

Synthetic Ionophores as Non-Resistant Antibiotic Adjuvants

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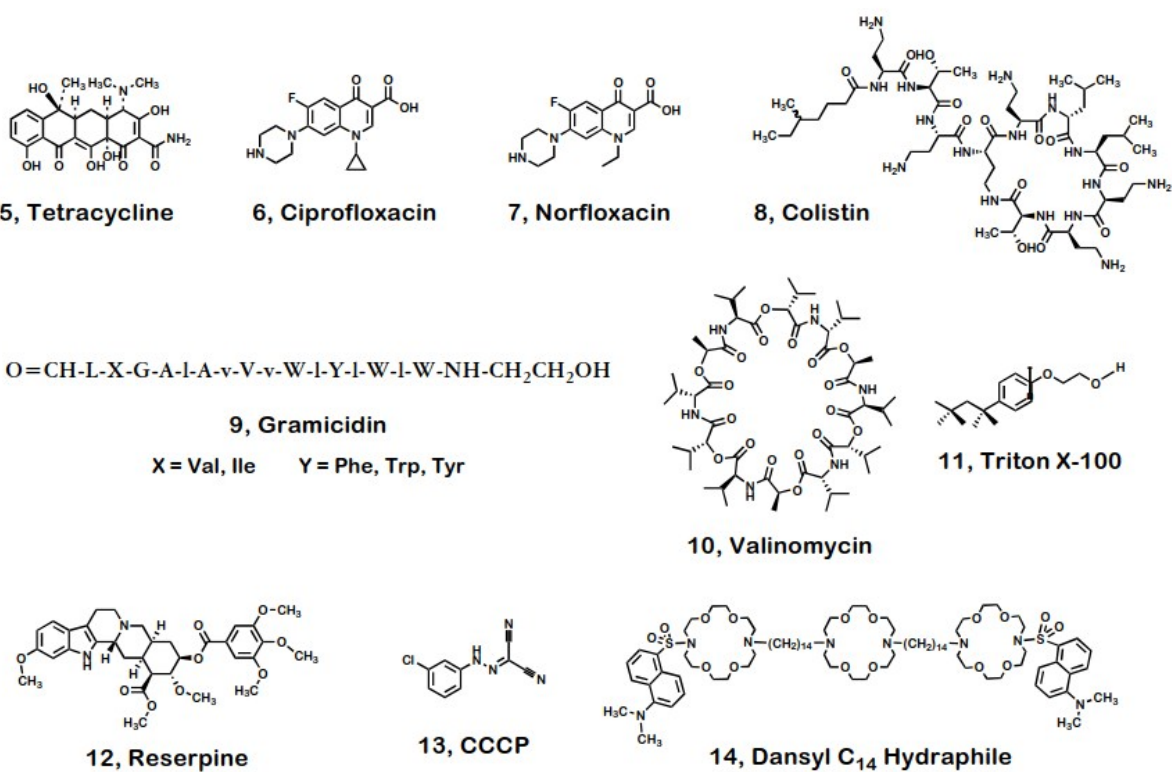


Figure S1. Structures of antibiotics and controls used for the study. The antibiotics studied were tetracycline (5), ciprofloxacin (6), norfloxacin (7) and colistin (8). The channel-former gramicidin-D (9), the carrier valinomycin (10), and the detergent triton X-100 (11) served as controls for ion transport. Reserpine (12) and CCCP (13) are known efflux pump inhibitors. Dansyl C₁₄ hydraphile (14) was synthesized to study localization of hydraphiles in bacteria.

Table S1: MIC of benzyl C₈-C₁₄ hydraphiles (**1** - **4**) against efflux pump expressing tet^R *E. coli*, *K. pneumoniae* (ATCC BAA 2146TM), and *S. aureus* 1199B.

Table S1: MIC in μM			
Compounds used	<i>K. pneumoniae</i> (ATCC BAA 2146TM)	<i>E. coli</i> (tet^R)	<i>S. aureus</i> 1199B
Benzyl C ₈ hydraphile (1)	200	250	128
Benzyl C ₁₀ hydraphile (2)	56	35	16
Benzyl C ₁₂ hydraphile (3)	35	5	1
Benzyl C ₁₄ hydraphile (4)	10	2	≤1
Tetracycline (5)	1000	900	N.D.
Ciprofloxacin (6)	700	0.5	N.D.
Norfloxacin (7)	N.D.	N.D.	64
Colistin (8)	0.25	0.25	N.D.
CCCP (13)	N.D.	56	N.D.
Reserpine (12)	>128	>128	>128

N.D. = not determined; Colistin is inactive against *S. aureus*

Figure S2: Combination of benzyl C₈-C₁₀ hydrophile (1-2) and tetracycline or norfloxacin against tet^R *E. coli*, *K. pneumoniae* and *S. aureus* 1199B.

