# Emulsion Polymerization Induced Self-Assembly of Thermoresponsive Polymer, Poly(N-vinylcaprolactam)

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# **Electronic Supplementary Information**

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## 1. Synthesis of mCTAs

1. Step: Monomethoxy poly(ethylene glycol) mPEG<sub>113</sub>-OH (20.16 g, 4 mmol) and triethylamine (1.238 g, 12 mmol) were dissolved in 100 ml of anhydrous  $CH_2Cl_2$  in a dry 250 ml three-necked flask placed in an ice bath. A solution of 2-bromopropionyl bromide (1.3 ml, 12 mmol) in 20 ml of anhydrous  $CH_2Cl_2$  was added dropwise to the solution over 2 h. After the addition, the mixture was taken off the ice and stirring was continued for 1 d.

The solution was filtered and 200 ml of  $CH_2CI_2$  was added. The organic layer was washed with 1 M HCl solution (3 × 80 ml), 1 M NaOH solution (3 × 80 ml), and deionized water (2 × 80 ml) and dried with anhydrous MgSO<sub>4</sub>. The solution was concentrated in a rotary evaporator. The polymer was precipitated in 400 ml of cold diethyl ether and dried in vacuum, giving mPEG<sub>113</sub>-Br. Gravimetric yield: 19.4 g (96 %). Product analyzed with <sup>1</sup>H-NMR in DMSO-d6, 20 mg ml<sup>-1</sup>.

The same procedure was used for the preparation of mPEG<sub>42</sub>-Br. Gravimetric yield: 89%.

2. Step: mPEG<sub>113</sub>-Br (5.00 g, 1 mmol) and potassium ethyl xanthogenate (0.480 g, 3 mmol) were mixed with 100 ml of anhydrous  $CH_2Cl_2$ , reaction time was 30 min. Solids were filtered off and 50 ml of  $CH_2Cl_2$  were added.

The solution was washed with saturated NaHCO<sub>3</sub> solution (3 × 70 ml) and deionized water (1 × 70 ml). The organic phase was dried with anhydrous MgSO<sub>4</sub> and concentrated to 20 ml at 30 °C using a rotary evaporator. The polymer was precipitated in 400 ml of cold diethyl ether. The product was dried under atmosphere overnight. mPEG<sub>113</sub>-X was obtained. Gravimetric yield: 4.64 g (93%). Product analyzed via <sup>1</sup>H-NMR in DMSO-d6, 20 mg ml-1.

In the preparation of mPEG<sub>42</sub>-X from mPEG<sub>42</sub>-Br, only a 10 min reaction time was used. Longer reaction times seemed to promote difficulties in removing unreacted xanthate from the products, as indicated by impurities in <sup>1</sup>H-NMR spectra. Gravimetric yield: 90%.



Chemical Shift (ppm)

Figure S1. <sup>1</sup>H-NMR spectra

<sup>1</sup>H-NMR (mPEG2000-OH, 500 MHz, DMSO,  $\delta$ ): 4.56 (t, 1H, OH), 3.51 CH<sub>2</sub> (m, 4H per repeating unit of PEG = 181 H), 3.24 (s, CH<sub>3</sub>O),

<sup>1</sup>H-NMR (mPEG2000-Br, 500 MHz, DMSO, δ): 1.72 (d, 3H, C*H*<sub>3</sub>), 3.24 (s, 3H, C*H*<sub>3</sub>O), 3.51 C*H*<sub>2</sub> (m, 4 per repeating unit of PEG), 4.2 (m, 2H, C*H*<sub>2</sub>OCO), 4.69 (q, 1H, C*H*Br)

<sup>1</sup>H-NMR (mPEG2000-X, 500 MHz, DMSO,  $\delta$ ): 1.34 (t, 3H, CH<sub>3</sub>CH<sub>2</sub>O), 1.48 (d, 3H, CH<sub>3</sub>), 3.24 (s, CH<sub>3</sub>O), 3.51 CH<sub>2</sub> (m, 4 per repeating unit of PEG), 4.21 (S, 2H, CH<sub>2</sub>OCO), 4.39 (q, 1H, CHS), 4.62 (q, 2H CH<sub>2</sub>CH<sub>3</sub>)

<sup>1</sup>H-NMR (mPEG5000-OH, 500 MHz, DMSO,  $\delta$ ): 4.56 (t, 1H, OH), 3.51 CH<sub>2</sub> (m, 4 per repeating unit of PEG = 452 H), 3.24 (s, 3H CH<sub>3</sub>O),

