

1 **Support information for**

2 **Fabrication and Characterization of DDAB/PLA-alginate**

3 **Composite microcapsules System as Single-shot Vaccine**

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16 **Methods**

17 **Modified Nanoprecipitation Method for DDAB/PLA Nanoparticles**

18 **Preparation**

19 A certain amount of DDAB and PLA was completely dissolved in 15 mL oil  
20 mixture of acetone and ethanol under ultrasonic. Then, the oil phase was slowly  
21 dropped into 90 mL deionized water under magnetic stirring with speed of 400-600  
22 rpm. The suspension was continuously stirred overnight for complete evaporation  
23 of organic solvent. The DDAB/PLA nanoparticles were finally washed and  
24 collected by centrifugation at 20000 g for 15 min for three times. Finally, the  
25 nanoparticles were lyophilized for further experiments.

26 **Single Factor Experiment**

27 In order to determine which factors had significant influence on alginate  
28 micorcapsules size and HBsAg encapsulation efficiency, single-factor experiment  
29 was designed first. Here, we mainly explored the effects of three factors including  
30 gas flow rate, liquid flow rate and the mass ratio of ALG to NPs on the average  
31 particle size of microcapsules and HBsAg encapsulation efficiency.

32 **Response Surface Methodology (RSM) Experiment**

33 According to Box-Behnken model, gas flow rate (A), liquid flow rate (B) and

34 the mass rate of ALG to NPs (C) were employed as factors, and the particle size of  
 35 alginate microcapsules and HBsAg encapsulation efficiency were used as response  
 36 variables. In the previous experiments, we explored the value range of every factor.  
 37 The factors and levels of RSM experiments were shown in Table S1. The different  
 38 responses changed with the initial gas flow rate (450, 600, and 750 L/h), liquid flow  
 39 rate (1.00, 2.00, and 3.00 mL/min) and mass ratio of ALG to NPs (C) (10:1, 15:1  
 40 and 20:1). 17 experimental runs in total decided by the 3<sup>3</sup> factorial Box-Behnken  
 41 models were employed. The quadratic polynomial equation predicted for  
 42 optimization of dependent variables(Y) was Eq. (1):

$$43 \quad Y = \alpha_0 + \sum_{i=1}^3 \alpha_i X_i + \sum_{i=1}^3 \alpha_{ii} X_i^2 + \sum_{i < j=2}^3 \alpha_{ij} X_i X_j \quad (1)$$

44 Where  $\alpha_0$ ,  $\alpha_i$ ,  $\alpha_{ii}$ , and  $\alpha_{ij}$  were regression coefficients;  $X_i$  and  $X_j$  were the input  
 45 variables, which affected the response variable Y. The appropriate program from  
 46 Design Expert™ 8.0.6 was employed.

47 **Table S1** The factors and levels of RSM experiments

Level	Factor		
	Factor A	Factor B	Factor C
	Gas flow (L/h)	Liquid flow Rate (mL/min)	The mass ratio of ALG to NPs (mg/mg)
-1	450	1.00	10:1
0	600	2.00	15:1
1	750	3.00	20:1

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## 49 Results and Discussion

### 50 Single-factor Experiment

51 As a vaccine delivery system, the immune response *in vivo* was significantly  
 52 affected by the antigen encapsulation efficiency and particle size of microcapsules.  
 53 During preparation of alginate microcapsules three factors including liquid flow  
 54 rate, gas flow rate and the mass ratio of ALG to NPs were employed for evaluating

55 their influence on the HBsAg encapsulation efficiency and the particle size of  
56 microcapsules. Other preparation parameters were consistent with follows: sodium  
57 alginate concentration was 1.5 wt%, and the concentration of solidifier of  $\text{CaCl}_2$   
58 was 0.5 M, and the nozzles with 0.7 mm diameter and 15 cm away from liquid  
59 surface was employed.

