

## Supplementary Information

### Hydrofunctionalization of Alkenols Triggered by Addition of Diverse Radicals to Unactivated Alkenes and Subsequent Remote Hydrogen Atom Translocation

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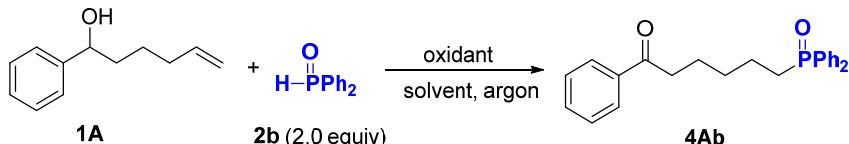
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## **General information.**

All reactions were carried out under argon using Schlenk techniques. Reagents were purchased at the commercial quality and used without further purification. Analytical thin layer chromatography (TLC) was performed on precoated silica gel 60 GF254 plates. Flash column chromatography was performed using Tsingdao silica gel (60, particle size 0.040-0.063 mm). Visualization on TLC was achieved by use of UV light (254 nm) or iodine. NMR spectra were recorded on a Bruker DPX 400 spectrometer at 400 MHz for <sup>1</sup>H NMR, 100 MHz for <sup>13</sup>C NMR and 376 MHz for <sup>19</sup>F NMR in CDCl<sub>3</sub> with tetramethylsilane (TMS) as internal standard. Microwave irradiation experiments were carried out in a dedicated Biotage Initiator Robot 8 auto microwave apparatus. The chemical shifts are expressed in ppm and coupling constants are given in Hz. Data for <sup>1</sup>H NMR are recorded as follows: chemical shift (ppm), multiplicity (s, singlet; d, doublet; t, triplet; q, quarter; m, multiplet), coupling constant (Hz), integration. Data for <sup>13</sup>C NMR are reported in terms of chemical shift ( $\delta$ , ppm). <sup>19</sup>F NMR spectra were recorded on a Bruker DPX 400 MHz spectrometer (CFCl<sub>3</sub> as an external reference (0 ppm)). Mass spectrometric data were obtained using Bruker Apex IV RTMS.

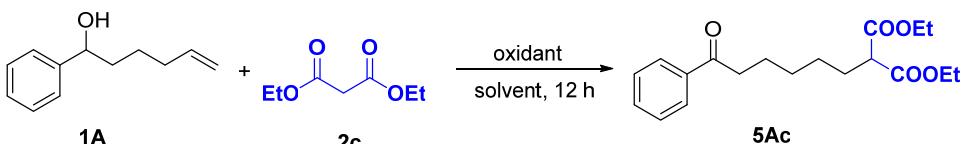
**Supplementary Table S1.** Screening Results of Reaction Conditions for Phosphonylation<sup>a</sup>



entry	Oxidant ( X equiv)	T (°C)	solvent	yield (%) <sup>b</sup>
1	CuCl (2)	80	CH <sub>3</sub> CN	No reaction
2	K <sub>2</sub> S <sub>2</sub> O <sub>8</sub> (2)	80	CH <sub>3</sub> CN/H <sub>2</sub> O (2/1)	68% (58%) <sup>c</sup>
3	K <sub>2</sub> S <sub>2</sub> O <sub>8</sub> (2)	80	CH <sub>3</sub> CN/H <sub>2</sub> O (2/1)	68% <sup>d</sup>
4	K <sub>2</sub> S <sub>2</sub> O <sub>8</sub> (2)	80	CH <sub>3</sub> CN/H <sub>2</sub> O (2/1)	60% <sup>e</sup>
5	AgOAc (3)	100	DMF	100% (90%) <sup>c</sup>
6	AgNO <sub>3</sub> (0.5)	100	EtOAc	63%
7	AgNO <sub>3</sub> (0.5)	100	CH <sub>3</sub> CN	65%
8	AgNO <sub>3</sub> (0.5)	100	Toluene	65%
9	AgNO <sub>3</sub> (0.5)	100	DMF	60%
10	AgNO <sub>3</sub> (1)	100	CH <sub>3</sub> CN	70%

<sup>a</sup> Reaction conditions: **1A** (0.2 mmol), **2b** (0.4 mmol), solvent (2.0 mL) at 80 °C for 12 h under argon. <sup>b</sup> Determined by NMR spectroscopy using 1,3,5-trimethylbenzene as an internal standard. <sup>c</sup> Isolated yield in bracket. <sup>d</sup> Ag<sub>2</sub>SO<sub>4</sub> (0.02 mmol) was added. <sup>e</sup> Shortening the reaction time to 5 h.

**Supplementary Table S2.** Screening Results of Reaction Conditions for Addition of Diethyl Malonate to Alkene<sup>a</sup>

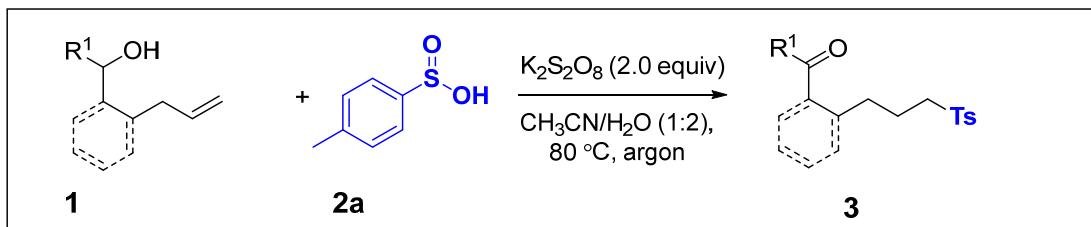


entry	Oxidant (X equiv)	2c (X equiv)	Solvent/T(°C)	yield (%) <sup>b</sup>
1	Cu(OAc) <sub>2</sub> (2)	1.5	TFE <sup>c</sup> (100)	No reaction
2	Ag <sub>2</sub> CO <sub>3</sub> (2)	3.0	DMF (80)	40% conversion 45% yield
3	K <sub>2</sub> S <sub>2</sub> O <sub>8</sub> (2)	1.5	CH <sub>3</sub> CN/H <sub>2</sub> O (80)	trace
4	Mn(OAc) <sub>3</sub> ·2H <sub>2</sub> O (3)	1.5	AcOH (80)	100% conversion 14% yield
5	Mn(OAc) <sub>3</sub> ·2H <sub>2</sub> O (3)	1.5	TFE (80)	60% conversion 34% yield
6	Mn(OAc) <sub>3</sub> ·2H <sub>2</sub> O (3)	1.5	TFE (110) <sup>d</sup>	74% conversion 57% yield
7	Mn(OAc) <sub>3</sub> ·2H <sub>2</sub> O (3.6)	1.8	TFE (110) <sup>d</sup>	100% conversion 50% yield

<sup>a</sup> Reaction conditions: **1A** (0.2 mmol), **2c**, solvent (2.0 mL) was heated for 16 h under argon. <sup>b</sup> Isolated yield. <sup>c</sup> TFE: 2,2,2-trifluoroethanol. <sup>d</sup> reaction time: 36 h.

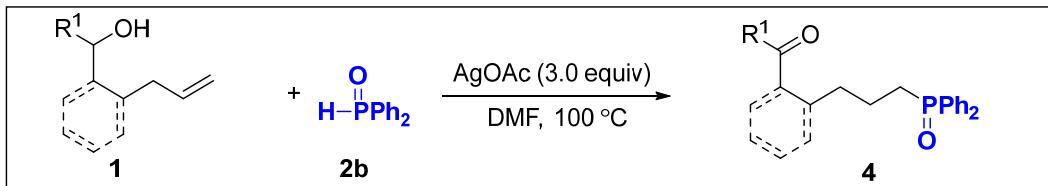
## General procedures

### General procedure for radical sulfonylation reaction system



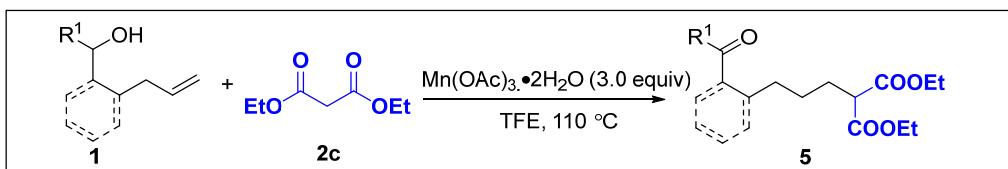
To a flame-dried Schlenk tube equipped with a magnetic stir bar were added **1** (0.2 mmol), **2a** (0.6 mmol) and  $\text{K}_2\text{S}_2\text{O}_8$  (0.4 mmol). The tube was evacuated and backfilled with argon for three times, and then  $\text{CH}_3\text{CN}$  (0.7 mL) and  $\text{H}_2\text{O}$  (1.4 mL) were added. The tube was stirred at  $80^\circ\text{C}$  for 18 h and then  $\text{H}_2\text{O}$  (5 mL) was added.  $\text{EtOAc}$  was used to extract the product from the aqueous layer ( $3 \times 20$  mL). The combined organic layer was dried over anhydrous  $\text{Na}_2\text{SO}_4$ , filtered and concentrated to afford the crude product, which was purified by flash column chromatography to afford the product **3**.

### General procedures for radical phosphorylation reaction system



To a flame-dried Schlenk tube equipped with a magnetic stir bar were added **1** (0.2 mmol), **2b** (0.4 mmol) and  $\text{AgOAc}$  (0.60 mmol). The tube was evacuated and backfilled with argon for three times, and then DMF (2 mL) were added. The tube was stirred at  $80^\circ\text{C}$  for 12 h. Diethyl ether (50 mL) was added, followed by washing with water ( $2 \times 5$  mL). The organic layer was dried over anhydrous  $\text{Na}_2\text{SO}_4$ , filtered and concentrated to afford the crude product, which was purified by flash column chromatography to afford the product **4**.

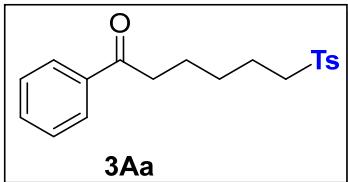
### General procedure for radical reaction with diethyl malonate



To a flame-dried Schlenk tube equipped with a magnetic stir bar were added **1** (0.2 mmol), diethyl malonate **2c** (0.3 mmol) and  $\text{Mn}(\text{OAc})_3 \cdot 2\text{H}_2\text{O}$  (0.6 mmol). The tube was evacuated and backfilled with argon for three times, and then 2,2,2-trifluoroethanol (2.0 mL) were added. The tube was stirred at  $110^\circ\text{C}$  for 36 h

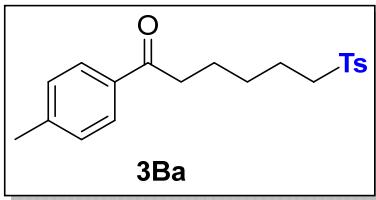
and then H<sub>2</sub>O (5 mL) was added. EtOAc was used to extract the product from the aqueous layer (3 × 20 mL). The combined organic layer was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated to afford the crude product, which was purified by flash column chromatography to afford the product **5**.

### **1-phenyl-6-tosylhexan-1-one (3Aa)**



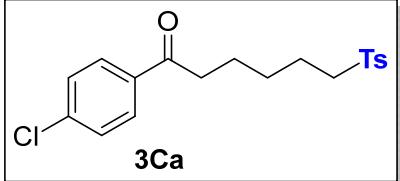
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.00 – 7.90 (m, 2H), 7.79 (d, *J* = 8.3 Hz, 2H), 7.66 – 7.54 (m, 1H), 7.47 (t, *J* = 7.6 Hz, 2H), 7.37 (d, *J* = 8.0 Hz, 2H), 3.10 (dd, *J* = 9.1, 6.9 Hz, 2H), 2.96 (t, *J* = 7.2 Hz, 2H), 2.46 (s, 3H), 1.91 – 1.57 (m, 4H), 1.51 – 1.43 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 199.78, 144.66, 136.84, 136.15, 133.10, 129.92, 128.63, 128.08, 127.99, 56.17, 37.93, 27.90, 23.45, 22.70, 21.64. HRMS (APCI) m/z calcd. for C<sub>19</sub>H<sub>23</sub>O<sub>3</sub>S [M+H]<sup>+</sup> 331.1368, found 331.1363.

### **1-(*p*-tolyl)-6-tosylhexan-1-one (3Ba)**



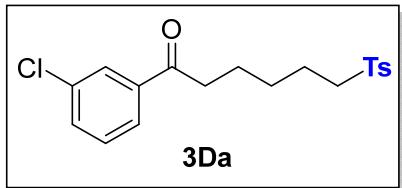
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.83 (d, *J* = 8.2 Hz, 2H), 7.79 (d, *J* = 8.2 Hz, 2H), 7.36 (d, *J* = 8.0 Hz, 2H), 7.25 (d, *J* = 8.0 Hz, 2H), 3.09 (dd, *J* = 9.1, 6.9 Hz, 2H), 2.92 (t, *J* = 7.2 Hz, 2H), 2.45 (s, 3H), 2.41 (s, 3H), 1.81 – 1.67 (m, 4H), 1.51 – 1.43 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 199.44, 144.63, 143.84, 136.19, 134.40, 129.91, 129.28, 128.11, 128.07, 56.17, 37.81, 27.91, 23.55, 22.69, 21.62. HRMS (APCI) m/z calcd. for C<sub>20</sub>H<sub>25</sub>O<sub>3</sub>S [M+H]<sup>+</sup> 345.1524, found 345.1521.

### **1-(4-chlorophenyl)-6-tosylhexan-1-one (3Ca)**



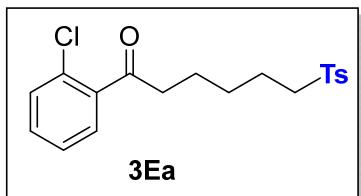
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.87 (d, *J* = 8.3 Hz, 2H), 7.78 (d, *J* = 8.0 Hz, 2H), 7.43 (d, *J* = 8.3 Hz, 2H), 7.36 (d, *J* = 8.0 Hz, 2H), 3.13 – 3.05 (m, 2H), 2.92 (t, *J* = 7.1 Hz, 2H), 2.45 (s, 3H), 1.80 – 1.67 (m, 4H), 1.51 – 1.43 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 198.50, 144.69, 139.49, 136.14, 135.13, 129.92, 129.42, 128.96, 128.06, 56.13, 37.91, 27.83, 23.34, 22.65, 21.64. HRMS (APCI) m/z calcd. for C<sub>19</sub>H<sub>22</sub>ClO<sub>3</sub>S [M+H]<sup>+</sup> 365.0978, found 365.0973.

**1-(3-chlorophenyl)-6-tosylhexan-1-one (3Da)**



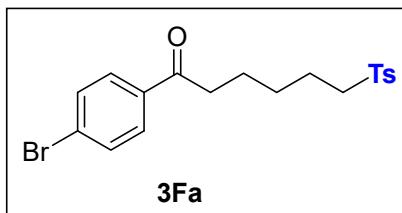
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.78 (d, *J* = 8.3 Hz, 2H), 7.42 – 7.29 (m, 6H), 3.08 (dd, *J* = 9.1, 6.9 Hz, 2H), 2.90 (t, *J* = 7.2 Hz, 2H), 2.45 (s, 3H), 1.78 – 1.64 (m, 4H), 1.50 – 1.40 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 203.11, 144.69, 139.42, 136.10, 131.69, 130.68, 130.51, 129.93, 128.71, 128.07, 126.99, 56.11, 42.35, 27.71, 23.39, 22.64, 21.64. HRMS (APCI) m/z calcd. for C<sub>19</sub>H<sub>22</sub>ClO<sub>3</sub>S [M+H]<sup>+</sup> 365.0978, found 365.0971.

**1-(2-chlorophenyl)-6-tosylhexan-1-one (3Ea):**



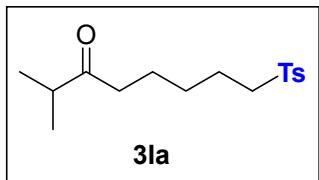
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.90 (t, *J* = 1.8 Hz, 1H), 7.82 – 7.78 (m, 3H), 7.54 (ddd, *J* = 8.0, 2.1, 1.0 Hz, 1H), 7.41 (t, *J* = 7.9 Hz, 1H), 7.37 (d, *J* = 7.9 Hz, 2H), 3.12 – 3.07 (m, 2H), 2.93 (t, *J* = 7.1 Hz, 2H), 2.46 (s, 3H), 1.81 – 1.68 (m, 4H), 1.51 – 1.42 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 198.39, 144.69, 138.36, 136.16, 134.90, 133.09, 129.93, 129.90, 128.17, 128.08, 126.08, 56.13, 38.05, 27.81, 23.26, 22.66, 21.64. HRMS (APCI) m/z calcd. for C<sub>19</sub>H<sub>22</sub>ClO<sub>3</sub>S [M+H]<sup>+</sup> 365.0978, found 365.0970.

**1-(4-bromophenyl)-6-tosylhexan-1-one (3Fa):**



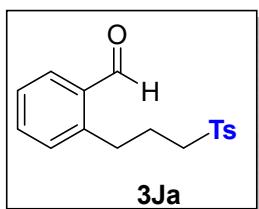
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.85 – 7.72 (m, 4H), 7.58 (d, *J* = 8.6 Hz, 2H), 7.35 (d, *J* = 8.0 Hz, 2H), 3.08 (t, *J* = 6.9 Hz, 2H), 2.90 (t, *J* = 7.2 Hz, 2H), 2.44 (s, 3H), 1.79 – 1.66 (m, 4H), 1.49 – 1.43 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 198.68, 144.69, 136.18, 135.55, 131.93, 129.90, 129.57, 128.16, 128.04, 56.13, 37.84, 27.81, 23.32, 22.66, 21.64. HRMS (APCI) m/z calcd. for C<sub>19</sub>H<sub>22</sub>BrO<sub>3</sub>S [M+H]<sup>+</sup> 409.0473, found 409.0467.

**2-methyl-8-tosyloctan-3-one (3Ia):**



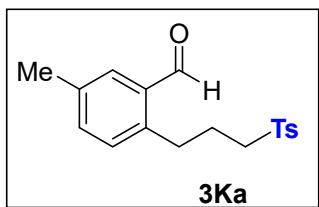
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.79 (d, *J* = 8.0 Hz, 2H), 7.38 (d, *J* = 7.9 Hz, 2H), 3.11 – 3.04 (m, 2H), 2.61 – 2.51 (m, 1H), 2.47 (s, 3H), 2.43 (t, *J* = 7.1 Hz, 2H), 1.78 – 1.68 (m, 2H), 1.60 – 1.50 (m, 2H), 1.41 – 1.32 (m, 2H), 1.08 (d, *J* = 6.9 Hz, 6H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 214.33, 144.63, 136.17, 129.89, 128.06, 56.15, 40.87, 39.61, 27.84, 22.95, 22.62, 21.63, 18.23. HRMS (APCI) m/z calcd. for C<sub>16</sub>H<sub>25</sub>O<sub>3</sub>S [M+H]<sup>+</sup> 297.1510, found 297.1519.

#### 2-(3-tosylpropyl)benzaldehyde (3Ja):



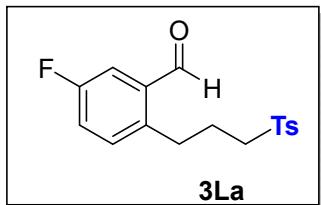
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 10.09 (s, 1H), 7.80 – 7.74 (m, 3H), 7.51 (td, *J* = 7.5, 1.6 Hz, 1H), 7.42 (td, *J* = 7.5, 1.3 Hz, 1H), 7.37 – 7.32 (m, 2H), 7.25 (dd, *J* = 7.6, 1.2 Hz, 1H), 3.16 – 3.10 (m, 4H), 2.44 (s, 3H), 2.06 – 1.97 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 192.96, 144.77, 142.62, 136.17, 134.43, 133.96, 133.80, 131.33, 130.01, 128.13, 127.27, 55.73, 31.23, 24.51, 21.73. HRMS (APCI) m/z calcd. for C<sub>17</sub>H<sub>19</sub>O<sub>3</sub>S [M+H]<sup>+</sup> 303.1042, found 303.1049.

#### 5-methyl-2-(3-tosylpropyl)benzaldehyde (3Ka):



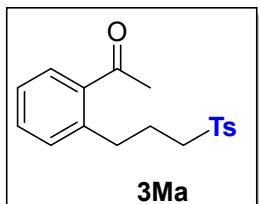
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 10.06 (s, 1H), 7.79 – 7.72 (m, 2H), 7.57 (d, *J* = 1.8 Hz, 1H), 7.37 – 7.28 (m, 3H), 7.13 (d, *J* = 7.7 Hz, 1H), 3.18 – 3.02 (m, 4H), 2.44 (s, 3H), 2.40 (s, 3H), 2.05 – 1.90 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 193.09, 144.75, 139.62, 137.06, 136.20, 134.83, 134.73, 133.65, 131.31, 130.00, 128.15, 55.74, 30.78, 24.61, 21.74, 20.87. HRMS (APCI) m/z calcd. for C<sub>18</sub>H<sub>21</sub>O<sub>3</sub>S [M+H]<sup>+</sup> 317.1198, found 317.1206.

#### 5-fluoro-2-(3-tosylpropyl)benzaldehyde (3La):



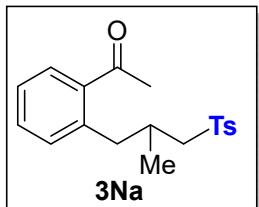
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 10.04 (s, 1H), 7.85 – 7.73 (m, 3H), 7.39 – 7.33 (m, 2H), 7.14 – 7.05 (m, 1H), 6.95 (dd, *J* = 9.5, 2.5 Hz, 1H), 3.20 – 3.09 (m, 4H), 2.45 (s, 3H), 2.08 – 1.95 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 191.09, 165.59 (d, *J* = 257.7 Hz), 146.13 (d, *J* = 8.8 Hz), 144.80, 137.02 (d, *J* = 10.1 Hz), 136.03, 129.97, 128.04, 118.20 (d, *J* = 21.7 Hz), 114.32 (d, *J* = 21.7 Hz), 55.50, 31.02, 24.15, 21.65. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) δ -102.99. HRMS (APCI) m/z calcd. for C<sub>17</sub>H<sub>18</sub>O<sub>3</sub>FS [M+H]<sup>+</sup> 321.0948, found 321.0955.

#### 1-(2-(3-tosylpropyl)phenyl)ethanone (3Ma):



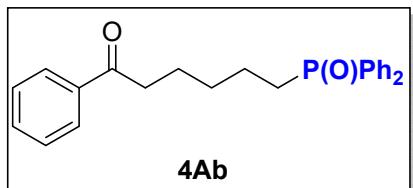
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.78 (d, *J* = 8.2 Hz, 2H), 7.73 – 7.69 (m, 1H), 7.42 (td, *J* = 7.5, 1.2 Hz, 1H), 7.37 – 7.28 (m, 3H), 7.23 (d, *J* = 7.6 Hz, 1H), 3.17 – 3.10 (m, 2H), 2.94 (t, *J* = 7.6 Hz, 2H), 2.55 (s, 3H), 2.45 (s, 3H), 2.06 – 1.96 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 201.47, 144.54, 140.72, 137.22, 136.21, 131.90, 131.45, 129.91, 129.87, 128.07, 127.98, 126.52, 55.82, 32.33, 29.58, 24.51, 21.63. HRMS (APCI) m/z calcd. for C<sub>18</sub>H<sub>21</sub>O<sub>3</sub>S [M+H]<sup>+</sup> 317.1211, found 317.1209.

#### 1-(2-(2-methyl-3-tosylpropyl)phenyl)ethan-1-one (3Na):



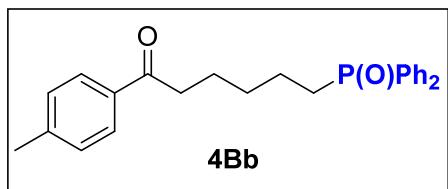
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.70 – 7.60 (m, 3H), 7.41 – 7.32 (m, 1H), 7.33 – 7.24 (m, 3H), 7.16 – 7.09 (m, 1H), 3.15 – 3.08 (m, 1H), 3.01 – 2.93 (m, 1H), 2.92 – 2.80 (m, 2H), 2.48 (s, 3H), 2.43 (s, 3H), 2.29 – 2.19 (m, 1H), 1.11 (d, *J* = 6.6 Hz, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 201.66, 144.29, 139.45, 137.74, 137.01, 132.11, 131.63, 129.88, 129.84, 127.91, 126.57, 61.49, 40.56, 30.89, 29.75, 21.70, 20.02. HRMS (APCI) m/z calcd. for C<sub>19</sub>H<sub>23</sub>O<sub>3</sub>S [M+H]<sup>+</sup> 331.1354, found 331.1362.

#### 6-(diphenylphosphoryl)-1-phenylhexan-1-one (4Ab):



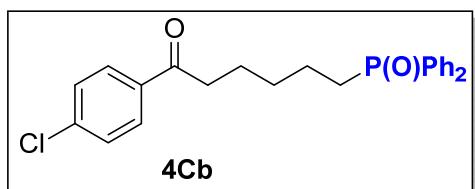
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.92 (d, *J* = 7.3 Hz, 2H), 7.78 – 7.70 (m, 5H), 7.54 – 7.43 (m, 7H), 7.38 (d, *J* = 6.7 Hz, 1H), 2.92 (t, *J* = 6.8 Hz, 2H), 2.28 (dd, *J* = 15.5, 10.4 Hz, 2H), 1.78 – 1.60 (m, 4H), 1.52 – 1.48 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 200.15, 136.93, 133.82 (d, *J<sub>C-P</sub>* = 97.7 Hz), 133.03, 131.74 (d, *J<sub>C-P</sub>* = 2.5 Hz), 130.86 (d, *J<sub>C-P</sub>* = 9.3 Hz), 128.76 (d, *J<sub>C-P</sub>* = 11.5 Hz), 128.52, 128.01, 38.08, 30.42 (d, *J<sub>C-P</sub>* = 14.6 Hz), 29.43 (d, *J<sub>C-P</sub>* = 71.3 Hz), 23.52, 21.21 (d, *J<sub>C-P</sub>* = 3.8 Hz). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>) δ 33.03. HRMS (APCI) m/z calcd. for C<sub>24</sub>H<sub>26</sub>O<sub>2</sub>P [M+H]<sup>+</sup> 377.1670, found 377.1667.

**6-(diphenylphosphoryl)-1-(*p*-tolyl)hexan-1-one (4Bb):**



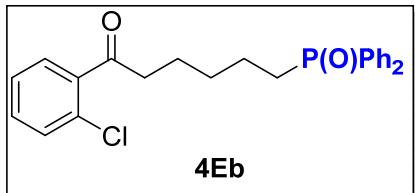
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.81 (d, *J* = 7.5 Hz, 2H), 7.78 – 7.68 (m, 4H), 7.50 – 7.43 (m, 6H), 7.22 (d, *J* = 7.3 Hz, 2H), 2.88 (t, *J* = 6.6 Hz, 2H), 2.38 (s, 3H), 2.32 – 2.26 (m, 2H), 1.72 – 1.65 (m, 4H), 1.51 – 1.47 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 199.83, 143.73, 134.44, 132.87 (d, *J<sub>C-P</sub>* = 98.0 Hz), 131.71 (d, *J<sub>C-P</sub>* = 2.4 Hz), 130.71 (d, *J<sub>C-P</sub>* = 9.7 Hz), 129.24, 128.72 (d, *J<sub>C-P</sub>* = 11.5 Hz), 128.13, 37.96, 30.40 (d, *J<sub>C-P</sub>* = 14.5 Hz), 29.42 (d, *J<sub>C-P</sub>* = 71.6 Hz), 23.62, 21.62, 21.21 (d, *J<sub>C-P</sub>* = 3.5 Hz). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>) δ 32.43. HRMS (APCI) m/z calcd. for C<sub>25</sub>H<sub>28</sub>O<sub>2</sub>P [M+H]<sup>+</sup> 391.1827, found 391.1823.

**1-(4-chlorophenyl)-6-(diphenylphosphoryl)hexan-1-one (4Cb):**



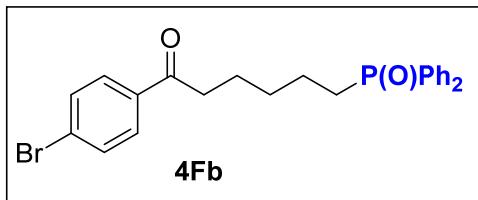
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.85 (d, *J* = 7.6 Hz, 2H), 7.75 – 7.71 (m, 4H), 7.55 – 7.36 (m, 8H), 2.89 (t, *J* = 7.1 Hz, 2H), 2.31 – 2.25 (m, 2H), 1.77 – 1.62 (m, 4H), 1.53 – 1.47 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 198.84, 139.33, 135.20, 132.92 (d, *J<sub>C-P</sub>* = 97.2 Hz), 131.72 (d, *J<sub>C-P</sub>* = 2.5 Hz), 130.71 (d, *J<sub>C-P</sub>* = 9.2 Hz), 129.44, 128.87, 128.75 (d, *J<sub>C-P</sub>* = 11.5 Hz), 38.02, 30.42 (d, *J<sub>C-P</sub>* = 14.3 Hz), 29.54 (d, *J<sub>C-P</sub>* = 71.7 Hz), 23.42, 21.25 (*J<sub>C-P</sub>* = 3.6 Hz). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>) δ 32.63. HRMS (APCI) m/z calcd. for C<sub>24</sub>H<sub>25</sub>ClO<sub>2</sub>P [M+H]<sup>+</sup> 411.1281, found 411.1274.

