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Supporting Information

for

Reductive N-Alkylation of Primary and Secondary Amines Using

Carboxylic Acids and Borazane under Mild Conditions

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Table of Contents

1. General information	2
2. Screening of Conditions	3
3. General process for the synthesis of 2	5
4. Characterization data for products	6
5. NMR spectroscopic data	18
6. Reference	72

1. General information

All chemicals were purchased from Adamas Reagent, energy chemical company, J&K Scientific Ltd, Bide Pharmatech Ltd and Tansoole. The reagents and solvents were purchased from commercial suppliers and used without further purification. Reactions were monitored by TLC or GC-MS analysis. Flash column chromatography was performed over silica gel (200-300 mesh).

¹H-NMR and ¹³C-NMR spectra were recorded in CDCl₃ on a Bruker Avance III 500MHz NMR spectrometer (500 MHz ¹H, 125 MHz ¹³C) at room temperature. Chemical shifts were reported in ppm on the scale relative to CDCl₃ (δ = 7.26 for ¹H-NMR, δ = 77.00 for ¹³C-NMR) as an internal reference. High resolution mass spectra were recorded using a Thermo Fisher Scientific LTQ FT Ultra or Waters Micromass GCT Premier instrument. Coupling constants (*J*) were reported in Hertz (Hz).

2. Screening of Conditions

H N N	+ MeCOOH CH	Additive <u>Reductant</u> ₃ CN, 60 °C, 5h		
1a		2a	3	a
Entry ^a	Reductant	Additive	Yielo 2a	^{lb} (%) 3a
1	B ₂ pin ₂	[RuCl ₂ (p-cymene)] ₂ (3 %)	trace	0
2	B ₂ (OH) ₄	[RuCl ₂ (p-cymene)] ₂ (3 %)	21	0
3	B ₂ (OH) ₄	Pd(OAc) ₂ (10 %)	14	0
4	B ₂ (OH) ₄	Fe(acac) ₃ (10 %)	23	0
5	B ₂ (OH) ₄	CuSO ₄ (10 %)	10	0
6	$H_3B \cdot NH_3$	[RuCl ₂ (p-cymene)] ₂ (3 %)	42	0
7	$H_3B \cdot NH_3$	CuSO ₄ (10 %)	52	0
8	$H_3B \cdot NH_3$	Cu(OAc) ₂ .H ₂ O(10 %)	62	0
9	$H_3B \cdot NH_3$	Cs ₂ CO ₃ (100 %)	0	0
10	$H_3B \cdot NH_3$	DBU(100 %)	trace	0
11	$H_3B \cdot NH_3$	HNTf ₂ (100 %)	trace	0
12	$H_3B \cdot NH_3$	BF ₃ •Et ₂ O(100 %)	59	trace
13	$H_3B \cdot NH_3$	HBF ₄ (100 %)	38	trace
14	$H_3B \cdot NH_3$	MSA(100 %)	66	trace
15	$H_3B \cdot NH_3$	MSA(200 %)	97(89 ^c)	0
^a Reaction conditions: 1a(0.2 mmol), MeCOOH(3.0 equiv), Reductant(2.0 equiv), CH ₃ CN(1 mL), under N ₂ . ^b Determined by GC using n-dodecane as an internal standard. ^c Isolated vield.				

Table S1 Optimization of the reaction with 1a and MeCOOH

Table S2 Screening of solvent

H N 1a	MeCOOH MSA, H ₃ B•NH		2a N N Ba
Entry ^a	Solvent	Yield ^t 2a	^o (%) 3a
1	Acetone	trace	0
2	CH₃OH	n.r.	0
3	HFIP	n.r.	0
4	Hexane	48	0
5	THF	55	0
6	Toluene	45	0
7	1,4-dioxane	n.r.	0
8	n-Bu ₂ O	n.d.	0
9	CH ₃ CN	97	0
10 ^c	CH ₃ CN	76	0
11 ^d	CH ₃ CN	83	0
12 ^e	CH₃CN	82	0
13 ^f	CH ₃ CN	88	0
14 ^g	CH₃CN	66	18
^a Reaction con (2.0 equiv), sc ^e 3 h. ^f MeCOO not detected.	nditions: 1a (0.2 mmol), MeCOOH (olvent (1 mL), 5 h, 50 ^o C, under N _{2.} 0H (2.5 equiv). ^g MSA (1.0 equiv). n.	3.0 equiv), H ^b under air. ^d r.=no reactio	₃ B•NH ₃ 40 °C. n. n.d.=

3. General process for the synthesis of 2

R ¹ R ² NH or	^r R ¹ NH ₂ +	R ³ CO ₂ H	MSA, H ₃ N•BH ₃	R^{1} $R^{2} \sim R^{3}$	
1				2	

To a mixture of $H_3N \cdot BH_3$ (30.9 mg, 1 mmol, 2.0 equiv) and 1 (0.5 mmol, 1.0 equiv) in CH₃CN (2 mL) was added RCOOH (1.25 or 1.5 mmol, 2.5 or 3.0 equiv). Then the Schlenk tube was evacuated with N₂ three times and finally MSA (1 mmol, 2.0 equiv) was added. The resulting mixture was stirred at 60 °C for 5 h. Upon completion of the reaction, the solvent was evaporated under reduced pressure and the residue was purified by flash column chromatograph (silica gel, petroleum ether:EtOAc = 50:1, v/v) to give the desired product **2**.

4. Characterization data for products

N-ethyl-N-methylaniline (2a) (CAS: No. 613-97-8)¹

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, /v) to give the product as yellow oil (60.1 mg, 89%). ¹H NMR (500 MHz, CDCl₃) δ 7.36 7.29 (m, 2H), 6.83 – 6.74 (m, 3H), 3.48 (q, *J* = 7.1 Hz, 2H), 2.98 (s, 3H), 1.20 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 149.1, 129.2, 116.2, 112.5, 46.9, 37.5, 11.3.

N-ethyl-N,4-dimethylaniline (2b) (CAS: No. 35113-87-2)³

he reaction was performed by following the general procedure **3**. The residue was purified by Mash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as yellow oil (65.6 mg, 88%). ¹H NMR (500 MHz, CDCl₃) δ 13 (d, *J* = 8.5 Hz, 2H), 6.77 – 6.73 (m, 2H), 3.44 (q, *J* = 7.1 Hz, 2H), 2.94 (s, 3H), 2.34 (s, 3H), 1.18 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 147.2, 129.7, 125.6, 113.1, 47.2, 37.8, 20.3, 11.1.

