## **Supporting Information**

# Open-cage silsesquioxane necklace polymers having closed-cage silsesquioxane pendants

Hiroaki, Imoto; Ryoichi, Katoh; Kensuke, Naka\*

Faculty of Molecular Chemistry and Engineering, Graduate School of Science and Technology, Kyoto Institute of Technology, Goshokaido-cho, Matsugasaki, Sakyo-ku, Kyoto 606-8585, Japan. E-mail: kenaka@kit.ac.jp

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### 1. Materials

Tetrahydrofuran (THF), toluene, triethylamine (NEt<sub>3</sub>), and magnesium sulfate anhydrous (MgSO<sub>4</sub>) were purchased from Nacalai Tesque (Kyoto, Japan). Distilled water was purchased from Wako Pure Chemical Industry (Osaka, Japan). Chlorodimethylvinylsilane, 1,1,3,3-tetramethyldisiloxane (**5b**) were purchased from Tokyo Chemical Industry (Tokyo, Japan). Xylene solution (0.1 M) of platinum(0)-1,3divinyl-1,1,3,3-tetramethyldisiloxane (Pt(dvs)), 1,1,3,3,5,5-hexamethyltrisiloxane (**5a**), and 1,4-bis(dimethylsilyl)benzene (**5c**) were purchased from Sigma-Aldrich (Hattiesburg, Mississippi, US). Heptaphenyl trisilanol POSS (**1**<sub>Ph</sub>) and heptaisobutyl trisilanol POSS (**1**<sub>IBu</sub>) was purchased from Hybrid Plastics Inc (Hattiesburg, Mississippi, US). SiliaMetS(R) Thiol (40-63 µmol 60 Å, functionalized Silica) was purchased from SiliCycle, Inc. Dimethylvinylsilane-substituted heptaphenyl IC-POSS (**2**<sub>Ph</sub>) and heptaisobutyl IC-POSS (**3**<sub>Ph</sub>) were prepared by following the literatures.

#### 2. Measurements

<sup>1</sup>H (400MHz) and <sup>13</sup>C (100MHz) nuclear magnetic resonance (NMR) spectra were recorded on a Bruker DPX-400 spectrometer (Bruker Biospin GmbH, Rheinstetten, Germany) in CDCl<sub>3</sub> using Me<sub>4</sub>Si as an internal standard. The following abbreviations are used: s, singlet; d, doublet; t, triplet; m, multiplet; br, broad. Molecular weights were determined by size exclusion chromatography (SEC) of LC-6AD (Shimadzu, Kyoto, Japan) with Shodex KF-805L (Showa Denko, Tokyo, Japan), and then analysed by refractive index with RID-20A (Shimadzu, Kyoto, Japan). Preparative highperformance liquid chromatography (HPLC) for purification was performed on LC-6AD (Shimadzu, Kyoto, Japan) with a KF-2002 (for polymers, Showa Denko, Tokyo, Japan) or a tandem column system of two columns selected from Shodex KF-2001 and KF-2002 (for monomers, Showa Denko, Tokyo, Japan) using chloroform as an eluent. Matrix assisted laser desorption ionization time-of-flight mass spectrometry (MALDI-TOF-MS) was recorded on a Bruker Autoflex II instrument (Bruker Daltonics, Billerica, MA, USA): trans-2-[3-(4-tert-butylphenyl)-2-methyl-2-propenylidene]malononitrile (DCTB) matrix (20 mg/mL in CHCl<sub>3</sub>) and sodium trifluoroacetate cationizing agents (1 mg/mL in THF). TGA and DSC measurements were performed by Shimadzu DTG-60 and DSC-60 Plus (Shimadzu, Kyoto, Japan), respectively, under nitrogen atmosphere at a heating rate of 10 °C/min. Transmittance spectra were recorded on a JASCO spectrophotometer V-670 KNN (JASCO, Tokyo, Japan). Refractive index (RI) values were measured by Abbe's method with an Atago refractometer (Atago, Tokyo, Japan).

