Electronic Supplementary Information (ESI):

A Real Time Bioimaging for Mitochondria by Taking Aggregating Process of Aggregation-Induced Emission Near-Infrared Dyes with Wash-Free Staining

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1. Experimental

1.1 Materials

These compounds were obtained by conventional synthetic routes, as shown in Schemes S1~S12. All chemicals were used by commercial suppliers without further purification unless otherwise stated. CuCl, Pd(PPh₃)₄ and ethyl 4-aminobenzoate were bought from J&K. POCl₃ bought from Xiya Reagent. 2, 5-dimethoxytetrahydrofuran, (4was (diphenylamino)phenyl)boronic acid, and 1-bromopyrrolidine-2, 5-dione were bought from Energy Chemical. KPF₆ was bought from Aladdin. Chloroform-d and DMSO-d₆ were bought from Innochem. DMEM, PBS and trpsin were purchased from Coring Co., Ltd. MitoTracker Green, MitoTracker Red and Annexin V-FITC/PI Apoptosis Detection Kit were purchased from Beyotime technologies Co., Ltd. The deionized water was used for all experiments.

1.2 Equipments

¹H and ¹³C NMR spectra were measured on a Bruker AV 400 spectrometer. Mass spectra were collected by using a Finnigan BIFLEX III mass spectrometer. UV-Vis spectra were recorded on a TU-1901 double beam UV-Vis spectrophotometer. Fluorescence spectra were measured on a Hitachi F-7000 fluorescence spectrophotometer. The size of aggregates was measured by a Malvern ZEN3600 Zetasizer. PL quantum yields were measured by using an integrating sphere on Nanolog FL3-2iHR fluorescence spectrometer (Horiba Jobinyvon) and PL time-resolved decays were measured by Deltaflex ultrafast lifetime spectrofluorometer (Horiba Jobinyvon). Morphologies of the aggregates were imaged on a Hitachi S-4800

scanning electron microscope (SEM) operating at an accelerating voltage of 5 kV. The fluorescence imaging in living cell was recorded with a Leica TCS SP5 laser scanning confocal microscope. Cell apoptosis experiments were conducted with a Beckman FC500MCL flow cytometer.

1.3 Synthesis



Scheme S1. Synthesis routes for TPP-1, TPP-2, and TPP-3

1.3.1 Preparation of TPP-1.



Scheme S2. The synthetic route to compound 1

1,2-diphenylethanediyne^[1] (2.0104 g, 10.04 mmol), ethyl *p*-aminobenzoate (6.6340 g, 40.16 mmol) and CuCl (0.0983 g, 1.00 mmol) were added into a 100 ml polymerization tube reacted at 120 °C for 24 h under nitrogen. After the reaction was completed, the mixture was dissolved in dichloromethane, then washed three times with water and dilute hydrochloric acid. The solution was dried over anhydrous MgSO4 and filtered by suction. The solvent was evaporated by vacuum distillation. The crude product was purified by gel chromatography using dichloromethane/petroleum ether mixture (1/5, V_d/V_p) as eluent. A white flocculent product **1** was obtained with a yield of 53.7%. ¹H NMR (400 MHz, CDCl₃): δ = 7.90 (d, *J* = 8.4 Hz, 2H), 7.22-7.12 (m, 6H), 7.09-7.00 (m, 6H), 6.48 (s, 2H), 4.35 (q, *J* = 7.2 Hz, 2H), 1.37 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃): δ = 166.00, 142.89, 135.75, 132.93, 130.10, 129.01, 128.86, 128.71, 128.07, 126.48, 110.60, 61.20, 14.31. MS (ESI, *m/z*) Calcd for C₂₅H₂₁NO₂ [M+H]⁺: 368.15, found: 368.16.



Figure S1. ¹H NMR spectrum of 1 in CDCl₃



Figure S2. ¹³C NMR spectrum of 1 in CDCl₃.



