Supporting Information

For

Comparative Study of Intrachain Cyclization and Solution Property of Long Subchain Hyperbranched Polymers Prepared via Y-type and V-type Macromonomer Approaches

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Sample	M _n / (g/mol)	M _w / (g/mol)	$M_{\rm w}/M_{\rm n}$	Sample	M _n / (g/mol)	M _w / (g/mol)	$M_{\rm w}/M_{\rm n}$
PS _{Y1}	2100	2400	1.16	PS _{V1}	2400	2700	1.13
PS _{Y2}	4000	4500	1.12	PS _{V2}	6900	7600	1.10
PS _{Y3}	6600	7500	1.13	PS _{V3}	11000	12100	1.10
PS _{Y4}	11700	13000	1.11	PS _{V4}	17600	19500	1.11
				PS _{V5}	29600	34000	1.15

Table S1. Molecular parameters of prepared polystyrene macromonomers



Figure S1. ¹H NMR spectra of ATRP functional initiators I (V-type) and II (V-type).



Figure S2. ¹H NMR spectra of V-type and Y-type polystyrene macromonomers.



Figure S3. (a) and (b) FTIR spectra of V-type and Y-type azide/alkyne functionalized polystyrene macromonomers. (c) and (d) FTIR spectra of HPS_V and HPS_Y hyperbranched samples.



Figure S4. SEC curves of the resultant hyperbranched polystyrenes obtained in our own lab, where the reaction was conducted at T = 35 °C and C = 0.30 g/mL, and the blue curves represent the Gaussian fitting curves for the macromonomer peaks, where only the right half side of the whole peaks are shown (the fitting parameters can be found in Figure S5).



Figure S5. Fitting parameters of macromonomer peaks in Figure S4 by Gaussian mode.



Figure S6. Macromonomer molar mass (M_m) dependence of weight fraction (A_c/A_e) of cyclized macromonomer for HPS_V and HPS_Y samples prepared at different reaction concentrations (C = 0.30 g/mL and C = 0.15 g/mL).



Figure S7. TD-SEC curves of HPS_{V5}, where the solid line, dashed line and hollow cycle represent the signals from RI, LS and viscosity detectors, respectively.



Figure S8. (a)-(d) Total molar mass (M_t) dependence of intrinsic viscosity ([η]) of HPS_V and HPS_Y samples measured by TD-SEC system in THF at T = 35 °C, where (a) and (b) present the datas in all ranges of molar mass, and (c) and (d) present the datas only in the valid regions of molar mass in TD-SEC measurement.



Figure S9. (a) and (b) Cumulative molar mass distributions of HPS_V and HPS_Y hyperbranched samples. (c) and (d) Differential molar mass distributions of HPS_V and HPS_Y hyperbranched samples.



Figure S10. Total molar mass (M_t) dependence of normalized intrinsic viscosity $([\eta]/M_m^{\mu})$ of HPS_V samples, where the intrinsic viscosity is normalized by a factor of M_m^{μ} , and $\mu = 0.15, 0.20, 0.25$ and 0.30 was tested as normalization exponent in (a)-(d), respectively.



Figure S11. Total molar mass (M_t) dependence of normalized intrinsic viscosity ([η]/ M_m^{μ}) of HPS_Y samples, where the intrinsic viscosity is normalized by a factor of M_m^{μ} , where $\mu = 0.15, 0.20, 0.25$ and 0.30 was tested as normalization exponent in (a)-(d), respectively.



Figure S12. (a) Retention volume ($V_{retention}$) dependence of SEC signals, molar mass (M) and radius of gyration (R_g) of HPS_{V2}. (b) Retention volume ($V_{retention}$) dependence of SEC signals, molar mass (M) and radius of gyration (R_g) of HPS_{V5}. (c) Total molar mass (M_t) dependence of radius of gyration (R_g) of HPS_V and HPS_Y samples measured by TD-SEC system in THF at T = 35 °C, where the data in the green region (Figure c) corresponds to the data in the green region in Figure a and Figure b.