SUPPORTING INFORMATION

Highly active Mg(II) and Zn(II) complexes for the ring

opening polymerisation of lactide

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General experimental methods

The preparation and characterisation of all metal complexes was carried out under inert argon atmosphere using standard Schlenk or glovebox techniques. All chemicals used were purchased from Aldrich and used as received except for rac-LA and L-LA which were recrystallised from dry toluene. ¹H and ¹³C{¹H} NMR spectra were recorded on a Bruker 400 MHz instrument and referenced to residual solvent resonances. CDCl₃ was dried over CaH₂ prior to use with metal complexes. C₆D₆ was degassed and stored over molecular sieves. Coupling constants are given in Hertz. Diffusional ordered spectroscopy (DOSY) NMR analysis was carried out on a Bruker 500 MHz instrument; the standard Bruker pulse sequence $ledgp2s^1$ was used, with d₁ of 5 seconds, 64k data points and 8 scans per gradient level. Typically the gradient pulse was $1750 \,\mu\text{s}$, with a diffusion time of 0.05 s. Ten gradient strengths were used between 10 and 90 %. Data were processed using DOSY methods.² All ligands were characterised by electron-spray ionisation-mass spectrometry (ESI-MS) in positive mode. CHN microanalysis was performed by Mr. Stephen Boyer of London Metropolitan University. In certain cases, satisfactory elemental analysis results were not achieved, with samples being consistently low in carbon. This is despite reasonable characterisation by other techniques $({}^{1}H/{}^{13}C{}^{1}H{}$ NMR spectroscopy and X-ray crystallography).

All crystallographic data was collected on a SuperNova or Excalibur, EOS detector diffractometer using radiation CuK α ($\lambda = 1.54184$ Å) or Mo-K α ($\lambda = 0.71073$ Å) radiation all recorded at 150(2) K. All structures were solved by direct methods and refined on all F^2 data using the SHELXL-2014 suite of programs. All hydrogen atoms were included in idealised positions and refined using the riding model, all refinement details are given in the .cif file. All models were straightforward with the following exceptions: $Zn(1)_2$ the methyl groups of one tBu (C29) were disordered over two positions in a 70:30 ratio; $Zn(2)_2$ refined as a 2-component inversion twin in the ratio 93:7. Two independent complex molecules are connected via hydrogen bonding. Disorder, of one ligand backbone, in the ratio 70:30 in one complex molecule is observed and the minor part has been refined with ADP restraints. All NH hydrogen atoms, with exception of the one involved in the disorder, have been refined freely with bond length restraints. One solvent toluene in the asymmetric unit which is disordered over two sites in the ratio 70:30. This has been refined with geometric constraints and ADP restraints; $Mg(1)_2$ the methyl groups of one tBu (C47) are disordered over two positions in a 60:40 ratio; $Mg(7)_2$ the ligand has disorder in the backbone over two positions in a ca. 70:30 ratio.

Polymerisations were carried out in a Young's ampoule under inert argon conditions. All polymerisations were carried out in the absence of solvent. Initial polymerisations were performed with *rac*-lactide (1 g, 6.94 mmol) at 130 °C. Immortal polymerisation was carried out with *L*LA (3 g, 20.8 mmol) at both 130 °C and 180 °C. After polymerisation, the flask was exposed to air to quench and the product was dissolved in CH₂Cl₂ which was then removed *in vacuo* and a crude ¹H NMR spectrum recorded. For immortal polymerisation, the polymer was precipitated from methanol to remove LA and initiator. IR kinetic measurements were recorded using a Bruker Matrix-MF FTIR spectrometer equipped with a diamond ATR probe (IN350 T) suitable for Mid-IR in situ reaction monitoring was used. These reactions were performed with *L*-LA (15 g) at 180 °C. The decrease of the C-O-C lactide peak was monitored. Several known LA:PLA ratios were measured to set up a calibration curve to relate peak area to LA concentration. Stereocomplexation was achieved by sequential addition of monomers under a flow of argon. Overhead stirring was used to ensure a homogeneous polymerisation.

All polymer molecular weights were characterised by gel permeation chromatography (GPC). GPC was carried out at 1 ml min⁻¹ at 35 °C with a THF eluent using a PLgel 5 μ m MIXED-D 300 × 7.5 mm column. The system was referenced against 11 narrow molecular weight standards polystyrene standards with detection *via* refractive index response. A correction factor of 0.58 was applied to measured values.³ Epimerisation was assessed *via* ¹H NMR spectroscopy (CDCl₃) analysis of the homonuclear decoupled methine region. MALDI-ToF mass spectra were determined on a Bruker Autoflex speed instrument using DCTB (trans-2-[3-(4-tert-Butylphenyl)-2-methyl-2-propenylidene]malononitrile) as the matrix and ionised using NaTFA. DSC analysis was recorded on a TA Instruments DSC Q20. The sample was held at 40 °C for 1 minute, heated to 250 °C at 10 °C/min held at this temperature for 1 minute, cooled to 40 °C at 10 °C/min held at this temperature for 1 minute, neared to 250 °C at 5 °C/min - the *T_m* values are quoted for the second heating cycle.

Materials characterisation (GPC, ESI-MS MALDI-TOF) facilities were provided through the Chemical Characterisation and Analysis Facility (CCAF) at the University of Bath.

Synthesis and characterisation

Ligand synthesis and characterisation

1-7H: All ligands were prepared in Schlenk tubes and made directly prior to complexation. Substituted aldehyde (2 mmol) was dissolved in methanol (3 ml) and diamine (2 mmol) was added dropwise to afford a yellow or orange solution. To ensure complete conversion, this solution was stirred at room temperature for 16 hours. Solvent was removed *in vacuo* to yield a quantitative yield of product.

