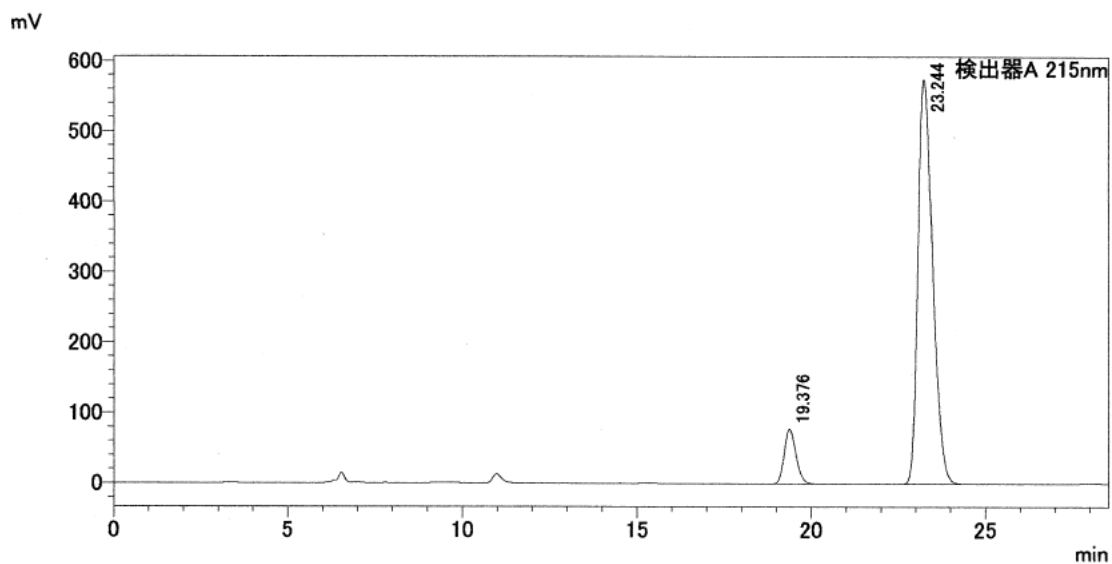
ピーク情報

ピークNo.	CH	tR [min]	面積 [$\mu\text{V}\cdot\text{sec}$]	高さ [μV]	面積%	高さ%
1	1	24.067	429255	14083	3.282	4.081
2	1	29.133	12649864	331023	96.718	95.919