Scheme S3. The synthetic route to compound 2

POCl₃ (200 µL, 2.20 mmol) was dropped slowly into DMF (24 mL) at 0 °C and stirred for another 1 h at room temperature. To the above solution was added a dichloromethane solution of **1** (0.7343 g, 2.00 mmol). After the mixture was stirred for 12 h at room temperature. The residue was poured into water (250 mL) and extracted with dichloromethane; then the solution of dichloromethane was dried over anhydrous MgSO₄ and filtered by suction. The solvent was evaporated by vacuum distillation. The crude product was purified by gel chromatography using dichloromethane/petroleum ether mixture (1/5, V_d/V_p) as eluent. Product **2** was obtained with the yield of 83.2%. ¹H NMR (400 MHz, DMSO-*d*₆): δ = 9.58 (s, 1H), 7.83 (d, *J* = 7.9 Hz, 2H), 7.37-7.27 (m, 6H), 7.25 (d, *J* = 7.5 Hz, 4H), 7.14 (d, *J* = 5.6 Hz, 2H), 6.91 (s, 1H), 4.27 (q, *J* = 6.9 Hz, 2H), 1.28 (t, *J* = 6.9 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃): δ = 186.91, 165.59, 141.19, 136.64, 131.38, 131.17, 130.21, 129.89, 129.06, 128.91, 128.81, 128.62, 128.32, 128.27, 127.58, 124.53, 107.93, 61.36, 14.28. MS (ESI, *m/z*) Calcd for C₂₆H₂₁NO₃ [M+Na]⁺: 418.14, found: 417.71.



Figure S4. ¹H NMR spectrum of 2 in DMSO- d_6 .



Figure S5. ¹³C NMR spectrum of 2 in CDCl₃.



Figure S6. MS spectrum of 2.



Scheme S4. The synthetic route to target compound TPP-1.

A solution of **2** (0.2377 g, 0.60 mmol) and indolium iodide salt^[2] (0.2068 g, 0.65 mmol) in dry ethanol (15 mL) was refluxed for 24 h. After the reaction mixture was cooled to room temperature, the solvent was evaporated under reduced pressure. The solid was dissolved in THF (5 mL) and a saturated aqueous solution of KPF6 (5 mL) was then added. After stirring for 30 min, the solvent was evaporated to get the crude product which was purified by a silica gel column chromatography using dichloromethane/methanol mixture (10/1, V_d/V_m) as eluent to give a red solid in 78.2% yield. ¹H NMR (400 MHz, DMSO-*d*₆): $\delta = 7.94$ (d, J = 16.0 Hz,

1H), 7.88 (d, J = 8.0, 2H), 7.82 (d, J = 8.0, 1H), 7.76 (d, J = 8.0, 1H), 7.68 (s, 1H), 7.61-7.47 (m, 6H), 7.32 (m, 7H), 7.20 (d, J = 6.9 Hz, 2H), 4.58 (q, J = 7.3 Hz, 2H), 4.28 (q, J = 8.9, 6.8 Hz, 2H), 1.53 (s, 6H), 1.42 (t, J = 7.3 Hz, 3H), 1.29 (t, J = 6.8 Hz, 3H). ¹³C NMR (100 MHz, DMSO- d_6): $\delta = 180.70, 165.27, 148.52, 143.14, 141.38, 141.10, 138.80, 131.24, 131.13, 130.39, 130.27, 129.89, 129.60, 129.10, 129.07, 129.04, 128.95, 128.87, 128.54, 123.44, 122.33, 114.66, 108.94, 108.73, 61.61, 51.63, 41.83, 26.78, 14.52, 13.82. HR-MS (ESI, <math>m/z$) Calcd for C₃₉H₃₇N₂O₂⁺ [M]⁺: 565.2849, found: 565.2846, error -0.6 ppm.

Figure S7. ¹H NMR spectrum of TPP-1 in DMSO-*d*₆.

Figure S8. ¹³C NMR spectrum of TPP-1 in DMSO-d6.

Figure S9. HR-MS spectrum of TPP-1.

1.3.2 Preparation of TPP-2.

Scheme S5. The synthetic route to compound 3.

Ethyl 4-aminobenzoate (1.6519 g, 10.00 mmol) and 2, 5-dimethoxytetrahydrofuran (1.3216 g, 10.00 mmol) were dissolved in 25 mL acetic acid, then refluxed for 12 h. After cooled to room temperature, the solvent was evaporated under reduced pressure. The residue was dissolved in dichloromethane and washed with water for three times, followed by extraction with dichloromethane. Removed the water from the solution with anhydrous MgSO₄, then filtered by suction and evaporated by vacuum distillation. The crude product was purified by a silica gel column using dichloromethane/petroleum ether mixture (1/6, V_d/V_p) as eluent. Product **3** was obtained with the yield of 83.2%. ¹H NMR (400 MHz, CDCl₃): δ = 8.10 (d, *J* = 8.2 Hz, 2H), 7.44 (d, *J* = 8.1 Hz, 2H), 7.16 (s, 2H), 6.38 (s, 2H), 4.39 (q, *J* = 7.0 Hz, 2H), 1.41 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃): δ = 165.96, 143.94, 131.29, 119.51, 119.28, 119.07, 111.49, 61.07, 14.40. MS (MALDI, *m*/*z*) Calcd for C₁₃H₁₃NO₂ [M+Na]⁺: 238.08, found: 237.61.