N-ethyl-4-methoxy-N-methylaniline (2c) (CAS: No. 6114-15-4)³

he reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 50:1, v/v) to give the product as yellow oil (70.1 mg, 85%). ¹H NMR (500 MHz, CDCl₃) δ 6.89 – 0.84 (m, 2H), 6.78 (d, *J* = 9.1 Hz, 2H), 3.79 (s, 3H), 3.34 (q, *J* = 7.1 Hz, 2H), 2.86 (s, 3H), 1.12 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 151.9, 144.0, 115.2, 114.8, 55.8, 48.2, 38.4, 11.1.

N-ethyl-4-fluoro-N-methylaniline (2d) (CAS: No. 67274-53-7)³

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 80:1, v/v) of give the product as yellow oil (63.5 mg, 83%). ¹H NMR (500 MHz, CDCl₃) δ 7.00 – 6.94 (m, 2H), 6.74 – 6.67 (m, 2H), 3.38 (q, *J* = 7.1 Hz, 2H), 2.89 (s, 3H), 1.13 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 155.4 (d, *J* = 235.0 Hz), 145.9, 115.5 (d, *J* = 21.9 Hz), 114.0 (d, *J* = 6.9 Hz), 47.7, 38.1, 11.0.

4-chloro-N-ethyl-N-methylaniline (2e) (CAS: No. 13519-85-2)³

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, **v**) to give the product as yellow oil (71.0 mg, 84%). ¹H NMR (500 MHz, CDCl₃) δ 7.23 – 7.18 (m, 2H), 6.68 – 6.65 (m, 2H), 3.41 (q, *J* = 7.1 Hz, 2H), 2.92 (s, 3H), 1.15 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 147.7, 128.9, 120.8, 113.5, 47.0, 37.6, 11.1.

4-bromo-N-ethyl-N-methylaniline (2f) (CAS: No. 67274-54-8)³

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v(p) to give the product as yellow oil (94.8 mg, 89%). ¹H NMR (500 MHz, CDCl₃) δ 7.37 = 7.30 (m, 2H), 6.66 – 6.57 (m, 2H), 3.41 (q, *J* = 7.1 Hz, 2H), 2.92 (s, 3H), 1.15 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 148.0, 131.8, 114.0, 107.9, 46.9, 37.6, 11.1.

N-ethyl-N-methyl-4-nitroaniline (2g) (CAS: No. 56269-48-8)³

The reaction was performed by following the general procedure **3**. The residue was purified by fash column chromatograph (silica gel, petroleum ether:AcOEt = 30:1, v/v) togive the product as a yellow solid (82.8 mg, 92%). M. P. 87 °C (Lit. 87-89 °C). ¹H NMR (500 MHz, CDCl₃) δ 8.11 – 8.01 (m, 2H), 6.60 – 6.54 (m, 2H), 3.49 (q, *J* = 7.2 Hz, 2H), 3.04 (s, 3H), 1.19 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 153.2, 136.4, 126.2, 110.1, 47.0, 37.8, 11.6.

N,N-diethylnaphthalen-1-amine (2h) (CAS: No. 84-95-7)⁶

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 50:1, v/v) to give the product as yellow oil (82.6 mg, 83%). ¹H NMR (500 MHz, CDCl₃) δ 8.42 – 8.33 (m, 11), 7.86 (dd, *J* = 6.6, 2.5 Hz, 1H), 7.60 (d, *J* = 8.1 Hz, 1H), 7.54 – 7.43 (m, 3H), 7.19 (d, *J* = 7.3 Hz, 1H), 3.25 (q, *J* = 7.0 Hz, 4H), 1.11 (t, *J* = 7.1 Hz, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 147.9, 134.9, 131.3, 128.1, 125.6, 125.5, 125.1, 124.3, 123.3, 117.9, 47.7, 12.3.

1-ethylindoline (2i) (CAS: No. 5876-09-5)³

The reaction was performed by following the general procedure **3**. The residue was purfied by flash column chromatograph (silica gel, petroleum ether:AcOEt = 50:1, v/v)

to give the product as yellow oil (63.2 mg, 86%). ¹H NMR (500 MHz, CDCl₃) δ 7.19 – 7.11 (m, 2H), 6.74 (td, J = 7.5, 0.8 Hz, 1H), 6.57 (d, J = 7.7 Hz, 1H), 3.40 (t, J = 8.3 Hz, 2H), 3.22 (d, J = 7.2 Hz, 2H), 3.03 (t, J = 8.3 Hz, 2H), 1.27 (t, J = 7.2 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 152.3, 130.4, 127.3, 124.4, 117.6, 107.3, 52.4, 43.3, 28.6, 12.0.

1-ethyl-1,2,3,4-tetrahydroquinoline (2j) (CAS: No. 16768-69-7)⁵

The seaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 50:1, v/v) to give the product as yellow oil (75.7 mg, 94%). ¹H NMR (500 MHz, CDCl₃) δ 7.17 td, *J* = 8.2, 1.7 Hz, 1H), 7.06 (dd, *J* = 7.3, 0.7 Hz, 1H), 6.74 – 6.66 (m, 2H), 3.45 (t, *J* = 7.1 Hz, 2H), 3.38 – 3.35 (m, 2H), 2.87 (t, *J* = 6.4 Hz, 2H), 2.09 – 2.05 (m, 2H), 1.26 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 145.1, 129.3, 127.2, 122.5, 115.5, 110.6, 48.5, 45.4, 28.3, 22.4, 10.9.

N-benzyl-N-ethylaniline (2k) (CAS: No. 92-59-1)³

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 50:1, v/v) of give the product as colorless oil (95.0 mg, 90%). ¹H NMR (500 MHz, CDCl₃) δ 7.55 – 7.48 (m, 2H), 7.47 – 7.37 (m, 5H), 6.94 – 6.86 (m, 3H), 4.71 (s, 2H), 3.67 (q, *J* = 7.1 Hz, 2H), 1.41 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 148.7, 139.5, 129.4, 128.7, 126.9, 126.8, 116.3, 112.4, 54.1, 45.3, 12.3.

