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# **Electronic Supplementary Information (ESI)**

## for

# Multicomponent Spiropolymerization of Diisocyanides, Alkynes and Carbon Dioxide for Constructing 1,6-Dioxospiro[4,4]nonane-3,8-diene as Structural Units under One-Pot Catalyst-Free Conditions

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## 1 Experimental

#### 1.1 Materials

Unless stated otherwise, all chemicals were obtained from commercial suppliers and used without further purification. The monomers **1a - d** were synthesized according to the commonly used synthetic routes as shown in Scheme S1. Benzyltriethylammonium chloride and trans-1,4-diaminocyclohexane, 4, 4' - methylenebis(cyclohexylamine) were bought from Heowns. 1,4-bis(aminomethyl) benzene, 4,4'-methylenebis(2,6-diethylaniline), dimethyl acetylenedicarboxylate and diethyl acetylenedicarboxylate were bought from Energy Chemical. Chloroform-d was bought from Innochem.

# 1.2 Equipments

Weight-average molecular weights ( $M_w$ ) and polydispersity indices ( $M_w/M_n$ ) of the polymers were obtained from gel permeation chromatography (GPC) system equipped with a Waters 1515 isocratic HPLC pump and Waters 2414 refractive index detector. Polystyrene standard was utilized and THF was used as the eluent at a flow rate of 1.0 mL min<sup>-1</sup>. Fourier transform infrared (FT-IR) spectra were measured on a Bruker (ALPHA) spectrometer. <sup>1</sup>H NMR spectra were measured on a Bruker AV 400 spectrometer. Mass spectra were collected by using a Finnigan BIFLEX III mass spectrometer. Thermogravimetric analysis (TGA) was carried out on a PerkinElmer STA 8000 at a heating rate of 10 °C min<sup>-1</sup> under a nitrogen flow. Differential scanning calorimeter (DSC) was measured on a TA DSC Q2000. UV-Vis spectra were recorded on a TU-1901 double beam UV-Vis spectrophotometer.

## 1.3 Synthesis of monomers 1a ~ d

Scheme S1. The synthetic route to monomers 1a - d

Monomers 1a - d were prepared by the same steps. The synthesis procedure of 1b - d is almost the same with that of 1a. The detailed method for the synthesis of 1a was given here as an example.

Trans-1,4-diisocyanocyclohexane (1a): To a 500 mL round-bottom flask were added trans-1,4-diaminocyclohexane (8.1200 g, 50 mmol), DCM (20 mL), chloroform (8.044 mL, 100 mmol) and TEBAC (0.2278 g, 1 mmol). A 50% aqueous solution of sodium hydroxide (15 mL) was added. The mixture was heated at 40 °C for 12 hours. The organic layer was combined and washed with DCM and water, and then dried over MgSO<sub>4</sub> for an hour. Product 1a was separated by flash column chromatograph.

#### 1.4 Characterization data of monomers 1a - d

Characterization data of trans-1,4-diisocyanocyclohexane **1a**: White powder of **1a** was obtained in 30.4% yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 3.80 (m, 2H), 2.02-1.99 (d, J = 10.4 Hz, 4H), 1.76-1.74 (m, J = 6.8 Hz, 4H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  = 156.21-156.11 (t, J = 4.9 Hz), 49.66-49.55 (t, J = 5.6 Hz), 26.86. HRMS (EI, m/z) Calcd for [M+H]<sup>+</sup> C<sub>8</sub>H<sub>10</sub>N<sub>2</sub>: 134.0844, Found: 134.0843, error: -0.7 ppm.

Characterization data of 4,4'-methylenebis(cyclohexylisocyano) **1b**: White powder of **1b** was obtained in 34.5% yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 3.90 (m, 1H), 3.37-3.33 (m, 2H), 2.17-2.14 (d, J = 12.0 Hz, 4H), 1.96-1.92 (d, J = 12.8 Hz, 2H), 1.76-1.73 (d, J = 10.8 Hz, 5H), 1.61-1.50 (m, 9H), 1.36-1.33 (m, 6H), 1.13 (s, 1H), 1.07-1.03 (t, J = 7.2 Hz, 1H), 0.92-0.84 (m, 4H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  = 154.94-154.78 (q, J = 10.4 Hz), 153.86-153.70 (q, J = 10.8 Hz), 52.31-52.14 (q, J = 10.8 Hz), 50.66-50.51 (q, J = 10.0 Hz), 43.58, 43.42, 43.17, 33.34-33.26 (t, J = 4.3 Hz), 33.04, 32.94, 32.73, 30.94, 30.61, 30.59, 27.00. HRMS (ESI, m/z) Calcd for [M+Na]<sup>+</sup> C<sub>15</sub>H<sub>22</sub>N<sub>2</sub>Na: 230.1681, Found: 230.1671, error: -3.9 ppm.