#### 60 **The Effect of Liquid Flow Rate**

61 The alginate solution containing HBsAg-loaded DDAB/PLA NP suspension  
62 was sprayed under liquid flow rate of 1.00, 2.00, 2.50, 3.00 mL/min, respectively.  
63 The gas flow rate was fixed at 600 L/h, and the mass ratio of ALG to NPs was 15:1.  
64 After solidification, the alginate microcapsules were employed for evaluating the  
65 antigen encapsulation efficiency using BCA assay kit and the particle size  
66 measurement using MasterSizer 2000 Laser Diffraction Particle Size Analyzer. As  
67 shown in Figure S1A, the antigen encapsulation efficiency and particle size of  
68 microcapsules increased with the increase of liquid flow rate. In order to meet the  
69 needs of vaccine formulation for injection, the particle size of alginate  
70 microcapsules should be smaller than 50  $\mu\text{m}$ , and the antigen encapsulation  
71 efficiency should be high enough. The obtained microcapsules size was smaller  
72 than 50  $\mu\text{m}$  and HBsAg encapsulation efficiency was 50-60% when liquid flow rate  
73 was 1.00 and 2.00 mL/min.

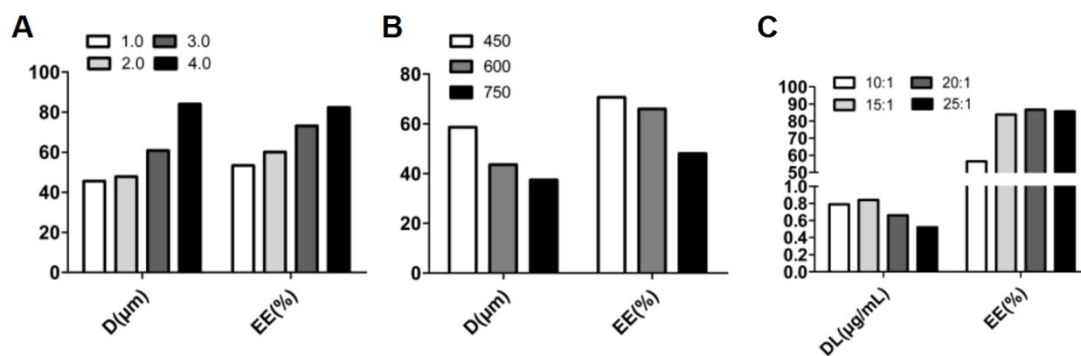
#### 74 **The Effect of Gas Flow Rate**

75 The alginate solution containing HBsAg-DDAB/PLA NPs suspension was also  
76 sprayed under various gas flow rate of  $\text{N}_2$  including 500, 600, 750 L/h, respectively.  
77 The liquid flow rate was fixed at 1.00 mL/min, and the mass ratio of ALG to NPs  
78 was 15:1. The evaluation results of antigen encapsulation efficiency and particle  
79 size of microcapsules were shown in Figure S1B. The antigen encapsulation  
80 efficiency and particle size of microcapsules both decreased with the increase of gas  
81 flow rate. The alginate capsules with size less than 50  $\mu\text{m}$  were obtained when the  
82 gas flow rate was 600 and 750 L/h. The higher shear force led to broken of  
83 microcapsules and leakage of HBsAg-loaded NPs due to higher gas flow rate,  
84 which further resulted in lower HBsAg encapsulation efficiency.

## 85 The Effect of Mass Ratio of ALG to NPs

86 The alginate microcapsules containing HBsAg-loaded DDAB/PLA NPs were  
87 also fabricated using different mass ratio of ALG to NPs as 5:1, 10:1, 15:1, 20:1  
88 and 25:1. The liquid flow rate and the gas flow rate were fixed at 1.00 mL/min and  
89 600 L/h, respectively. The results proved that the mass ratio of ALG to NPs  
90 exhibited almost no effect on particle size of alginate microcapsules, but exhibited  
91 great influence on encapsulation efficiency and loading efficiency of antigen  
92 (Figure S1C in Support information). The results demonstrated that the antigen  
93 loading efficiency was decreased with the increase of mass ratio of ALG to NPs,  
94 while the encapsulation efficiency was increased with augment of mass ratio of  
95 ALG to NPs.

96 The above single-factor experiments demonstrated that the liquid flow rate, the  
97 gas flow rate, and the mass ratio of ALG to NPs had significant influence on  
98 antigen encapsulation efficiency and particle size of microcapsules. Therefore,  
99 these factors were selected for the further response surface method optimization  
100 experiments.



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103 **Figure S1** Single-factor experiments-the influence of the liquid flow rate (A), the gas flow  
104 rate (B), and the mass ratio of ALG to NPs (C) on the HBsAg encapsulation efficiency and  
105 the particle size of microcapsules.

