

A FRET based ratiometric fluorescent probe for detection of sulfite in food

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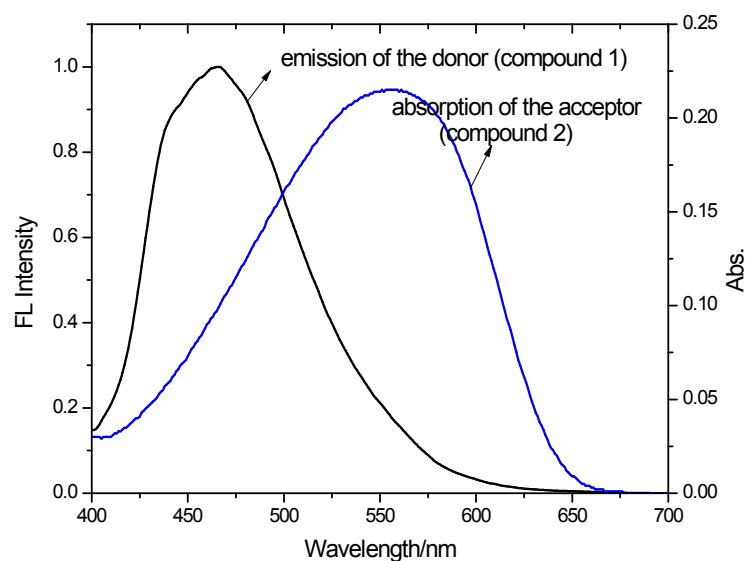


Fig. S1 Normalized emission spectra of donor (compound 1, black line) and normalized absorption spectra of acceptor (compound 2, blue line) Condition: donor, 5 μ M, acceptor, 5 μ M, Condition: DMF-PBS buffer solution (3/7, v/v, pH = 7.20). ($\lambda_{\text{ex}} = 380$ nm, slit = 10 nm/10 nm).

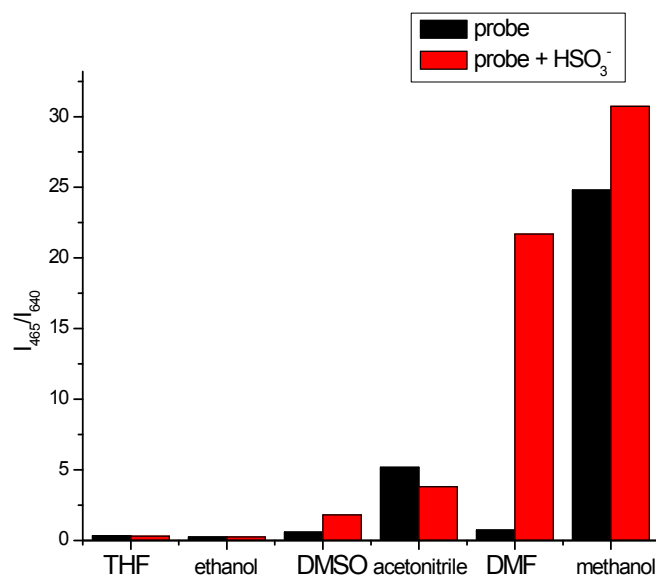


Fig. S2 The FL intensity ratio (I_{465}/I_{640}) of the probe with and without HSO₃⁻ in different solvent. The black line is the probe (5 μ M) and the red line is probe (5 μ M)

after addition of HSO_3^- (30 eq.) Condition: $\lambda_{\text{ex}} = 380 \text{ nm}$, slit = 10 nm/10 nm