N-allyl-N-ethylaniline (2l) (CAS: No. 16078-91-4)³

The reaction was performed by following the general procedure **3**. The residue was putified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, vv) to give the product as yellow oil (74.9 mg, 93%). ¹H NMR (500 MHz, CDCl₃) δ 7.37 – 7.21 (m, 2H), 6.76 (dd, J = 19.3, 7.7 Hz, 3H), 5.94 (ddt, J = 17.1, 10.0, 4.9 Hz, 1H), 5.24 (ddq, J = 19.0, 10.3, 1.7 Hz, 2H), 3.97 (dt, J = 4.7, 1.7 Hz, 2H), 3.46 (q, J = 7.1 Hz, 2H), 1.25 (t, J = 7.1 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 148.3, 134.5, 129.2, 115.9, 115.8, 112.1, 52.7, 44.8, 12.3.

2-(ethyl(phenyl)amino)ethan-1-ol (2m) (CAS: No. 92-50-2)¹

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 50:1, v/v) to give the product as yellow oil (73.4 mg, 89%). ¹H NMR (500 MHz, CDCl₃) δ 7.31 – 7.24 (m, 2H), 6.83 (d, *J* = 8.0 Hz, 2H), 6.77 (t, *J* = 7.3 Hz, 1H), 3.80 (t, *J* = 5.9 Hz, 2H),

3.50 - 3.42 (m, 4H), 2.41 (s, 1H), 1.19 (t, J = 7.0 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 148.2, 129.4, 116.9, 113.1, 60.1, 52.7, 45.8, 11.9.

N,N-dibenzylethanamine (2n) (CAS: No. 10479-25-1)¹

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 50:1, v/v) to give the product as yellow oil (77.6 mg, 69%). ¹H NMR (500 MHz, CDCl₃) δ 7.43 (d, J = 7.4 Hz, 4H), 7.35 (dd, J = 10.3, 4.7 Hz, 4H), 7.29 – 7.25 (m, 2H), 3.63 (s, 4H), 2.56 (q, J = 7.0 Hz, 2H), 1.12 (t, J = 7.1 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 128.8, 128.8, 128.2, 126.8, 57.7, 47.1, 11.8.

N-ethylaniline (20) (CAS: No. 103-69-5)²

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 30:1, v/v) to give the product as yellow oil (50.8 mg, 84%). ¹H NMR (500 MHz, CDCl₃) δ 7.41 – 7.S1 (m, 2H), 6.86 (tt, *J* = 7.3, 1.0 Hz, 1H), 6.79 – 6.72 (m, 2H), 3.63 (s, 1H), 3.29 (q, *J* = 7.1 Hz, 2H), 1.39 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 148.6, 129.4, 117.3, 112.9, 38.6, 15.0.

N-ethyl-3-methylaniline (2p) (CAS: No. 102-27-2)¹

The reaction was performed by following the general procedure **3**. The residue was purified **5**, flash column chromatograph (silica gel, petroleum ether:AcOEt = 30:1, v/v) to give the product as yellow oil (56.7 mg, 78%). ¹H NMR (500 MHz, CDCl₃) δ 7.27 – 7.22 (m, 1H), 6.70 (d, J = 7.6 Hz, 1H), 6.58 (d, J = 6.5 Hz, 2H), 3.62 (s, 1H), 3.29 (q, J = 7.1 Hz, 2H), 2.46 (s, 3H), 1.39 (t, J = 7.2 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 148.7, 139.0, 129.2, 118.3, 113.7, 110.1, 38.6, 21.8, 15.1.

N,4-diethylaniline (2q) (CAS: No. 4960-26-3)³

The reaction was performed by following the general procedure **3**. The residue was purified by fash column chromatograph (silica gel, petroleum ether:AcOEt = 30:1, v/v) to give the product as a white solid (60.3 mg, 81%). ¹H NMR (500 MHz, CDCl₃) δ 7.04 (d, J = 8.3 Hz, 2H), 6.59 (d, J = 8.4 Hz, 2H), 3.17 (q, J = 7.1 Hz, 2H), 2.57 (q, J = 7.6 Hz, 2H), 1.27 (t, J = 7.1 Hz, 3H), 1.21 (t, J = 7.6 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 146.4, 133.2, 128.5, 113.0, 38.8, 27.9, 16.0, 15.0.

N,N-diethylaniline (2r) (CAS: No. 91-66-7)²

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, vph give the product as yellow oil (64.8 mg, 87%). ¹H NMR (500 MHz, CDCl₃) δ 7.29 – 7.24 (m, 2H), 6.82 – 6.64 (m, 3H), 3.40 (d, *J* = 7.1 Hz, 4H), 1.21 (t, *J* = 7.1 Hz, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 147.8, 129.3, 115.4, 111.9, 44.4, 12.6.

N-methylaniline (2s) (CAS: No. 100-61-8)³

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 30:1, v/v) to give the product as yellow oil (43.3 mg, 81%). ¹H NMR (500 MHz, CDCl₃) δ 7.39 – 7.34 (m, 2H), 6.89 (tt, J = 7.3, 1.0 Hz, 1H), 6.78 – 6.73 (m, 2H), 3.66 (s, 1H), 2.94 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 149.5, 129.4, 117.3, 112.6, 30.8.

N,N-dimethylaniline (2t) (CAS: No. 121-69-7)³

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v. **Pb** give the product as yellow oil (50.8 mg, 84%). ¹H NMR (500 MHz, CDCl₃) δ 7.48 – 7.40 (m, 2H), 6.97 – 6.90 (m, 3H), 3.11 (s, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 150.8, 129.2, 116.8, 112.8, 40.7.

N-butylaniline (2u) (CAS: No. 1126-78-9)¹

The reaction was performed by following the general procedure **3**. The residue was purpled by flash column chromatograph (silica gel, petroleum ether:AcOEt = 30:1, v/v) to give the product as yellow oil (64.1 mg, 86%). ¹H NMR (500 MHz, CDCl₃) δ 7.34 – 7.26 (m, 2H), 6.82 (tt, J = 7.4, 1.0 Hz, 1H), 6.72 (dt, J = 8.9, 1.6 Hz, 2H), 3.24 – 3.19 (m, 2H), 1.71 (ddd, J = 12.6, 8.4, 6.4 Hz, 2H), 1.55 (dq, J = 14.5, 7.3 Hz, 2H), 1.09 (t, J = 7.4 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 148.7, 129.3, 117.1, 112.8, 43.8, 31.8, 20.4, 14.1.