**1-(2-chlorophenyl)-6-(diphenylphosphoryl)hexan-1-one (4Eb):**



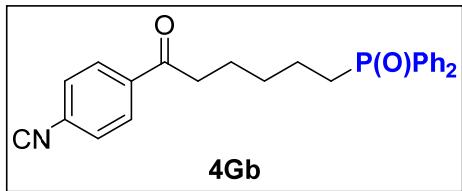
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.87 – 7.86 (m, 1H), 7.79 – 7.69 (m, 5H), 7.51 – 7.42 (m, 7H), 7.36 (t, *J* = 7.9 Hz, 1H), 2.88 (t, *J* = 7.2 Hz, 2H), 2.33 – 2.23 (m, 2H), 1.75 – 1.61 (m, 4H), 1.52 – 1.44 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 198.73, 138.41, 134.88, 132.90, 132.83 (d, *J<sub>C-P</sub>* = 97.7 Hz), 131.74 (d, *J<sub>C-P</sub>* = 2.5 Hz), 130.75 (d, *J<sub>C-P</sub>* = 9.2 Hz), 129.96, 128.75 (d, *J<sub>C-P</sub>* = 11.5 Hz), 128.08, 126.12, 38.16, 30.34 (d, *J<sub>C-P</sub>* = 14.4 Hz), 29.31 (d, *J<sub>C-P</sub>* = 71.6 Hz), 23.32, 21.23 (d, *J<sub>C-P</sub>* = 3.7 Hz). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>) δ 33.00. HRMS (APCI) m/z calcd. for C<sub>24</sub>H<sub>25</sub>ClO<sub>2</sub>P [M+H]<sup>+</sup> 411.1281, found 411.1278.

#### 1-(4-bromophenyl)-6-(diphenylphosphoryl)hexan-1-one (4Fb):



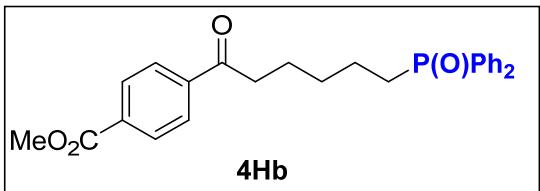
(47% yield, 40% starting was recovered); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.81 – 7.69 (m, 6H), 7.60 – 7.54 (m, 2H), 7.50 – 7.43 (m, 6H), 2.88 (t, *J* = 8.1 Hz, 2H), 2.31 – 2.24 (m, 2H), 1.76 – 1.60 (m, 4H), 1.52 – 1.47 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 199.00, 135.60, 133.04 (d, *J<sub>C-P</sub>* = 98.0 Hz), 131.87, 131.71 (d, *J<sub>C-P</sub>* = 2.6 Hz), 130.72 (d, *J<sub>C-P</sub>* = 9.9 Hz), 129.55, 128.72 (d, *J<sub>C-P</sub>* = 11.5 Hz), 128.16, 38.00, 30.32 (d, *J<sub>C-P</sub>* = 14.5 Hz), 29.52 (d, *J<sub>C-P</sub>* = 71.6 Hz), 23.41, 21.29 (d, *J<sub>C-P</sub>* = 3.7 Hz). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>) δ 32.49. HRMS (APCI) m/z calcd. for C<sub>24</sub>H<sub>25</sub>BrO<sub>2</sub>P [M+H]<sup>+</sup> 455.0776, found 455.0772.

#### 4-(6-(diphenylphosphoryl)hexanoyl)benzonitrile (4Gb):



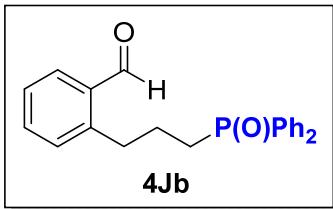
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.01 (d, *J* = 8.4 Hz, 2H), 7.78 – 7.70 (m, 6H), 7.58 – 7.44 (m, 6H), 2.95 (t, *J* = 7.2 Hz, 2H), 2.38 – 2.25 (m, 2H), 1.80 – 1.65 (m, 4H), 1.57 – 1.46 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 198.62, 139.82, 132.85 (d, *J* = 98.3 Hz), 132.50, 131.78 (d, *J* = 2.7 Hz), 130.74 (d, *J* = 9.3 Hz), 128.69 (d, *J* = 11.6 Hz), 128.42, 117.95, 116.26, 38.36, 30.26 (d, *J* = 14.4 Hz), 29.44 (d, *J* = 72.0 Hz), 23.21, 21.24 (d, *J* = 3.8 Hz). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>) δ 32.77. HRMS (APCI) m/z calcd. for C<sub>25</sub>H<sub>25</sub>N<sub>2</sub>O<sub>2</sub>P [M+H]<sup>+</sup> 402.1623, found 402.1615.

**methyl 4-(6-(diphenylphosphoryl)hexanoyl)benzoate (4Hb):**



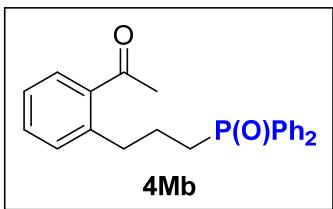
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.11 (d, *J* = 8.5 Hz, 2H), 7.96 (d, *J* = 8.5 Hz, 2H), 7.77 – 7.70 (m, 4H), 7.54 – 7.43 (m, 6H), 3.95 (s, 3H), 2.96 (t, *J* = 7.2 Hz, 2H), 2.34 – 2.23 (m, 2H), 1.78 – 1.63 (m, 4H), 1.55 – 1.45 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 199.56, 166.23, 140.10, 133.76, 132.93 (d, *J* = 98.2 Hz), 131.73 (d, *J* = 2.8 Hz), 130.75 (d, *J* = 9.2 Hz), 129.82, 128.67 (d, *J* = 11.5 Hz), 127.90, 52.47, 38.43, 30.36 (d, *J* = 14.4 Hz), 29.48 (d, *J* = 71.9 Hz), 23.33, 21.28 (d, *J* = 3.8 Hz). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>) δ 32.65. HRMS (APCI) m/z calcd. for C<sub>26</sub>H<sub>28</sub>O<sub>4</sub>P [M+H]<sup>+</sup> 435.1725, found 435.1719.

**2-(3-(diphenylphosphoryl)propyl)benzaldehyde (4Jb):**



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 10.14 (s, 1H), 7.80 – 7.69 (m, 5H), 7.55 – 7.42 (m, 7H), 7.42 – 7.36 (m, 1H), 7.29 – 7.22 (m, 1H), 3.17 (t, *J* = 7.7 Hz, 2H), 2.42 – 2.34 (m, 2H), 2.01 – 1.89 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 192.67, 143.59, 133.79, 133.73, 133.39, 132.84 (d, *J*<sub>C-P</sub> = 98.3 Hz), 131.75 (d, *J*<sub>C-P</sub> = 2.8 Hz), 131.12, 130.75 (d, *J*<sub>C-P</sub> = 9.3 Hz), 128.68 (d, *J*<sub>C-P</sub> = 11.5 Hz), 126.85, 33.46 (d, *J*<sub>C-P</sub> = 14.9 Hz), 29.29 (d, *J*<sub>C-P</sub> = 71.8 Hz), 23.67 (d, *J*<sub>C-P</sub> = 3.4 Hz). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>) δ 32.53. HRMS (APCI) m/z calcd. for C<sub>22</sub>H<sub>22</sub>O<sub>2</sub>P [M+H]<sup>+</sup> 349.1345, found 349.1352.

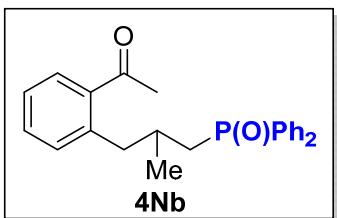
**1-(2-(3-(diphenylphosphoryl)propyl)phenyl)ethanone (4Mb):**



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.69 – 7.65 (m, 4H), 7.56 (d, *J* = 7.7 Hz, 1H), 7.45 – 7.32 (m, 6H), 7.28 (t, *J* = 7.4 Hz, 1H), 7.19 – 7.13 (m, 2H), 2.98 – 2.86 (m, 2H), 2.42 (s, 3H), 2.33 – 2.26 (m, 2H), 1.94 – 1.80 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 201.71, 141.29, 137.44, 133.02 (d, *J*<sub>C-P</sub> = 97.3 Hz), 131.62 (d, *J*<sub>C-P</sub> = 2.7 Hz), 131.63, 131.20, 130.70 (d, *J*<sub>C-P</sub> = 9.2 Hz), 129.49, 128.65 (d, *J*<sub>C-P</sub> = 11.5 Hz), 126.15, 34.82 (d, *J*<sub>C-P</sub> = 15.5 Hz), 29.70, 29.31 (d, *J*<sub>C-P</sub> = 70.0 Hz), 23.52 (d, *J*<sub>C-P</sub> = 3.2 Hz). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>) δ 32.46. HRMS (APCI) m/z calcd. for C<sub>23</sub>H<sub>24</sub>O<sub>2</sub>P [M+H]<sup>+</sup>

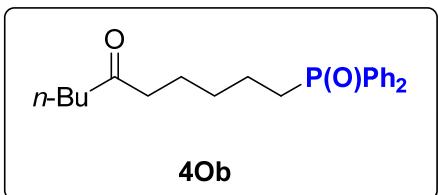
363.1514, found 363.1510.

**1-(2-(3-(diphenylphosphoryl)-2-methylpropyl)phenyl)ethan-1-one (4Nb):**



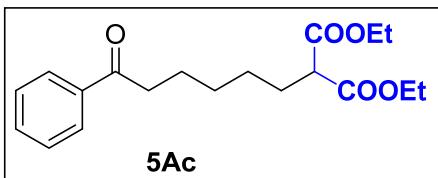
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.72 – 7.66 (m, 2H), 7.64 – 7.55 (m, 3H), 7.51 – 7.35 (m, 7H), 7.33 – 7.27 (m, 1H), 7.17 (d, *J* = 7.6 Hz, 1H), 2.98 – 2.87 (m, 2H), 2.47 – 2.39 (m, 4H), 2.19 – 2.12 (m, 2H), 1.08 (d, *J* = 6.1 Hz, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 202.16, 140.10, 138.25, 134.56 (d, *J<sub>C-P</sub>* = 97.8 Hz), 132.80 (d, *J<sub>C-P</sub>* = 97.5 Hz), 132.14, 131.46 (d, *J<sub>C-P</sub>* = 2.7 Hz), 131.26, 131.40 (d, *J<sub>C-P</sub>* = 2.7 Hz), 130.87 (d, *J<sub>C-P</sub>* = 9.1 Hz), 130.46 (d, *J<sub>C-P</sub>* = 9.2 Hz), 129.33, 128.61 (d, *J<sub>C-P</sub>* = 3.0 Hz), 128.50 (d, *J<sub>C-P</sub>* = 3.1 Hz), 126.21, 42.57 (d, *J<sub>C-P</sub>* = 13.9 Hz), 35.63 (d, *J<sub>C-P</sub>* = 71.3 Hz), 30.81 (d, *J<sub>C-P</sub>* = 3.5 Hz), 29.85, 21.33 (d, *J<sub>C-P</sub>* = 3.0 Hz). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>) δ 31.82. HRMS (APCI) m/z calcd. for C<sub>24</sub>H<sub>26</sub>O<sub>2</sub>P [M+H]<sup>+</sup> 377.1653, found 377.1665.

**10-(diphenylphosphoryl)decan-5-one (4Ob):**



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.75 – 7.67 (m, 4H), 7.52 – 7.42 (m, 6H), 2.33 (td, *J* = 7.4, 1.8 Hz, 4H), 2.29 – 2.18 (m, 2H), 1.63 – 1.58 (m, 2H), 1.56 – 1.46 (m, 4H), 1.42 – 1.32 (m, 2H), 1.31 – 1.21 (m, 2H), 0.87 (t, *J* = 7.2 Hz, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 211.26, 132.98 (d, *J<sub>C-P</sub>* = 98.0 Hz), 131.71 (d, *J<sub>C-P</sub>* = 2.6 Hz), 130.73 (d, *J<sub>C-P</sub>* = 9.9 Hz), 128.65 (d, *J<sub>C-P</sub>* = 11.5 Hz), 42.57, 42.20, 30.34 (d, *J<sub>C-P</sub>* = 14.5 Hz), 29.44 (d, *J<sub>C-P</sub>* = 71.6 Hz), 25.92, 23.05, 22.32, 21.21 (d, *J<sub>C-P</sub>* = 3.7 Hz), 13.86. <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>) δ 32.70. HRMS (APCI) m/z calcd. for C<sub>22</sub>H<sub>30</sub>O<sub>2</sub>P [M+H]<sup>+</sup> 357.1983, found 357.1977.

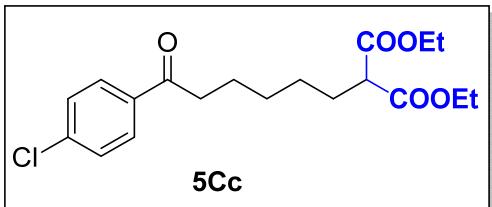
**diethyl 2-(6-oxo-6-phenylhexyl)malonate (5Ac):**



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.96 (d, *J* = 7.6 Hz, 2H), 7.57 (t, *J* = 7.4 Hz, 1H), 7.47 (t, *J* = 7.6 Hz, 2H), 4.25 – 4.16 (m, 4H), 3.33 (t, *J* = 7.5 Hz, 1H), 2.97 (t, *J* = 7.3 Hz, 2H), 1.96 – 1.90 (m, 2H), 1.80 – 1.72 (m, 2H), 1.48 – 1.37 (m, 4H), 1.28 (t, *J* = 7.1 Hz,

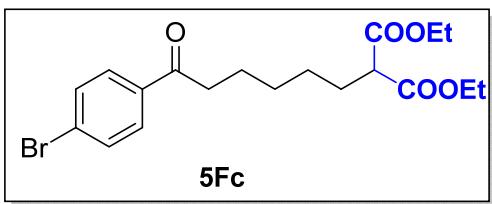
6H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  200.29, 169.53, 137.01, 132.94, 128.58, 128.04, 61.31, 51.99, 38.37, 28.93, 28.58, 27.20, 23.98, 14.09. HRMS (APCI) m/z calcd. for  $\text{C}_{19}\text{H}_{27}\text{O}_5$   $[\text{M}+\text{H}]^+$  335.1858, found 335.1857.

**diethyl 2-(6-(4-chlorophenyl)-6-oxohexyl)malonate (5Cc):**



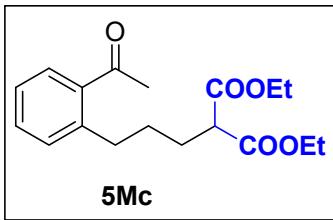
$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.91 (d,  $J = 8.6$  Hz, 2H), 7.45 (d,  $J = 8.6$  Hz, 2H), 4.27 – 4.15 (m, 4H), 3.33 (t,  $J = 7.5$  Hz, 1H), 2.94 (t,  $J = 7.3$  Hz, 2H), 1.92 (dd,  $J = 14.9$ , 7.5 Hz, 2H), 1.80 – 1.70 (m, 2H), 1.44 – 1.40 (m, 4H), 1.28 (t,  $J = 7.1$  Hz, 6H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  198.99, 169.51, 139.36, 135.30, 129.47, 128.89, 61.32, 51.98, 38.34, 28.87, 28.55, 27.16, 23.87, 14.09. HRMS (APCI) m/z calcd. for  $\text{C}_{19}\text{H}_{26}\text{O}_5\text{Cl}$   $[\text{M}+\text{H}]^+$  369.1469, found 369.1466.

**diethyl 2-(6-(4-bromophenyl)-6-oxohexyl)malonate (5Fc):**



$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.81 (d,  $J = 8.5$  Hz, 2H), 7.59 (d,  $J = 8.5$  Hz, 2H), 4.24 – 4.13 (m, 4H), 3.31 (t,  $J = 7.5$  Hz, 1H), 2.92 (t,  $J = 7.3$  Hz, 2H), 1.93 – 1.88 (m, 2H), 1.77 – 1.69 (m, 2H), 1.42 – 1.38 (m, 4H), 1.26 (t,  $J = 7.1$  Hz, 6H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  199.14, 169.49, 135.68, 131.87, 129.58, 128.05, 61.31, 51.95, 38.30, 28.84, 28.53, 27.15, 23.83, 14.09. HRMS (APCI) m/z calcd. for  $\text{C}_{19}\text{H}_{26}\text{O}_5\text{Br}$   $[\text{M}+\text{H}]^+$  413.0964, found 413.0955.

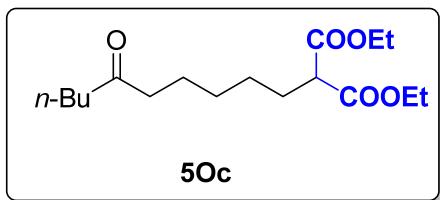
**diethyl 2-(3-(2-acetylphenyl)propyl)malonate (5Mc):**



$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.66 (dd,  $J = 7.7$ , 1.1 Hz, 1H), 7.38 (dt,  $J = 7.5$ , 3.8 Hz, 1H), 7.30 – 7.19 (m, 2H), 4.26 – 4.10 (m, 4H), 3.36 (t,  $J = 7.5$  Hz, 1H), 2.93 – 2.79 (m, 2H), 2.57 (s, 3H), 1.99 – 1.93 (m, 2H), 1.66 – 1.58 (m, 2H), 1.25 (t,  $J = 7.1$  Hz, 6H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  201.86, 169.47, 141.96, 137.65, 131.52, 131.26, 129.36, 125.97, 61.26, 51.81, 33.69, 29.78, 29.22, 28.65, 14.07. HRMS (APCI) m/z

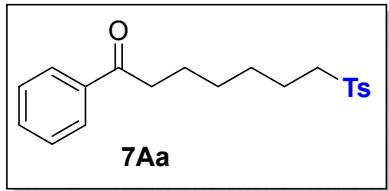
calcd. for C<sub>18</sub>H<sub>25</sub>O<sub>5</sub> [M+H]<sup>+</sup> 321.1702, found 321.1695.

**diethyl 2-(6-oxodecyl)malonate (5Oc)**



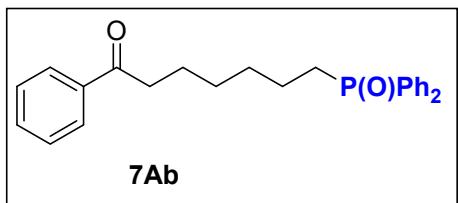
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 4.19 (qd, *J* = 7.1, 1.0 Hz, 4H), 3.30 (t, *J* = 7.5 Hz, 1H), 2.39 (t, *J* = 7.5 Hz, 4H), 1.88 (dd, *J* = 15.0, 7.5 Hz, 2H), 1.60 – 1.50 (m, 4H), 1.36 – 1.24 (m, 12H), 0.90 (t, *J* = 7.3 Hz, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 211.41, 169.51, 61.30, 51.96, 42.55, 42.53, 28.82, 28.53, 27.12, 25.96, 23.48, 22.36, 14.08, 13.86. HRMS (ESI) m/z calcd. for C<sub>17</sub>H<sub>31</sub>O<sub>5</sub> [M+H]<sup>+</sup> 315.2171, found 315.2166.

**1-phenyl-7-tosylheptan-1-one (7Aa)**



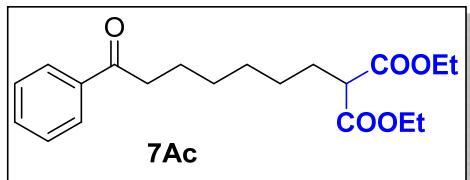
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.93 (d, *J* = 7.5 Hz, 2H), 7.77 (d, *J* = 8.2 Hz, 2H), 7.55 (t, *J* = 7.3 Hz, 1H), 7.45 (t, *J* = 7.6 Hz, 2H), 7.35 (d, *J* = 8.0 Hz, 2H), 3.12 – 3.03 (m, 2H), 2.94 (t, *J* = 7.2 Hz, 2H), 2.44 (s, 3H), 1.75 – 1.66 (m, 4H), 1.43 – 1.33 (m, 4H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 200.12, 144.60, 136.96, 136.24, 133.00, 129.90, 128.60, 128.17, 128.01, 56.30, 38.24, 28.71, 28.17, 23.82, 22.65, 21.62. HRMS (APCI) m/z calcd. for C<sub>20</sub>H<sub>25</sub>O<sub>3</sub>S [M+H]<sup>+</sup> 345.1524, found 345.1520.

**7-(diphenylphosphoryl)-1-phenylheptan-1-one (7Ab):**



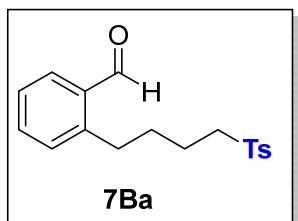
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.90 (d, *J* = 7.4 Hz, 2H), 7.78 – 7.64 (m, 4H), 7.55 – 7.34 (m, 9H), 2.89 (t, *J* = 7.3 Hz, 2H), 2.31 – 2.17 (m, 2H), 1.68 – 1.62 (m, 4H), 1.48 – 1.28 (m, 4H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 200.28, 136.95, 133.12 (d, *J*<sub>C-P</sub> = 97.3 Hz), 132.93, 131.61 (d, *J*<sub>C-P</sub> = 2.6 Hz), 130.74 (d, *J*<sub>C-P</sub> = 9.2 Hz), 128.64 (d, *J*<sub>C-P</sub> = 11.6 Hz), 128.56, 127.99, 38.34, 30.83 (d, *J*<sub>C-P</sub> = 14.3 Hz), 29.61 (d, *J*<sub>C-P</sub> = 71.6 Hz), 28.75, 23.96, 21.33 (d, *J*<sub>C-P</sub> = 3.9 Hz). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>) δ 32.52. HRMS (APCI) m/z calcd. for C<sub>25</sub>H<sub>28</sub>O<sub>2</sub>P [M+H]<sup>+</sup> 391.1827, found 391.1824.

**diethyl 2-(7-oxo-7-phenylheptyl)malonate (7Ac):**



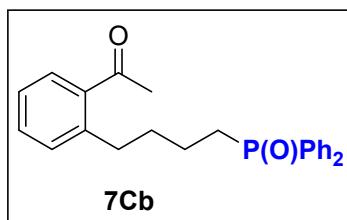
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.99 – 7.94 (m, 2H), 7.59 – 7.53 (m, 1H), 7.48 – 7.45 (m, 2H), 4.23 – 4.16 (m, 4H), 3.32 (t, *J* = 7.5 Hz, 1H), 2.97 (t, *J* = 7.4 Hz, 2H), 1.93 – 1.88 (m, 2H), 1.76 – 1.70 (m, 2H), 1.43 – 1.34 (m, 6H), 1.27 (t, *J* = 7.1 Hz, 6H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 200.41, 169.56, 137.06, 132.90, 128.56, 128.03, 61.27, 52.03, 38.48, 29.05, 29.02, 28.67, 27.18, 24.17, 14.08. HRMS (APCI) m/z calcd. for C<sub>20</sub>H<sub>29</sub>O<sub>5</sub> [M+H]<sup>+</sup> 349.2015, found 349.2013.

**2-(4-tosylbutyl)benzaldehyde (7Ba):**



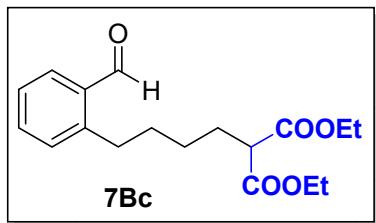
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 10.16 (s, 1H), 7.88 – 7.74 (m, 3H), 7.51 (t, *J* = 7.0 Hz, 1H), 7.44 – 7.35 (m, 3H), 7.23 (d, *J* = 7.6 Hz, 1H), 3.17 – 3.09 (m, 2H), 3.04 – 2.99 (m, 2H), 2.48 (s, 3H), 1.88 – 1.76 (m, 2H), 1.74 – 1.62 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 192.79, 144.67, 143.97, 136.16, 133.83, 133.64, 131.13, 129.93, 129.89, 128.10, 126.81, 56.09, 32.31, 30.29, 22.60, 21.65. HRMS (APCI) m/z calcd. for C<sub>18</sub>H<sub>21</sub>O<sub>3</sub>S [M+H]<sup>+</sup> 317.1199, found 317.1206.

**2-(4-(diphenylphosphoryl)butyl)benzaldehyde (7Cb):**



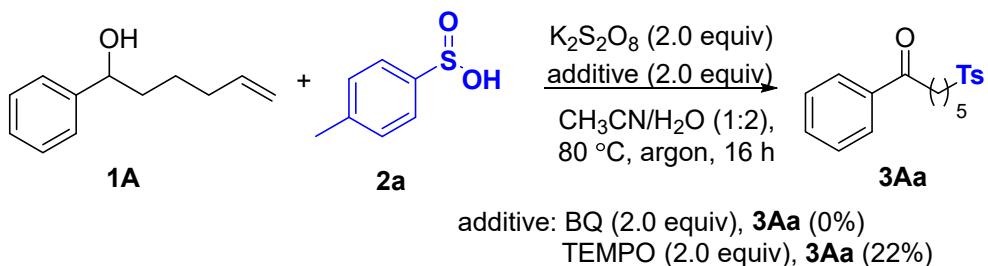
<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.74 (ddd, *J* = 11.4, 8.1, 1.6 Hz, 4H), 7.67 – 7.60 (m, 1H), 7.57 – 7.41 (m, 6H), 7.36 (td, *J* = 7.5, 1.4 Hz, 1H), 7.31 – 7.20 (m, 1H), 7.19 (dd, *J* = 7.6, 1.3 Hz, 1H), 2.83 (t, *J* = 7.2 Hz, 2H), 2.55 (s, 3H), 2.37 – 2.27 (m, 2H), 1.77 – 1.66 (m, 4H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 202.01, 142.15, 137.59, 133.07 (d, *J*<sub>C-P</sub> = 98.2 Hz), 131.65 (d, *J*<sub>C-P</sub> = 2.8 Hz), 131.52, 131.27, 130.79 (d, *J*<sub>C-P</sub> = 9.3 Hz), 129.29, 128.63 (d, *J*<sub>C-P</sub> = 11.6 Hz), 125.86, 33.53, 32.88 (d, *J*<sub>C-P</sub> = 14.8 Hz), 29.82, 29.11, 21.48 (d, *J*<sub>C-P</sub> = 3.9 Hz). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>) δ 32.73. HRMS (APCI) m/z calcd. for C<sub>24</sub>H<sub>26</sub>O<sub>2</sub>P [M+H]<sup>+</sup> 377.1658, found 377.1665.

**diethyl 2-(4-(2-formylphenyl)butyl)malonate (7Bc):**

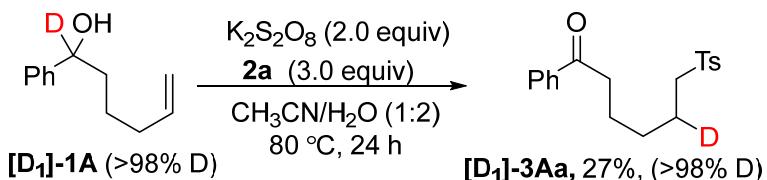


$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  10.27 (s, 1H), 7.84 (dd,  $J = 7.6, 1.4$  Hz, 1H), 7.52 (td,  $J = 7.5, 1.5$  Hz, 1H), 7.39 (td,  $J = 7.5, 1.2$  Hz, 1H), 7.31 – 7.24 (m, 1H), 4.21 (qd,  $J = 7.1, 1.2$  Hz, 4H), 3.34 (t,  $J = 7.5$  Hz, 1H), 3.08 – 3.02 (m, 2H), 2.01 – 1.89 (m, 2H), 1.72 – 1.61 (m, 2H), 1.50 – 1.38 (m, 2H), 1.28 (t,  $J = 7.1$  Hz, 6H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  192.40, 169.50, 145.12, 133.78, 133.66, 132.13, 131.01, 126.55, 63.24, 51.92, 32.28, 31.68, 28.51, 27.17, 14.08. HRMS (APCI) m/z calcd. for  $\text{C}_{18}\text{H}_{25}\text{O}_5\text{P} [\text{M}+\text{H}]^+$  321.1689, found 321.1697.

## Mechanistic Studies:

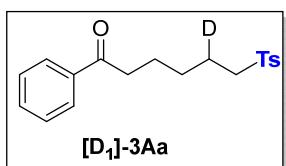


**Procedure:** To a flame-dried Schlenk tube equipped with a magnetic stir bar were added **1A** (0.2 mmol), **2a** (0.6 mmol) and  $\text{K}_2\text{S}_2\text{O}_8$  (0.4 mmol) and additive (0.4 mmol). The tube was evacuated and backfilled with argon for three times, and then  $\text{CH}_3\text{CN}$  (0.7 mL) and  $\text{H}_2\text{O}$  (1.4 mL) were added. The tube was stirred at  $80^\circ\text{C}$  for 18 h and then  $\text{H}_2\text{O}$  (5 mL) was added.  $\text{EtOAc}$  was used to extract the product from the aqueous layer ( $3 \times 20$  mL). The combined organic layer was dried over anhydrous  $\text{Na}_2\text{SO}_4$ , filtered and concentrated to afford the crude product. When TEMPO was used as the additive, the  $^1\text{H}$  NMR yield of **3Aa** is 22%. When *p*-benzoquinone was used as the additive, no **3Aa** was observed and **1A** was recovered.