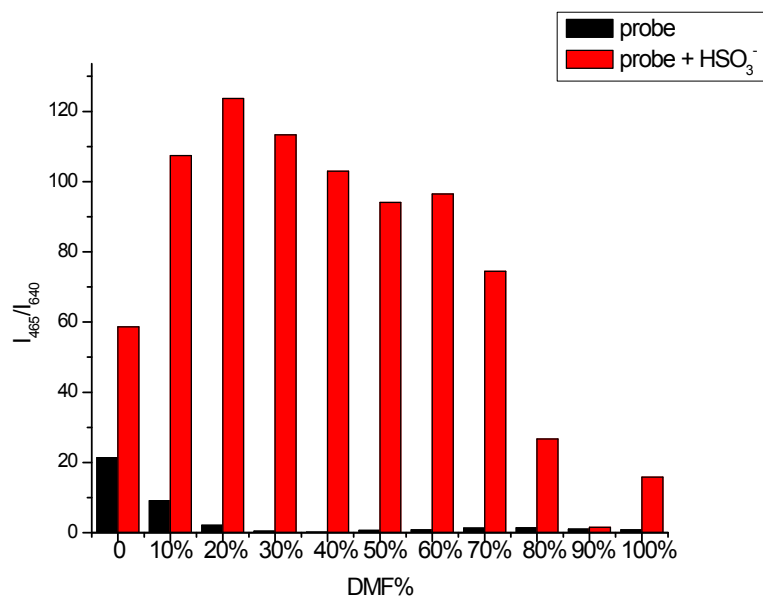


Fig. S3 The effect of water (PBS buffer solution, pH = 7.20) content on fluorescence spectra. The black line is the probe (5 μM) and the red line is probe (5 μM) after addition of HSO_3^- (30 eq.) Condition: $\lambda_{\text{ex}} = 380 \text{ nm}$, slit = 10 nm/10 nm

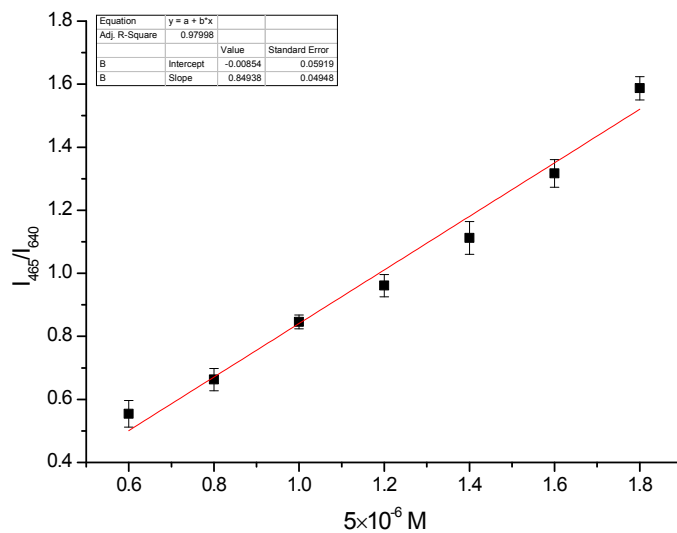


Fig. S4 Fluorescence intensity ratio changes (I_{640}/I_{465}) of probe (5 μM) upon gradual addition of HSO_3^- in DMF-PBS buffer solution (3/7, v/v, pH = 7.20) ($\lambda_{\text{ex}} = 380 \text{ nm}$, slit = 10 nm/10 nm).

The results were presented as means \pm SE with replicates $n = 5$.

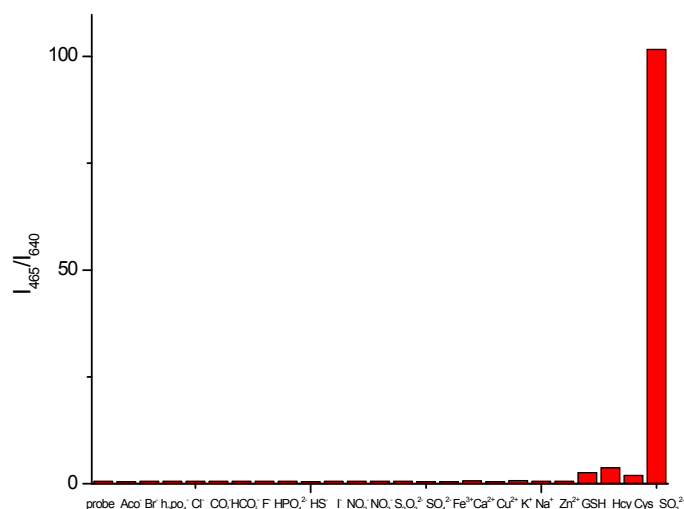


Fig. S5 Fluorescent intensity ratio changes at 465 nm and 640 nm (I_{465}/I_{640}) for each analyte (150 μ M, probe **PBI-S** (5 μ M), $\lambda_{ex} = 380$ nm, slit = 10 nm/10 nm).