Figure S10. ¹H NMR spectrum of 3 in CDCl₃.

Figure S11. ¹³C NMR spectrum of 3 in CDCl₃.

Figure S12. MS spectrum of 3.

Scheme S6. The synthetic route to compound 4.

The compound **3** (0.9024 g, 5.10 mmol) was added into 25 mL DMF. 1-Bromopyrrolidine-2, 5-dione (1.0755 g, 5.00 mmol) was dissolved in 5 mL DMF, and then slowly added dropwise into the DMF solution of compound **3** with stirring. The mixture reacted at room temperature for 12 h. The solution was washed by water and extracted with dichloromethane. The dichloromethane solution was dried over anhydrous MgSO₄, then filtered by suction and evaporated by vacuum distillation. The crude product was purified by gel chromatography using dichloromethane/petroleum ether mixture (1/6, V_d/V_p) as eluent. Product **4** was obtained with 78.2% yield. ¹H NMR (400 MHz, CDCl₃): $\delta = 8.10$ (d, J = 8.1 Hz, 2H), 7.34 (d, J = 8.1Hz, 2H), 7.13 (s, 2H), 4.39 (d, J = 7.2 Hz, 2H), 1.41 (t, J = 7.1 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃): $\delta = 165.56, 142.43, 131.48, 128.56, 119.19, 119.16, 102.37, 61.29, 14.38. MS (APCI,$ *m/z*) Calcd for C₁₃H₁₁Br₂NO₂ [M+H]⁺: 373.91, found: 373.92.

Figure S13. ¹H NMR spectrum of 4 in CDCl₃.

Figure S14. ¹³C NMR spectrum of 4 in CDCl₃.

Figure S15. MS spectrum of 4.

Scheme S7. The synthetic route to compound 5.

Compound **4** (0.7458 g, 2.00 mmol), (4-(diphenylamino)phenyl)boronic acid (1.3011 g, 4.50 mmol) and Pd(PPh₃)₄ (0.0924 g, 0.08 mmol) were dissolved in degassed acetonitrile (20 ml) and then a saturated aqueous solution of K_2CO_3 (5 mL) was added. After stirring for 24 h at 80 °C under nitrogen protection, the solution was cooled to room temperature, and then removed the solvent under reduced pressure. The residue was extracted with dichloromethane and washed with water, then the organic layer was dried over anhydrous MgSO₄. The solvent was removed under reduced pressure, and the crude product was purified by silica gel column

using dichloromethane/petroleum mixture (1/6, V_d/V_p) as eluent to give the compound **5** with 59.5% yield. ¹H NMR (400 MHz, CDCl₃): $\delta = 7.94$ (d, J = 8.0 Hz, 2H), 7.29-7.17 (m, 8H), 7.12 (d, J = 8.1 Hz, 2H), 7.09-7.02 (m, 8H), 7.02-6.95 (m, 4H), 6.92-6.76 (m, 8H), 6.43 (s, 2H), 4.37 (q, J = 7.2 Hz, 2H), 1.40 (t, J = 6.8 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃): $\delta = 165.98$, 147.56, 146.25, 143.23, 135.32, 130.01, 129.50, 129.27, 128.85, 124.56, 122.99, 122.73, 109.93, 61.21, 14.38. MS (MALDI, m/z) Calcd for C₄₉H₃₉N₃O₂ [M]⁺: 701.30, found: 701.07.

Figure S16. ¹H NMR spectrum of 5 in CDCl₃.

Figure S17. ¹³C NMR spectrum of 5 in CDCl₃.

Figure S18. MS spectrum of 5.

Scheme S8. The synthetic route to compound 6.