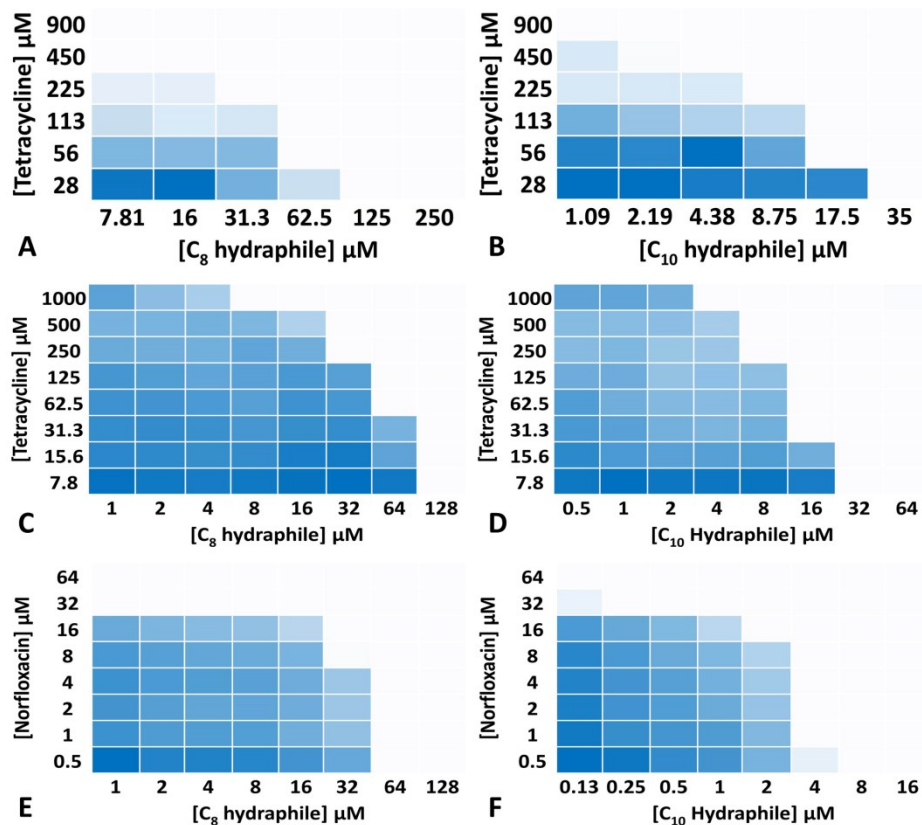


Figure S2. Checkerboard experiments with 1-2 (a-b) with tetracycline against tet^R *E. coli*; (c-d) with tetracycline against *K. pneumoniae* (e-f) with norfloxacin against *S. aureus* 1199B. Color change from blue (0% inhibition) to white (>90% inhibition) indicates increasing growth inhibition.

Figure S3: Tet^R *E. coli* growth curve with benzyl C₈-C₁₄ hydraphile (1-4), tetracycline (5) and their combination.

