Electronic Supplementary Information (ESI) for

## A miraculous chiral Ir-Rh bimetallic nanocatalyst for asymmetric hydrogenation of activated ketones

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#### 1. General Information

Unless otherwise noted, all chemicals were purchased from commercial sources without further purification. IrCl<sub>3</sub> (99.99%), RhCl<sub>3</sub>•xH<sub>2</sub>O (99.99%) and 1,1-dimethoxyacetone **1a** (97%) were purchased from Alfa Aesar. 1-(1,3-Dioxan-2-yl)ethanone 1b, 1,1-diethoxy-2-propanone 1c, 1,1-dipropoxy-2-propanone 1d, 1,1-dibutoxy-2propanone 1e, 1,1-diisopropoxy-2-propanone 1f and 1,1-ditertbutoxy-2-propanone 1g were synthesized as described in the literatures.<sup>1-2</sup> 2,2-Diethoxyacetophenone 1h (96%) was purchased from Acros. 3,3-Dimethoxybutan-2-one 1i (97%) and 3-hydroxy-2-butanone 1k (97%) were from Adamas.1-Hydroxybutan-2-one 1j (98%), 2-hydroxy-1-phenylethanone 1l (97%) and (1-hydroxycyclohexyl)(phenyl)methanone 1m (98%) were from Ark. Pyruvic aldehyde (30 wt.% in H<sub>2</sub>O) was from Accela. 1,3-Propanediol (98%) was from Innochem. Chiral ionic liquid CIL<sub>PEG-CD750</sub> ([CH<sub>3</sub>(OCH<sub>2</sub>CH<sub>2</sub>)<sub>16</sub>CD]<sup>+</sup>[CH<sub>3</sub>SO<sub>3</sub>]<sup>-</sup> (CD = cinchonidine)) was synthesized as reported<sup>3</sup>. Ethyl alcohol, propanol, isopropanol, n-butanol, t-butyl alcohol, toluene, glacial acetic acid, n-heptane and cyclohexane were all analytical reagents and purchased from Kermel. <sup>1</sup>H NMR was recorded on a Varian (400 MHz). Chemical shifts ( $\delta$ ) were denoted in ppm using residual solvent peaks as internal standard (CDCl<sub>3</sub>,  $\delta$  = 7.26 ppm). Chiral GC analyses were carried out on Fuli 9790 GC instrument equipped with an Agilent CP-Chirasil-Dex (25 m × 0.25 mm × 0.25 μm) and an FID detector (N<sub>2</sub> as a carrier gas). HRTEM (Transmission electron microscopic) images were carried out by using a Tecnai F 30 (300 kV) instrument. ICP-AES (Inductively coupled plasma atomic emission spectrometer) analyses were performed on Optima 2000 DV (detection limit is 10 ppm).

#### 2. Experiment Procedures

#### 2.1 Preparation of chiral Ir-Rh bimetallic nanocatalyst

The chiral Ir-Rh bimetallic nanocatalyst was prepared as a typical example: a mixture of  $IrCl_3$  (2.13 mg, 7.13 ×  $10^{-3}$  mmol), RhCl<sub>3</sub>•xH<sub>2</sub>O (0.47 mg,  $1.78 \times 10^{-3}$  mmol) and ClL<sub>PEG-CD750</sub> (30 mg,  $26.7 \times 10^{-3}$  mmol) was added to a 75 mL stainless-steel autoclave. Then the autoclave was flushed for three times with 2.0 MPa H<sub>2</sub> and inflated to 4.0 MPa (H<sub>2</sub>). After being stirred at 70 °C for 7 h, the reactor was cooled to room temperature and depressurized. At

this time, the color of the mixture changed from claret to black, which indicated that the chiral Ir-Rh bimetallic nanocatalyst formed. The preparation of other chiral Ir-Rh bimetallic nanocatalysts was carried out according to the same procedure.

#### 2.2 Asymmetric hydrogenation of activated ketones

1,1-Dimethoxyacetone **1a** was used as a model reaction substrate: the autoclave was charged with the aboveprepared chiral Ir-Rh bimetallic nanocatalyst ,  $CIL_{PEG-CD750}$  (12 mg), glacial acetic acid (0.85 g), toluene (0.3 g), *n*heptane (0.15 g) and cyclohexane (50 mg, internal standard) and 1,1-dimethoxyacetone (53 mg, **1a**/Ir-Rh = 50, mol/mol). Then the reactor was flushed three times with 2.0 MPa H<sub>2</sub> and pressurized to the required hydrogen pressure at an appointed temperature for a designated time. After reaction, the autoclave was cooled in an icewater bath and then depressurized. The lower chiral Ir-Rh bimetallic nanocatalyst phase was easily separated from the upper product phase by simple phase separation and directly reused in next catalytic cycle. The upper phase was directly analyzed by chiral GC.