POCl₃ (50 µL, 0.55 mmol) was slowly dropped into DMF (24 mL) at 0 °C, then the mixture was stirred for 1 h at room temperature. To the above solution was added a dichloromethane solution of compound **5** (0.3507 g, 0.50 mmol). After the mixture was stirred for 12 h at room temperature, the solution was poured into water (250 mL) and extracted with dichloromethane. The dichloromethane solution was dried over anhydrous MgSO₄ and filtered by suction, and then the solvent was evaporated by vacuum distillation. The residue was purified by gel chromatography using dichloromethane/petroleum mixture (1/2, V_d/V_p) as eluent. Product **6** was obtained with a yield of 73.2%. ¹H NMR (400 MHz, CDCl₃): δ = 9.76 (s, 1H), 7.95 (d, *J* = 8.0 Hz, 2H), 7.30-7.21 (m, 9H), 7.12-6.99 (m, 14H), 6.96 (d, *J* = 8.3 Hz, 2H), 6.93-6.85 (m, 6H), 4.39 (q, *J* = 7.1 Hz, 2H), 1.41 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃): δ = 187.08, 165.68, 148.28, 147.31, 147.21, 147.03, 141.60, 136.38, 131.92, 130.09, 129.83, 129.60, 129.47, 129.35, 128.77, 125.23, 124.89, 124.26, 123.81, 123.36, 122.16, 121.67, 121.26, 107.09, 61.38, 14.36. MS (MALDI, *m/z*) Calcd for C₅₀H₃₉N₃O₃ [M]⁺ : 729.30, found: 729.07.

Figure S19. ¹H NMR spectrum of 6 in CDCl₃.

Figure S20. ¹³C NMR spectrum of 6 in CDCl₃.

Figure S21. MS spectrum of 6.

Scheme S9. The synthetic route to target compound TPP-2.

A solution of **6** (0.2917 g, 0.40 mmol) and indolium iodide salt (0.1553 g, 0.45 mmol) in ethanol (15 mL) was refluxed for 24 h. After the reaction mixture was cooled to room temperature, the solvent was evaporated under reduced pressure. The residue was dissolved in THF (5 mL) and a saturated aqueous solution of KPF₆ (5 mL) was then added. After stirring for 30 min, the solution was evaporated to get crude product that was purified by a silica gel column chromatography using dichloromethane/methanol mixture (10/1, V_d/V_m) as eluent to give a red solid in 52.2% yield. ¹H NMR (400 MHz, DMSO-*d*₆): δ = 7.98 (d, *J* = 16.0 Hz, 1H), 7.94 (s, 2H), 7.82 (t, *J* = 7.5 Hz, 2H), 7.66-7.49 (m, 3H), 7.46-7.25 (m, 11H), 7.20-7.02 (m, 12H), 6.99 (d, *J* = 7.8 Hz, 4H), 6.92 (d, *J* = 8.0 Hz, 2H), 6.84 (d, *J* = 8.1 Hz, 2H), 4.56 (q, *J* =

7.0 Hz, 2H), 4.31 (q, J = 6.9 Hz, 2H), 1.59 (s, 6H), 1.42 (t, J = 6.9 Hz, 3H), 1.32 (t, J = 7.0 Hz, 3H). ¹³C NMR (100 MHz, DMSO- d_6): $\delta = 180.43$, 165.35, 148.62, 147.12, 146.84, 145.20, 143.03, 141.52, 141.17, 138.66, 132.32, 130.45, 130.30, 130.13, 130.08, 129.58, 125.40, 124.95, 124.69, 124.53, 124.15, 123.54, 122.46, 122.23, 121.54, 121.31, 108.33, 108.17, 61.63, 51.56, 26.87, 14.57, 13.77. HR-MS (ESI, m/z) Calcd for C₆₄H₅₆N₃O₂⁺ [M+H]⁺: 899.4320, found: 899.4293, error -3.0 ppm.

Figure S22. ¹H NMR spectrum of TPP-2 in DMSO-*d*₆.

Figure S23. ¹³C NMR spectrum of TPP-2 in DMSO-*d*₆.

Figure S24. HR-MS spectrum of TPP-2.

1.3.3 Preparation of TPP-3.

Scheme S10. The synthetic route to compound 7.

