

## Supplementary information

### **Enhanced production of optical (*S*)-acetoin by recombinant *Escherichia coli* whole-cell biocatalyst with NADH regeneration**

Jian-Xiu Li <sup>1,2</sup>, Yan-Yan Huang <sup>2</sup>, Xian-Rui Chen <sup>2</sup>, Qi-Shi Du <sup>2</sup>, Jian-Zong Meng <sup>1</sup>,  
Neng-Zhong Xie <sup>1,2, \*</sup> and Ri-Bo Huang <sup>1,2, \*</sup>

<sup>1</sup> *State Key Laboratory for Conservation and Utilization of Subtropical Agro-Bioresources, Life Science and Biotechnology College, Guangxi University, 100 Daxue Road, Nanning, 530004, China*

<sup>2</sup> *State Key Laboratory of No-Food Biomass and Enzyme Technology, National Engineering Research Center for No-Food Biorefinery, Guangxi Key Laboratory of Biorefinery, Guangxi Academy of Sciences, 98 Daling Road, Nanning, 530007, China.*

\* Corresponding author.

*E-mail addresses:* xienengzhong@gxas.cn (N.Z. Xie), guruace@163.com (R.B. Huang).

**Table S1. Sequences of codon-optimized genes**

> *fdh*

atgtctaaaggcaaagtctgctggtcctgtatgaaggcggcaaacacgctgaagaacaagaaaaactgctgggctgtatt  
gaaaatgaactgggcattcgaactttatcgaagaacagggttatgaactggttaccacgatcgataaagaccggaaccg  
acctcaacgggtgatcgtgaactgaaagacgcagaaattgtcatcaccacgccgttttcccggcttacattagtcgtaaccg  
catcgcagaagctccgaatctgaaactgtgctcaccgccggcgtgggttccgatcatgttgacctggaagcggccaacg  
aacgcaaaattaccgttacggaagtcaccggctcaaatgtggttccggttcagaacatgtcatggctacgattctggtgctg  
atccgtaactataatggcggtcaccagcaagcaattaacggcgaatgggatatcgcgggtgtggccaaaaacgaatacga  
tetggaagacaaaatcatctccaccgttggcgcaggtcgtattggttaccgctgctggaacgtctggttgccttaaccga  
aaaaactgctgtattacgattaccaggaactgccggcggaagccattaaccgctgaatgaagcatcaaaaactgttcaatgg  
ccgcggtgatcgtgcagcgtgttgaaaaactggaagatatggtggcgaatcggacgtcgtgaccattaactgccgct  
gcataaagatagccgcggcctgtcaacaaaaactgatctctcacatgaaagacggcgcgtatctggttaacacggcacg  
tggtgctatttgtgtggccgaagatgttcagaagctgtcaaaagcggcaaacactggcgggttacggcggtgatgtgtggga  
caaacagccggcaccgaaagatcaccgtggcgtacgatggataacaagaccatgtcggcaatcgatgacctgcac  
atcagcggtagctctctggatgctgcagaaacgttatgcccaaggtgtcaaaaacatcctgaatagttacttctccaaaaaatt  
cgattaccgcccgcaggacattatcgtgcaaaatggctcttatgcgaccctgcctacggtcagaaaaataa

> *gdh*

atgtaccggacctgaagggtaaagtgggtgcgatcaccggtgcggcgagcggctctgggcaaggcgaatggcgatccgttt  
cggcaaggagcaggcgaaggtggttatcaactactacagcaacaagcaagatccgaacgaggttaaggaagaggtgat  
caaagcgggtggcgaagcgggtggttgcagggtgacgttaccaggaagaggacgttaaaaacatcgtgcaaacgc  
gattaaggaatttggcacctggacatcatgattaacaacgcgggtctggagaaccggtccgagccacgaaatgccgct  
gaaggactgggataaagtgatcggcaccaacctgaccggtgcgttctgggcagccgtgaggcgaatcaagtactcgttg  
aaaacgatataagggaacgtgattaacatgagcagcgttcacgaggtgatcccgtggccgctgttcgtgactacgcg  
gagcaaggggtggcattaaactgatgaccgagaccctggcgtggaatgcgccgaagggtatccgtgtaacaacat  
tggtccgggcgatcaacacccgattaacgcggagaaatttgcggaccgaagcagaagcggatgtggagagcat  
gatcccgatgggttatattggcgaaccggaggaaatcgcggcggttgcggcgtggctggcgagcaagaagcagcga  
cgtgaccggtattaccctgttcgaggacgggtggcatgaccagatccgagctttcaagcgggtcgtggctaa

