

SUPPORTING INFORMATION

Synthesis and potassium K_v7 channel opening activity of thioether analogues of the analgesic flupirtine

Christian Bock^a, Kristin Beirow^a, Abdr Rahman Shemsu Surur^a, Lukas Schulig^a, Anja Bodtke^a, Patrick J. Bednarski^a and Andreas Link^a

^aPharmaceutical and Medicinal Chemistry, Institute of Pharmacy, University of Greifswald, Friedrich-Ludwig-Jahn-Str. 17, 17487 Greifswald, Germany

Contents

6-Chloro-3-nitropyridin-2-amine.....	2
6-Amino-5-nitropyridine-2-thiol (5)	2
6-[(4-Fluorobenzyl)thio]-3-nitropyridin-2-amine (5a).....	3
6-(Benzylthio)-3-nitropyridin-2-amine (5b).....	3
6-[(3,4-Difluorobenzyl)thio]-3-nitropyridin-2-amine (5c)	3
3-Nitro-6-[[4-(trifluoromethyl)benzyl]thio]pyridin-2-amine (5d)	3
Ethyl-{2-amino-6-[(4-fluorobenzyl)thio]pyridin-3-yl}carbamate (6a)	3
Ethyl-{2-amino-6-(benzylthio)pyridin-3-yl}carbamate (6b).....	4
Ethyl-{2-amino-6-[(3,4-difluorobenzyl)thio]pyridin-3-yl}carbamate (6c).....	4
Ethyl-{2-amino-6-[[4-(trifluoromethyl)benzyl]thio]pyridin-3-yl}carbamate (6d):.....	5
Ethyl-{2-amino-6-[(4-fluorobenzyl)sulfinyl]pyridin-3-yl}carbamate (7a)	6
Ethyl-{2-amino-6-(benzylsulfinyl)pyridin-3-yl}carbamate (7b)	6
Ethyl-{2-amino-6-[(3,4-difluorobenzyl)sulfinyl]pyridin-3-yl}carbamate (7c)	6
Ethyl-{2-amino-6-[[4-(trifluoromethyl)benzyl]sulfinyl]pyridin-3-yl}carbamate (7d).....	7
Ethyl-{2-amino-6-[(4-fluorobenzyl)sulfonyl]pyridin-3-yl}carbamate (8a)	7
Ethyl-{2-amino-6-(benzylsulfonyl)pyridin-3-yl}carbamate (8b)	7
Ethyl-{2-amino-6-[(3,4-difluorobenzyl)sulfonyl]pyridin-3-yl}carbamate (8c)	8
Ethyl-{2-amino-6-[[4-(trifluoromethyl)benzyl]sulfonyl]pyridin-3-yl}carbamate (8d).....	8

Spectra of 6-chloro-3-nitropyridin-2-amine.....	9
Spectra of 5	10
Spectra of 5a	11
Spectra of 5b	12
Spectra of 5c	13
Spectra of 5d	14
Spectra of 6a	15
Spectra of 6b	16
Spectra of 6c	17
Spectra of 6d	18
Spectra of 7a	19
Spectra of 7b	20
Spectra of 7c	21
Spectra of 7d	22
Spectra of 8a	23
Spectra of 8b	24
Spectra of 8c	25
Spectra of 8d	26
Biological Evaluations.....	27
Cell culture	27
MTT assay.....	27
Kv7.2/3 channel opening assay	27
LogD _{7.4} determination	28
Table of reference compounds with known logP and measured logk'	28
Table of synthesized compounds with measured logk' and calculated logD _{7.4}	29
Cyclic voltammetry.....	30
Computational methods	31
Heteromeric Kv7.2/7.3 multimer homology modeling.....	31
Docking.....	31

Experimental

All commercial reagents and solvents were used without further purification.

Reaction control via TLC was performed on silica gel 60 F₂₅₄ aluminium plates from Merck using UV-light (254 nm and 366 nm) for visualization.

NMR spectra were recorded with a Bruker Avance III instrument at 400 MHz (¹H-NMR) and 101 MHz (¹³C-NMR), respectively. The samples were found in DMSO-*d*₆ solution with tetramethylsilane as an internal standard at 25 °C.

HRMS were obtained with an LCMS-IT-TOF (Shimadzu) by using electrospray ionization.

IR spectra were recorded on an ALPHA FT-IR spectrometer from Bruker with the help of ATR technology.

Melting points were found automatically with the Melting Point M-565 (Büchi).

The compound purities were determined by HPLC with DAD (100 % method at 220 nm).

Flash chromatography was performed (unless stated otherwise) on a glass column using silica gel 60 from Carl Roth GmbH + Co. KG with a particle size of 20–45 µm. The flow rate was adjusted to approximately 25 mL/min by applying pressure with compressed air.

6-Chloro-3-nitropyridin-2-amine: 2,6-Dichloro-3-nitropyridine (100 mmol, 19.2 g) was suspended in 200 mL ethanol and cooled to 0 °C. Ammonia gas was bubbled into the suspension for 3 hours while the temperature was kept at 0 °C. Afterwards the flask was sealed with a stopper and the reaction stirred at r.t. overnight. The product was precipitated with 400 mL water and filtered off. The yellow coloured solid was washed with 200 mL water, 200 mL *n*-hexane and dried at 70 °C in the oven. Yellow solid (m.p.: 190–192 °C). Yield: 16.2 g (93.6%). *R*_f = 0.80 (cyclohexane/ethanol/Et₃N 6:2:2). ¹H-NMR [ppm]: δ = 6.77 (d, 1H, *J* = 8.6 Hz), 8.25 (s, 2H), 8.39 (d, 1H, *J* = 8.6 Hz). ¹³C-NMR [ppm]: δ = 112.0, 126.1, 138.3, 153.4, 155.0. IR [cm⁻¹]: $\tilde{\nu}$ = 1552, 1594, 3157, 3364, 3442.

6-Amino-5-nitropyridine-2-thiol (5): A mixture of 6-chloro-3-nitropyridin-2-amine (30 mmol, 5.2 g), sulfur (0.9 eq, 27 mmol, 0.86 g), Na₂S·9H₂O (0.9 eq, 27 mmol, 6.49 g) and NaOH (1 eq, 30 mmol, 1.2 g) were suspended in 125 mL ethanol and refluxed for 2.5 hours. The crude product was evaporated to dryness and the residue dissolved in 400 mL water. The aqueous phase was extracted twice with 200 mL dichloromethane and subsequently acidified with 1 M HCl until pH 2. The product precipitated as an orange coloured solid and could be collected by filtration. Drying was achieved by heating to 70 °C in the oven for 5 hours. Orange solid (m.p.: 180–183 °C). Yield: 4.76 g (92.7%). *R*_f = 0.36 (*n*-hexane/ethyl acetate 4:6). ¹H-NMR [ppm]: δ = 6.49 (d, 1H, *J* = 9.5 Hz), 7.53 (bs, 1H), 7.92 (d, 1H, *J* = 9.5 Hz), 8.66 (bs, 1H), 12.61 (s, 1H). ¹³C-NMR [ppm]: δ = 118.8, 119.5, 131.9, 149.5, 183.8. IR [cm⁻¹]: $\tilde{\nu}$ = 1087, 1570, 3247, 3387.

General procedure for synthesizing compounds 5a–5d: 1 eq of **5** was dissolved in DMF (4 mL per mmol of **5**) and 1 eq of NaOH (as 3 M solution in water) was added to form a dark brown coloured solution. 1 eq of a suitable substituted benzyl bromide was added whereby the solution changed to orange. The reaction was stirred at r.t.

and monitored by TLC. After completion the mixture was poured into water. The resulting yellow precipitate was filtered off, washed with water and dried under reduced pressure at 40 °C.

6-[(4-Fluorobenzyl)thio]-3-nitropyridin-2-amine (5a): **5** (10 mmol, 1.71 g) was reacted with 4-fluorobenzyl bromide (10 mmol, 1.25 mL) and 4.3 mL 3 M NaOH. The reaction was completed after 3 hours. Yellow solid (m.p.: 139–141 °C). Yield: 1.98 g (70.0%). R_f = 0.35 (*n*-hexane/ethyl acetate 9:1). $^1\text{H-NMR}$ [ppm]: δ = 4.46 (s, 2H), 6.61 (d, 1H, J = 8.8 Hz), 7.13 (m, 2H), 7.53 (m, 2H), 8.15 (d, 1H, J = 8.8 Hz), 8.18 (bs, 2H). $^{13}\text{C-NMR}$ [ppm]: δ = 32.2, 110.3, 115.1 (d, 2C, $^2J_{\text{C,F}}$ = 22 Hz), 123.5, 131.1 (d, 2C, $^3J_{\text{C,F}}$ = 8 Hz), 134.2 (d, $^4J_{\text{C,F}}$ = 3 Hz), 134.5, 153.1, 160.1 and 162.5 (d, $^1J_{\text{C,F}}$ = 242 Hz), 166.3. IR [cm^{-1}]: $\tilde{\nu}$ = 1342, 1558, 3332, 3456.

6-(Benzylthio)-3-nitropyridin-2-amine (5b): **5** (3 mmol, 0.51 g) was reacted with benzyl bromide (3 mmol, 0.36 mL) and 1.3 mL 3 M NaOH. The reaction was completed after 3 hours. Yellow solid (m.p.: 151–153 °C). Yield: 0.65 g (82.9%). R_f = 0.53 (*n*-hexane/ethyl acetate 9:1). $^1\text{H-NMR}$ [ppm]: δ = 4.47 (s, 2H), 6.62 (d, 1H, J = 8.8 Hz), 7.22–7.26 (m, 1H), 7.29–7.33 (m, 2H), 7.47 (d, 2H, J = 7.2 Hz), 8.08–8.23 (bs, 2H), 8.16 (d, 1H, J = 8.7 Hz). $^{13}\text{C-NMR}$ [ppm]: δ = 33.1, 110.2, 123.4, 127.1, 128.4 (2C), 129.1 (2C), 134.4, 137.7, 153.1, 166.6. IR [cm^{-1}]: $\tilde{\nu}$ = 1144, 1345, 1560, 3335, 3459.

6-[(3,4-Difluorobenzyl)thio]-3-nitropyridin-2-amine (5c): **5** (5 mmol, 0.85 g) was reacted with 3,4-difluorobenzyl bromide (5 mmol, 0.64 mL) and 2.2 mL 3 M NaOH. After 2 hours additional 3,4-difluorobenzyl bromide (1 mmol, 0.13 mL) were added. After further 2 hours the reaction was completed. Yellow solid (m.p.: 135–138 °C). Yield: 1.44 g (97.0%). R_f = 0.75 (cyclohexane/ethanol/ Et_3N 6:2:2). $^1\text{H-NMR}$ [ppm]: δ = 4.44 (s, 2H), 6.61 (d, 1H, J = 8.8 Hz), 7.34–7.38 (m, 2H), 7.59–7.65 (m, 2H), 8.16 (d, 1H, J = 8.8 Hz), 8.21 (bs, 2H). $^{13}\text{C-NMR}$ [ppm]: δ = 31.9, 110.3, 117.3 (d, $^2J_{\text{C,F}}$ = 17 Hz), 118.2 (d, $^2J_{\text{C,F}}$ = 17 Hz), 123.5, 126.1 (dd, $^3J_{\text{C,F}}$ = 6 Hz, $^4J_{\text{C,F}}$ = 3 Hz), 134.6, 136.1 (dd, $^3J_{\text{C,F}}$ = 6 Hz, $^4J_{\text{C,F}}$ = 4 Hz), 147.3 and 149.7 (dd, $^1J_{\text{C,F}}$ = 245 Hz, $^2J_{\text{C,F}}$ = 13 Hz), 147.8 and 150.2 (dd, $^1J_{\text{C,F}}$ = 245 Hz, $^2J_{\text{C,F}}$ = 13 Hz), 153.1, 165.8. IR [cm^{-1}]: $\tilde{\nu}$ = 1149, 1165, 1346, 1560, 3331, 3464.

3-Nitro-6-[[4-(trifluoromethyl)benzyl]thio]pyridin-2-amine (5d): **5** (5 mmol, 0.85 g) was reacted with 4-(trifluoromethyl)benzyl bromide (5 mmol, 0.77 mL) and 2.2 mL 3 M NaOH. After 45 min additional of 4-(trifluoromethyl)benzyl bromide (1.5 mmol, 0.23 mL) were added. The reaction was completed after further 30 min. Yellow solid (m.p.: 128–130 °C). Yield: 1.67 g (101.2%, due to some bisalkylated by-product). $^1\text{H-NMR}$ [ppm]: δ = 4.55 (s, 2H), 6.63 (d, 1H, J = 8.8 Hz), 7.67 (d, 2H, J = 8.2 Hz), 7.74 (d, 2H, J = 8.2 Hz), 8.16 (d, 1H, J = 8.8 Hz), 8.19 (bs, 2H). $^{13}\text{C-NMR}$ [ppm]: δ = 32.3, 110.2, 122.8 and 125.5 (d, 2H, $^1J_{\text{C,F}}$ = 270 Hz), 123.5, 125.1 (q, 2C, $^4J_{\text{C,F}}$ = 4 Hz), 127.6 (q, 2C, $^2J_{\text{C,F}}$ = 32 Hz), 129.9 (2C), 134.5, 143.1, 153.0, 165.7. IR [cm^{-1}]: $\tilde{\nu}$ = 1122, 1341, 1560, 3335, 3460.

Ethyl-{2-amino-6-[(4-fluorobenzyl)thio]pyridin-3-yl}carbamate (6a):

Method A: A three-neck round-bottom flask was equipped with **5a** (3.65 mmol, 1.02 g), $\text{SnCl}_2 \cdot 2\text{H}_2\text{O}$ (5.1 eq, 18.6 mmol, 4.2 g) and 40 mL ethanol. The mixture was put under an argon atmosphere and heated to 70 °C for 25 hours. Afterwards, the temperature was lowered to 40 °C and Et_3N (10.2 eq, 37.2 mmol, 5.16 mL), as well as ethyl chloroformate (1.26 eq, 4.6 mmol 0.44 mL), were added slowly to the suspension. After 30 min the temperature was increased to 55 °C and kept for one additional hour. After cooling to r.t. the mixture was filtered

through a pad of celite and washed with ethanol. The filtrate was evaporated to dryness and distributed between water and dichloromethane. The organic layer was evaporated to dryness and the residue washed with *n*-hexane. White solid (m.p.: 138–140 °C). Yield: 0.86 g (73.0%).

Method B: **5a** (5 mmol, 1.4 g) was suspended in 30 mL water and 50 mL 2-propanol. Iron powder (3 eq, 15 mmol, 0.84 g) and NH₄Cl (4.5 eq, 22.5 mmol, 1.2 g) were added and the suspension refluxed for 5 hours. The resulting dark brown mixture was filtered and evaporated to dryness. The residue was diluted with 150 mL water and extracted three times with 60 mL ethyl acetate. The combined organic layers were washed with 150 mL saturated brine and dried over Na₂SO₄. Et₃N (1.4 eq, 7 mmol, 0.97 mL) was added and the solution cooled to 0 °C. A solution of ethyl chloroformate (1.2 eq, 6 mmol, 0.57 mL) in 5 mL ethyl acetate was added drop wise. After 2 hours additional ethyl chloroformate (1.2 eq, 6 mmol, 0.57 mL) were added at r.t., 5 hours later, ethyl chloroformate (0.6 eq, 3 mmol, 0.29 mL) and Et₃N (0.7 eq, 3.5 mmol, 0.49 mL) were added. After 2 hours the reaction was completed and the solution washed with 200 mL water and 200 mL saturated brine. After drying with Na₂SO₄ and evaporation to dryness the resulting solid was recrystallized from DCM. The product was dried at 40 °C under reduced pressure. Brown solid (m.p.: 140–142). Yield: 0.75 g (47%). Compound purity: 100.0%. *R_f* = 0.83 (cyclohexane/ ethanol/Et₃N 6:2:2). ¹H-NMR [ppm]: δ = 1.22 (t, 3H, *J* = 7.1 Hz), 4.09 (q, 2H, *J* = 7.1 Hz), 4.29 (s, 2H), 5.94 (s, 2H), 6.43 (d, 1H, *J* = 8.0 Hz), 7.07–7.12 (m, 2 H), 7.40–7.45 (m, 3H), 8.57 (m, 1H). ¹³C-NMR [ppm]: δ = 14.5, 32.6, 60.3, 109.3, 115.0 (d, 2C, ²*J*_{C,F} = 21 Hz), 115.2, 130.7 (d, 2C, ³*J*_{C,F} = 8 Hz), 131.8, 135.0 (d, ⁴*J*_{C,F} = 3 Hz), 150.2, 152.8, 154.4, 159.9 and 162.3 (d, ¹*J*_{C,F} = 243 Hz). IR [cm⁻¹]: $\tilde{\nu}$ = 1454, 1678, 2984, 3145, 3260, 3403. HRMS: [C₁₅H₁₆N₃O₂FS + H]⁺ req.: 322.1020, found: 322.1027.

Ethyl-[2-amino-6-(benzylthio)pyridin-3-yl]carbamate (6b): **5b** (2.31 mmol, 0.60 g) was suspended in a mixture of 15 mL water and 25 mL 2-propanol. Iron powder (3 eq, 6.93 mmol, 0.39 g) and NH₄Cl (4.5 eq, 10.4 mmol, 0.56 g) were added and the suspension refluxed for 6 hours. The resulting dark brown mixture was filtered and the residue washed with 50 mL 2-propanol. Et₃N (6.0 eq, 13.9 mmol, 1.91 mL) and ethyl chloroformate (2.4 eq, 5.54 mmol, 0.53 mL) were added. The solution was stirred at 40 °C and reaction progress monitored by TLC. After 1 hour and after 2 hours ethyl chloroformate (2.4 eq, 5.54 mmol, 0.53 mL) were added to complete the reaction. The solution was evaporated to dryness and the resulting solid dissolved in 100 mL ethyl acetate. The organic phase was washed with 2×100 mL water as well as 100 mL saturated brine. After drying with Na₂SO₄ the crude product was purified by flash chromatography (mobile phase: *n*-hexane/ethyl acetate 7:3). The resulting brown solid was washed with small quantities of ice-cold diethyl ether and dried at 40 °C under reduced pressure. Light blue solid (m.p.: 100–101 °C). Yield: 0.23 g (33.0%). Compound purity: 100.0%. *R_f* = 0.72 (cyclohexane/ethanol/Et₃N 6:2:2). ¹H-NMR [ppm]: δ = 1.22 (t, 3H, *J* = 7.1 Hz), 4.09 (q, 2H, *J* = 7.1 Hz), 4.30 (s, 2H), 5.93 (s, 2H), 6.44 (d, 1H, *J* = 8.0 Hz), 7.21 (t, 1H, *J* = 7.3 Hz), 7.28 (t, 2H, *J* = 7.3 Hz), 7.39–7.41 (m, 3H), 8.57 (s, 1H). ¹³C-NMR [ppm]: δ = 14.5, 33.5, 60.3, 109.2, 115.2, 126.8, 128.3 (2C), 128.8 (2C), 131.8, 138.6, 150.5, 152.7, 154.4. IR [cm⁻¹]: $\tilde{\nu}$ = 1457, 1681, 2975, 3140, 3281, 3405. HRMS: [C₁₅H₁₇N₃O₂S + H]⁺ calculated: 304.1114, found: 304.1099.

Ethyl-[2-amino-6-[(3,4-difluorobenzyl)thio]pyridin-3-yl]carbamate (6c): **5c** (5 mmol, 1.49 g) was suspended in a mixture of 30 mL water and 50 mL 2-propanol. Iron powder (3 eq, 15 mmol, 0.84 g) and NH₄Cl (4.5 eq,

22.5 mmol, 1.2 g) were added and the suspension refluxed for 3.5 hours. The resulting dark brown mixture was filtered and the residue washed with 100 mL 2-propanol. Et₃N (3.0 eq, 15 mmol, 2.07 mL) and ethyl chloroformate (1.2 eq, 6 mmol, 0.57 mL) were added. The solution was stirred for 2 hours at 40 °C. Since TLC monitoring showed no complete conversion ethyl chloroformate (0.6 eq, 3 mmol, 0.29 mL) were added and additional ethyl chloroformate (0.5 eq, 2.5 mmol, 0.24 mL) as well as Et₃N (1 eq, 5 mmol, 0.69 mL) after further 2 hours. After 5 hours the reaction was completed and the solution evaporated to dryness. The solid was dissolved in 100 mL ethyl acetate and the solution washed with 2×100 mL water as well as 100 mL saturated brine. After drying with Na₂SO₄ the crude product was dissolved in 50 mL isopropanol and precipitated with 100 mL water. The crude product was purified by flash chromatography (mobile phase: *n*-hexane/ethyl acetate 7:3) and dried at 40 °C under reduced pressure. White solid (m.p.: 137–140 °C). Yield: 0.95 g (56.0%). Compound purity: 100.0%. *R*_f = 0.39 (*n*-hexane/ethyl acetate 7:3). ¹H-NMR [ppm]: δ = 1.22 (t, 3H, *J* = 7.1 Hz), 4.09 (q, 2H, *J* = 7.1 Hz), 4.29 (s, 2H), 5.99 (s, 2H), 6.42 (d, 1H, *J* = 8.0 Hz), 7.24–7.36 (m, 2H), 7.4 (d, 1H, *J* = 6.3 Hz), 7.45–7.50 (m, 1H), 8.57 (s, 1H). ¹³C-NMR [ppm]: δ = 14.5, 32.2, 60.4, 109.3, 115.3, 117.1 (d, ²*J*_{C,F} = 17 Hz), 117.8 (d, ²*J*_{C,F} = 17 Hz), 125.7 (dd, ³*J*_{C,F} = 6 Hz, ⁴*J*_{C,F} = 3 Hz), 132.0, 137.0 (dd, ³*J*_{C,F} = 6 Hz, ⁴*J*_{C,F} = 4 Hz), 147.2 and 149.6 (dd, ¹*J*_{C,F} = 245 Hz, ²*J*_{C,F} = 13 Hz), 147.8 and 150.2 (dd, ¹*J*_{C,F} = 245 Hz, ²*J*_{C,F} = 13 Hz), 152.9, 154.4. IR [cm⁻¹]: $\tilde{\nu}$ = 1203, 1454, 1510, 1674, 2991, 3149, 3272, 3396. HRMS: [C₁₅H₁₅N₃O₂F₂S + H]⁺ calculated: 340.0926, found: 340.0919.