### 3. Synthetic procedure and characterization data

**Bis(dimethylvinylsilyloxy) heptaphenyl IC-POSS with heptaphenyl CC-POSS (4**<sub>Ph-Ph</sub>). A Toluene solution (8.4 mL) of **2**<sub>Ph</sub> (1.00 g, 0.84 mmol), **3**<sub>Ph</sub> (0.96 g, 1.00 mmol) and Pt(dvs) (0.1 M in xylene, 0.05mL,  $5.0 \times 10^{-3}$  mmol) was stirred at 50 °C for 6 h under N<sub>2</sub> atmosphere. The solvents were removed in *vacuo*, and the residue was subjected to preparative HPLC to give **4**<sub>Ph-Ph</sub> (0.94 g, 0.44 mmol, 52%). <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz) δ 7.78-7.68 (m, 14H), 7.59-7.56 (m, 2H), 7.48-7.40 (m, 14H), 7.40-7.24 (m, 28H), 7.18-7.08 (m, 10H), 6.96-6.91 (t, *J* = 2.8Hz, 2H), 6.24-5.77 (m, 6H), 0.75 (s, 4H), 0.30-0.28 (m, 12H), 0.24-0.22 (m, 6H) ppm. <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100 MHz) δ 138.7, 134.4, 134.3, 134.2, 134.1, 134.0, 139.9, 132.9, 132.8, 132.6, 131.2, 130.8, 130.6, 130.5, 130.2, 130.1, 128.0, 127.7, 127.6, 9.1, 3.6, 0.5, -0.2 ppm. <sup>29</sup>Si-NMR (CDCl<sub>3</sub>, 80 MHz) δ 12.3, 0.1, -64.6, -77.4, -77.6, -77.8, -78.1, -78.2, -78.4, -78.7 ppm. MALDI-TOF MS (m/z): calcd for C<sub>96</sub>H<sub>98</sub>O<sub>24</sub>Si<sub>18</sub>Na [M+Na]<sup>+</sup>: 2161.2; found: 2161.2.

Bis(dimethylvinylsilyloxy) heptaphenyl IC-POSS with heptaisobutyl CC-POSS (4<sub>Ph-iBu</sub>). The same procedure as that of 4<sub>Ph-Ph</sub> using 3<sub>iBu</sub> instead of 3<sub>Ph</sub>. The isolated yield was 52%. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz) δ 7.59-7.56 (m, 2H), 7.43-7.38 (m, 7H), 7.38-7.24 (m, 14H), 7.17-7.08 (m, 12H), 6.27-5.81 (m, 6H), 1.91-1.80 (m, 7H), 0.98-0.91 (m, 42H), 0.63-0.56 (m, 18H), 0.34-0.32 (m, 12H), 0.26-0.24 (m, 6H) ppm. <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100 MHz) δ 138.6, 134.1, 134.0, 133.9, 132.9, 132.8, 132.5, 132.9, 132.8, 132.5, 131.2, 130.7, 130.6, 130.1, 130.0, 129.9, 127.6, 127.5, 25.8, 25.7, 23.9, 22.6, 22.5, 8.9, 3.7, 0.4, -0.3 ppm. <sup>29</sup>Si-NMR (CDCl<sub>3</sub>, 80 MHz) δ 12.5, -0.2, -66.9, -67.5 -67.6, -67.8, -67.9, -77.3, -77.7, -77.8, -78.0, -78.1 ppm. MALDI-TOF MS (m/z): calcd for  $C_{82}H_{126}O_{24}Si_{18}Na [M+Na]^+$ : 2021.4; found: 2021.4.

Bis(dimethylvinylsilyloxy) heptaisobutyl IC-POSS with heptaphenyl CC-POSS ( $4_{iBu-Ph}$ ). The same procedure as that of  $4_{Ph-Ph}$  using  $2_{iBu}$  instead of  $2_{Ph}$ . The isolated yield was 54%. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.86-7.78 (m, 14H), 7.55-7.45 (m, 7H), 7.45-7.38 (m, 14H), 6.25-5.75 (m, 6H), 1.99-1.82 (m, 7H), 1.08-0.95 (m, 42H), 0.90-

0.75 (m, 4H), 0.67-0.57 (m, 14H), 0.26-0.24 (m, 12H), 0.20-0.18 (m, 6H) ppm. <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  139.1, 134.3, 134.2, 131.9, 130.8, 130.6, 130.4, 130.3, 130.2, 129.7, 127.9, 26.1, 26.0, 25.9, 25.7, 25.0, 24.1, 24.0, 23.9, 23.8, 22.5, 9.1, 3.7, 0.4, -0.3 ppm. <sup>29</sup>Si-NMR (CDCl<sub>3</sub>, 80 MHz)  $\delta$  9.8, -2.6, -64.3, -67.2 -67.6, -67.7, -67.8, -68.0, -77.8, -78.2, -78.3, -78.6 ppm. MALDI-TOF MS (m/z): calcd for C<sub>82</sub>H<sub>126</sub>O<sub>24</sub>Si<sub>18</sub>Na [M+Na]<sup>+</sup>: 2021.4; found: 2021.4.