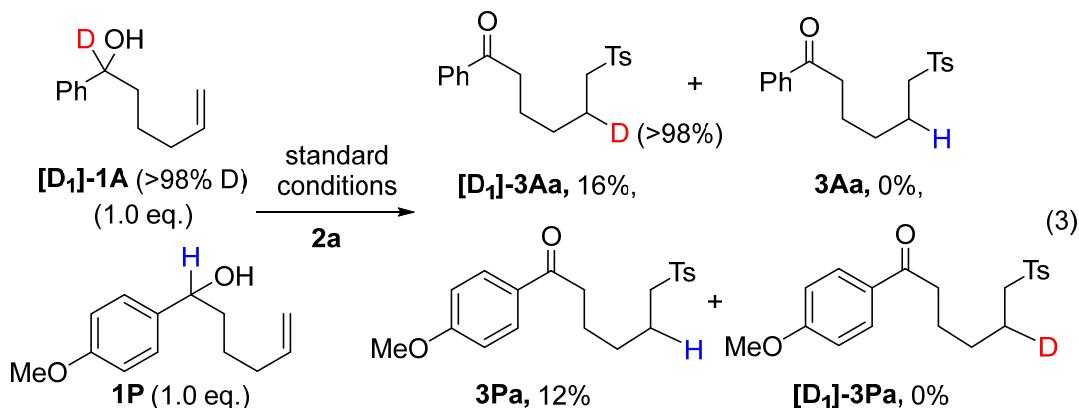
**Procedure:** To a flame-dried Schlenk tube equipped with a magnetic stir bar were added **[D<sub>1</sub>]-1A** (0.2 mmol), **2a** (0.6 mmol) and  $\text{K}_2\text{S}_2\text{O}_8$  (0.4 mmol). The tube was evacuated and backfilled with argon for three times, and then  $\text{CH}_3\text{CN}$  (0.7 mL) and  $\text{H}_2\text{O}$  (1.4 mL) were added. The tube was stirred at  $80^\circ\text{C}$  for 24 h and then  $\text{H}_2\text{O}$  (5 mL) was added.  $\text{EtOAc}$  was used to extract the product from the aqueous layer ( $3 \times 20$  mL). The combined organic layer was dried over anhydrous  $\text{Na}_2\text{SO}_4$ , filtered and concentrated to afford the crude product, which was purified by flash column chromatography to afford the product **[D<sub>1</sub>]-3Aa** (27%) and 35% starting material **[D<sub>1</sub>]-1A** was recovered.

### Compound **[D<sub>1</sub>]-3Aa**:



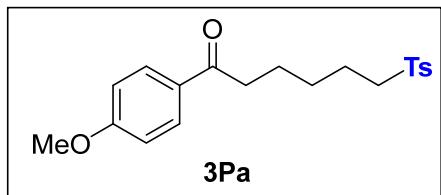
(35% starting recovered, 27% yield)  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.97 – 7.89 (m, 2H), 7.80 (d,  $J = 8.2$  Hz, 2H), 7.59 – 7.56 (m, 1H), 7.47 (t,  $J = 7.6$  Hz, 2H), 7.37 (d,  $J = 8.0$  Hz, 2H), 3.10 (d,  $J = 8.0$  Hz, 2H), 2.96 (t,  $J = 7.2$  Hz, 2H), 2.46 (s, 3H), 1.78 – 1.69 (m, 3H), 1.50 – 1.44 (m, 2H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  199.77, 144.66,

136.85, 136.18, 133.09, 129.92, 128.63, 128.08, 127.99, 56.10, 37.94, 27.81, 23.42, 22.36 (*t*,  $J = 20$  Hz), 21.64. HRMS (APCI)  $m/z$  calcd. for  $C_{19}H_{22}DO_3S$   $[M+H]^+$  332.1432, found 332.1428.



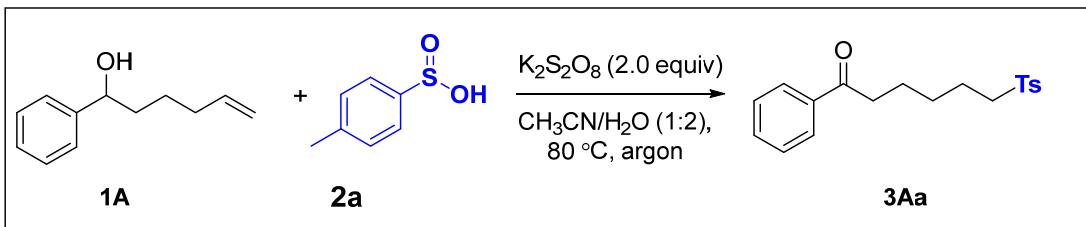
**Procedure:** To a flame-dried Schlenk tube equipped with a magnetic stir bar were added **[D<sub>1</sub>]-1A** (0.2 mmol), **1P** (0.2 mmol), **2a** (1.2 mmol) and  $K_2S_2O_8$  (0.8 mmol). The tube was evacuated and backfilled with argon for three times, and then  $CH_3CN$  (0.7 mL) and  $H_2O$  (1.4 mL) were added. The tube was stirred at 80 °C for 24 h and then  $H_2O$  (5 mL) was added. EtOAc was used to extract the product from the aqueous layer (3 × 20 mL). The combined organic layer was dried over anhydrous  $Na_2SO_4$ , filtered and concentrated to afford the crude product, which was purified by flash column chromatography to afford **3Pa** (12%) and **[D<sub>1</sub>]-3Aa** (16%). Meanwhile, about 35% **[D<sub>1</sub>]-1A** was recovered.

### 1-(4-methoxyphenyl)-6-tosylhexan-1-one (3Pa):



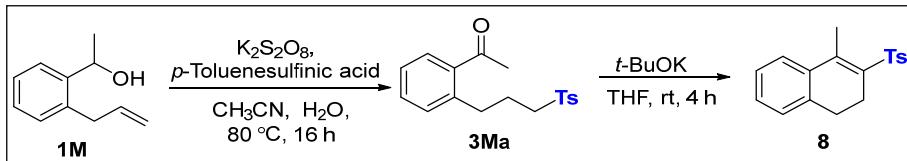
$^1H$  NMR (400 MHz,  $CDCl_3$ )  $\delta$  7.92 (d,  $J = 8.9$  Hz, 2H), 7.79 (d,  $J = 8.2$  Hz, 2H), 7.37 (d,  $J = 8.0$  Hz, 2H), 6.94 (d,  $J = 8.9$  Hz, 2H), 3.88 (s, 3H), 3.15 – 3.06 (m, 2H), 2.91 (t,  $J = 7.2$  Hz, 2H), 2.47 (s, 3H), 1.81 – 1.68 (m, 4H), 1.50 – 1.44 (m, 2H).  $^{13}C$  NMR (100 MHz,  $CDCl_3$ )  $\delta$  198.38, 163.47, 144.64, 136.18, 130.26, 129.96, 129.91, 128.08, 113.74, 56.18, 55.49, 37.57, 27.95, 23.67, 22.69, 21.62. HRMS (APCI)  $m/z$  calcd. for  $C_{20}H_{25}O_4S$   $[M+H]^+$  361.1474, found 361.1470.

## Large scale preparation and transformations



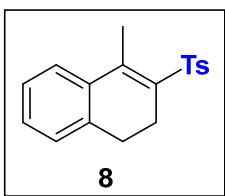
To a flame-dried Schlenk tube equipped with a magnetic stir bar were added **1A** (7.2 mmol), **2a** (21.6 mmol) and  $\text{K}_2\text{S}_2\text{O}_8$  (14.4 mmol). The tube was evacuated and backfilled with argon for three times, and then  $\text{CH}_3\text{CN}$  (20 mL) and  $\text{H}_2\text{O}$  (40 mL) were added. The tube was stirred at  $80^\circ\text{C}$  for 18 h and then  $\text{H}_2\text{O}$  (50 mL) was added.  $\text{EtOAc}$  was used to extract the product from the aqueous layer ( $3 \times 40$  mL). The combined organic layer was dried over anhydrous  $\text{Na}_2\text{SO}_4$ , filtered and concentrated to afford the crude product, which was purified by flash column chromatography to afford 1.7 g product of **3Aa** (70%).

## Transformation of **3Ma**



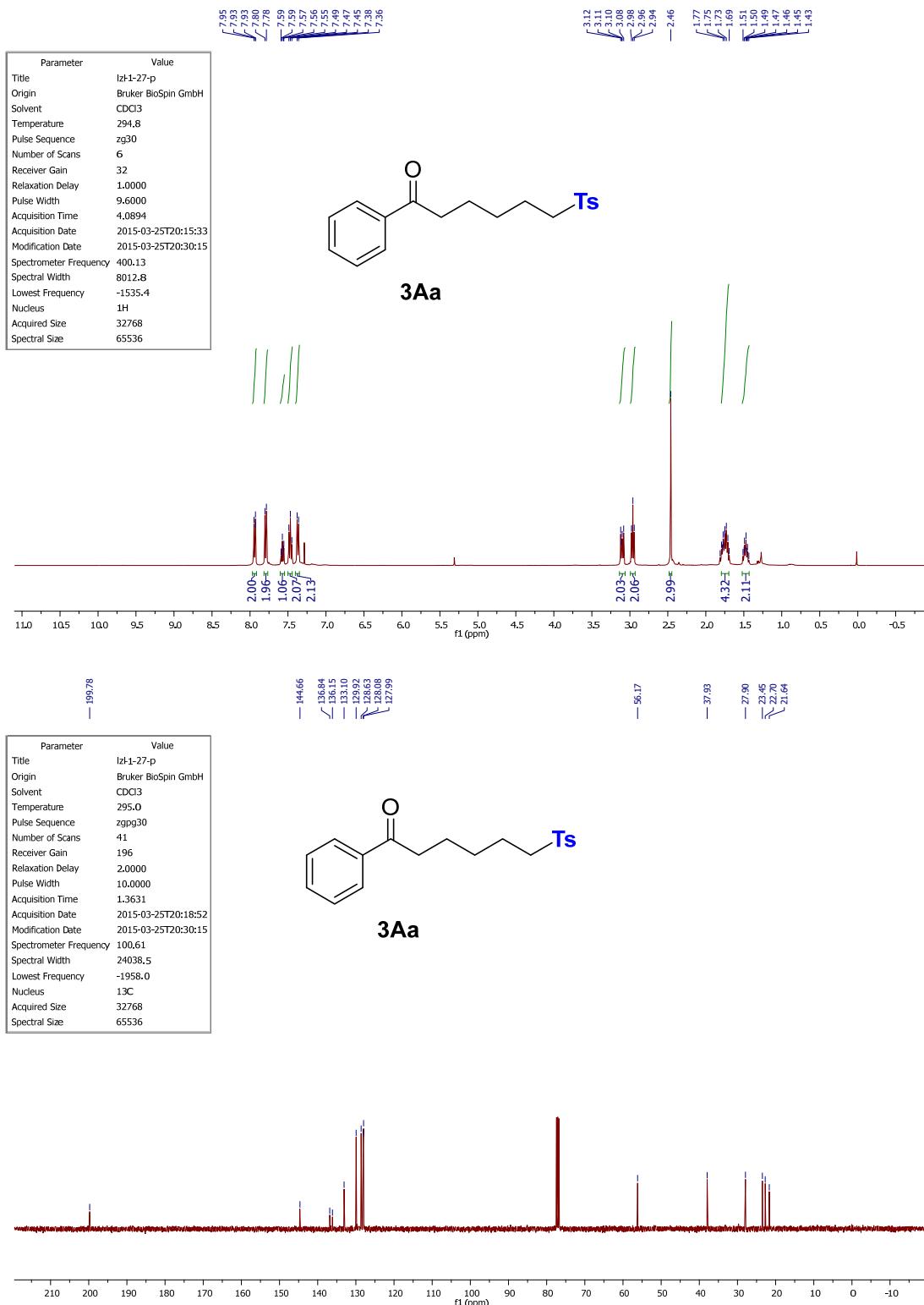
Compound **3Ma** was obtained by following the reported sulfonylation procedure.

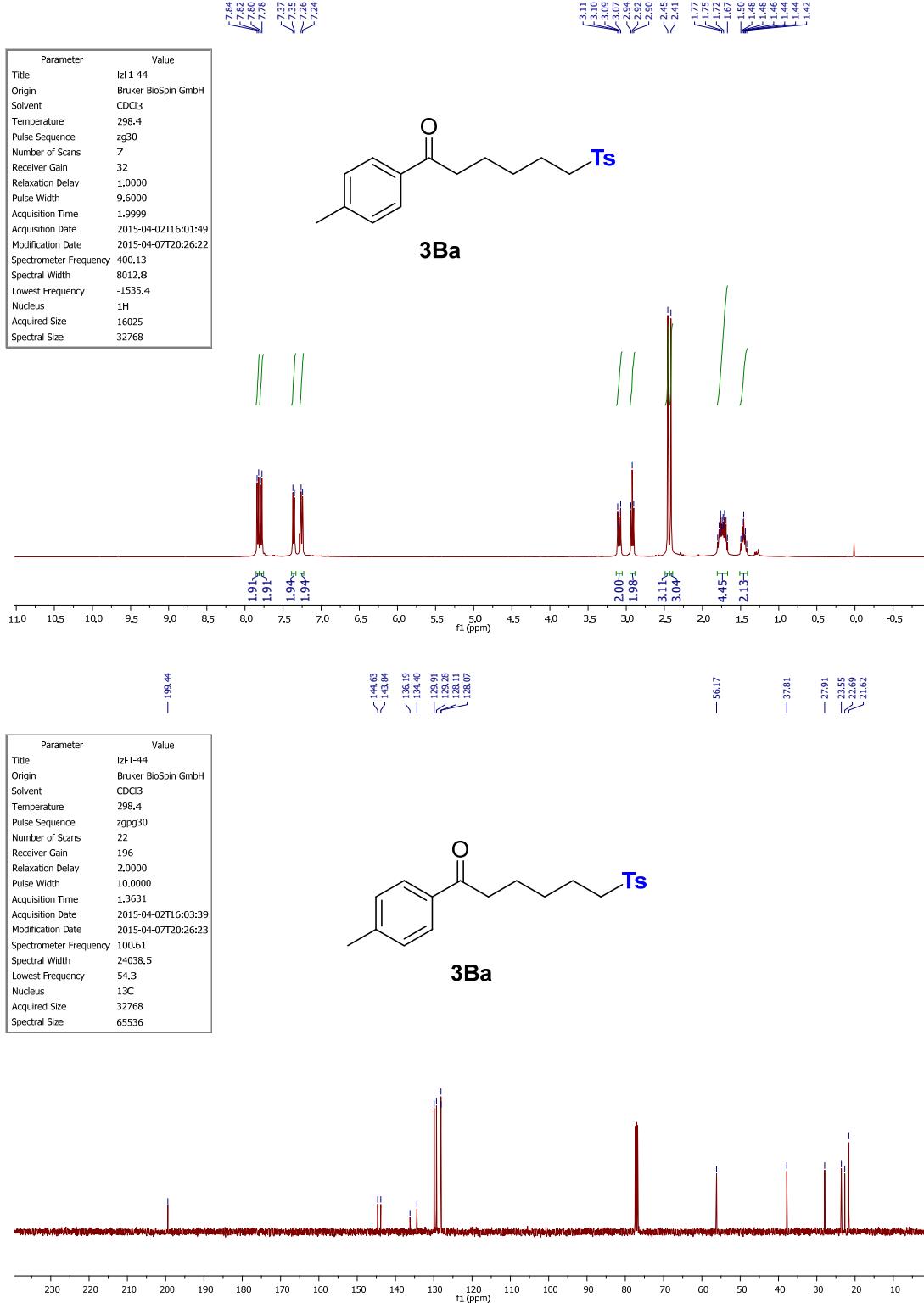
**Procedure:** To a solution of  $t\text{-BuOK}$  (74 mg, 0.66 mmol) in anhydrous THF (8 mL) was added a solution of **3Ma** (70 mg, 0.22 mmol) in THF dropwisely. The reaction was stirred 4 h and quenched with water (5 mL).  $\text{EtOAc}$  was used to extract the product from the aqueous layer ( $3 \times 20$  mL). The combined organic layer was dried over anhydrous  $\text{Na}_2\text{SO}_4$ , filtered and concentrated to afford the crude product, which was purified by flash column chromatography to afford the product **8** in 73% yield.

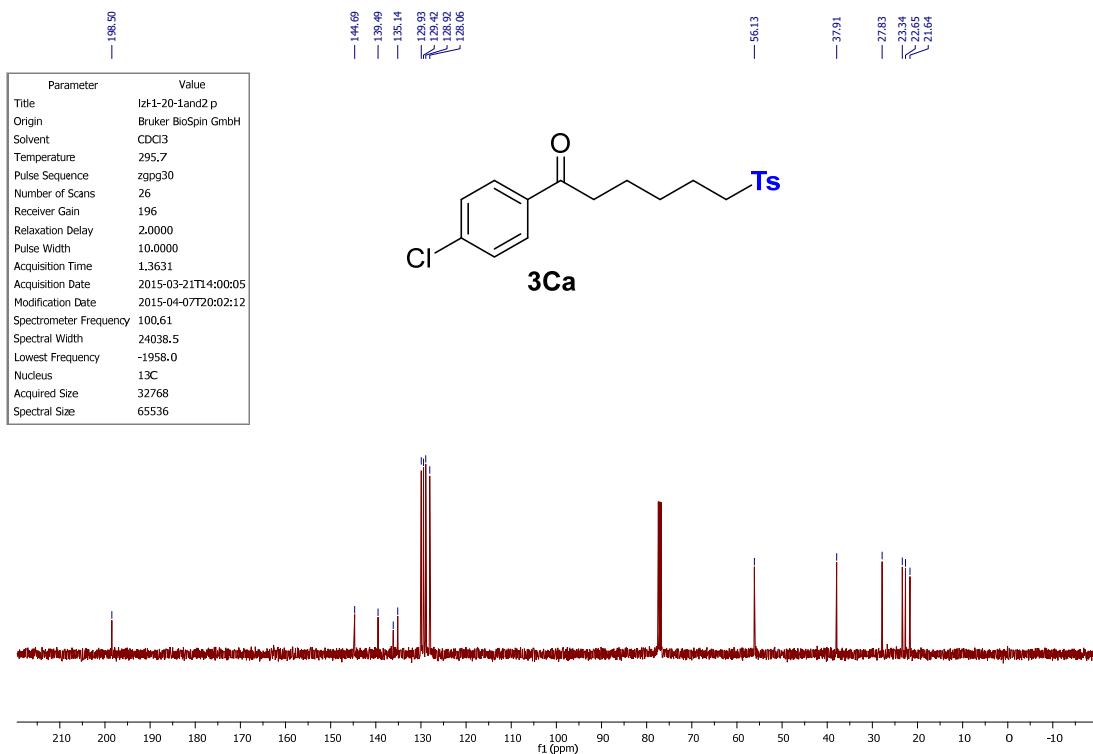
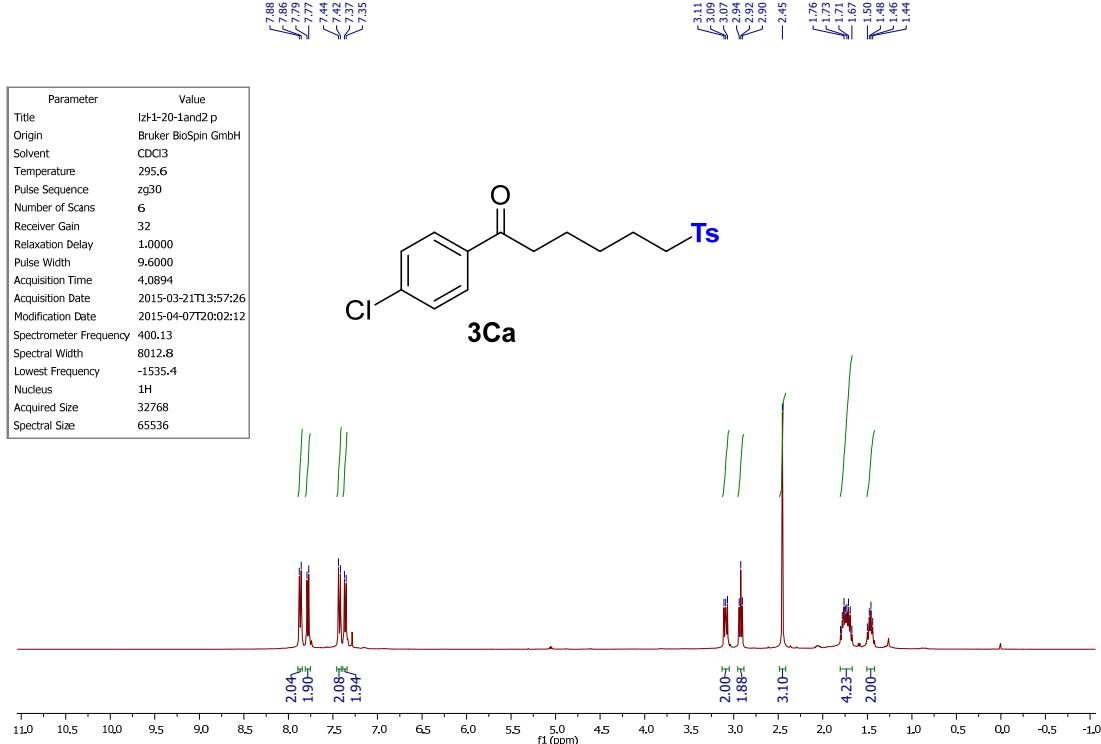


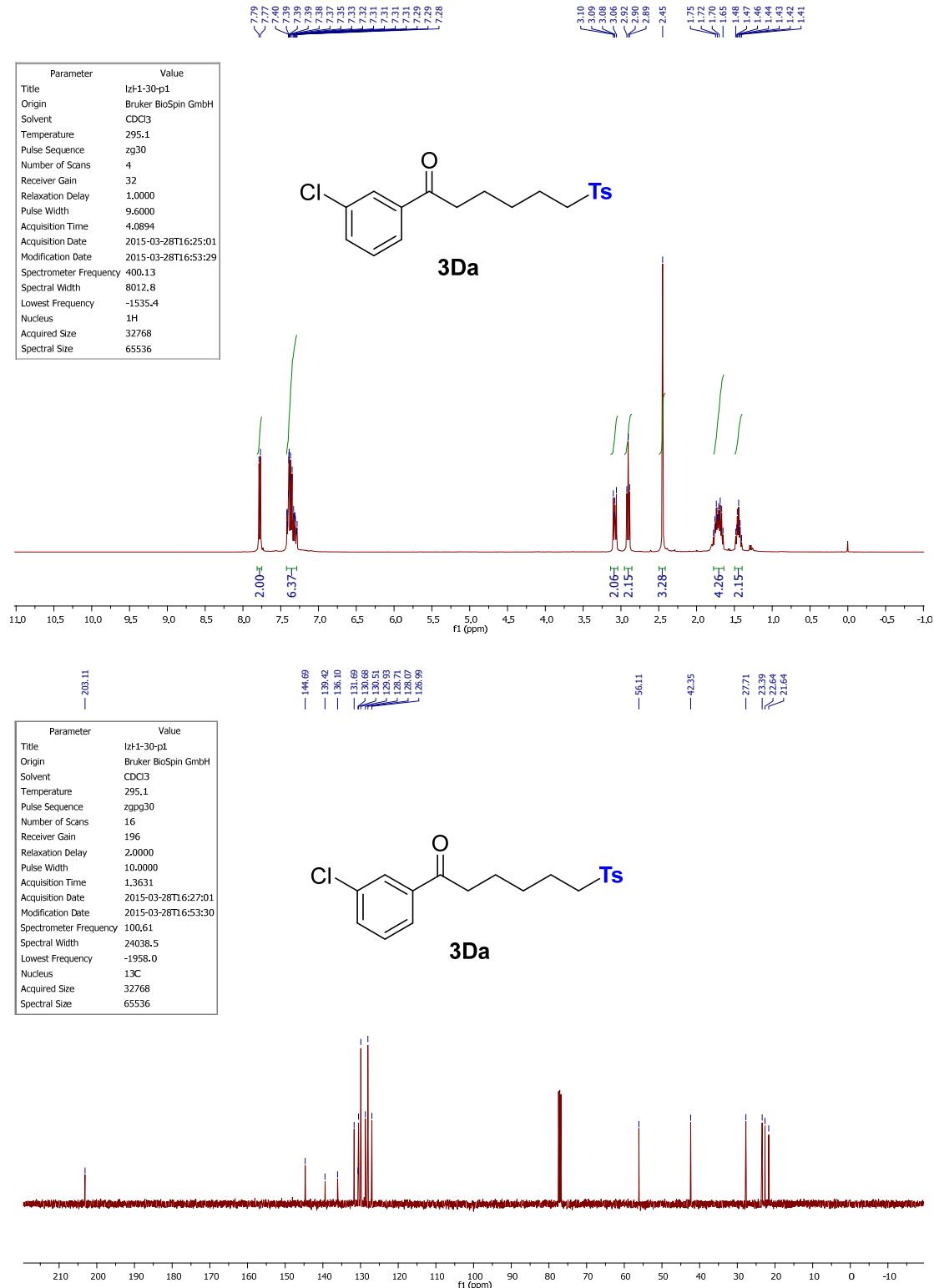
**4-methyl-3-tosyl-1,2-dihydronaphthalene (8):** (73% isolated yield);  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.83 (d,  $J = 8.1$  Hz, 2H), 7.49 – 7.45 (m, 1H), 7.34 (d,  $J = 8.0$  Hz, 2H), 7.30 – 7.26 (m, 2H), 7.19 – 7.15 (m, 1H), 2.82 – 2.75 (m, 2H), 2.71 – 2.68 (m, 2H), 2.57 (s, 3H), 2.45 (s, 3H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  143.92, 142.49, 139.29, 136.77, 135.26, 135.10, 129.75, 129.66, 127.40, 127.06, 126.90, 125.26, 28.37, 24.78, 21.61, 15.70. HRMS (APCI) m/z calcd. for  $\text{C}_{18}\text{H}_{19}\text{O}_2\text{S}$  [ $\text{M}+\text{H}]^+$  299.1106, found 299.1100.