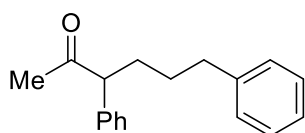




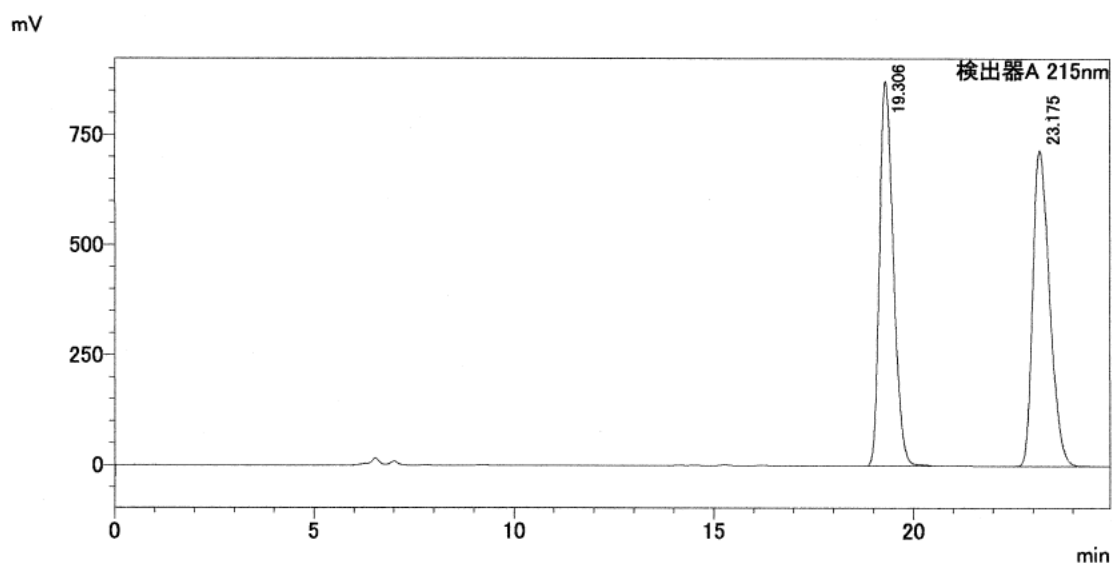
2n

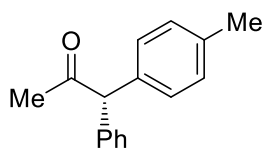


検出器A 215nm							
ピーク#	保持時間	面積	高さ	濃度	単位	マーク	化合物名
1	19.376	1838038	77569	9.809			
2	23.244	16900772	574708	90.191			
合計		18738810	652277				

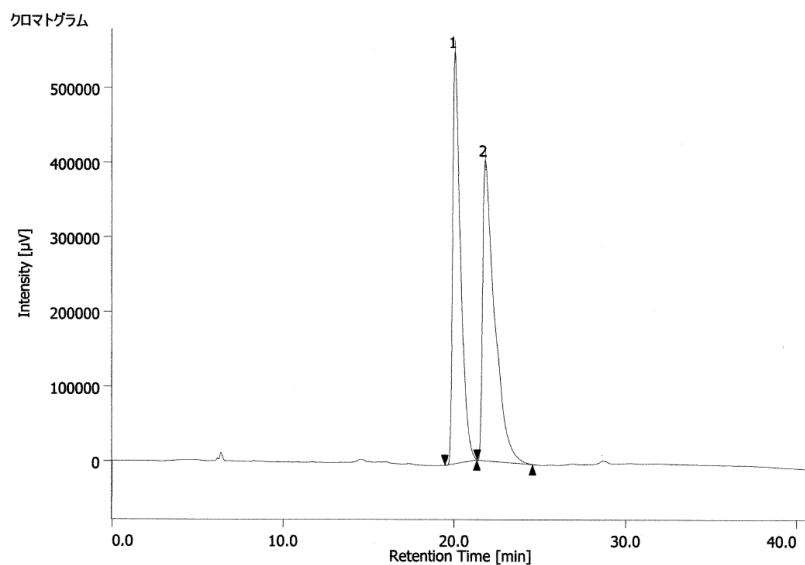


rac-2n



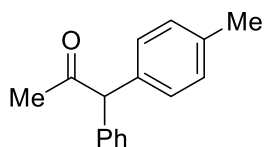


2o

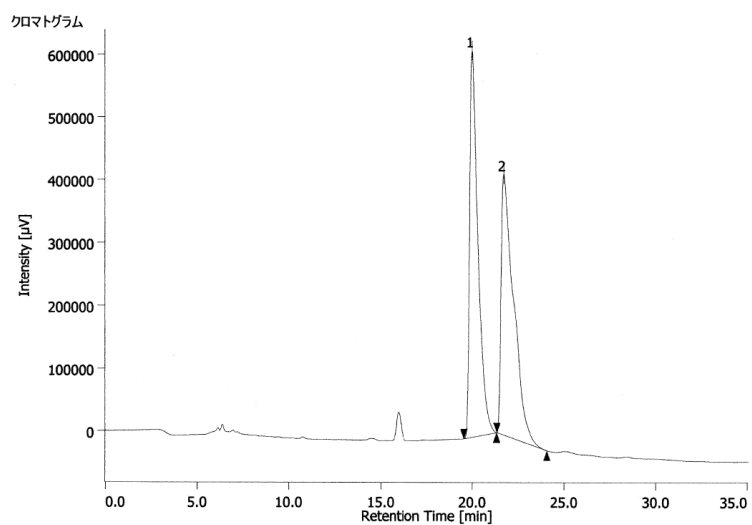


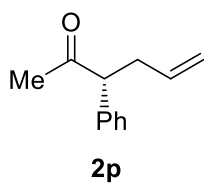
ピーク情報

ピークNo.	CH	tR [min]	面積 [μ V \cdot sec]	高さ [μ V]	面積%	高さ%
1	1	20.050	17567995	552155	47.319	57.808
2	1	21.800	19558944	402996	52.681	42.192