1H (3,5-di-*tert*-butyl-2-hydroxybenzaldehyde + N-methylethylenediamine, yellow oil)

¹H NMR (CDCl₃, 400 MHz) δ = 13.65 (s, 1H; OH), 8.41 (s, 1H; ArCHN), 7.40 (d, *J* = 2.3 Hz, 1H; ArH), 7.10 (d, *J* = 2.4 Hz, 1H; ArH), 3.74 (t, *J* = 5.7 Hz, 2H; CH₂), 2.93 (t, *J* = 5.7 Hz, 2H; CH₂). 2.48 (s, 3H; CH₃) 1.46 (s, 9H; (CH₃)₃), 1.32 (s, 9H; (CH₃)₃).¹³C{¹H} NMR (CDCl₃, 100 MHz) δ = 167.1 (ArCHN), 158.0, 140.1, 136.7, 127.0, 125.9, 117.8 (Ar), 59.5, 51.9 (CH₂), 36.3 (*C*(CH₃)₃), 35.0 (CH₃), 34.1 (*C*(CH₃)₃), 31.5, 29.4 (C(*C*H₃)₃).



ESI-MS (MeOH): Calcd m/z [C₁₈H₃₀N₂ONa]⁺ = 313.2250, found m/z = 313.2246.

Figure SI1: ¹H NMR spectra (400 MHz, CDCl₃) of **1**H. Inset: OH resonance.



Figure SI2: ¹³C{¹H} NMR spectra (100 MHz, CDCl₃) of **1**H.

2H (Salicylaldehyde + N-methylethylenediamine, orange oil)

¹H NMR (CDCl₃, 400 MHz) δ = 13.27 (s, 1H; OH), 8.39 (s, 1H; ArCHN), 7.31 (m, 1H; ArH), 7.25 (m, 1H; ArH), 6.96 (br d, *J* = 7.5 Hz, 1H; ArH), 6.88 (br t, *J* = 7.4 Hz, 1H; ArH), 3.74 (br s, 2H; CH₂), 2.92 (br s, 2H; CH₂). 2.47 (br,s, 4H; CH₃/NH).¹³C{¹H} NMR (CDCl₃, 100 MHz) δ = 166.0 (ArCHN), 161.1, 132.3, 131.3, 120.0, 117.0 (Ar), 59.5, 51.8 (CH₂), 36.3 (CH₃). Broadness of spectra due to ligand cyclisation.

ESI-MS (MeOH): Calcd m/z [C₁₀H₁₅N₂O]⁺ = 179.1179, found m/z = 179.1199.



Figure SI4: ¹³C{¹H} NMR spectra (100 MHz, CDCl₃) of **2**H.

3H (3,5-Dichlorosalicylaldehyde + N-methylethylenediamine, orange oil).

¹H NMR (CDCl₃, 400 MHz) δ = 7.35 (br s, 1H; ArH), 7.09 (br s, 1H; ArH), 3.49 (br s, 2H; CH₂), 2.96 (br s, 2H; CH₂). 2.44 (s, 3H; CH₃).¹³C{¹H} NMR (CDCl₃, 100 MHz) δ = 128.5, 122.7 (Ar).

Notes: No OH and imine resonance observable in ¹H NMR spectra. Broadness of spectra due to ligand cyclisation.

ESI-MS (MeOH): Calcd m/z [C₁₀H₁₃Cl₂N₂O]⁺ = 247.0399, found m/z = 247.0382.



Figure SI5: ¹H NMR spectra (400 MHz, CDCl₃) of **3**H.

4H (3-adamantyl-2-hydroxy-5-*tert*-butylbenzylaldehyde⁴ + N-methylethylenediamine, yellow oil).

¹H NMR (CDCl₃, 400 MHz) δ = 13.69 (s, 1H; OH), 8.40 (s, 1H; ArCHN), 7.33 (d, *J* = 2.5 Hz, 1H; ArH), 7.09 (d, *J* = 2.4 Hz, 1H; ArH), 3.73 (t, *J* = 5.8 Hz, 2H; CH₂), 2.92 (t, *J* = 5.8 Hz, 2H; CH₂). 2.48 (s, 3H; CH₃), 2.21 (m, 6H; CH_{2,ad}), 2.10 (m, 3H; CH_{ad}), 1.81 (m, 6H; CH_{2,ad}), 1.32 (s, 9H; (CH₃)₃). ¹³C{¹H} NMR (CDCl₃, 100 MHz) δ = 167.2 (ArCHN), 158.2, 140.2, 136.9,

126.9, 125.8, 117.7 (Ar), 59.4, 51.8 (CH₂), 40.3 (CH_{2,ad}), 37.2 (C_{ad}), 37.1 (CH_{2,ad}), 36.2 (CH₃), 34.1 (*C*(CH₃)₃), 31.4, 29.1 (C(*C*H₃)₃).



ESI-MS (MeOH): Calcd m/z [C₂₄H₃₇N₂O]⁺ = 369.2900, found m/z = 369.2933.

Figure SI6: ¹H NMR spectra (400 MHz, CDCl₃) of 4H. Inset: OH resonance.



¹H NMR (CDCl₃, 400 MHz) δ = 13.77 (s, 1H; OH), 8.39 (s, 1H; ArCHN), 7.38 (d, *J* = 2.5 Hz, 1H; ArH), 7.09 (d, *J* = 2.4Hz, 1H; ArH), 3.71 (td, *J* = 7.0, 1.1 Hz, 2H; CH₂), 2.66 (t, *J* = 7.2 Hz, 2H; CH₂), 2.32 (s, 6H; 2 × CH₃) 1.45 (s, 9H; (CH₃)₃), 1.32 (s, 9H; (CH₃)₃).¹³C{¹H} NMR (CDCl₃, 100 MHz) δ = 166.6 (ArCHN), 158.1, 139.9, 136.7, 126.8, 125.8,117.9 (Ar), 60.1, 57.8 (CH₂), 45.8 (CH₃), 35.0, 34.1 (*C*(CH₃)₃), 31.5, 29.4 (C(*C*H₃)₃).

5H (3,5-di-*tert*-butyl-2-hydroxybenzaldehyde + N,N-dimethylethylenediamine, yellow solid)

ESI-MS (MeOH): Calcd m/z [C₁₉H₃₃N₂O]⁺ = 305.2593, found m/z = 305.2607.



Figure SI8: ¹H NMR spectra (400 MHz, CDCl₃) of **5**H. Inset: OH resonance.



Figure SI9: ${}^{13}C{}^{1}H$ NMR spectra (100 MHz, CDCl₃) of **5**H.