Ethyl-(2-amino-6-[[4-(trifluoromethyl)benzyl]thio]pyridin-3-yl)carbamate (6d):

Method A: A three-neck round-bottom flask was equipped with **5d** (1.52 mmol, 0.50 g), SnCl₂•2H₂O (5.2 eq, 7.9 mmol, 1.79 g) and 20 mL ethanol. The mixture was put under an argon atmosphere and heated to 70 °C for 23 hours. Afterwards, the temperature was lowered to 40 °C and Et₃N (10.2 eq, 15.4 mmol, 2.14 mL), as well as ethyl chloroformate (1.27 eq, 1.9 mmol, 0.18 mL), were added slowly to the suspension. After 30 min the temperature was increased to 55 °C and kept for one additional hour. After cooling to r.t. the mixture was filtered through a pad of celite and washed with ethanol. The filtrate was evaporated to dryness and distributed between water and dichloromethane. The organic layer was evaporated to dryness and the residue washed with *n*-hexane. The product was dried at 40 °C under reduced pressure. White solid (m.p.: 166–168 °C). Yield: 0.42 g (74.0%).

Method B: **5c** (5 mmol, 1.65 g) was suspended in a mixture of 30 mL water and 50 mL 2-propanol. Iron powder (3 eq, 15 mmol, 0.84 g) and NH₄Cl (4.5 eq, 22.5 mmol, 1.2 g) were added and the suspension refluxed for 4 hours. The resulting dark brown mixture was filtered and the residue washed with 100 mL 2-propanol. Et₃N (5.0 eq, 25 mmol, 3.45 mL) and ethyl chloroformate (1.2 eq, 6 mmol, 0.57 mL) were added. The solution was stirred for 3 hours at 40 °C. Since TLC monitoring showed no complete conversion ethyl chloroformate (1.2 eq, 6 mmol, 0.57 mL) and Et₃N (1 eq, 5 mmol, 0.69 mL) were added. After 7 hours ethyl chloroformate (0.6 eq, 3 mmol, 0.29 mL) and Et₃N (0.6 eq, 3 mmol, 0.41 mL) were added again. The reaction was completed after 2 more hours and the solution evaporated to dryness. The solid was dissolved in 150 mL ethyl acetate and the solution washed with 2×100 mL water as well as 100 mL saturated brine. After drying with Na₂SO₄ the crude product was purified by flash chromatography (mobile phase: *n*-hexane/ethyl acetate 7:3). The resulting brown solid was washed with small quantities of ice-cold diethyl ether and dried at 40 °C under reduced pressure. White solid (m.p.: 170–171 °C). Yield: 0.71 g (38.2%). Compound purity: 100.0%. *R*_f = 0.58 (*n*-hexane/ethyl acetate 7:3). ¹H-NMR [ppm]: δ = 1.22 (t, 3H, *J* = 7.1 Hz), 4.09 (q, 2H, *J* = 7.1 Hz), 4.39 (s, 2H), 5.97 (s, 2H), 6.44 (d, 1H, *J* = 8.0 Hz), 7.41 (d, 1H,

$J = 5.6$ Hz), 7.64 (s, 4H), 8.57 (s, 1H). ^{13}C -NMR [ppm]: $\delta = 14.5, 32.8, 60.4, 109.3, 115.3, 122.9$ and $125.6, 125.1$ (q, 2C, $^4J_{\text{C,F}} = 4$ Hz), 127.4 (q, $^2J_{\text{C,F}} = 32$ Hz), 129.6 (2C), 131.8, 144.1, 149.6, 152.7, 154.4. IR [cm^{-1}]: $\tilde{\nu} = 1113, 1460, 1685, 2980, 3174, 3307, 3423$. HRMS: $[\text{C}_{16}\text{H}_{16}\text{N}_3\text{O}_2\text{F}_3\text{S} + \text{H}]^+$ calculated: 372.0988, found: 372.0970.

Ethyl-{2-amino-6-[(4-fluorobenzyl)sulfinyl]pyridin-3-yl}carbamate (7a): **6a** (0.5 mmol, 0.17 g) was dissolved in 10 mL DCM and cooled to 0 °C. *meta*-Chloroperoxybenzoic acid (*m*CPBA, 1.1 eq, 0.55 mmol, 0.17 g of a 55% mixture with water) were dissolved in 10 mL DCM and added dropwise within 30 min. The solution was stirred for 3 hours while the product precipitated as a white solid. The suspension was evaporated to dryness and purified by flash chromatography (mobile phase: DCM/MeOH 95:5). The product was washed with diethyl ether, sat. NaHCO_3 solution and water and dried under reduced pressure at 40 °C. Light pink solid (m.p.: 202–204 °C). Yield: 0.11 g (65.1%). Compound purity: 100.0%. $R_f = 0.58$ (*n*-hexane/ethyl acetate 5:5). ^1H -NMR [ppm]: $\delta = 1.25$ (t, 3H, $J = 7.1$), 3.99 (d, 1H, $J = 13.0$ Hz), 4.14 (q, 2H, $J = 7.0$ Hz), 4.32 (d, 1H, $J = 13.0$ Hz), 6.38 (s, 2H), 6.72 (d, 1H, $J = 7.9$ Hz), 7.05–7.12 (m, 4H), 7.82 (d, 1H, $J = 7.5$ Hz), 8.86 (s, 1H). ^{13}C -NMR [ppm]: $\delta = 14.5, 57.6, 60.7, 108.7, 114.9$ (d, 2C, $^2J_{\text{C,F}} = 21$ Hz), 120.5, 126.6 (d, $^4J_{\text{C,F}} = 3$ Hz), 129.5, 132.2 (d, 2C, $^3J_{\text{C,F}} = 8$ Hz), 152.0, 154.2, 154.6, 160.7 and 163.0 (d, $^1J_{\text{C,F}} = 244$ Hz). IR [cm^{-1}]: $\tilde{\nu} = 1219, 1466, 1532, 1731, 2978, 3213, 3294, 3347$. HRMS: $[\text{C}_{15}\text{H}_{16}\text{N}_3\text{O}_3\text{FS} + \text{H}]^+$ req.: 338.0969, found: 338.0978.

Ethyl-{2-amino-6-(benzylsulfinyl)pyridin-3-yl}carbamate (7b): **6b** (1 mmol, 0.30 g) was dissolved in 20 mL DCM and cooled to 0 °C. *m*CPBA (1.1 eq, 1.1 mmol, 0.34 g of a 55% mixture with water) was dissolved in 30 mL DCM and added drop wise over a period of 2 hours. The resulting solution was stirred for further 3 hours. After evaporation of approximately 50% of the solvent a white precipitate was formed. The precipitation was completed by storage of the suspension at 4 °C for 72 hours. The solid was filtered off and washed with sat. NaHCO_3 solution. The crude product was purified by flash chromatography (mobile phase: *n*-hexane/ethyl acetate 5:5 until the starting material eluted, afterwards the mobile phase was changed to methanol and the product eluted). The solid was dried under reduced pressure at 40 °C. White solid (m.p.: 183–186 °C). Yield: 0.05 g (16.0%). Compound purity: 100.0%. $R_f = 0.21$ (cyclohexane/ethanol/ Et_3N 6:2:2). ^1H -NMR [ppm]: $\delta = 1.26$ (t, 3H, $J = 7.1$ Hz), 3.97 (d, 1H, $J = 12.9$ Hz), 4.14 (q, 2H, $J = 7.1$ Hz), 4.31 (d, 1H, $J = 12.9$ Hz), 6.38 (s, 2H), 6.78 (d, 1H, $J = 7.9$ Hz), 7.08–7.10 (m, 2H), 7.28–7.29 (m, 3H), 7.82 (d, 1H, $J = 7.8$ Hz), 8.87 (s, 1H). ^{13}C -NMR [ppm]: $\delta = 14.5, 59.0, 60.7, 108.6, 120.5, 127.7, 128.2$ (2C), 129.6, 130.2 (2C), 130.7, 152.0, 154.2, 154.9. IR [cm^{-1}]: $\tilde{\nu} = 1220, 1463, 1533, 1728, 2975, 3212, 3341$. HRMS: $[\text{C}_{15}\text{H}_{17}\text{N}_3\text{O}_3\text{S} + \text{H}]^+$ calculated: 320.1063, found: 320.1052.

Ethyl-{2-amino-6-[(3,4-difluorobenzyl)sulfinyl]pyridin-3-yl}carbamate (7c): **6d** (1 mmol, 0.34 g) was dissolved in 20 mL DCM and cooled to 0 °C. *m*CPBA (1.1 eq, 1.1 mmol, 0.35 g of a 55% mixture with water) was dissolved in 20 mL DCM and added drop wise within 30 min. The solution was stirred for further 20 min and afterwards diluted with 60 mL DCM. The organic phase was washed with 3×100 mL sat. NaHCO_3 solution as well as 100 mL saturated brine and dried with Na_2SO_4 . After evaporation the crude product was recrystallized from ethyl acetate and dried under reduced pressure at 40 °C. Beige solid (m.p.: 196–198 °C). Yield: 0.13 g (37.3%). Compound purity: 100.0%. $R_f = 0.27$ (*n*-hexane/ethyl acetate 4:6). ^1H -NMR [ppm]: $\delta = 1.25$ (t, 3H, $J = 7.1$ Hz), 4.00 (d, 1H, $J = 13.0$ Hz), 4.33 (d, 1H, $J = 13.0$ Hz), 6.37 (s, 2H), 6.70 (d, 1H, $J = 7.9$ Hz), 6.82–6.85 (m, 1H), 7.03–7.08 (m, 1H), 7.29–7.36 (m, 1H), 7.82 (d, 1H, $J = 7.6$ Hz), 8.87 (s, 1H). ^{13}C -NMR [ppm]: $\delta = 14.4, 57.0, 60.7, 108.8, 117.0$ (d,

$^2J_{C,F} = 17$ Hz), 119.1 (d, $^2J_{C,F} = 17$ Hz), 120.6, 127.2 (dd, $^3J_{C,F} = 6$ Hz, $^4J_{C,F} = 3$ Hz), 128.0 (dd, $^3J_{C,F} = 6$ Hz, $^4J_{C,F} = 4$ Hz), 129.4, 147.6 and 150.0 (dd, $^1J_{C,F} = 245$ Hz, $^2J_{C,F} = 12$ Hz), 147.9 and 150.3 (dd, $^1J_{C,F} = 245$ Hz, $^2J_{C,F} = 12$ Hz), 151.9, 154.1, 154.2. IR [cm^{-1}]: $\tilde{\nu} = 1221, 1463, 1672, 1724, 2979, 3215, 3343$. HRMS: $[\text{C}_{15}\text{H}_{15}\text{N}_3\text{O}_3\text{F}_2\text{S} + \text{H}]^+$ calculated: 356.0875, found: 356.0881.

Ethyl-(2-amino-6-[[4-(trifluoromethyl)benzyl]sulfinyl]pyridin-3-yl)carbamate (7d): **6c** (0.40 mmol, 0.15 g) was dissolved in 12 mL DCM and cooled to 0 °C. *m*CPBA (1.0 eq, 0.40 mmol, 0.13 g of a 55% mixture with water) was added to the solution. Within 30 minutes the product started to precipitate. Since the reaction was not completed after one hour, *m*CPBA (0.5 eq, 0.20 mmol, 0.07 g of a 55% mixture with water) was added. The suspension was stirred for one more hour, filtrated and the product dried under reduced pressure at 40 °C. White solid (m.p.: 187–189 °C). Yield: 0.05 g (35.0%). Compound purity: 100.0%. $R_f = 0.19$ (*n*-hexane/ ethyl acetate 4:6). $^1\text{H-NMR}$ [ppm]: $\delta = 1.26$ (t, 3H, $J = 7.1$ Hz), 4.11 (d, 1H, $J = 12.8$ Hz), 4.15 (q, 2H, $J = 7.0$ Hz), 4.45 (d, 1H, $J = 12.8$ Hz), 6.37 (s, 2H), 6.72 (d, 1H, $J = 7.9$ Hz), 7.26 (d, 2H, $J = 8.0$ Hz), 7.63 (d, 2H, $J = 8.1$ Hz), 7.82 (d, 1H, $J = 7.9$ Hz), 8.85 (s, 1H). $^{13}\text{C-NMR}$ [ppm]: $\delta = 14.4, 57.8, 60.7, 108.7, 120.6, 122.9$ and 125.6 (q, $^1J_{C,F} = 270$ Hz), 124.8 (q, $^4J_{C,F} = 4$ Hz), 128.2 (q, 2C, $^2J_{C,F} = 31.8$), 129.5, 131.1 (2C), 135.3, 152.0, 154.1, 154.2. IR [cm^{-1}]: $\tilde{\nu} = 1122, 1466, 1659, 1730, 2979, 3216, 3305, 3343$. HRMS: $[\text{C}_{15}\text{H}_{16}\text{N}_3\text{O}_3\text{FS} - \text{H}]^-$ calculated: 386.0792, found: 386.0781.

Ethyl-(2-amino-6-[(4-fluorobenzyl)sulfonyl]pyridin-3-yl)carbamate (8a): **6a** (1 mmol, 0.32 g) was dissolved in 20 mL DCM and cooled to 0 °C. *m*CPBA (2 eq, 2 mmol, 0.63 g of a 55% mixture with water) was dissolved in 20 mL DCM and added drop wise over a period of 1 hour. The solution was stirred for further 45 min at 0 °C. The resulting suspension was filtered and the residue washed with DCM. The filtrate was washed with 3×100 mL sat. NaHCO_3 solution and 100 mL brine. The organic phase was evaporated and purified by flash chromatography (puriFlash® 450 system from Interchim, 25 g silica column, mobile phase: *n*-hexane/ethyl acetate, gradient: 1 CV 10% ethyl acetate, 10 CV 10–80% ethyl acetate, 5 CV 80% ethyl acetate). The solid was washed with small quantities of diethyl ether and dried under reduced pressure at 40 °C. Pink solid (m.p.: 168–171 °C). Yield: 0.09 g (25.5%). Compound purity: 100.0%. $R_f = 0.41$ (*n*-hexane/ethyl acetate). $^1\text{H-NMR}$ [ppm]: $\delta = 1.26$ (t, 3H, $J = 7.0$ Hz), 4.16 (q, 2H, $J = 7.0$ Hz), 4.61 (s, 2H), 6.64 (s, 2H), 7.00 (d, 1H, $J = 8.0$ Hz), 7.13–7.17 (m, 2H), 7.22–7.25 (m, 2H), 7.95 (d, 1H, $J = 7.8$ Hz), 9.05 (s, 1H). $^{13}\text{C-NMR}$ [ppm]: $\delta = 14.4, 56.1, 60.9, 111.5, 115.2$ (d, 2C, $^2J_{C,F} = 22$ Hz), 123.3, 124.8 (d, $^4J_{C,F} = 3$ Hz), 127.4, 133.0 (d, 2C, $^3J_{C,F} = 8$ Hz), 147.3, 151.3, 153.9, 160.9 and 163.3 (d, $^1J_{C,F} = 245$ Hz). IR [cm^{-1}]: $\tilde{\nu} = 1232, 1462, 1511, 1634, 1714, 2979, 3305, 3445$. HRMS: $[\text{C}_{15}\text{H}_{16}\text{N}_3\text{O}_4\text{FS} + \text{H}]^+$ req.: 354.0918, found: 354.0930.

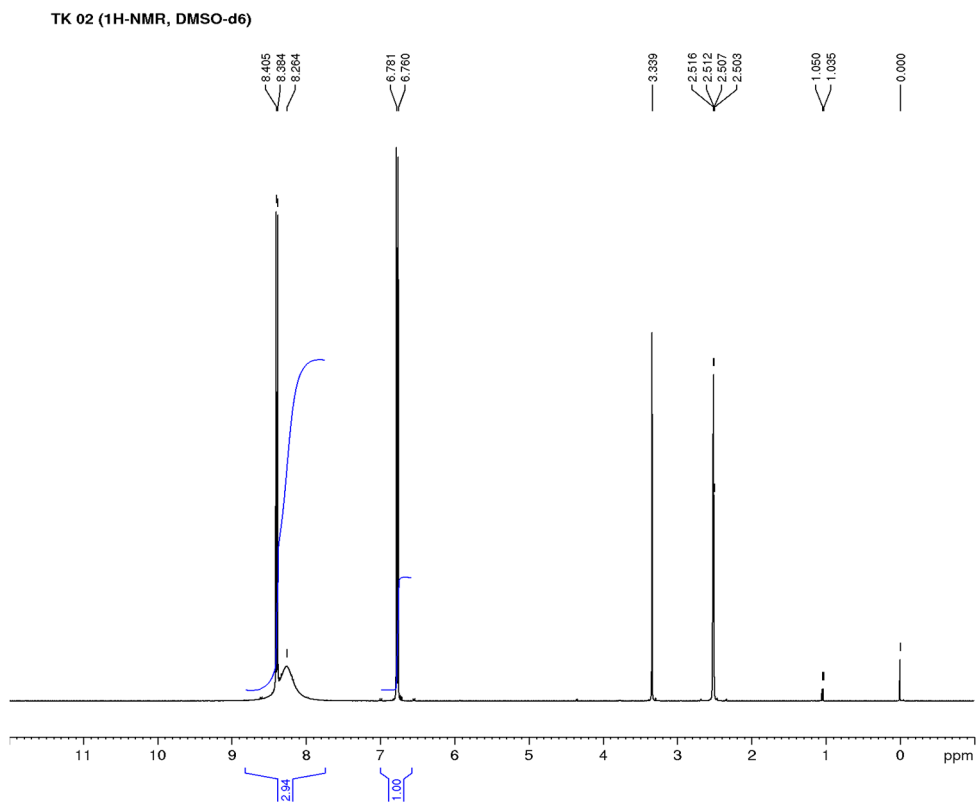
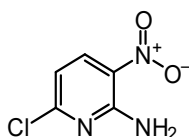
Ethyl-(2-amino-6-(benzylsulfonyl)pyridin-3-yl)carbamate (8b): **6b** (1 mmol, 0.30 g) was dissolved in 20 mL DCM and cooled to 0 °C. *m*CPBA (2.2 eq, 2.2 mmol, 0.69 g of a 55% mixture with water) was dissolved in 40 mL DCM and added drop wise within 1 hour. Since TLC monitoring showed no complete conversion after 1 additional hour *m*CPBA (0.5 eq, 0.5 mmol, 0.35 g of a 55% mixture with water) was dissolved in 5 mL DCM and added drop wise. After 30 min the solution was extracted with 2×100 mL sat. NaHCO_3 solution and 100 mL saturated brine. After drying with NaSO_4 the product was purified via flash chromatography (mobile phase: *n*-hexane/ethyl acetate 5:5), washed with diethyl ether and dried under reduced pressure at 40 °C. Light pink solid (m.p.: 164–166 °C).

Yield: 0.19 g (55.2%). Compound purity: 100.0%. R_f = 0.46 (*n*-hexane/ethyl acetate 4:6). $^1\text{H-NMR}$ [ppm]: δ = 1.26 (t, 3H, J = 7.1 Hz), 4.16 (q, 2H, J = 7.1 Hz), 4.60 (s, 2H), 6.64 (s, 2H), 7.00 (d, 1H, J = 8.0 Hz), 7.18–7.20 (m, 2H), 7.30–7.31 (m, 2H), 7.94 (d, 1H, J = 8.0 Hz), 9.05 (s, 1H). $^{13}\text{C-NMR}$ [ppm]: δ = 14.4, 57.0, 60.9, 111.4, 123.3, 127.4, 128.2, 128.3 (2C), 128.4, 131.0 (2C), 147.5, 151.4, 153.9. IR [cm^{-1}]: $\tilde{\nu}$ = 1229, 1464, 1518, 1633, 1713, 2972, 3299, 3449. HRMS: $[\text{C}_{15}\text{H}_{17}\text{N}_3\text{O}_4\text{S} + \text{H}]^+$ calculated: 336.1013, found: 336.1029.