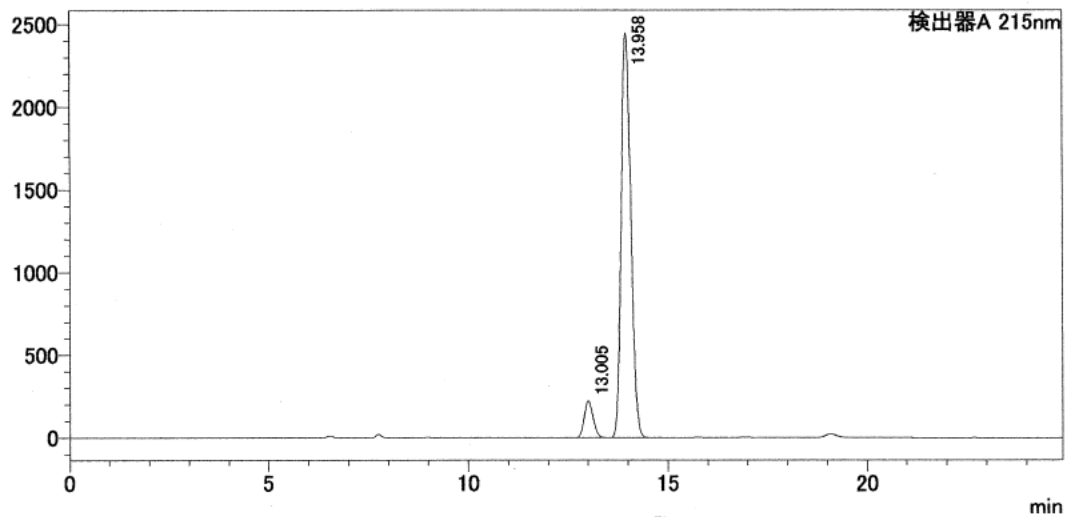


rac-2o



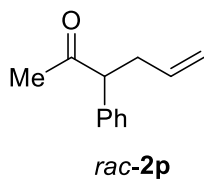


mV

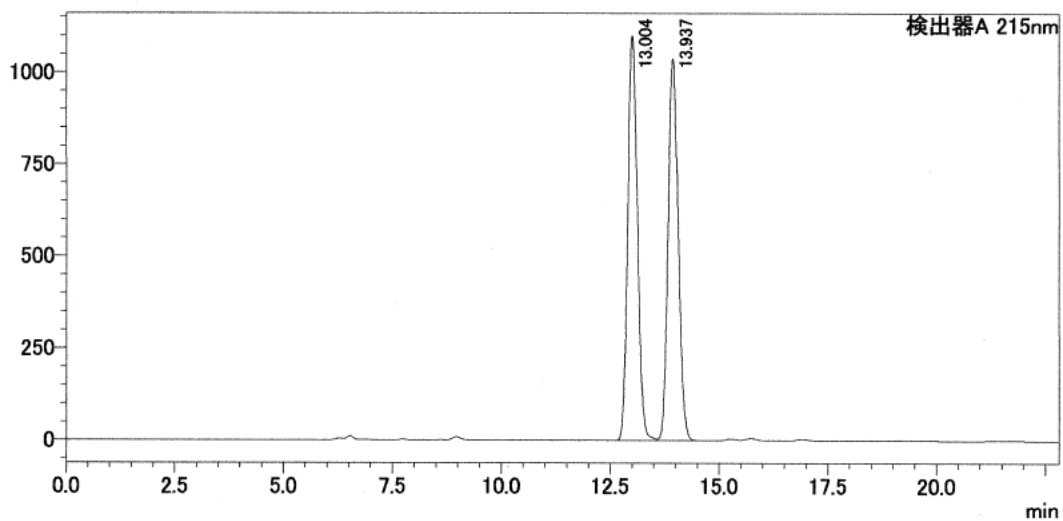


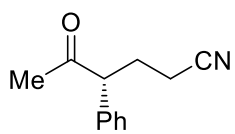
検出器A 215nm

ピーク#	保持時間	面積	高さ	濃度	単位	マーク	化合物名
1	13.005	3402608	222128	7.536			
2	13.958	41748293	2446955	92.464		V	
合計		45150901	2669083				