1,4-Bis(4-bromophenyl) buta-1,3-diyne^[3] (3.5990 g, 10.00 mmol), ethyl p-aminobenzoate (6.6076 g, 40.00 mmol) and CuCl (0.0983 g, 1.00 mmol) were added into a 100 ml Schlenk tube reacted at 130 °C for 24 h under nitrogen. After the reaction was completed, dichloromethane was added into the tube and then washed with dilute hydrochloric acid for three times. The organic solution was dried over anhydrous MgSO4 and filtered by suction, and then the solvent was evaporated by vacuum distillation. The crude product was purified by gel chromatography using dichloromethane/petroleum ether mixture (1/5, V_d/V_p) as eluent. Product **7** was obtained with a yield of 50.6%. ¹H NMR (400 MHz, CDCl₃): δ = 7.94 (d, *J* = 8.0 Hz, 2H), 7.38-7.30 (m, 4H), 7.07 (d, *J* = 8.4 Hz, 2H), 7.01-6.80 (m, 4H), 6.50 (s, 2H), 4.40 (q, *J* = 7.2 Hz, 2H), 1.42 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃): δ = 165.72, 134.89, 131.29, 130.39, 130.18, 129.90, 129.57, 128.62, 120.86, 110.99, 61.32. MS (APCI, *m/z*) Calcd for C₂₅H₁₉Br₂NO₂ [M+H]⁺: 526.0, found: 526.0.

Figure S25. ¹H NMR spectrum of 7 in CDCl₃.

Figure S26. ¹³C NMR spectrum of 7 in CDCl₃.

Figure S27. MS spectrum of 7.

Scheme S11. The synthetic route to compound 8.

Compound 7 (1.0500 g, 2.00 mmol), (4-(diphenylamino)phenyl)boronic acid (1.3011 g, 4.50 mmol), Pd(PPh₃)₄ (0.0924 g, 0.08 mmol) were dissolved in degassed acetonitrile (20 ml) and a saturated aqueous solution of K₂CO₃ (5 mL) was then added. After stirring for 24 h at 80 °C under nitrogen, the solution was cooled to room temperature, removed the solvent under reduced pressure. The residue was extracted by dichloromethane and then washed with water. The organic solution was dried over anhydrous MgSO₄, and then the solvent was removed under reduced pressure. The crude product was purified by silica gel column using dichloromethane/petroleum mixture (1/6, V_d/V_p) as eluent to give compound **8** with 68.5%

yield. ¹H NMR (400 MHz, CDCl₃): $\delta = 7.94$ (d, J = 8.0 Hz, 2H), 7.43 (dd, J = 8.0, 15.2 Hz, 8H), 7.31-7.20(m, 9H), 7.18-7.06 (m, 17H), 7.02 (t, J = 7.5 Hz, 4H), 6.54 (s, 2H), 4.36 (q, J = 7.2 Hz, 2H), 1.37 (t, J = 7.2 Hz, 3H).¹³C NMR (100 MHz, CDCl₃): $\delta = 166.00$, 147.55, 146.28, 143.25, 130.03, 129.54, 129.28, 129.05, 128.86, 124.59, 123.01, 122.74, 109.96, 61.22, 14.40. MS (APCI, m/z) Calcd for C₆₁H₄₇N₃O₂ [M+H]⁺: 854.37, found: 854.37.

Figure S28. ¹H NMR spectrum of 8 in CDCl₃.

Figure S29. ¹³C NMR spectrum of 8 in CDCl₃.

Figure S30. MS spectrum of 8.

Scheme S12. The synthetic route to compound 9.

POCl₃ (100 µL, 1.10 mmol) was slowly dropped into DMF (24 mL) at 0 °C and stirred for 1 h at room temperature. Then dichloromethane solution of compound **8** (0.8534 g, 1.00 mmol) was added into the above solution. After the mixture was stirred for 12 h at room temperature, the resultant solution was poured into water (250 mL) and extracted by dichloromethane. The dichloromethane solution was dried over anhydrous MgSO₄ and filtered by suction, and then the solvent was evaporated by vacuum distillation. The crude product was purified by gel chromatography using dichloromethane/petroleum mixture (1/2, V_d/V_p) as eluent. Product **9** was obtained with a yield of 73.8%. ¹H NMR (400 MHz, CDCl₃): $\delta = 9.79$ (s, 1H), 7.92 (d, J = 8.1 Hz, 2H), 7.53-7.46 (m, 3H), 7.46-7.38 (m, 5H), 7.32-7.17 (m, 11H), 7.16-7.07 (m, 16H), 7.07-6.97 (m, 5H), 4.34 (q, J = 7.2 Hz, 2H), 1.36 (t, J = 7.2 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃): δ D= 186.98, 165.64, 147.62, 147.56, 147.50, 141.35, 133.91, 131.59, 130.39, 129.99, 129.74, 129.39, 129.23, 128.75, 127.72, 127.60, 127.29, 126.39, 126.26, 124.68, 124.63, 124.54, 123.82, 123.72, 123.22, 123.10, 108.10, 61.42, 14.34. MS (APCI, *m*/z) Calcd for C₆₂H₄₇N₃O₃ [M+H]⁺: 882.36, found: 882.37.