Characterization data of 1,4-bis(isocyanomethyl)benzene **1c**: White powder of **1c** was obtained in 42.0% yield.  $^{1}$ H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 7.39 (s, 4H), 4.66 (m, 4H);  $^{13}$ C NMR (100 MHz, CDCl<sub>3</sub>): 158.26-158.16 (t, J = 5.1 Hz), 132.75, 121.31, 45.27-45.12 (t, J = 7.3 Hz). HRMS (EI, m/z) Calcd for [M+H]<sup>+</sup> C<sub>10</sub>H<sub>8</sub>N<sub>2</sub>: 156.0687, Found: 156.0684, error: -1.9 ppm.

Characterization data of 1,1-bis(3-ethyl-4-isocyano-5-ethylphenyl)methane **1d**: White powder of **1d** was obtained in 44.7% yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 6.91 (s, 4H), 3.92 (s, 2H), 2.78-2.72 (q, J = 7.6 Hz, 8H), 1.26-1.22 (t, J = 7.6 Hz, 12H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  = 167.92, 141.51, 141.06, 126.82, 41.53, 25.87, 13.97. HRMS (EI, m/z) Calcd for [M+H]<sup>+</sup> C<sub>23</sub>H<sub>26</sub>N<sub>2</sub>: 330.2096, Found: 330.2096, error: 0 ppm.

# 1.5 Synthetic route to MC

The synthetic route has shown in Scheme 2. To a 30 mL Schlenk tube equipped with a magnetic stir bar were added dimethyl acetylenedicarboxylate **2a** (0.1705 g,

1.2 mmol) and 6 mL toluene. Then, cyclohexyl isocyanide (0.1091 g, 1.0 mmol) was slowly added. The reaction mixture was heated in an oil bath at 80 °C for 12 h under constant stirring under CO<sub>2</sub> atmosphere, cooled to room temperature. The organic layer was combined and washed with DCM and water, and then dried over MgSO<sub>4</sub> for an hour. **MC** was separated by flash column chromatograph.

## 1.6 Characterization data of MC

White powder of **MC** was obtained in 54.0% yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  = 3.96 (s, 6H), 3.78 (s, 6H), 3.63 (m, 2H), 1.74-1.71 (m, 4H), 1.62-1.57 (m, 4H), 1.43-1.16 (m, 12H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  = 161.44, 159.68, 150.63, 139.74, 135.87, 111.33, 57.26, 53.36, 53.33, 52.98, 52.95, 33.30, 32.95, 25.59, 24.59, 24.49. HRMS (EI, m/z) Calcd for [M+H]<sup>+</sup> C<sub>27</sub>H<sub>34</sub>N<sub>2</sub>O<sub>10</sub>: 546.2213, Found: 546.2222, error: +1.6 ppm.

# 2 FT-IR spectra of spiropolymers

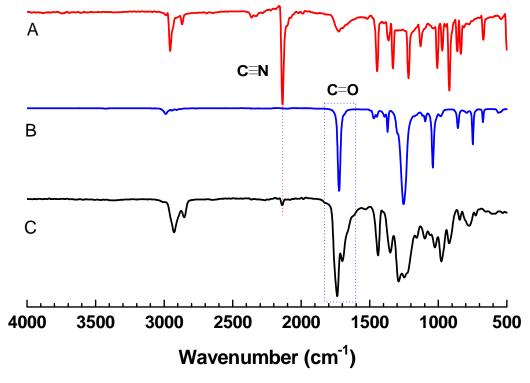


Figure S1. FT-IR spectra of (A) 1a, (B) 2b, and (C) P1a2b.