Ethyl-(2-amino-6-[(3,4-difluorobenzyl)sulfonyl]pyridin-3-yl)carbamate (8c): **6d** (1 mmol, 0.34 g) was dissolved in 20 mL DCM and cooled to 0 °C. *m*CPBA (2 eq, 2 mmol, 0.63 g of a 55% mixture with water) was dissolved in 20 mL DCM and added drop wise. Since TLC still showed starting material after 3 hours *m*CPBA (0.5 eq, 0.5 mmol, 0.32 g of a 55% mixture with water) was dissolved in 5 mL DCM and added drop wise again. After 4 additional hours the suspension was extracted with 2×50 mL sat. NaHCO_3 solution, 50 mL water and 50 mL saturated brine. The product was purified by flash chromatography (*n*-hexane/ethyl acetate 5:5) washed with diethyl ether and dried under reduced pressure at 40 °C. White solid (m.p.: 186–188 °C). Yield: 0.19 g (51.2%). Compound purity: 100.0%. R_f = 0.57 (*n*-hexane/ethyl acetate 4:6). $^1\text{H-NMR}$ [ppm]: δ = 1.26 (t, 3H, J = 7.1 Hz), 4.16 (q, 2H, J = 7.1 Hz), 4.65 (s, 2H), 6.64 (s, 2H), 7.02 (d, 1H, J = 8.0 Hz), 7.01–7.05 (m, 1H), 7.24–7.29 (m, 1H), 7.35–7.42 (m, 1H), 7.96 (d, 1H, J = 8.0 Hz), 9.06 (s, 1H). $^{13}\text{C-NMR}$ [ppm]: δ = 14.4, 55.9, 60.9, 111.5, 117.4 (d, $^2J_{\text{C,F}}$ = 17 Hz), 119.9 (d, $^2J_{\text{C,F}}$ = 17 Hz), 123.4, 126.3 (dd, $^3J_{\text{C,F}}$ = 6 Hz, $^4J_{\text{C,F}}$ = 4 Hz), 127.4, 128.1 (dd, $^3J_{\text{C,F}}$ = 7 Hz, $^4J_{\text{C,F}}$ = 3 Hz), 147.1, 147.7 and 150.2 (dd, $^1J_{\text{C,F}}$ = 249 Hz, $^2J_{\text{C,F}}$ = 16 Hz), 148.2 and 150.7 (dd, $^1J_{\text{C,F}}$ = 255 Hz, $^2J_{\text{C,F}}$ = 20 Hz), 151.4, 153.9. IR [cm^{-1}]: $\tilde{\nu}$ = 1231, 1464, 1519, 1713, 2975, 3299, 3449. HRMS: $[\text{C}_{15}\text{H}_{15}\text{F}_2\text{N}_3\text{O}_4\text{S} + \text{H}]^+$ calculated: 372.0824, found: 372.0832.

Ethyl-(2-amino-6-[[4-(trifluoromethyl)benzyl)sulfonyl]pyridin-3-yl)carbamate (8d): **6c** (0.75 mmol, 0.28 g) was dissolved in 15 mL DCM and cooled to 0 °C. *m*CPBA (2 eq, 1.5 mmol, 0.47 g of a 55% mixture with water) was dissolved in 15 mL DCM and added drop wise within 1 hour. After additional 100 min the reaction is completed and the solution extracted with 3×50 mL sat. NaHCO_3 solution as well as 50 mL saturated brine. After drying with Na_2SO_4 the product was purified by flash chromatography (puriFlash® 450 system from Interchim, 25 g silica column, mobile phase: *n*-hexane/ethyl acetate, gradient: 1 CV 10% ethyl acetate, 10 CV 10–70% ethyl acetate, 5 CV 70% ethyl acetate). The product was washed with small quantities diethyl ether. Light orange solid (m.p.: 184–186 °C). Yield: 0.13 g (41.3%). Compound purity: 100.0%. R_f = 0.49 (*n*-hexane/ethyl acetate 5:5). $^1\text{H-NMR}$ [ppm]: δ = 1.26 (t, 3H, J = 7.1 Hz), 4.17 (q, 2H, J = 7.1 Hz), 4.76 (s, 2H), 6.66 (s, 2H), 7.03 (d, 1H, J = 8.0 Hz), 7.44 (d, 2H, J = 7.9 Hz), 7.70 (d, 2H, J = 8.0 Hz), 7.97 (d, 1H, J = 8.0 Hz), 9.07 (s, 1H). $^{13}\text{C-NMR}$ [ppm]: δ = 14.4, 56.5, 61.0, 111.4, 120.1 + 122.7 + 125.4 + 128.2 (q, $^1J_{\text{C,F}}$ = 272 Hz), 123.5, 125.1 (q, 2C, $^4J_{\text{C,F}}$ = 4 Hz), 127.3, 128.7 (q, $^2J_{\text{C,F}}$ = 32 Hz), 131.8 (2C), 133.4, 147.2, 151.4, 153.9. IR [cm^{-1}]: $\tilde{\nu}$ = 1234, 1470, 1529, 1649, 1700, 2939, 3345, 3430. HRMS: $[\text{C}_{16}\text{H}_{16}\text{F}_3\text{N}_3\text{O}_4\text{S} - \text{H}]^-$ calculated: 402.0741, found: 402.0750.

Spectra of 6-chloro-3-nitropyridin-2-amine

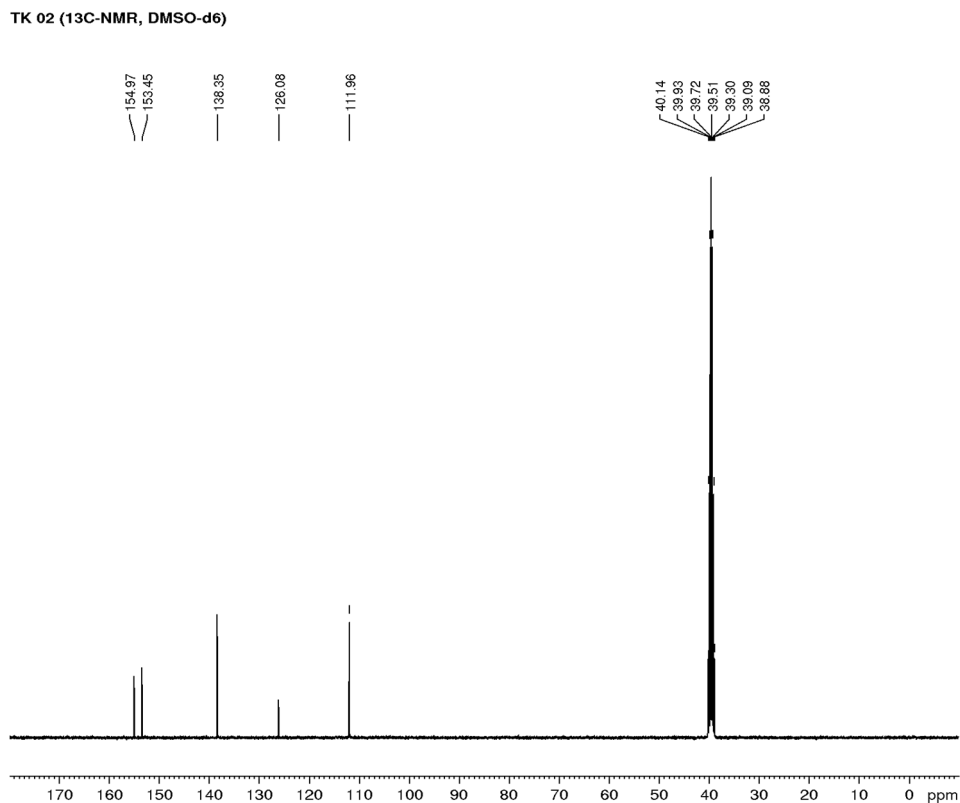


Current Data Parameters
NAME T. Kasch
EXPNO 5
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150512
Time 10.38
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 8223.665 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 101
DW 60.800 usec
DE 10.00 usec
TE 296.2 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 9.75 usec
PL1 -4.50 dB
PL1W 26.90193939 W
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 32768
SF 400.1299984 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



Current Data Parameters
NAME T. Kasch
EXPNO 6
PROCNO 1

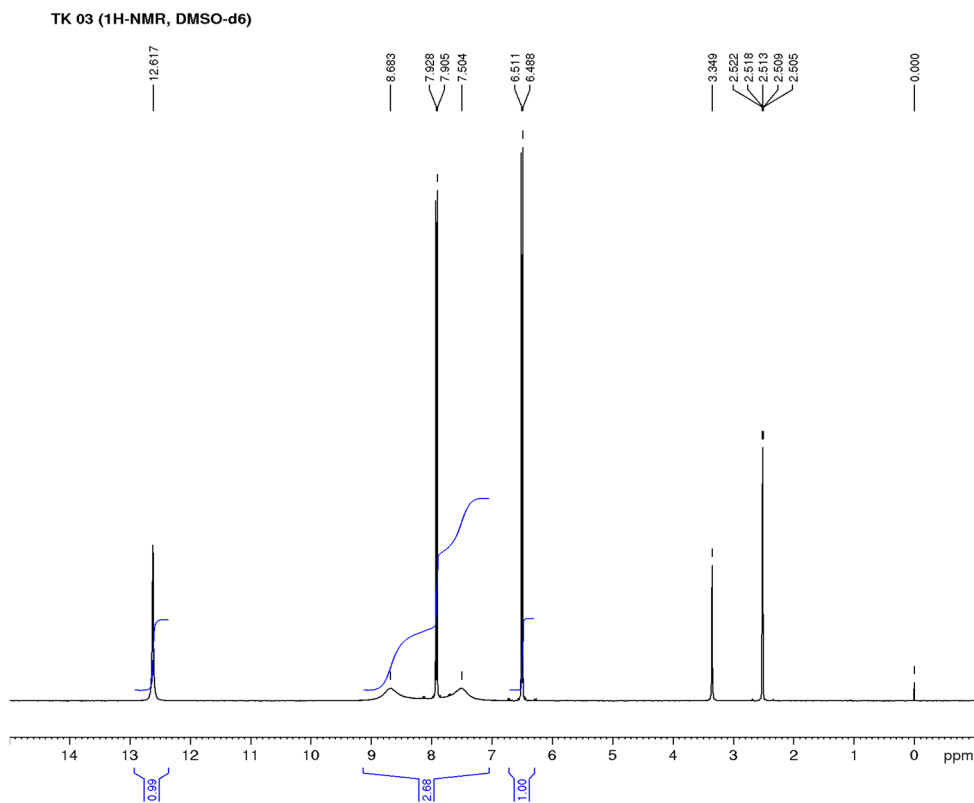
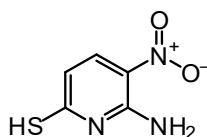
F2 - Acquisition Parameters
Date_ 20150512
Time 13.54
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 1024
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 2050
DW 20.800 usec
DE 10.00 usec
TE 298.5 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.80 usec
PL1 -2.00 dB
PL1W 53.34635925 W
SFO1 100.6228298 MHz

===== CHANNEL f2 =====
CPDPRG[2] waltz16
NUC2 1H
PCPD2 60.00 usec
PL2 -4.50 dB
PL12 11.28 dB
PL13 15.00 dB
PL2W 26.90193939 W
PL12W 0.71085924 W
PL13W 0.30184472 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6128179 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Spectra of 5

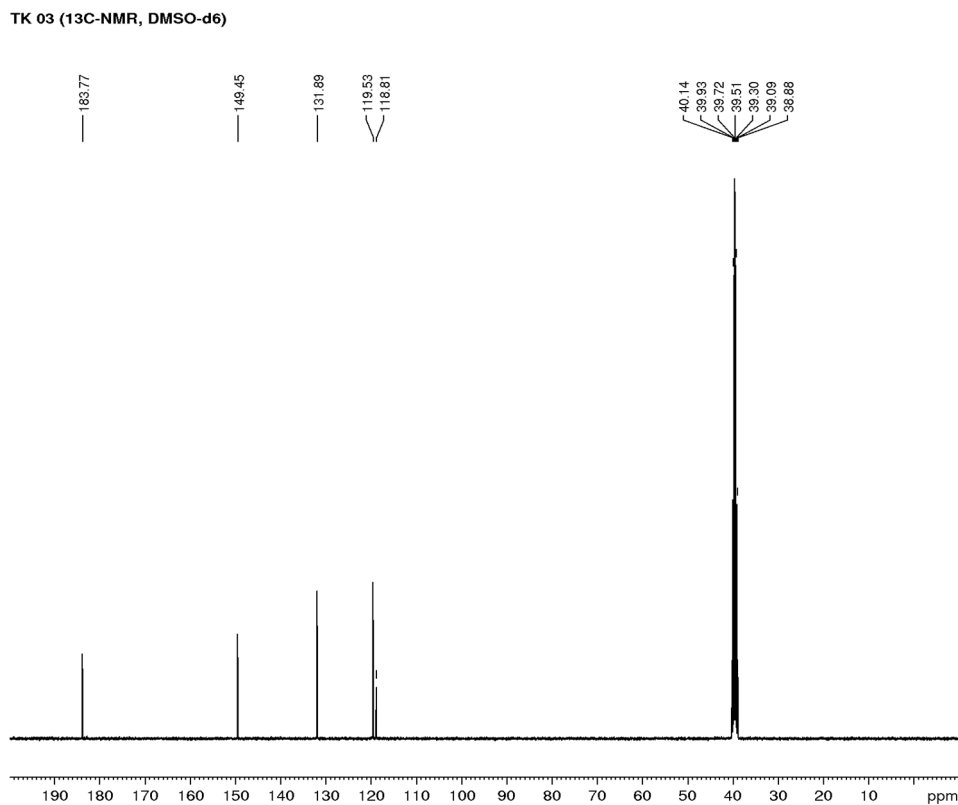


Current Data Parameters
NAME T. Kasch
EXPNO 7
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150518
Time 10.36
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 8223.665 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 90.5
DW 60.800 usec
DE 10.00 usec
TE 296.3 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 9.75 usec
PL1 -4.50 dB
PL1W 26.90193939 W
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 32768
SF 400.1299981 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



Current Data Parameters
NAME T. Kasch
EXPNO 8
PROCNO 1

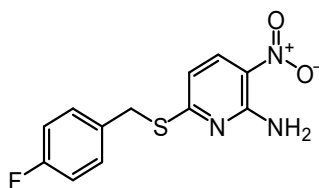
F2 - Acquisition Parameters
Date_ 20150518
Time 14.01
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 1024
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 2050
DW 20.800 usec
DE 10.00 usec
TE 298.1 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.80 usec
PL1 -2.00 dB
PL1W 53.34635925 W
SFO1 100.6228298 MHz

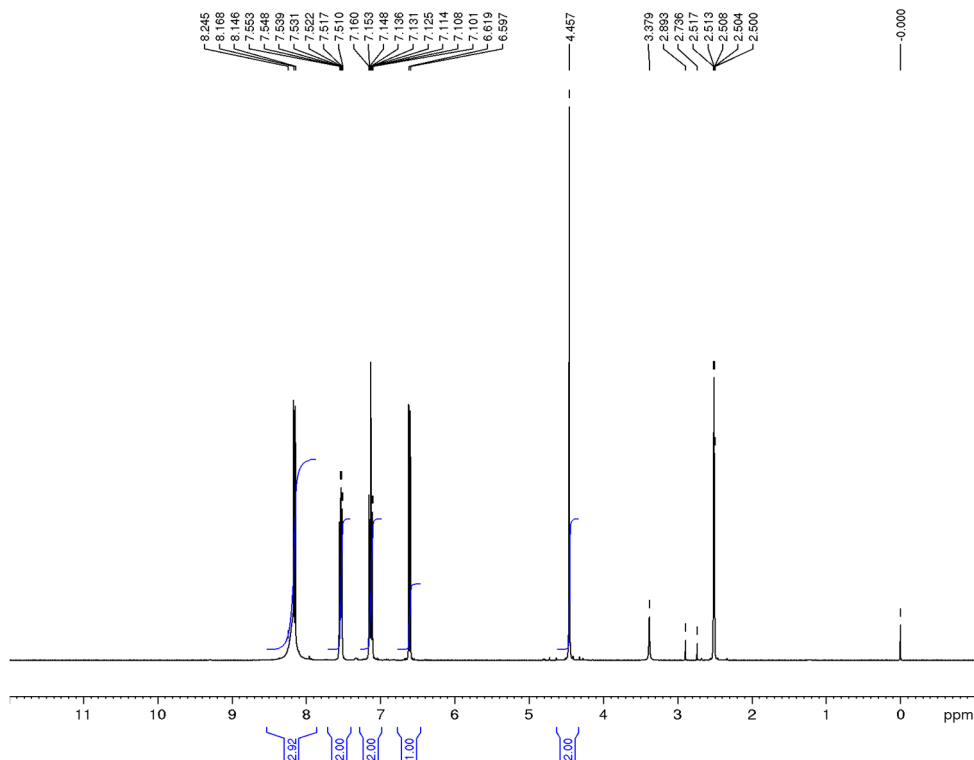
===== CHANNEL f2 =====
CPDPRG12 waltz16
NUC2 1H
PCPD2 60.00 usec
PL2 -4.50 dB
PL12 11.28 dB
PL13 15.00 dB
PL12W 26.90193939 W
PL12W 0.71085924 W
PL13W 0.30184472 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6128167 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Spectra of 5a



TK 04 (1H-NMR, DMSO-d6)



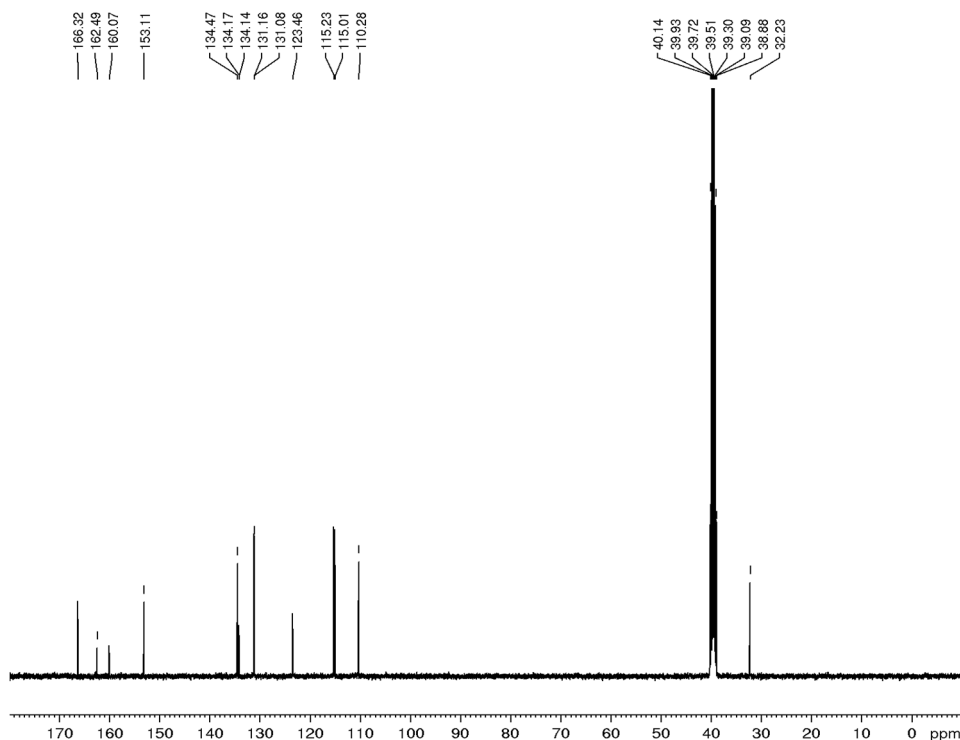
Current Data Parameters
NAME T. Kasch
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150519
Time 13.39
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 8223.665 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 101
DW 60.800 usec
DE 10.00 usec
TE 296.1 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 9.75 usec
PL1 -4.50 dB
PL1W 26.90193939 W
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 32768
SF 400.1300001 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

TK 04 (13C-NMR, DMSO-d6)



Current Data Parameters
NAME T. Kasch
EXPNO 15
PROCNO 1

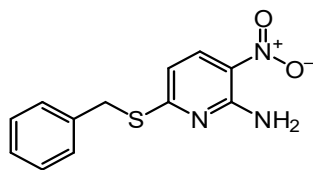
F2 - Acquisition Parameters
Date_ 20150527
Time 12.18
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 1024
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 2050
DW 20.800 usec
DE 10.00 usec
TE 297.9 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.80 usec
PL1 -2.00 dB
PL1W 53.34635925 W
SFO1 100.6228298 MHz