6H (Salicylaldehyde + N,N-dimethylethylenediamine, orange oil).

¹H NMR (CDCl₃, 400 MHz) δ = 13.42 (s, 1H; OH), 8.37 (s, 1H; ArCHN), 7.30 (td, *J* = 7.3, 1.8 Hz, 1H; ArH), 7.25 (dd, *J* = 7.7, 1.6 Hz, 1H; ArH), 6.96 (d, *J* = 8.3 Hz, 1H; ArH), 6.87 (td, *J* = 7.5, 1.1 Hz, 1H; ArH), 3.72 (dt, *J* = 6.8, 1.2 Hz, 2H; CH₂), 2.65 (t, *J* = 6.7 Hz, 2H; CH₂). 2.31 (s, 6H; 2 × CH₃).¹³C{¹H} NMR (CDCl₃, 100 MHz) δ = 165.5 (ArCHN), 161.2, 132.1, 131.2, 118.8, 118.4, 117.0 (Ar), 59.9, 57.8 (CH₂), 45.8 (CH₃). ESI-MS (MeOH): Calcd *m*/*z* [C₁₁H₁₆N₂ONa]⁺ = 215.1155, found *m*/*z* = 215.1145.

 $251 \text{ MS} (110011). Calca m <math>\chi [0, 111, 0, 0, 201, 0, 1] = 210.11100, 100000 \text{ m} \chi = 210.11100.$



Figure SI10: ¹H NMR spectra (400 MHz, CDCl₃) of **6**H. Inset: OH resonance.



Figure SI11: ¹³C{¹H} NMR spectra (100 MHz, CDCl₃) of **6**H.

7H (3,5-Dichlorosalicylaldehyde + N,N-dimethylethylenediamine, orange oil).

¹H NMR (CDCl₃, 400 MHz) δ = 14.55 (s, 1H; OH), 8.25 (s, 1H; ArCHN), 7.40 (d, *J* = 2.5 Hz, 1H; ArH), 7.15 (d, *J* = 2.6 Hz, 1H; ArH), 3.73 (dt, *J* = 6.4, 1.1 Hz, 2H; CH₂), 2.64 (t, *J* = 6.4 Hz, 2H; CH₂). 2.29 (s, 6H; 2 × CH₃).¹³C{¹H} NMR (CDCl₃, 100 MHz) δ = 164.0 (ArCHN), 158.3, 132.4, 129.0, 123.5, 121.8, 119.0 (Ar), 59.4, 55.9 (CH₂), 45.7 (CH₃). ESI-MS (MeOH): Calcd *m*/*z* [C₁₁H₁₅Cl₂N₂O]⁺ = 261.0562, found *m*/*z* = 261.0541.



Figure SI12: ¹H NMR spectra (400 MHz, CDCl₃) of **7**H. Inset: OH resonance.



Figure SI13: ¹³C{¹H} NMR spectra (100 MHz, CDCl₃) of **7**H.

Zn(II) complexes

Synthesis of imino monophenolate zinc complexes, $Zn(1-7)_2$: $Zn(Et)_2$ (1M, 1 ml, 1 mmol) was added to a solution of ligand (2 mmol) in toluene (10 ml). After complete addition, the solution was stirred for 1 hour before solvent removal. The desired complex was purified *via* washing or recrystallisation from hexane.

Zn(*1*)₂: Recrystallised from hexane. Isolated as pale yellow crystals (0.40 g, 0.62 mmol, 62%). ¹H NMR (C₆D₆, 400 MHz) δ = 7.88 (s, 1H; ArCHN), 7.57 (d, *J* = 2.5 Hz, 1H; ArH), 6.96 (d, *J* = 2.5 Hz, 1H; ArH), 3.26 (m, 1H; CH₂), 2.91 (m, 1H; CH₂), 2.59 (m, 1H; CH₂), 2.15 (m, 1H; CH₂), 1.81 (d, *J* = 6.2 Hz, 3H; CH₃), 1.61 (s, 9H; C(CH₃)₃), 1.41 (s, 9H; C(CH₃)₃), 0.93 (m, 1H; NH). ¹³C{¹H} NMR (C₆D₆, 100 MHz) δ = 171.1 (ArCHN), 169.9, 144.8, 132.8, 129.6, 118.5 (Ar), 58.0, 51.7 (CH₂), 36.3 (CH₃), 36.2, 34.4 (*C*(CH₃)₃), 32.4, 30.2 (*C*(*C*H₃)₃). Elemental analysis (C₃₆H₅₈ZnN₄O₂) Calcd in %: C, 67.11; H, 9.07; N, 8.70. Found: C, 66.94; H, 8.91; N, 8.64.

Note: Complexation scaled up to 54 mmol in reduced solvent volume (toluene, 50 ml). Solution was cooled with an ice batch during $Zn(Et)_2$ addition. Solvent was removed and product washed with hexane (2 x 50 ml). Isolated as yellow powder (25.0 g, 39 mmol, 72 %).



Figure SI14: ¹H NMR spectra (400 MHz, C_6D_6) of Zn(1)₂.



Figure SI15: ${}^{13}C{}^{1}H$ NMR spectra (100 MHz, C₆D₆) of Zn(1)₂.



Figure SI16: ¹H NMR spectra (400 MHz, CDCl₃) of Zn(1)₂.



Figure SI17: ${}^{13}C{}^{1}H$ NMR spectra (100 MHz, CDCl₃) of Zn(1)₂.

Zn(2)₂: Recrystallised from hexane. Isolated as pale yellow crystals (0.20 g, 0.47 mmol, 47%). ¹H NMR (CDCl₃, 400 MHz) δ = 8.19 (s, 1H; ArCHN), 7.15 (m, 1H; ArH), 7.02 (dd, *J* = 7.8, 1.5 Hz, 1H; ArH), 6.75 (d, *J* = 8.5 Hz, 1H; ArH), 6.42 (d, *J* = 7.4 Hz, 1H; ArH), 3.69 (br s, 2H; CH₂), 2.83 (br s, 2H; CH₂), 2.24 (d, *J* = 6.4 Hz, 3H; CH₃), 0.93 (hept, *J* = 6.5 Hz, 1H; NH). ¹³C{¹H} NMR (CDCl₃, 100 MHz) δ = 172.0 (Ar), 168.8 (ArCHN), 135.1, 133.5, 123.6, 119.0, 112.2 (Ar), 57.7, 51.2 (CH₂), 36.2 (CH₃).