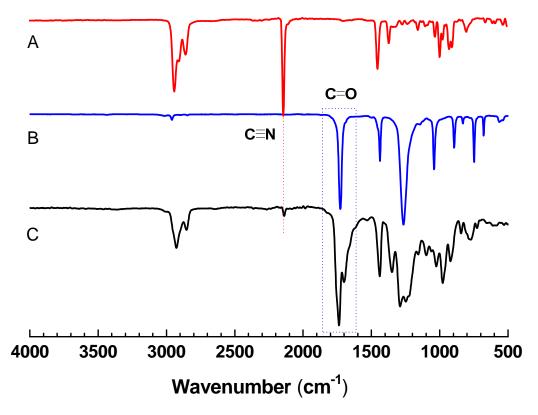


Figure S2. FT-IR spectra of (A) 1b, (B) 2a, and (C) P1b2a.

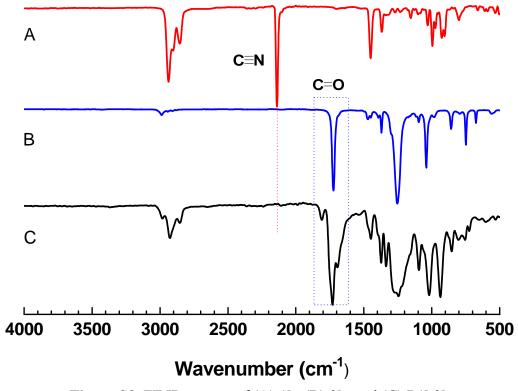


Figure S3. FT-IR spectra of (A) 1b, (B) 2b, and (C) P1b2b.

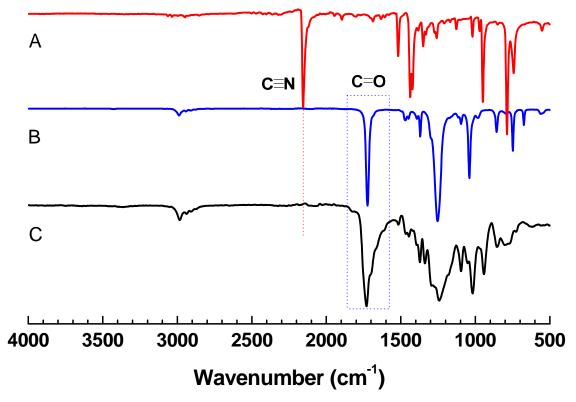


Figure S4. FT-IR spectra of (A) 1c, (B) 2b, and (C) P1c2b.

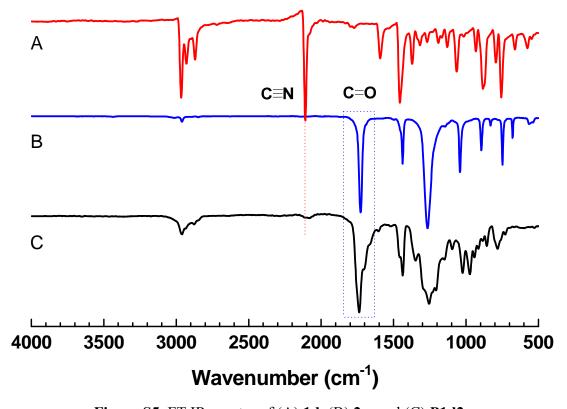


Figure S5. FT-IR spectra of (A) 1d, (B) 2a, and (C) P1d2a.

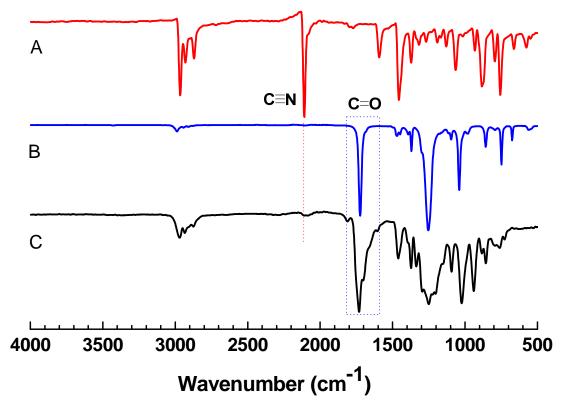


Figure S6. FT-IR spectra of (A) 1d, (B) 2b, and (C) P1d2b.

