Supporting Information

Simple and Sensitive Colorimetric Detection of Trace Amount of 2,4,6-

trinitrotoluene (TNT) with QD multilayers-modified micro-channel assays

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Part S1 Experimental section

Synthesis of MPA-capped CaTe QDs and L-cysteine-capped CdTe QDs

CdTe NPs were prepared by reacting Cd²⁺ and H₂Te gas in the presence of stabilizers according to the Rogach-Weller method^{S1} with slight modification.^{S2-S3} A 0.498 g (0.59 mmol) sample of Cd(ClO₄)₂·6H₂O and 0.498g (0.14 mmol) of L-cysteine or 0.012 mL (0.14 mmol) 3-mercaptopropionic acid (MPA) was dissolved in 62 mL distilled water. The pH was then adjusted to 11.2 by adding 1 M NaOH. The solution was placed in the three-neck flask and deaerated by N₂ gas for about 30 min. H₂Te gas generated by the mixture of 0.2 g of Al₂Te₃ and 15 mL of 0.5 M H₂SO₄ in another threeneck flask was directed into the prepared solution under a N2 atmosphere. The precursors were converted to CdTe nanocrystals by refluxing the reaction mixture at 100 °C for several hours in N2 gas to obtain appropriate size. Figure S1 shows typical absorption and photoluminescence spectra of CdTe QDs. The green QDs shows a fluorescence maximum at 520 nm and red QDs show a fluorescence peak at 620 nm, respectively. The sharp absorbance and photoluminescence peaks indicate the relatively narrow size distribution of both QDs samples. The size of CdTe QDs could be calculated through the wavelength of the first excitonic absorption peak of the QDs. The average particle size of above-mentioned CdTe QDs was estimated to be 1.69 nm (green QDs) and 3.47 nm (red QDs) with the UV-Vis spectrum according to the empirical equations proposed by Peng and co-workers,^{S4} respectively. These results are consistent with that obtained from typical TEM image of the CdTe QDs (Figure S2).



Figure S1. UV-vis absorption spectra and fluorescence spectra of green QDs(a) and red QDs(b).



Figure S2 TEM images of green QDs and red QDs.

Fabrication of QD multilayers-coated microfluidic chips

Glass substrates and microfluidic chips were used as substrates. Glass substrates, and glass substrates of microfluidic chips was cleaned with Piranha solution $(H_2O_2:H_2SO_4=3:1)$ at 70 °C,^{S5} followed by washed by deionized water and dried under N₂ flow.

QD multilayers-coated glass substrates were fabricated by a typical layer-by-layer (LbL) self-assembly method.^{S6-S7} Firstly, the substrates were alternatively immersed into 1mg mL⁻¹ positively-charged PEI solutions (pH 7.4, 1 M NaCl) and 1mg mL⁻¹ negatively-charged PSS solutions (pH 7.4, 1 M NaCl) to form (PEI/PSS)₃ multilayerscoated glass substrates, using an immersion time of 5min, and rinsed with pure water and dried under N₂ flow after each layer deposited. Secondly, ten PAH/green QDs bilayers were alternatively deposited onto (PEI/PSS)₃ multilayers-coated glass substrates from 1mg mL⁻¹ positively-charged PAH (pH 7.4, 1 M NaCl) and green fluorescent MPA-capped CdTe QDs solution to obtain (PEI/PSS)₃(PAH/green QDs)₁₀ bilayers of PAH/PSS multilayers. Thirdly, three were deposited onto (PEI/PSS)₃(PAH/green QDs)₁₀ multilayers from 1mg mL⁻¹ positively-charged PAH solutions (pH 7.4, 1 M NaCl) and 1mg/mL negative-charged PSS solutions (pH 7.4, 1 M NaCl). Finally, five bilayers of PAH/Red QDs were deposited onto (PEI/PSS)₃(PAH/green QDs)₁₀(PAH/PSS)₃ multilayers from 1mg/mL positivelycharged PAH (pH 7.4, without NaCl) and red fluorescent L-cysteine-capped CdTe QDs solution and one more PAH was deposited at the outside of the sensor film. The configuration of the multilayer was (PEI/PSS)₃(PAH/green QDs)₁₀(PAH/PSS)₃ (PAH/red QDs)₅PAH.

A PDMS microfluidic chips used in this work had a simple layout. Microfluidic channels were formed in PDMS by a photolithography technique.^{S8-S9} The microfluidic chips had brief channels geometry (5 parallel lines, 50 µm wide, 30 µm deep, 3 cm long), then this chips was followed by alternate deposition of polyelectrolytes and QDs

to form the hybrid film sensor in the microfluidic channels following the same procedure as QD multilayers-coated glass substrates. The micro channels were first filled with sample solution using syringe pump and allowed to incubate (no flow) for 10 min. The channels were then rinsed with water and dried. As same, the configuration of film in micro channels were also (PEI/PSS)₃(PAH/green QDs)₁₀(PAH/PSS)₃ (PAH/red QDs)₅PAH.



Part S2 Characterization of QD multilayers-coated glass substrates and microfluidic chips

Figure S3. UV-vis (a) and fluorescent (b) spectra of QD multilayers deposited on glass substrates (PEI/PSS)₃(PAH/green during LBL assembly process: $QDs)_2$ (blank solid line), (PEI/PSS)₃(PAH/green QDs)₄ (red solid line), (PEI/PSS)₆(PAH/green QDs)₆ (blue solid line), (PEI/PSS)₃(PAH/green QDs)₈ (Dark Cyan solid line), (PEI/PSS)₃ (PAH/green QDs)₁₀ (Magenta solid line), (PEI/PSS)₃(PAH/green QDs)₁₀(PAH/PSS)₃ (PAH/red QDs)₁ (Dark Yellow Dash line), (PEI/PSS)₃(PAH/green QDs)₁₀(PAH/PSS)₃ (PAH/red QDs)₂ line), (Navy Dash (PEI/PSS)₃(PAH/green QDs)₁₀(PAH/PSS)₃ (PAH/red QDs)₃ (Wine Dash line), (PEI/PSS)₃(PAH/green QDs)₁₀(PAH/PSS)₃ (PAH/red QDs)₄ (Pink Dash line), (PEI/PSS)₃(PAH/green QDs)₁₀(PAH/PSS)₃ (PAH/red QDs)₅ (Olive Dash line).