Elemental analysis (C₂₀H₂₆ZnN₄O₂) Calc %, C 57.22, H 6.24, N 13.35; found %, C 57.31, H 6.33, N 13.16.



Figure SI18: ¹H NMR spectra (400 MHz, CDCl₃) of Zn(2)₂.



Figure SI19: ¹H NMR spectra (400 MHz, 233 K, CDCl₃) of Zn(2)₂.



Figure SI20: ${}^{13}C{}^{1}H$ NMR spectra (100 MHz, CDCl₃) of Zn(2)₂.

 $Zn(3)_2$: Recrystallised from hexane/toluene mixture. Isolated as pale yellow crystals (0.34 g, 0.61 mmol, 61%).

¹H NMR (CDCl₃, 400 MHz) δ = 8.16 (s, 1H; ArCHN), 7.30 (br s, 1H; ArH), 6.95 (br s, 1H; ArH), 3.82 (br s, 2H; CH₂), 2.91 (br s, 2H; CH₂), 2.27 (s, 3H; CH₃), 1.73 (br s, 1H; NH). Note: Due to low solubility, ¹³C{¹H} NMR was not recorded.

Elemental analysis (C₂₀H₂₂Cl₄ZnN₄O₂) : Calc %, C 43.08, H 3.98, N 10.05; found %, C 43.23, H 4.21, N 9.86.



Figure SI21: ¹H NMR spectra (400 MHz, CDCl₃) of Zn(**3**)₂.

Zn(*4*)₂: Recrystallised from mixture of hexane/toluene. Isolated as pale yellow crystals (0.33 g, 0.41 mmol, 41%).

¹H NMR (C₆D₆, 400 MHz) $\delta = 8.01$ (s, 1H; ArCHN), 7.47 (d, J = 2.5 Hz, 1H; ArH), 6.96 (d, J = 2.5 Hz, 1H; ArH), 3.08 (br t, J = 4.6 Hz, 2H; CH₂), 2.76 (m, 1H; CH₂), 2.29 (br s, 6H; CH_{2ad}), 2.20 (m, 1H; CH₂), 2.06 (br s, 3H; CH_{ad}), 1.87 (d, J = 6.2 Hz, 3H; CH₃), 1.84 (br s, 6H; CH_{2ad}), 1.44 (s, 9H; C(CH₃)₃), 0.83 (m, 1H; NH). ¹³C{¹H} NMR (C₆D₆, 100 MHz) $\delta = 171.7$ (Ar), 169.8 (ArCHN), 141.8, 132.6, 129.2, 128.3, 119.0 (Ar), 58.2, 51.7, 40.9, 38.6 (CH₂), 38.4 (CH₃), 36.0 (C_{ad}), 34.4 (*C*(CH₃)₃), 32.3 (*C*(*C*H₃)₃), 30.4 (CH).



Figure SI22: ¹H NMR spectra (400 MHz, C₆D₆) of Zn(4)₂.



Figure SI23: ${}^{13}C{}^{1}H$ NMR spectra (100 MHz, C₆D₆) of Zn(4)₂.

Zn(*5*)₂: Recrystallised from hexane. Isolated as yellow crystals (0.33 g, 0.49 mmol, 49%). ¹H NMR (C₆D₆, 400 MHz) δ = 7.80 (s, 1H; ArCHN), 7.64 (d, *J* = 2.6 Hz, 1H; ArH), 6.91 (d, *J* = 2.5 Hz, 1H; ArH), 3.14 (2, *J* = 6.3 Hz, 1H; CH₂), 2.37 (sept, *J* = 6.4 Hz, 1H; CH₂), 2.11 (sept, *J* = 6.3 Hz, 1H; CH₂), 1.88 (s, 6H; 2 × CH₃), 1.70 (s, 9H; C(CH₃)₃), 1.39 (s, 9H; C(CH₃)₃). ¹³C{¹H} NMR (C₆D₆, 100 MHz) δ = 172.0 (Ar), 170.7 (ArCHN), 142.2, 134.2, 129.9, 129.6, 118.2 (Ar), 56.0, 57.7 (CH₂), 45.9 (CH₃), 36.3, 34.4 (*C*(CH₃)₃), 32.2, 30.5 (C(*C*H₃)₃). Elemental analysis (C₃₈H₆₂ZnN₄O₂) Calcd in %: C, 67.89; H, 9.30; N, 8.33. Found: C, 67.98; H, 9.20; N, 8.33.



Figure SI24: ¹H NMR spectra (400 MHz, C_6D_6) of Zn(5)₂.



Zn(*6*)₂: Washed with hexane. Isolated as pale yellow powder (0.23 g, 0.51 mmol, 51%). ¹H NMR (CDCl₃, 400 MHz) $\delta = 8.19$ (s, 1H; ArCHN), 7.21 (m, 1H; ArH), 7.05 (dd, *J* = 7.8, 1.5 Hz, 1H; ArH), 6.81 (d, *J* = 8.5 Hz, 1H; ArH), 6.48 (t, *J* = 7.3 Hz, 1H; ArH), 3.69 (br t, *J* = 6.2 Hz, 2H; CH₂), 2.54 (m, 2H; CH₂), 2.09 (s, 6H; 2 × CH₃).¹³C{¹H} NMR (CDCl₃, 100 MHz) $\delta = 171.7$ (Ar), 170.8 (ArCHN), 135.2, 134.3, 123.6, 118.5, 113.1 (Ar), 59.4, 57.2 (CH₂), 45.6 (CH₃).

Elemental analysis (C₂₂H₃₀ZnN₄O₂) Calc %, C 59.00, H 6.75, N 12.51; found %, C 58.96, H 6.79, N 12.38.



Figure SI26: ¹H NMR spectra (400 MHz, CDCl₃) of Zn(6)₂.



Figure SI27: ¹³C{¹H} NMR spectra (100 MHz, CDCl₃) of Zn(6)₂.

