Electronic Supplementary Material

Construction of multi-hollow polymer microspheres with tailored mesoporous wall

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I. Synthesis of Macro@Meso-SPS microspheres

The TEM images of PS and SPS microspheres were obtained on transmission electron microscope (H-7650) at an accelerate voltage of 100 kV. The Samples were prepared by dropping the ethanol dispersion of microspheres onto a piece of copper grids and being dried at ambient temperature. The weight average diameter (D_w), number average diameter (D_n), and polydispersion index of the diameters (PDI_D) were calculated by the following equations. At least 100 microspheres were measured in the TEM images.

$$D_{w} = \frac{\sum n_{i} D_{i}^{4}}{\sum n_{i} D_{i}^{3}}, D_{n} = \frac{\sum n_{i} D_{i}}{\sum n_{i}}, PDI_{D} = \frac{D_{w}}{D_{n}}$$

where n_i is the number of particles with a diameter of D_i . The D_n and PDI are listed in the Table S1.



Fig. S1. TEM images of PS-R1 (a), SPS-R1 (b), PS-R2 (c), SPS-R2 (d), PS-C (e), and SPS-C microspheres (f).

Table S1 The synthetic condition and the size of different original PS microspheres									
	St	PVP	Solv	Solvent		Ď	D	D_n	DDI
	(mL)	(g)	ethanol (mL)	water (mL)	(mg)	(Gy min ⁻¹)	(kGy)	(µm)	
PS-R1	4	1	15	1	0	60	54	0.81	1.004
PS-R2	4	1	15	1	0	80	72	1.15	1.004
PS-C	10	1	95	5	80	-	-	0.83	1.010

	original PS	DS (0/)§	DS (0/)§	D_n (µm)	DDI
	microspheres	DS_{XPS} (70) ³	$DS_{EA}(70)^{\circ}$		PDI _D
SPS-R1	PS-R1	45.45	2.04	1.14	1.004
SPS-C	PS-C	54.80	2.33	0.84	1.020

Table S2 The degree of sulfonation and the size of different SPS seed microspheres

[§] The degree of sulfonation (*DS*) of the SPS microspheres measured from XPS (*DS*_{XPS}) and EA (*DS*_{EA}) analysis was obtained according to the following Eqs S1 and S2, respectively:

$$DS_{XPS} = 8 \left(\frac{atom\% S}{atom\% C}\right) \times 100\%$$
(S1)

$$DS_{EA} = {\binom{96}{32}} {\binom{wt\%}{5}} \times 100\%$$
(S2)

 DS_{EA} can be considered as the *DS* of the whole SPS microspheres, while DS_{XPS} is the *DS* of the surface layer of SPS microspheres since the detection depth of XPS is only about 5 nm. The data of SPS microspheres measured by XPS and EA were listed in Table S3 and Table S4, respectively.

Table S3 The XPS data of different SPS microspheres C ratio (at.%) O ratio (at.%) N ratio (at.%) S ratio (at.%) SPS-R1 77.62 4.41 15.25 2.71 SPS-C 74.60 17.26 3.03 5.11 Table S4 The EA data of different SPS microspheres. C ratio H ratio S ratio O ratio N ratio (wt.%) (wt.%) (wt.%) (wt.%) (wt.%) SPS-R1 86.40 7.62 1.49 0.23 1.31 SPS-C 87.48 0.68 8.13 1.39 0.30

Seed	Swelling-osmosis	PIPS process					
microspheres	process	Sample identity	MMA/SPS weight ratio	Polymerization time of MMA (h)			
		Macro@Meso-SPS-R1-1	1:5	10			
		Macro@Meso-SPS-R1-2	1:2	10			
		Macro@Meso-SPS-R1-3	1:1	10			
SPS-R1	Macro-SPS-R1	Macro@Meso-SPS-R1-4	2:1	10			
		Macro@Meso-SPS-R1-5	5:1	10			
		Macro@Meso-SPS-R1-6	1:1	5			
		Macro@Meso-SPS-R1-7	1:1	20			
SPS-R2	Macro-SPS-R2	Macro@Meso-SPS-R2	1:1	10			
SPS-C	Macro-SPS-C	Macro-SPS-C-P	1:1	10			

Table S5 The identifications (IDs) of the samples obtained after the SPS seed microspheres were treated in the swelling-osmosis and the PIPS processes with different conditions

Samula identity	BET surface area	Pore volume	Most probable surface pore size calculated from BJH method
Sample identity	$(m^2 g^{-1})$	$(cm^3 g^{-1})$	(nm)
Macro@Meso-SPS-R1-1	14.1	0.0431	~ 2
Macro@Meso-SPS-R1-2	15.6	0.0463	11.2
Macro@Meso-SPS-R1-3	5.8	0.0261	44.7
Macro@Meso-SPS-R1-4	12.0	0.0468	45.3
Macro@Meso-SPS-R1-5	5.3	0.0178	> 50
Macro@Meso-SPS-R1-6	14.5	0.0431	14.9
Macro@Meso-SPS-R1-7	16.3	0.1981	> 50
Macro@Meso-SPS-R2	15.0	0.0613	3.1
Macro-SPS-C-P	59.7	0.4217	> 50