Figure S31. ¹H NMR spectrum of 9 in CDCl₃.

Figure S32. ¹³C NMR spectrum of 9 in CDCl₃.

Scheme S13. The synthetic route to target compound TPP-3.

A solution of **9** (0.2377 mg, 0.60 mmol) and indolium iodide salt (0.2068 mg, 0.65 mmol) in dry ethanol (15 mL) were fluxed for 24 h. After the reaction mixture was cooled to room temperature, the solvent was evaporated under reduced pressure. The residue was dissolved in THF (5 mL) and a saturated aqueous solution of KPF₆ (5 mL) was then added. After stirring for 30 min, the solvent was removed to give the crude porduct. The crude product was purified by a silica gel column chromatography using dichloromethane/methanol mixture (10/1, V_d/V_m) as eluent to give a red solid in 78.2% yield. ¹H NMR (400 MHz, DMSO-*d*₆): δ = 8.02 (d, *J* = 16.0 Hz, 1H), 7.93 (d, *J* = 8.0 Hz, 2H), 7.82 (d, *J* = 8.0 Hz, 1H) 7.75 (t, *J* = 7.5 Hz, 4H), 7.68 (d, *J* = 8.0 Hz, 2H), 7.63-7.47 (m, 7H), 7.42 (d, *J* = 8.0 Hz, 2H), 7.37-7.31 (m,

10H), 7.25 (d, J = 7.6 Hz, 2H), 7.11-7.01 (m, 16H), 4.59 (q, J = 6.4 Hz, 2H), 4.28 (q, J = 6.4 Hz, 2H), 1.57 (s, 6H), 1.43 (t, J = 6.4 Hz, 3H), 1.28 (t, J = 7.2 Hz, 3H). ¹³C NMR (100 MHz, DMSO-*d*₆): $\delta = 180.66$, 165.29, 147.52, 147.40, 147.35, 141.51, 140.41, 139.33, 132.54, 131.80, 130.57, 130.37, 130.15, 130.11, 129.67, 129.43, 128.12, 127.99, 126.44, 126.35, 124.83, 124.73, 124.00, 123.88, 123.52, 123.44, 61.63, 51.65, 26.83, 14.52, 13.84. HR-MS (ESI, *m/z*) Calcd for C₇₅H₆₃N₄O₂ [M]⁺: 1051.4945, found: 1051.4933, error -1.2 ppm.

Figure S34. ¹H NMR spectrum of TPP-3 in DMSO-*d*₆.

Figure S36. HR-MS spectrum of TPP-3.

Figure S37. ¹H-¹H COSY NMR spectra of (A) TPP-1, (B) TPP-2, and (C) TPP-3 in DMSO- d_{6} .

2. PL spectra of TPP-1~3

Figure S38. PL spectra of DMSO solution of (A) TPP-1, (B) TPP-2 and (C) TPP-3 at different time. Excitation wavelengths: $\lambda_{ex} = 460$ nm for TPP-1, 520 nm for TPP-2, and 500 nm for TPP-3. [TPP-1] = [TPP-2] = [TPP-3] = 1.0×10^{-5} mol/L.

Figure S39. (A) PL spectra of organic solutions of TPP-1~3 in DMSO ([TPP-1] = [TPP-2] = $[TPP-3] = 1.0 \times 10^{-5} \text{ mol/L}$). (B) PL spectra of TPP-1~3 in solid. Excitation wavelength: $\lambda_{ex} = 500 \text{ nm}$.

Figure S40. PL spectra of (A) TPP-1, (B) TPP-2 and (C) TPP-3 in the mixture of DMSO/H₂O with different water fractions (f_w). Excitation wavelengths: $\lambda_{ex} = 460$ nm for TPP-1, 520 nm for TPP-2, and 500 nm for TPP-3. [TPP-1] = [TPP-2] = [TPP-3] = 1.0 × 10⁻⁵ mol/L.