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110 **Table S2** Immunization Study in Mice (n=6)

<b>Groups</b>	<b>Formulation</b>	<b>Immunization at</b>
<b>A</b>	PBS	Days 0, 14, and 28
<b>B</b>	PBS plus HBsAg	Days 0, 14, and 28
<b>C</b>	Aluminum plus HBsAg	Days 0, 14, and 28
<b>D</b>	Microcapsules	Day 0

111

112 **Table S3** ANOVA for response surface reduced quadratic model

<b>Std. Dev.</b>	6.37	<b>R-Squared</b>	0.9294
<b>Mean</b>	58.79	<b>Adj R-Squared</b>	0.8386
<b>C.V. %</b>	10.84	<b>Pred R-Squared</b>	0.1290
<b>PRESS</b>	3509.25	<b>Adeq Precision</b>	10.905

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114

115 **Table S4** ANOVA for response surface reduced quadratic model

<b>Std. Dev.</b>	3.91	<b>R-Squared</b>	0.9592
<b>Mean</b>	34.28	<b>Adj R-Squared</b>	0.9068
<b>C.V. %</b>	11.41	<b>Pred R-Squared</b>	0.6770
<b>PRESS</b>	848.07	<b>Adeq Precision</b>	15.039

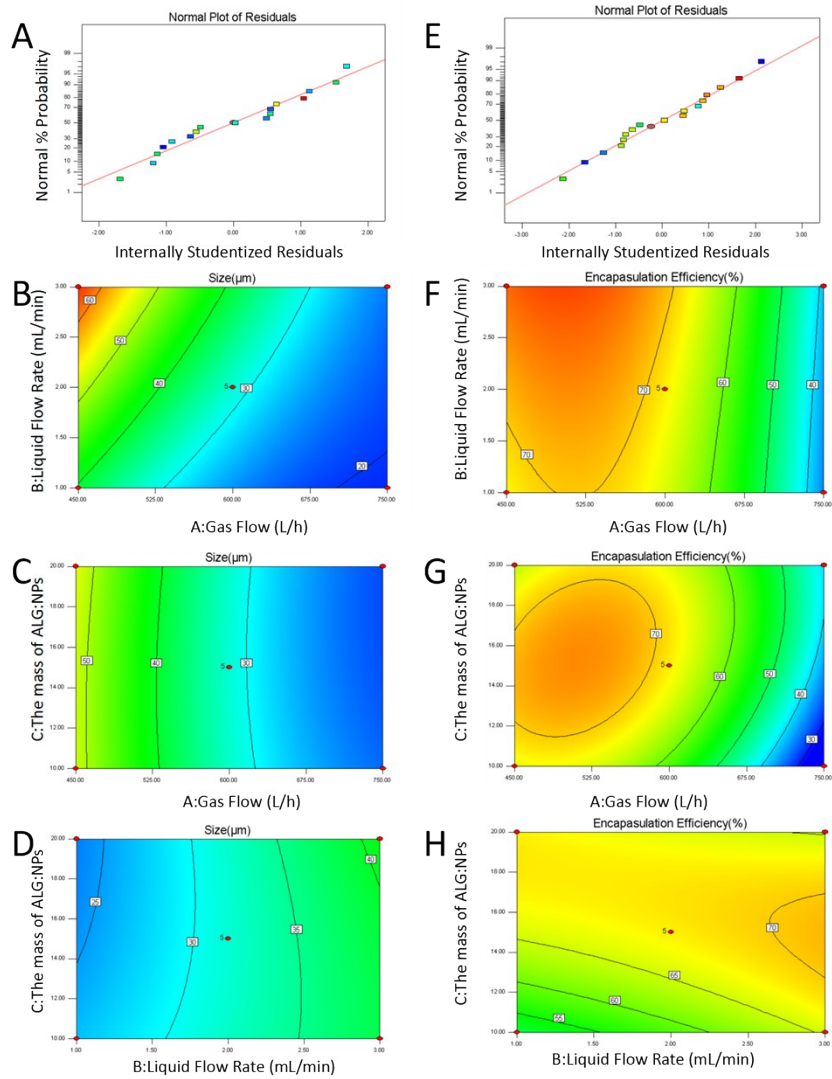
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117 The effects of gas flow, liquid flow rate and the mass ratio of ALG to NPs on  
 118 the microcapsules size are shown in Figure S2B-D. The factors as gas flow rate and  
 119 liquid flow rate have an interaction effect on microcapsules size, and the effect of  
 120 liquid flow rate on microcapsule size was greater than that of gas flow rate (Figure  
 121 S2B). We found that the mass ratio of ALG to NPs had little impact on  
 122 microcapsules size (Figure S2C-D). The factor of liquid flow rate has little impact on  
 123 antigen encapsulation efficiency, and the factor of gas flow rate showed important  
 124 influence on antigen encapsulation efficiency, and the encapsulation efficiency was  
 125 decreased with increase of gas flow rate (Figure S2F). The factors as the mass ratio  
 126 of ALG to NPs and gas flow rate have an interaction effect on encapsulation  
 127 efficiency, and the effect of gas flow rate on encapsulation efficiency was greater