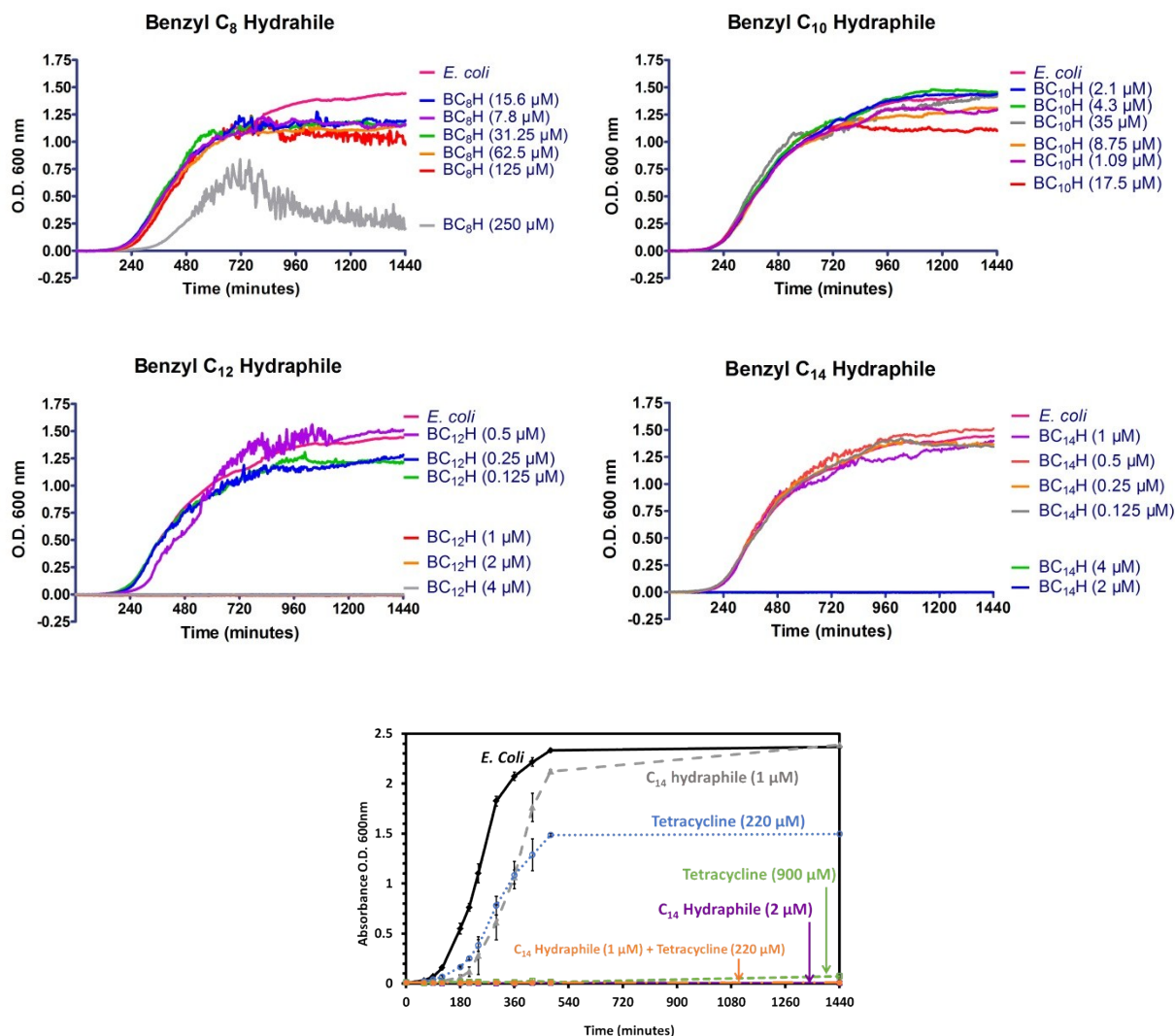


Figure S3. Growth of Tet^R *E. coli* in the presence MIC, and fractional MICs of Benzyl C₈-C₁₄ Hydraphile (1-4) over 24 hours (Four graphs on the top). Readings were taken every 4 minute, average of 2 trials was plotted and error bars were omitted for clarity.

Bottom graph shows the growth of Tet^R *E. coli* in the presence of 1 μM (dashed grey line) and 2 μM (solid purple) benzyl C₁₄ hydraphile, 220 μM (dotted blue line) and 900 μM (dashed green line), and combination of 1 μM C₁₄ hydraphile and 220 μM tetracycline (orange line). The data represents average optical density (600 nm) from three separate trials. The error bars represent standard deviation in the results.

Table S2: Recovery of tetracycline, ampicillin and ciprofloxacin activity against *Klebsiella pneumoniae* by benzyl C₈-C₁₄ hydraphiles.