> *Ppdar*

atggagctgaagaacaaaaccgcatcattaccggtgcgagcaagggtatcggcaaacgattgaggagaccctggcg  
aaggaaggtgtaacctgggcctgatcagccgtaccctgaccgacctgcagaagctgcaagatagcctgggtagcacct



> *adh*

atgggtagcaccatgcgtgccgtgcaggtgggttggtatcatcaaacctggaaatgaaagaagtccggttccgaccccg  
accggtccgtttgatgtcgtggttaaaattggcgggtgcgggtgtctgtcgtaccgacctgcatacctggaaggccagtggg  
ccgaaaaatctcaggtgcaactgccgtacacgattggccacgaaaacgctggttgggttgatgcggtcgggtgcggccgtg  
accaatgttcgtgaaggcgacaaagtcattgtgcacccgctgattacctgtggtctgtcgtgcctgccgttcaggtgatga  
cgttcactgtgaagctaacgcgttcccgggtattgataccaatggcgggtatgccgaatacctgaaaacgagcgcacgctct  
gtcgtgaaaatc gatgacaccctggaaccgtcggatgtggcagctctggcggatgcaggtctgaccgcctatcatgcggc  
ggcaaaagcggcccgtcctgacgccgcgtgatcgtgtgttgctcattggtgcgggcggctctgggccacattggtatcca  
ggttctgaaagccctgtccccggcagaactgattgtggttgatcgtaatccggaagcgtgaaactggctgaagcgtcgg  
cgctgatcatggcgtcgtggcggatggcaccaggtggatcaggtgtgacctgacgggcggtcacggcgtgaaacc  
gttatcgttttgcggcgaaggcgggtgcaacgagccaggggtgtggcaatgctgcgtcgcgcagggcattatcacgttgc  
ggctacggtgaaaacattaatgtccgaccatcgatatcatcagtacgaaatcaacttcacggcaatctggttggttctat  
aacgatctgtgtgacctgatggcactggcggcacgcggcgcagttaatctgcataccagaaatacgcactggatgacttc  
caaagtctattgatgatctggatgccggctatgtgcgtggctcgcgcgatcctgacgccgtaa