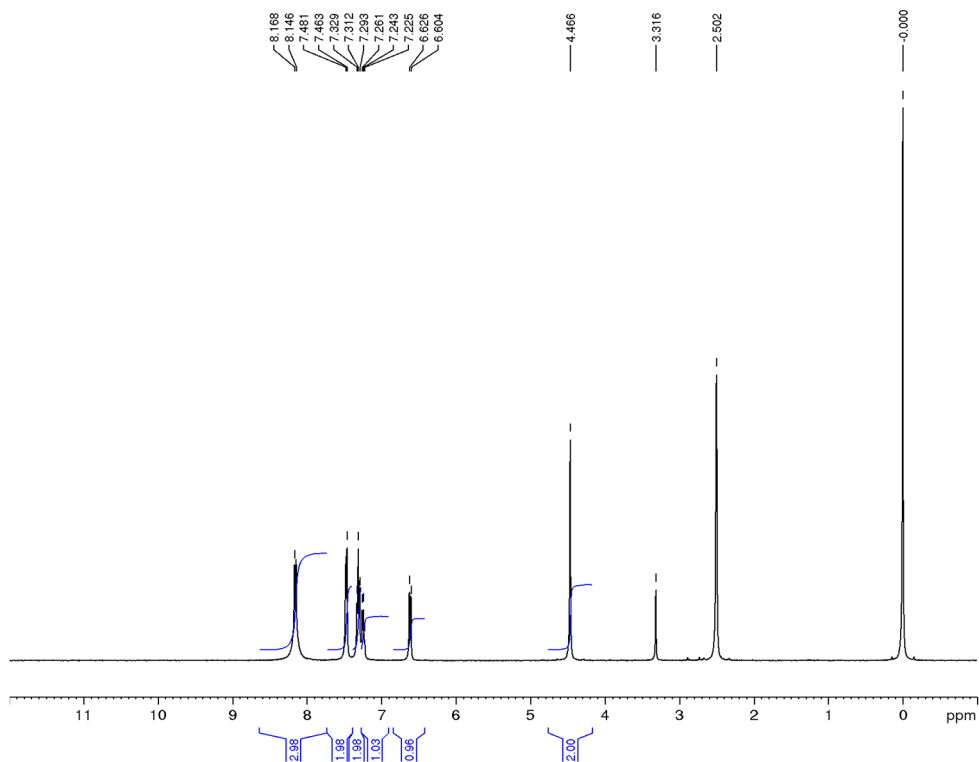
===== CHANNEL f2 =====
CPDPRG[2] waltz16
NUC2 1H
PCPD2 60.00 usec
PL2 -4.50 dB
PL12 11.28 dB
PL13 15.00 dB
PL2W 26.90193939 W
PL12W 0.71085924 W
PL13W 0.30184472 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6128178 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Spectra of 5b



AS-19 (1H-NMR, DMSO-d6)



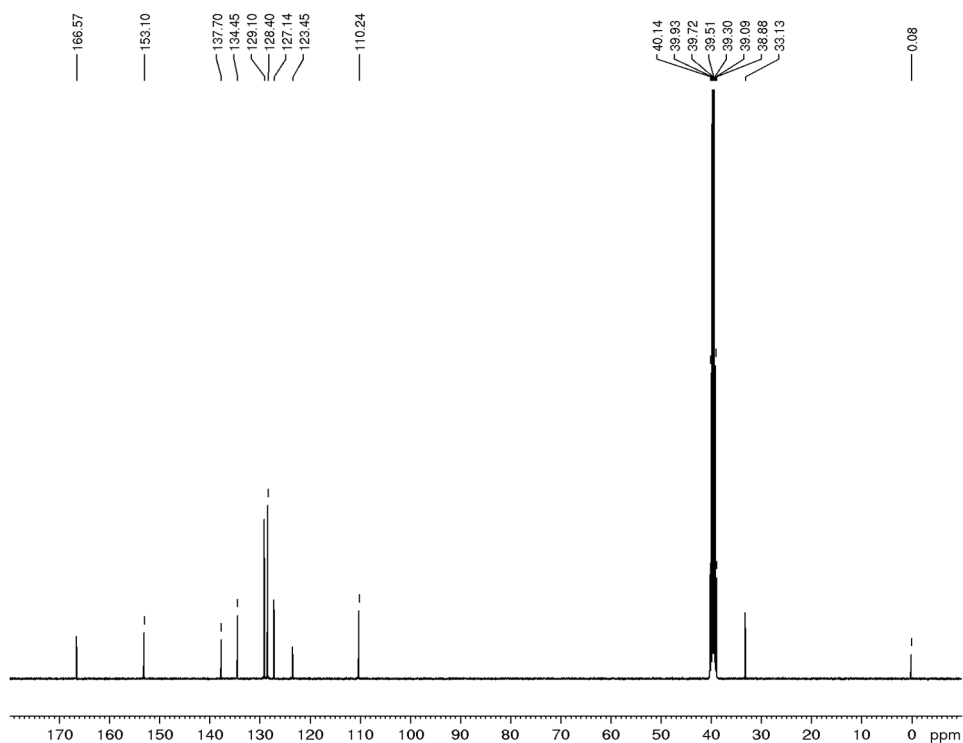
Current Data Parameters
NAME A. Surur
EXPNO 111
PROCNO 1

F2 - Acquisition Parameters
Date_ 20170111
Time 14.24
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 114
DW 60.800 usec
DE 10.00 usec
TE 297.2 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 9.75 usec
PL1 -5.00 dB
PL1W 30.18447304 W
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 32768
SF 400.1300017 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

AS-19 (13C-NMR, DMSO-d6)



Current Data Parameters
NAME A. Surur
EXPNO 113
PROCNO 1

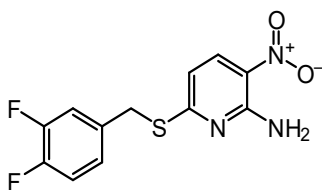
F2 - Acquisition Parameters
Date_ 20170113
Time 6.52
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 4000
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 2050
DW 20.800 usec
DE 10.00 usec
TE 298.2 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
PL1 -2.70 dB
PL1W 62.67650986 W
SFO1 100.6228298 MHz

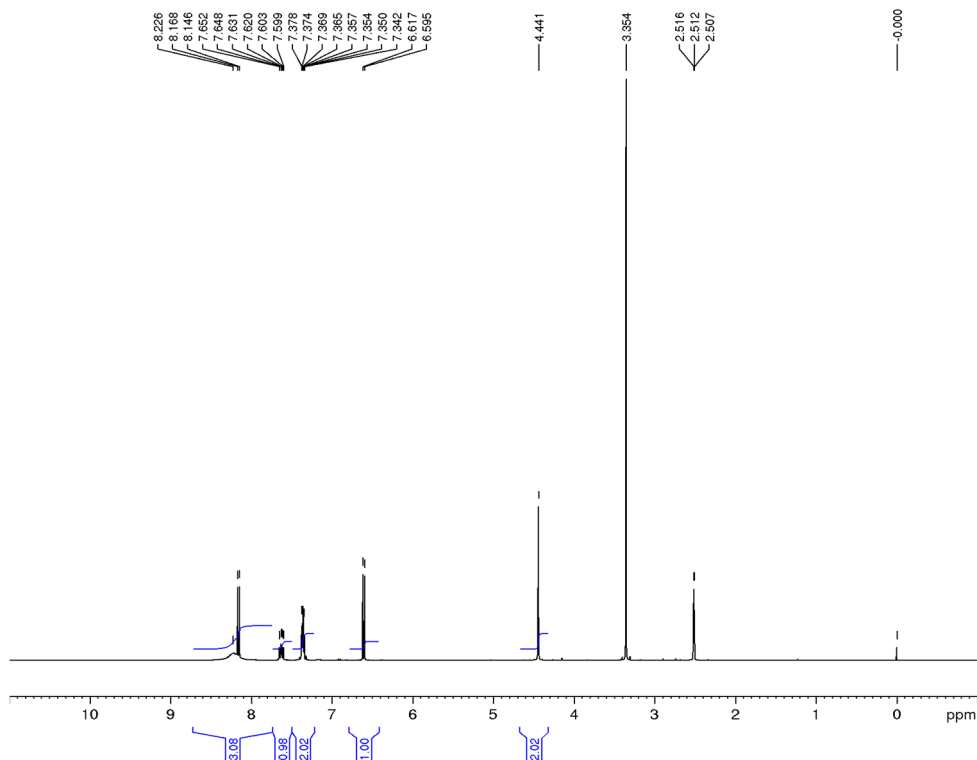
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 60.00 usec
PL2 -5.00 dB
PL12 10.78 dB
PL13 15.00 dB
PL2W 30.18447304 W
PL12W 0.79759723 W
PL13W 0.30184472 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6128183 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Spectra of 5c



TK23.1 (1H-NMR, DMSO-d6)



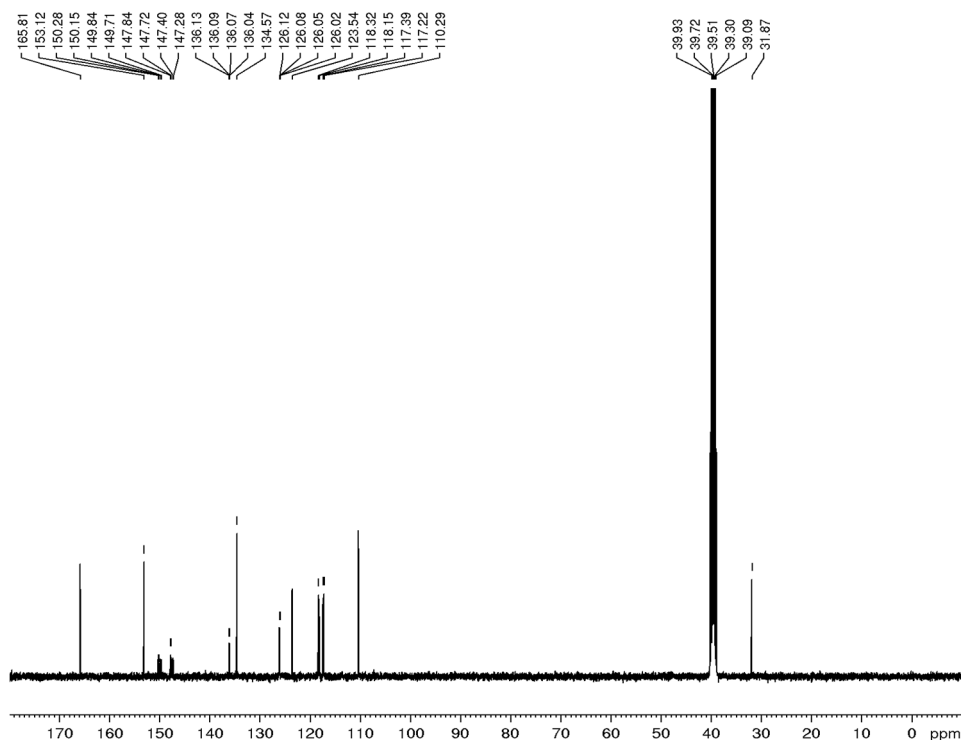
Current Data Parameters
NAME T. Kasch
EXPNO 99
PROCNO 1

F2 - Acquisition Parameters
Date_ 20151009
Time 8.37
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 101
DW 60.800 usec
DE 10.00 usec
TE 283.8 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 9.75 usec
PL1 -4.50 dB
PL1W 26.90193939 W
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 32768
SF 400.1299984 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

TK23.1 (13CNMR, DMSO-d6)



Current Data Parameters
NAME T. Kasch
EXPNO 100
PROCNO 1

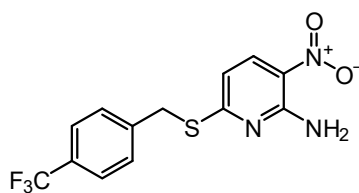
F2 - Acquisition Parameters
Date_ 20151009
Time 9.39
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 1024
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 2050
DW 20.800 usec
DE 10.00 usec
TE 285.3 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.80 usec
PL1 -2.00 dB
PL1W 53.34635925 W
SFO1 100.6228298 MHz

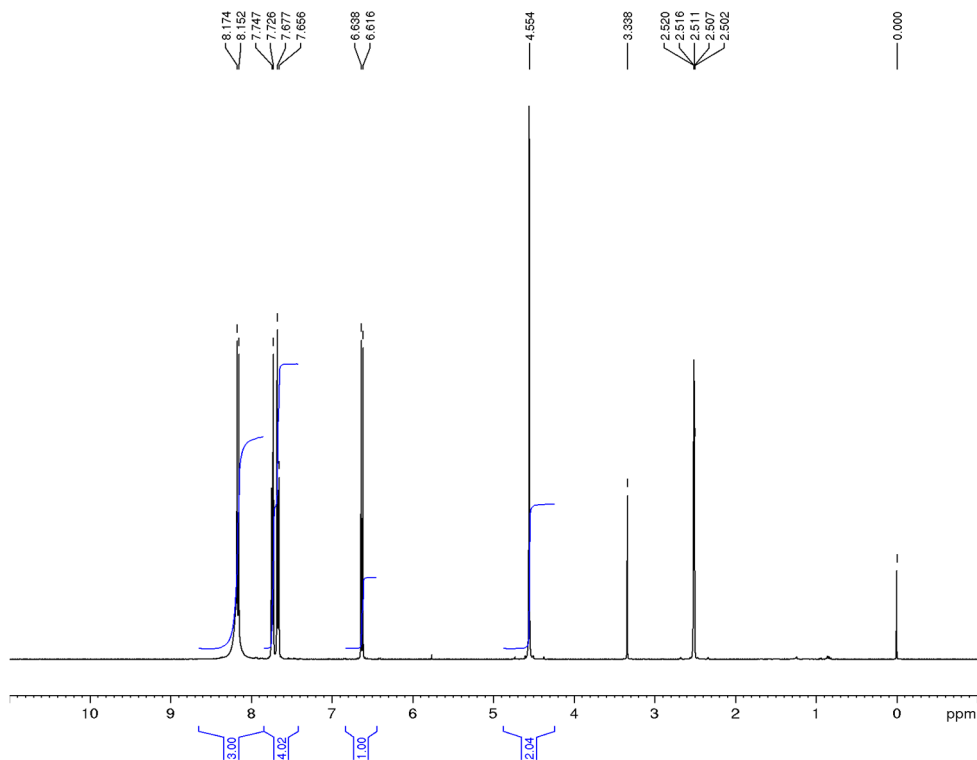
===== CHANNEL f2 =====
CPDPRG[2] waltz16
NUC2 1H
PCPD2 60.00 usec
PL2 -4.50 dB
PL12 11.28 dB
PL13 15.00 dB
PL2W 26.90193939 W
PL12W 0.71085924 W
PL13W 0.30184472 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6128165 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Spectra of 5d



ELB 010 UK (1H-NMR, DMSO-d6)



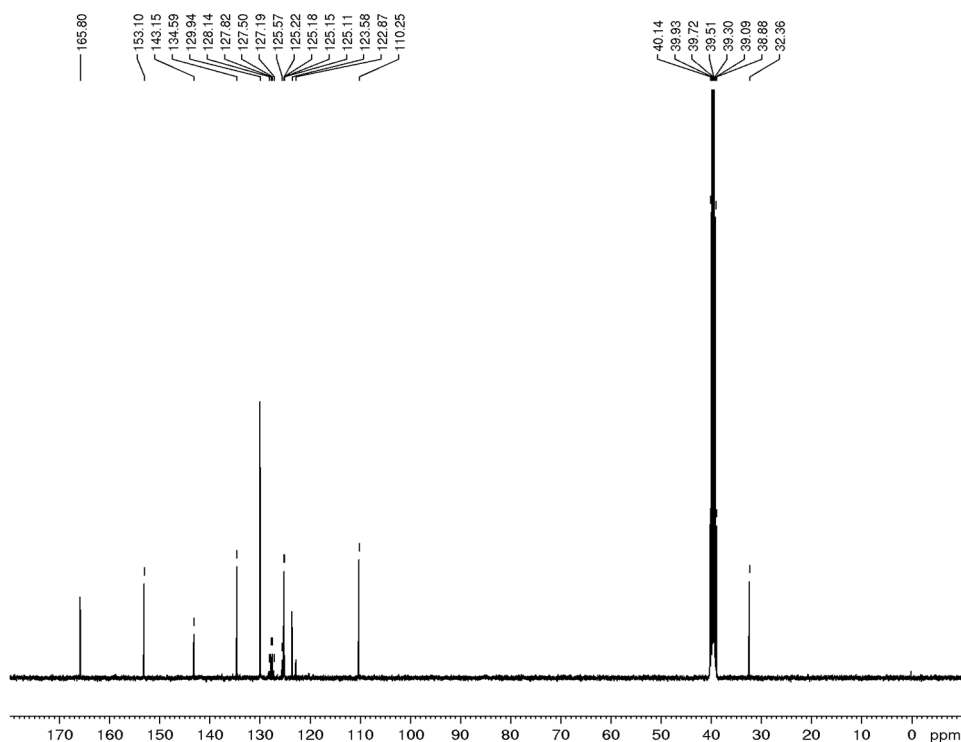
Current Data Parameters
NAME C. Bock
EXPNO 32
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150323
Time 13.39
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 101
DW 60.800 usec
DE 10.00 usec
TE 297.4 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 9.75 usec
PL1 -4.50 dB
PL1W 26.90193939 W
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 32768
SF 400.1299984 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

ELB 010 UK (13C-NMR, DMSO-d6)



Current Data Parameters
NAME C. Bock
EXPNO 38
PROCNO 1

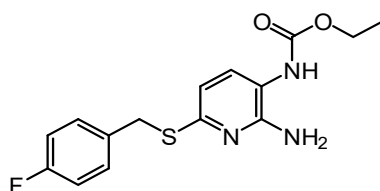
F2 - Acquisition Parameters
Date_ 20150324
Time 10.56
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 1024
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 2050
DW 20.800 usec
DE 10.00 usec
TE 297.7 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 9.80 usec
PL1 -2.00 dB
PL1W 53.34635925 W
SFO1 100.6228298 MHz

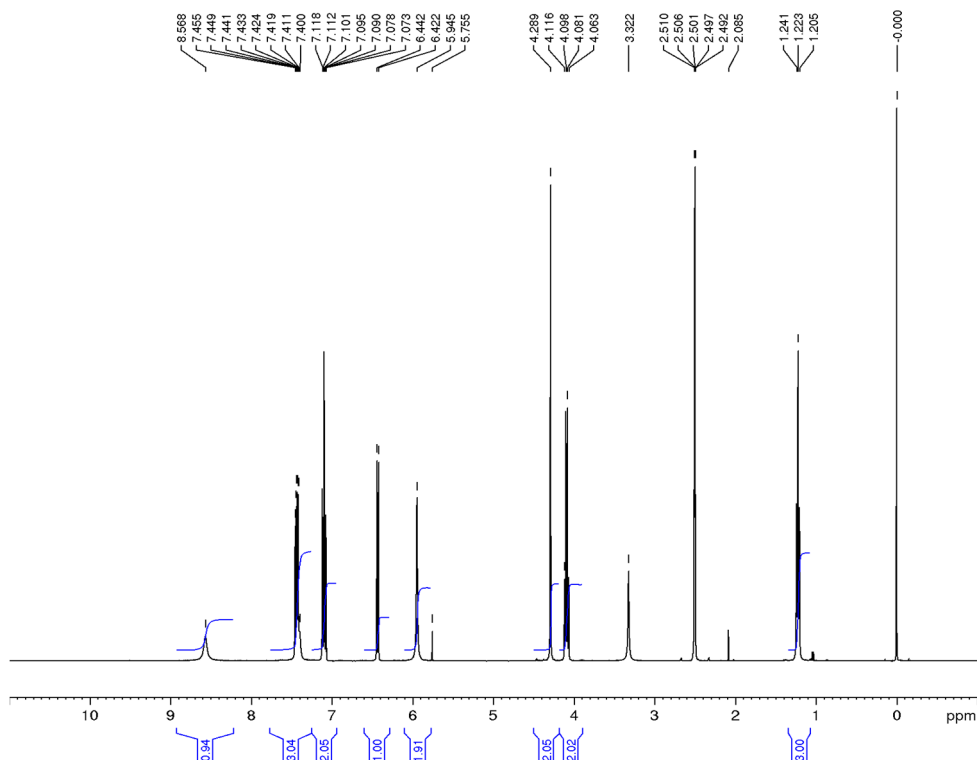
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 60.00 usec
PL2 -4.50 dB
PL12 11.28 dB
PL13 15.00 dB
PL2W 26.90193939 W
PL12W 0.71085924 W
PL13W 0.30184472 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6128175 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Spectra of 6a



ELB079 (1H-NMR, DMSO-d6)



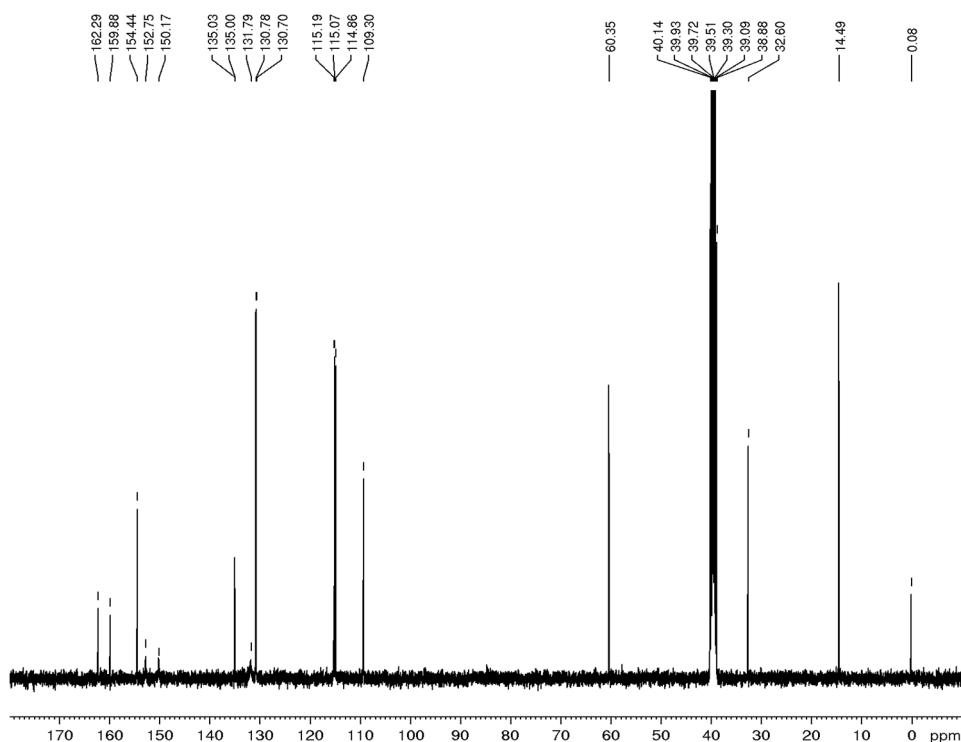
Current Data Parameters
NAME C. Bock
EXPNO 348
PROCNO 1