Table S2: Benzyl C₈-C₁₄ hydraphiles recover antibiotic potency against <i>K. pneumoniae</i>.				
Amphiphile used	[Amphiphile] μM	[Antibiotic] μM	Fold enhancement	FIC index
Antibiotic used: Tetracycline				
No amphiphile	-	1000 \pm 100	n/a	n/a
Benzyl C ₈ hydraphile	2.5	1000 \pm 100	1-fold	1
Benzyl C ₈ hydraphile	50 ($\frac{1}{4}$ MIC)	250 \pm 50	4-fold	0.5
Benzyl C ₈ hydraphile	100 ($\frac{1}{2}$ MIC)	25 \pm 10	40-fold	0.53
Benzyl C ₁₀ hydraphile	2.5	900 \pm 100	1.1-fold	1
Benzyl C ₁₀ hydraphile	14 ($\frac{1}{4}$ MIC)	300 \pm 50	3-fold	0.58
Benzyl C ₁₀ hydraphile	28 ($\frac{1}{2}$ MIC)	125 \pm 25	8-fold	0.63
Benzyl C ₁₂ hydraphile	2.5	500 \pm 50	2-fold	0.57
Benzyl C ₁₂ hydraphile	8.75 ($\frac{1}{4}$ MIC)	300 \pm 25	3-fold	0.58
Benzyl C ₁₂ hydraphile	17.5 ($\frac{1}{2}$ MIC)	125 \pm 25	8-fold	0.63
Benzyl C ₁₄ hydraphile	2.5 ($\frac{1}{4}$ MIC)	350 \pm 50	3-fold	0.58
Benzyl C ₁₄ hydraphile	5 ($\frac{1}{2}$ [MIC])	62.5 \pm 25	16-fold	0.56
Antibiotic used: Ampicillin (beta-lactamase)				
Benzyl C ₁₄ hydraphile	5 ($\frac{1}{2}$ MIC)	> 1000	0-fold	n/a
Antibiotic used: Ciprofloxacin				
No amphiphile	-	700 \pm 100	n/a	n/a
Benzyl C ₈ hydraphile	100 ($\frac{1}{2}$ MIC)	70 \pm 20	10-fold	0.6
Benzyl C ₁₂ hydraphile	8.75 ($\frac{1}{4}$ MIC)	300 \pm 25	2-fold	0.67
Benzyl C ₁₂ hydraphile	17.5 ($\frac{1}{2}$ MIC)	175 \pm 25	4-fold	0.75
Benzyl C ₁₄ hydraphile	2.5 ($\frac{1}{4}$ MIC)	400 \pm 50	2-fold	0.82
Benzyl C ₁₄ hydraphile	5 ($\frac{1}{2}$ MIC)	250 \pm 25	3-fold	0.85

Table S3: Recovery of tetracycline activity against tet^R *E. coli* by gramicidin-D, valinomycin, triton X-100, CCCP, reserpine, and colistin.

Table S3: Combination studies with controls against Tet^R <i>E. coli</i>				
Amphiphile used	[Amphiphile] μM	[Tetracycline] μM	Fold enhancement	FIC index
No amphiphile	-	900 \pm 100	n/a	
CCCP	1	900	1-fold	1
CCCP	21	450	2-fold	0.75
CCCP	42	225	4-fold	0.75
Reserpine	64	450	2-fold	1
Reserpine	128	225	4-fold	1
Colistin	0.03125	225	4-fold	0.52
Colistin	0.0625	56.25	16-fold	0.31
Gramicidin-D	20	900 \pm 100	1-fold	1
Valinomycin	20	450 \pm 100	2-fold	1
Triton X-100	20	450 \pm 100	2-fold	1
Triton X-100	1700 (0.1%)	450 \pm 100	2-fold	1
Colistin	0.0625	56.25	16-fold	0.8
Colistin	0.03125	225	4-fold	0.5
Colistin	0.0156	450	2-fold	0.6

Figure S4: Scanning electron micrographs of tet^R *E. coli* treated with benzyl C₁₄ hydraphile.