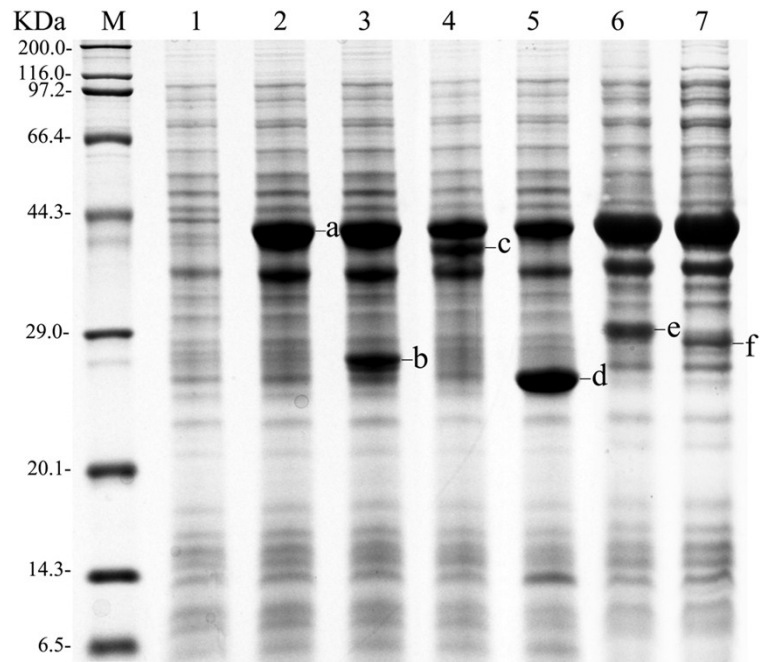
> *adr*

atgagcattacgggcaaagtcgttctggttacgggcgctggtaaggtattggtcgtggtatcgactgcgtctggcacatg  
atggcgtgatatcgcgtggtcgtatctggaccagaccaaactggatgccgtggcagacgaaattcgtcgcacgtggccgt  
cgcgccaccacgtttgtggcagatgtagcaccgtgcacaagtccatcgggccgtggaacatcgcacagtgaaactgtc  
cggtttcgatgtcattgtgaacaatgctggcatcgcgtggttggtccgatttccgatgccaccccggaagaagtctcaaaaa  
tctggtcgggtaacgtggatggcgtgctgtgggtattcaggcagctgcggccaaattcaggcactgggccaacgtggtgta  
aaattatcaatgctagctctattgcgggccacgatggtttgctatgctggcgtgtatagtcgaccaaattcggcgttcgcg  
cactgacgcaagcagctgcgaaagaatatgcttcagacggtatcaccgttaacgcctactgcccgggcgtggttggtaccg  
atatgtgggtgacgattgacaaacgttttcagaactgaccgggtgcaccggaaggtgcaacgtatgaaaaattcgttggtgg  
tatcgtctgggtcgtgcagaaaccccggatgacgttgcggctcgtcagctatctggcaggtccggattctgactacatg  
acgggccaggcgggtctgattgatggcggctcgtgtaccgtaa

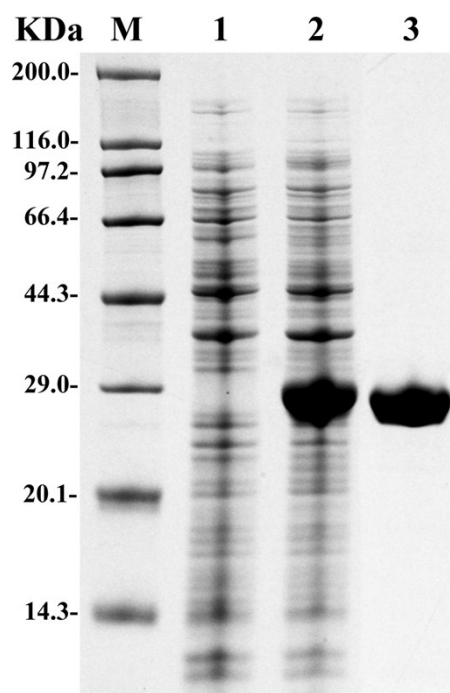
**Table S2. Substrate specificities of KpDAR**

Reaction	Substrate	Relative activity (%)
Reductive reaction <sup>a</sup>	Diacetyl	100 ± 9.07
	( <i>R</i> )/( <i>S</i> )-Acetoin	33.3 ± 4.02
	Ethyl pyruvate	14.9 ± 1.30
	Acetophenone	0.4 ± 0.02
Oxidative reaction <sup>b</sup>	<i>meso</i> -2,3-Butanediol	100 ± 5.6
	(2 <i>S</i> ,3 <i>S</i> )-2,3-Butanediol	0.6 ± 0.06
	(2 <i>R</i> ,3 <i>R</i> )-2,3-Butanediol	0.3 ± 0.01
	( <i>R</i> )/( <i>S</i> )-Acetoin	0.7 ± 0.07
	Phenethyl alcohol	0.7 ± 0.02
	Ethyl acetate	0.7 ± 0.03