The FRET efficiency of $(PEI/PSS)_3(PAH/green QDs)_{10}(PAH/PSS)_3(PAH/red QDs)_y$ increases with *y*, and can be calculated according to equation:^{S10} E=1-F_{DA}/F_D

Where F_{DA} is the 520nm QD fluorescence of (PEI/PSS)₃(PAH/green QDs)₁₀(PAH/PSS)₃ in the presence of (PAH/red QDs)_y, and F_D is the 520nm QD fluorescence of (PEI/PSS)₃(PAH/green QDs)₁₀(PAH/PSS)₃ in the absence of (PAH/red QDs)_y. The FRET efficiency of (PEI/PSS)₃(PAH/green QDs)₁₀(PAH/PSS)₃(PAH/green QDs)₁₀(PAH/PSS)₃(PAH/red QDs)_y is 11.7%, 21.6%, 31.7%, 42.4% and 45.6% for y=1~5, respectively.



Fig. S4 AFM micrograph (tapping mode) (a) and cross-sectional analysis (b) of the QD multilayerscoated film on glass.



Fig. S5 FL spectra of different concentrations of TNT with 365 nm as excitation wavelength.



Figure S6¹³C-NMR spectrum of 2,4,6-trinitrotoluene (TNT) (2 mM) (a), cysteine (6 mM) (b), and

the mixture of TNT and cysteine (c) in 1/1 (by vol.) H_2O/d_6 -acetone at room temperature.



Figure S7. Energy level diagram of green CdTe QDs and the reduction potential of TNT.



Figure S8. Photo images of QD multilayers-coated microfluidic chips.

Part S3 Performance of QD multilayers-coated microfluidic chips for TNT detection



Figure S9. Stability of QD multilayers-coated microfluidic chips.



Figure S10. Response of (PEI/PSS)₃(PAH/green QDs)₁₀(PAH/PSS)₃(PAH/red QDs)₅ multilayers-

based microchannel assays toward different common ions.



Figure S11. Response of (PEI/PSS)₃(PAH/green QDs)₁₀(PAH/PSS)₃(PAH/red QDs)₅ multilayersbased microchannel assays toward different TNT analogs spiked in Jiulong Lake water. The concentration of TNT analogs is 10⁵ ppt.

Detection method	LOD	Linear range	Detection time	Reference
Fluorescence of MIP-capped	6.16×10 ⁴ ppt	1.76×10 ⁵ - 6.6×10 ⁷	5 mins	S11
CdSe QDs		ppt		
Fluorescence of amine-				
Capped ZnS-Mn ²⁺ QDs	200 ppt	-	5 mins	S12
Fluorescence of creatinine-	3.37×10 ³ ppt	10 ⁴ - 3×10 ⁵ ppt	5 mins	S13
capped CdSe/ZnS QDs				
Fluorescence of MoO _x QDs	2.64×10 ³ ppt	1.1×10 ⁴ - 5.3×10 ⁵ ppt	5 mins	S14
Fluorescence of histidine-	4.1×10 ⁴ ppt	4.1×10 ⁴ -10 ⁷ ppt	60 mins	S15
capped CdSe/ZnS QDs				
Fluorescence of L-cysteine-	400 ppt	-	120 mins	S16
capped CdTe QDs				
Fluorescence of L-cysteine-				
capped Co ²⁺ dopped ZnS	2.5×10 ³ ppt	2.5×10 ³ - 1.1×10 ⁵ ppt	5 mins	S17
QDs				
FRET between				
QDs/Antibody Fragment	$2 \times 10^4 \text{ ppt}$	9.8×10 ³ - 10 ⁷ ppt	10 mins	S18
hybrid				
FRET between gold	22 ppt	22 - 13.2×10 ³ ppt	10 mins	S19
nanorod-CdTe/CdS QDs				

 Table S1 Comparison the proposed method and other TNT sensors.

4×10 ³ ppt	4×10 ³ - 10 ⁴ ppt	3 mins	S20
1.1×105 mmt	9.8×10^{7} 9.8×10^{8} mmt	10 mins	\$21
1.1×10° ppt	8.8×10 [°] - 8.8×10 [°] ppt	10 mins	521
7.5×10 ³ ppt	2.2×10 ³ -1.7×10 ⁶ ppt	10 mins	S22
5.24 ppt	10 - 10 ⁷ ppt	5 mins	This study
	4×10 ³ ppt 1.1×10 ⁵ ppt 7.5×10 ³ ppt 5.24 ppt	 4×10³ ppt 4×10³ - 10⁴ppt 1.1×10⁵ ppt 8.8×10⁷ - 8.8×10⁸ ppt 7.5×10³ ppt 2.2×10³ - 1.7×10⁶ ppt 5.24 ppt 10 - 10⁷ ppt 	 4×10³ ppt 4×10³ - 10⁴ppt 3 mins 1.1×10⁵ ppt 8.8×10⁷ - 8.8×10⁸ ppt 10 mins 7.5×10³ ppt 2.2×10³ - 1.7×10⁶ ppt 10 mins 5.24 ppt 10 - 10⁷ ppt 5 mins

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