F2 - Acquisition Parameters
Date_ 20171103
Time 10.42
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 90.5
DW 60.800 usec
DE 10.00 usec
TE 298.4 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 9.75 usec
PL1 -5.00 dB
PL1W 30.18447304 W
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 32768
SF 400.1300027 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

ELB079 (13C-NMR, DMSO-d6)



Current Data Parameters
NAME C. Bock
EXPNO 423
PROCNO 1

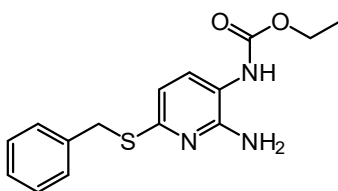
F2 - Acquisition Parameters
Date_ 20180123
Time 14.11
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 1024
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 2050
DW 20.800 usec
DE 10.00 usec
TE 298.3 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
PL1 -2.70 dB
PL1W 62.67650986 W
SFO1 100.6228298 MHz

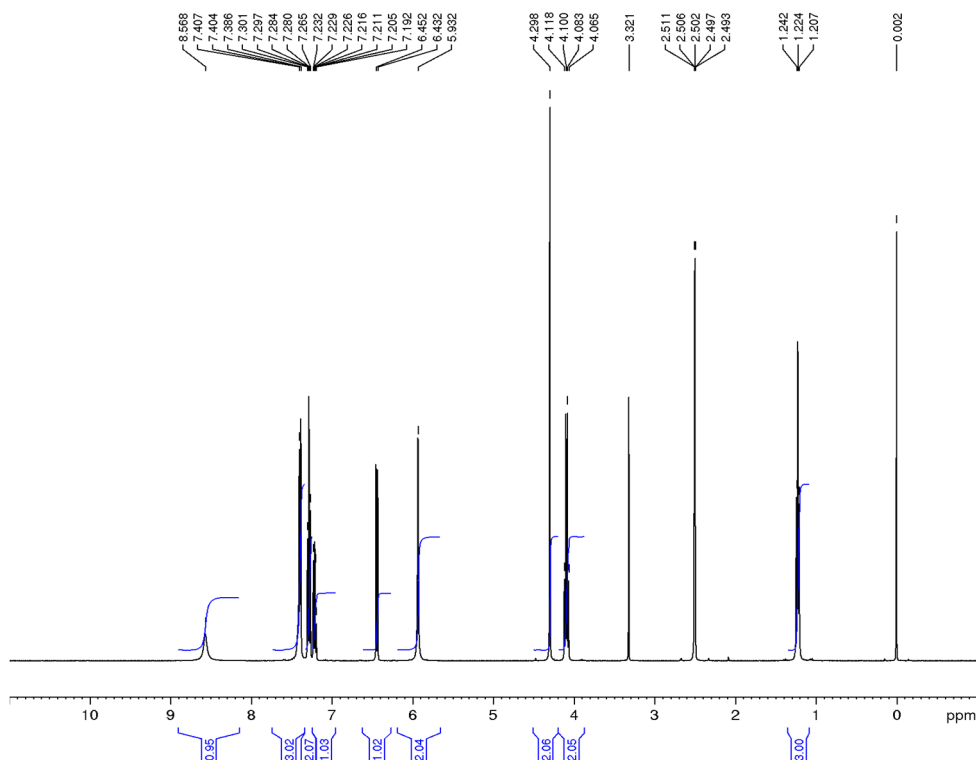
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 60.00 usec
PL2 -5.00 dB
PL12 10.78 dB
PL13 15.00 dB
PL2W 30.18447304 W
PL12W 0.79759723 W
PL13W 0.30184472 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6128182 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

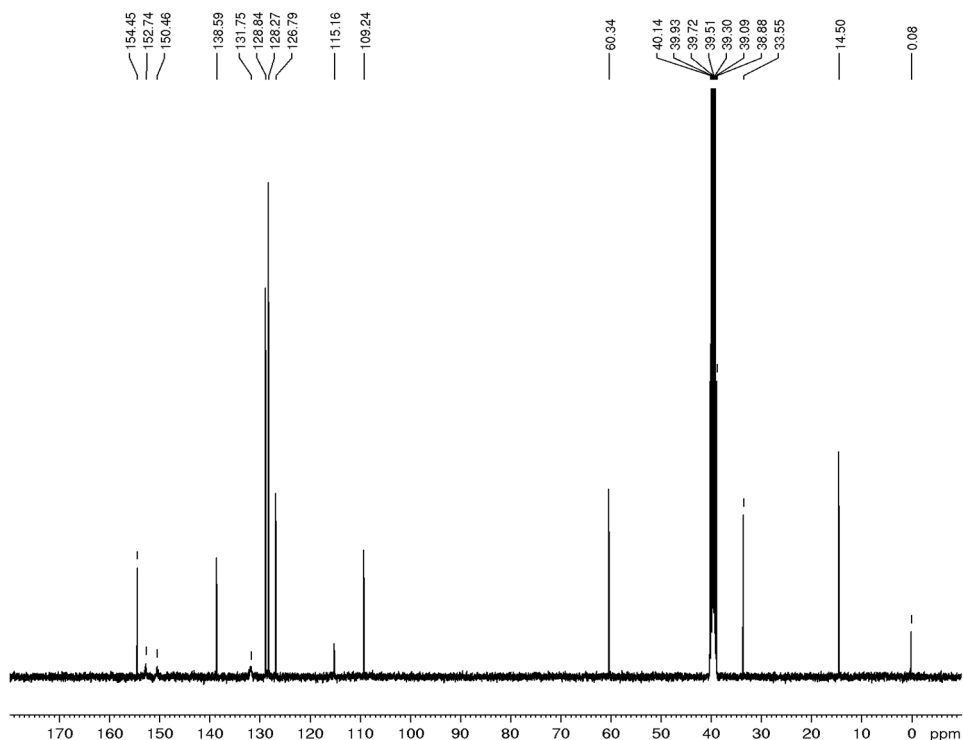
Spectra of 6b



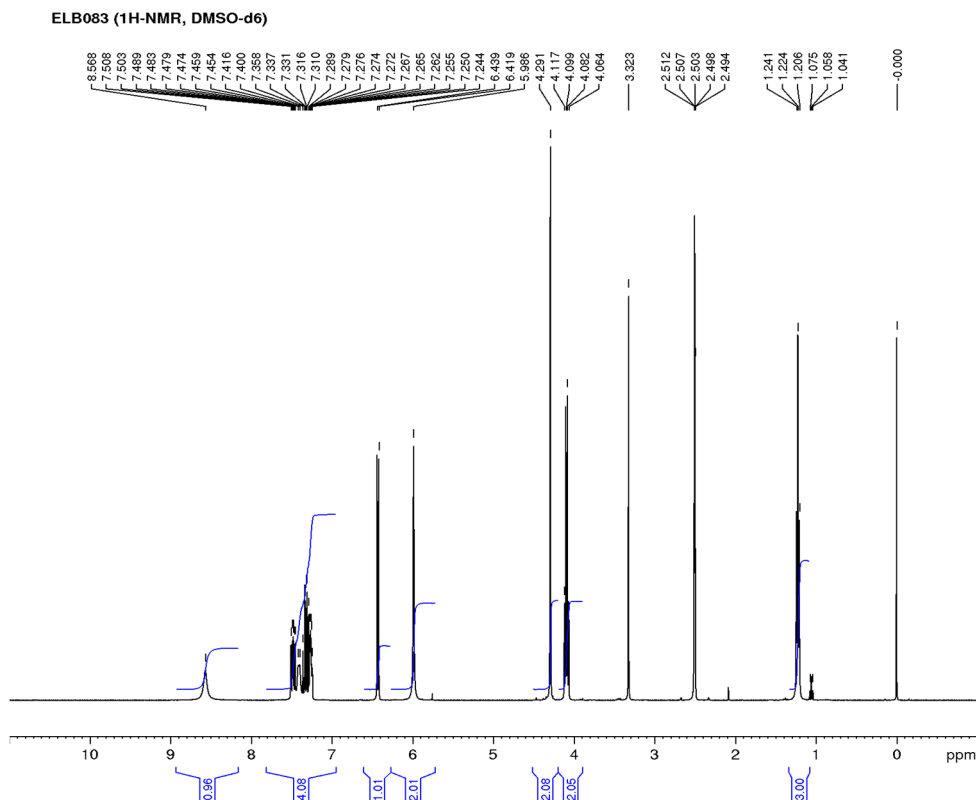
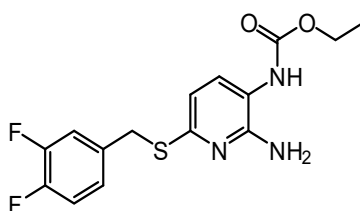
ELB108 (1H-NMR, DMSO-d6)



ELB108 (13C-NMR, DMSO-d6)



Spectra of 6c

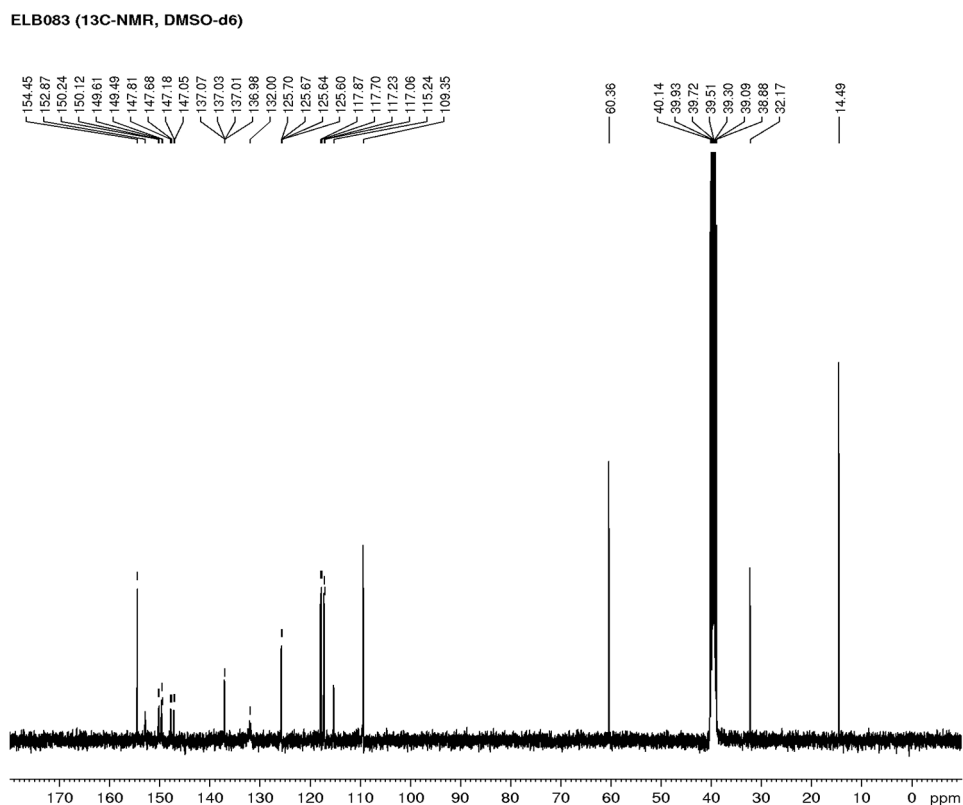


Current Data Parameters
NAME C. Bock
EXPNO 353
PROCNO 1

F2 - Acquisition Parameters
Date_ 20171113
Time 9.47
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 90.5
DW 60.800 usec
DE 10.00 usec
TE 298.1 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 9.75 usec
PL1 -5.00 dB
PL1W 30.18447304 W
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 32768
SF 400.1300020 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



Current Data Parameters
NAME C. Bock
EXPNO 362
PROCNO 1

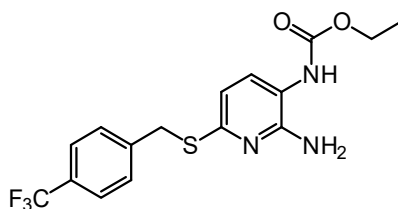
F2 - Acquisition Parameters
Date_ 20171121
Time 9.36
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 1024
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 2050
DW 20.800 usec
DE 10.00 usec
TE 298.3 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
PL1 -2.70 dB
PL1W 62.67650986 W
SFO1 100.6228298 MHz

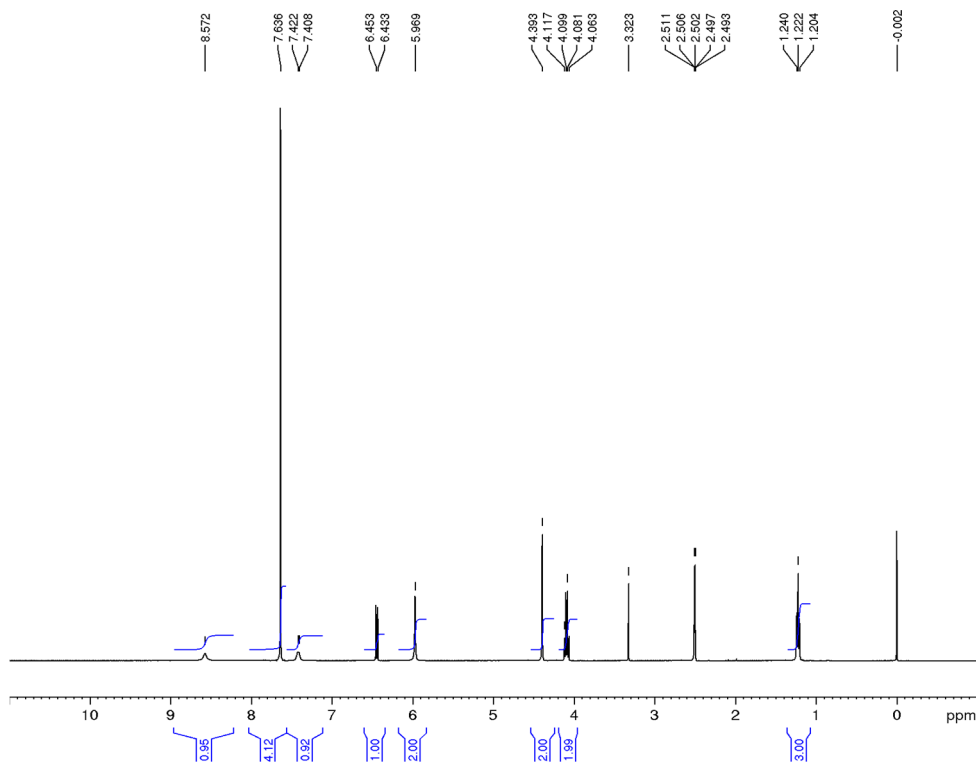
===== CHANNEL f2 =====
CPDPRG[2] waltz16
NUC2 1H
PCPD2 60.00 usec
PL2 -5.00 dB
PL12 10.78 dB
PL13 15.00 dB
PL2W 30.18447304 W
PL12W 0.79759723 W
PL13W 0.30184472 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6128178 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Spectra of 6d



ELB017.2 (1H-NMR, DMSO-d6)



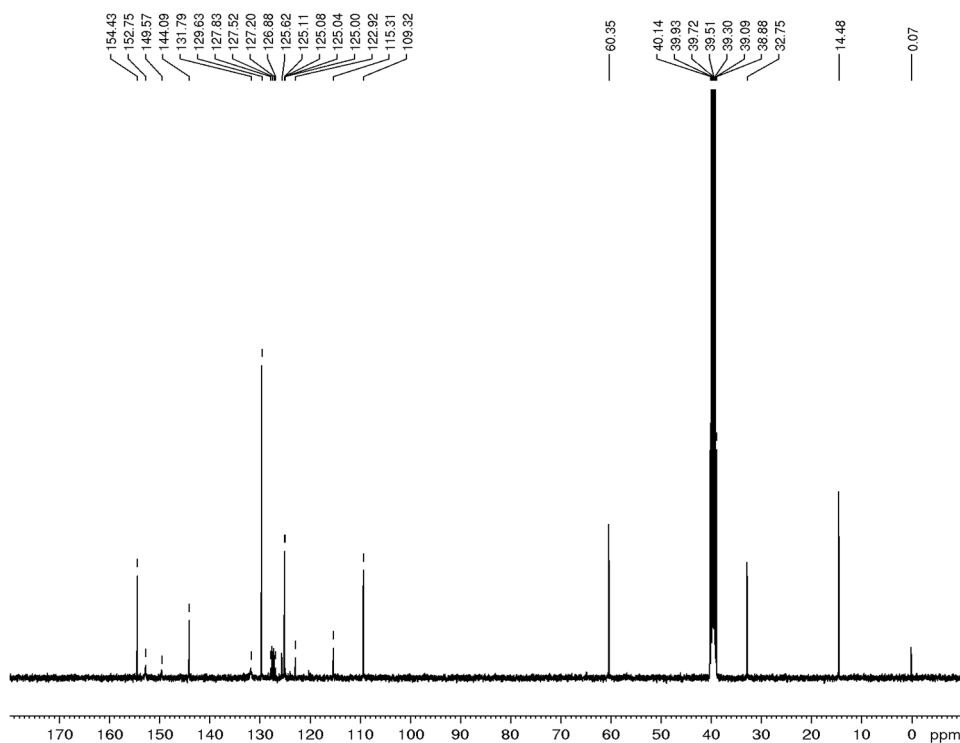
Current Data Parameters
NAME C. Bock
EXPNO 442
PROCNO 1

F2 - Acquisition Parameters
Date_ 20180208
Time 15.16
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 114
DW 60.800 usec
DE 10.00 usec
TE 298.4 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 9.75 usec
PL1 -5.00 dB
PL1W 30.18447304 W
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 32768
SF 400.1300023 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

ELB017.2 (13C-NMR, DMSO-d6)



Current Data Parameters
NAME C. Bock
EXPNO 443
PROCNO 1

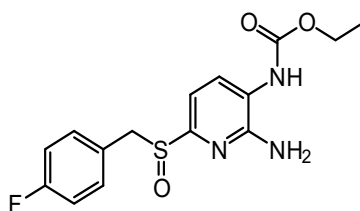
F2 - Acquisition Parameters
Date_ 20180209
Time 9.41
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 1500
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 2050
DW 20.800 usec
DE 10.00 usec
TE 298.1 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
PL1 -2.70 dB
PL1W 62.67650986 W
SFO1 100.6228298 MHz

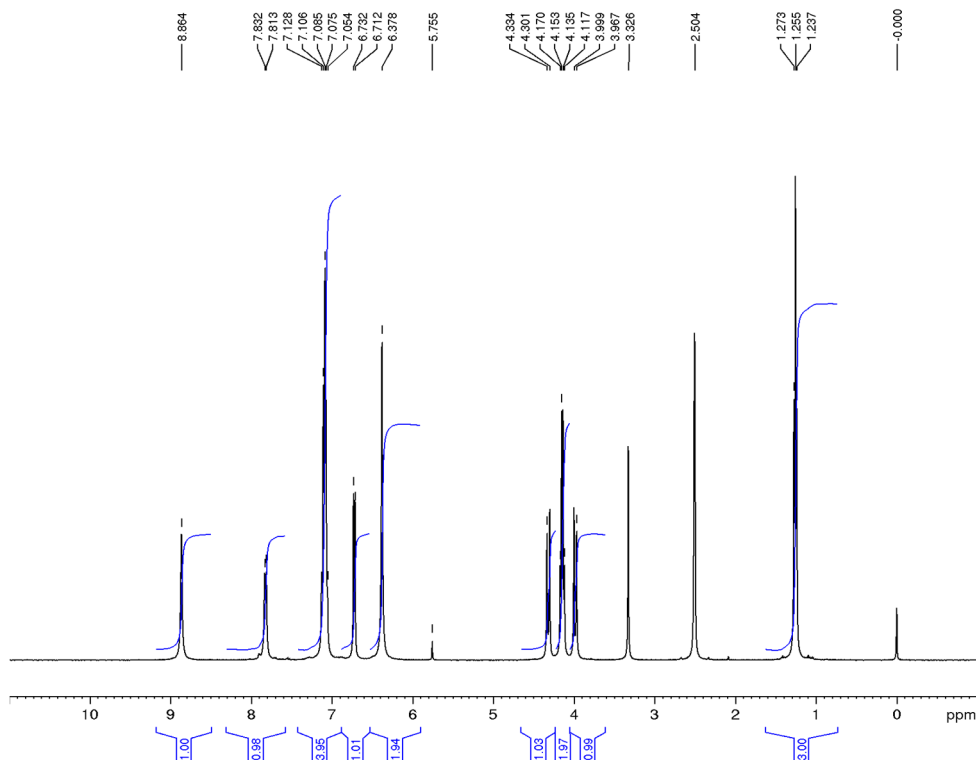
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 60.00 usec
PL2 -5.00 dB
PL12 10.78 dB
PL13 15.00 dB
PL12W 30.18447304 W
PL12W 0.79759723 W
PL13W 0.30184472 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6128181 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Spectra of 7a