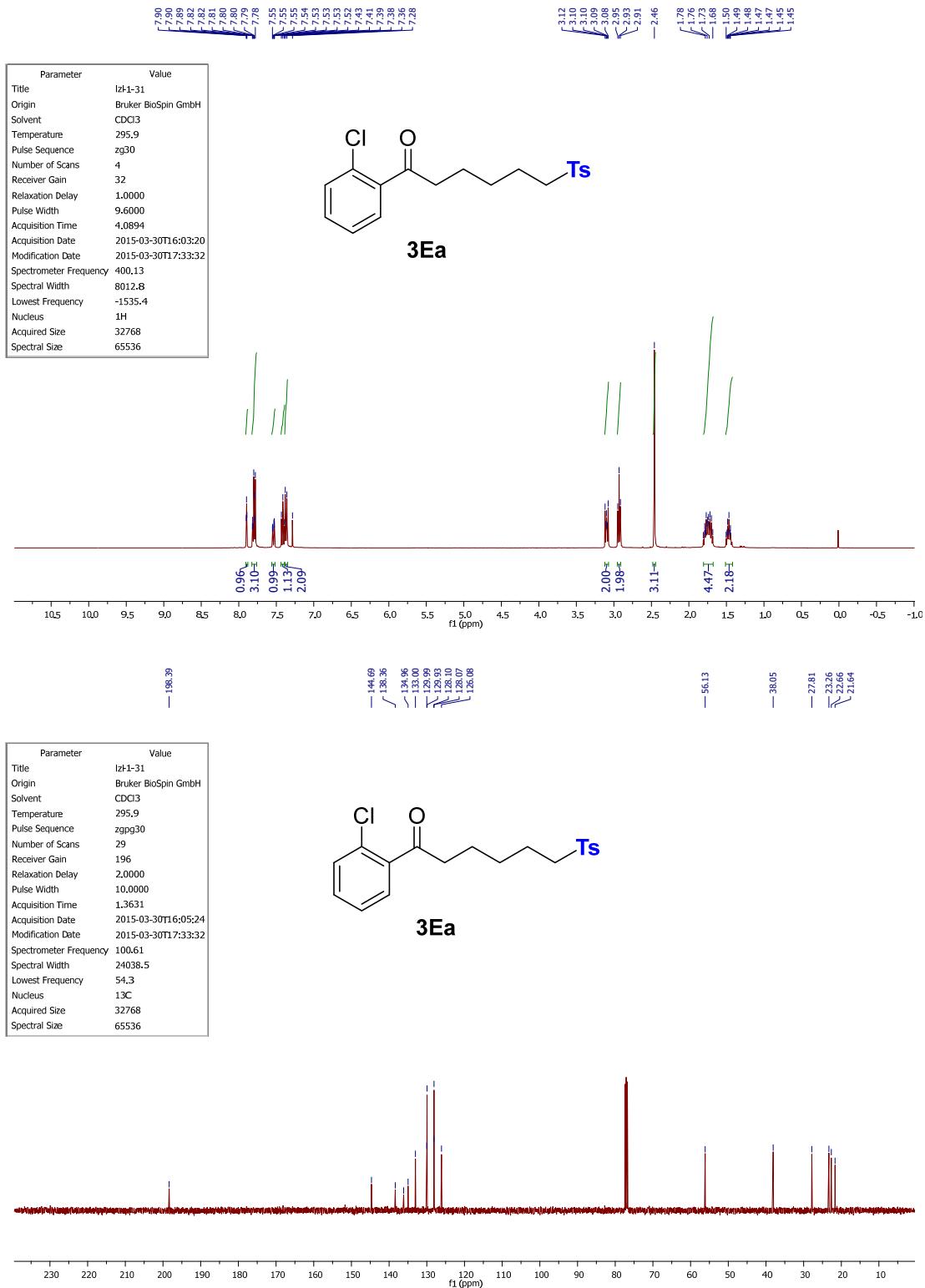
## NMR Spectra:

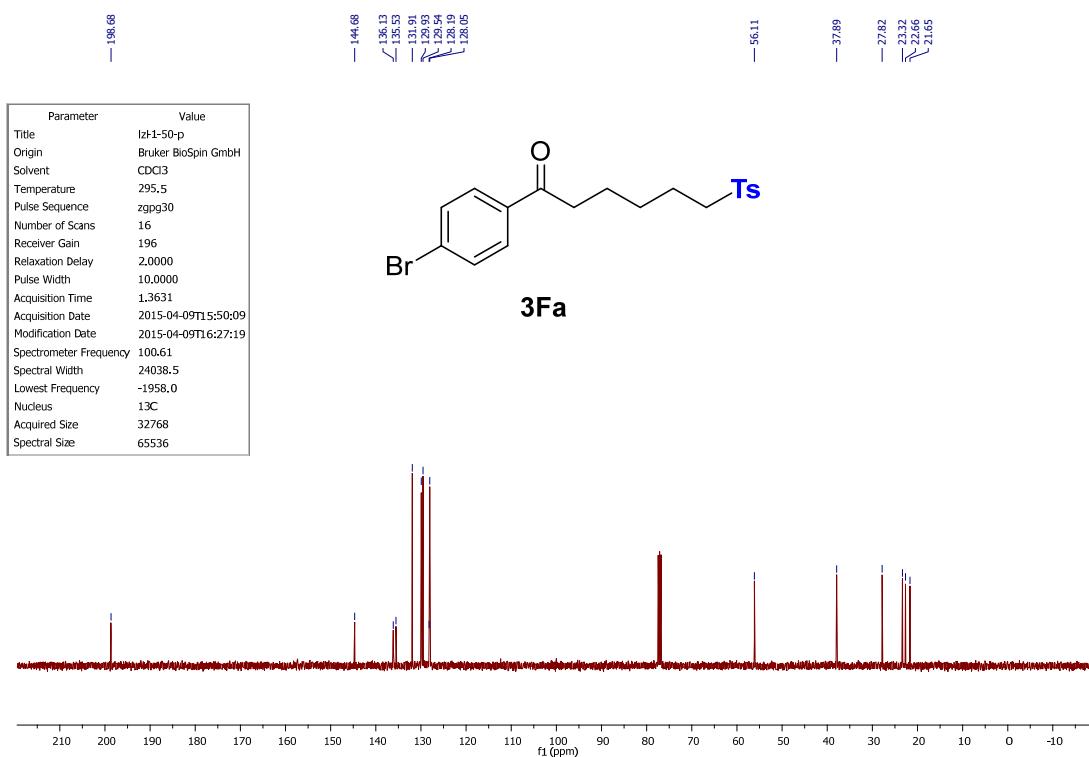
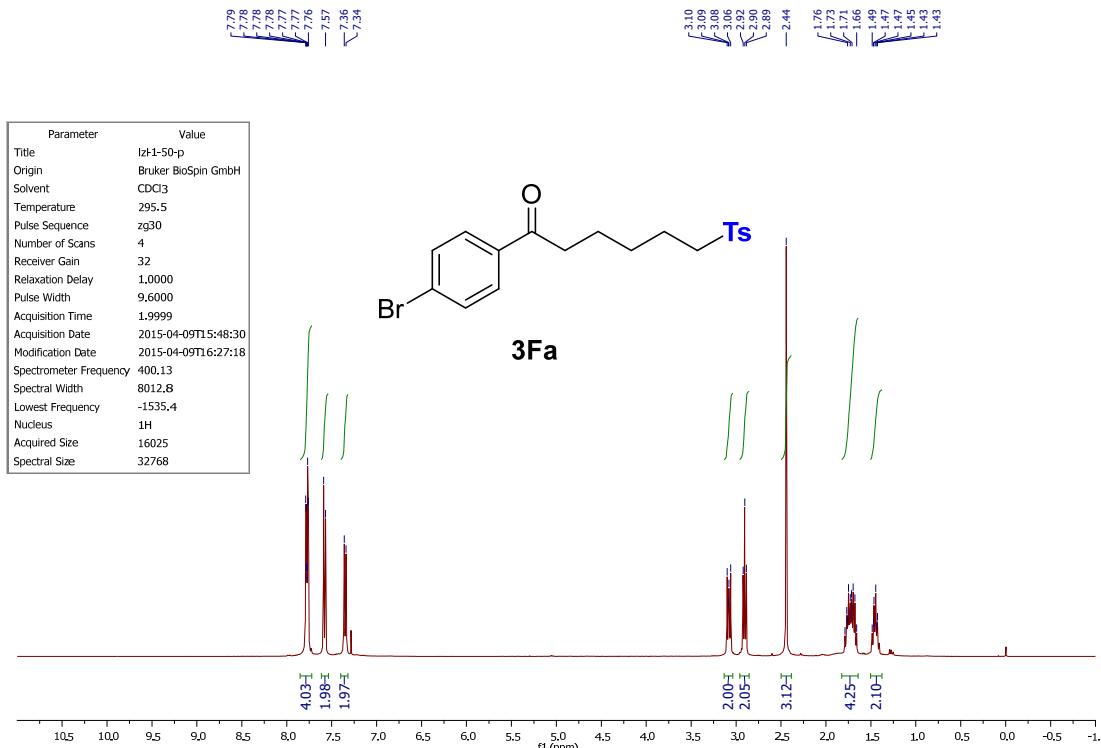


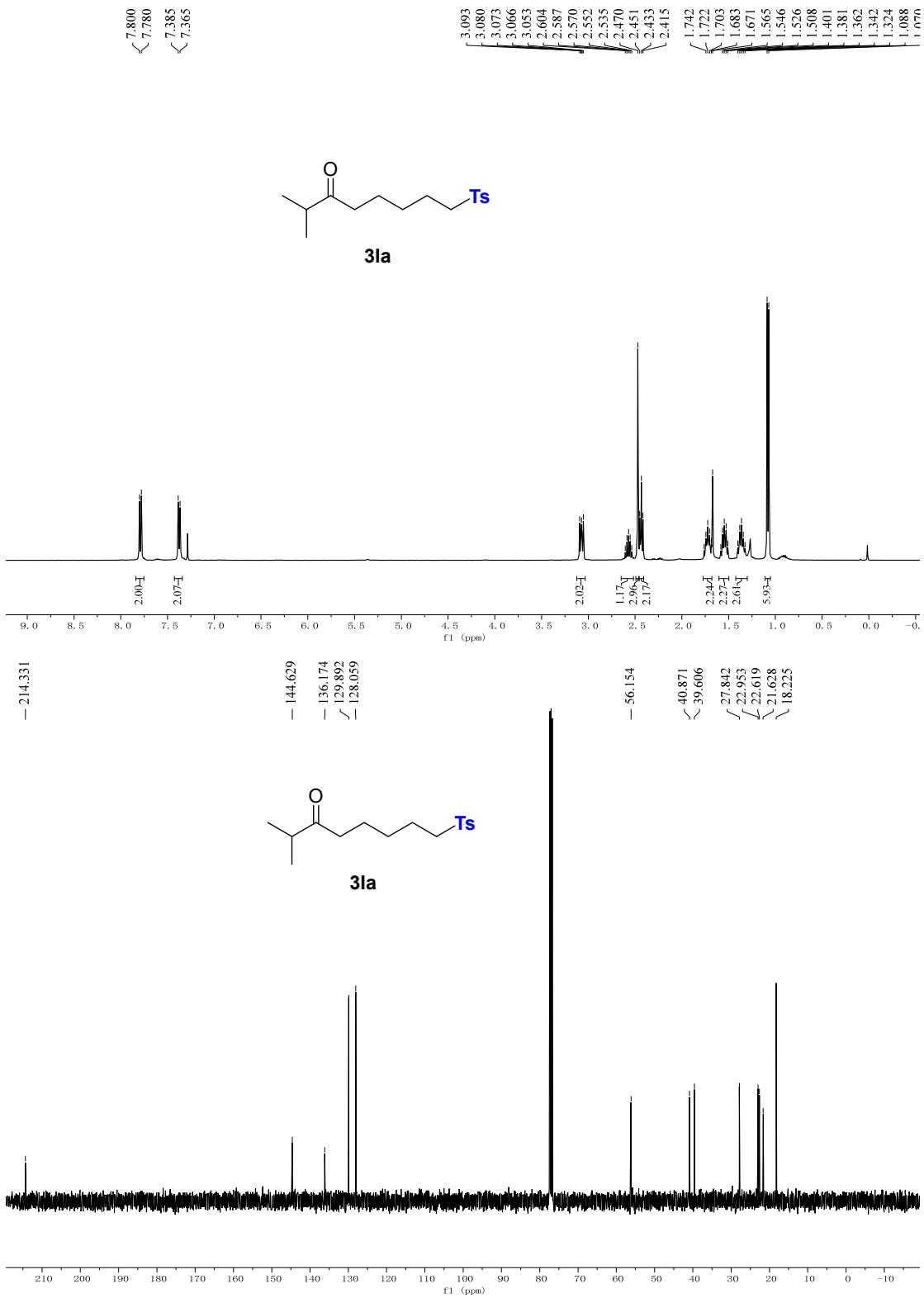


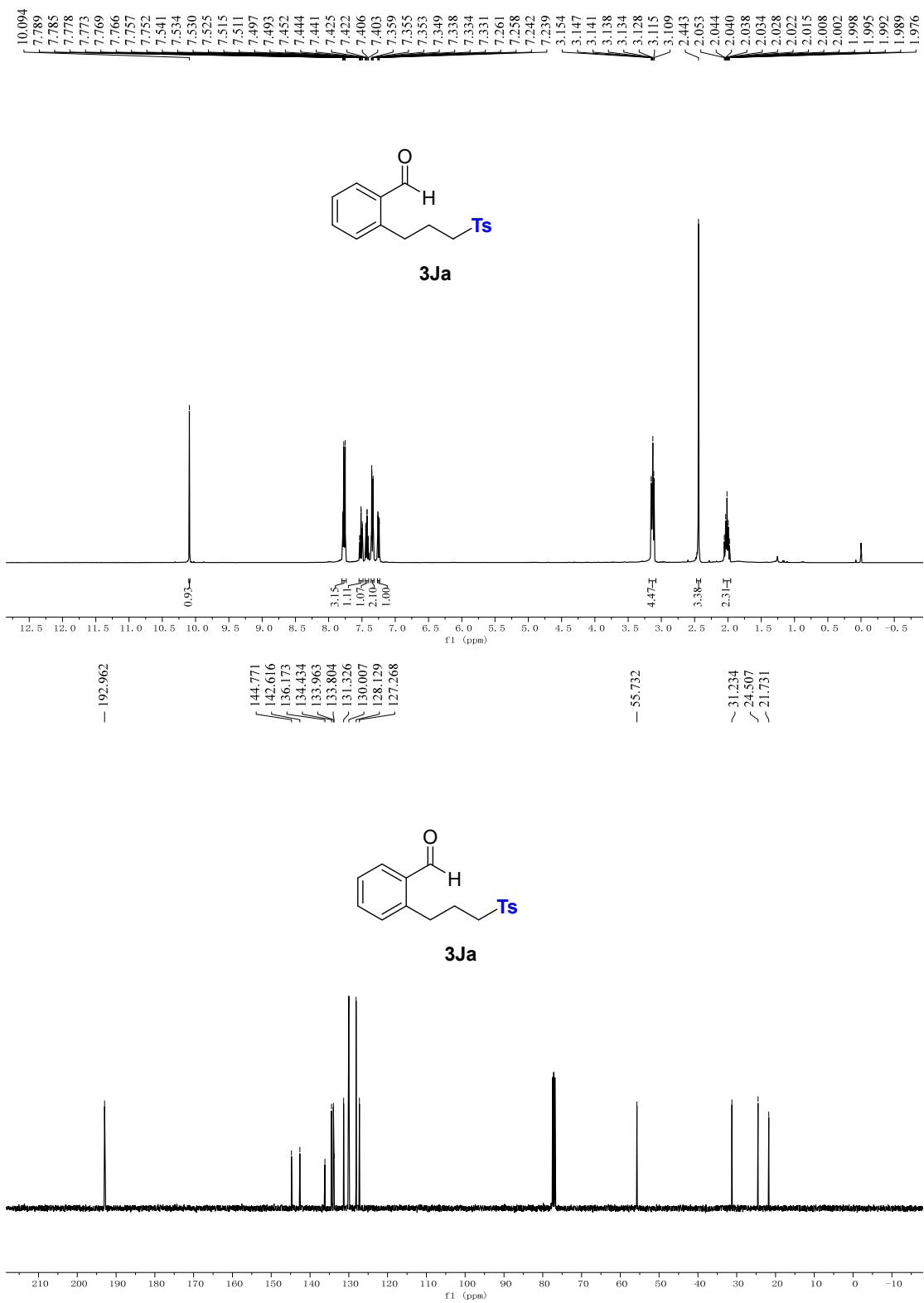


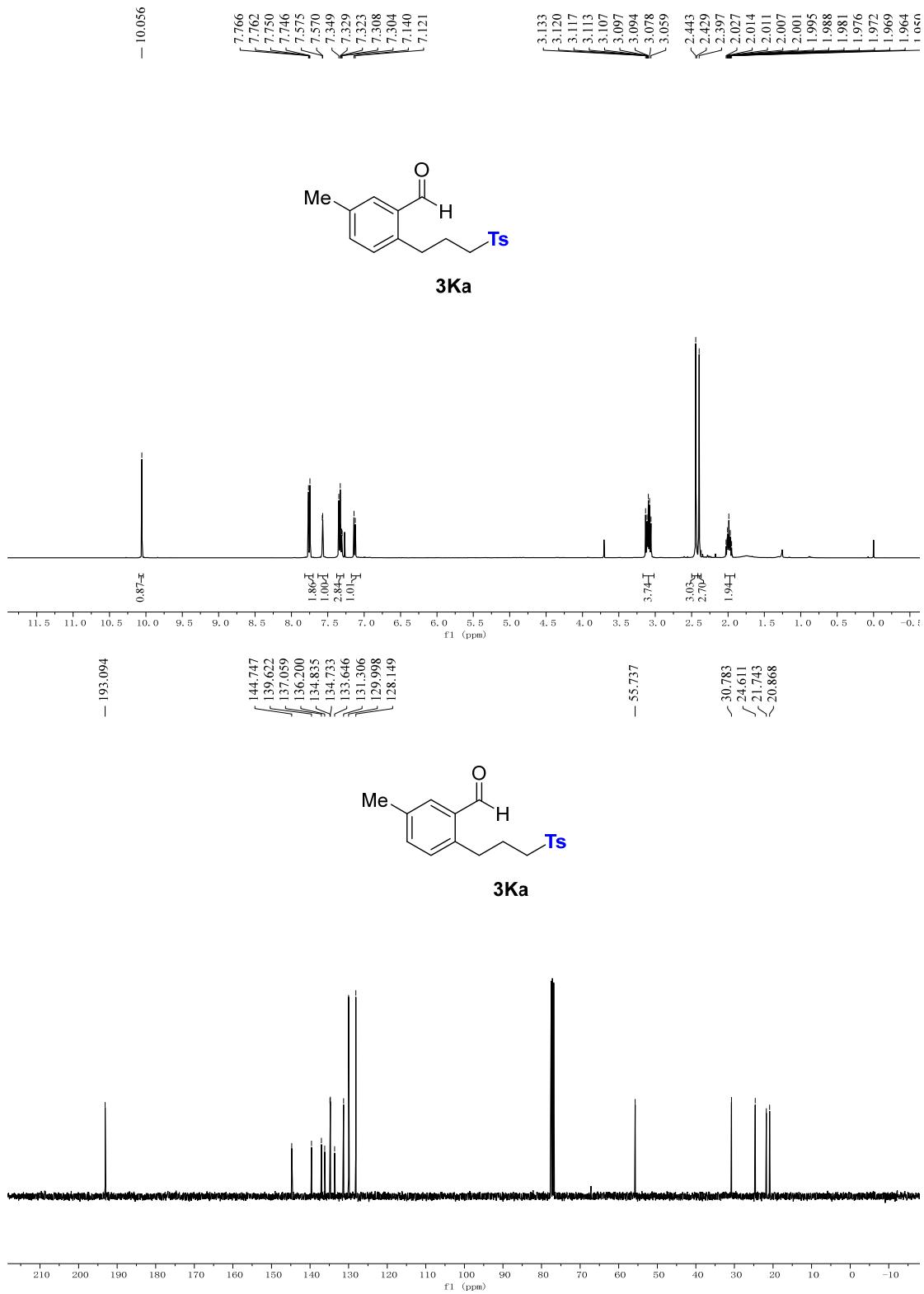


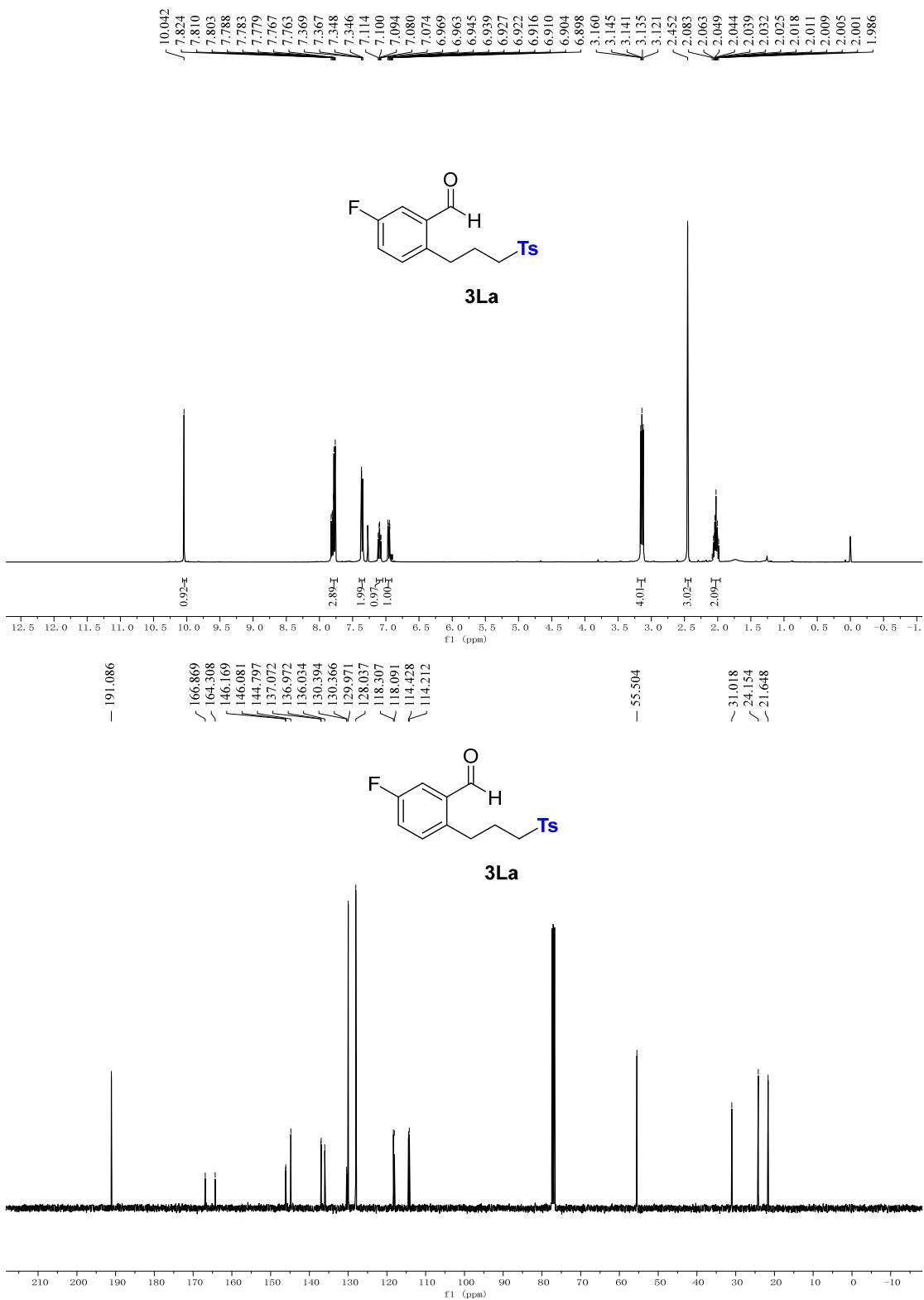


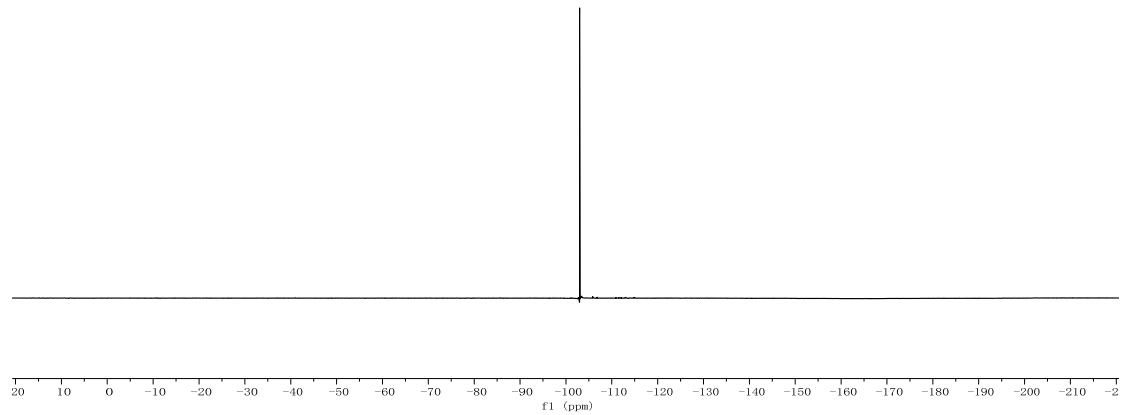
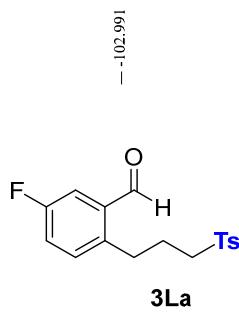




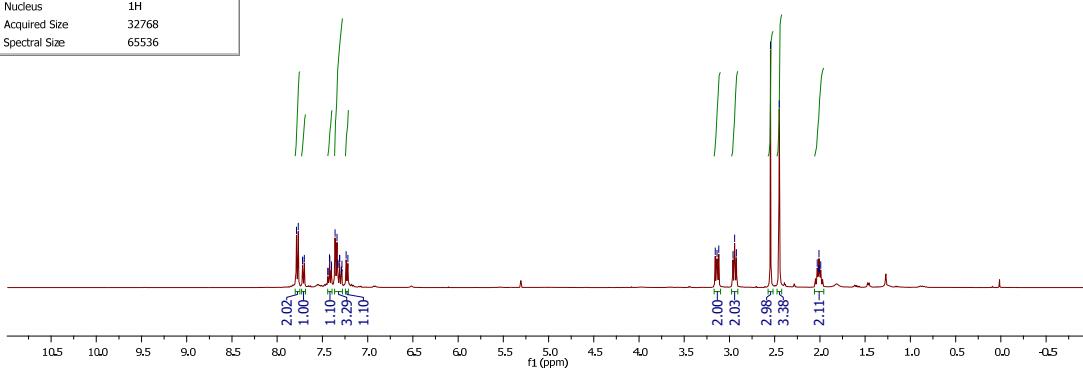
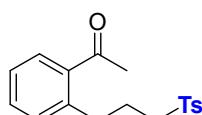






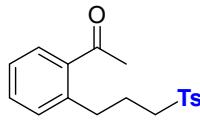


Parameter	Value
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Temperature	296.1
Pulse Sequence	zg30
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Receiver Gain	32
Relaxation Delay	1.0000
Pulse Width	9.6000
Acquisition Time	4.0894
Acquisition Date	2015-03-17T11:14:11
Modification Date	2015-03-17T11:37:16
Spectrometer Frequency	400.13
Spectral Width	8012.8
Lowest Frequency	-1535.4
Nucleus	1H
Acquired Size	32768
Spectral Size	65536

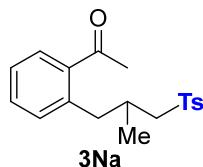
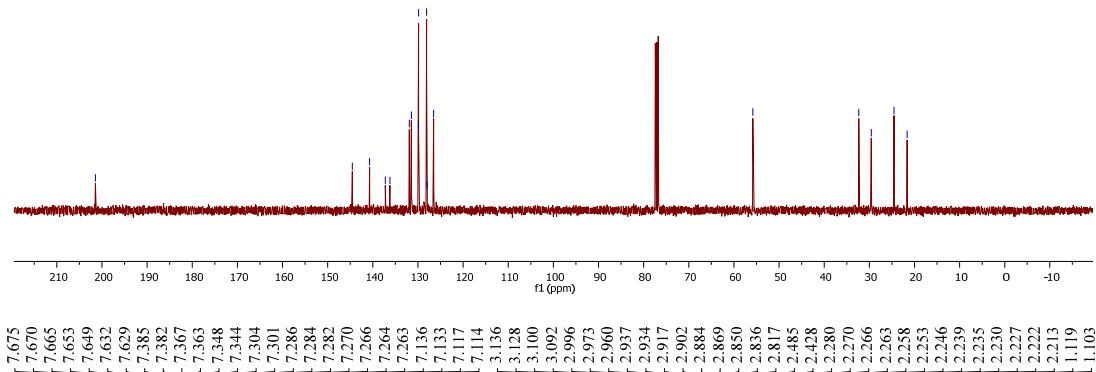


— 201.47

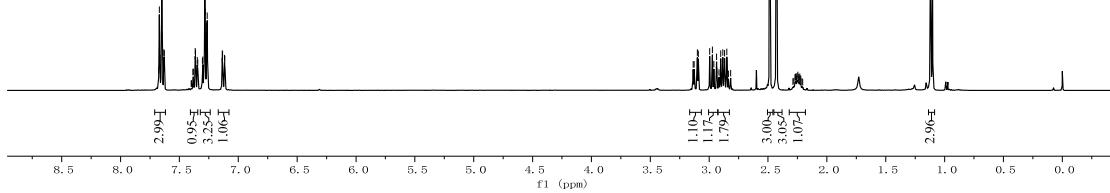
Parameter	Value
Title	IzH-1-13-1p
Origin	Bruker BioSpin GmbH
Solvent	CDCl <sub>3</sub>
Temperature	296.3
Pulse Sequence	zgpg30
Number of Scans	32
Receiver Gain	196
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2015-03-17T11:18:09
Modification Date	2015-03-17T11:13:17
Spectrometer Frequency	100.61
Spectral Width	24038.5
Lowest Frequency	-1958.0
Nucleus	<sup>13</sup> C
Acquired Size	32768
Spectral Size	65536