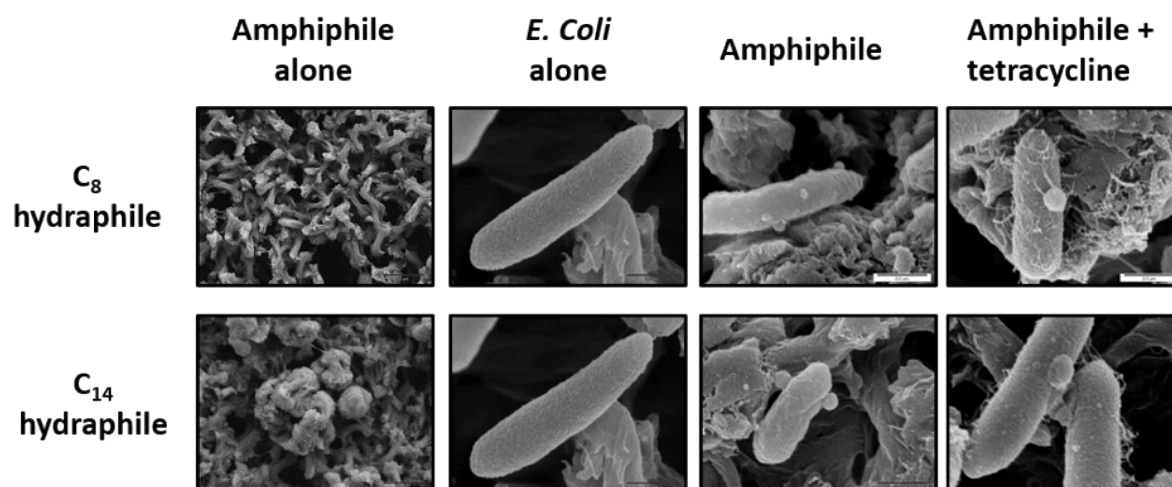


Figure S4. Scanning electron microscopy images of tet^R *E. coli* treated with benzyl C₈ hydraphile (top panel, scale = 0.5 μm) and benzyl C₁₄ hydraphile (bottom panel, scale = 0.5 μm). The column labeled ‘amphiphiles alone’ shows the nylon membrane (top, scale = 2 μm) and benzyl C₁₄ hydraphile aggregate (bottom, scale = 2 μm).

Figure S5: Localization of dansyl labeled C₁₄ hydrophile in human embryonic kidney (HEK-293) cells.

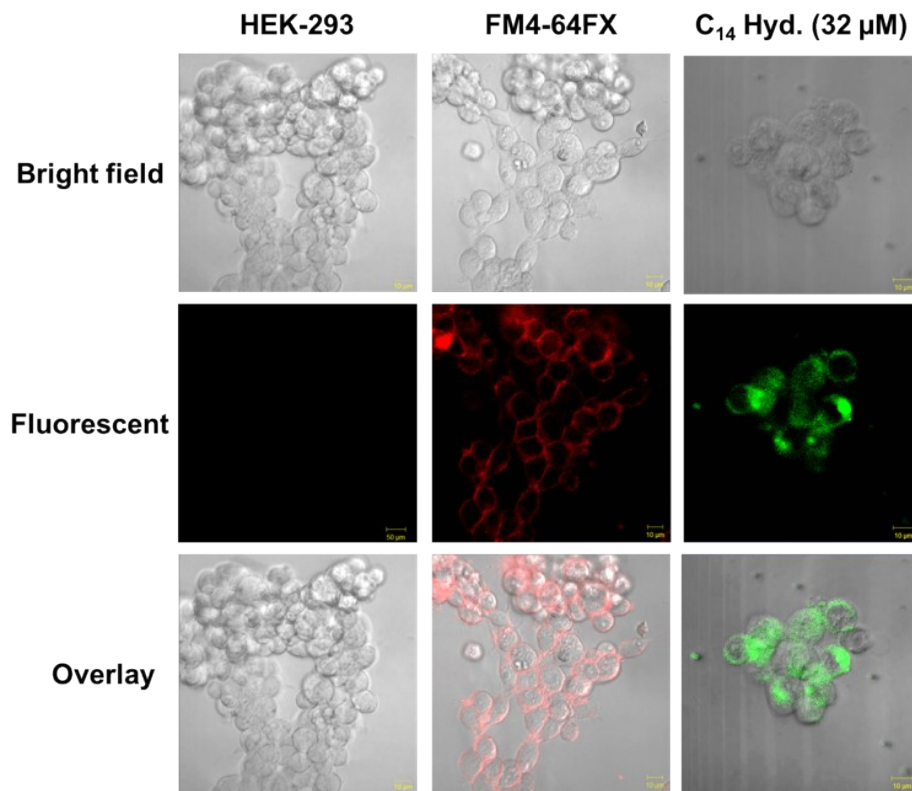


Figure S5. Localization of FM4-64FX (red), and dansyl labeled C₁₄ hydrophile (green) in mammalian cells. The magnification is 400-X and the scale is 10 μm.

Figure S6: Change in membrane permeability of tet^R *E. coli* and *S. aureus* 1199B in the presence of compounds 1-4, triton X-100 and colistin.