ELB084 (1H-NMR, DMSO-d6)



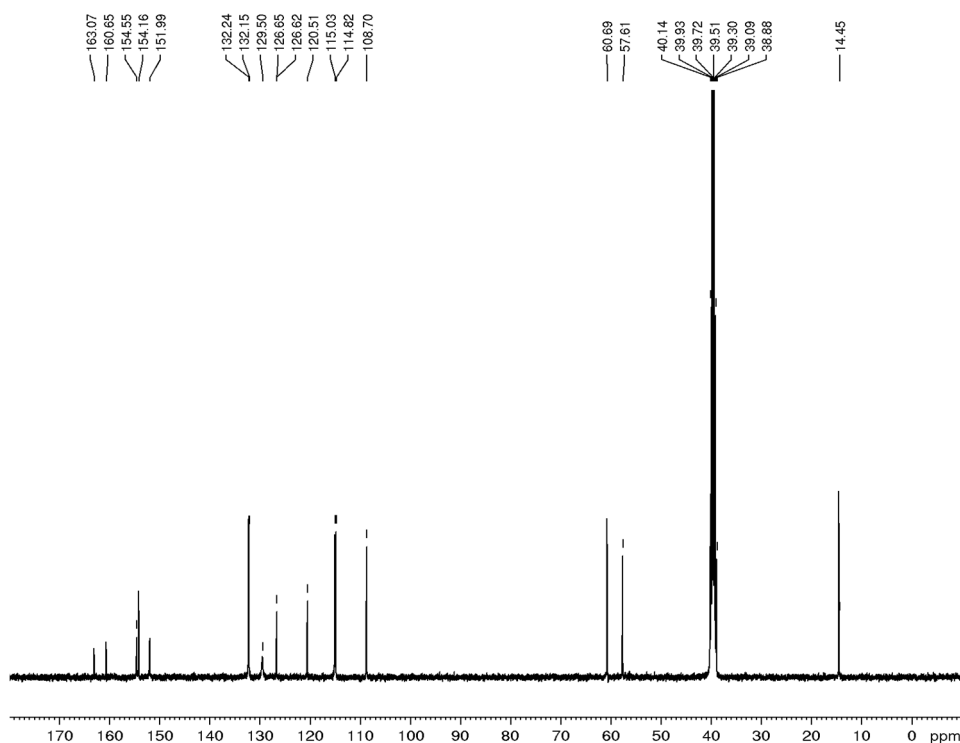
Current Data Parameters
NAME C. Bock
EXPNO 374
PROCNO 1

F2 - Acquisition Parameters
Date_ 20171205
Time 14.50
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 90.5
DW 60.800 usec
DE 10.00 usec
TE 298.3 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 9.75 usec
PL1 -5.00 dB
PL1W 30.18447304 W
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 32768
SF 400.1300014 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

ELB084 (13C-NMR, DMSO-d6)



Current Data Parameters
NAME C. Bock
EXPNO 383
PROCNO 1

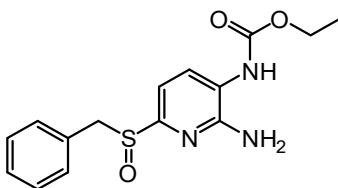
F2 - Acquisition Parameters
Date_ 20171218
Time 9.12
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 1024
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 2050
DW 20.800 usec
DE 10.00 usec
TE 298.3 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
PL1 -2.70 dB
PL1W 62.67650986 W
SFO1 100.6228298 MHz

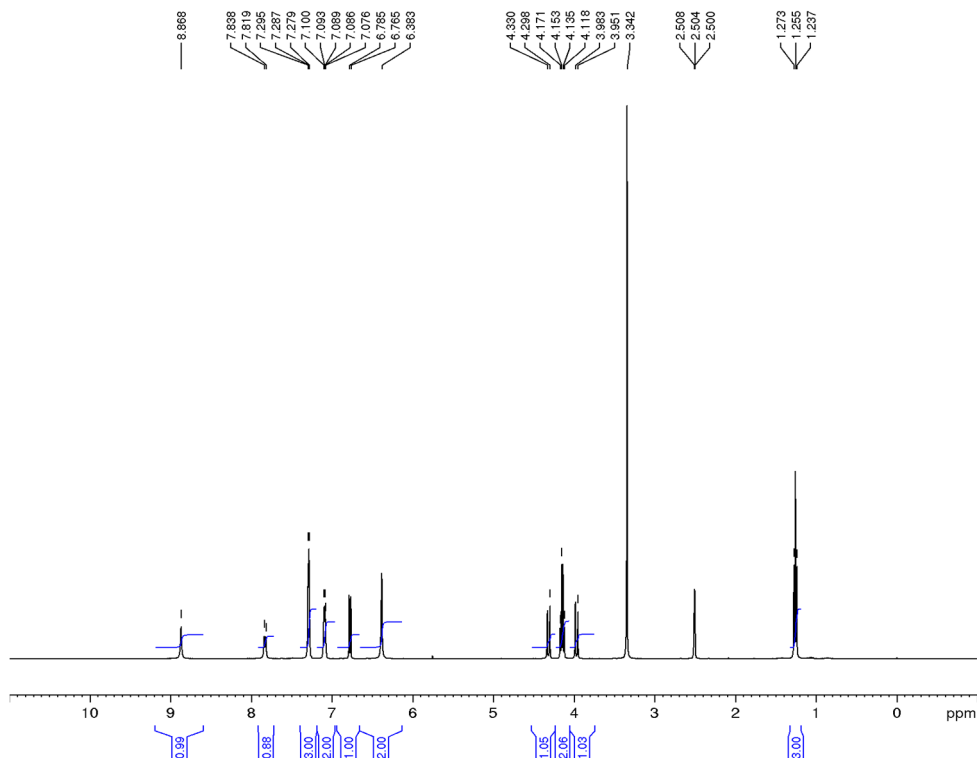
===== CHANNEL f2 =====
CPDPRG[2] waltz16
NUC2 1H
PCPD2 60.00 usec
PL2 -5.00 dB
PL12 10.78 dB
PL13 15.00 dB
PL2W 30.18447304 W
PL12W 0.79759723 W
PL13W 0.30184472 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6128163 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Spectra of 7b



AS-29 (1H-NMR, DMSO-d6)



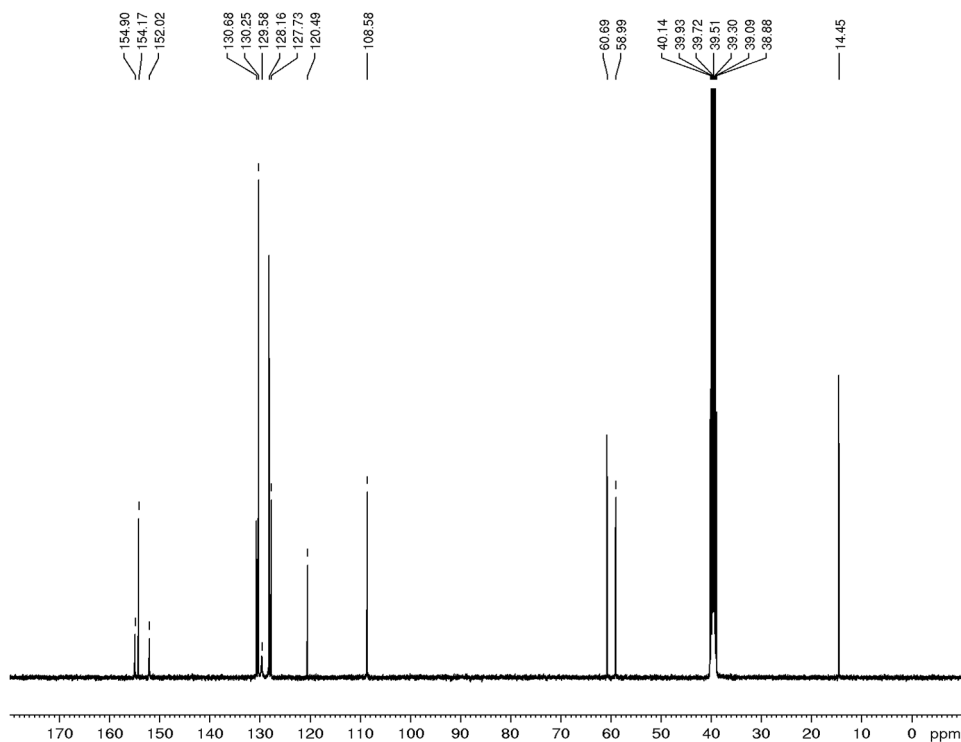
Current Data Parameters
NAME A. Surur
EXPNO 262
PROCNO 1

F2 - Acquisition Parameters
Date_ 20170505
Time 8.08
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 71.8
DW 60.800 usec
DE 10.00 usec
TE 298.2 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 9.75 usec
PL1 -5.00 dB
PL1W 30.18447304 W
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 32768
SF 400.1300016 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

AS-29 (13C-NMR, DMSO-d6)



Current Data Parameters
NAME A. Surur
EXPNO 265
PROCNO 1

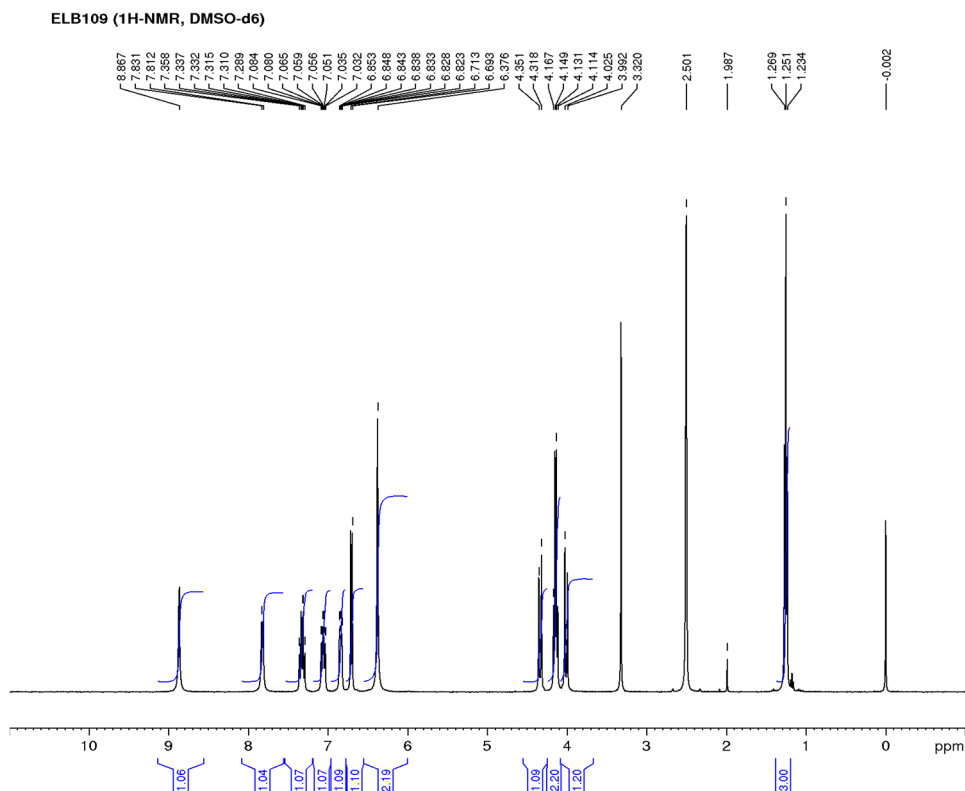
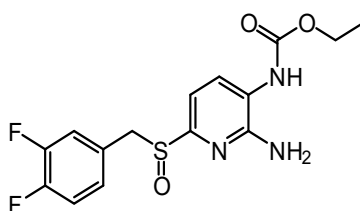
F2 - Acquisition Parameters
Date_ 20170505
Time 17.54
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 4
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 2050
DW 20.800 usec
DE 10.00 usec
TE 299.2 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
PL1 -2.70 dB
PL1W 62.67650986 W
SFO1 100.6228298 MHz

===== CHANNEL f2 =====
CPDPRG[2] waltz16
NUC2 1H
PCPD2 60.00 usec
PL2 -5.00 dB
PL12 10.78 dB
PL13 15.00 dB
PL2W 30.18447304 W
PL12W 0.79759723 W
PL13W 0.30184472 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6128172 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Spectra of 7c

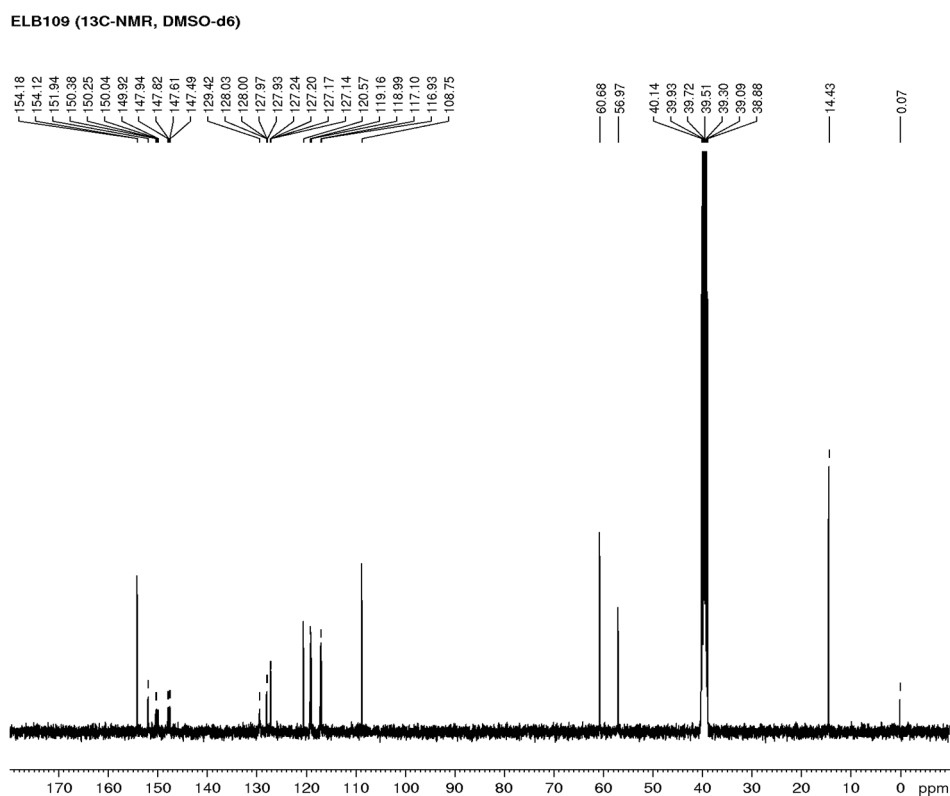


Current Data Parameters
NAME C. Bock
EXPNO 454
PROCNO 1

F2 - Acquisition Parameters
Date_ 20180213
Time 9.49
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 114
DW 60.800 usec
DE 10.00 usec
TE 298.5 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 9.75 usec
PL1 -5.00 dB
PL1W 30.18447304 W
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 32768
SF 400.1300023 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



Current Data Parameters
NAME C. Bock
EXPNO 455
PROCNO 1

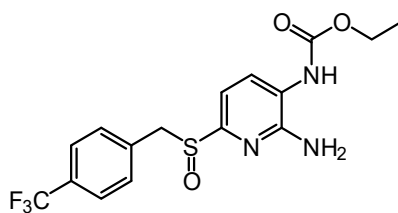
F2 - Acquisition Parameters
Date_ 20180213
Time 12.47
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 1500
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 2050
DW 20.800 usec
DE 10.00 usec
TE 298.2 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
PL1 -2.70 dB
PL1W 62.67650986 W
SFO1 100.6228298 MHz

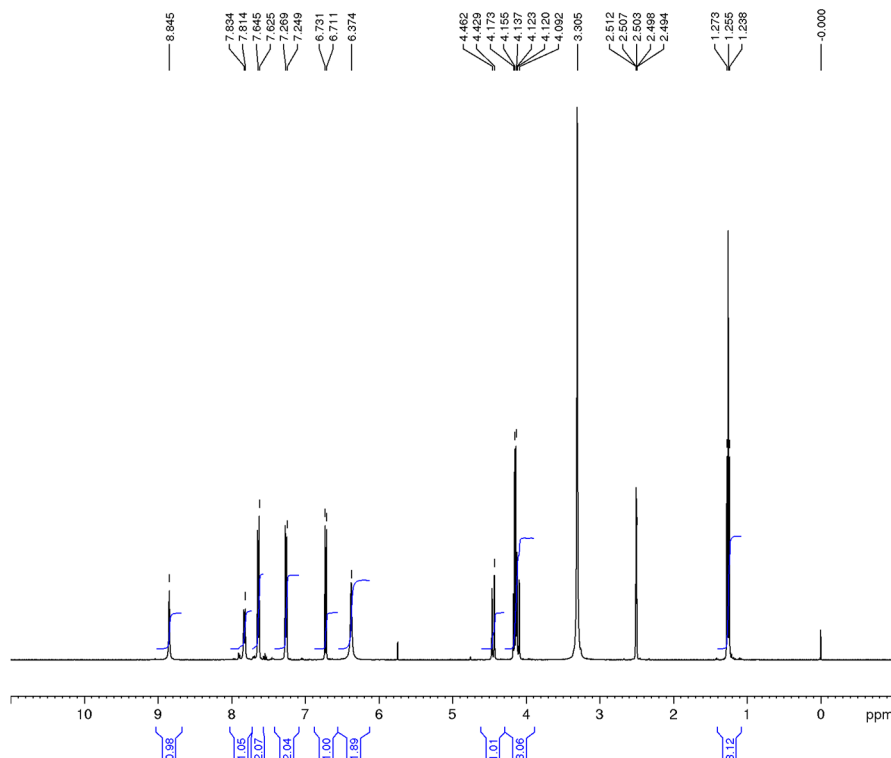
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 60.00 usec
PL2 -5.00 dB
PL12 10.78 dB
PL13 15.00 dB
PL2W 30.18447304 W
PL12W 0.79759723 W
PL13W 0.30184472 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6128180 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Spectra of 7d



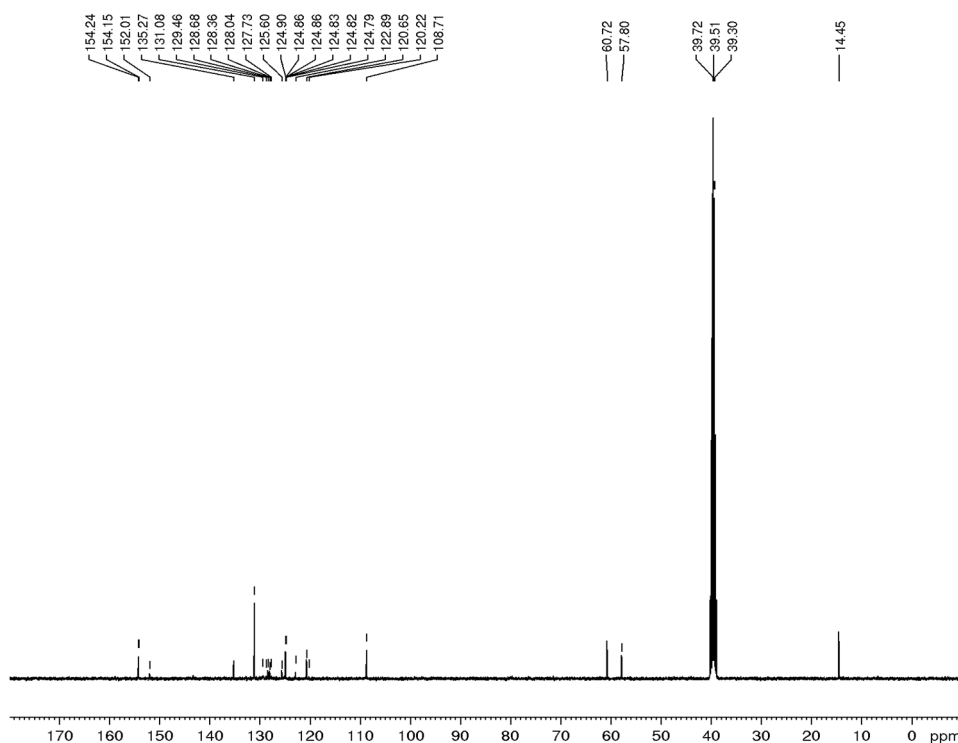
TK21 (1H-NMR, DMSO-d6)



Current Data Parameters
NAME T. Kasch
EXPNO 90
PROCNO 1
F2 - Acquisition Parameters
Date_ 20151001
Time 11.00
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 90.5
DW 60.800 usec
DE 10.00 usec
TE 295.1 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 9.75 usec
PL1 -4.50 dB
PL1W 26.90193939 W
SFO1 400.1324710 MHz
F2 - Processing parameters
SI 32768
SF 400.1300024 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

TK21 (13C-NMR, DMSO-d6)