# 3 <sup>1</sup>H NMR spectra of spiropolymers

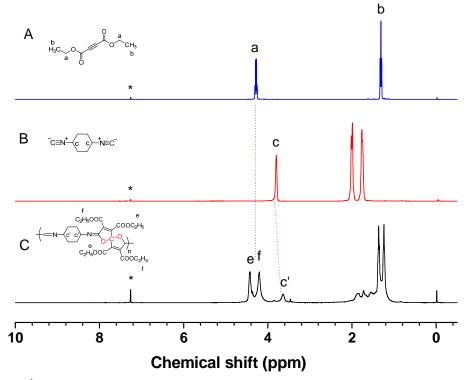
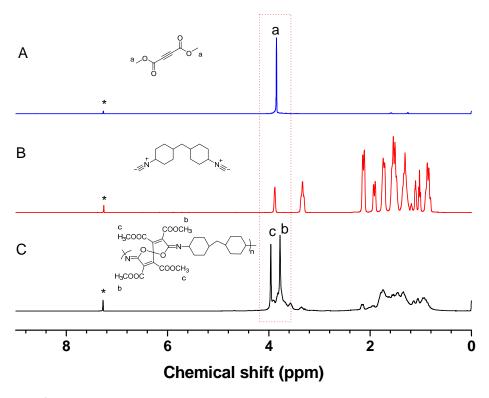
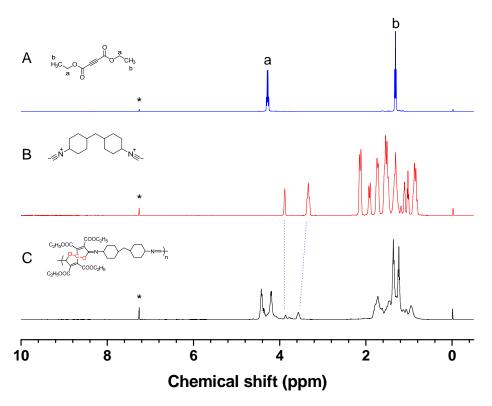


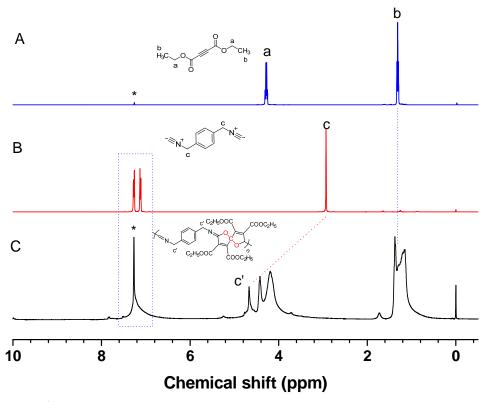
Figure S7. <sup>1</sup>H NMR spectra of (A) 1a, (B) 2b, and (C) P1a2b in CDCl<sub>3</sub>. The solvent peaks are marked with asterisks.



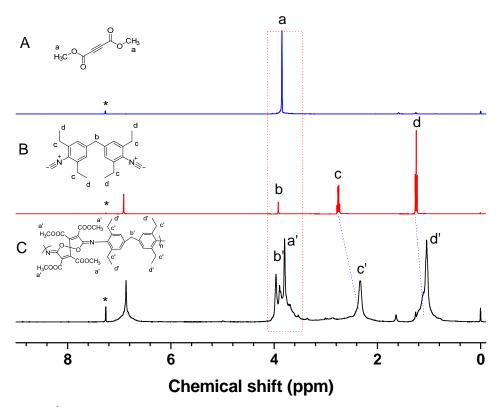
**Figure S8.** <sup>1</sup>H NMR spectra of (A) **1b**, (B) **2a**, and (C) **P1b2a** in CDCl<sub>3</sub>. The solvent peaks are marked with asterisks.



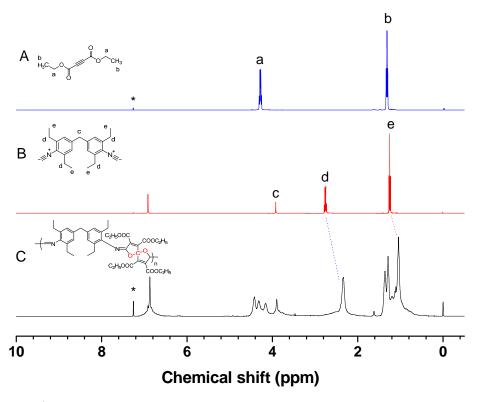
**Figure S9.** <sup>1</sup>H NMR spectra of (A) **1b**, (B) **2b**, and (C) **P1b2b** in CDCl<sub>3</sub>. The solvent peaks are marked with asterisks.