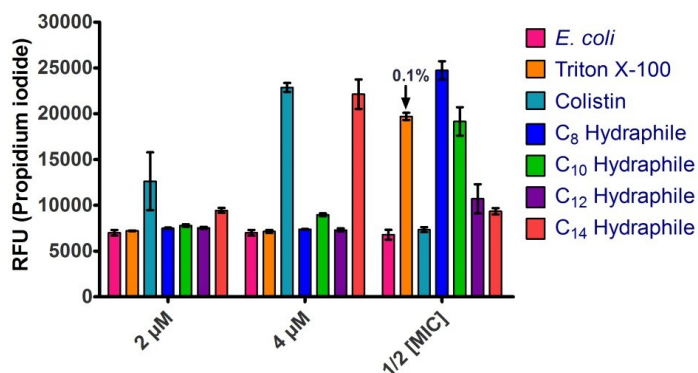


Figure S6a. Permeability of propidium iodide to *E. coli* cell membranes mediated by compounds 1-4, triton X-100 and colistin. The X-axis represents the concentration of the compounds used whereas the y-axis represents the relative fluorescence unit (RFU). Error bars represent standard deviation in three trials.

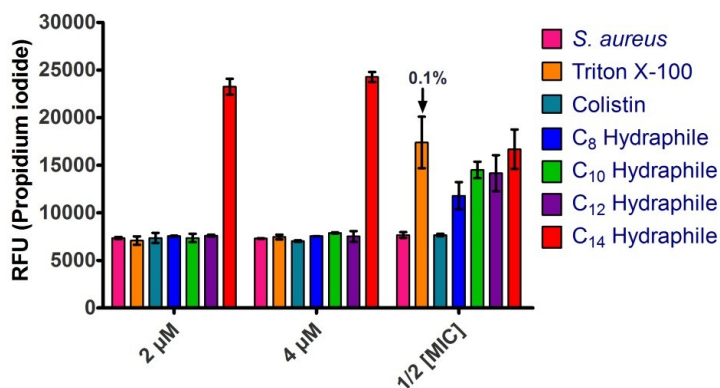


Figure S6b. Permeability of propidium iodide to *S. aureus* cell membranes mediated by compounds 1-4, triton X-100 and colistin. The X-axis represents the concentration of the compounds used whereas the y-axis represents the relative fluorescence unit (RFU). Error bars represent standard deviation in three trials.

Figure S7. Confocal images showing permeability of PI and FDA in tet^R *E. coli* and HEK-293 cells mediated by compounds 1-4 and triton X-100.

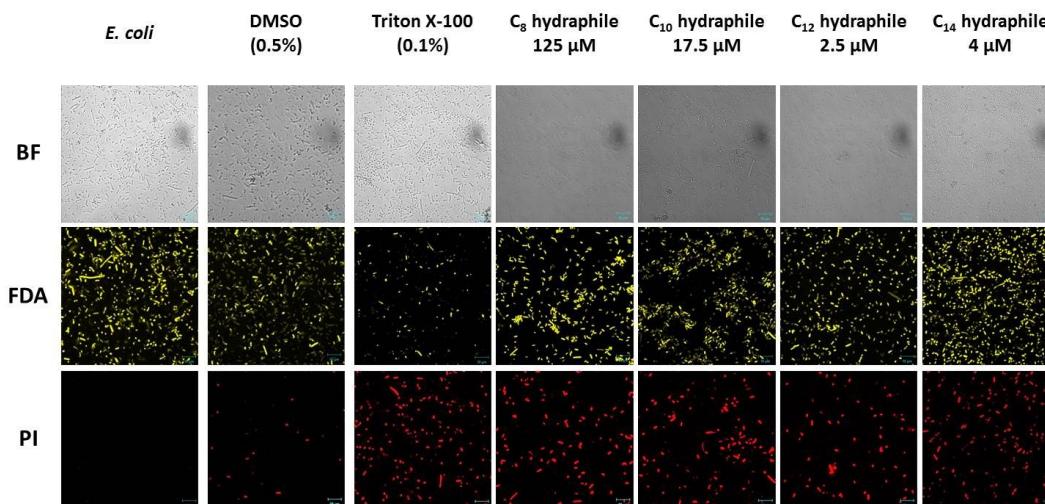


Figure S7a. Benzyl C₈-C₁₄ hydraphiles (¹/₂ [MIC]) mediated permeability of propidium iodide and fluorescein diacetate in tet^R *E. coli*. *E. coli* alone, DMSO (0.5% v/v) and triton X-100 (0.1% v/v) were used as controls.