Synthesis of polymer 6a. A Toluene solution (0.3 mL) of  $4_{Ph-Ph}$  (300 mg, 0.144 mmol), 5a (29 mg, 0.14 mmol) and Pt(dvs) (0.1 M in xylene, 8 × 10<sup>-3</sup> mL, 8.0 × 10<sup>-4</sup> mmol) was stirred at 50 °C for 2 h under N<sub>2</sub> atmosphere. The solvents were removed in *vacuo*, and the residue was subjected to preparative HPLC to give 5 (157 mg, 48%). <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.89-7.80 (m, 14H), 7.70-7.65 (m, 2H), 7.56-7.46 (m, 14H), 7.46-7.32 (m, 28H) , 7.27-7.16 (m, 10H) ,7.05-6.97 (m, 2H), 0.85 (s, 4H), 0.64-0.47 (m, 8H), 0.42-0.34 (m, 18H) , 0.24-0.02 (m, 18H) ppm. <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  134.3, 134.2, 134.1, 134.0, 133.9, 133.1, 132.8, 130.9, 130.6, 130.4, 130.2, 130.0, 130.0, 127.7, 127.6, 9.5, 9.1, 3.6, 1.5, -0.2, -0.5 ppm. <sup>29</sup>Si-NMR (CDCl<sub>3</sub>, 80 MHz)  $\delta$  12.7, 12.3, 8.3, -21.1, -64.5, -77.3, -77.6, -77.7, -78.0, -78.2, -78.5 ppm.

**Synthesis of polymer 6b.** The same procedure as that of **6a** using **5b** instead of **5a**. The isolated yield was 62%. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.95-7.82 (br, 14H), 7.74-7.68 (br, 2H), 7.60-7.53 (br, 14H), 7.53-7.34 (br, 28H), 7.29-7.18 (br, 10H), 7.09-7.01 (br, 2H), 0.92-0.84 (br, 4H), 0.65-0.47 (br, 8H), 0.47-0.35 (br, 18H), 0.20-0.01 (br, 12H) ppm. <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  134.4, 134.3, 134.2, 134.1, 134.0, 133.1, 132.8, 131.4, 131.3, 130.9, 130.8, 130.6, 130.5, 130.4, 130.2, 130.0 128.0, 127.7, 9.6, 9.5, 9.1, 3.6, -0.2, -0.3 ppm. <sup>29</sup>Si-NMR (CDCl<sub>3</sub>, 80 MHz)  $\delta$  12.7, 12.4, 8.4, -64.4, -77.2, -77.6, -77.7, -77.9, -78.2, -78.2, -78.5 ppm.

**Synthesis of polymer 6c.** The same procedure as that of **6a** using **5c** instead of **5a**. The isolated yield was 58%. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.94-7.85 (m, 14H), 7.74-7.70 (m, 2H), 7.60-7.53 (m, 14H), 7.53-7.36 (m, 32H), 7.30-7.19 (m, 10H), 7.12-7.05 (m, 2H), 0.92 (s, 4H), 0.83-0.64 (m, 8H), 0.47-0.37 (m, 18H), 0.34-0.26 (m, 12H) ppm. <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  134.4, 134.3, 134.2, 134.0, 133.1, 133.0, 132.9, 131.4, 131.3, 130.9, 130.8, 130.6, 130.5, 130.3, 130.2, 130.1 128.0, 127.7, 10.3, 9.1, 7.0, 3.7, -0.1, -3.5 ppm. <sup>29</sup>Si-NMR (CDCl<sub>3</sub>, 80 MHz)  $\delta$  12.6, 12.4, -1.5, -64.4, -77.2, -77.5, -77.7, -77.9, -78.1, -78.5 ppm.

