Electronic supplementary information

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Syntheses of preussin until 2002

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General Information

Reactions were generally performed under argon in flame-dried flasks and the components were added by syringe. Solvents were dried using standard procedures (abbreviations in the following text: DMSO = dimethylsulfoxide, THF = tetrahydrofuran). Reagents were purchased and used without further purification. Potassium *t*-butoxide was freshly sublimed in vacuo before use.

Products were purified by flash chromatography on neutral aluminum oxide (neutral, 6% water, activity III, Merck-Schuchardt or Fluka) or on silica gel (32-63 μ m, Merck-Schuchardt or Fluka). HPLC was performed with nucleosil 50-5 columns (dimensions: analytical, 4x245 mm; preparative, 16x244 mm or 32x237 mm); detection by Knauer variable UV detector (λ = 255 nm) and Knauer refractometer. Unless otherwise stated, yields refer to analytically pure samples.

¹H NMR [CHCl₃ (δ = 7.26 ppm), TMS (δ = 0.00 ppm) as internal standard] and ¹³C NMR spectra [CDCl₃ (δ = 77.0 ppm) as internal standard] were recorded with Bruker WH 270 and DRX 500 instruments in CDCl₃ solutions. Integrals are in accordance with assignments; coupling constants are given in Hz. Multiplicity of signals is indicated as follows: s (singlet), s_{br} (broad singlet), d (doublet), br d (broad doublet), dd (doublet of doublet), t (triplet), m (multiplet), m_c (centered multiplet). ¹H/¹³C-correlated spectra (HSQC, HMBC) and ¹H/¹H-correlated spectra (COSY, NOESY) were recorded for assignments. Optical rotations [α]_D were measured with a Perkin Elmer 241 polarimeter in a 1 mL microcuvette at the temperature given. IR spectra were measured with Nicolet 5 SXC FTIR or Nicolet 205 FTIR spectrometers. MS analyses were performed with MAT 711, MAT 112 S and with HP 5890 II instruments. The elemental analyses were recorded with "Elemental-Analyzers" (Perkin-Elmer or Carlo Erba). Melting points were measured with a Reichert apparatus (Thermovar) or a Gallenkamp apparatus (MPD 350) and are uncorrected.

General procedure for the addition of lithiated alkoxyallenes to aldehydes or imines (GP1): For generation of the lithiated alkoxyallene, the corresponding alkoxyallene was dissolved in THF and *n*-butyllithium was added at -40 °C. After 30 min, the solution was cooled to -80 °C and the corresponding electrophile was added. The mixture was allowed to warm up to -30 °C and stirred for the time given in the individual experiments. After quenching with saturated aqueous NaHCO₃ solution (10 mL/), the organic phase was separated and the aqueous phase was extracted with diethyl ether (3 x 15 mL/mmol of alkoxyallene). The combined organic phases were dried (Na₂SO₄), filtered and evaporated in vacuo to provide the crude product that was used directly or purified as given in the individual experiment. The yields refer to the amount of electrophile used.

General procedure for silver nitrate-promoted cyclization (GP2): To a solution of the corresponding allenyl alcohol or amine in acetonitrile (5 mL/mmol) were added under a stream of argon via a funnel potassium carbonate and silver nitrate. The resulting mixture was stirred at room temperature under light exclusion for 12 h, then filtered and evaporated. The residue was dissolved in a small amount ethyl acetate and filtered through a pad of Celite (elution with ethyl acetate). After removal of the solvents in vacuo, the crude product was purified as indicated in the individual experiments.

N-(3-Methoxy-1-phenylpenta-3,4-dien-2-yl)-*p*-toluenesulfonamide (10): According to **GP1**, methoxyallene **7**^[1] (0.12 mL, 1.44 mmol), *n*-butyllithium (0.59 mL, 1.44 mmol of 2.44 M solution in hexanes) and **9** (0.308 g, 0.72 mmol) in THF (15 mL) provided after 2 h crude **10** (0.237 g) as light yellow solid (melting range 80-90 °C, decomposition). The product was not purified but directly used in the cyclization step. ¹H NMR (CDCl₃, 270 MHz): δ = 2.39 (s, 3 H, Tos-Me), 2.90, 2.96 (AB part of ABX system, *J*_{AB} = 13.8 Hz, *J*_{AX} = 7.4 Hz, *J*_{BX} = 6.3 Hz, 1 H each, 1-H), 3.12 (s, 3 H, OMe), 4.05–4.17 (m, 1 H, 2-H), 4.83 (d, *J* = 9.6 Hz, 1 H, NH), 5.03, 5.08 (AB system, *J*_{AB} = 7.4 Hz, 2 H, 5-H), 7.08 (d, *J* = 8.1 Hz, 2 H, Tos), 7.18–7.35 (m, 5 H, Ph), 7.65 ppm (d, *J* = 8.1 Hz, 2 H, Tos). ¹³C NMR (CDCl₃, 67.9 MHz): δ = 21.4 (q, Tos-Me), 39.9 (t, C-1), 55.9, 56.2 (q, d, OMe, C-2), 91.8 (t, C-5), 126.5, 127.2, 128.1, 129.1, 129.5 (5 d, Ph, Tos), 130.9 (s, C-3), 136.6, 137.8, 143.0 ppm (3 s, Ph, Tos), 197.1 (s, C-4). IR (KBr): $\tilde{\nu}$ = 3260 (N-H), 3065–2830 (C-H), 1960 (C=C=C), 1325, 1160 cm⁻¹ (Tos-N). MS (EI, 80 eV): *m/z* (%) = 343 (0.3) [M⁺], 274 (2) [BnCHNHTos⁺], 252 (36) [M⁺ - Bn], 172 (49), 155 (24), 91 (100) [Bn⁺].