3. Particle diameter size distributions

Figure S41. Particle diameter size distributions of (A) TPP-1 in DMSO, (B) TPP-1 in DMSO/H₂O (1:9), (C) TPP-1 in DMSO/H₂O (1:99), (D) TPP-2 in DMSO, (E) TPP-2 in DMSO/H₂O (3:7), (F) TPP-3 in DMSO, (G) TPP-3 in DMSO/H₂O (4:6). [TPP-1] = [TPP-2] = [TPP-3] = 1.0×10^{-5} mol/L.

4. UV-visible spectral changes on addition of H₂O

Figure S42. Absorption spectra of (A) TPP-1, (B) TPP-2 and (C) TPP-3 in the mixture of DMSO/H₂O with different f_{w} . [TPP-1] = [TPP-2] = [TPP-3] = 1.0 × 10⁻⁵ mol/L.

5. SEM images of the aggregates in the mixture of DMSO/H₂O

Figure S43. SEM images of the aggregates of (A) TPP-1, (B) TPP-2 and (C) TPP-3. The aggregates were formed in the mixture of DMSO/H₂O (1:99 for TPP-1, 3:7 for TPP-2, 4:6 for TPP-3) and then filtered by pure carbon coated grids and dried in vacuum at room temperature.

6. Color changes in the molecular and aggregate state

Figure S44. Photographs of fluorescent (A) TPP-1, (B) TPP-2 and (C) TPP-3 in the mixture of DMSO/H₂O with different f_w under UV radiation at 365 nm. [TPP-1] = [TPP-2] = [TPP-3] = 1.0 × 10⁻⁵ mol/L.

7. The simulated molecular conformation

Figure S45. The optimized molecular conformation of (A) TPP-1, (B) TPP-2 and (C) TPP-3 by using the B3LYP/6-31G basis set with G09 program.

8. Fluorescence decay data and curves

Compounds	Solution (DMSO 10 ⁻⁵ M)							
	τ_1 (ns)	τ_2 (ns)	\mathbf{B}_1	B ₂	χ2	$\tau_{ave} (ns)$		
TPP-1	0.98	1.34	0.80	0.20	1.58	1.05		
TPP-2	-	-	-	-	-	-		
TPP-3	-	-	-	-	-	-		

Table S1. The original lifetime data of TPP-1~3 in solution

Table S2. The original lifetime data of TPP-1~3 in aggregation and solid state

	Aggregation					Solid						
Compounds	τ_1	τ_2	B1	Ba	~?	τ_{ave}	τ_1	τ_2	B1	Ba	~2	τ_{ave}
	(ns)	(ns)	D	\mathbf{D}_2	χ2	(ns)	(ns)	(ns)	$\mathbf{D}_{\mathbf{I}}$	D 2	λ2	(ns)
TPP-1	0.58	2.08	0.89	0.11	1.73	0.75	0.78	2.99	0.77	0.23	1.67	1.30
TPP-2	0.52	2.31	0.92	0.08	1.85	0.65	0.63	2.04	0.82	0.18	1.90	0.89
TPP-3	0.74	3.60	0.82	0.18	1.78	1.25	1.06	2.88	0.77	0.23	1.61	1.49

Figure S46. The fluorescence decay curves of TPP-1, TPP-2 and TPP-3 in aggregated state (A) and in solid (B). The aggregates were obtained from the mixture of DMSO and water at f_w = 99 vol% for TPP-1, f_w = 70 vol% for TPP-2, and f_w = 60 vol% for TPP-3 and [TPP-1] = [TPP-2] = [TPP-3] = 1.0 × 10⁻⁵ mol/L.

9. Cell apoptosis test

HeLa cells were stained with 1.0×10^{-7} mol/L of TPP-1, TPP-2 and TPP-3 solution for 12 h, 24 h, and 48 h, respectively. Cell apoptosis was measured by Annexin V-FITC/PI Apoptosis Detection Kit. Cells were digested with 0.05% Trypsin and stained with annexin V-FITC and propidium iodide in the dark at room temperature according to the manufacturer's instructions. After treatment at a given time, 400µL 1 x Annexin V binding solution was added into each sample. Apoptotic cells were measured by a flow cytometer.