**Figure S10.** <sup>1</sup>H NMR spectra of (A) **1c**, (B) **2b**, and (C) **P1c2b** in CDCl<sub>3</sub>. The solvent peaks are marked with asterisks.

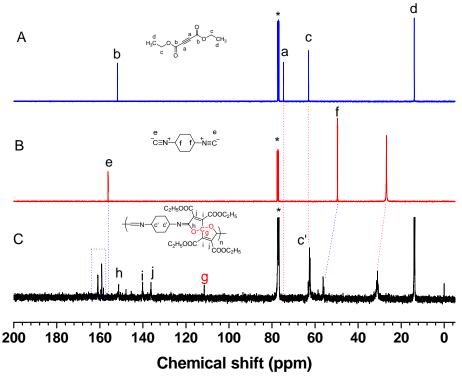


**Figure S11.** <sup>1</sup>H NMR spectra of (A) **1d**, (B) **2a**, and (C) **P1d2a** in CDCl<sub>3</sub>. The solvent peaks are marked with asterisks.

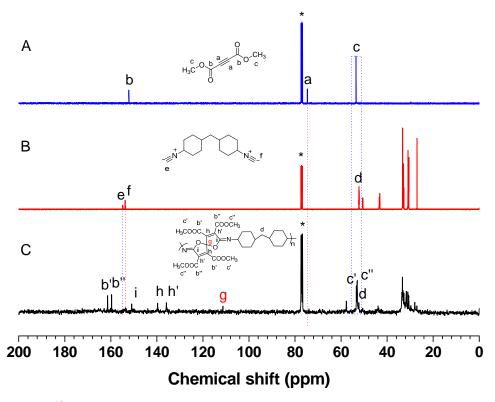


**Figure S12.** <sup>1</sup>H NMR spectra of (A) **1d**, (B) **2b**, and (C) **P1d2b** in CDCl<sub>3</sub>. The solvent peaks are marked with asterisks.

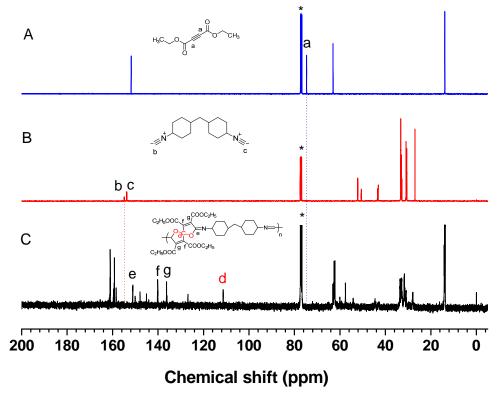
# 4 <sup>13</sup>C NMR spectra of spiropolymers



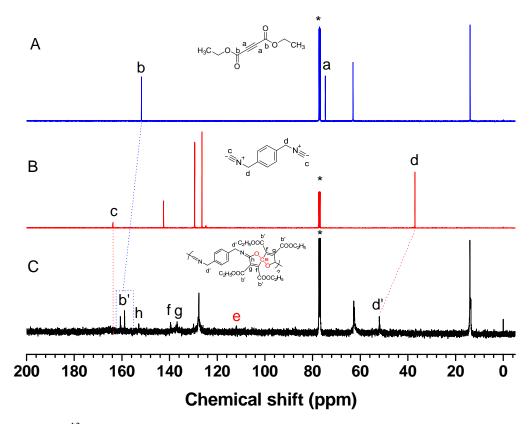
**Figure S13.** <sup>13</sup>C NMR spectra of (A) **1a**, (B) **2b**, and (C) **P1a2b** in CDCl<sub>3</sub>. The solvent peaks are marked with asterisks.



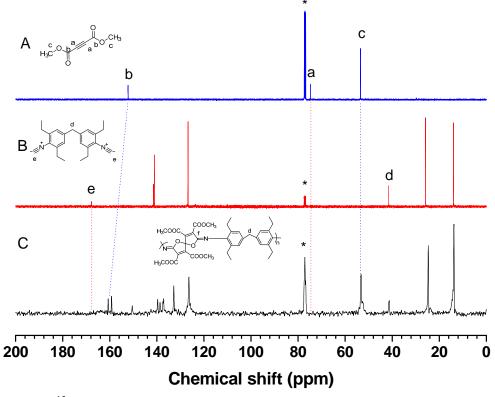
**Figure S14.** <sup>13</sup>C NMR spectra of (A) **1b**, (B) **2a**, and (C) **P1b2a** in CDCl<sub>3</sub>. The solvent peaks are marked with asterisks.