2-Benzyl-3-methoxy-1-tosyl-2,5-dihydropyrrole (11): According to **GP2**, crude allenyl amine **10** (0.300 g, 0.88 mmol) in acetonitrile (10 mL) was treated with AgNO₃ (0.029 g, 0.17 mmol) and K₂CO₃ (0.243 g, 1.76 mmol) for 12 h to provide crude **11**. Purification by column chromatography (silica gel, hexanes/ethyl acetate 8:1) furnished **11** (0.180 g, 60%) as colorless crystals (m.p. 128 °C). ¹H NMR (CDCl₃, 500 MHz): δ = 2.43 (s, 3 H, Tos-Me), 3.01, 3.31 (AB part of ABX system, *J*_{AB} = 13.4 Hz, *J*_{AX} = 2.6 Hz, *J*_{BX} = 4.4 Hz, 1 H each, PhC*H*₂), 3.50 (s, 3 H, OMe), 3.52 (dd, *J* = 12.5 Hz, *J* = 5.0 Hz, 1 H, 5-H) 3.81 (dd, *J* = 12.5 Hz, *J* = 1.7 Hz, 1 H, 5-H), 4.14 (s_{br}, 1 H, 4-H), 4.45–4.55 (m, 1 H, 2-H), 7.17–7.32 (m, 5 H, Ph), 7.32, 7.73 ppm (2 d, *J* = 8.1 Hz, 2 H each, Tos). ¹³C NMR (CDCl₃, 125.8 MHz): δ = 21.5 (q, Tos-Me), 38.9 (t, Ph*C*H₂), 52.4 (t, C-5), 56.6 (q, OMe), 64.3 (d, C-2), 90.0 (d, C-4), 126.1, 127.3, 127.6, 129.7, 130.4 (5 d, Ph, Tos), 134.3, 136.2, 143.4 (3 s, Ph, Tos), 154.4 ppm (s, C-3). IR (KBr): $\tilde{\nu}$ = 3030–

2860 (C-H), 1670 (C=C), 1335, 1165 cm⁻¹ (Tos-N). $C_{19}H_{21}NO_3S$ (343.4): calcd. C 66.45, H 6.16, N 4.08; found C 6

Attempt to epimerize *trans*-14 into *cis*-14: *trans*-14 (0.011 g, 0.02 mmol) was dissolved in dichloromethane (4 mL) and stirred with DBU (0.033 g, 0.2 mmol) for 2 h at room temperature. The proceeding of the reaction was followed by TLC. The mixture was filtered through a short pad of silica gel (elution with hexanes/ethyl acetate 4:1) and after evaporation of the solvents a yellow oil (0.007 g) was obtained, that was identified as a mixture of *cis*-14 and compound 15 (ratio ca. 1:1).

Conversion of *cis*-14 into compound 15 and 16: *cis*-14 (0.016 g, 0.04 mmol) was dissolved in dichloromethane (15 mL) and stirred with DBU (0.1 g, 0.66 mmol) for 5.5 h at room temperature. The proceeding of the reaction was followed by TLC. The mixture was filtered through a short pad of silica gel (elution with hexanes/ethyl acetate 3:1) and after evaporation of the solvents a yellow oil (0.005 g, ca. 48%) was obtained. ¹H NMR analysis showed that a mixture of 15 and 16 (ratio ca. 3:1, 16 as 1:1 mixture of *E/Z* isomers). ¹H NMR (CDCl₃, 270 MHz), signals of 15: δ = 0.89 (t, *J* = 7.0 Hz, 3 H, Me), 0.90–1.50 (m, 16 H, CH₂), 2.05, 2.53 (AB part of ABX system, J_{AB} = 19.9 Hz, J_{AX} = 2.5 Hz, J_{BX} = 5.9 Hz, 1 H each, 4-H), 3.82 (s, 2 H, PhC*H*₂), 4.20–4.32 (m, 1 H, 5-H), 7.20–7.40 ppm (m, 5 H, Ph); characteristic signals of the *E/Z* isomers of 16: (*E/Z* ca. 1:1): δ = 2.25, 2.72 (AB system of ABX system, J_{AB} = 18.4 Hz, J_{AX} = 5.9 Hz, J_{BX} = 6.6 Hz, 1 H each, 4-H), 4.98 (s, 1 H, NH), 5.10 (d, *J* = 4.1 Hz, 1 H, NH), 6.07 (s, 0.5 H, =CPh*H*), 6.60 ppm (d, *J* = 4.1 Hz, 0.5 H, =CPh*H*). ¹³C NMR (CDCl₃, 67.9 MHz), signals of 15: δ = 14.2 (q, Me), 22.6, 26.1, 29.3*, 29.5*, 31.9, 33.9, 36.3, 40.1 (8 t, CH₂, PhCH₂, C-4), 65.3 (d, C-5), 126.7, 128.6, 128.8, 131.0 (3 d, s, Ph), 171.4 (s, C-2), 203.0 ppm (s, C-3); * signals with higher intensity; the signals of *E/Z*-16 are too weak to be unambiguously identified in this mixture. MS (EI, 80 eV): *m/z* (%) = 299 (1) [M⁺], 298 (14) [M⁺ - H], 297 (38), 198 (36), 185 (57), 57 (89). 44(100).