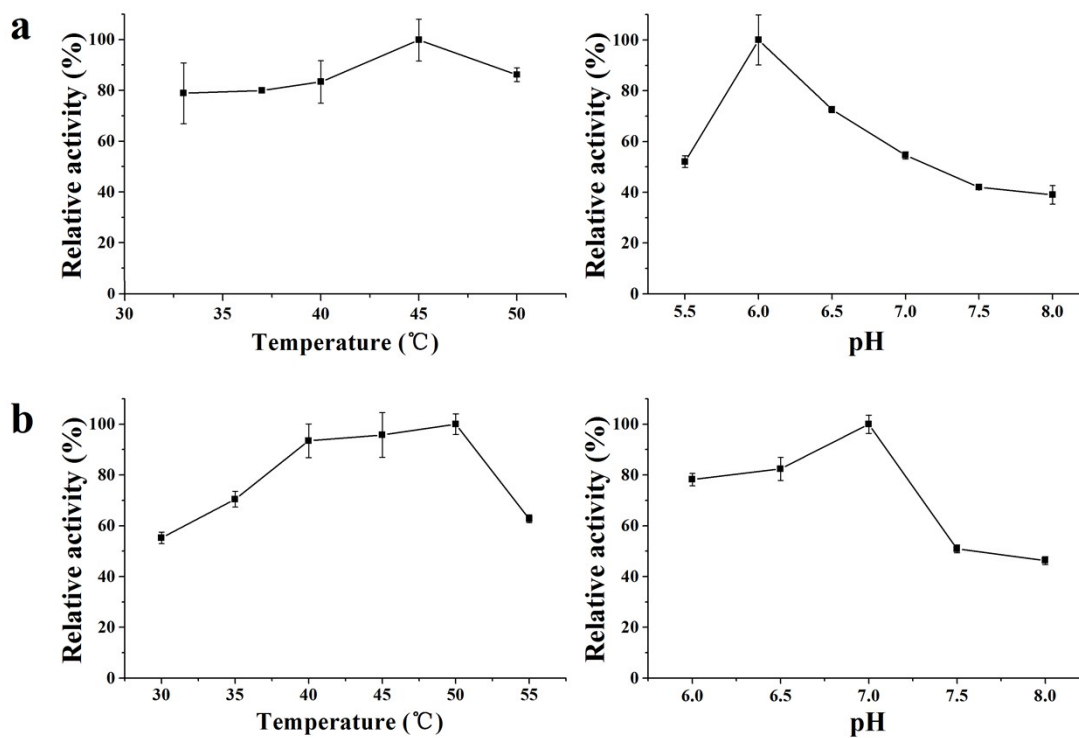
<sup>a</sup> Enzyme activities in ketone reduction were measured with 5 mM substrate and 0.2 mM NADH in 100 mM phosphate buffer, and the relative activity of 100% represents 2887.6 U/mg for diacetyl reduction. <sup>b</sup> Alcohol oxidation reactions were measured with 5 mM substrate and 0.2 mM NAD<sup>+</sup> in 100 mM phosphate buffer, and the relative activity of 100% represents 108.8 U/mg for *meso*-2,3-butanediol oxidation. All reactions were carried out at 45°C and pH 6.0. Data are the averages standard deviations (*n*=3).



**Fig. S1** SDS-PAGE analyses the expression of various DARS. Lane M, marker proteins; lane 1, *E. coli* BL21 (DE3) /pETDuet-1; lane 2, *E. coli* BL21/ pETDuet-*fdh*; lane 3, *E. coli* BL21/ pETDuet-*adr-fdh*; lane 4, *E. coli* BL21/ pETDuet-*adh-fdh*; lane 5, *E. coli* BL21/ pETDuet-P*pdar-fdh*; lane 6, *E. coli* BL21/ pETDuet-*ardII-fdh*; lane 7, *E. coli* BL21/ pETDuet-K*pdar-fdh*. a: FDH; b-f: DAR encoded by *adr*, *adh*, P*pdar*, *ardII* and K*pdar*, respectively.



**Fig. S2** SDS-PAGE analysis of purified KpDAR. Lane M, maker; lane 1, crude extract of *E. coli* BL21 (DE3)/pET-28a(+); lane 2, crude extract of *E. coli* BL21/pET 28a(+)-Kpdar; lane 3, purified KpDAR.



**Fig. S3** Effect temperature and pH on the activities of KpDAR. **(a)** Reductive reaction towards diacetyl; **(b)** Reductive reaction towards (*R*)/(*S*)-acetoin. The optimal temperature for diacetyl and (*R*)/(*S*)-acetoin were 45°C and 50°C, respectively. Maximum activities were observed at pH 6.0 and pH 7.0 respectively.