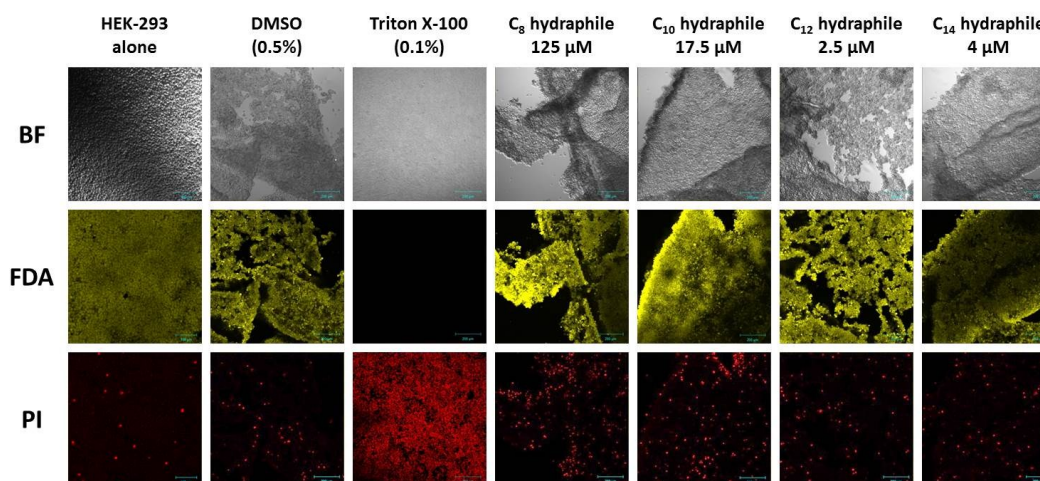


Figure S7b. Benzyl C₈-C₁₄ hydraphiles (¹/₂ [MIC]) mediated permeability of propidium iodide and fluorescein diacetate in HEK-293 cells. HEK-293 alone, DMSO (0.5%) and triton X-100 (0.1%) were used as controls.

Figure S8: Release of potassium ions from tet^R *E. coli* in the presence of benzyl C₈-C₁₄ hydraphiles and Gramicidin.

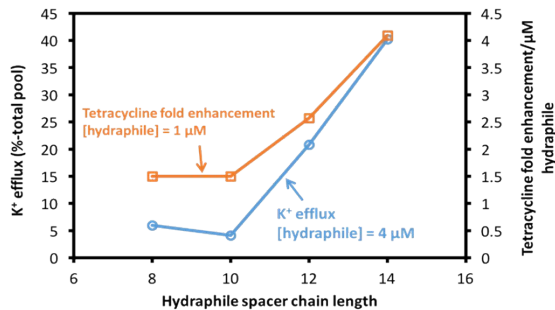


Figure S8(a). Comparison of benzyl C₈-C₁₄ hydraphiles to transport potassium ions (open circles) and recovery of tetracycline activity (open squares) against tet^R *E. coli*. Potassium release is expressed in percent of total potassium content of tet^R *E. coli* released (y-axis, left) and tetracycline recovery in fold enhancement (y-axis, right). The x-axis shows the number of carbon atoms in the spacer chains of hydraphiles.