N-(cyclopropylmethyl)aniline (2v) (CAS: No. 36178-60-6)⁴

he reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 30:1, v/v) to give the product as yellow oil (64.7 mg, 88%). ¹H NMR (500 MHz, CDCl₃) δ 7.28 – 7.18 (m, 2H), 6.81 – 6.73 (m, 1H), 6.67 (dd, J = 8.6, 0.9 Hz, 2H), 3.84 (s, 1H), 3.01 (d, J = 6.9 Hz, 2H), 1.20 – 1.11 (m, 1H), 0.65 – 0.57 (m, 2H), 0.30 (dt, J = 9.6, 4.6 Hz, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 148.6, 129.3, 117.3, 112.8, 49.1, 11.0, 3.5.

N-(cyclohexylmethyl)aniline (2w) (CAS: No. 79952-92-4)¹

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 30:1, v/v) to give the product as yellow oil (85.1 mg, 90%). ¹H NMR (500 MHz, CDCl₃) δ 7.26 – 7.16 (m, 2H), 6.72 (t, J = 7.3 Hz, 1H), 6.67 – 6.61 (m, 2H), 3.74 (s, 1H), 3.00 (d, J = 6.7 Hz, 2H), 1.90 – 1.83 (m, 2H), 1.76 (dddd, J = 6.3, 4.9, 4.2, 2.4 Hz, 3H), 1.67 – 1.59 (m, 1H), 1.34 – 1.22 (m, 3H), 1.03 (qd, J = 12.3, 3.1 Hz, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 148.7, 129.2, 116.9, 112.6, 50.6, 37.6, 31.4, 26.6, 26.0.

N-(2-phenylpropyl)aniline (2x) (CAS: No. 56165-31-2)¹¹

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 30:1, v/v) to give the product as a white solid (93.9 mg, 89%). ¹H NMR (500 MHz, CDCl₃) δ 7.39 – 7.35 (m, 2H), 7.30 – 7.25 (m, 3H), 7.23 – 7.18 (m, 2H), 6.75 (dd, J = 11.5, 4.2 Hz, 1H), 6.67 – 6.60 (m, 2H), 3.38 (dd, J = 12.4, 6.2 Hz, 1H), 3.28 (dd, J = 12.4, 8.2 Hz, 1H), 3.10 (dq, J = 14.0, 6.9 Hz, 1H), 1.38 (d, J = 7.0 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 147.8, 144.5, 129.3, 128.7, 127.3, 126.7, 117.6, 113.2, 51.1, 39.2, 19.8.

N-(2-phenoxyethyl)aniline (2y) (CAS: No. 622-18-4)¹

The reaction was performed by following the general procedure **3**. The residue was performed by flash column chromatograph (silica gel, petroleum ether:AcOEt = 30:1, v/v) to give the product as a orange solid (87.3 mg, 82%). M. P. 48 °C (Lit. 48-50 °C). ¹H NMR (500 MHz, CDCl₃) δ 7.39 – 7.33 (m, 2H), 7.28 – 7.24 (m, 2H), 7.05 – 6.96 (m, 3H), 6.80 (tt, J = 7.4, 1.0 Hz, 1H), 6.73 (dt, J = 8.8, 1.6 Hz, 2H), 4.22 (t, J = 5.3 Hz, 2H), 3.58 (t, J = 5.3 Hz, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 158.7, 147.9, 129.6, 129.4, 121.1, 117.9, 114.6, 113.2, 66.4, 43.4.

N-(((3r,5r,7r)-adamantan-1-yl)methyl)aniline (2aa) (CAS: No. 802038-

41-1)⁸

The reaction period by following the general procedure **3**. The residue was purified by hash column chromatograph (silica gel, petroleum ether:AcOEt = 30:1, v/v) to give the product as a white solid (102.4 mg, 85%). M. P. 49 °C (Lit. 49-51 °C). ¹H NUR (500 MHz, CDCl₃) δ 7.25 – 7.19 (m, 2H), 6.72 (t, *J* = 7.3 Hz, 1H), 6.70 – 6.66 (m, 2H), 3.71 (s, 1H), 2.86 (s, 2H), 2.07 (s, 3H), 1.81 (d, *J* = 12.2 Hz, 3H), 1.73 (d, *J* = 11.4 Hz, 3H), 1.65 (d, *J* = 2.5 Hz, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 1493, 129.2, 116.8, 112.6, 56.3, 40.8, 37.2, 33.9, 28.4.

N-benzylaniline (2ab) (CAS: No. 103-32-2)¹

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 30:1, v/v) to give the product as a grey solid (76.9 mg, 84%). ¹H NMR (500 MHz, CDCl₃) δ 7.45 (dt, J = 15.0, 7.4 Hz, 4H), 7.37 (t, J = 6.9 Hz, 1H), 7.30 – 7.25 (m, 2H), 6.82 (td, J = 7.3, 0.6 Hz, 1H), 6.73 (d, J = 8.5 Hz, 2H), 4.41 (s, 2H), 4.10 (s, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 148.2, 139.5, 129.4, 128.7, 127.6, 127.3, 117.6, 112.9, 48.4.

N-(4-fluorobenzyl)aniline (2ac) (CAS: No. 83444-25-1)⁴

the reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 30:1, v/v) oPhye the product as verified with equal of (84.4 mg, 84%). ¹H NMR (500 MHz, CDCl₃) δ 7.42 – 7.36 (m, 2H), 7.27 – 7.22 (m, 2H), 7.13 – 7.05 (m, 2H), 6.80 (tt, J = 7.4, 1.0 Hz, 1H), 6.69 (dt, J = 8.9, 1.6 Hz, 2H), 4.35 (s, 2H), 4.07 (s, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 163.1, 161.1, 148.0, 135.18 (d, J = 3.1 Hz), 129.4, 129.05 (d, J = 8.0 Hz), 117.8, 115.6, 115.4, 112.9, 47.6.