Zn(7)₂: Washed with hexane. Isolated as pale yellow powder (0.28 g, 0.48 mmol, 48%). ¹H NMR (CDCl₃, 400 MHz) δ = 8.17 (s, 1H; ArCHN), 7.21 (d, *J* = 2.8 Hz, 1H; ArH), 6.98 (d, *J* = 2.8 Hz, 1H; ArH), 3.72 (m, 2H; CH₂), 2.65 (t, *J* = 5.5 Hz, 2H; CH₂), 2.15 (s, 6H; 2 × CH₃). ¹³C{¹H} NMR (CDCl₃, 100 MHz) δ = 169.0 (ArCHN), 164.7, 133.2, 132.2 127.7, 119.3, 115.9 (Ar), 59.2, 56.4 (CH₂), 45.7 (CH₃).

Elemental analysis (C₂₂H₂₆Cl₄ZnN₄O₂) Calc %, C 45.12, H 4.47, N 9.57; found %, C 45.21, H 4.33, N 9.38.



Figure SI28: ¹H NMR spectra (400 MHz, CDCl₃) of Zn(7)₂.



Figure SI29: ${}^{13}C{}^{1}H$ NMR spectra (100 MHz, CDCl₃) of Zn(7)₂.

DOSY NMR spectra



Figure SI30: $Zn(1)_2$ DOSY NMR spectra (CDCl₃, 500 MHz, D = $7.39 \times 10^{-10} \text{ m}^2\text{s}^{-1}$).



Figure SI31: $Zn(2)_2$ DOSY NMR spectra (CDCl₃, 500 MHz, D = $8.49 \times 10^{-10} \text{ m}^2\text{s}^{-1}$).



Figure SI32: $Zn(3)_2$ DOSY NMR spectra (CDCl₃, 500 MHz, D = $8.25 \times 10^{-10} \text{ m}^2\text{s}^{-1}$). Note: Complex precipitated during experiment.





Figure SI34: $Zn(5)_2$ DOSY NMR spectra (CDCl₃, 500 MHz, D = $7.16 \times 10^{-10} \text{ m}^2\text{s}^{-1}$).



Figure SI35: $Zn(6)_2$ DOSY NMR spectra (CDCl₃, 500 MHz, D = $8.36 \times 10^{-10} \text{ m}^2\text{s}^{-1}$).



Figure SI36: $Zn(7)_2$ DOSY NMR spectra (CDCl₃, 500 MHz, D = $8.16 \times 10^{-10} \text{ m}^2\text{s}^{-1}$).

Mg(II) complexes

Synthesis of imino monophenolate magnesium complexes, $Mg(1-3,5-7)_2$: Mg(n-Bu)₂ (1M, 1 ml, 1 mmol) was added to a solution of ligand (2 mmol) in toluene (10 ml). After complete addition, the solution was stirred for 1 hour before solvent removal. The desired complex was purified *via* washing or recrystallisation from hexane.

Mg(*1*)₂: Recrystallised from hexane. Isolated as pale yellow crystals (0.45 g, 0.75 mmol, 75%). ¹H NMR (C₆D₆, 400 MHz) δ = 7.95 (s, 1H; ArCHN), 7.57 (d, *J* = 2.5 Hz, 1H; ArH), 7.05 (d, *J* = 2.5 Hz, 1H; ArH), 3.27 (m, 1H; CH₂), 2.90 (m, 1H; CH₂), 2.57 (m, 1H; CH₂), 2.05 (m, 1H; CH₂), 1.72 (d, *J* = 6.3 Hz, 3H; CH₃), 1.59 (s, 9H; C(CH₃)₃), 1.42 (s, 9H; C(CH₃)₃), 0.91 (m, 1H; NH). ¹³C{¹H} NMR (C₆D₆, 100 MHz) δ = 169.78 (Ar), 169.76 (ArCHN), 141.1, 132.4,129.3, 120.1 (Ar), 57.8, 51.8 (CH₂), 36.2 (CH₃), 36.1, 34.3 (*C*(CH₃)₃), 32.4, 30.5 (C(*C*H₃)₃).



Figure SI37: ¹H NMR spectra (400 MHz, C_6D_6) of Mg(1)₂.



Figure SI38: ${}^{13}C{}^{1}H$ NMR spectra (100 MHz, C₆D₆) of Zn(1)₂.

Mg(2)₂: Recrystallised from hexane. Isolated as pale yellow crystals (0.33 g, 0.87 mmol, 87%). ¹H NMR (CDCl₃, 400 MHz) δ = 8.19 (s, 1H; ArCHN), 7.14 (t, *J* = 8.4 Hz, 1H; ArH), 7.06 (d, *J* = 7.7 Hz, 1H; ArH), 6.70 (d, *J* = 8.5 Hz, 1H; ArH), 6.40 (t, *J* = 7.4 Hz, 1H; ArH), 3.91 (m, 1H; CH₂), 3.45 (m, 1H; CH₂), 3.06 (m, 1H; CH₂), 2.60 (m, 1H; CH₂), 2.22 (d, *J* = 6.4 Hz, 3H; CH₃), 1.70 (m, 1H; NH). ¹³C{¹H} NMR (CDCl₃, 100 MHz) δ = 170.7 (Ar), 168.5 (ArCHN), 134.7, 133.3,123.2, 120.8, 111.6 (Ar), 57.1, 51.4 (CH₂), 36.3 (CH₃).







Figure SI40: ¹³C{¹H} NMR spectra (100 MHz, CDCl₃) of Mg(**2**)₂.

Mg(*3*)₂:Precipitated from toluene. Isolated as pale yellow powder (0.38 g, 0.74 mmol, 74%). ¹H NMR (CDCl₃, 400 MHz) δ = 8.15 (s, 1H; ArCHN), 7.29 (t, *J* = 2.8 Hz, 1H; ArH), 6.98 (t, *J* = 2.8 Hz, 1H; ArH), 3.91 (m, 1H; CH₂), 4.03 (br s, 1H; CH₂), 3.52 (br s, 1H; CH₂), 3.20 (br s, 1H; CH₂), 2.66 (br s, 1H; CH₂), 2.24 (d, *J* = 6.4 Hz, 3H; CH₃), 1.71 (m, 1H; NH). ¹³C{¹H} NMR (CDCl₃, 100 MHz) δ = 166.6 (ArCHN), 163.5 (ArCHN), 132.3, 131.7,127.2, 121.2, 114.6 (Ar), 57.3, 51.4 (CH₂), 36.1 (CH₃).