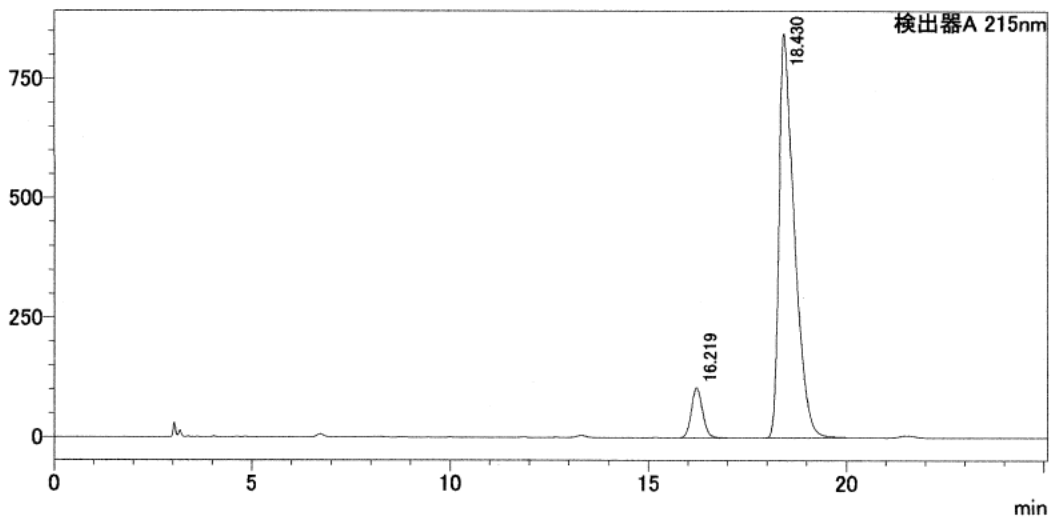
mV





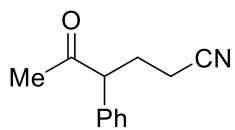
2q

mV



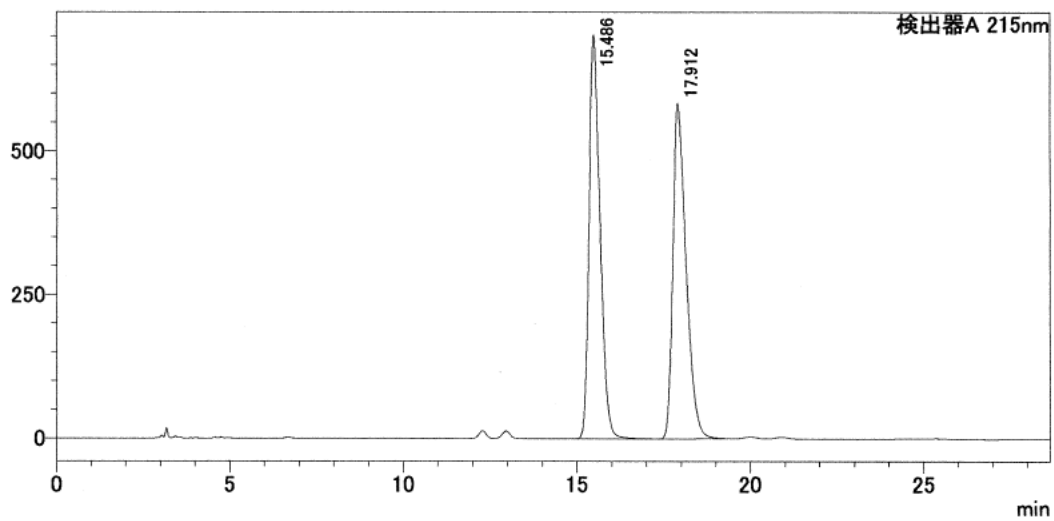
検出器A 215nm

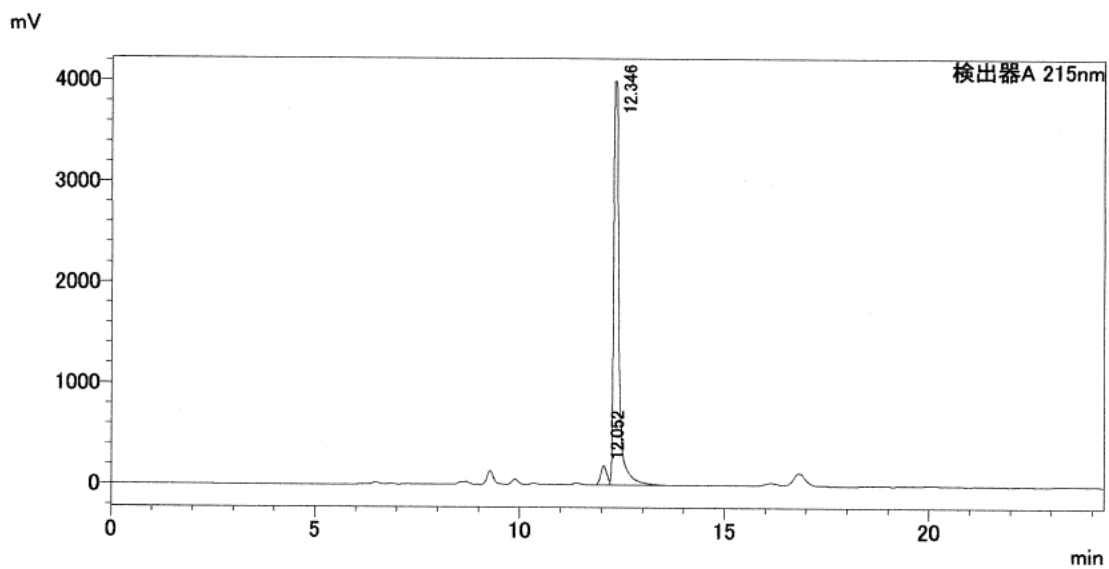
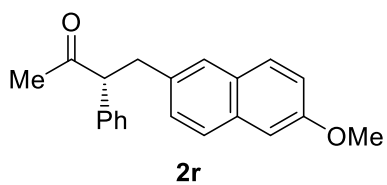
ピーク#	保持時間	面積	高さ	濃度	単位	マーク	化合物名
1	16.219	2070363	104688	8.316			
2	18.430	22826527	844659	91.684			
合計		24896890	949347				