N-(4-methoxybenzyl)aniline (2ad) (CAS: No. 3526-43-0)⁴

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 20:1, v/v) Physe the product as yellow oil (84.1 mg, 79%). ¹H NMR (500 MHz, CDCl₃) δ 7.38 – 7.31 (m, 2H), 7.26 – 7.21 (m, 2H), 6.96 – 6.92 (m, 2H), 6.81 – 6.75 (m, 1H), 6.69 (dd, J = 8.6, 1.0 Hz, 2H), 4.30 (s, 2H), 4.00 (s, 1H), 3.85 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 158.9, 148.3, 131.5, 129.3, 128.9, 117.5, 114.1, 112.9, 55.3, 47.8.

N-(4,5-dimethoxy-2-nitrobenzyl)aniline (2ae)

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 10:1, v/v) o give the product as yellow oil (118.1 mg, 82%). ¹H NMR (500 MHz, CDCl₃) δ 7.74 s, ¹H), 7.19 + 7.15 (m, 3H), 6.75 (t, *J* = 7.3 Hz, 1H), 6.60 (dd, *J* = 8.5, 0.9 Hz, 2H), 4.28 (ddd, *J* = 28.6, 22.9, 13.5 Hz, 1H), 3.96 (s, 3H), 3.84 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 153.7, 147.6, 147.6, 140.1, 131.3, 129.3, 118.1, 113.1, 111.0, 108.5, 56.4, 56.4, 46.5.

HRMS (ESI, m/z) calcd for C₁₅H₁₇N₂O₄[M+H]⁺: 289.1183; found: 289.1185.

N-(2-(4-isobutylphenyl)propyl)aniline (2af) (CAS: No. 1553525-77-1)²

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 30:1, v/v) to give the product as yellow oil (117.5 mg, 88%). ¹H NMR (500 MHz, CDCl₃) δ 7.19 (dd, J = 23.4, 11.4, 5.1 Hz, 6H), 6.74 (t, J = 7.3 Hz, 1H), 6.63 (dd, J = 8.5, 0.9 Hz,

2H), 3.37 (dd, J = 12.3, 6.2 Hz, 1H), 3.27 (dd, J = 12.3, 8.2 Hz, 1H), 3.12 – 3.04 (m, 1H), 2.51 (d, J = 7.2 Hz, 2H), 1.91 (dt, J = 13.6, 6.8 Hz, 1H), 1.38 (d, J = 7.0 Hz, 3H), 0.97 (d, J = 6.6 Hz, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 148.2, 141.7, 140.0, 129.4, 129.3, 127.0, 117.3, 113.0, 51.0, 45.1, 38.8, 30.3, 22.5, 19.8.

N-(furan-2-ylmethyl)aniline (2ag) (CAS: No. 4439-56-9)¹³

The reaction oval performed by following the general procedure **3**. The residue was burified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 30:1, v/v) to give the product as yellow oil (74.4 mg, 86%). ¹H NMR (500 MHz, CDCl₃) δ 7.40 (d, J = 1.5 Hz, 1H), 7.24 – 7.20 (m, 2H), 6.77 (t, J = 7.3 Hz, 1H), 6.73 – 6.69 (m, 2H), 6.35 (dd, J = 3.1, 1.9 Hz, 1H), 6.27 (d, J = 3.1 Hz, 1H), 4.35 (s, 2H), 4.05 (s, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 152.7, 147.6, 141.9, 129.2, 118.0, 113.2, 110.3, 107.0, 41.5.

N-(thiophen-2-ylmethyl)aniline (2ah) (CAS: No. 40625-28-3)⁴

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 30:1, v/v) to give the product as colorless oil (82.2 mg, 87%). ¹H NMR (500 MHz, CDCl₃) δ 7.24 (ddd, J = 8.5, 6.2, 1.6 Hz, 3H), 7.07 – 7.04 (m, 1H), 7.01 (dd, J = 5.0, 3.5 Hz, 1H), 6.82 – 6.77 (m, 1H), 6.72 (dd, J = 8.5, 0.9 Hz, 2H), 4.55 (s, 2H), 4.08 (s, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 147.6, 143.0, 129.3, 126.9, 125.1, 124.6, 118.1, 113.2, 43.5.

N-(hex-3-en-1-yl)aniline (2ai) (CAS: No. 634181-56-9)9

The reaction was performed by following the general procedure **3**. The residue was performed by fash column chromatograph (silica gel, petroleum ether:AcOEt = 30:1, v/v) to give the product as yellow oil (75.3 mg, 86%). ¹H NMR (500 MHz, CDCl₃) δ 7.24 – 7.17 (m, 2H), 6.76 – 6.70 (m, 1H), 6.64 (dd, J = 8.6, 0.9 Hz, 2H), 5.63 (dtt, J = 13.8, 6.3, 1.2 Hz, 1H), 5.48 – 5.39 (m, 1H), 3.69 (s, 1H), 3.16 (t, J = 6.7 Hz, 2H), 2.35 (qd, J = 6.7, 1.0 Hz, 2H), 2.07 (pd, J = 7.4, 1.1 Hz, 2H), 1.03 (t, J = 7.5 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 148.4, 135.0, 129.2, 125.9, 117.3, 112.9, 43.4, 32.4, 25.7, 13.9.

N-((2E,4E)-hexa-2,4-dien-1-yl)aniline (2aj) (CAS: No. 115477-06-0)¹⁰

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 30:1, v/v) to give the product as yellow oil (71.8 mg, 83%). ¹H NMR (500 MHz, CDCl₃) δ 7.22 (t, J = 7.4 Hz, 2H), 6.76 (t, J = 7.0 Hz, 1H), 6.67 (d, J = 7.7 Hz, 2H), 6.33 – 6.20 (m, 1H), 6.17 – 6.03 (m, 1H), 5.72 (dd, J = 13.8, 7.7 Hz, 2H), 3.81 (d, J = 5.4 Hz, 2H), 3.47 (s, 1H), 1.79 (d, J = 6.3 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 148.1, 132.2, 130.9, 129.4, 129.2, 127.6, 117.6, 113.1, 46.0, 18.1.