Figure SI41: ¹H NMR spectra (400 MHz, CDCl₃) of Mg(**3**)₂.



Figure SI42: ${}^{13}C{}^{1}H$ NMR spectra (100 MHz, CDCl₃) of Mg(**3**)₂.

Mg(*5*)₂: Recrystallised from toluene. Isolated as yellow crystals (0.14 g, 0.22 mmol, 22%). ¹H NMR (CDCl₃, 400 MHz) δ = 8.19 (s, 1H; ArCHN), 7.28 (s, 1H; ArH), 6.88 (s, 1H; ArH), 3.71 (m, 1H; CH₂), 3.47 (m, 1H; CH₂), 2.68 (m, 1H; CH₂), 2.34 (m, 1H; CH₂), 2.19 (s, 6H; 2 × CH₃), 1.28 (s, 18H; 2 × C(CH₃)₃).¹³C{¹H} NMR (CDCl₃, 100 MHz) δ = 171.3 (ArCHN), 168.4, 140.6, 132.6, 128.4, 128.3, 119.2 (Ar), 60.0, 56.2 (CH₂), 46.2 (CH₃), 35.2, 33.7 (*C*(CH₃)₃), 31.5, 29.5 (C(*C*H₃)₃).



Figure SI43: ¹H NMR spectra (400 MHz, CDCl₃) of Mg(**5**)₂.



Figure SI44: ${}^{13}C{}^{1}H$ NMR spectra (100 MHz, CDCl₃) of Mg(**5**)₂.

Mg(*6*)₂: Washed with hexane. Isolated as pale yellow crystals (0.13 g, 0.32 mmol, 32%). ¹H NMR (CDCl₃, 400 MHz) δ = 8.09 (s, 1H; ArCHN), 7.13 (m, 1H; ArH), 6.99 (d, *J* = 7.2 Hz, 1H; ArH), 6.74 (br s, 1H; ArH), 6.39 (br t, *J* = 6.9 Hz, 1H; ArH), 3.54 (br s, 2H; CH₂), 2.38 (br s, 2H; CH₂), 2.03 (s, 6H; 2 × CH₃). ¹³C{¹H} NMR (CDCl₃, 100 MHz) δ = 168.4 (ArCHN), 163.2, 132.3, 131.8, 126.9, 121.0, 114.4 (Ar), 59.2, 55.0 (CH₂), 46.8 (CH₃).



Figure SI45: ¹H NMR spectra (400 MHz, CDCl₃) of Mg(6)₂.



Figure SI46: ${}^{13}C{}^{1}H$ NMR spectra (100 MHz, CDCl₃) of Mg(6)₂.

Mg(7)₂: Recrystallised from a mixture of hexane/CH₂Cl₂.Isolated as pale yellow crystals (0.29 g, 0.53 mmol, 53%).

¹H NMR (CDCl₃, 400 MHz) δ = 8.16 (s, 1H; ArCHN), 7.21 (m, 1H; ArH), 6.95 (s, 1H; ArH), 3.68 (br s, 2H; CH₂), 2.82 (br s, 2H; CH₂), 2.16 (s, 6H; 2 × CH₃). ¹³C{¹H} NMR (CDCl₃, 100 MHz) δ = 170.4 (ArCHN), 134.7, 133.5, 129.0, 128.2, 123.2, 121.1 (Ar), 59.8, 56.0 (CH₂), 46.2 (CH₃).

Elemental analysis ($C_{22}H_{26}MgN_4O_2$) Calcd in %: C, 48.52; H, 4.81; N, 10.29. Found: C, 50.81; H, 5.33; N, 9.42 (Toluene present in crystal structure).



Figure SI47: ¹H NMR spectra (400 MHz, CDCl₃) of Mg(**7**)₂.



Figure SI48: ${}^{13}C{}^{1}H$ NMR spectra (100 MHz, CDCl₃) of Mg(7)₂.

Polymer characterisation

Mechanistic studies



Figure SI49: ¹H NMR (400 MHz, C_6D_6) of $Zn(4)_2 + L$ -LA. Polymer is formed after several days with no change in $Zn(4)_2$ resonances.



Figure SI50: ¹H NMR (400 MHz, C_6D_6) of $Zn(5)_2 + L$ -LA. Polymer is formed after several days with no change in $Zn(5)_2$ resonances.



Figure SI51: ¹H NMR (400 MHz, CDCl₃) of 1. Red spectrum: $Zn(1)_2$; 2. Blue spectrum: $Zn(1)_2 + L$ -LA after 1 minute (180°C, 1:5).



Figure SI52: a) Observed ESI-MS peaks and assignment from solvent free polymerisation $\{180^{\circ}C, [Zn(1)_2:[L-LA] 1:5\}$.b) Possible propagation species $Zn(1)\{Lactyl\}$. No BnOH added to the polymerisation.

IR-Kinetic data



Figure SI53: Semi-logarithmic first order plot with respect to lactide consumption for $Zn(1)_2$ (180°C, 10000:1:100) (Table 5, entry 1).



Figure SI54: Change in intensity of IR spectrum due to LA ROP for Zn(1)₂ (Table 5, entry 1).



Figure SI55: DSC trace of PLLA produced by Zn(1)₂ (180°C, 10000:1:100) (Table 5, entry 1).



Figure SI56: PLLA produced by Zn(1)₂ (180°C, 10000:1:100) (Table 5, entry 1).



Figure SI57: Semi-logarithmic first order plot with respect to lactide consumption for $Zn(4)_2$ (180°C, 10000:1:100) (Table 5, entry 2).



Figure SI58: Semi-logarithmic first order plot with respect to lactide consumption for $Zn(5)_2$ (180°C, 10000:1:100) (Table 5, entry 3).



Figure SI59: PLLA produced by Zn(**4**)₂ (left) and Zn(**5**)₂ (right) (180°C, 10000:1:100) (Table 5, entries 2 and 3).