<sup>1</sup>H-NMR (mPEG5000-Br, 500 MHz, DMSO, δ): 1.72 (d, 3H, C*H*<sub>3</sub>), 3.24 (s, 3H C*H*<sub>3</sub>O), 3.51 C*H*<sub>2</sub> (m, 4H per repeating unit of PEG), 4.2 (m, 2H, C*H*<sub>2</sub>OCO), 4.69 (q, 1H, C*H*Br)

<sup>1</sup>H-NMR (mPEG5000-X, 500 MHz, DMSO, δ): 1.34 (t, 3 H, CH<sub>3</sub>CH<sub>2</sub>O), 1.48 (d, 3 H, CH<sub>3</sub>), 3.24 (s, 3H CH<sub>3</sub>O), 3.51 CH<sub>2</sub> (m, 4 per repeating unit of PEG), 4.21 (S, 2H, CH<sub>2</sub>OCO), 4.39 (q, 1H, CHS), 4.62 (q, 2H CH<sub>2</sub>CH<sub>3</sub>)

#### 2. <sup>1</sup>H-NMR of a typical product and starting materials



Figure S2. <sup>1</sup>H-NMR spectra of PEG<sub>113</sub>-X (top), NVCL (middle), PEG-b-PNVCL (bottom) with peak assignations

3. Conversion analysis of a typical product via <sup>1</sup>H-NMR



Figure S3. <sup>1</sup>H-NMR spectra of conversion samples taken at different time intervals from reaction mixture (D<sub>2</sub>O)

Conversion was calculated from the integrals of <sup>1</sup>H-NMR spectra presented in figure according to equation 1. In short integral coming from a proton of NVCL (at 7 ppm) was compared to a peak assigned to 2 protons from either NVCL or PNVCL repeating unit (2.8 to 2 ppm).

$$Conversion = 1 - \frac{\sum_{2ppm}^{2.8ppm} \delta(x) dx}{0.5 x \int_{6.8ppm}^{7.2ppm} \delta(x) dx}$$
[S1]

4. Kinetics of polymerizations with varying mCTA and initiator concentration



Figure S4. Conversion versus time for polymerizations with varying mCTA and initiator concentrations; DP = ▲ 194, ● 340, ■ 485, or ▼ 980

#### 0 а b` b, S $\cap$ m II S റ b` $\mathbf{c}$ d a,d,e f с b L 0.0 10.00 1.960 1.000 3.2 | 2.0 4.4 4.0 3.6 2.8 2.4 1.6 0.8 0.4 1.2 PPM 4.8

# 5. $M_n$ of the high molecular weight PNVCL via <sup>1</sup>H-NMR

Figure S5. <sup>1</sup>H-NMR spectra of high molecular weight PNVCL in D<sub>2</sub>O

$$Mn = \frac{n \text{ of repeating units of VCL}}{n \text{ of PEG chains}} * M(NVCL) + Mn(mCTA) = \frac{\int b \, dx}{\int b' dx/(113 * 4)} * 139.19 \text{ g/mol} + 5000 \text{ g/moL}$$
[S2]
$$Mn = \frac{1}{0.078/(113 * 4)} * 139.19 \text{ g/mol} + 5000 \text{ g/mol} = 810 000 \text{ g/mol}$$

# 6. SEC Chromatogram of the high molecular weight PNVCL



Figure S6. SEC chromatogram of the high molecular weight PNVCL in THF

### 7. $M_w$ of the high molecular weight PNVCL via SLS

Refractive index increment (dn/dc) was determined for the high molecular weight polymer in THF with Wyatt Optilab rEX differential refractometer at 25 °C 633 nm. The obtained dn/dc value was 0.137.

The Mw measurements were performed using a setup with Brookhaven Instruments BI-200SM goniometer, a BIC-TurboCorr digital pseudo-cross-correlator, and a BI-CrossCorr detector and a red 637 nm BI-mini L140 opetated with 50 mW power was used as light source and. Sample cell was connected to a Lauda RC 6 CP thermostat and temperature was set to 25 °C. Goniometer alignment was checked before measurements and calibration was done at 90° with toluene. Zimm-plot software was used for Mw determination.

M<sub>w</sub> = 1 270 000 ± 49 000 g/mol.