#### 3. <sup>1</sup>H NMR Data



**1-(1,3-Dioxane-2-yl)ethanone 1b:** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  = 4.75 (1H, s, OCHO), 3.96-3.78 (4H, m, OCH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>O), 2.24 (3H, s, COCH<sub>3</sub>), 2.15 (2H, m, OCH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>O). The <sup>1</sup>H NMR data were consistent with those reported.<sup>4</sup>



**1,1-Diethoxy-2-propanone 1c:** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  = 4.34 (1H, s, OCHO), 3.43-3.36 (4H, q, OCH<sub>2</sub>CH<sub>3</sub>), 2.24 (3H, s, COCH<sub>3</sub>), 1.27 (6H, t, CH<sub>2</sub>CH<sub>3</sub>). The <sup>1</sup>H NMR data were consistent with those reported.<sup>5</sup>



**1,1-Dipropoxy-2-propanone 1d:** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  = 4.26 (1H, s, OCHO), 3.43-3.36 (4H, t, OCH<sub>2</sub>CH<sub>2</sub>), 2.05 (3H, s, COCH<sub>3</sub>), 1.53 (4H, m, CH<sub>2</sub>CH<sub>3</sub>), 0.90 (6H, t, CH<sub>2</sub>CH<sub>3</sub>). The <sup>1</sup>H NMR data were consistent with those reported.<sup>6</sup>



**1,1-Dibutoxy-2-propanone 1e:** <sup>1</sup>H NMR (400 MHz,  $CDCI_3$ ),  $\delta = 4.26$  (1H, s, OCHO), 3.45 (4H, t,  $OCH_2CH_2$ ), 2.05 (3H, s,  $COCH_3$ ), 1.53-1.40 (8H, m,  $CH_2CH_2CH_3$ ), 0.90 (6H, t,  $CH_2CH_2CH_3$ ). The <sup>1</sup>H NMR data were consistent with those reported.<sup>4</sup>



**1,1-Diisopropoxy-2-propanone 1f:** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  = 4.39 (1H, s, OCHO), 3.83-3.75 (2H, m, OCH(CH<sub>3</sub>)<sub>2</sub>), 2.07 (3H, s, COCH<sub>3</sub>), 1.27 (12H, d, OCH(CH<sub>3</sub>)<sub>2</sub>). The <sup>1</sup>H NMR data were consistent with those reported.<sup>6</sup>



**1,1-Ditertbutoxy-2-propanone 1g:** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  = 4.70 (1H, s, OCHO), 2.24 (3H, s, COCH<sub>3</sub>), 1.26 (18H, s, C(CH<sub>3</sub>)<sub>3</sub>). The <sup>1</sup>H NMR data were consistent with those reported.<sup>6</sup>



**1,1-Dimethoxy-2-propanol 2-a:** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  = 4.08 (1H, d, OCHO), 3.75 (1H, m, CH<sub>3</sub>CHOH), 3.45 (6H, s, OCH<sub>3</sub>), 2.17 (1H, br s, OH), 1.15 (3H, d, CHCH<sub>3</sub>). The <sup>1</sup>H NMR data were consistent with those reported.<sup>7</sup>



**1-(1,3-Dioxane-2-yl)ethanol 2-b:** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  = 4.27 (1H, d, OCHO), 4.15-3.92 (4H, m, OCH<sub>2</sub>), 3.83 (1H, m, CH<sub>3</sub>CHOH), 2.17 (1H, br s, OH), 1.90 (2H, m, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 1.15 (3H, d, CHCH<sub>3</sub>). The <sup>1</sup>H NMR data were consistent with those reported.<sup>4</sup>



**1,1-Diethoxy-2-propanol 2-c:** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  = 4.10 (1H, d, OCHO), 3.75 (1H, m, CH<sub>3</sub>CHOH), 3.53 (4H, q, OCH<sub>2</sub>CH<sub>3</sub>), 2.16 (1H, br s, OH), 1.21 (6H, t, CH<sub>2</sub>CH<sub>3</sub>), 1.15 (3H, d, CHCH<sub>3</sub>). The <sup>1</sup>H NMR data were consistent with those reported.<sup>8</sup>



**1,1-Dipropoxy-2-propanol 2-d:** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  = 4.12 (1H, d, OCHO), 3.75 (1H, m, CH<sub>3</sub>CHOH), 3.57 (4H, t, OCH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 2.16 (1H, br s, OH), 1.50 (4H, m, OCH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 1.15 (3H, d, CHCH<sub>3</sub>), 0.99 (6H, t, OCH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>). The <sup>1</sup>H NMR data were consistent with those reported.<sup>6</sup>



**1,1-Dibutoxy-2-propanol 2-e:** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  = 4.29 (1H, d, OCHO), 3.83 (1H, m, CH<sub>3</sub>CHOH), 3.47 (4H, t, OCH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 2.17 (1H, br s, OH), 1.41-1.27 (8H, m, OCH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 1.15 (3H, d, CHCH<sub>3</sub>), 0.96 (6H, t, OCH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>). The <sup>1</sup>H NMR data were consistent with those reported.<sup>4</sup>



