# High sensitive sensing of hydroquinone and catechol based on $ß-$ cyclodextrin modified carbon dots <br> Zhong-Yi Lin ${ }^{\text {a }}$, Yuan-Jie Guo ${ }^{\text {a }}$, Yu-Syuan Lin ${ }^{\text {a }}$, Chih-Jui Chang ${ }^{\text {c }}$, Tai-Chia Chiu ${ }^{\mathrm{a}, \mathrm{b}}$, Cho-Chun Hu ${ }^{\mathrm{a}, \mathrm{b} *}$ 

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Fig. S1. Fluorescence spectra of C-dot and amono-6-OTs- $\beta-\mathrm{CD}$ mixed with the different ratio ([mono-6-OTs- $\beta-\mathrm{CD}] /[\mathrm{C}-\mathrm{dot}] \mathrm{v} / \mathrm{v}$ : (black) $3: 1$, (red) $4: 1$, (blue) $6: 1$, (pink) $9: 1$, (green) $12: 1$.


Fig. S2. Fluorescence spectra of the synthesized C-dot@ $\beta$-CD at variety incubate time.


Fig. S3. (A) The stability of $\mathrm{C}-\mathrm{dot} @ \beta$ - CD at pH ranges from 5 to 12 , (B) Stability of C - $\operatorname{dot} @ \beta$ - CD in different concentrations of NaCl ranging from 0.1 mM to 1 M , (C) Photostability of C-dot $@ \beta$-CD, (D)Influence of different solvents ( $80 \%$ of total volume) on the fluorescence properties.


Fig. S4. UV-vis absorption of C-dot@ $\beta$-CD(black), CC (red), $\mathrm{HQ}($ blue) and C-dot@ $\beta$-CD fluorescent emission(green).


Fig. S5. Time-resolved decay of C-dot $@ \beta$-CD (black) with HQ (red), with CC(blue).


Fig. S6. Cell viability assay of human HeLa cells against C-dot@ $\beta$-CD at arranged concentrations from $0.35-1.4 \mathrm{mg} / \mathrm{mL}$.

TableS1. Time-resolved decay of C-dots in the absence and presence of catechol and hydroquinone

| Sample | $\tau_{1}(\mathrm{~ns})$ | Area (\%) | $\tau_{2}(\mathrm{~ns})$ | Area (\%) | Average $\tau(n s)$ |
| :---: | :---: | :---: | :---: | :---: | :---: |
| C-dot@ $\beta$-CD | 13.65 | 64.08 | 3.38 | 35.92 | 9.96 |
| C-dot@ $\beta$-CD | 5.11 | 62.80 | 1.73 | 37.20 | 3.85 |
| + CC |  |  |  |  |  |
| C-dot@ $\beta$-CD | 5.63 | 63.68 | 1.96 | 36.32 | 4.30 |
| + HQ |  |  |  |  |  |