 $Table \ S6 \ BET \ surface \ areas, \ pore \ volumes, \ and \ the \ most \ probable \ surface \ pore \ sizes \ of \ different \ porous \ microspheres \ measured \ by \ N_2 \ adsorption-desorption \ isotherms$



II. Loading and sustained release behaviors of MO from Macro@Meso-SPS microspheres

Fig. S2. Standard work curves (a) of MO in DMF (black) and PBS buffer (red). UV-vis spectra of different MO-loaded Macro@Meso-SPS microspheres in DMF (b). UV-vis spectra of MO in PBS buffer released from Macro@Meso-SPS microspheres with different surficial pore sizes: 3.1 (c), 11.2 (d), 44.7 (e), and > 50 nm (f).

III. ¹H NMR spectra of different PS microspheres



Fig. S3 ¹H NMR spectra of PS-R1, PS-C, and PVP in CDCl₃ (the insets are the corresponding magnified spectra from $\delta = 1.5 - 1.6$, and 3.0 - 4.0 ppm). The small peak at $\delta = 1.53$ of PS-C should be attributed to AIBN residues (Fordham, P. J., Gramshaw, J. W., & Castle, L., Food Addit. Contam., 2001, 18, 461.)



Fig. S4 Low resolution (**a**) and N 1s (**b**) XPS spectra of PS-R1 microspheres; XPS spectra of SPS-R1 (**c**) and SPS-C (**d**) microspheres.

V. Release kinetic models of MO from Macro@Meso-SPS microspheres

The cumulative release ratios (*R*) of MO from hierarchically porous SPS microspheres with different surficial pore sizes (3.1, 11.2, 44.7, and > 50 nm) were fitted by the following three release kinetic models: first-order,¹ Korsmeyer–Peppas,² and Higuchi models.³ Because of burst effect for the highly soluble molecules^{4,5} and the rapid release for the residual MO on surface, the above three models are modified as follows:

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$$R = (R_e - b_1) \times (1 - e^{-k_1 t}) + b_1 \quad \text{modified first-order model}$$
$$R = k_{KP} \times t^n + b_{KP} \quad \text{modified Korsmeyer-Peppas model}$$
$$R = k_H \times \sqrt{t} + b_H \quad \text{modified Higuchi model}$$

where *R* and R_e are the cumulative release ratios of MO at time *t* and at equilibrium. k_1 , k_{KP} and k_H are the release rate coefficients of first-order, Korsmeyer-Peppas and Higuchi kinetic models, respectively. *n* is the diffusivity coefficient. b_1 , b_{KP} , and b_H are constants related to the burst effect for the highly soluble molecules and the rapid release of the residual MO on surface. The corresponding kinetic parameters are listed in Table S7.

Table S7. The fitting parameters of different kinetic models for the release of MO from different

 hierarchically porous SPS microspheres

Madal	Doromotor	Size of surficial pores				
WIOdel	Parameter	3.1 nm	11.2 nm	44.7 nm	> 50 nm	
	R ²	0.992	0.996	0.959	0.945	
Tinet and a	R_e	5.35×10^{-1}	7.30×10^{-1}	7.77×10^{-1}	7.91×10^{-1}	
First-order	k_{l}	7.12×10^{-2}	2.75×10^{-1}	3.16×10 ⁻¹	5.67×10^{-1}	
	b_{I}	6.94×10^{-2}	1.13×10^{-1}	2.79×10^{-1}	3.06×10^{-1}	
	R ²	0.994	0.865	0.830	0.813	
<i>V</i> D	k_{KP}	7.45×10^{-2}	1.59×10^{-1}	1.46×10^{-1}	2.14×10^{-1}	
Korsmeyer–Peppas	b_{KP}	3.31×10 ⁻²	1.46×10^{-1}	3.00×10^{-1}	3.08×10^{-1}	
	п	0.551	0.473	0.424	0.305	
	R ²	0.993	0.932	0.904	0.864	
Higuchi	k_H	8.97×10^{-2}	1.43×10^{-1}	1.08×10^{-1}	9.12×10 ⁻²	
	b_H	1.74×10 ⁻²	1.64×10 ⁻¹	3.41×10 ⁻¹	4.38×10 ⁻¹	

If *R* is plotted versus the square root of *t*, it is evident that the release curve of Macro@Meso-SPS microspheres with surficial pores around 3.1 nm can fit Higuchi model well (Figure 10B), while the other three porous microspheres also show Higuchi release profiles at the beginning (approximately R < 60%). The fitting results are listed in Table S8.

Table S8. The fitting parameters of the release curves for MO from different hierarchically porousSPS microspheres at R is approximately below 60%.

Doromotora -	Size of surficial pores					
Parameters -	3.1 nm	11.2 nm	44.7 nm	> 50 nm		
R ²	0.993	0.996	0.982	0.991		
k_H	8.97×10^{-2}	2.50×10^{-1}	2.57×10 ⁻¹	3.57×10 ⁻¹		
b_H	1.74×10 ⁻²	2.10×10 ⁻²	1.68×10 ⁻¹	1.74×10^{-1}		

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