128 than that of the mass ratio of ALG to NPs (Figure S2G). Both factors as the mass  
129 ratio of ALG to NPs and liquid flow rate showed little impact on encapsulation  
130 efficiency when the mass ratio of ALG to NPs was more than 15. However, when  
131 the mass ratio of ALG to NPs was between 10 and 15, the factors as the mass ratio of  
132 ALG to NPs and liquid flow rate demonstrated interaction effect on encapsulation  
133 efficiency, and the effect of the mass ratio of ALG to NPs on encapsulation  
134 efficiency was greater than that of liquid flow rate (Figure S2H). In addition, there is  
135 no strong evidence for the departures from the normal plots of the residuals for size  
136 and encapsulation efficiency. As seen in Figure S2A and E, all the points in the plot  
137 form a straight line. As a result, we can say that the model is fairly suitable.

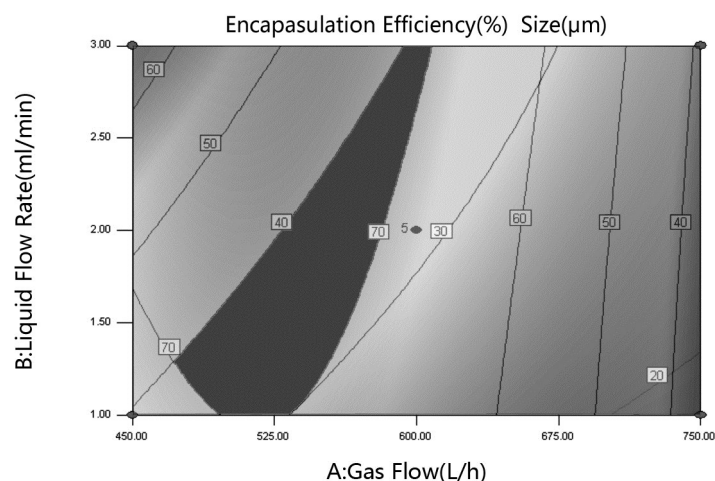
138       The ideal composite microcapsules not only show high encapsulation efficiency,  
139 but also have smaller particle size. Based on the analysis of the response surface  
140 model, we found that the mass ratio of ALG to NPs had little impact on  
141 microcapsules size and antigen encapsulation efficiency. The mass ratio of ALG to  
142 NPs was fixed at 15:1, and the effects of gas flow rate and liquid flow rate on  
143 microcapsules size and antigen encapsulation efficiency were investigated. With  
144 multiple responses, the optimal condition where all parameters simultaneously meet  
145 the desirable criteria could be visually searched by overlaying plot (Figure S3).  
146 Graphical optimization showed the shadow region could meet the above conditions  
147 at the same time, and the encapsulation efficiency of composite microcapsules was  
148 more than 60% and the size was about 30  $\mu\text{m}$ . According to overlay plot, the shadow  
149 region was determined as optimum process parameters.

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151

152 **Figure S2** DOE-RSM analysis of size as a function of gas flow rate, liquid flow rate and the  
 153 mass ratio of ALG to NPs (B; C; D), perturbation analyses (A); DOE-RSM analysis of final  
 154 HBsAg encapsulation efficiency as a function of gas flow rate, liquid flow rate, and the mass  
 155 ratio of ALG to NPs (F; G; H), perturbation analyses (B).