rac-**2q**

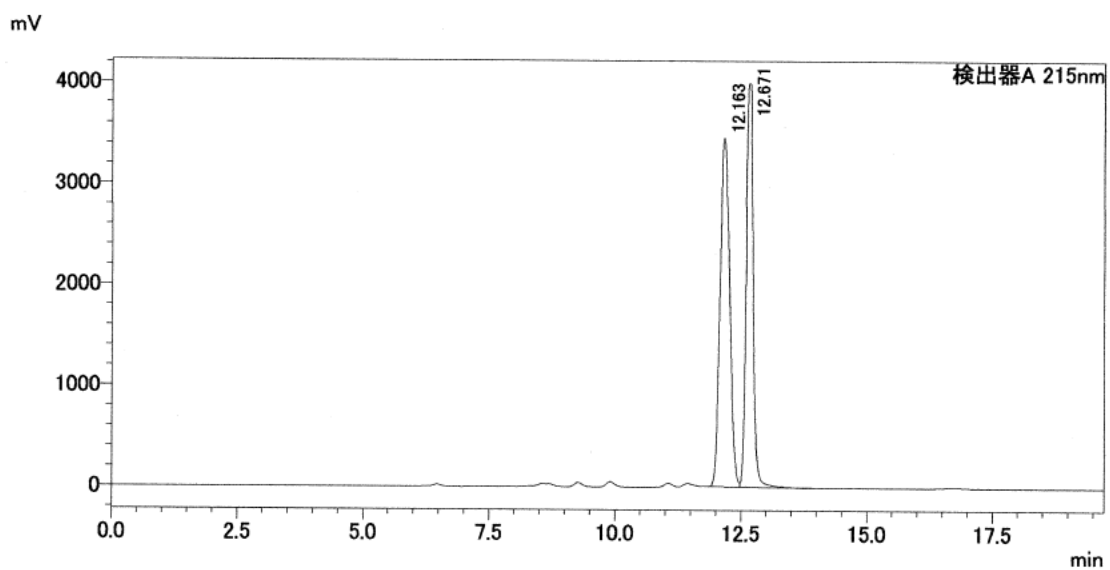
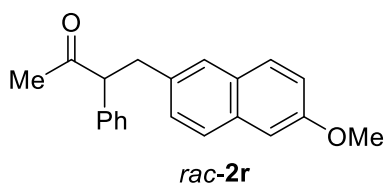
mV

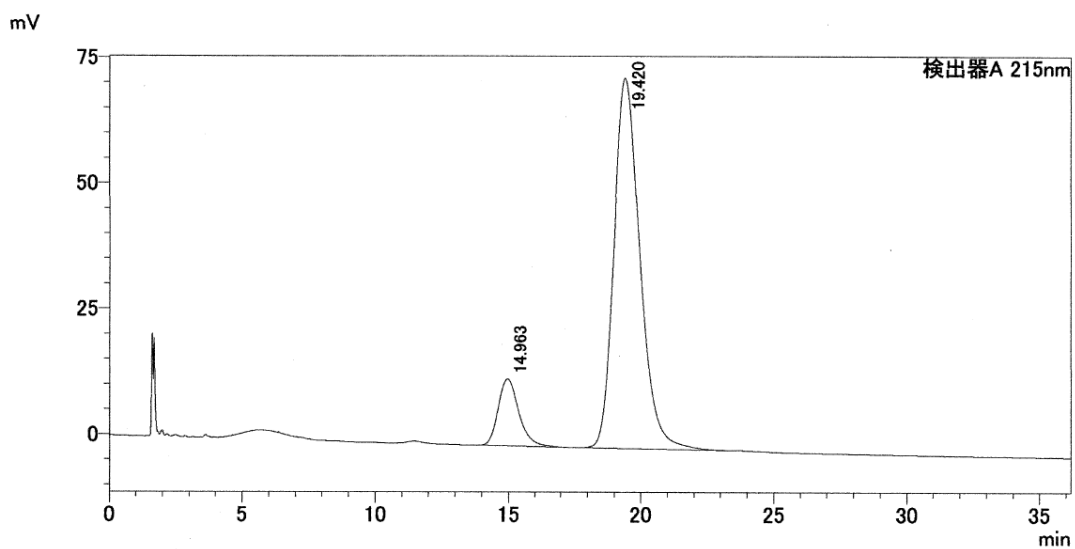
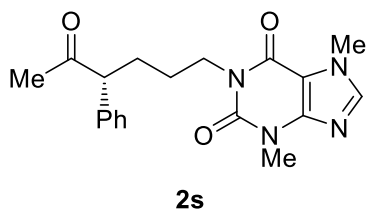