2-methyl-1-phenylpyrrolidine (2ak) (CAS: No. 33342-99-3)12

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 10:1, v/v) to give the product as yellow oil (58.8 mg, 73%). ¹H NMR (500 MHz, CDCl₃) δ 7.21 (dd, J = 8.4, 7.5 Hz, 2H), 6.73 (t, J = 7.3 Hz, 1H), 6.65 (d, J = 7.7 Hz, 2H), 3.93 – 3.78 (m, 1H), 3.17 (t, J = 6.9 Hz, 2H), 1.79 – 1.68 (m, 2H), 1.62 – 1.56 (m, 2H), 1.24 (d, J = 6.2 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 148.4, 129.3, 117.4, 112.9, 67.9, 44.1, 36.8, 25.9, 23.7.

1,2-diphenylpyrrolidine (2al) (CAS: No. 72709-29-6)¹²

he fraction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 10:1, v/v) or product as yellow oil (97.0 mg, 87%). ¹H NMR (500 MHz, CDCl₃) δ 7.32 (dd, J = 10.1, 4.9 Hz, 2H), 7.25 (d, J = 7.8 Hz, 3H), 7.17 (dd, J = 8.4, 7.4 Hz, 2H), 6.66 (t, J = 7.3 Hz, 1H), 6.52 (d, J = 8.2 Hz, 2H), 4.76 (dd, J = 8.2, 1.7 Hz, 1H), 3.77 – 3.71 (m, 1H), 3.44 (td, J = 9.1, 6.9 Hz, 1H), 2.46 – 2.37 (m, 1H), 2.08 – 1.94 (m, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 147.2, 144.6, 129.0, 128.5, 126.6, 125.9, 115.7, 112.3, 62.9, 49.1, 36.1, 23.1.

N-benzyl-4-methoxyaniline (2am) (CAS: No. 17377-95-6)⁴

horeaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 50:1, v/v) o give the product as vellow oil (78.8 mg, 74%). ¹H NMR (500 MHz, CDCl₃) δ 7.45 – 7.36 (m, 4H), 7.31 (t, *J* = 7.1 Hz, 1H), 6.85 – 6.79 (m, 2H), 6.68 – 6.62 (m, 2H), 4.32 (s, 2H), 3.78 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 152.2, 142.5, 139.7, 128.6, 127.6, 127.2, 114.9, 114.2, 55.8, 49.3.

4-methoxy-N-neopentylaniline (2an) (CAS: No. 65570-14-1)⁴

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 20:1, v/v) to give the product as yellow oil (83.0 mg, 86%). ¹H NMR (500 MHz, CDCl₃) δ 6.84 – 6.79 (m, 2H), 6.65 – 6.60 (m, 2H), 3.78 (s, 3H), 3.40 (s, 1H), 2.88 (s, 2H), 1.02 (s, 9H). ¹³C NMR (126 MHz, CDCl₃) δ 151.8, 143.5, 114.9, 113.9, 57.1, 55.9, 31.8, 27.7.

N-benzyl-N-methylaniline (2ao) (CAS: No. 614-30-2)³

The reaction was performed by following the general procedure **3**. The residue was purified by fight column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, $\frac{1}{1000}$ / \frac 3H). ¹³C NMR (126 MHz, CDCl₃) δ 149.8, 139.1, 129.2, 128.6, 126.9, 126.8, 116.6, 112.4, 56.7, 38.6.

N-methyl-N-phenethylaniline (2ap) (CAS: No. 28059-49-6)⁷

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v) to give the product as yellow oil (97.1 mg, 92%). ¹H NMR (500 MHz, CDCl₃) δ 7.45 – 7.31 (m, 7H), 6.86 (dd, J = 16.1, 7.9 Hz, 3H), 3.71 – 3.66 (m, 2H), 3.01 (s, 3H), 3.00 – 2.96 (m, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 148.9, 139.9, 129.4, 128.9, 128.6, 126.3, 116.3, 112.3, 54.9, 38.6, 33.0.

N-ethyl-N-(2-(trifluoromethyl)phenethyl)aniline (2aq)

The reaction was performed by following the general procedure **3**. The residue was purified by this column chromatograph (silica gel, petroleum ether:AcOEt = 200:1, v/v) to give the product as a yellow oil (128.9 mg, 88%). ¹H NMR (500 MHz, CDCl₃) δ 778 (d, J_{CF} 7.8 Hz, 1H), 7.59 (t, J = 7.5 Hz, 1H), 7.51 – 7.32 (m, 4H), 6.93 – 6.86 (m, 2H), 6.83 (dd, J = 0.4, 4.0 Hz, 1H), 3.65 (dd, J = 9.9, 5.6 Hz, 2H), 3.45 (q, J = 7.0 Hz, 2H), 3.20 (t, J = 7.5 Hz, 2H), 1.27 (ddd, J = 7.1, 4.4, 1.8 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 147.6, 138.4, 132.0 (d, J = 7.4 Hz), 129.5, 129.3, 128.9, 128.7, 126.4, 126.2 (q, J = 5.7 Hz), 116.0, 111.9, 52.2, 45.1, 30.9, 12.4. ¹⁹F NMR (471 MHz, CDCl₃) δ - 59.32 – -59.38 (m).

HRMS (ESI, m/z) calcd for C₁₇H₁₉F₃N[M+H]⁺: 294.1464; found: 294.1467.

N-butyl-N-(4,4,4-trifluorobutyl)aniline (2ar)

The reaction was performed by following the general procedure **3**. The residue was purified ^{Ph}_{Dy} flash column chromatograph (silica gel, petroleum ether:AcOEt = 200:1, v/v) to give the product as a yellow oil (114.0 mg, 88%). ¹H NMR (500 MHz, CDCl₃) 53 -34 (dd, *J* = 11.0, 4.4 Hz, 2H), 6.85 – 6.77 (m, 3H), 3.45 (t, *J* = 7.2 Hz, 2H), 3.37 (dd, *J* = 10.4, 4.8 Hz, 2H), 2.29 – 2.17 (m, 2H), 2.01 – 1.93 (m, 2H), 1.72 – 1.66 (m, 2H), 1.48 (dt, *J* = 15.0, 7.4 Hz, 2H), 1.11 – 1.06 (m, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 148.0, 129.4, 127.3 (q, *J* = 275.0 Hz), 116.2, 112.4, 51.1, 49.8, 31.4 (q, *J* = 28.8 Hz), 29.4, 20.4, 20.1 (d, *J* = 2.1 Hz), 14.0. ¹⁹F NMR (471 MHz, CDCl₃) δ -65.93 (s). HRMS (ESI, m/z) calcd for C₁₄H₂₁F₃N[M+H]⁺: 260.1621; found: 260.1623.