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157 **Figure S3** Overlay plot showing optimal values of gas flow rate, liquid flow rate, and the  
 158 mass ratio of ALG to NPs for achieving an optimal compromise between size and  
 159 encapsulation efficiency

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161 **Table S5** Verification test of RSM (Encapsulation efficiency)

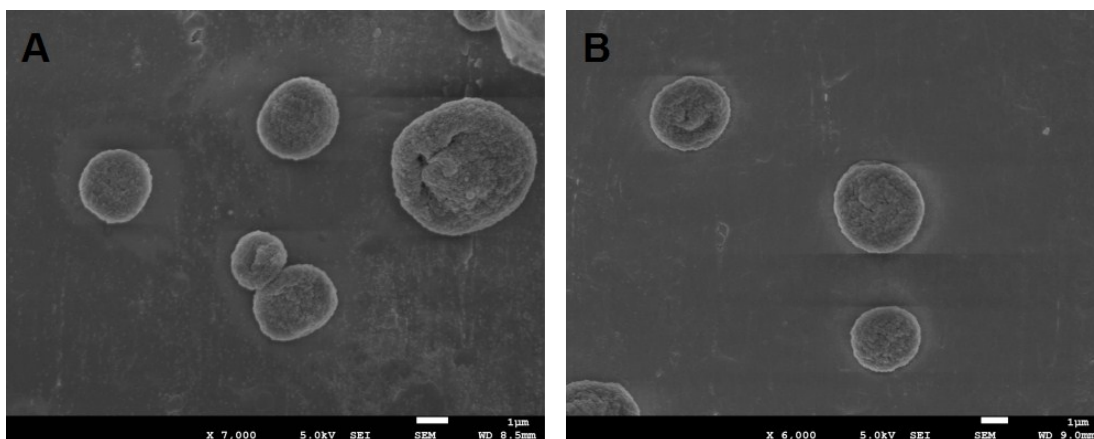
No.	A/L/h	B/ml/min	C/mg/mg	Predicted value/ %	Actual value/ %	Error/ %
1	591.5	1	18.68:1	69.18	70.52	1.9
2	591.5	1	18.68:1	69.18	64.64	6.5
3	591.5	1	18.68:1	69.18	63.7	7.9

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163 **Table S6** Verification test of RSM (Size)

No.	A/L/h	B/ml/min	C/mg/mg	Predicted value/ μm	Actual value/ μm	Error/ %
1	591.5	1	18.68:1	24.249	27.060	11.5
2	591.5	1	18.68:1	24.249	20.473	15.5
3	591.5	1	18.68:1	24.249	26.780	10.4





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165 **Figure S4** The SEM images of alginate microcapsules containing HBsAg-loaded  
 166 DDAB/PLA nanoparticles solidified by  $\text{CaCl}_2$  with concentrations of 0.5 M (A) and 1.0 M  
 167 (B)

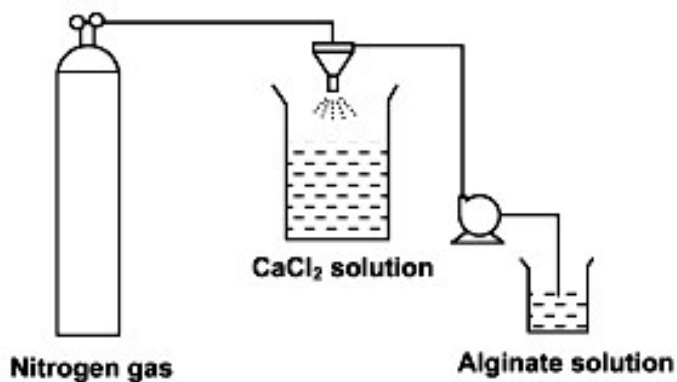
168

169 The microcapsules were prepared employing the equipment as shown in  
 170 Figure S5, which could continuously prepare microcapsules to meet large scale  
 171 productions. We had prepared three batches microcapsules and measured the  
 172 encapsulation efficiency of HBsAg, particle size and morphology of the  
 173 microcapsules. The measurements suggested that three batches microcapsules  
 174 showed similar particle size, morphology and antigen encapsulation efficiency as  
 175 shown in Figure S6 and Table S7.