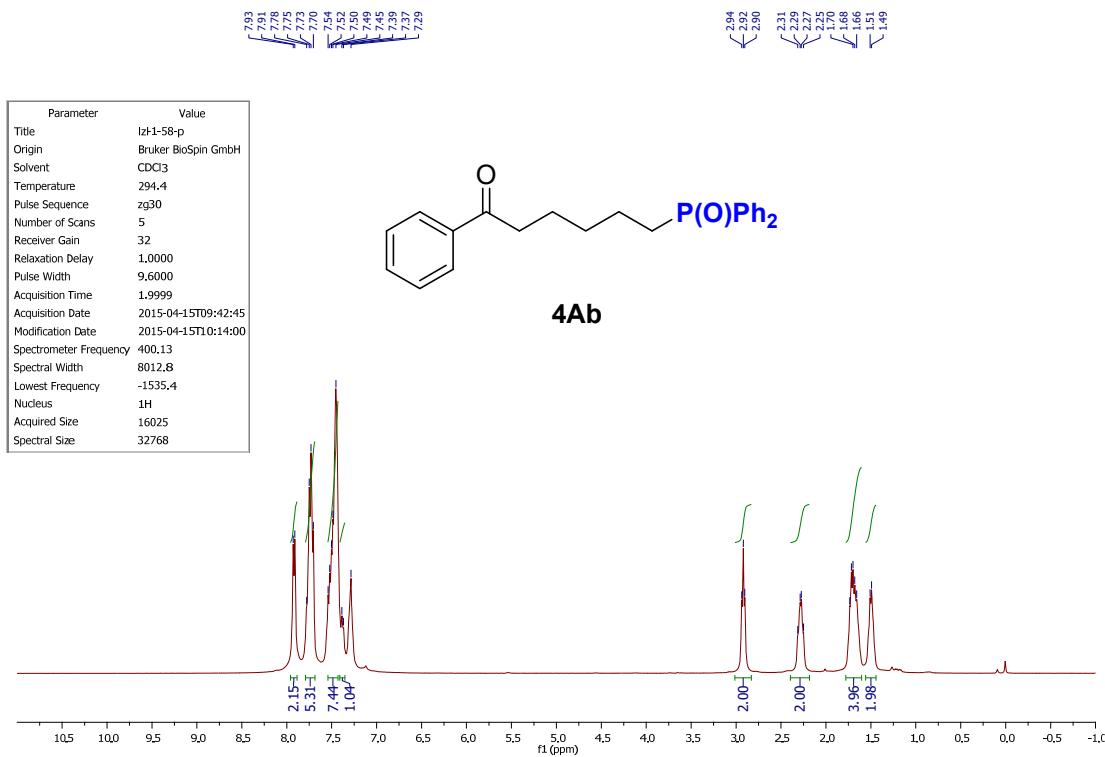
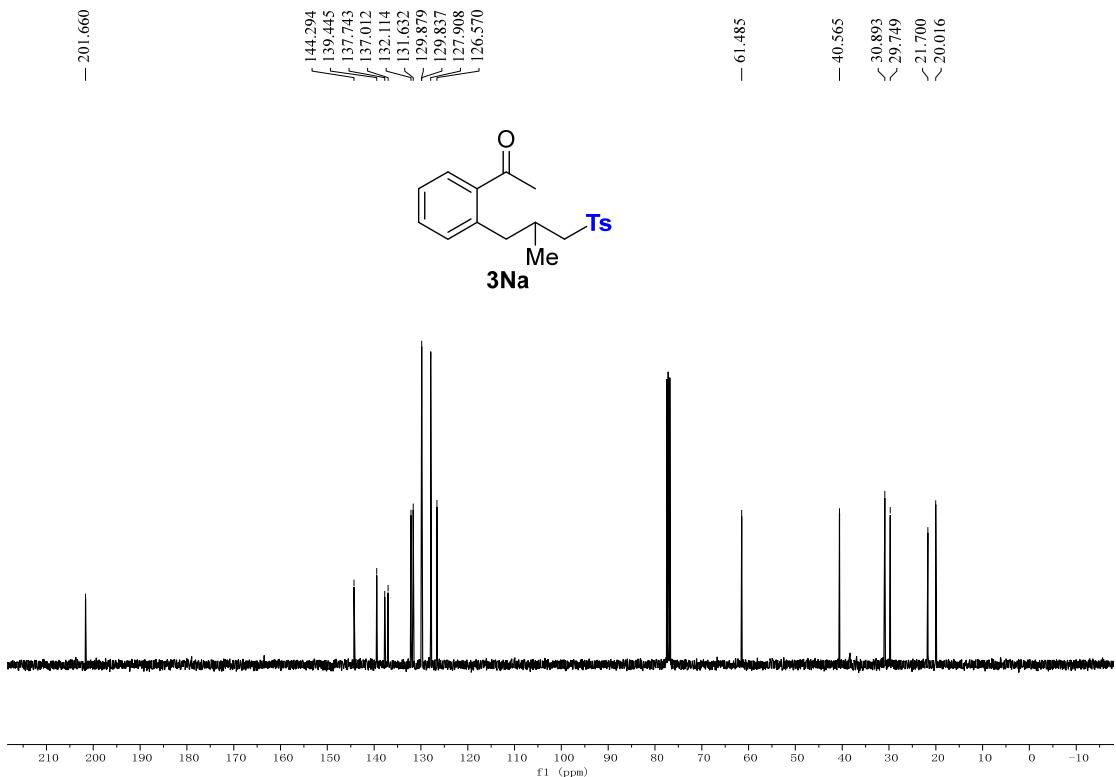


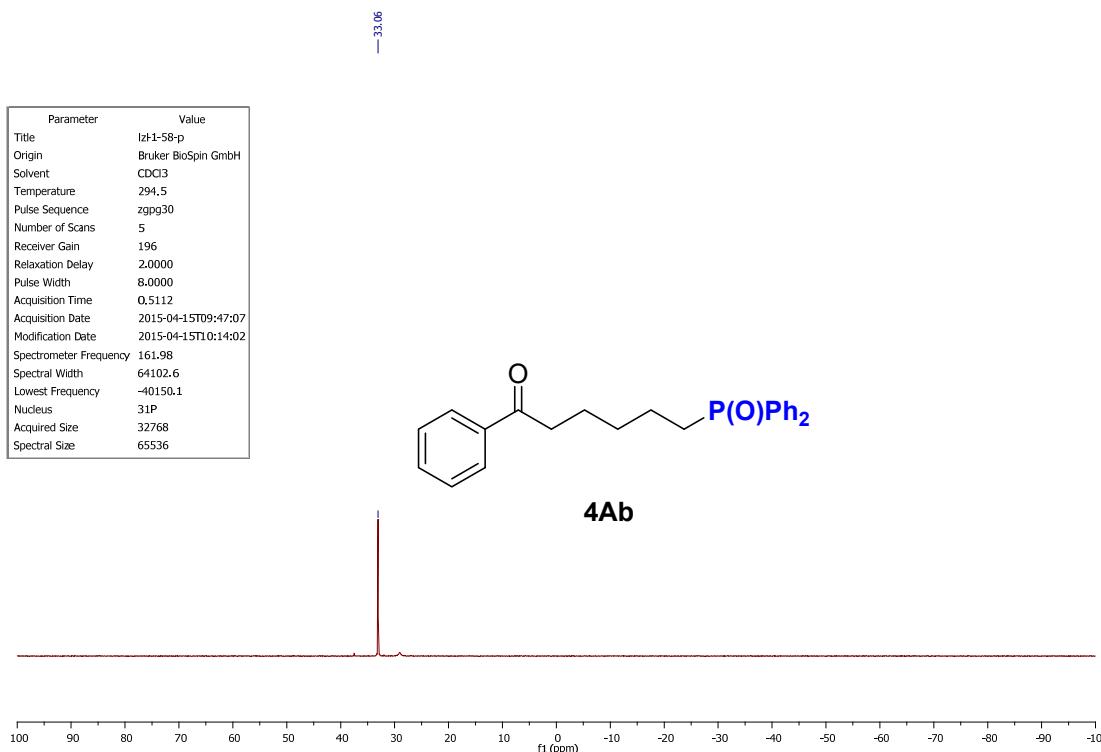
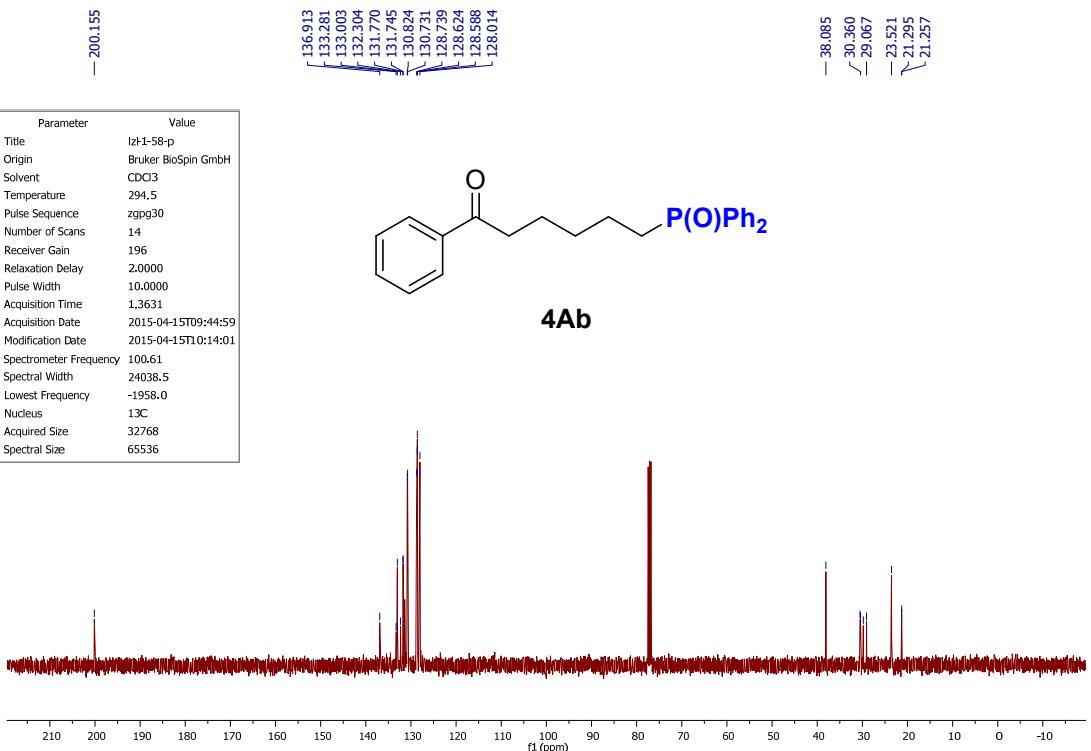
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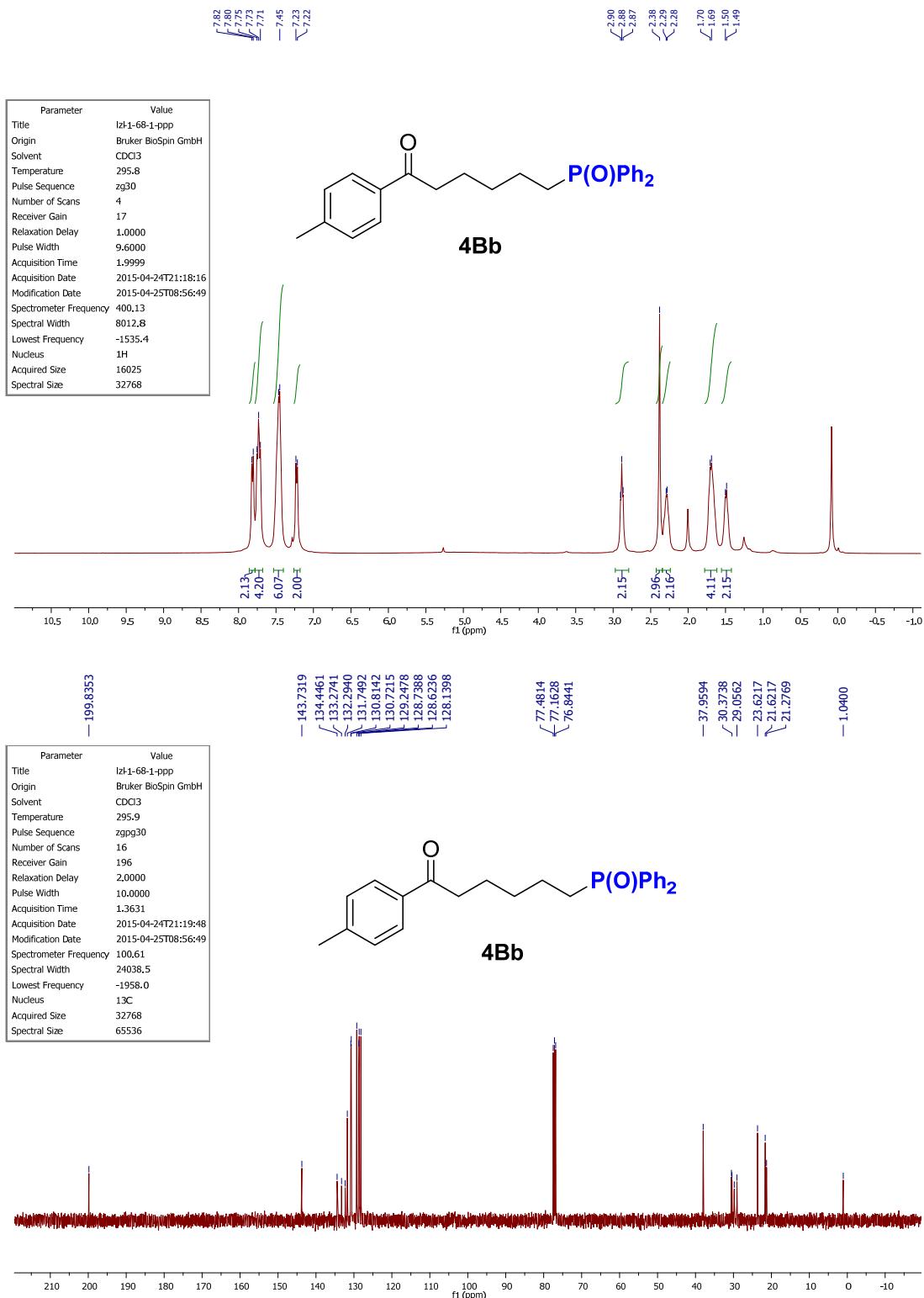


3Na

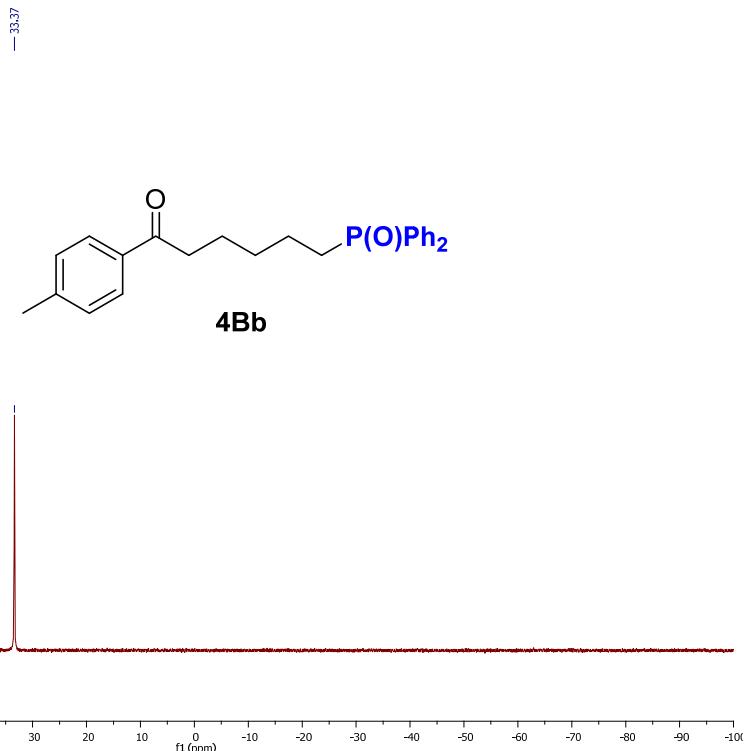




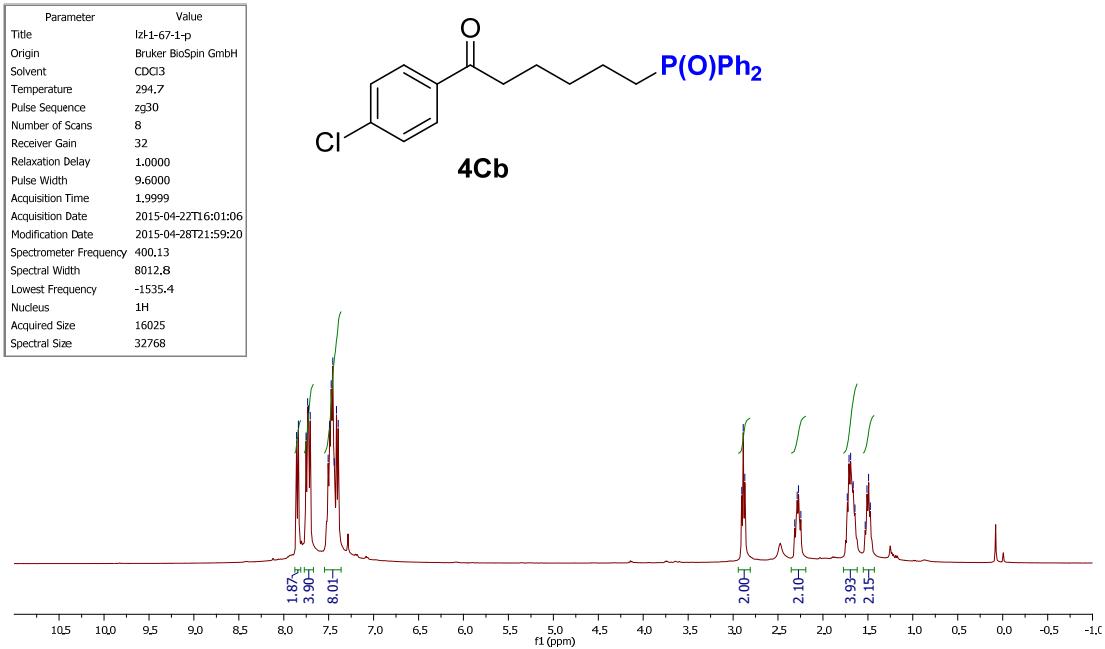


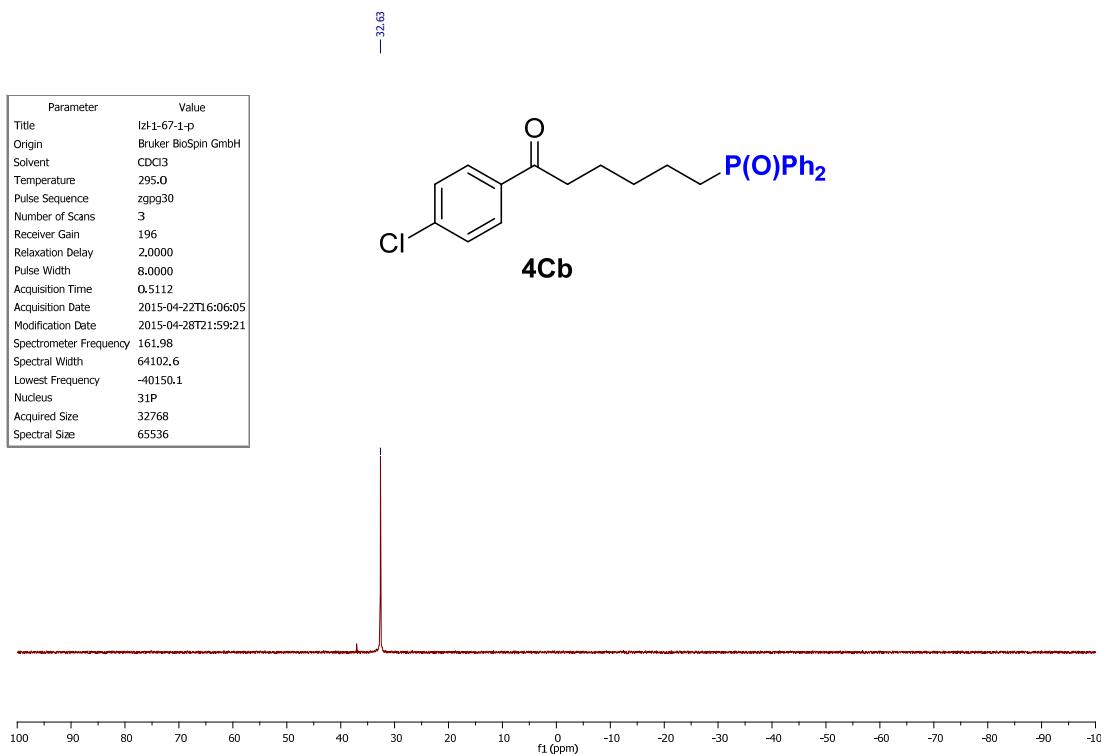
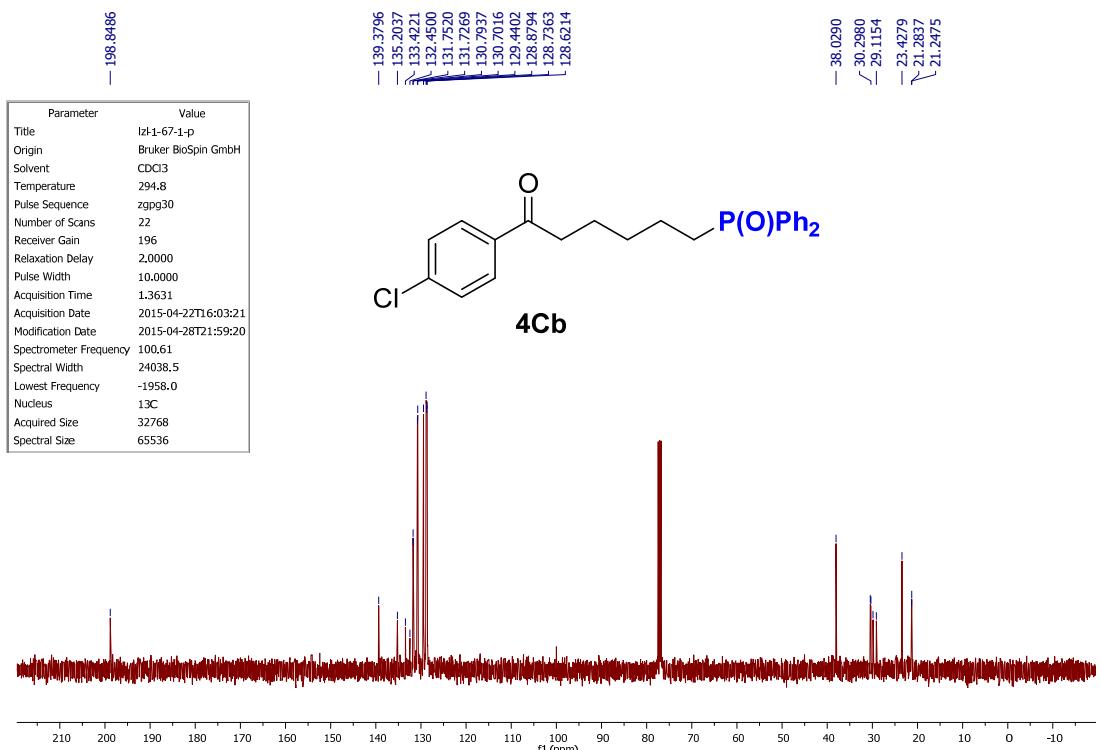


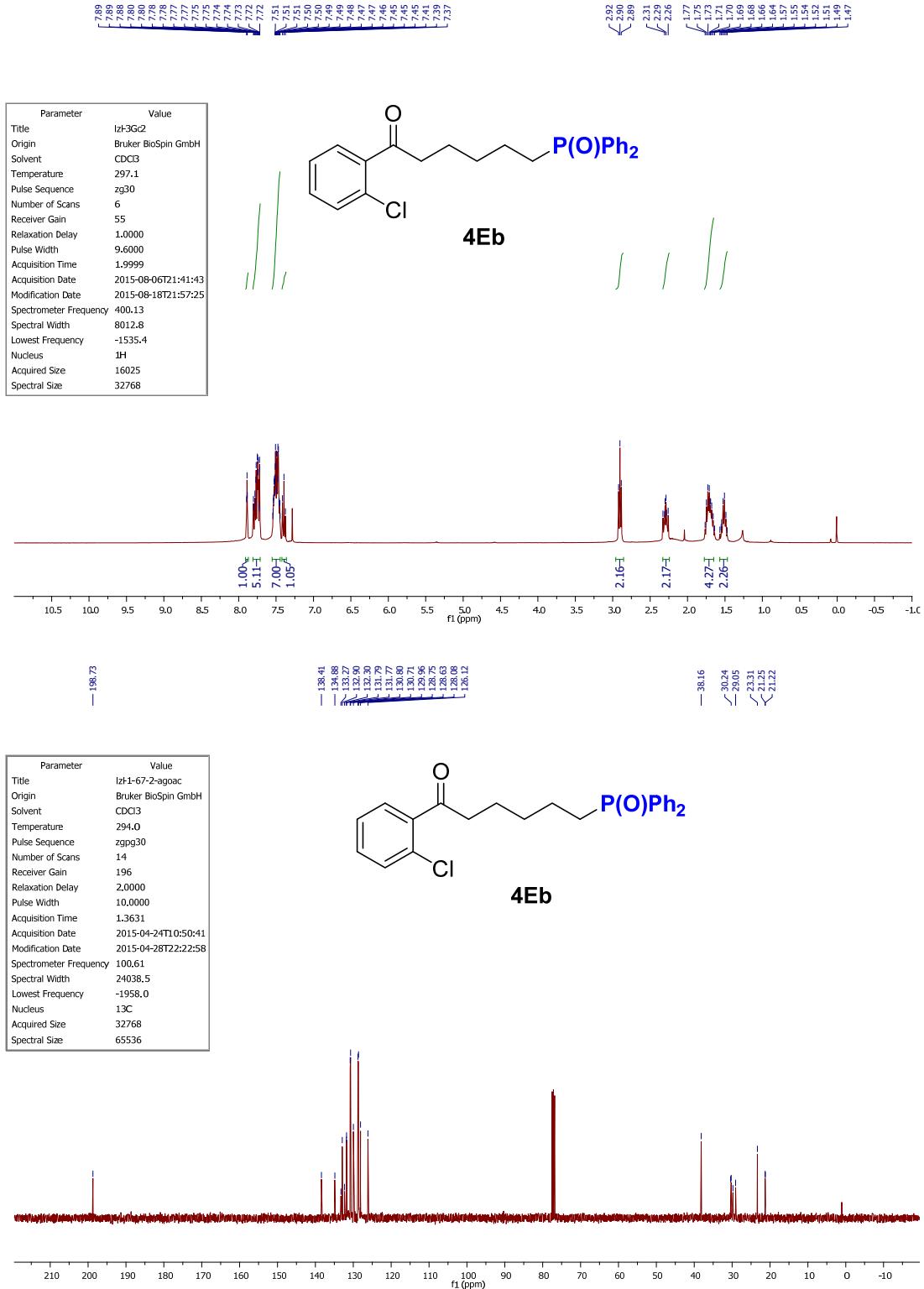
Parameter	Value
Title	lzl1-68-1-pppp
Origin	Bruker BioSpin GmbH
Solvent	CDCl <sub>3</sub>
Temperature	294.0
Pulse Sequence	zgpg30
Number of Scans	3
Receiver Gain	196
Relaxation Delay	2.0000
Pulse Width	8.0000
Acquisition Time	0.5112
Acquisition Date	2015-04-29T21:05:49
Modification Date	2015-04-30T19:30:48
Spectrometer Frequency	161.98
Spectral Width	64102.6
Lowest Frequency	-40150.1
Nucleus	<sup>31</sup> P
Acquired Size	32768
Spectral Size	65536



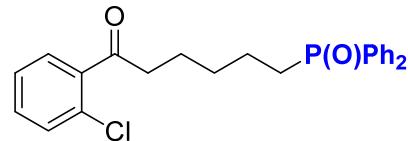
Parameter	Value
Title	lzl1-67-1-p
Origin	Bruker BioSpin GmbH
Solvent	CDCl <sub>3</sub>
Temperature	294.7
Pulse Sequence	zg30
Number of Scans	8
Receiver Gain	32
Relaxation Delay	1.0000
Pulse Width	9.6000
Acquisition Time	1.9999
Acquisition Date	2015-04-22T16:01:06
Modification Date	2015-04-28T21:59:20
Spectrometer Frequency	400.13
Spectral Width	8012.8
Lowest Frequency	-1535.4
Nucleus	<sup>1</sup> H
Acquired Size	16025
Spectral Size	32768



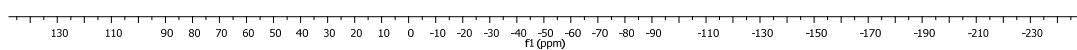




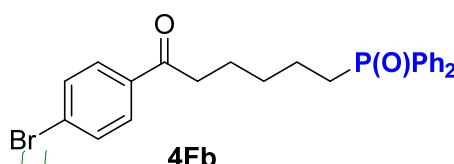
Parameter	Value
Title	IzI-3Gc2
Origin	Bruker BioSpin GmbH
Solvent	CDCB
Temperature	297.2
Pulse Sequence	zgpg30
Number of Scans	4
Receiver Gain	196
Relaxation Delay	2.0000
Pulse Width	8.0000
Acquisition Time	0.5112
Acquisition Date	2015-08-06T21:43:28
Modification Date	2015-08-18T21:57:26
Spectrometer Frequency	161.98
Spectral Width	64102.6
Lowest Frequency	-40150.1
Nucleus	31P
Acquired Size	32768
Spectral Size	65536