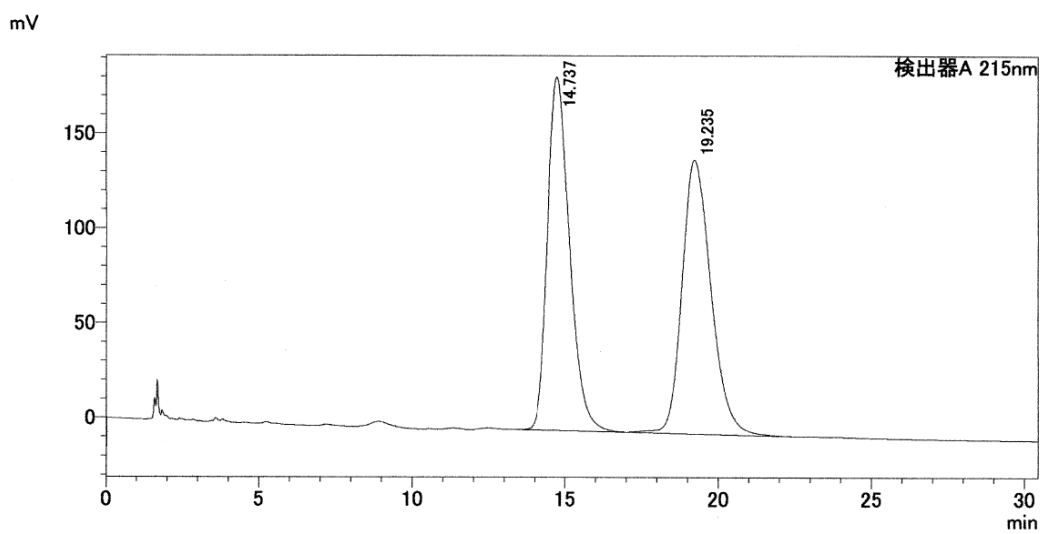
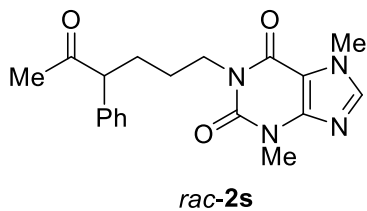
検出器A 215nm

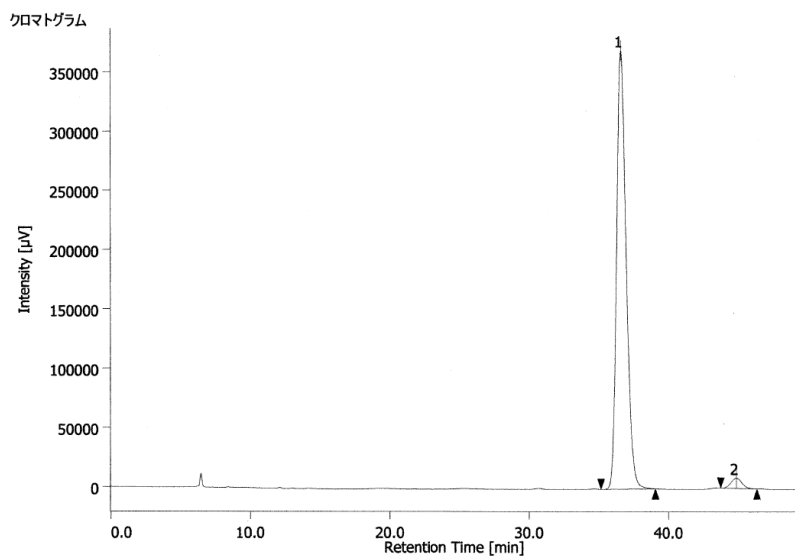
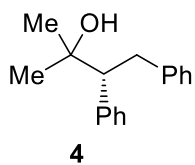
ピーク#	保持時間	面積	高さ	濃度	単位	マーク	化合物名
1	12.052	2088119	194969	5.421			
2	12.346	36434207	3999171	94.579		V	
合計		38522326	4194140				