**Synthesis of polymer 7.** The same procedure as that of **6a** using  $4_{Ph-iBu}$  instead of  $4_{Ph-Ph}$ , and reaction time was changed from 2 h to 4 h. The isolated yield was 31 %. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.64-7.56 (m, 2H), 7.48-7.38 (m, 7H), 7.38-7.24 (m, 14H), 7.22-7.06 (m, 12H), 1.95-1.83 (m, 7H), 1.02-0.95 (m, 42H), 0.66-0.59 (m, 18H), 0.58-0.40 (m, 8H), 0.34-0.26 (m, 18H), 0.13-0.04 (m, 18H) ppm. <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  134.0, 133.9, 133.0, 131.2, 130.1, 129.9, 127.9, 127.5, 25.8, 25.7, 23.9, 22.6, 22.5, 9.4, 8.9, 3.7, 1.2, 1.1, 1.0, -0.3, -0.6 ppm. <sup>29</sup>Si-NMR (CDCl<sub>3</sub>, 80 MHz)  $\delta$  12.6, 12.4, 8.1, -21.2, -67.0. -67.6, -67.8, -67.9, -77.4, -77.8, -78.1 ppm.

Synthesis of polymer 8. The same procedure as that of 6a using  $4_{iBu-Ph}$  instead of  $4_{Ph-Ph}$ , and reaction time was changed from 2 h to 4 h. The isolated yield was 31 %. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.84-7.74 (m, 14H), 7.52-7.44 (m, 7H), 7.44-7.34 (m, 14H), 1.96-1.78 (m, 7H), 1.06-0.90 (m, 42H), 0.88-0.72 (m, 4H), 0.64-0.54 (m, 14H), 0.52-0.44 (m, 8H), 0.23-0.13 (m, 18H), 0.13-0.04 (m, 18H) ppm. <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  134.3, 134.2, 130.7, 130.6, 130.4, 127.9, 26.1, 26.0, 25.9, 25.8, 25.7, 25.1, 25.0, 24.1, 24.0, 23.9, 23.8, 22.5, 9.5, 9.1, 3.6, 1.4, 1.3, 1.1, 0.4, 0.2, -0.4, -0.5 ppm. <sup>29</sup>Si-NMR (CDCl<sub>3</sub>, 80 MHz)  $\delta$  9.9, 9.7, 8.2, -2.6, -21.3, -64.4, -67.3, -67.6, -67.7, -67.8, -67.9, -68.1, -78.3, -78.6 ppm.

# 4. NMR spectra



Figure S1. <sup>1</sup>H NMR spectrum (400 MHz) of 4<sub>Ph-Ph</sub> in CDCl<sub>3</sub>.



Figure S2. <sup>13</sup>C NMR spectrum (100 MHz) of  $4_{Ph-Ph}$  in CDCl<sub>3</sub>.



Figure S3. <sup>29</sup>Si NMR spectrum (80 MHz) of 4<sub>Ph-Ph</sub> in CDCl<sub>3</sub>.



Figure S4. <sup>1</sup>H NMR spectrum (400 MHz) of 4<sub>Ph-*i*Bu</sub> in CDCl<sub>3</sub>.



Figure S5. <sup>13</sup>C NMR spectrum (100 MHz) of 4<sub>Ph-*i*Bu</sub> in CDCl<sub>3</sub>.



Figure S6. <sup>29</sup>Si NMR spectrum (80 MHz) of 4<sub>Ph-*i*Bu</sub> in CDCl<sub>3</sub>.



Figure S7. <sup>1</sup>H NMR spectrum (400 MHz) of 4<sub>*i*Bu-Ph</sub> in CDCl<sub>3</sub>.



Figure S8. <sup>13</sup>C NMR spectrum (100 MHz) of 4<sub>*i*Bu-Ph</sub> in CDCl<sub>3</sub>.



Figure S9. <sup>29</sup>Si NMR spectrum (80 MHz) of 4<sub>*i*Bu-Ph</sub> in CDCl<sub>3</sub>.



Figure S10. <sup>1</sup>H NMR spectrum (400 MHz) of 6a in CDCl<sub>3</sub>.



Figure S11. <sup>13</sup>C NMR spectrum (100 MHz) of 6a in CDCl<sub>3</sub>.



Figure S12. <sup>29</sup>Si NMR spectrum (80 MHz) of 6a in CDCl<sub>3</sub>.



Figure S13. <sup>1</sup>H NMR spectrum (400 MHz) of 6b in CDCl<sub>3</sub>.



Figure S14. <sup>13</sup>C NMR spectrum (100 MHz) of 6b in CDCl<sub>3</sub>.



Figure S15. <sup>29</sup>Si NMR spectrum (80 MHz) of 6b in CDCl<sub>3</sub>.



Figure S16. <sup>1</sup>H NMR spectrum (400 MHz) of 6c in CDCl<sub>3</sub>.



Figure S17. <sup>13</sup>C NMR spectrum (100 MHz) of 6c in CDCl<sub>3</sub>.