N-benzyl-N-(2,2-difluoroethyl)aniline (2as)

The reaction was performed by following the general procedure 3. The residue was purified by trash column chromatograph (silica gel, petroleum ether:AcOEt = 200:1, to give the product as a yellow oil (105.0 mg, 85%). ¹H NMR (500 MHz, CDCl₃)

δ 7.49 – 7.31 (m, 7H), 6.90 (d, J = 6.7 Hz, 3H), 6.25 – 5.94 (m, 1H), 4.78 (s, 2H), 3.88 (td, J = 14.0, 4.2 Hz, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 148.2, 137.9, 129.6, 128.8, 127.2, 126.7, 118.0, 114.9 (t, J = 242.5 Hz), 112.8, 55.1, 53.6 (t, J = 26.3 Hz). ¹⁹F NMR (471 MHz, CDCl₃) δ -120.27 (dd, J = 9.9, 5.2 Hz).

HRMS (ESI, m/z) calcd for $C_{15}H_{16}F_2N[M+H]^+$: 248.1245; found: 248.1245.

N-(2,2,2-trifluoroethyl)aniline (2at) (CAS: No. 351-61-1)³

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 30:1, v/v) to give the product as yellow oil (63.0 mg, 72%). ¹H NMR (500 MHz, CDCl₃) δ 7.28 – 7.23 (m, 2H), 6.84 (t, J = 7.4 Hz, 1H), 6.72 (d, J = 7.8 Hz, 2H), 3.95 (s, 1H), 3.80 (qd, J = 8.9, 5.6 Hz, 2H). ¹³C NMR (126 MHz, CDCl₃) δ 146.2, 129.4, 125.1(q, J = 278.8 Hz), 119.1, 113.1, 46.0 (q, J = 33.8 Hz).

N-(((3r,5r,7r)-adamantan-1-yl)methyl)-N-(2,2-difluoroethyl)aniline

(2au)

The reaction Physics performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether: AcOEt = 200:1, 1/10 to give the product as light green oil (122.1 mg, 80%). ¹H NMR (500 MHz, CDCl₃) 7.30 – 7.25 (m, 2H), 6.90 (d, J = 8.2 Hz, 2H), 6.78 (t, J = 7.3 Hz, 1H), 5.96 (tt, J = 56.1, 4.3 Hz, 1H), 3.82 (td, J = 13.5, 4.3 Hz, 2H), 3.23 (s, 2H), 2.03 (s, 3H), 1.76 (d, J = 12.1 Hz, 3H), 1.69 (d, J = 11.4 Hz, 3H), 1.64 (d, J = 2.6 Hz, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 148.7, 129.2, 117.0, 114.1(t, J = 243.8 Hz), 113.1, 62.9, 54.9(t, J = 26.3 Hz), 41.5, 37.3, 37.0, 28.4. ¹⁹F NMR (471 MHz, CDCl₃) δ -120.39 (d, J = 3.5 Hz).

HRMS (ESI, m/z) calcd for C₁₉H₂₆F₂N[M+H]⁺: 306.2028; found: 306.2029.

(4-chlorophenyl)(5-methoxy-2-methyl-3-(2-

(methyl(phenyl)amino)ethyl)-1H-indol-1-yl)methanone (2av)

The reaction was performed by following the general procedure **3**. The residue was purified by flash-column chromatograph (silica gel, petroleum ether:AcOEt = 15:1, v/v) by the product as yellow oil (175.0 mg, 81%). ¹H NMR (500 MHz, CDCl₃) δ 7.59 – 7.54 (m, 2H), 7.50 – 7.44 (m, 2H), 7.26 (dd, J = 10.3, 5.6 Hz, 2H), 7.02 (d, J = 9.0 Hz, 1H), 6.95 (d, J = 2.5 Hz, 1H), 6.78 – 6.70 (m, 4H), 3.84 (s, 3H), 3.65 (t, J = 7.0 Hz, 2H), 2.95 (d, J = 7.1 Hz, 2H), 2.93 (s, 3H), 2.25 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 168.3, C156.0, 148.7 139.0, 134.5, 134.2, 131.1, 131.0, 129.3, 129.1, 117.3, 116.1, 115.1, 111.8, 111.3, 101.0, 55.7, 52.1, 38.6, 21.6, 13.4.

HRMS (ESI, m/z) calcd for C₂₆H₂₆ClN₂O₂[M+H]⁺: 433.1677; found: 433.1679.

N-methyl-3-phenyl-3-(4-(trifluoromethyl)phenoxy)propan-1-amine

(2aw) (CAS: No. 54910-89-3)14

The reaction \mathbf{K}_{33} performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether: AcOEt = 20:1, v/v) to give the product as a white solid (80.3 mg, 52%). M. P. 157 °C (Lit. 157-159 °C). ¹H NMR (590 MHz, DMSO) δ 7.56 (d, J = 8.2 Hz, 2H), 7.49 – 7.33 (m, 4H), 7.29 (t, J = 6.8 Hz, 1Hp), 7.09 (d, J = 8.1 Hz, 2H), 5.75 (s, 1H), 2.97 (s, 2H), 2.51 (s, 3H), 2.32 (dd, J = 13.5, 6.4 Hz, 1H), 2.20 (d, J = 6.2 Hz, 1H). ¹³C NMR (126 MHz, DMSO) δ 160.6, 140.6, 129.2, 128.5, 128.1, 127.3 (d, J = 3.6 Hz), 126.4, 124.9(d, J = 270.0 Hz), 121.8(q, J = 32.5 Hz), 116.8, 77.0, 45.7, 34.8, 33.1. ¹⁹F NMR (471 MHz, DMSO) δ - 59.98 (s).