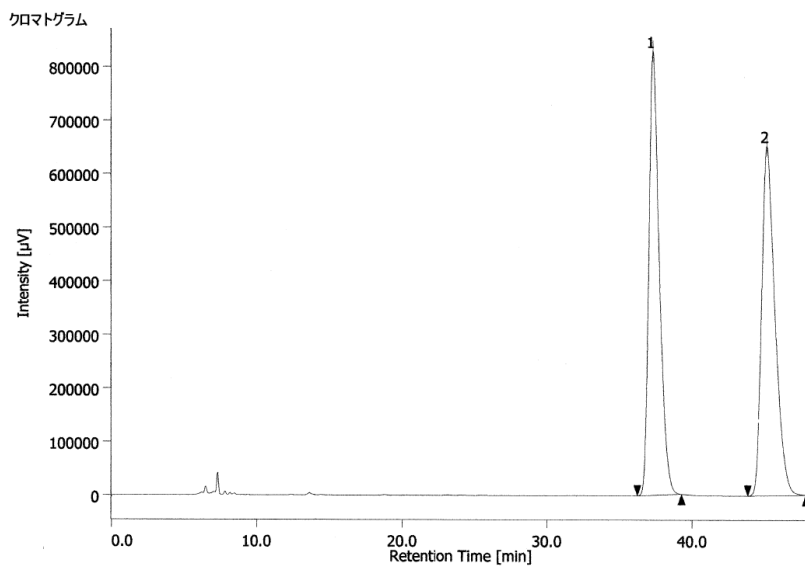
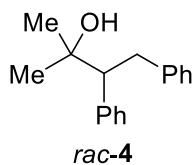
検出器A 215nm							
ピーク#	保持時間	面積	高さ	濃度	単位	マーク	化合物名
1	14.963	724244	13312	12.635			
2	19.420	5007997	73609	87.365			
合計		5732241	86921				

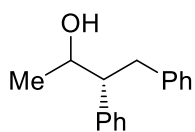




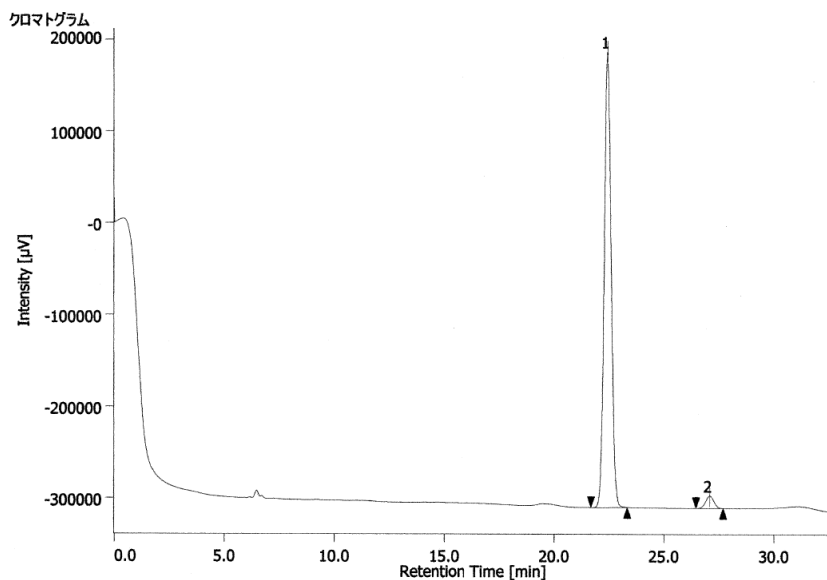
ピーク情報

ピークNo.	CH	tR [min]	面積 [$\mu\text{V}\cdot\text{sec}$]	高さ [μV]	面積%	高さ%
1	1	36.575	17417317	369939	97.380	97.716
2	1	44.833	468621	8647	2.620	2.284



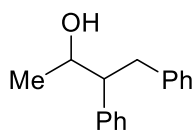


5 (major)



ピーク情報

ピークNo.	CH	tR [min]	面積 [μV·sec]	高さ [μV]	面積%	高さ%
1	1	22.425	11082620	496291	96.843	97.332
2	1	27.067	361249	13603	3.157	2.668



rac-5 (major)