Second viral coefficient (A<sub>2</sub>) =  $2.43*10^{-4}$  cm<sup>3</sup>mol/g<sup>2</sup>

R<sub>g</sub> = 59.8 nm



Figure S7. Zimm plot for high  $M_n$  PVCL

8. LS data of polymerizations with varied total concentration

[M<sub>0</sub>] / [mCTA] / [I] = 500 / 1 / 0.3

[NVCL]₀ (g/ml)	Conv. (%)	Mn <sub>theory</sub> (g/mol)	Mn <sub>SEC</sub> (g/mol)	Ð (M <sub>n</sub> /M <sub>w</sub> )	Dh* (nm]	R <sub>g</sub> / R <sub>h</sub> *	Table S1. Effect of polymerization concentration, with LS data
0.1	97	72 500	72 300	1.5	225	0.76	
0.2	98	73 200	71 400	1.4	220	0.74	
0.3	98	73 200	72 100	1.3	245	0.78	

\*measured at 50 °C from diluted particle dispersion before cooling

# 9. DLS data of polymerizations with varied initiator concentration

Table S2. Synthesis of polymers with different initiator concentrations, with LS data

C <sub>0</sub> (NVCL)	) = 0.1 g / n	nl <i>i.e</i> [M]₀∶	=0.718 mol / I
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[M <sub>0</sub> ]:[CTA]:[I]	Conv. (%)	Mn <sub>theory</sub> (g/mol)	Mn <sub>SEC</sub> (g/mol)	Ð (M <sub>n</sub> /M <sub>w</sub> )	Dh* (nm)	Rg / Rh*
500/1/0.3	97	72 500	72 300	1.5	220	0.77
500/1/0.6	99	73 900	74 700	1.2	225	0.80
500/1/0.9	100	74 600	75 700	1.2	210	0.82

\*measured at 50 °C from particle diluted dispersion before cooling

# 10. Cryo-TEM images of the high M<sub>n</sub> polymer particles

This cryo-EM sample was imaged with FEI Tecnai F20 microscope and micrographs were recorded with Gatan US4000 CCD camera.



Figure S8. Cryo-TEM image of high M<sub>n</sub> polymer particle, scale bar is 200 nm wide.



Figure S9. Another cryo-TEM image of high M<sub>n</sub> polymer particle, scale bar is 200 nm wide.

# 11. Solubility of NVCL to $H_2O$ and to TRIS buffer (100 mM, pH = 7.4)

Solubility of NVCL to TRIS buffer (100 mM, pH = 7.4) at 22 °C and at 50 °C were tested. At 22 °C, 5 ml of TRIS was mixed with 500 mg of NVCL at room temperature for 2 days to get saturated solution. Insoluble material was separated via centrifugation (2h, 10 000 rcf). 0.1 ml of supernatant was diluted with 1 ml  $D_2O$ . The amount of dissolved NVCL was determined via <sup>1</sup>H-NMR by comparing integral from TRIS protons with integral from NVCL (1 of the <sup>1</sup>H-NMR spectra presented as figure S10). Experiment was performed in triplicate.

Solubility of NVCL to 100mM TRIS at 22 °C is  $35 \pm 2$  g/L.

Solubility of NVCL to TRIS buffer at 50 °C was tested by mixing 5 ml of TRIS with 500 mg of NVCL for 4 h to get saturated solution. At 50 C, the monomer is liquid and the mixing time was deemed sufficient via number of test with different mixing times. Supernatant was separated via centrifugation (30 min, 3773 rcf, at 50 °C). 0.1 ml of supernatant was diluted with 1 ml D<sub>2</sub>O. The amount of dissolved NVCL was determined by comparing integral from TRIS protons with integral from NVCL. Experiment was performed in triplicate.

Solubility of NVCL to 100mM TRIS at 50 °C is  $36 \pm 2 \text{ g/L}$ .

Solubility of NVCL to  $H_20$  was tested at 22 °C. Test was same as with TRIS at 22 °C. Only difference was that after centrifugation supernatant was mixed in 1 to 1 ration with 100 mM TRIS. 0.1 ml of the mixture was diluted with 1 ml  $D_2O$ . The amount of dissolved NVCL was determined via <sup>1</sup>H-NMR by comparing integral from TRIS protons with integral from NVCL. Experiment was done in triplicate.

Solubility of NVCL to  $H_2O$  at 22 °C is 15 ± 2 g/L.



Figure S10. <sup>1</sup>H-NMR spectrum from solubility test