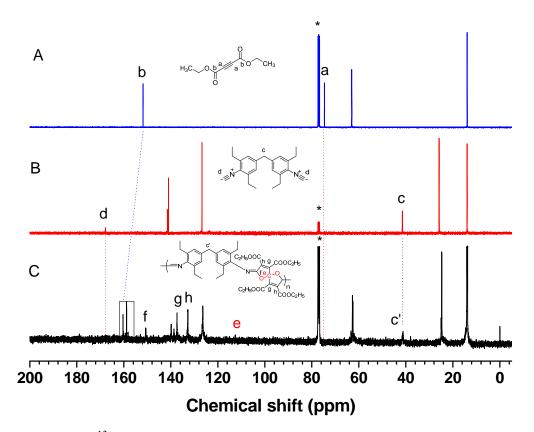
**Figure S15.** <sup>13</sup>C NMR spectra of (A) **1b**, (B) **2b**, and (C) **P1b2b** in CDCl<sub>3</sub>. The solvent peaks are marked with asterisks.



**Figure S16.** <sup>13</sup>C NMR spectra of (A) **1c**, (B) **2b**, and (C) **P1c2b** in CDCl<sub>3</sub>. The solvent peaks are marked with asterisks.



**Figure S17.** <sup>13</sup>C NMR spectra of (A) **1d**, (B) **2a**, and (C) **P1d2a** in CDCl<sub>3</sub>. The solvent peaks are marked with asterisks.



**Figure S18.** <sup>13</sup>C NMR spectra of (A) **1d**, (B) **2b**, and (C) **P1d2b** in CDCl<sub>3</sub>. The solvent peaks are marked with asterisks.

# 5 The DSC results of spiropolymers

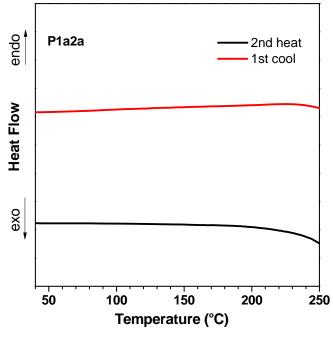


Figure S19. DSC curve of P1a2a. The measurement was performed under  $N_2$  at the heating rate of 10 °C/min.

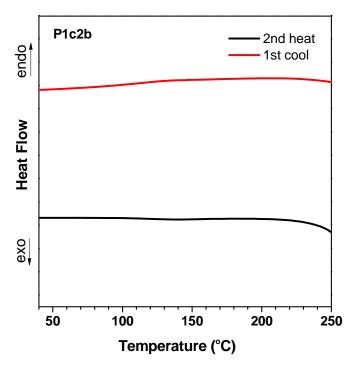


Figure S20. DSC curve of P1c2b. The measurement was performed under  $N_2$  at the heating rate of 10 °C/min.

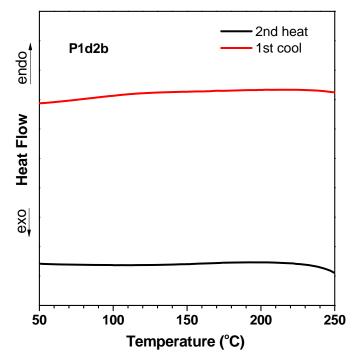


Figure S21. DSC curve of P1d2b. The measurement was performed under  $N_2$  at the heating rate of 10 °C/min.

# 6 Normalized UV-vis absorption spectra of spiropolymers

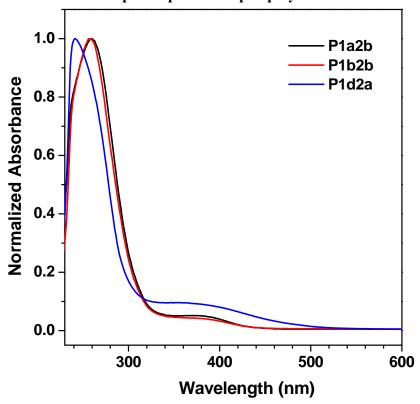


Figure S22. Normalized UV-vis absorption spectra of P1a2b, P1b2b, and P1d2a in THF (10  $\mu$ mol/L of the repeating structure unit as a reference).