Figure SI60: ${}^{1}H{}^{1}H{}$ NMR spectra of PLLA produced by Zn(1)₂ (180°C, 10000:1:100) (Table 5, entry 1).



Figure SI61: ${}^{1}H{}^{1}H{}$ NMR spectra of PLLA produced by Zn(4)₂ (180°C, 10000:1:100) (Table 5, entry 2).



Figure SI62: ${}^{1}H{}^{1}H{}$ NMR spectra of PLA produced by Zn(**5**)₂ (180°C, 10000:1:100)(Table 5, entry 3).

Stereocomplexation data

Table SI1. Stereocomplexation for polymerisation of L/D-lactide at 180°C with Zn(II) complexes.

| Init. | [LA]:[I]:[BnOH] ^a | Time / min | Conv % ^a | $M_{n/theo}{}^b$ | M_n^c | Т | $T_m / {}^{\circ}\mathrm{C}^{\mathrm{d}}$ |
|--------|------------------------------|------------|---------------------|------------------|---------|------|---|
| Zn(1)2 | (3000):1:10 | 5 | 95 | 42000 | 33400 | 1.62 | 190 |

Conditions: ([L-LA]+[D-LA]):[I]:[BnOH] = (3000):1:10, solvent free $(180^{\circ}C)^{a}$ Determined *via* ¹H NMR spectroscopy. ^b Theoretical molecular weight based on conversion and co-initiator added { $(M_{r,LA} \times 3 \times \%_{conv}) + M_{n,BnOH}$ }. ^c Molecular weight via GPC analysis (in THF). A correction factor of 0.58 has been applied. ^d Determined by DSC.



Figure SI63: DSC trace of stereocomplex produced by Zn(1)₂ (Table SI1).



Figure SI64: ${}^{1}H{}^{1}H{}$ NMR spectra of stereocomplexed PLA produced by Zn(1)₂ (180°C, 3000:1:10) (Table SI1).



Figure SI65: PLLA-PDLA produced by Zn(1)₂ (180°C, 3000:1:10) (Table SI1, entry 1).

Select homonuclear decoupled data



1, entry 6).





Figure SI67: ${}^{1}H{}^{1}H{}$ NMR spectra of PLLA produced by Zn(1)₂ (180°C, 10000:1) (Table 4, entry 4).



Figure SI68: ¹H{¹H} NMR spectra of PLLA produced by Zn(**7**)₂ (180°C, 10000:1:100) (Table 4, entry 10).



Figure SI69: ${}^{1}H{}^{1}H{}$ NMR spectra of PLLA produced by Mg(1)₂ (180°C, 10000:1:100) (Table 4, entry 12).



Figure SI70: ¹H{¹H} NMR spectra of PLLA produced by Mg(**5**)₂ (180°C, 10000:1:100) (Table 4, entry 13).





Figure SI71: MALDI-ToF spectra of PLA produced by $Zn(\mathbf{6})_2$ (130°C, 300:1:1)(Table 1, entry 7).



Figure SI72: MALDI-ToF spectra of PLA produced by Zn(7)₂ (130°C, 300:1:1) (Table 1, entry 8).



Figure SI73: MALDI-ToF spectra of PLA produced by $Zn(1)_2$ (130°C, 10000:1:100)(Table 3, entry 1).



Figure SI74: MALDI-ToF spectra of PLA produced by Zn(4)₂ (130°C, 10000:1:100) (Table 3, entry 2).



Figure SI75: MALDI-ToF spectra of PLA produced by $Zn(5)_2$ (130°C, 10000:1:100) (Table 3, entry 3).



Figure SI76: MALDI-ToF spectra of PLA produced by $Mg(1)_2$ (130°C, 10000:1:100) (Table 3, entry 4).



Figure SI77: MALDI-ToF spectra of PLA produced by $Mg(5)_2$ (130°C, 10000:1:100) (Table 3, entry 5).



Figure SI78: MALDI-ToF spectra of PLA produced by Zn(**5**)₂ (180°C, 10000:1:100) (Table 4, entry 8).



Figure SI79: MALDI-ToF spectra of PLA produced by $Zn(7)_2$ (180°C, 10000:1:100) (Table 4, entry 10).



Figure SI80: MALDI-ToF spectra of PLA produced by Mg(**1**)₂ (180°C, 10000:1:100) (Table 4, entry 12).



Figure SI81: MALDI-ToF spectra of PLA produced by Mg(**5**)₂ (180°C, 10000:1:100) (Table 4, entry 13).

Select GPC spectra



Figure SI82: GPC spectra of PLA produced by Zn(1)₂ (180°C, 10000:1:100) (Table 4, entry 1).



Figure SI83: GPC spectra comparing PLA produced by $Zn(1)_2$ (red) and commercial PLA (black) (NatureWorks, 6202D) (180°C, 10000:1:15) (Table 4, entry 3).



Figure SI84: GPC spectra of PLA produced by Zn(1)₂ (180°C, 10000:1) (Table 4, entry 3).



Figure SI85: GPC spectra comparing PLA produced by $Zn(1)_2$ with (green) and (magenta) without co-initiator (180°C, 10000:1:{100}) (Table 4, entry 1 vs. 4).



Figure SI86: GPC spectra comparing PLA produced by Zn(1)₂ (green) and commercial PLA (black/gray) (NatureWorks, 4043D/2500HP) (180°C, 10000:1) (Table 4, entry 5).



Figure SI87: GPC spectra of PLA produced by $Zn(4)_2$ (180°C, 10000:1:100) (Table 4, entry 6).



Figure SI88: GPC spectra comparing PLA produced by $Zn(4)_2$ (green) with and without (magenta) co-initiator (180°C, 10000:1:{100}) (Table 4, entry 6 vs. 7).



Figure SI89: GPC spectra of PLA produced by Zn(**5**)₂ (180°C, 10000:1:100) (Table 4, entry 8).



Figure SI90: GPC spectra of PLA produced by Zn(7)₂ (180°C, 10000:1:100) (Table 4, entry 10).

Crystallographic data



Figure SI91: Solid-state structure of $Zn(5)_2$. Ellipsoids shown at 30 % probability level and all hydrogens have been omitted for clarity.