2-Benzyl-3-methoxy-5-nonyl-2,5-dihydropyrrole (cis-17): A solution of sodium naphthalenide (0.63 mL of a 1 M solution in DME, prepared according to ref. 2) was added to a solution of *cis*-14 (0.099 g, 0.21 mmol) in DME (10 mL) at -78 °C. After 1 h the mixture was quenched with water (5 mL) and concentrated in vacuo. The residue was taken up in saturated aqueous NaHCO3 solution (10 mL) and extracted with ethyl acetate (3 x 10 mL). The combined organic phases were dried (Na₂SO₄), filtered and evaporated in vacuo to obtain the crude product (0.150 g) as yellow solid. Purification by column chromatography (alumina III, hexanes/ethyl acetate 10:1 + 1% of NEt₃) gave *cis*-17 (0.050 g, 75%) as yellow oil. ¹H NMR (CDCl₃, 270 MHz): δ = 0.81 (t, J = 6.3 Hz, 3 H, Me), 1.00–1.30 (m, 16 H, CH₂), 1.42 (s_{br}, 1 H, NH), 2.63, 2.93 (AB part of ABX system, J_{AB} = 14.0 Hz, J_{AX} = 7.4 Hz, J_{BX} = 3.7 Hz, 1 H each, PhCH₂), 3.58 (s, 3 H, OMe), 3.65–3.72, 3.95–4.05 (2 m, 1 H each, 2-H, 5-H), 4.46 (s, 1 H, 4-H), 7.08– 7.25 ppm (m, 5 H, Ph). ¹³C NMR (CDCl₃, 67.9 MHz): δ = 14.1 (q, Me), 22.7, 26.1, 29.3, 29.6*, 29.7, 31.9, 38.8 (7 t, CH₂), 40.7 (t, PhCH₂), 57.0 (q, OMe), 61.4, 63.0 (2 d, C-2, C-5), 97.2 (d, C-4), 126.1, 128.1, 129.6, 138.6 (3 d, s, Ph), 159.9 ppm (s, C-3); * signal with higher intensity. IR (film): $\tilde{v} = 3400$ (N-H), 3060, 3025, 2925, 2855 (C-H), 1655 cm⁻¹ (C=C). MS (FAB): m/z (%) = 316 (81) [M⁺ + H], 284 (14) [M⁺ -OMe], 224 (52) [M⁺ - Bn], 188 (23) [C₉H₁₉⁺], 91 (100) [Bn⁺]. C₂₁H₃₃NO (315.5): calcd. C 79.95, H 10.54, N 4.44; found C 79.34, H 10.51, N 4.08.

Reaction of (-)-preussin with Mosher's chloride:

To a solution of (-)-preussin (8.3 mg, 26 μ mol) in a 1:1 mixture of dichloromethane and pyridine (1 mL) Mosher's chloride (14 mg, 54 μ mol) was added dropwise. After 3 d of stirring dichloromethane was added (5 mL) and the solution was extracted with aqueous 2 M HCl (2 x 5 mL), with saturated aqueous NaHCO₃ solution (2 x 5 mL) and water (5 mL). The organic phase was dried (Na₂SO₄), filtered and evaporated in vacuo to give the crude product (10 mg) as orange oil. ¹H NMR spectroscopy indicated a conversion of ca. 30% and analytical HPLC showed that the formed Mosher's ester product is diastereomerically pure. Purification by HPLC (hexanes/ethyl acetate 95:5) provided pure product (4 mg, 28 %). ¹H NMR (CDCl₃, 270 MHz): δ = 0.89 (t, *J* = 6.8 Hz, 3 H, Me), 1.15–1.40 (m, 14 H, CH₂), 1.46 (ddd, *J* = 14.6 Hz, *J* = 7.8 Hz, *J* = 2.4 Hz, 1 H, 4-H), 1.65–1.75, 2.10–2.20 (2 m, 1 H each, CH₂), 2.28 (s, 3 H, NMe), 2.40–2.55, 2.67–2.80 (2 m, 5 H, PhCH₂, 2-H, 4-H, 5-H), 3.61 (s, 3 H, OMe), 6.87–6.93, 7.09–7.14, 7.40–7.45, 7.57–7.63 ppm (4 m, 2 H, 3 H, 3 H, 2 H, Ar).

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Copies of NMR spectra























