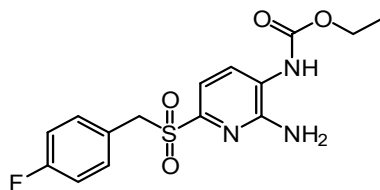


Current Data Parameters
NAME T. Kasch
EXPNO 93
PROCNO 1
F2 - Acquisition Parameters
Date_ 20151006
Time 10.01
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 1024
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 2050
DW 20.800 usec
DE 10.00 usec
TE 285.4 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

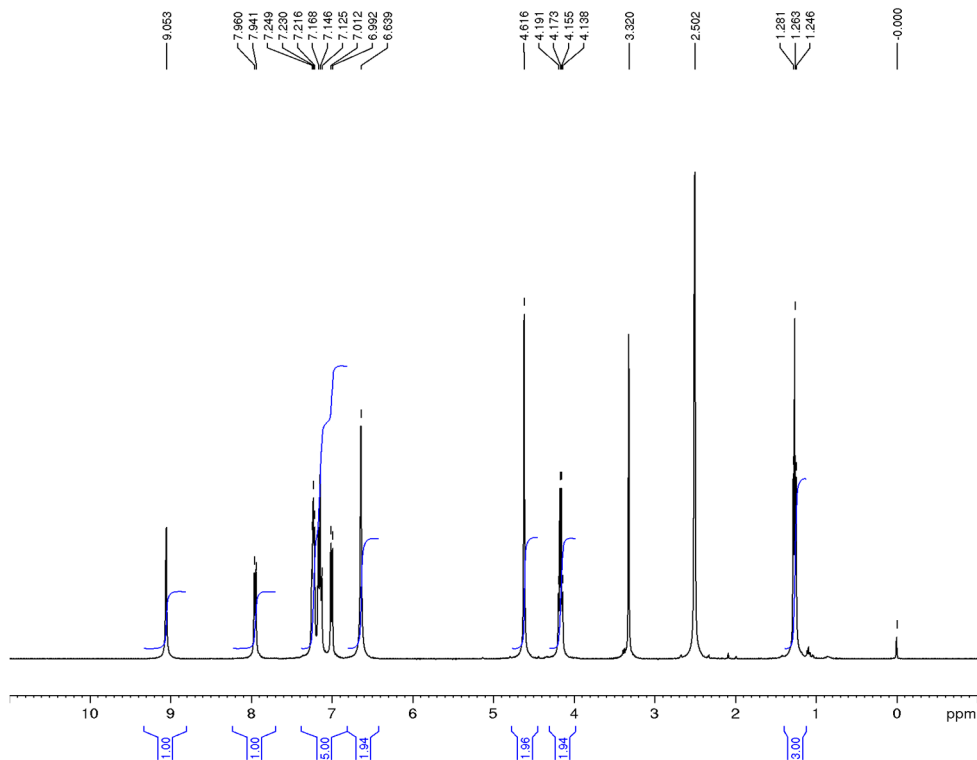
===== CHANNEL f1 =====
NUC1 13C
P1 9.80 usec
PL1 -2.00 dB
PL1W 53.34635925 W
SFO1 100.6228298 MHz
===== CHANNEL f2 =====
CPDPRG[2] waltz16
NUC2 1H
PCPD2 60.00 usec
PL2 -4.50 dB
PL12 11.28 dB
PL13 15.00 dB
PL2W 26.90193939 W
PL12W 0.71088924 W
PL13W 0.30184472 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6128155 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Spectra of 8a



ELB100 (1H-NMR, DMSO-d6)



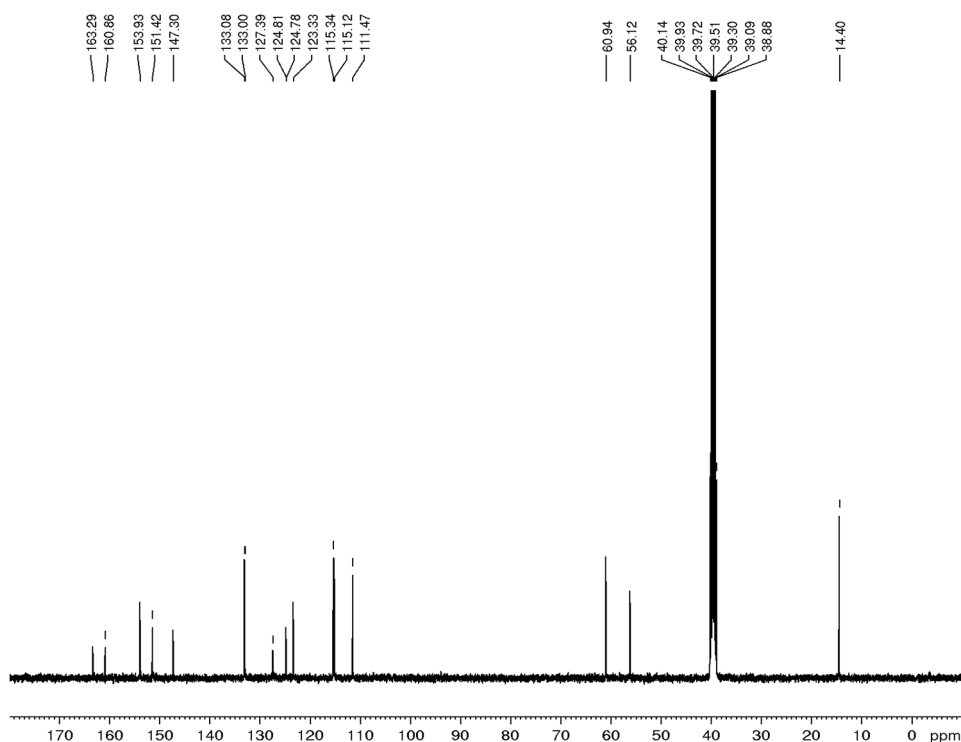
Current Data Parameters
NAME C. Bock
EXPNO 389
PROCNO 1

F2 - Acquisition Parameters
Date_ 20171219
Time 13.45
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 90.5
DW 60.800 usec
DE 10.00 usec
TE 298.5 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 9.75 usec
PL1 -5.00 dB
PL1W 30.18447304 W
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 32768
SF 400.1300017 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

ELB100 (13C-NMR, DMSO-d6)



Current Data Parameters
NAME C. Bock
EXPNO 398
PROCNO 1

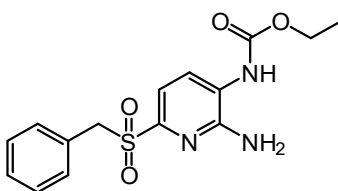
F2 - Acquisition Parameters
Date_ 20180108
Time 14.33
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 1024
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 2050
DW 20.800 usec
DE 10.00 usec
TE 298.2 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
PL1 -2.70 dB
PL1W 62.67650986 W
SFO1 100.6228298 MHz

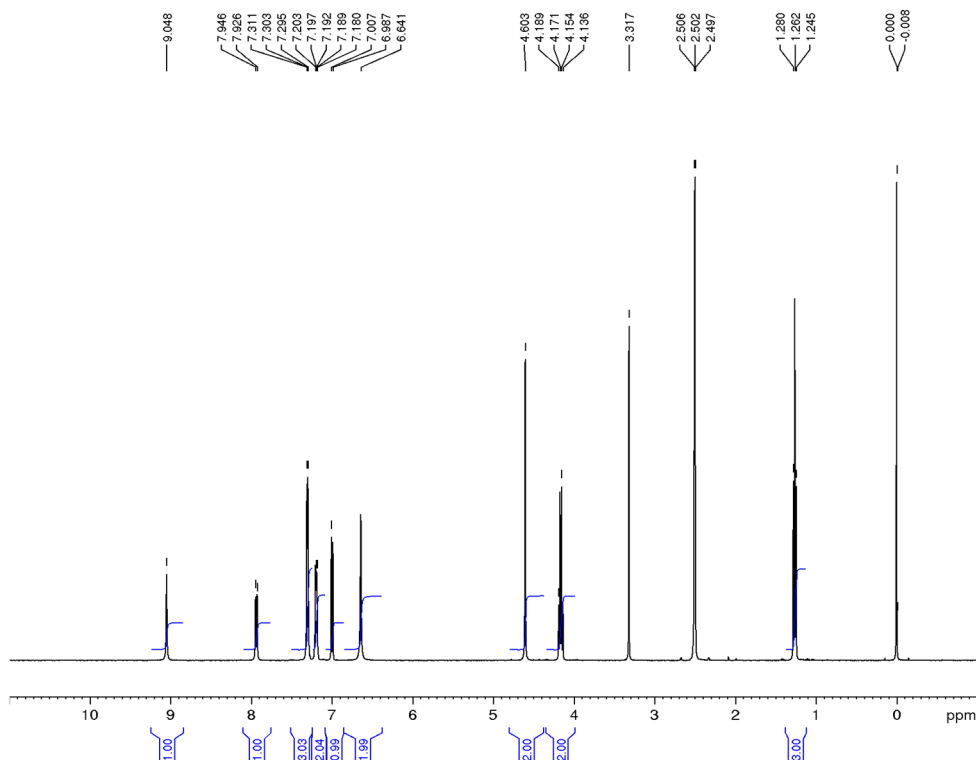
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 60.00 usec
PL2 -5.00 dB
PL12 10.78 dB
PL13 15.00 dB
PL2W 30.18447304 W
PL12W 0.79759723 W
PL13W 0.30184472 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6128162 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Spectra of 8b



ELB110 (1H-NMR, DMSO-d₆)



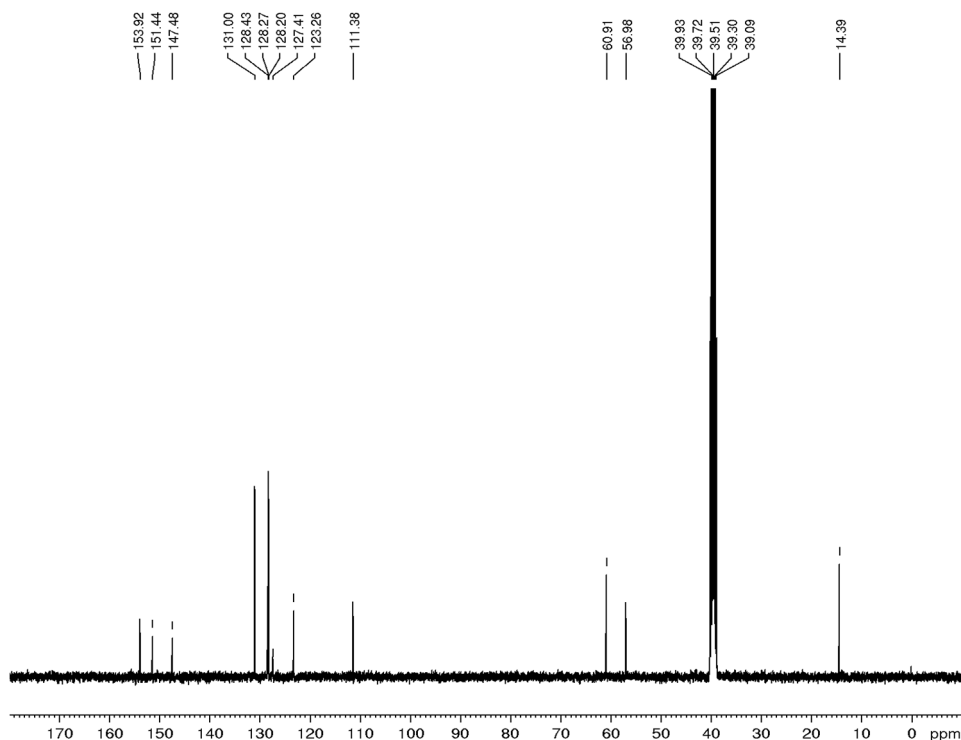
Current Data Parameters
NAME C. Bock
EXPNO 452
PROCNO 1

F2 - Acquisition Parameters
Date_ 20180213
Time 9.37
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 114
DW 60.800 usec
DE 10.00 usec
TE 298.4 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 9.75 usec
PL1 -5.00 dB
PL1W 30.18447304 W
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 32768
SF 400.1300023 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

ELB110 (13C-NMR, DMSO-d₆)



Current Data Parameters
NAME C. Bock
EXPNO 457
PROCNO 1

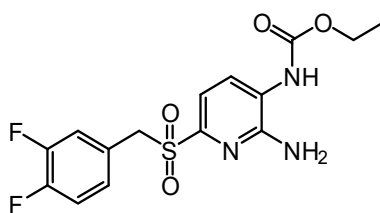
F2 - Acquisition Parameters
Date_ 20180215
Time 13.58
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 1500
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 2050
DW 20.800 usec
DE 10.00 usec
TE 298.2 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
PL1 -2.70 dB
PL1W 62.67650986 W
SFO1 100.6228298 MHz

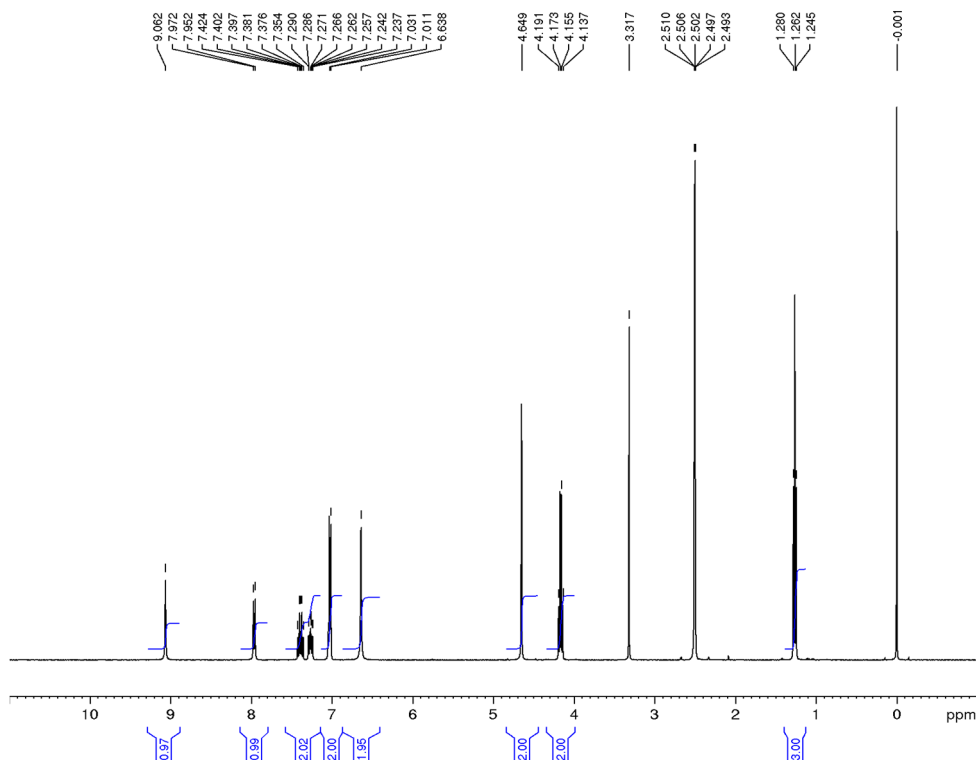
===== CHANNEL f2 =====
CPDPRG[2] waltz16
NUC2 1H
PCPD2 60.00 usec
PL2 -5.00 dB
PL12 10.78 dB
PL13 15.00 dB
PL2W 30.18447304 W
PL12W 0.79759723 W
PL13W 0.30184472 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6128182 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Spectra of 8c



ELB097.2 (1H-NMR, DMSO-d6)



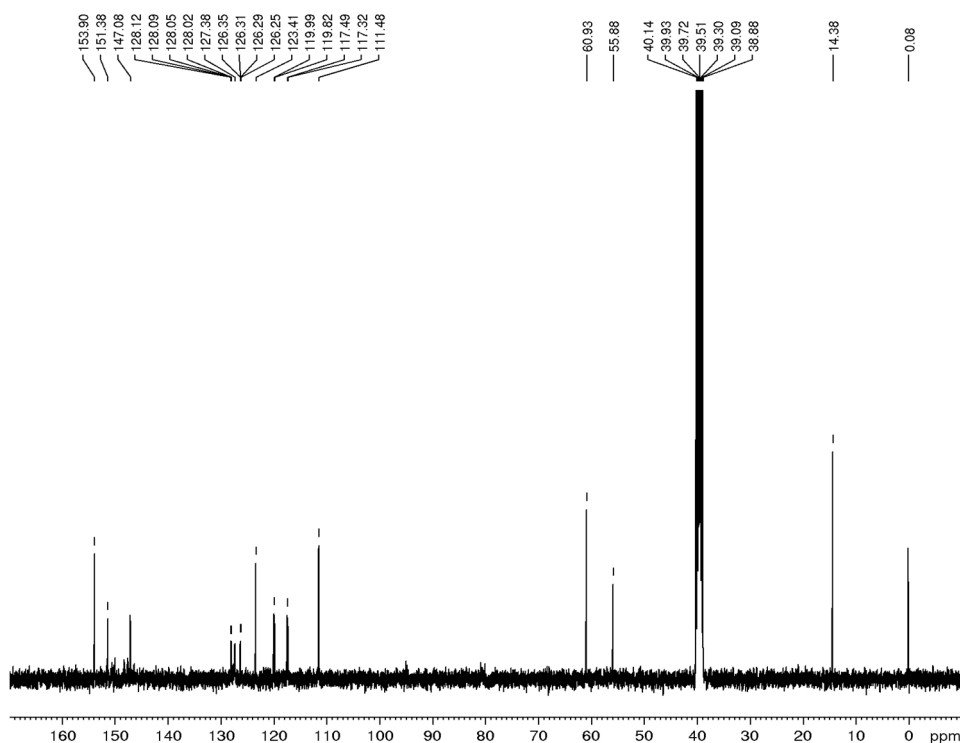
Current Data Parameters
NAME C. Bock
EXPNO 438
PROCNO 1

F2 - Acquisition Parameters
Date_ 20180206
Time 11.25
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 128
DW 60.800 usec
DE 10.00 usec
TE 298.5 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 9.75 usec
PL1 -5.00 dB
PL1W 30.18447304 W
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 32768
SF 400.1300023 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

ELB097.2 (13C-NMR, DMSO-d6)



Current Data Parameters
NAME C. Bock
EXPNO 439
PROCNO 1

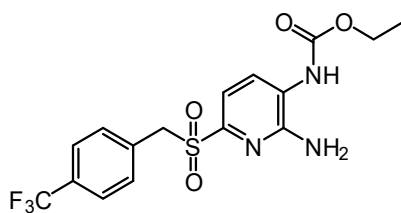
F2 - Acquisition Parameters
Date_ 20180206
Time 12.51
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 1500
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 2050
DW 20.800 usec
DE 10.00 usec
TE 298.4 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
PL1 -2.70 dB
PL1W 62.67650986 W
SFO1 100.6228298 MHz

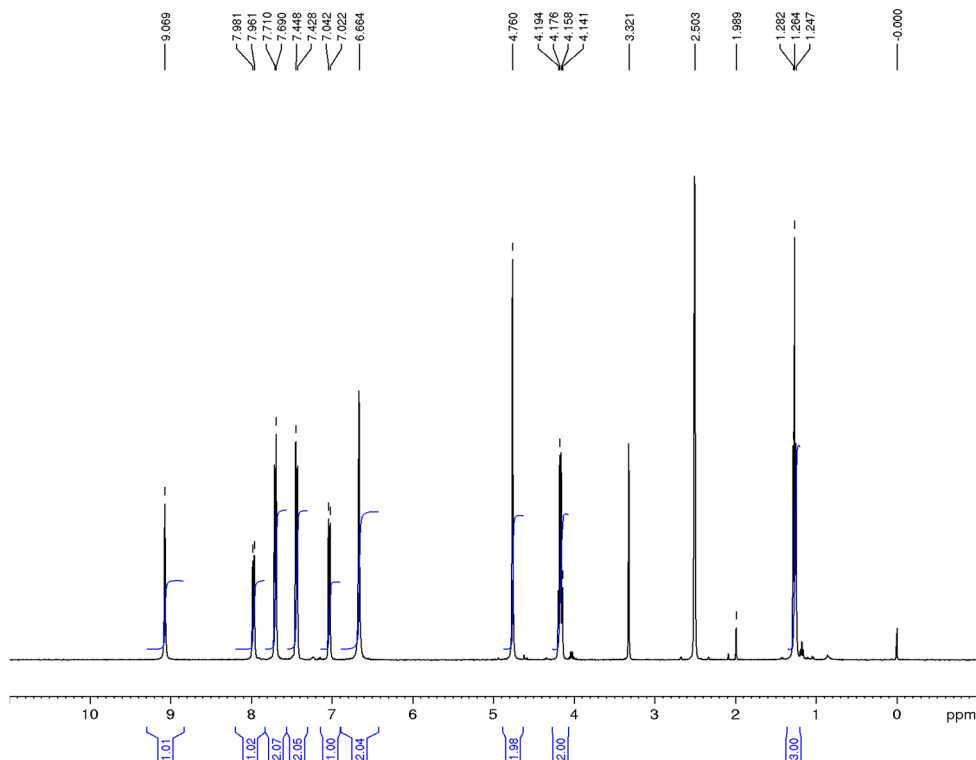
===== CHANNEL f2 =====
CPDPRG[2] waltz16
NUC2 1H
PCPD2 60.00 usec
PL2 -5.00 dB
PL12 10.78 dB
PL13 15.00 dB
PL2W 30.18447304 W
PL12W 0.79759723 W
PL13W 0.30184472 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6128183 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Spectra of 8d