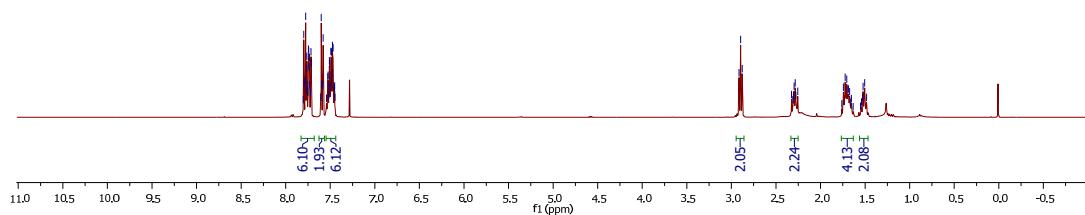
4E**b**

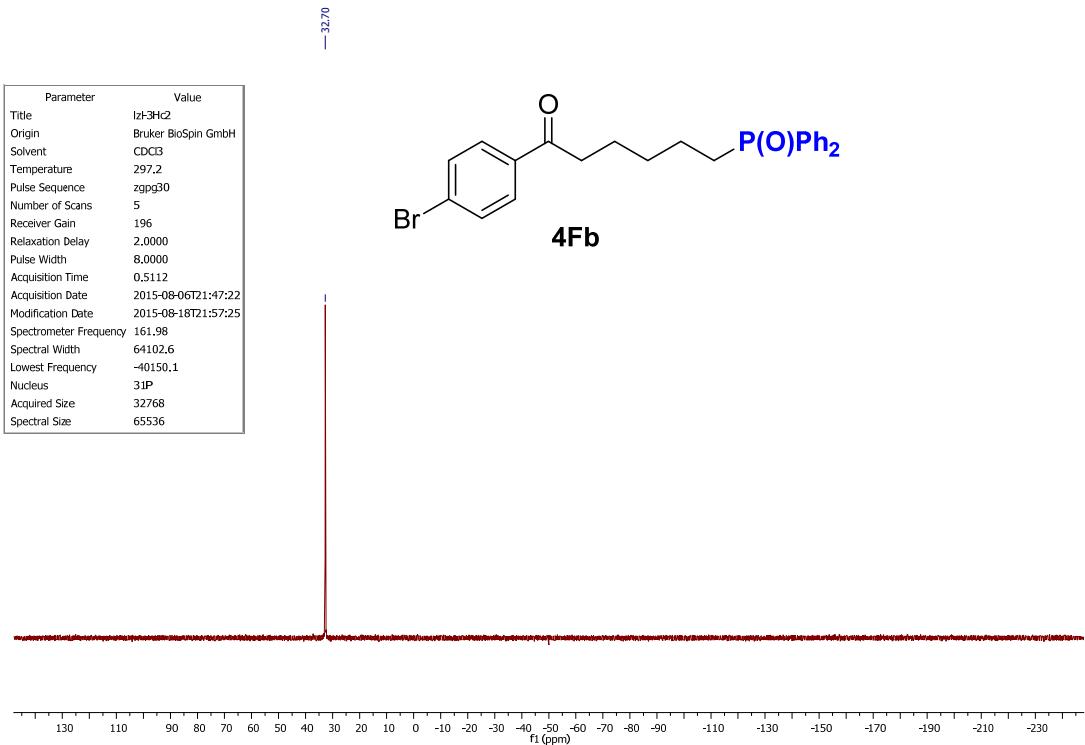
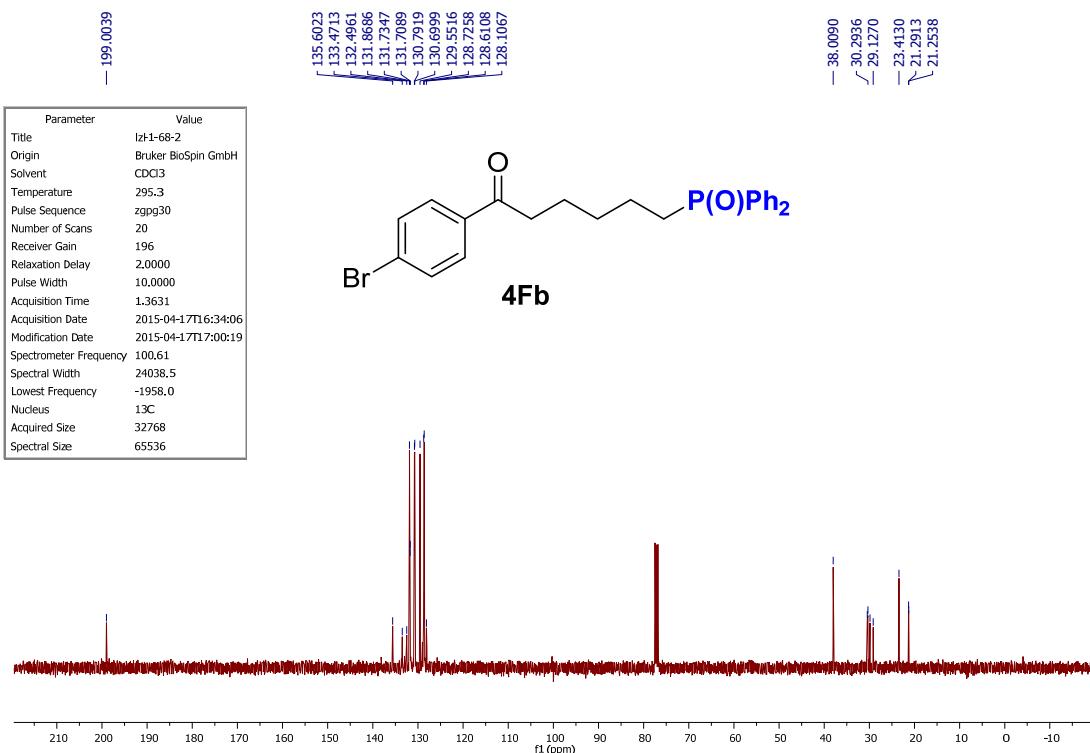


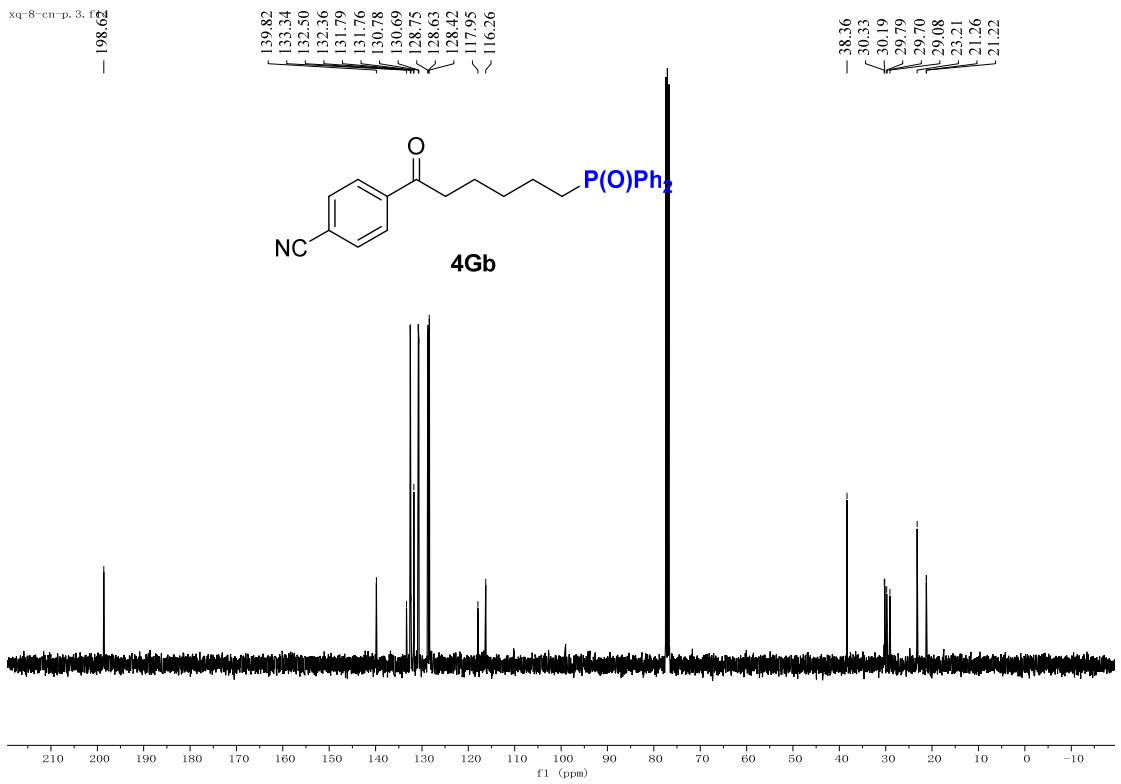
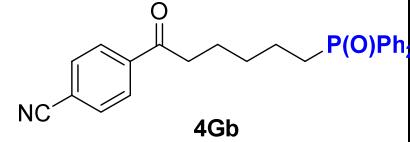
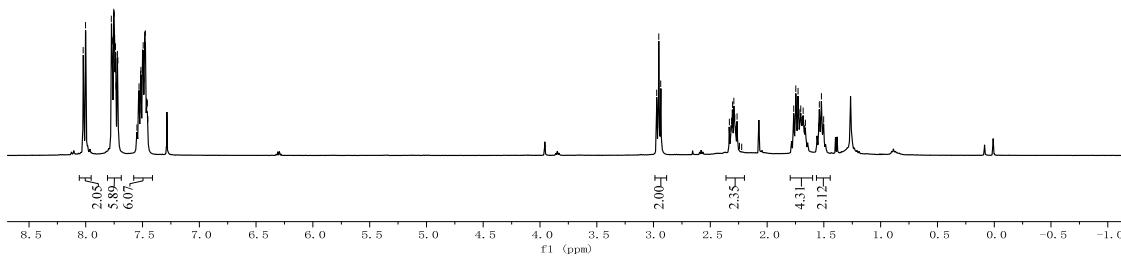
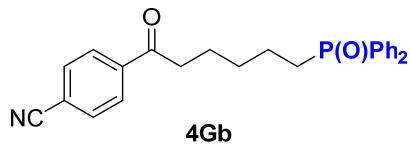
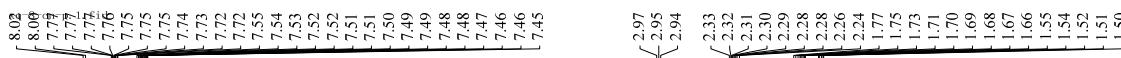
Parameter	Value
Title	IzH3Hc-1
Origin	Bruker BioSpin GmbH
Solvent	CDCB3
Temperature	295.6
Pulse Sequence	zg30
Number of Scans	4
Receiver Gain	62
Relaxation Delay	1.0000
Pulse Width	9.6000
Acquisition Time	1.9999
Acquisition Date	2015-08-06T19:38:33
Modification Date	2015-08-06T19:47:22
Spectrometer Frequency	400.13
Spectral Width	8012.8
Lowest Frequency	-1535.4
Nucleus	1H
Acquired Size	16025
Spectral Size	32768



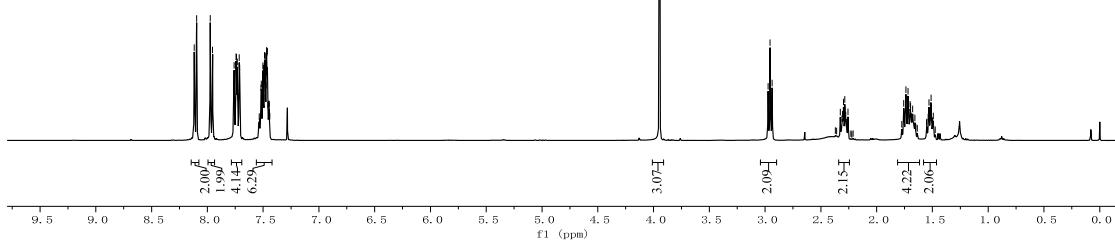
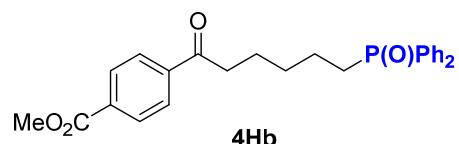
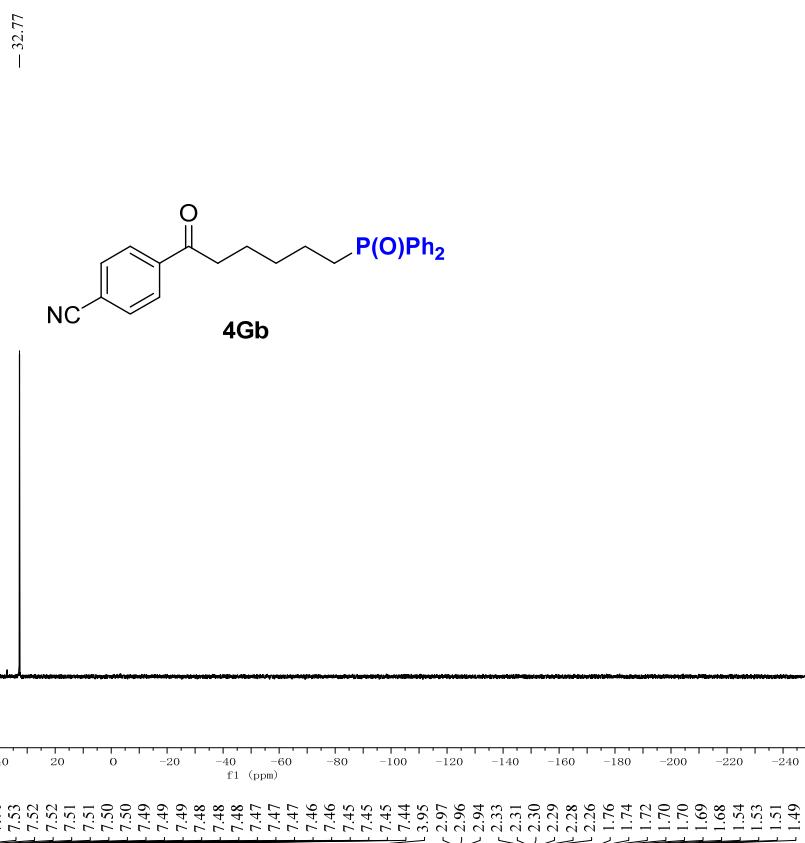
4Fb



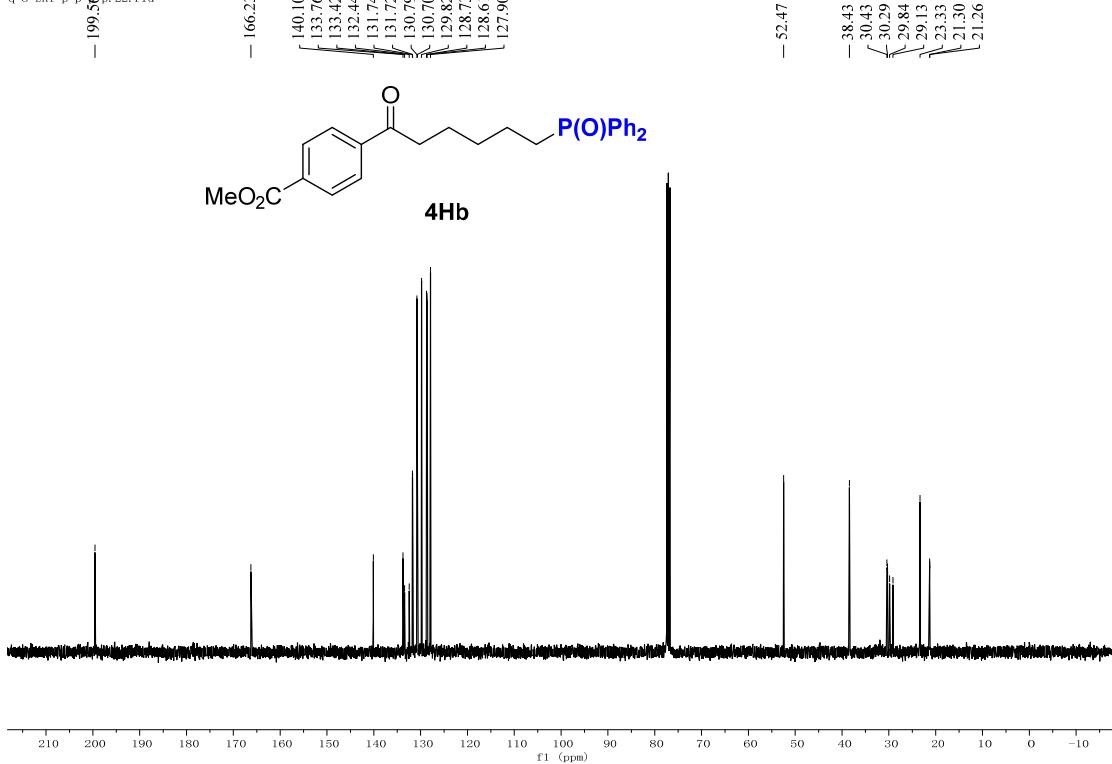




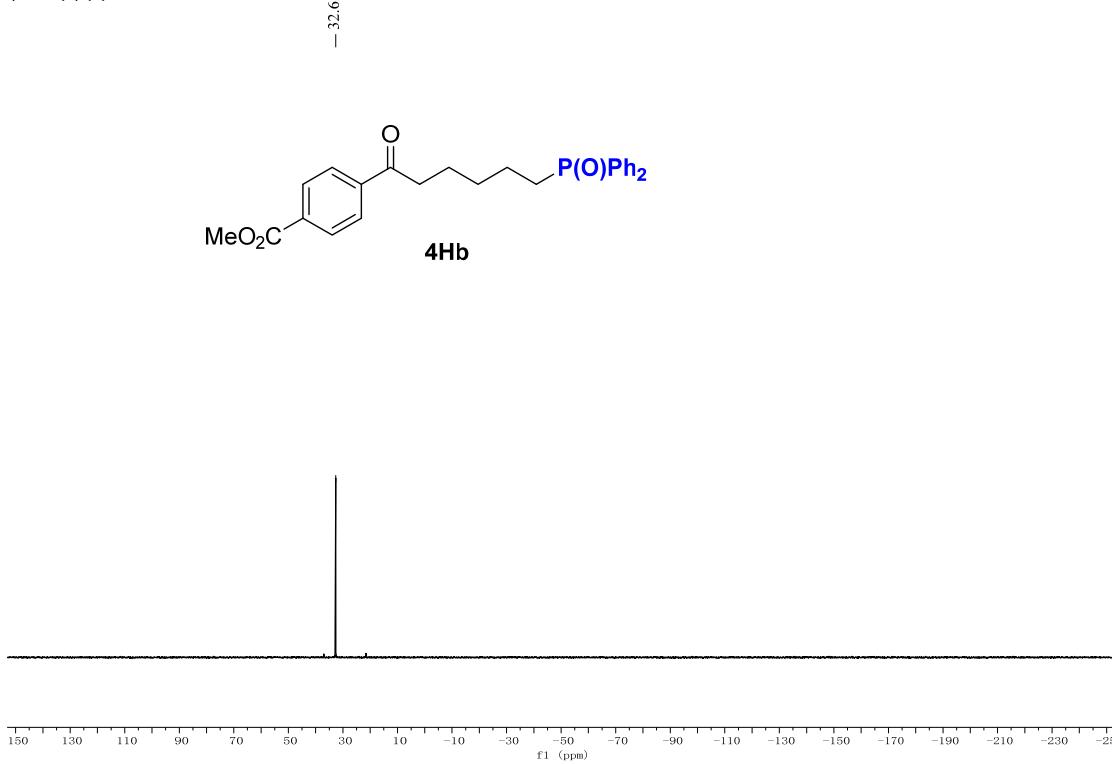
xq-8-cn-p, 2, fid

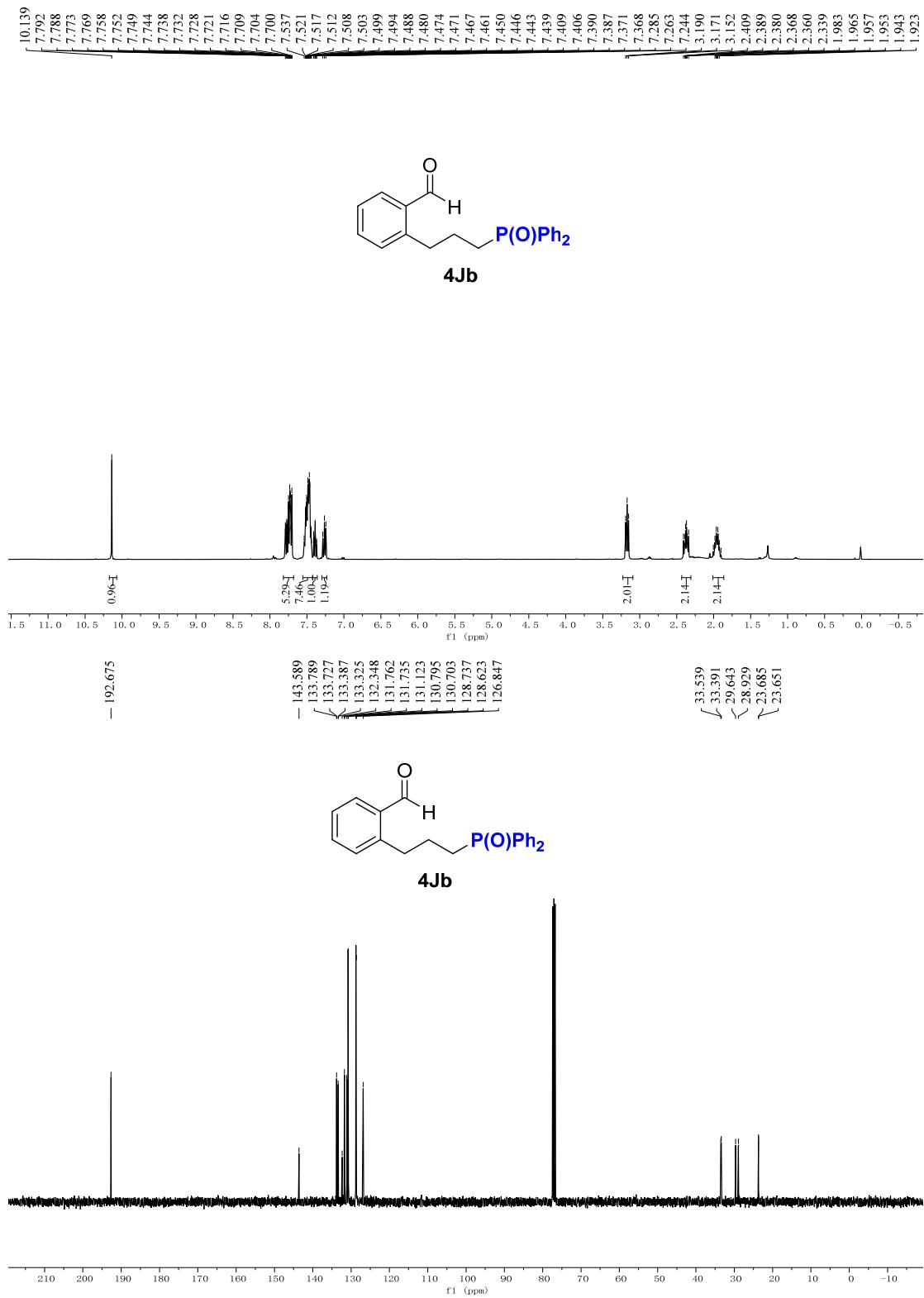


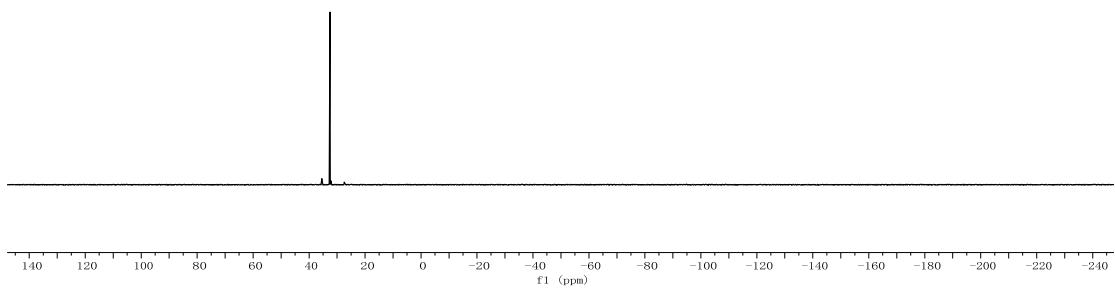
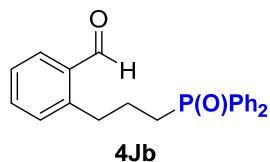
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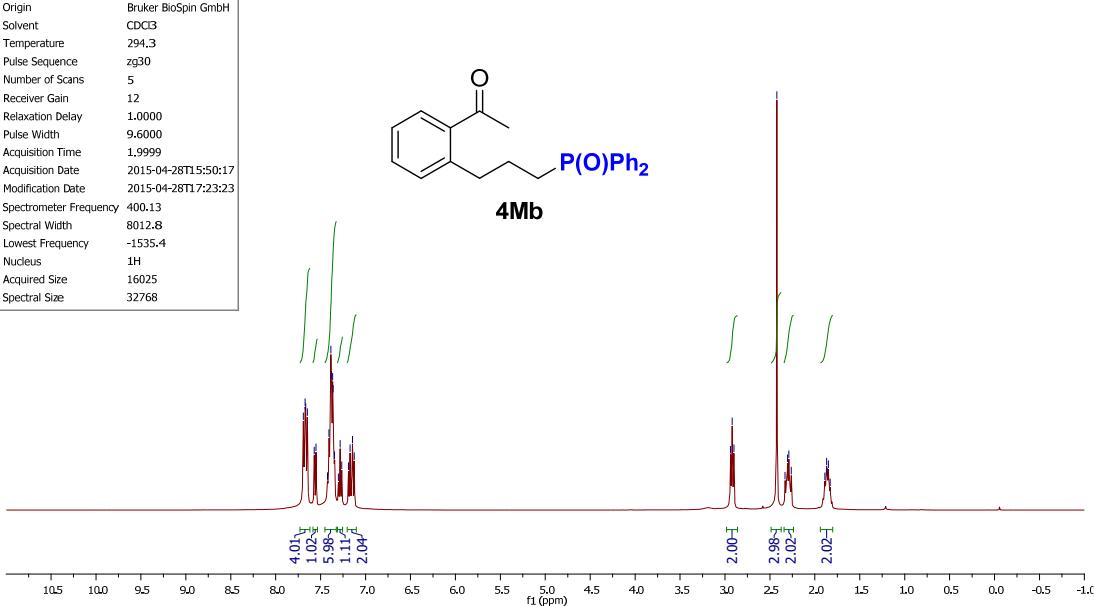
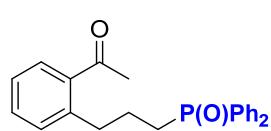
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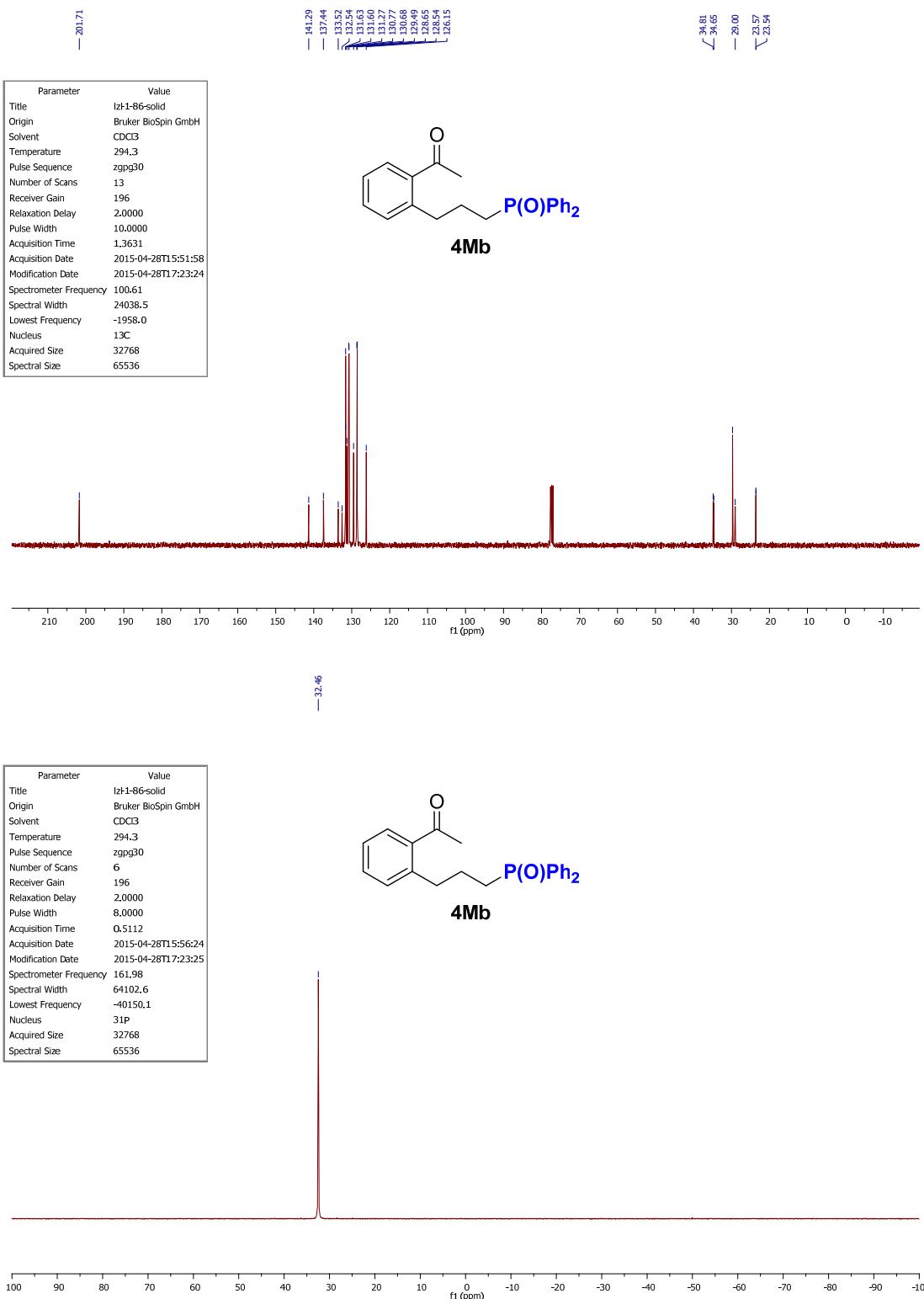