Figure S47. The apoptosis assay of (A) TPP-1, (B) TPP-2 and (C) TPP-3 evaluated on HeLa cells. $[TPP-1] = [TPP-2] = [TPP-3] = 1.0 \times 10^{-7} \text{ mol/L}.$

10. The test of amount diffused in HeLa cells

Cells were cultured after overnight culture in a humidified incubator at 37°C with 5% CO₂. 1.0×10^{-6} mol/L of TPP-1, TPP-2 and TPP-3 (1 mL) was added into Hela cells with 9 mL of DMEM. 1.0×10^{-6} mol/L of TPP-1, TPP-2 and TPP-3 (1 mL) was added into 9 mL DMEM without Hela cells as control group. After 15 min incubation, all solutions were taken out and the UV tests were measured.

Figure S48. The amount of TPP-1, TPP-2 and TPP-3 in HeLa cells after 15 min incubation in the same concentration at $[TPP-1] = [TPP-2] = [TPP-3] = 1.0 \times 10^{-7} \text{ mol/L}.$

Figure S49. Particle diameter size distributions of (A) DMEM, (B) TPP-1 in DMEM, (C)

TPP-2 in DMEM, (D) TPP-3 in DMEM at [TPP-1] = [TPP-2] = [TPP-3] = 1.0×10^{-7} mol/L.

Figure S50. PL spectra of (A) TPP-1, (B) TPP-2 and (C) TPP-3 in DMSO/ aqueous solution of NaCl (1:99) with different NaCl contents (*wt*%). Excitation wavelengths: $\lambda_{ex} = 460$ nm for

TPP-1, 520 nm for TPP-2, and 500 nm for TPP-3. [TPP-1] = [TPP-2] = [TPP-3] = 1.0×10^{-7} mol/L.

11. Confocal fluorescent images of TPP-1, TPP-2 and TPP-3 in living cells

HeLa, MCF-7, HepG2 and NIH3T3 cells were seeded in $\Phi 20$ mm glass bottom cell culture dishes (7.2 × 10⁵ ± 0.05 × 10⁵ cells in each dish). After overnight culture in a humidified incubator at 37 °C with 5% CO₂, culture medium was removed and cells were stained with 1.0 × 10⁻⁷ mol/L of TPP-1, TPP-2 and TPP-3 for 5 min in DMSO/DMEM (1‰ DMSO) solution at 37 °C. Before imaging, no washing processed were needed. The fluorescence imaging of TPP-1, TPP-2 and TPP-3 in living cells was recorded with a Leica TCS SP5 laser scanning confocal microscope.

Figure S51. Fluorescent images of TPP-1, TPP-2 and TPP-3 in HeLa cells after 5 min incubation in the same concentration at [TPP-1] = [TPP-2] = [TPP-3] = 1.0×10^{-7} mol/L.

Figure S52. Fluorescent images of TPP-1, TPP-2 and TPP-3 in HepG2 cells after 5 min incubation in the same condition at [TPP-1] = [TPP-2] = [TPP-3] = 1.0×10^{-7} mol/L.

Figure S53. Fluorescent images of TPP-1, TPP-2 and TPP-3 in MCF-7 cells after 5 min incubation in the same condition at [TPP-1] = [TPP-2] = [TPP-3] = 1.0×10^{-7} mol/L.

Figure S54. Fluorescent images of TPP-1, TPP-2 and TPP-3 in NIH 3T3 cells after 5 min incubation in the same condition at [TPP-1] = [TPP-2] = [TPP-3] = 1.0×10^{-7} mol/L.

12. Photostability of TPP-2

HeLa Cells were incubated in 5% CO₂ and 1.0×10^{-7} mol/L of TPP-2 at 37 °C for 5 min. In colocalization experiments, the cells were first washed with PBS, and then incubated with 1.0 $\times 10^{-7}$ mol/L of MitoTracker Red at 37 °C for 30 min. The medium was removed and the cells were rinsed with PBS. The cells were imaged immediately and at an interval by using Leica TCS SP5 laser scanning confocal microscope.

Figure S55. Fluorescent images of MitoTracker Red and TPP-2 after 6 h continuous irradiation. The concentration of MitoTracker Red and TPP-2 were both 1.0×10^{-7} mol/L.

13. References

- [1] T. Han, X. Feng, B. Tong, J. Shi, L. Chen, J. Zhi, Y. Dong, *Chem. Commun.* 2012, 48, 416.
- [2] N. Zhao, S. Chen, Y. Hong, B. Z. Tang, Chem. Commun. 2015, 51, 13599.
- [3] G. Liu, D. Chen, L. Kong, J. Shi, B. Tong, J. Zhi, X. Feng, Y. Dong, *Chem. Commun.* 2015, 51, 8555.