ELB099 (1H-NMR, DMSO-d6)



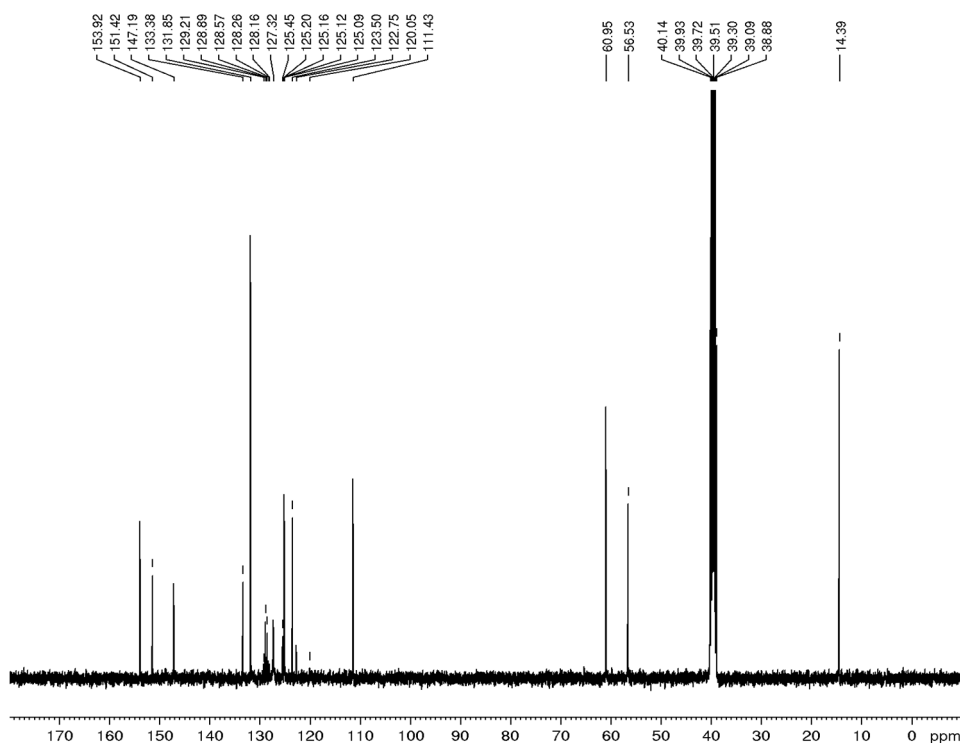
Current Data Parameters
NAME C. Bock
EXPNO 388
PROCNO 1

F2 - Acquisition Parameters
Date_ 20171219
Time 13.40
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 90.5
DW 60.800 usec
DE 10.00 usec
TE 297.9 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 9.75 usec
PL1 -5.00 dB
PL1W 30.18447304 W
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 32768
SF 400.1300015 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

ELB099 (13C-NMR, DMSO-d6)



Current Data Parameters
NAME C. Bock
EXPNO 400
PROCNO 1

F2 - Acquisition Parameters
Date_ 20180109
Time 8.50
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 1024
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 2050
DW 20.800 usec
DE 10.00 usec
TE 298.1 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
PL1 -2.70 dB
PL1W 62.67650986 W
SFO1 100.6228298 MHz

===== CHANNEL f2 =====
CPDPRG[2] waltz16
NUC2 1H
PCPD2 60.00 usec
PL2 -5.00 dB
PL12 10.78 dB
PL13 15.00 dB
PL2W 30.18447304 W
PL12W 0.79759723 W
PL13W 0.30184472 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6128162 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Biological Evaluations

Cell culture

TGF- α transgenic mouse hepatocytes (TAMH) were provided by Sidney D. Nelson (formerly Department of Medicinal Chemistry, University of Washington, Seattle, USA). HEK293 cells expressing Kv7.2/3 were purchased from SB Drug Discovery (Glasgow, UK). TAMH cells were maintained in a T75 flask with Dulbecco's Modified Eagle's Medium/Nutrient Mixture F-12 (DMEM/F12 1:1 mix) (PAN Biotech), supplemented with 5 % PANEXIN NTA (PAN Biotech), 10 mM nicotinamide (Sigma Aldrich) and 10 μ g/mL gentamicin sulfate (PAN Biotech). HEK293 cells were cultured in a T75 flask with minimum essential medium with non-essential amino acids (MEM) (Thermo Fisher Scientific), supplemented with 10% heat-inactivated fetal bovine serum (FBS) (Sigma Aldrich), 2 mM L-glutamine (Sigma Aldrich), 4 μ g/mL blasticidin S-HCl (Merck), 1% penicillin/streptomycin (Thermo Fisher Scientific) and 0.78 mg/mL G418 sulfate (Carl Roth). Both cell lines were incubated at 37 °C in a humidified incubator with 95% air/5% CO₂.

MTT assay

TAMH cells were detached from flask when ~90% confluent with Trypsin-EDTA (Sigma Aldrich), resuspended in the presence of 0.25 mg/mL soybean trypsin inhibitor (Sigma Aldrich) in DMEM/F12. After centrifugation, the cell pellet was resuspended in DMEM/F12 media. Cells were counted with an EVE™ Automated Cell Counter (NanoEnTek) and 20 000 cells/well/200 μ L DMEM/F12 media were seeded into 96 well plates (#83.3924.300, Sarstedt). Plates were incubated until cells became confluent (usually 48 h after seeding). In triplicate, medium was removed and replaced by 200 μ L/well DMEM/F12 media containing a serial dilution series of test compounds (5-9 concentrations) prepared in DMSO (BioScience-grade) (Carl Roth). Untreated controls contained 1% DMSO and wells lacking cells were used to determine background signal. After 24 h exposure to the test compounds, medium was aspirated off, 125 μ L DMEM/F12 media supplemented with 20% (V/V) of 2.5 mg/mL solution of 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-tetrazolium bromide (MTT) (Alfa Aesar) in phosphate-buffered saline (PBS) was added to wells and plates were incubated 2.5-4 h in a humidified incubator at 37 °C. After removing the medium and generated formazan crystals were dissolved in 50 μ L DMSO (for synthesis) (Carl Roth). Absorbance was measured at λ = 570 nm with SpectraMax 190 microplate reader (Molecular Devices). Cell viability (T/C_{corr}) was obtained by dividing absorbance values for each treatment condition through that measured for control wells expressed in % (both corrected for background without cells). The LD₅₀ is the estimated concentration that reduces the T/C_{corr} by 50%. This value was determined by plotting (log)concentration of test compound versus T/C_{corr} and performing linear regression analysis in Microsoft Excel (2013). When even the highest tested soluble concentration of a compound could not reduce cell viability to or below 50%, LD₅₀ was indicated as greater than the highest tested concentration. Determined values are the mean of at least three independent experiments $\pm$ standard deviation (SD).

Kv7.2/3 channel opening assay

HEK293 cells were detached from flask by TrypLE Express (Thermo Fisher Scientific) when ~80% confluent and resuspended in MEM media. After centrifugation the cell pellet was again resuspended in MEM media. Cells were counted with an EVE™ Automated Cell Counter and 60 000 cells/well/100 μ L MEM media were seeded into

black walled 96 well plates with clear bottom (4titude). Cells were allowed to attach for 24 h in a humidified incubator at 37 °C, forming a near confluent monolayer. FLIPR® Potassium Assay Kit (Molecular Devices) was used to determine $K_{v7.2/3}$ channel opening activity of test compounds according to manufacturer's instructions. Briefly, equal amount of Loading Buffer containing 5 mM probenecid (Santa Cruz Biotechnology) (pH 7.4) was added to wells and incubated for 1 h at room temperature in the dark. Serial diution series of test compounds (5-9 concentrations) were prepared in DMSO (BioScience-grade) (Carl Roth) and incubated with cells for another 30 min at room temperature in the dark. Control wells contained 1% DMSO. Plates were placed in an Infinite® F200 Pro plate reader (Tecan) and background fluorescence (baseline) for each well was bottom read at excitation/emmission wavelength: 485nm/535 nm for approximately 20 s. Then, 50 μ L Stimulus Buffer ($c(K^+) = 25$ mM, $c(Tl^+) = 15$ mM) was added to each well and fluorescence intensity was recorded immediately every 2.5-3 s continuously for further 2 min. The recorded data were transfered to Microsoft Excel (2013). The ratio readout of maximal fluorescence intensity change of a compound's concentration at a specific timepoint was divided through the average of fluorescence intensities of baseline ($\Delta F/F$) and analogous determined value for control was substracted (corr. $\Delta F/F$). At the chosen specific time point the sum of $\Delta F/F$ of all concentrations belonging to a dilution series of a compound became maximal. Corr. $\Delta F/F$ were plotted versus log(concentration) of a compound, fitted to a four parameter logistic equation and EC_{50} values were calculated according to the software GraphPad Prism 6 (La Jolla, California, USA). Indicated EC_{50} values are the mean of at least three independent experiments $\pm$ standard deviation (SD).

Log $D_{7.4}$ determination

Log $D_{7.4}$ values were determined using a HPLC method. Column: Phenomenex Kinetex PFP 2.6 μ M 75 $\times$ 4.6 mm, flow rate 0.5 mL/min, mobile phase: **A**: 95 % ammonium formiate buffer (21.05 mM, pH 7.4) and 5 % methanol, **B** 5 % ammonium formiate buffer (0.36 mM, pH 7.4) and 95 % methanol, gradient: 0-50 min 0 $\rightarrow$ 100 % **B**, 50-52 min 100 $\rightarrow$ 0 % **B**, 52-60 min 0 % **B**.

Table of reference compounds with known log P and measured log k'

reference	ret1	ret2	ret3	ret4	mean	k'	log k'	log P (lit.)
uracil	2.71	2.72	2.72	2.72	2.72			
pyridine	11.92	11.92	11.92	11.92	11.92	3.39	0.53	0.65
benzyl alcohol	14.95	14.96	14.95	14.95	14.95	4.50	0.65	1.1
acetanilide	16.26	16.26	16.26	16.25	16.26	4.98	0.70	1.16
picoline	19.09	19.10	19.09	19.08	19.09	6.02	0.78	1.2
acetophenone	24.15	24.16	24.16	24.15	24.15	7.89	0.90	1.7
methyl benzoate	29.39	29.40	29.39	29.39	29.39	9.82	1.00	2.1
ethyl benzoatee	33.65	33.65	33.65	33.65	33.65	11.38	1.06	2.6
benzophenone	37.19	37.18	37.18	37.18	37.18	12.68	1.10	3.2
phenyl benzoate	39.18	39.17	39.18	39.18	39.18	13.42	1.13	3.6
diphenyl ether	40,85	40,85	40,85	40,85	40,85	14,03	1,15	4,2
bibenzyl	43,95	43,94	43,95	43,95	43,95	15,17	1,18	4,8
triphenylamine	47,87	47,86	47,87	47,87	47,87	16,61	1,22	5,7

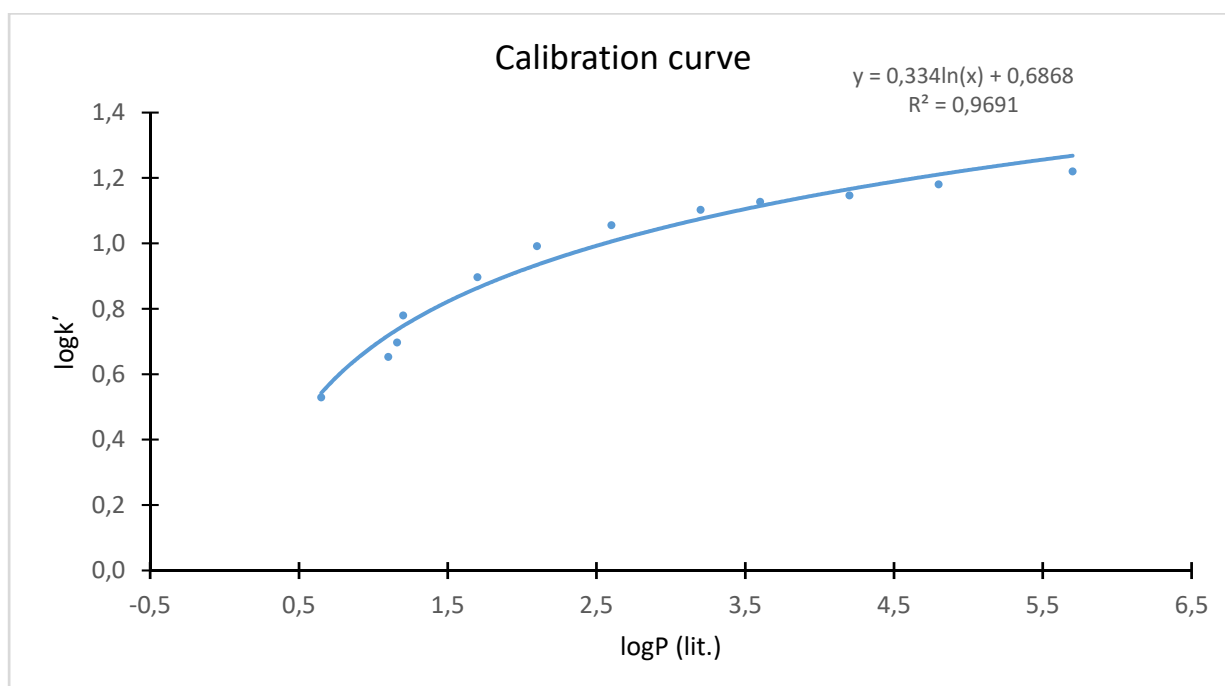
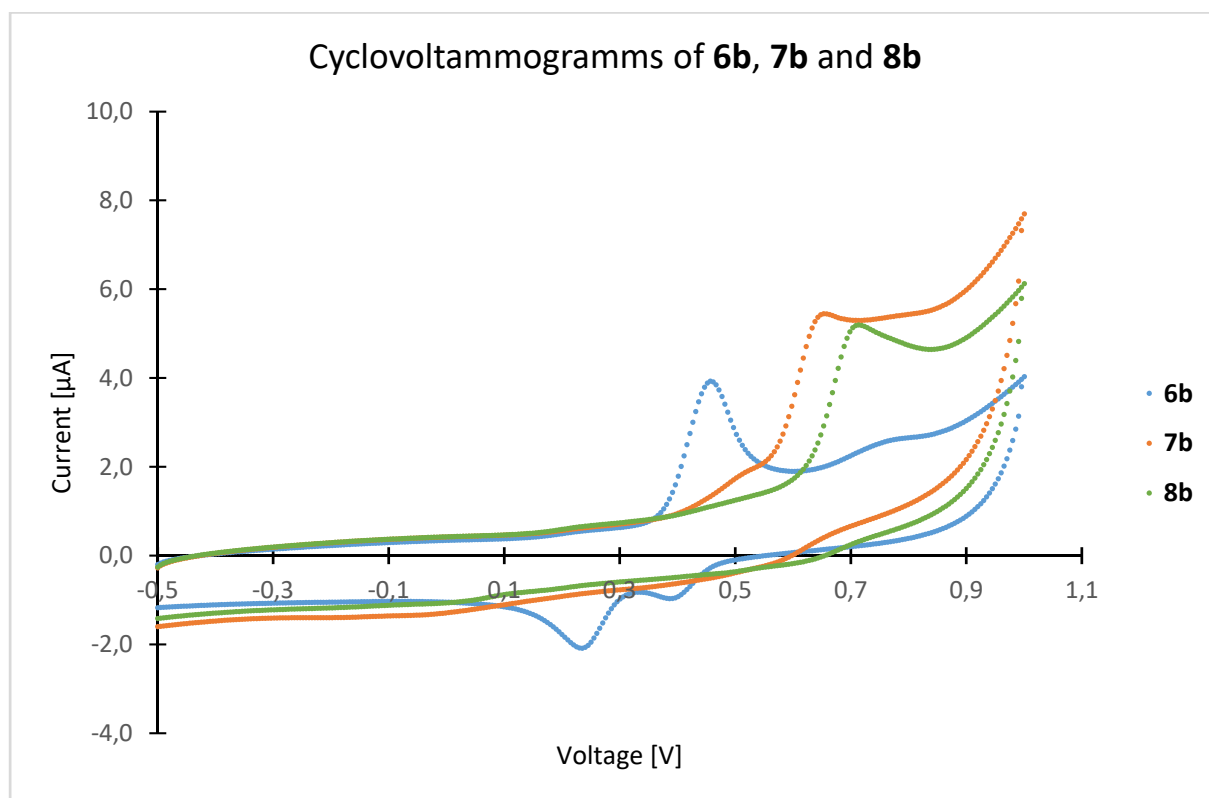


Table of synthesized compounds with measured $\log k'$ and calculated $\log D_{7.4}$

cmpd	ret 1	ret 2	ret3	mean	k'	$\log k'$	$\log D_{7.4}$
1	33.78	33.66	n.d.	33.72	11.21	1.05	2.96
6a	38.22	38.23	38.23	38.226	12.84	1.11	3.54
6b	37.54	37.53	37.54	37.54	12.60	1.10	3.45
6c	39.19	39.21	n.d.	39.20	13.20	1.12	3.66
6d	41.08	41.08	41.07	41.08	13.88	1.14	3.91
7a	31.07	31.08	31.08	31.08	10.26	1.01	2.64
7b	29.94	29.95	29.96	29.95	9.85	0.99	2.50
7c	32.37	32.37	32.38	32.37	10.72	1.03	2.80
7d	35.63	35.63	35.62	35.63	11.90	1.08	3.20
8a	33.26	33.25	33.25	33.25	11.04	1.04	2.91
8b	32.02	32.01	32.02	32.02	10.60	1.03	2.75
8c	34.59	34.59	34.58	34.59	11.53	1.06	3.07
8d	37.29	37.29	37.29	37.29	12.51	1.10	3.42

Cyclic voltammetry

Oxidation potentials of the compounds were obtained using the 797 VA Computrace from Metrohm. Working electrode: glassy carbon, reference electrode: Ag/AgCl/KCl 3 M, auxiliary electrode: Pt. 10 μmol of the compound were suspended in 10 mL of TRIS buffer (pH 7.4) and sonicated for 3 min. The mixture was poured into the glass vial and degassed by nitrogen for 3 min. Starting at -0.5 V the voltage was increased to 1.0 V with a speed of 0.1 V/s, while the current was recorded in 0.05 V steps. Once 1.0 V was reached, the voltage was inverted until -0.5 V with the same speed to obtain a full cycle. 5 of those cycles were recorded but only the first sweep was used to determine the anodic peak potential ($E_{p,A}$) as it gave the strongest signal. The consecutive cycles showed lower currents, most likely because of polymerization of the oxidized compounds on the surface of the working electrode. Thus the graphite surface had to be polished after every measurement with an alumina oxide suspension in water which was distributed on a patch of cloth. The $E_{p,A}$ was determined automatically using the software: 797 VA Computrace 1.3.1.



	1. measurement		2. measurement		3. measurement		mean	s.d.
cmpd.	E_{pa} [mV]	I_{pa} [μA]	E_{pa} [mV]	I_{pa} [μA]	E_{pa} [mV]	I_{pa} [μA]	E_{pa} [mV]	E_{pa} [mV]
1	255	6.21	220	0.46	225	1.36	233.33	18.93
6a	472	1.05	467	0.355	452	0.106	463.67	10.41
6b	457	0.34	462	0.41	457	0.48	458.67	2.89
6c	457	0.81	462	0.67	462	0.4	460.33	2.89
6d	457	2.74	457	1.42	452	1.89	455.33	2.89
7a	653	1.13	648	0.61	658	0.48	653.00	5.00

7b	628	0.14	628	0.15	623	0.29	626.33	2.89
7c	633	0.35	633	0.38	652	0.28	639.33	10.97
7d	633	1.34	653	0.69	648	2.2	644.67	10.41
8a	714	1.35	709	1.39	729	1.24	717.33	10.41
8b	709	0.95	714	1.07	709	1.49	710.67	2.89
8c	729	2.59	714	3.11	714	4.21	719.00	8.66
8d	709	1.75	714	2.27	704	2.32	709.00	5.00

Computational methods

All calculations were performed using the Molecular Operating Environment (MOE) software suite (version 2018.01). The default parameters were used if not explicitly specified.

Heteromeric K_v7.2/7.3 multimer homology modeling

The sequences of the voltage-gated potassium channels K_v7.2 and K_v7.3 were taken from the UniProt entries O43526 and O43525. Both were aligned to the Kv1.2 template (PDB 2R9R) in an alternating manner. Only the first 351 amino acids, containing the membrane-associated S1-S6 subunits, were taken into account. All four, pore-forming chains were modeled at once to prevent unusual sidechain positions. A total of 10 main chain models with 10 sidechain models each were calculated at a temperature of 300K. The final homology model was protonated with Protonate3D and energy-optimized.

Docking

The binding site was defined as all residues within 4.5 Å of residue W265, which was previously described as an important amino acid for the binding of closely related drug retigabine. The placement of 30 poses was performed using the Triangle Matcher with London dG scoring and flexible ligands. All generated binding poses were refined with an Induced Fit and MM/GBVI scoring. A total of 10 final poses for each ligand was calculated, visually inspected and cross-checked with additional 20 compounds from literature and patents using the same procedure and the obtained retigabine binding mode as template.