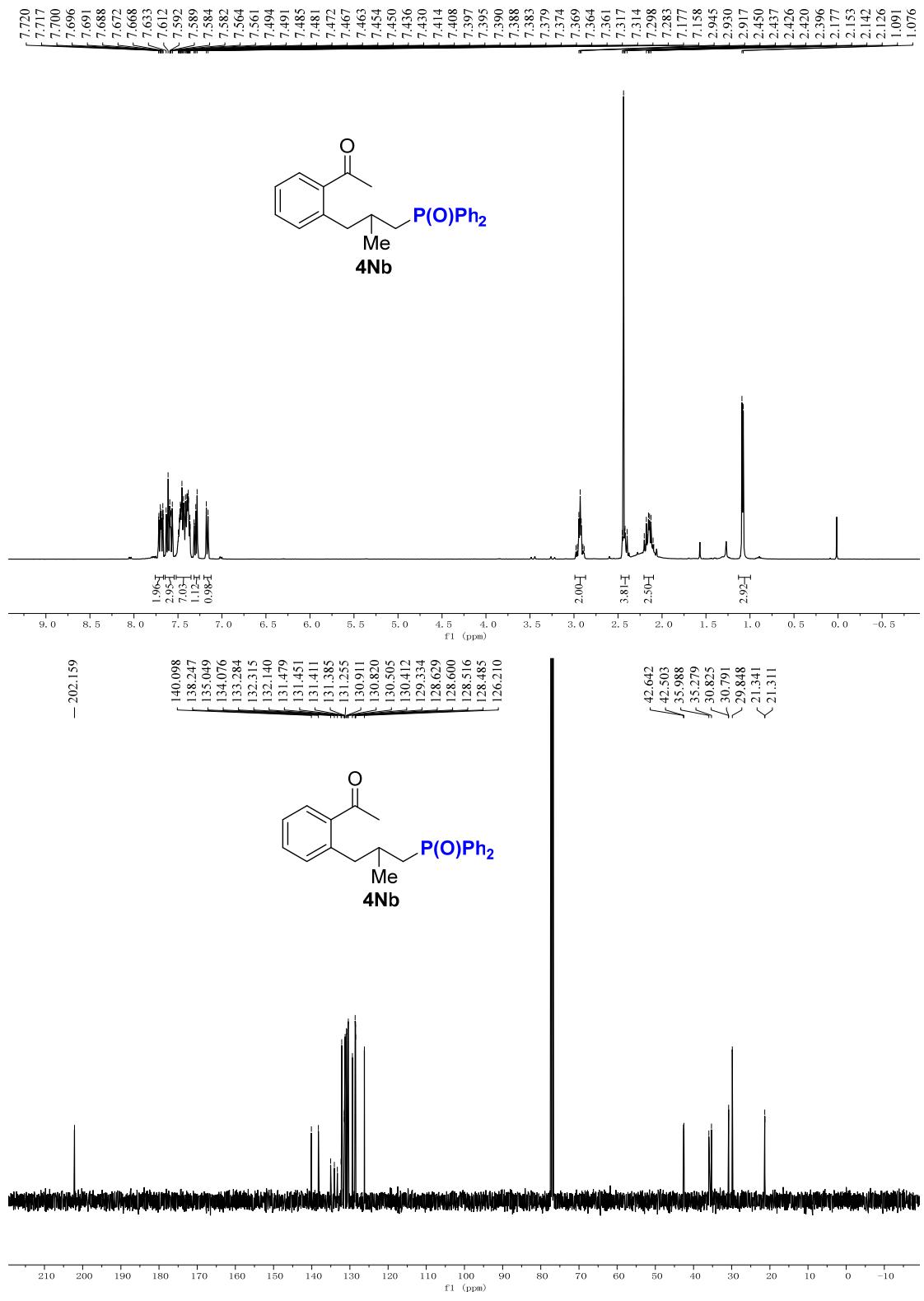




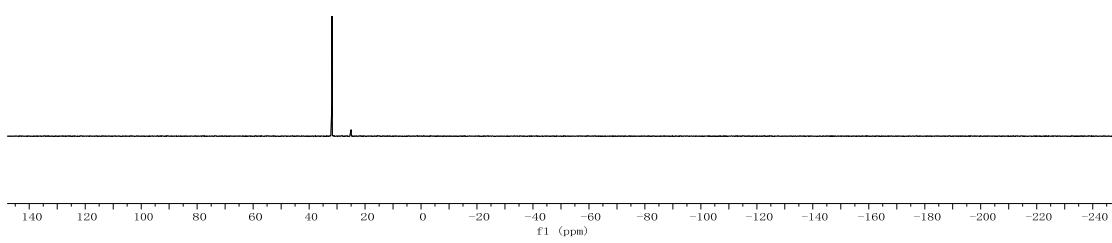
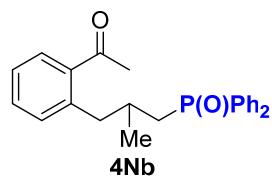
Parameter	Value
Title	lzh1-86-solid
Origin	Bruker BioSpin GmbH
Solvent	CDC13
Temperature	294.3
Pulse Sequence	zg30
Number of Scans	5
Receiver Gain	12
Relaxation Delay	1.0000
Pulse Width	9.6000
Acquisition Time	1.9999
Acquisition Date	2015-04-28T15:50:17
Modification Date	2015-04-28T17:23:23
Spectrometer Frequency	400.13
Spectral Width	8012.8
Lowest Frequency	-1535.4
Nucleus	1H
Acquired Size	16025
Spectral Size	32768



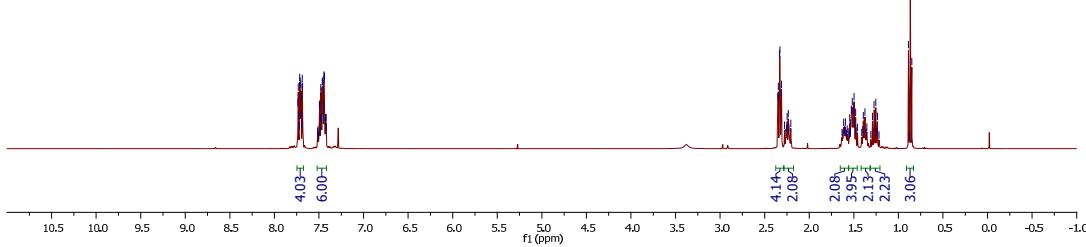
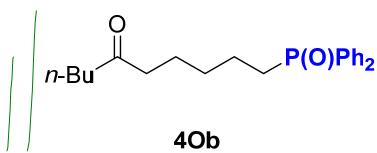




-31.821



Parameter	Value
Title	IzI-1-170-1-p
Origin	Bruker BioSpin GmbH
Solvent	$\text{CDCl}_3$
Temperature	294.4
Pulse Sequence	zg30
Number of Scans	4
Receiver Gain	32
Relaxation Delay	1.0000
Pulse Width	9.6000
Acquisition Time	1.9999
Acquisition Date	2015-07-03T11:13:27
Modification Date	2015-07-03T16:08:59
Spectrometer Frequency	400.13
Spectral Width	8012.8
Lowest Frequency	-1535.4
Nucleus	$^1\text{H}$
Acquired Size	16025
Spectral Size	32768

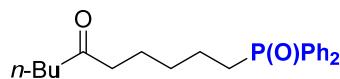


— 211.26

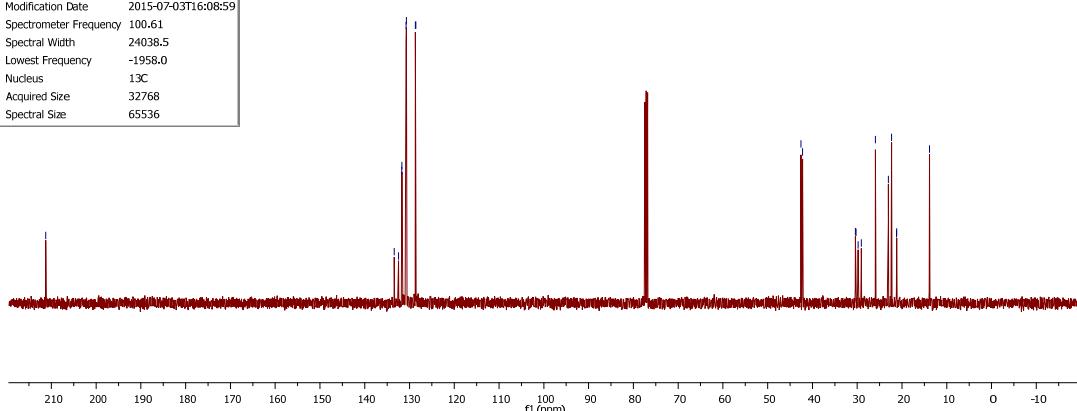
133.42  
133.44  
131.72  
131.69  
131.69  
131.69  
128.71  
128.59

42.57  
42.20  
30.41  
29.80  
29.69  
25.92  
22.32  
21.19  
— 13.86

Parameter	Value
Title	Iz-1-170-1-p
Origin	Bruker BioSpin GmbH
Solvent	CDCl <sub>3</sub>
Temperature	294.5
Pulse Sequence	zpgpg30
Number of Scans	20
Receiver Gain	196
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2015-07-03T11:15:20
Modification Date	2015-07-03T16:08:59
Spectrometer Frequency	100.61
Spectral Width	24038.5
Lowest Frequency	-1958.0
Nucleus	<sup>13</sup> C
Acquired Size	32768
Spectral Size	65536

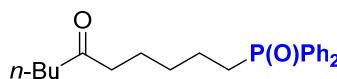


**4Ob**

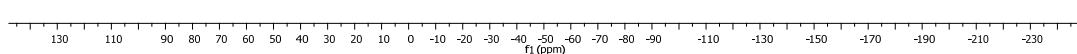


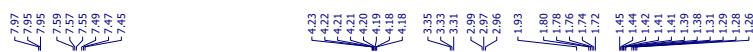
— 32.70

Parameter	Value
Title	Iz-1-170-1-p
Origin	Bruker BioSpin GmbH
Solvent	CDCl <sub>3</sub>
Temperature	294.5
Pulse Sequence	zpgpg30
Number of Scans	5
Receiver Gain	196
Relaxation Delay	2.0000
Pulse Width	8.0000
Acquisition Time	0.5112
Acquisition Date	2015-07-03T11:17:44
Modification Date	2015-07-03T16:09:00
Spectrometer Frequency	161.98
Spectral Width	64102.6
Lowest Frequency	-40150.1
Nucleus	<sup>31</sup> P
Acquired Size	32768
Spectral Size	65536

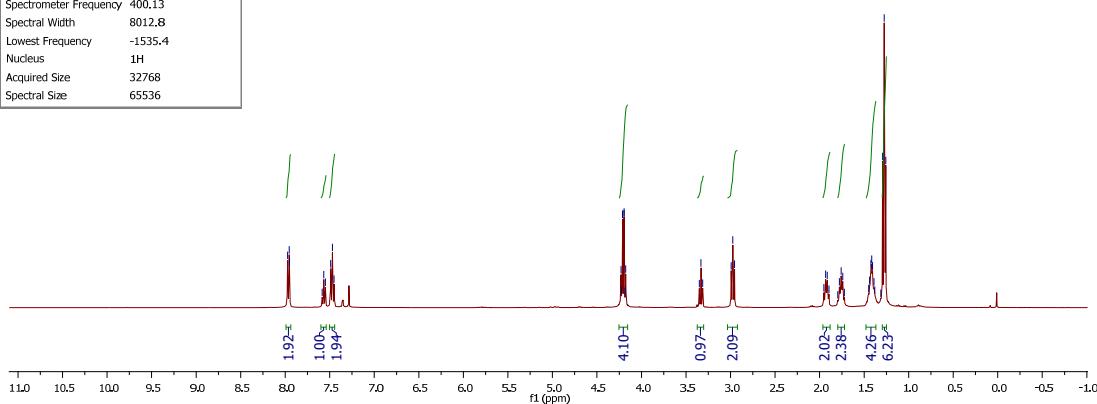


**4Ob**

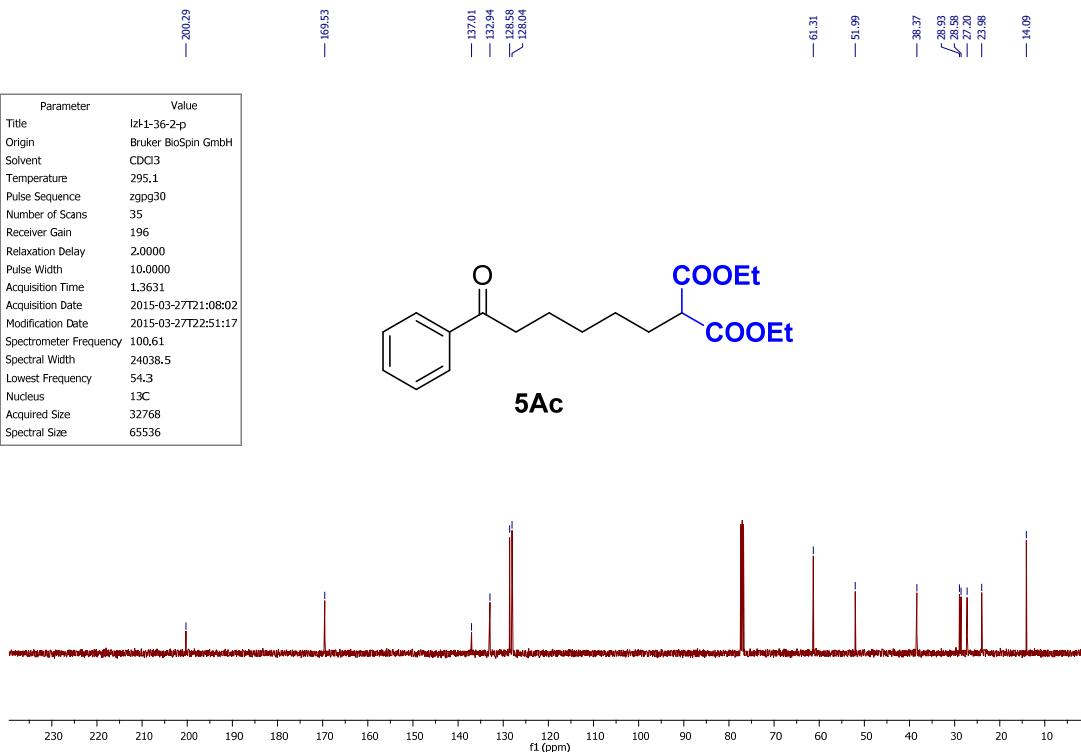


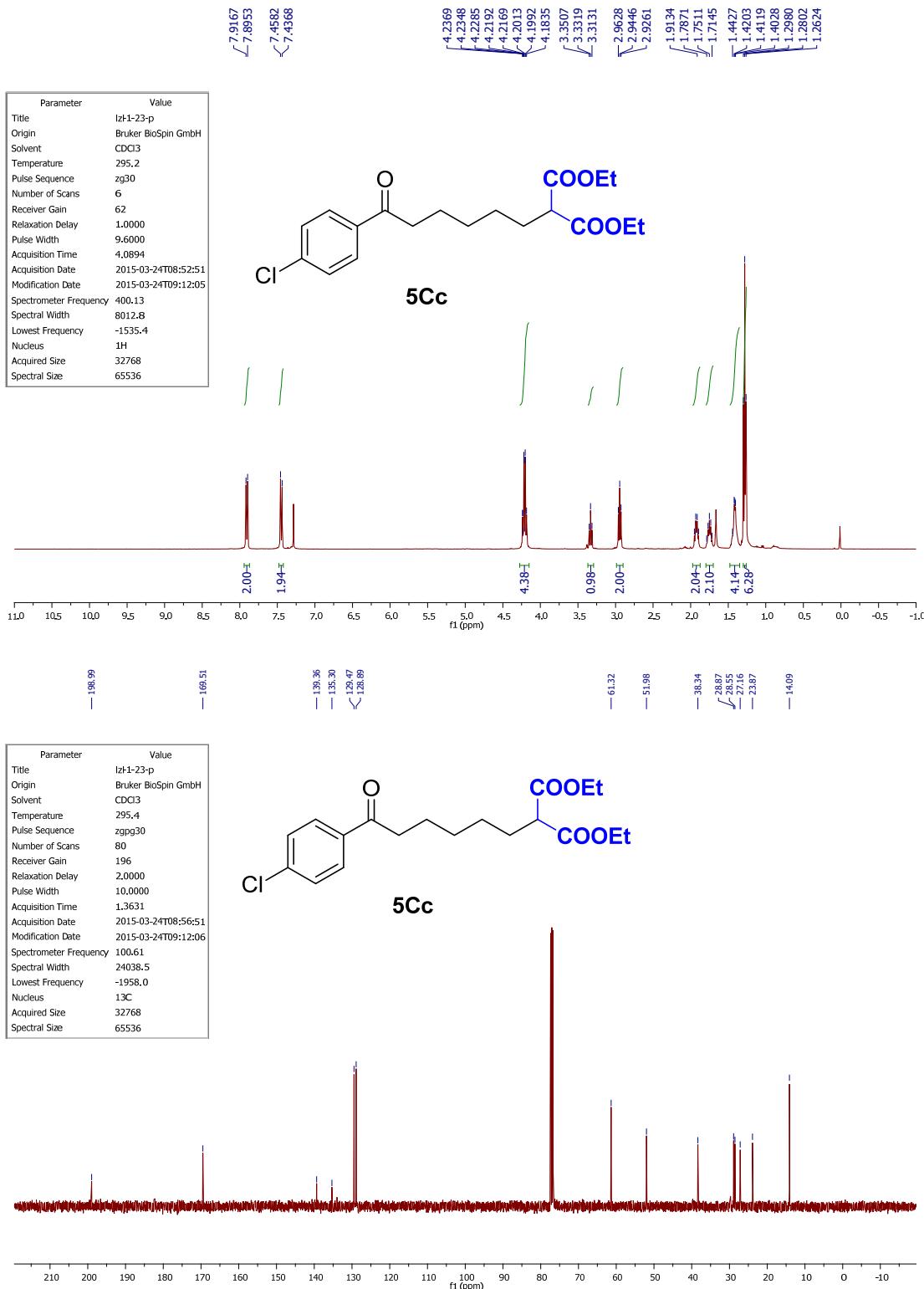


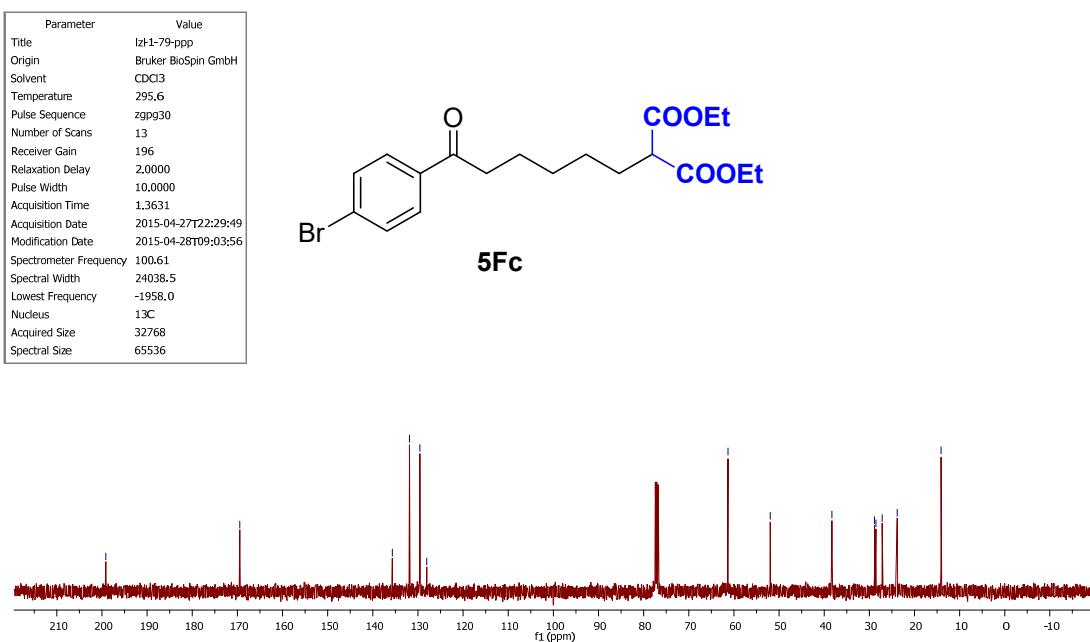
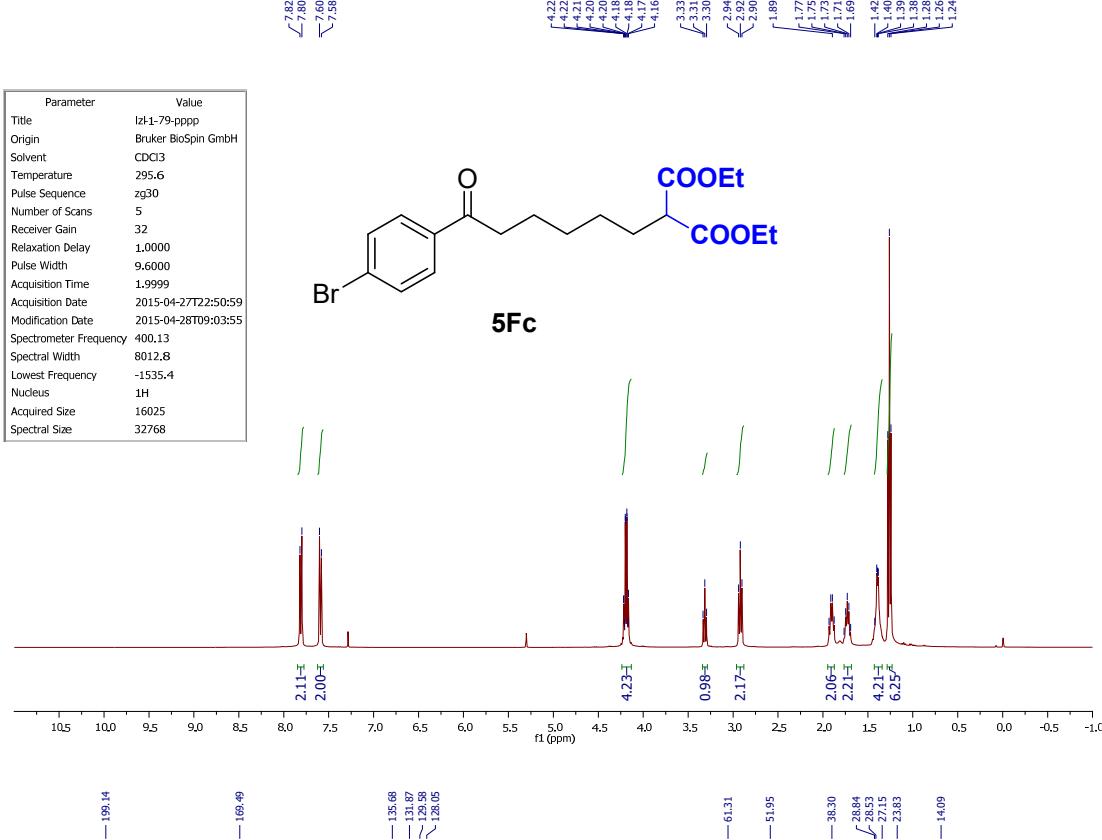
Parameter	Value
Title	IzI-1-36-2-p
Origin	Bruker BioSpin GmbH
Solvent	CDCl <sub>3</sub>
Temperature	295.0
Pulse Sequence	zg30
Number of Scans	5
Receiver Gain	32
Relaxation Delay	1.0000
Pulse Width	9.6000
Acquisition Time	4.0894
Acquisition Date	2015-03-27T21:06:01
Modification Date	2015-03-27T22:51:16
Spectrometer Frequency	400.13
Spectral Width	8012.8
Lowest Frequency	-1535.4
Nucleus	<sup>1</sup> H
Acquired Size	32768
Spectral Size	65536

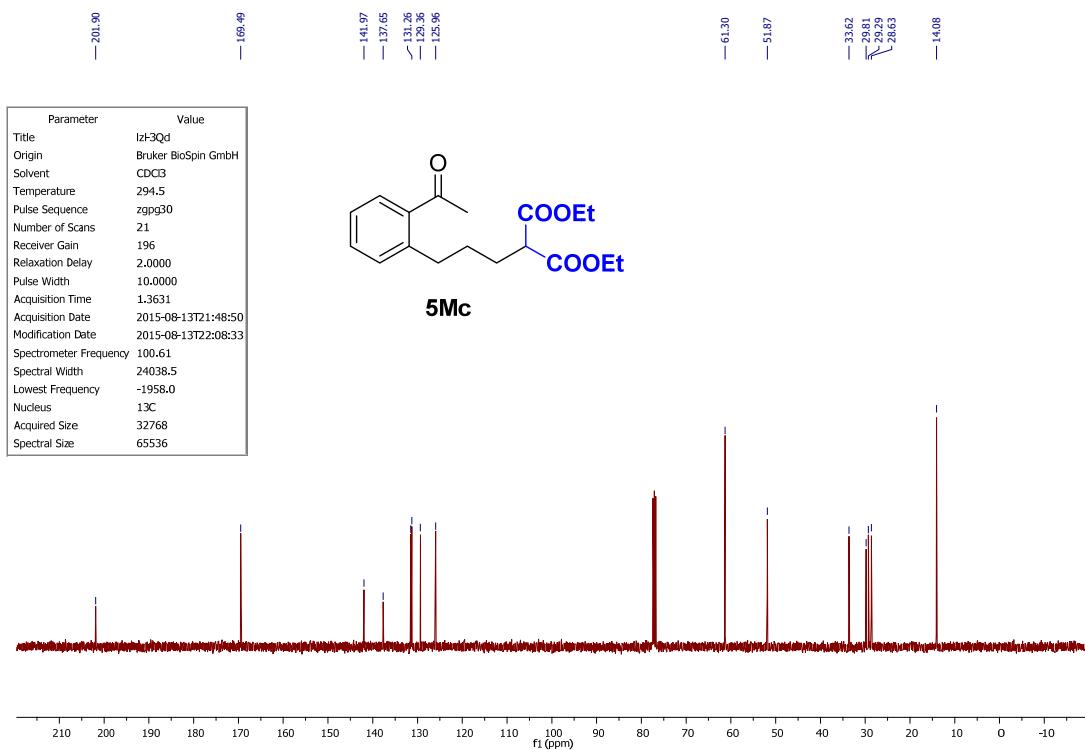
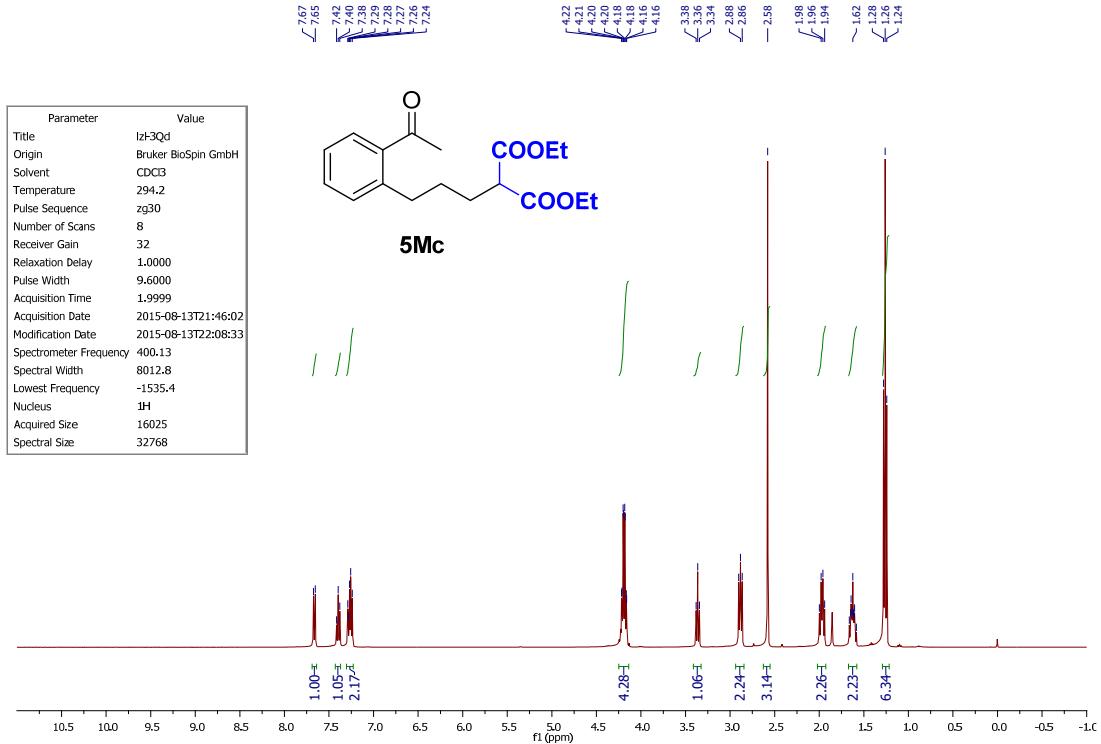