Figure S18. <sup>29</sup>Si NMR spectrum (80 MHz) of 6c in CDCl<sub>3</sub>.



Figure S19. <sup>1</sup>H NMR spectrum (400 MHz) of 6d in CDCl<sub>3</sub>.



Figure S20. <sup>13</sup>C NMR spectrum (100 MHz) of 6d in CDCl<sub>3</sub>.



Figure S21. <sup>29</sup>Si NMR spectrum (80 MHz) of 6d in CDCl<sub>3</sub>.



Figure S22. <sup>1</sup>H NMR spectrum (400 MHz) of 6e in CDCl<sub>3</sub>.



Figure S23. <sup>13</sup>C NMR spectrum (100 MHz) of 6e in CDCl<sub>3</sub>.



Figure S24. <sup>29</sup>Si NMR spectrum (80 MHz) of 6e in CDCl<sub>3</sub>.



Figure S25. <sup>29</sup>Si NMR spectra (80 MHz in CDCl<sub>3</sub>) and assignments of  $4_{Ph-iBu}$  and  $4_{iBu-Ph}$ .

# 5. MALDI-TOF-MASS spectra



Figure S26. MALDI TOF MS spectrum of  $4_{Ph-iBu}$ . Matrix: DCTB (20 mg/mL in CHCl<sub>3</sub>), cationizing agents: TFANa (1 mg/mL in THF). (a) Full spectrum and (b) expanded view.



Figure S27. MALDI TOF MS spectrum of  $4_{iBu-Ph}$ . Matrix: DCTB (20 mg/mL in CHCl<sub>3</sub>), cationizing agents: TFANa (1 mg/mL in THF).

### 6. X-ray diffraction patterns



Figure S28. Powder X-ray diffraction patterns of 6a, 7, and 8.

### 7. Detail of cyclic compound

In the course of polymerization, low molecular weight by-products were observed in the SEC charts though the conversions of the monomers were approximately 100%. Thus, the by-product was isolated by preparative HPLC in the case of polymer **6a**. The <sup>1</sup>H-NMR spectrum of the isolated by-product showed no signals due to vinyl (**4**<sub>Ph-Ph</sub>, 6.19-5.80 ppm) and Si-H (**5a**, 4.71 ppm) groups (Figure S29). In the <sup>29</sup>Si-NMR spectrum, the signals due to dimethylvinylsilyl (**4a**, -0.1 ppm) and dimethylsilyl (**5a**, -6.6 ppm) groups also disappeared (Figure S30). The MALDI-TOF MS spectra showed a peak at 2369.3 Da (C<sub>102</sub>H<sub>118</sub>O<sub>26</sub>Si<sub>21</sub>Na: [M+Na]<sup>+</sup> calcd. 2369.3) (Figure S31). These results indicate that the isolated by-product is cyclic compound as shown in Chart S1.



Chart S1. Chemical structure of the cyclic by-product.



Figure S29. <sup>1</sup>H-NMR spectra (400 MHz in CDCl<sub>3</sub>) of  $4_{Ph-Ph}$ , 5a, and the cyclic by-product.



Figure S30. <sup>29</sup>Si-NMR spectra (80 MHz in CDCl<sub>3</sub>) of  $4_{Ph-Ph}$ , 5a, and the cyclic by-product.



**Figure S31.** MALDI TOF MS spectrum of the cyclic by-product. Matrix: DCTB (20 mg/mL in CHCl<sub>3</sub>), cationizing agents: TFANa (1 mg/mL in THF).

### 8. Study on M<sub>n</sub> values

For the evaluation of  $M_n$  by <sup>1</sup>H-NMR spectroscopy, vinyl groups were introduced to the end groups. That is, polymerization of 1.5 equivalent **4**<sub>Ph-Ph</sub> with **5a** was carried out under the optimized condition (Run 4 in Figure 2). The low molecular weight molecules were removed by preparative HPLC (eluent: chloroform), and the obtained polymer products were subjected to <sup>1</sup>H-NMR and SEC measurements. The <sup>1</sup>H-NMR spectrum indicated that the  $M_n$  was approximately 21000 (n  $\approx$  8 in Figure S32), while the  $M_n$ estimated by the SEC was 7200. This result means that the molecular weight of the polymer was much underestimated using SEC.



Figure S32. <sup>1</sup>H-NMR (400 MHz in CDCl<sub>3</sub>) of the end-labeled polymer.