**1,1-Diisopropoxy-2-propanol 2-f:** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  = 4.12 (1H, d, OCHO), 3.75 (1H, m, CH<sub>3</sub>CHOH), 3.63 (2H, m, OCH(CH<sub>3</sub>)<sub>2</sub>), 2.17 (1H, br s, OH), 1.15 (3H, d, CHCH<sub>3</sub>), 0.91 (12H, d, CH(CH<sub>3</sub>)<sub>2</sub>). The <sup>1</sup>H NMR data were consistent with those reported.<sup>6</sup>



**1,1-Ditertbutoxy-2-propanol 2-g:** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  = 4.12 (1H, d, OCHO), 3.75 (1H, m, CH<sub>3</sub>CHOH), 2.17 (1H, br s, OH), 1.15 (3H, d, CHCH<sub>3</sub>), 0.89 (18H, s, C(CH<sub>3</sub>)<sub>3</sub>). The <sup>1</sup>H NMR data were consistent with those reported.<sup>6</sup>



**α-(Diethoxymethyl)benzenemethanol 2-h:** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  = 7.42 (2H, m, CHCCH), 7.24 (3H, m, CHCHCH), 4.60 (1H, d, OCHO), 4.39 (1H, d, CHOH), 3.76 (4H, q, OCH<sub>2</sub>CH<sub>3</sub>), 2.77 (1H, br s, OH), 1.27 (6H, t, OCH<sub>2</sub>CH<sub>3</sub>). The <sup>1</sup>H NMR data were consistent with those reported.<sup>9</sup>

OH

**3,3-Dimethoxy-2-butanol 2-i**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>), δ = 3.85 (1H, q, CH<sub>3</sub>CHOH), 3.28 (6H, s, OCH<sub>3</sub>), 2.17 (1H, br s, OH), 1.27 (3H, s, CCH<sub>3</sub>), 1.15 (3H, d, CHCH<sub>3</sub>). <sup>1</sup>H NMR data were consistent with those reported.<sup>6</sup>



reported.<sup>10</sup>

**2,3-Butanediol 2-k:** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>), δ = 3.89 (2H, m, CHOH), 3.53 (2H, br s, OH), 1.17 (6H, d, CH<sub>3</sub>). <sup>1</sup>H NMR data were consistent with those reported.<sup>11</sup>

OH

OH

OH

**1-Phenyl-1,2-ethanediol 2-I:** <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>),  $\delta$  = 7.32 (2H, m, CHCCH), 7.24 (3H, m, CHCHCH), 4.76 (1 H, t, CHOH), 3.69-3.60 (2H, dd, CH<sub>2</sub>OH), 3.07 (1 H, br s, CHOH), 2.83 (1 H, br s, CH<sub>2</sub>OH),. <sup>1</sup>H NMR data were consistent with those reported.<sup>12</sup>

m, CHCHCH), 4.47 (1H, s, CHOH), 2.48 (1H, br s, CHOH), 1.79 (1H, br s, OH), 1.63-1.21 (10H, m, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>). <sup>1</sup>H NMR data were consistent with those reported.<sup>13</sup>

# $\alpha$ -(1-Hydroxycyclohexyl)benzenemethanol 2-m: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>), $\delta$ = 7.36 (2H, m, CHCCH), 7.25 (3H,



OH





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#### 4. GC Charts





GC analytical conditions for **2a**: the temperature of gasify room, column, and detector were 200 °C, 70 °C and 200 °C, respectively.





GC analytical conditions for **2b**: the temperature of gasify room, column, and detector were 200 °C, 90 °C and 200 °C, respectively.





GC analytical conditions for **2c**: the temperature of gasify room, column, and detector were 200 °C, 90 °C and 200 °C, respectively.





GC analytical conditions for **2d**: the temperature of gasify room, column, and detector were 200 °C, 90 °C and 200 °C, respectively.





GC analytical conditions for **2e**: the temperature of gasify room, column, and detector were 200 °C, 100 °C and 200 °C, respectively.





GC analytical conditions for **2f**: the temperature of gasify room, column, and detector were 200 °C, 90 °C and 200 °C, respectively.





GC analytical conditions for **2g**: the temperature of gasify room, column, and detector were 200 °C, 120 °C and 200 °C, respectively.





GC analytical conditions for **2h**: the temperature of gasify room, column, and detector were 200 °C, 140 °C and 200 °C, respectively.





GC analytical conditions for **2i**: the temperature of gasify room, column, and detector were 200 °C, 70 °C and 200 °C, respectively.





GC analytical conditions for **2j**: the temperature of gasify room, column, and detector were 200 °C, 70 °C and 200 °C, respectively.





GC analytical conditions for **2k**: the temperature of gasify room, column, and detector were 200 °C, 70 °C and 200 °C, respectively.





GC analytical conditions for **2**I: the temperature of gasify room, column, and detector were 200 °C, 140 °C and 200 °C, respectively.





GC analytical conditions for **2m**: the temperature of gasify room, column, and detector were 200 °C, 180 °C and 200 °C, respectively.

### 5. Copies of <sup>1</sup>H NMR Spectra

























#### 6. References

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