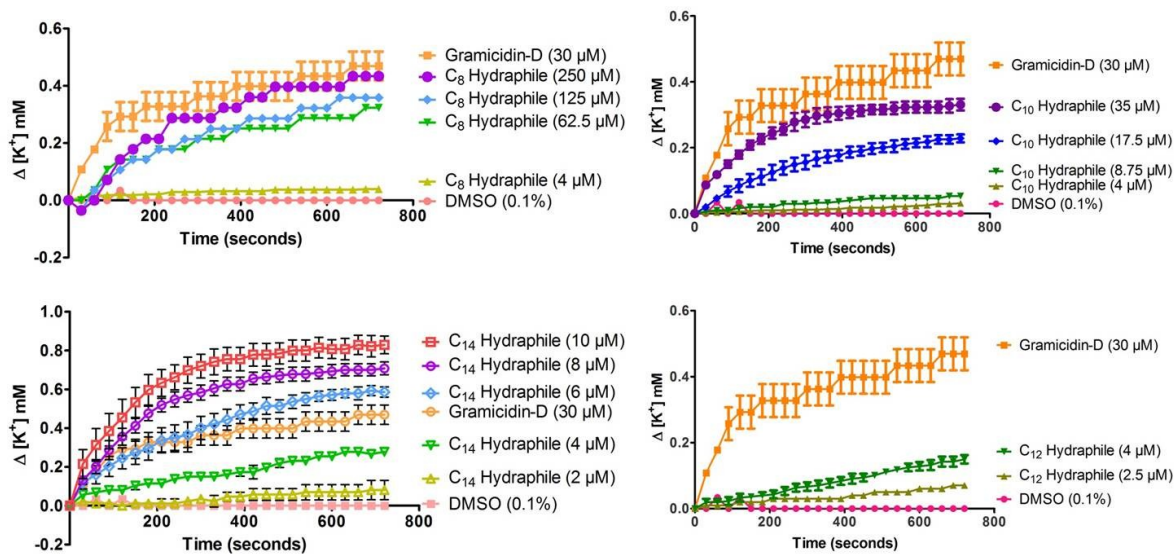


Figure S8 (b-e). Potassium release from *E. coli* over 720 seconds in the presence of various concentrations of benzyl C₈ hydraphile (a), benzyl C₁₀ hydraphile (b), benzyl C₁₄ hydraphile (c), and benzyl C₁₂ hydraphile (d). Error bars represent standard deviation in three trials.

Table S4. Combination studies with hydraphiles and ethidium bromide against *S. aureus* 1199B.

Table S4. Recovery of ethidium bromide activity against <i>S. aureus</i> 1199B			
Amphiphile used	[Amphiphile] μM	[EthBr] μM	Fold enhancement
No amphiphile	-	16	n/a
C ₈ hydraphile	32 ($\frac{1}{4}$ MIC)	16	1-fold
C ₈ hydraphile	64 ($\frac{1}{2}$ MIC)	8	2-fold
C ₁₀ hydraphile	4 ($\frac{1}{4}$ MIC)	16	1-fold
C ₁₀ hydraphile	8 ($\frac{1}{2}$ MIC)	1	16-fold
C ₁₂ hydraphile	0.25 ($\frac{1}{4}$ MIC)	16	1-fold
C ₁₂ hydraphile	0.5 ($\frac{1}{2}$ MIC)	2	8-fold
C ₁₄ hydraphile	0.25 ($\frac{1}{4}$ MIC)	4	4-fold
C ₁₄ hydraphile	0.5 ($\frac{1}{2}$ MIC)	0.5	32-fold

Figure S9: Survival of human hepatocytes in the presence of 1-100 μM benzyl C_{14} hydrophile and CCCP.

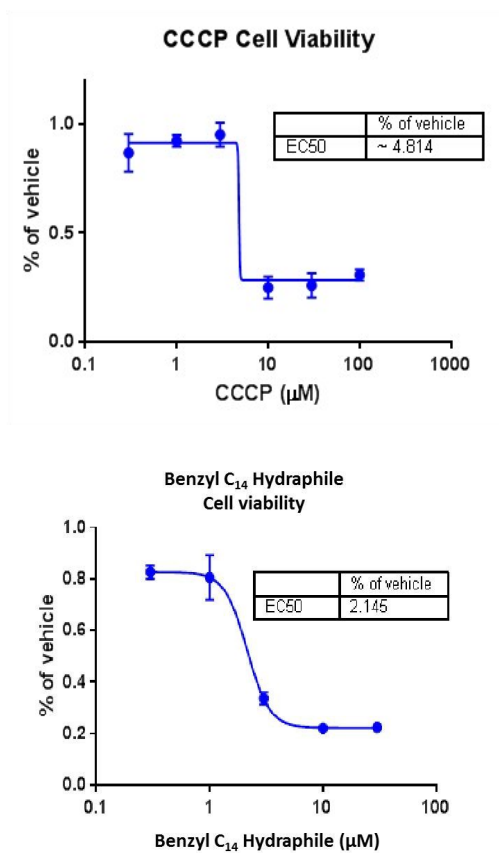


Figure S9. Cytotoxicity of benzyl C_{14} hydrophile and CCCP at 1-100 μM against human hepatocytes. The x-axis represents the concentration of the compound used. The y-axis represents the percent of cells survived as compared to the vehicle (DMSO) in the presence of the respective compounds used. The error bars represent standard deviation in three trials.