1-(2-(dimethylamino)-1-(4-methoxyphenyl)ethyl)cyclohexan-1-ol (2ax)

(CAS: No. 93413-69-5)¹⁵

The reaction was performed by following the general procedure **3**. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 10:1, v/v) to give the product as a white solid (84.5 mg, 61%). M. P. 102 °C (Lit. 102-104 °C). ¹H NMR (500 MHz, DMSO) δ 7.23 (d, *J* = 8.5 Hz, 2H), 6.88 (d, *J* = 8.6 Hz, 2H), 3.74 (s, 3H), 5.50 (d, *J* = 10.3 Hz, 1H), 3.22 (s, 1H), 3.03 (dd, *J* = 8.4, 4.2 Hz, 1H), 2.50 (s, 6H), 1.12 - 1.52 (m, 2H), 1.49 - 1.24 (m, 5H), 1.18 - 1.02 (m, 2H), 0.95 (q, *J* = 11.5 Hz, 1H). ¹³C NMR (126 MHz, DMSO) δ 158.5, 132.1, 131.0, 113.9, 72.6, 59.1, 55.4, 50.8, 44.0, 36.8, 33.5, 25.8, 21.8, 21.5.

5. NMR spectroscopic data

N-ethyl-N-methylaniline (2a)





N-ethyl-N,4-dimethylaniline (2b)



N-ethyl-4-methoxy-N-methylaniline (2c)



N-ethyl-4-fluoro-N-methylaniline (2d)



4-chloro-N-ethyl-N-methylaniline (2e)





4-bromo-N-ethyl-N-methylaniline (2f)



N-ethyl-N-methyl-4-nitroaniline (2g)



N,N-diethylnaphthalen-1-amine (2h)





1-ethylindoline (2i)





1-ethyl-1,2,3,4-tetrahydroquinoline (2j)







4-allyl-N-ethyl-N-methylaniline (2l)


2-(4-(ethyl(methyl)amino)phenyl)ethan-1-ol (2m)



N,N-dibenzylethanamine (2n)



N-ethylaniline (20)





N-ethyl-3-methylaniline (2p)





N,4-diethylaniline (2q)



N,N-diethylaniline (2r)





N-methylaniline (2s)





N,N-dimethylaniline (2t)



N-butylaniline (2u)



N-(cyclopropylmethyl)aniline (2v)





N-(cyclohexylmethyl)aniline (2w)



N-(2-phenylpropyl)aniline (2x)





N-(2-phenoxyethyl)aniline (2y)





N-(((3r,5r,7r)-adamantan-1-yl)methyl)aniline (2aa)





N-benzylaniline (2ab)





N-(4-fluorobenzyl)aniline (2ac)







N-(4-methoxybenzyl)aniline (2ad)



N-(4,5-dimethoxy-2-nitrobenzyl)aniline (2ae)







N-(2-(4-isobutylphenyl)propyl)aniline (2af)





N-(furan-2-ylmethyl)aniline (2ag)



N-(thiophen-2-ylmethyl)aniline (2ah)





N-(hex-3-en-1-yl)aniline (2ai)







N-((2E,4E)-hexa-2,4-dien-1-yl)aniline (2aj)



2-methyl-1-phenylpyrrolidine (2ak)





1,2-diphenylpyrrolidine (2al)





N-benzyl-4-methoxyaniline (2am)





4-methoxy-N-neopentylaniline (2an)


N-benzyl-N-methylaniline (2ao)





N-methyl-N-phenethylaniline (2ap)







N-ethyl-N-(2-(trifluoromethyl)phenethyl)aniline (2aq)



N-butyl-N-(4,4,4-trifluorobutyl)aniline (2ar)

















N-(((3r,5r,7r)-adamantan-1-yl)methyl)-N-(2,2-difluoroethyl)aniline



(2au)





(4-chlorophenyl)(5-methoxy-2-methyl-3-(2-



(methyl(phenyl)amino)ethyl)-1H-indol-1-yl)methanone (2av)

N-methyl-3-phenyl-3-(4-(trifluoromethyl)phenoxy)propan-1-amine









1-(2-(dimethylamino)-1-(4-methoxyphenyl)ethyl)cyclohexan-1-ol (2ax)



6. Reference

- 1. I. Sorribes, K. Junge and M. Beller, J. Am. Chem. Soc. 2014, 136, 14314.
- 2. I. Sorribes, J. R. Cabrero-Antonino, C. Vicent, K. Junge and M. Beller, J. Am. Chem. Soc. 2015, 137, 13580.
- 3. M.-C. Fu, R. Shang, W.-M. Cheng and Y. Fu, *Angew. Chem. Int. Ed.* 2015, **54**, 9042.
- 4. Y. Wei, C. Zhao, Q. Xuan, Q. Song, Org. Chem. Front. 2017, 4, 2291.
- 5. B. Abarca, R. Adam and Ballesteros, R. Org. Biomol. Chem. 2012, 10, 1826.
- T. D. Svejstrup, A. Ruffoni, F. Juliá, V. M. Aubert and D. Leonori, Angew. Chem. Int. Ed. 2017, 56, 14948.
- 7. H. Kim, Y. Yonekura and J. Yoshida, Angew. Chem. Int. Ed. 2018, 57, 4063.
- R. Leiva, M. Barniol-Xicota, S. Codony, T. Ginex, E. Vanderlinden, M. Montes, M. Caffrey, F. J. Luque, L. Naesens and S. Vazquez, *J. Med. Chem.* 2018, 61, 98.
- 9. F. Hermant, E. Nicolas and Y. Six, Tetrahedron, 2014, 70, 3924.
- 10. K. Hemming, M. J. Bevan, C. Loukou, S. D. Patel and D. Renaudeau, *Synlett.* 2000, **11**, 1565.
- 11. B. Sreedhar and V. S. Rawat, Synthetic Communications, 2012, 42, 2490.
- 12. E. A. Leo, L. R. Domingo, M. A. Miranda and R. Tormos, *J. Org. Chem.* 2006, **71**, 4439.
- 13. Y. Corre, W. Iali, M. Hamdaoui, X. Trivelli, J.-P. Djukic, F. Agbossou-Niedercorn and C. Michon, *Catal. Sci. Technol.* 2015, **5**, 1452.
- 14. R. Tao, Y. Yin, Y. Duan, Y. Sun, Y. Sun, F. Cheng, J. Pan, C. Lu and Y. Wang, *Tetrahedron* 2017, **73**, 1762.
- 15. N. B. Gowda, G. K. Rao and R. A. Ramakrishna, *Tetrahedron Letters*, 2010, **51**, 5690.