Figure SI92: Solid-state structure of $Zn(6)_2$. Ellipsoids shown at 30 % probability level and all hydrogens have been omitted for clarity.



Figure SI93: Solid-state structure of $Mg(2^*)(2)$. Ellipsoids shown at 30 % probability level and all hydrogens have been omitted for clarity.

| Table SI2 | X-ray | crystall | ographic | parameters |
|-----------|-------|----------|----------|------------|
| | | | | |

| Compound reference Chemical formula Formula Mass Crystal system | $\begin{array}{l} Zn(1)_2 \\ C_{36}H_{58}N_4O_2Zn \\ 644.23 \\ Monoclinic \end{array}$ | $\begin{array}{l} Zn({\bf 2})_2 \\ C_{47}H_{60}N_8O_4Zn_2 \\ 931.77 \\ Monoclinic \end{array}$ | $\begin{array}{l} Zn(\textbf{3})_2 \\ C_{47}H_{52}Cl_8N_8O_4Zn_2 \\ 1207.30 \\ Triclinic \end{array}$ | Zn(4) ₄ C ₅₄ H ₈₄ N ₄ O ₂ Zn 886.62 Monoclinic | $\begin{array}{l} Zn(\textbf{5})_2\\ C_{38}H_{62}N_4O_2Zn\\ 672.28\\ Monoclinic \end{array}$ | $\begin{array}{l} Zn(6)_2 \\ C_{22}H_{30}N_4O_2Zn \\ 447.87 \\ Tetragonal \end{array}$ | $\begin{array}{l} Mg(1)_2 \\ C_{36}H_{58}MgN_4O_2 \\ 603.17 \\ Monoclinic \end{array}$ | $\begin{array}{l} Mg({\bf 2})({\bf 2}^*) \\ C_{40}H_{52}Mg_2N_8O_4 \\ 757.51 \\ Monoclinic \end{array}$ | $\begin{array}{l} Mg({\bf 3})_2 \\ C_{51}H_{60}Cl_8Mg_2N_8O_4 \\ 1181.29 \\ Monoclinic \end{array}$ | $Mg(5)_2$ $C_{38}H_{62}MgN_4O_2$ 631.22 Monoclinic |
|--|--|--|---|---|--|--|--|---|---|---|
| a/Å | 17.3189(3) | 12.0930(10) | 11.3788(3) | 13.4614(7) | 17.0865(3) | 24.06310(10) | 19.8993(3) | 9.4092(5) | 39.731(4) | 17.1130(2) |
| b/Å | 12.4391(3) | 14.8544(12) | 11.6856(2) | 11.9750(7) | 15.0333(2) | 24.06310(10) | 14.4435(2) | 13.8925(12) | 11.9213(9) | 15.0517(2) |
| c/Å | 17.5819(3) | 13.2064(15) | 21.1747(5) | 31.0390(13) | 30.0246(4) | 7.46520(10) | 25.6572(4) | 14.8093(10) | 23.8229(19) | 29.9562(4) |
| a/° | 90 | 90 | 105.712(2) | 90 | 90 | 90 | 90 | 90 | 90 | 90 |
| γ° | 104.339(2) | 105.887(11) | 94.301(2) | 90.965(4) | 91.1930(10) | 90 | 92.1430(10) | 95.647(5) | 99.266(8) | 90.8720(10) |
| γ° | 90 | 90 | 104.647(2) | 90 | 90 | 90 | 90 | 90 | 90 | 90 |
| Unit cell volume/Å ³ | 3669.70(13) | 2281.7(4) | 2590.97(11) | 5002.8(4) | 7710.6(2) | 4322.60(7) | 7369.12(19) | 1926.4(2) | 11136.4(16) | 7715.22(17) |
| Temperature/K | 150(2) | 150(2) | 150(2) | 150(2) | 150(2) | 150(2) | 150(2) | 150(2) | 150(2) | 150(2) |
| Space group | $P2_1/n$ | P2 ₁ | P1 | $P2_1/n$ | <i>I</i> 2/ <i>a</i> | P4 2 ₁ c | $P2_{1}/c$ | $P2_{1}/n$ | <i>C2/c</i> | <i>I</i> 2/ <i>a</i> |
| No. of formula units per unit cell, Z | 4 | 2 | 2 | 4 | 8 | 8 | 8 | 2 | 8 | 8 |
| Radiation type | CuKα | ΜοΚα | CuKα | ΜοΚα | CuKα | CuKα | CuKα | CuKα | ΜοΚα | CuKα |
| Absorption coefficient, μ/mm^{-1} | 1.184 | 1.103 | 5.354 | 0.534 | 1.146 | 1.774 | 0.671 | 0.981 | 0.479 | 0.660 |
| No. of independent reflections | 82933 | 33276 | 20075 | 68799 | 30120 | 29566 | 32351 | 11021 | 38040 | 28402 |
| | 7370 | 9784 | 10229 | 9486 | 7675 | 4104 | 13991 | 3401 | 10155 | 7598 |
| R_{int} | 0.0439 | 0.0649 | 0.0233 | 0.0733 | 0.0317 | 0.0335 | 0.0363 | 0.0466 | 0.1014 | 0.0253 |
| Final R_I values $(I > 2\sigma(I))$ | 0.0349 | 0.0538 | 0.0360 | 0.0501 | 0.0335 | 0.0210 | 0.0572 | 0.0394 | 0.0610 | 0.0505 |
| Final $wR(F^2)$ values $(I > 2\sigma(I))$ | 0.0896 | 0.0725 | 0.0876 | 0.1137 | 0.0893 | 0.0547 | 0.1450 | 0.0932 | 0.0854 | 0.1415 |
| Final R_I values (all data) | 0.0384 | 0.0824 | 0.0399 | 0.0822 | 0.0377 | 0.0215 | 0.0773 | 0.0564 | 0.1590 | 0.0557 |
| Final $WK(F^{-})$ values (all data) | 0.0922 | 0.0800 | 0.0901 | 0.12/1 | 0.0924 | 0.0551 | 0.1585 | 0.1005 | 0.1139 | 0.1444 |

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