176



A



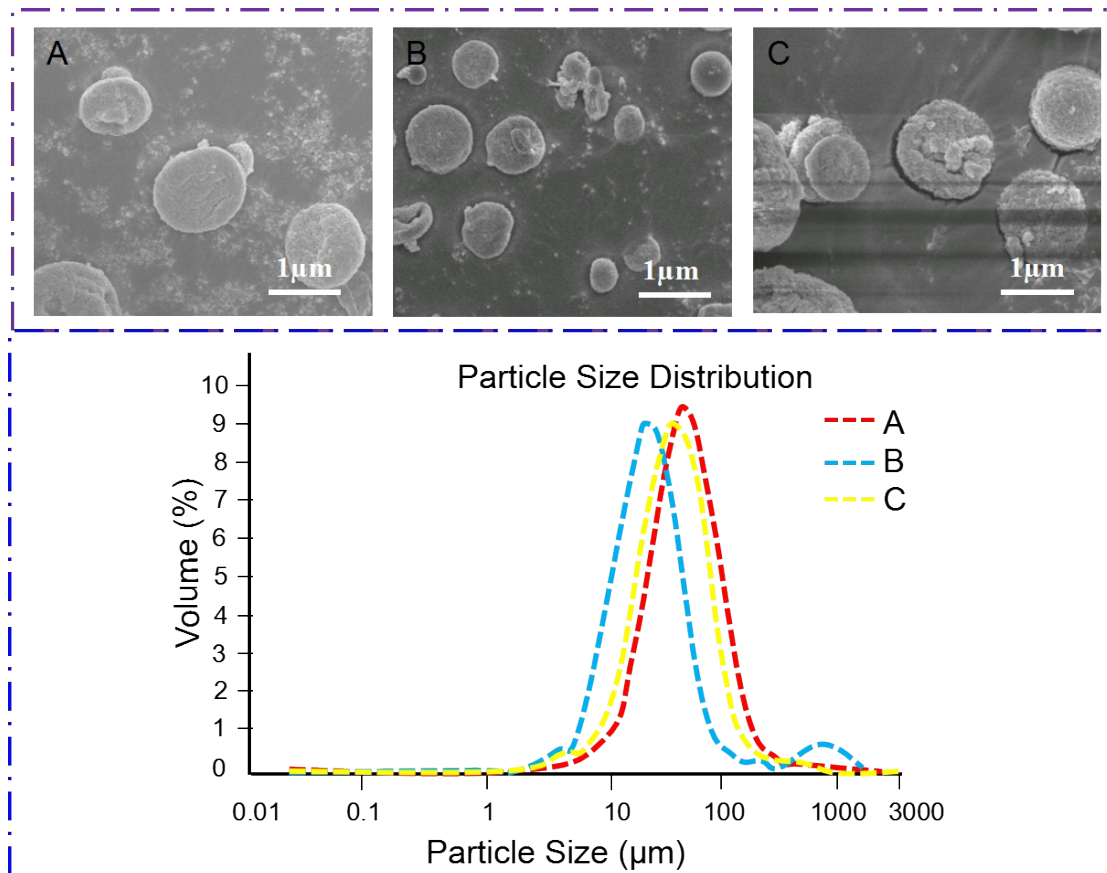
B

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178

179 **Figure S5** The photograph (A) and schematic diagram (B) of equipment for microcapsules  
 180 preparation

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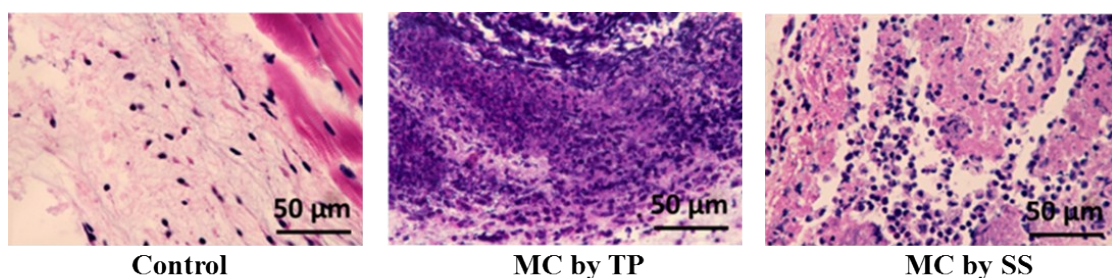


182  
 183 **Figure S6** The SEM micrographs (A-C) and size distributions of three batches HBsAg-  
 184 DDAB/PLA-alginate microcapsules

185  
 186  
 187 **Table S7** The characteristics of three batches HBsAg-DDAB/PLA-alginate microcapsules  
 188

Batches.	Size/ $\mu\text{m}$	Span value	Encapsulation efficiency/%
1	27.060	1.774	70.52
2	20.473	2.465	64.64
3	26.780	1.947	63.70

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190  
 191 **Figure S7** The local inflammation of HBsAg-DDAB/PLA-alginate microcapsules by  
 192 traditional preparation (MC by TP) and spray-solidification method (MC by SS) employing

193 in this study