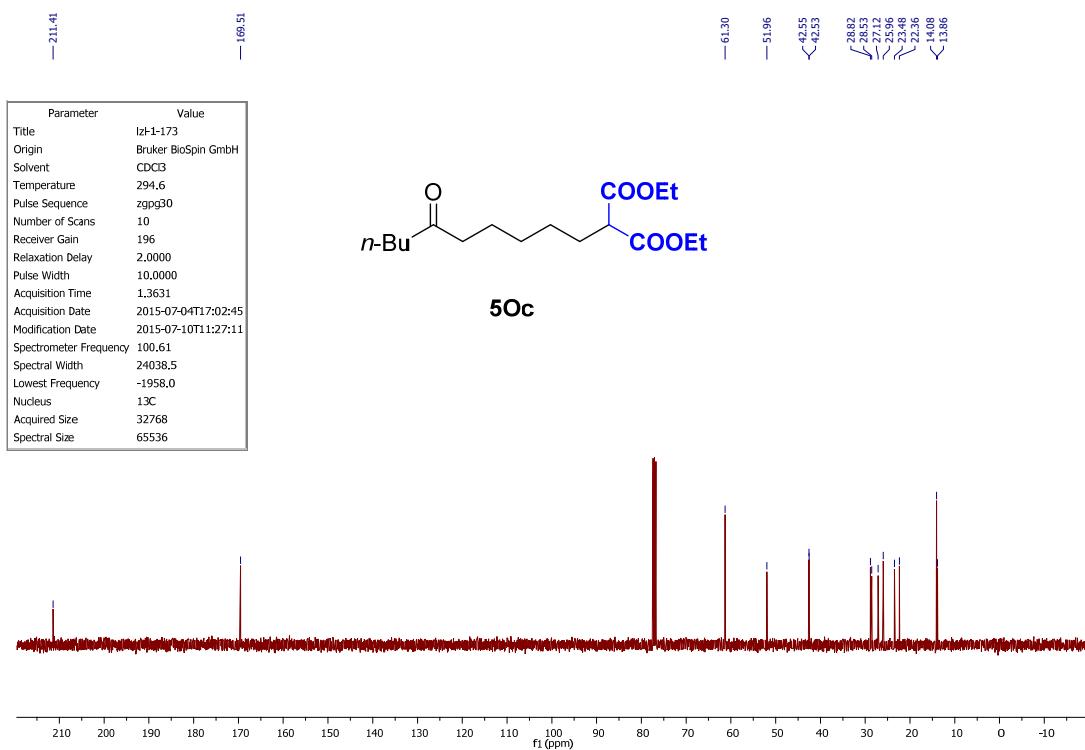
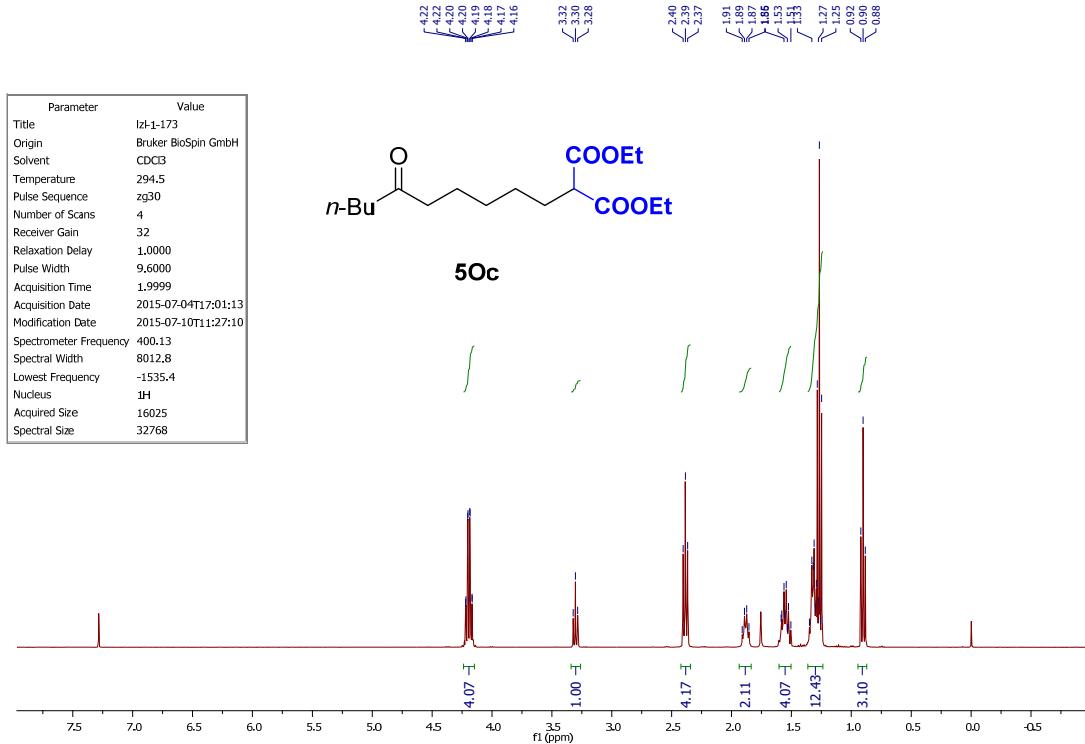
Parameter	Value
Title	IzI-1-36-2-p
Origin	Bruker BioSpin GmbH
Solvent	CDCl <sub>3</sub>
Temperature	295.1
Pulse Sequence	zgpg30
Number of Scans	35
Receiver Gain	196
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2015-03-27T21:08:02
Modification Date	2015-03-27T22:51:17
Spectrometer Frequency	100.61
Spectral Width	24038.5
Lowest Frequency	54.3
Nucleus	<sup>13</sup> C
Acquired Size	32768
Spectral Size	65536



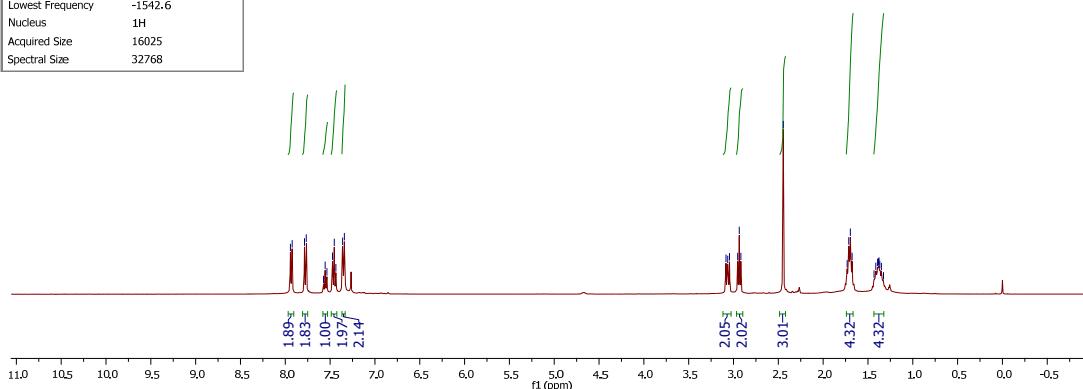
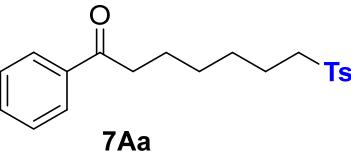




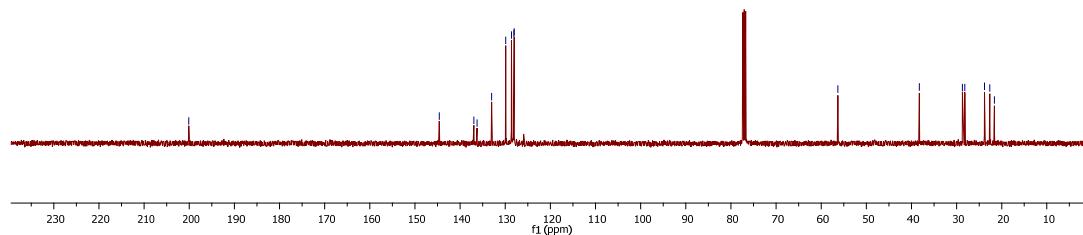
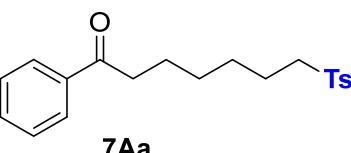


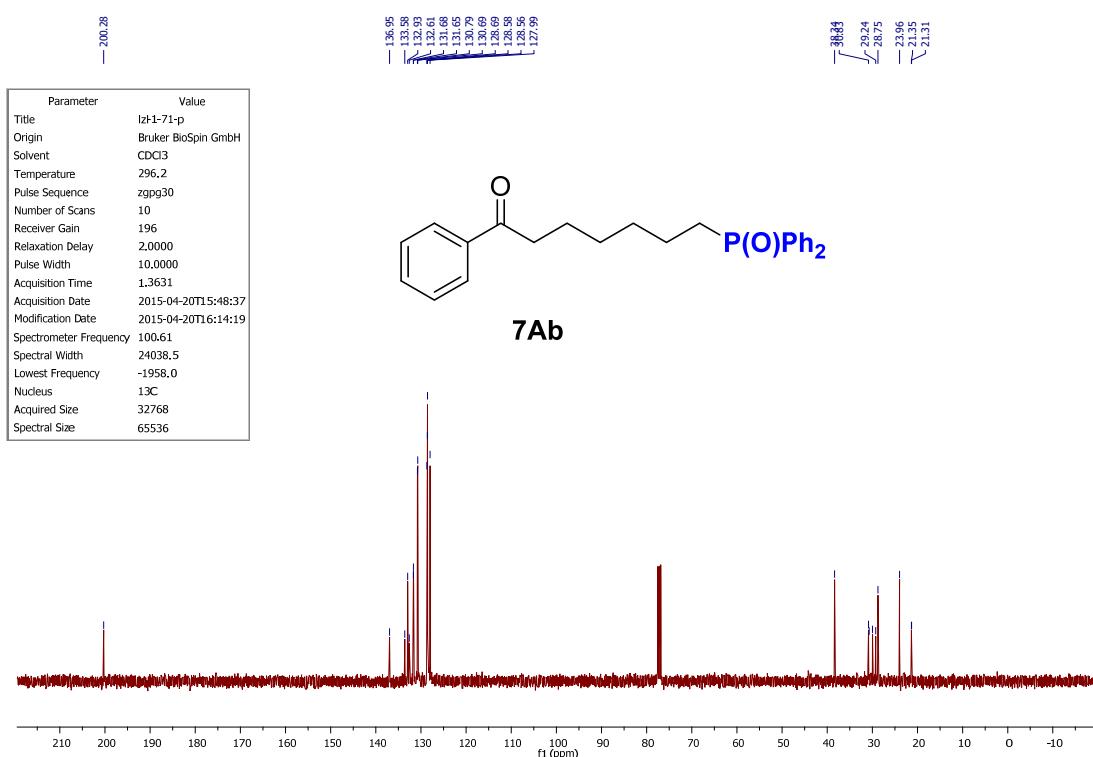
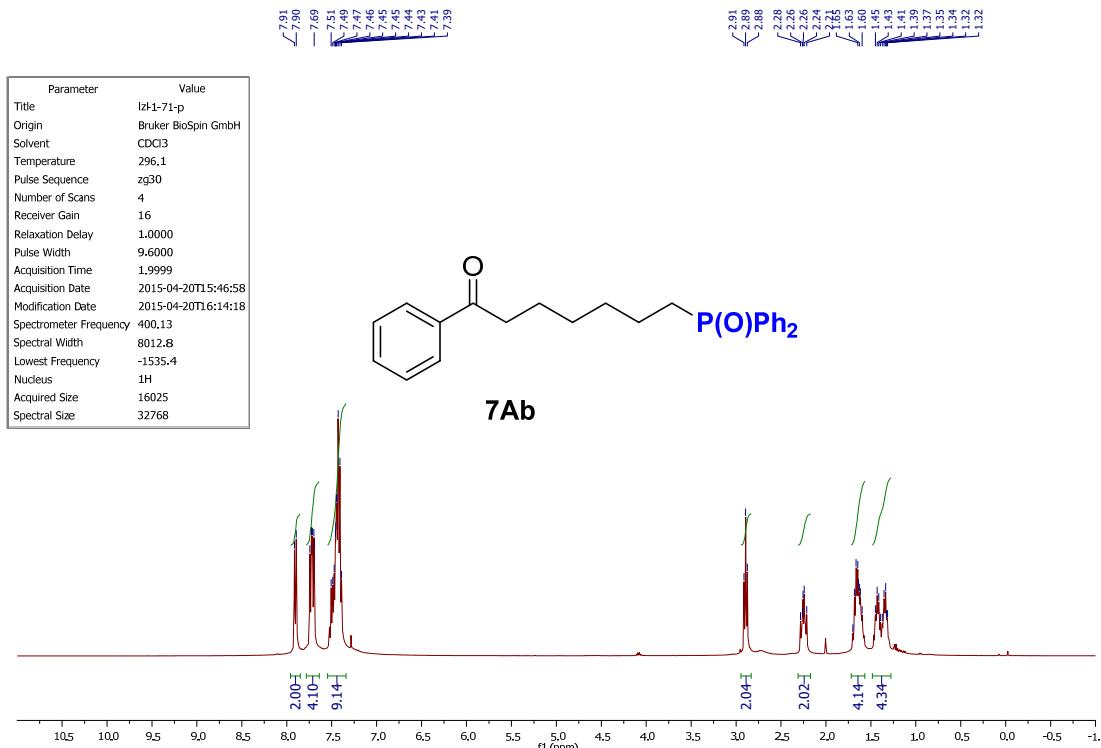


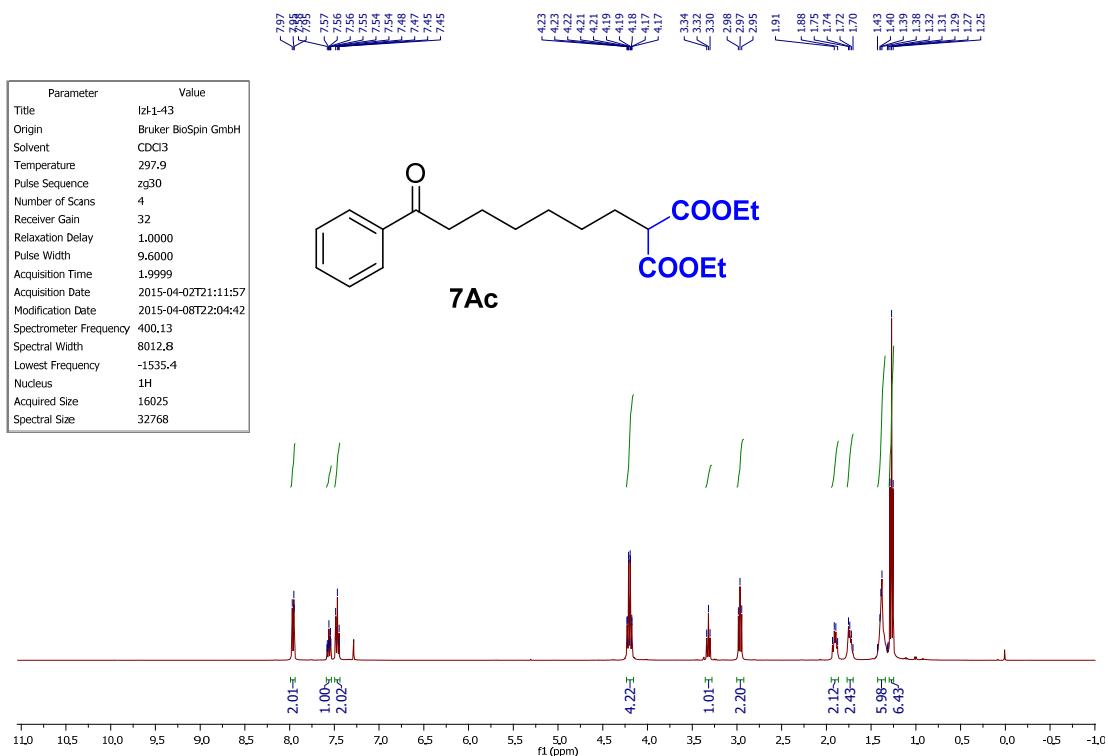
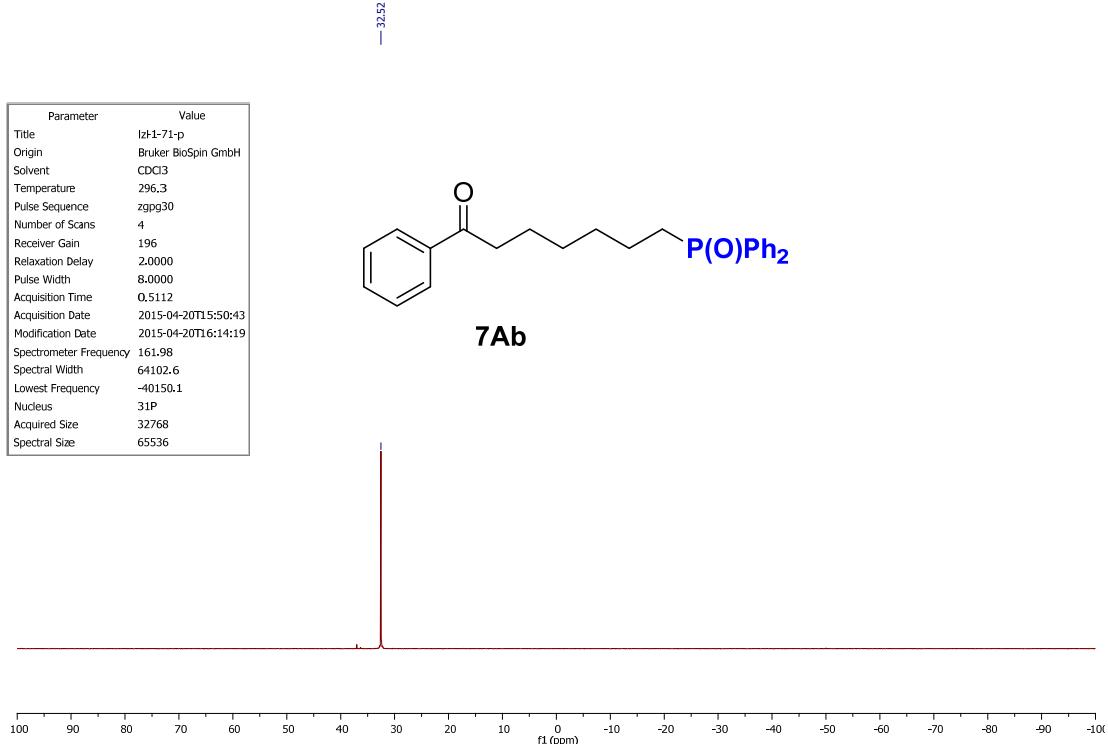
Parameter	Value
Title	IzI-141-pp
Origin	Bruker BioSpin GmbH
Solvent	CDCl3
Temperature	297.4
Pulse Sequence	zg30
Number of Scans	5
Receiver Gain	49
Relaxation Delay	1.0000
Pulse Width	9.6000
Acquisition Time	1.9999
Acquisition Date	2015-04-01T22:06:42
Modification Date	2015-04-02T09:22:40
Spectrometer Frequency	400.13
Spectral Width	8012.8
Lowest Frequency	-1542.6
Nucleus	1H
Acquired Size	16025
Spectral Size	32768

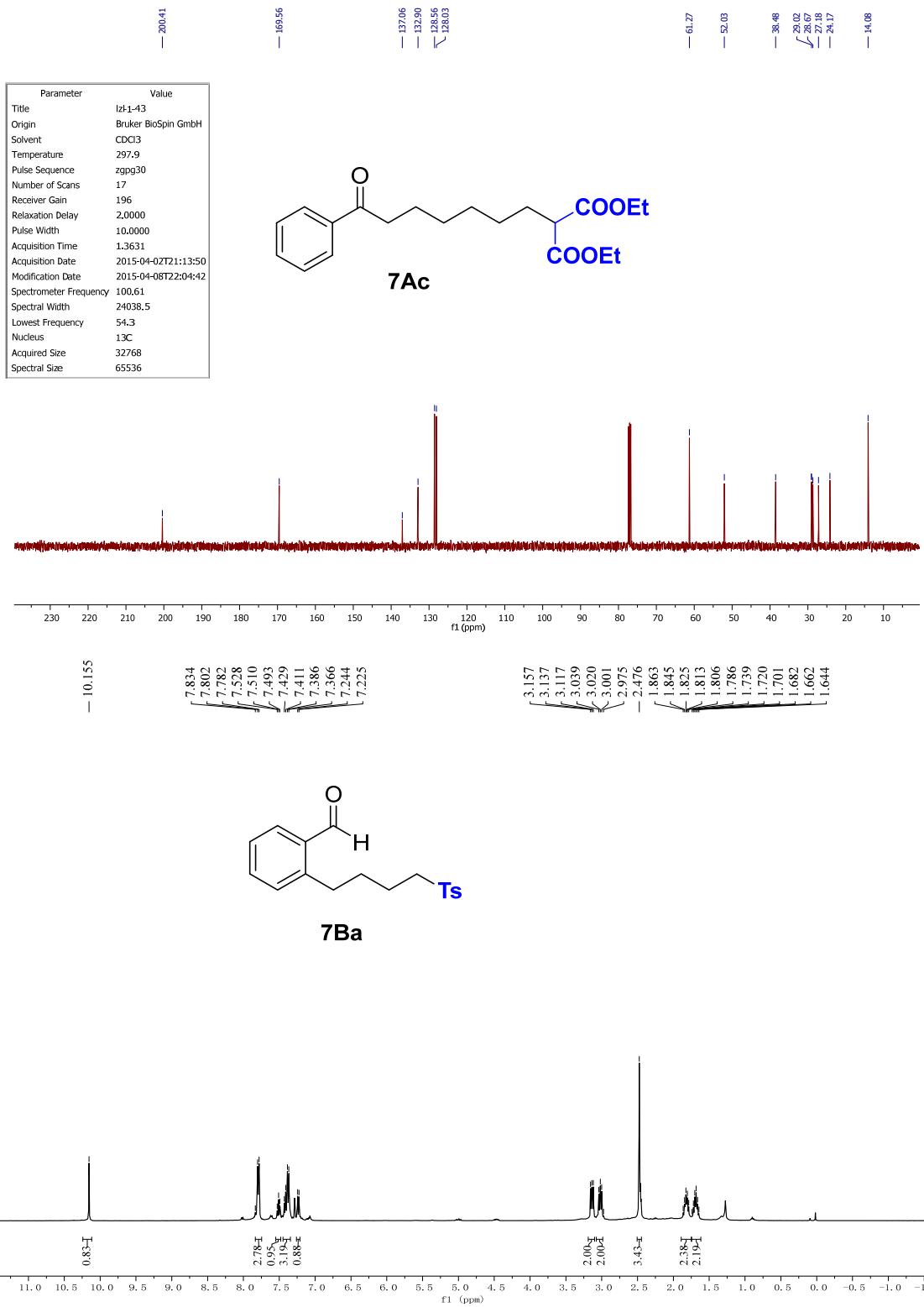


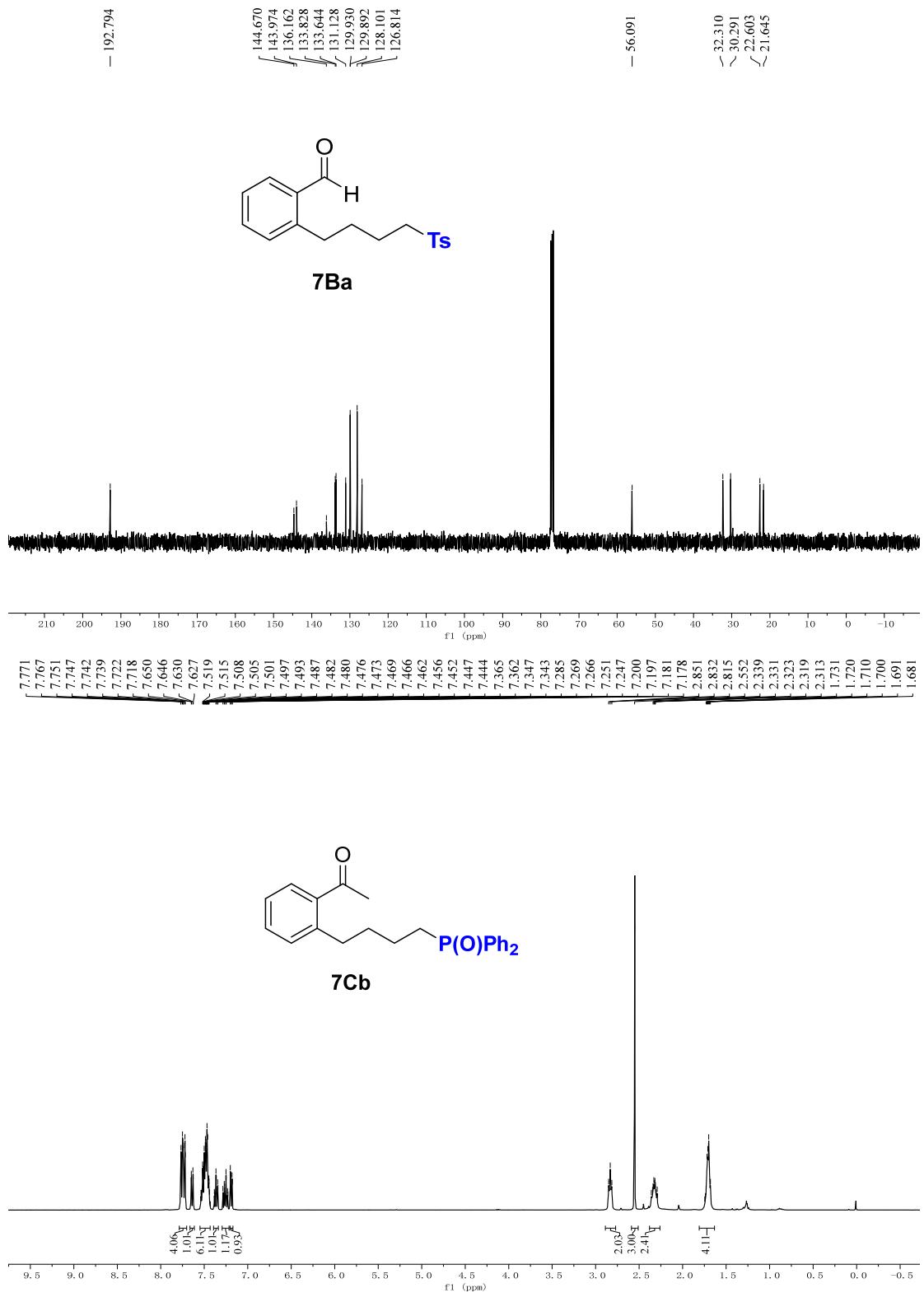
Parameter	Value
Title	lzl-1-41-pp
Origin	Bruker BioSpin GmbH
Solvent	CDCl3
Temperature	297.5
Pulse Sequence	zpgp30
Number of Scans	138
Receiver Gain	196
Relaxation Delay	2.0000
Pulse Width	10.0000
Acquisition Time	1.3631
Acquisition Date	2015-04-01T21:08:30
Modification Date	2015-04-02T09:22:41
Spectrometer Frequency	100.61
Spectral Width	24038.5
Lowest Frequency	54.3
Nucleus	13C
Acquired Size	32768
Spectral Size	65536



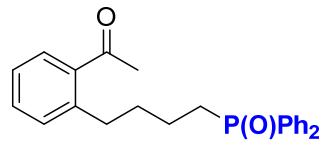




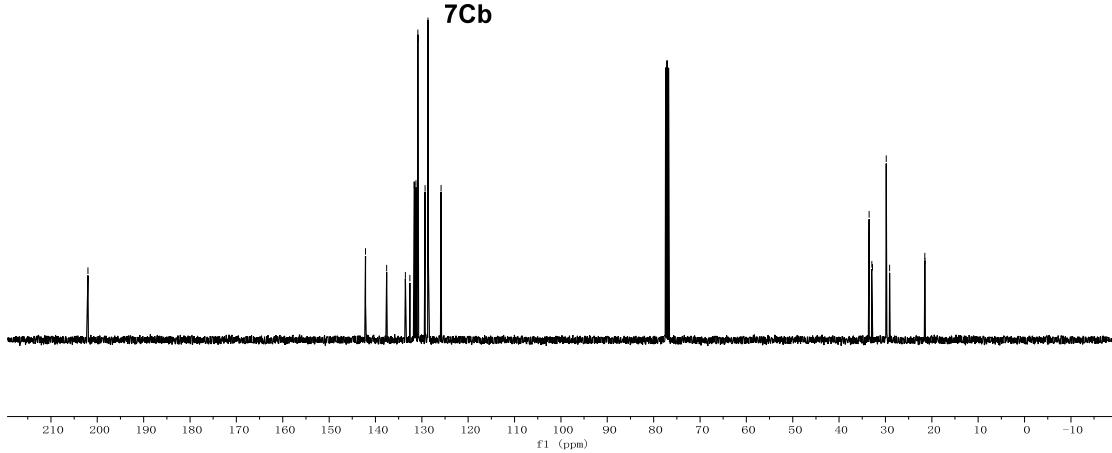




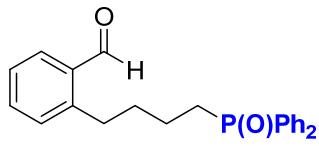
- 202.010



**7Cb**



- 32.726



**7Cb**