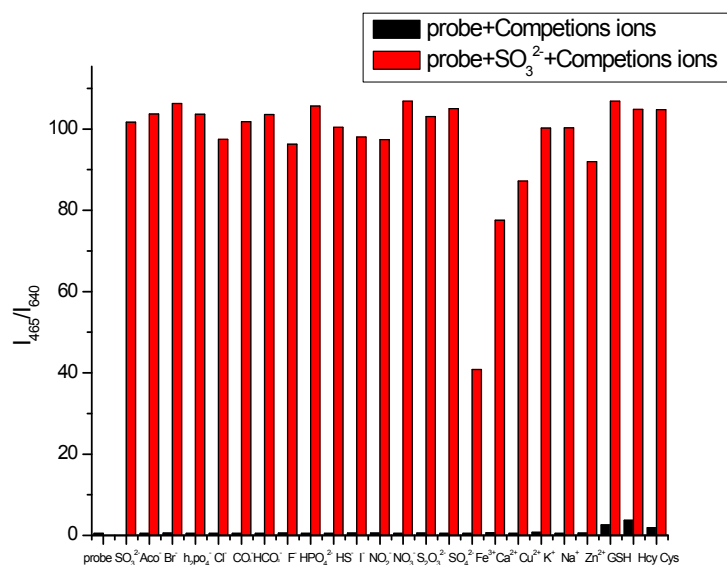


Fig. S6 The selectivity of probe **PBI-S** (5 μ M) in DMF-PBS buffer solution (3/7, v/v, pH = 7.20). The black bars represent the fluorescence emission ratio of **PBI-S** and 30 equiv. of different anions (150 μ M). The red bars show the ratio after the addition of 30 equiv. of HSO₃⁻ to the solution containing probe and different anions (150 μ M).

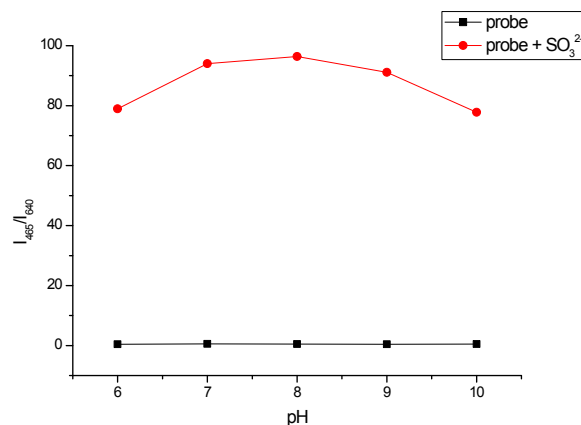


Fig. S7 The ratiometric fluorescence responses (I_{465}/I_{640}) of free probe ($5 \mu\text{M}$) and in the presence of 50 equiv of SO_3^{2-} in DMF-PBS buffer solution (3/7, v/v) solution with different pH conditions ($\lambda_{\text{ex}} = 380 \text{ nm}$, slit = 10 nm/10 nm).

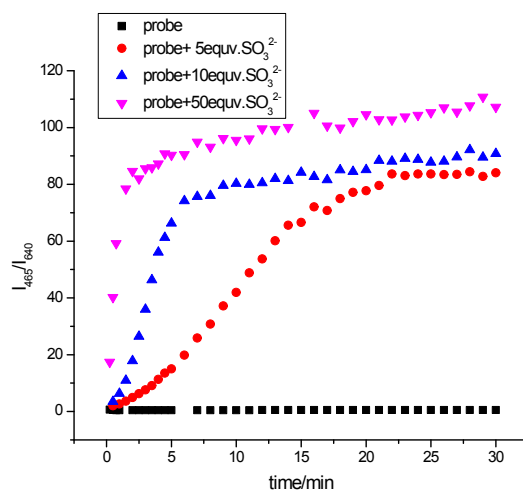


Fig. S8 Time dependent increase of probe ($5 \mu\text{M}$) fluorescence intensities after addition of various amounts of SO_3^{2-} in DMF-PBS buffer solution (3/7, v/v, pH = 7.20, $\lambda_{\text{ex}} = 380 \text{ nm}$, slit = 10 nm/10 nm).

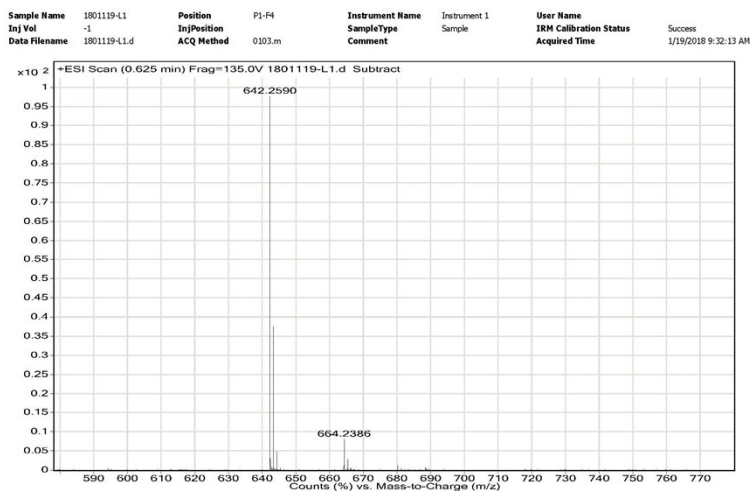


Fig. S9 HRMS of probe **PBI-S**.

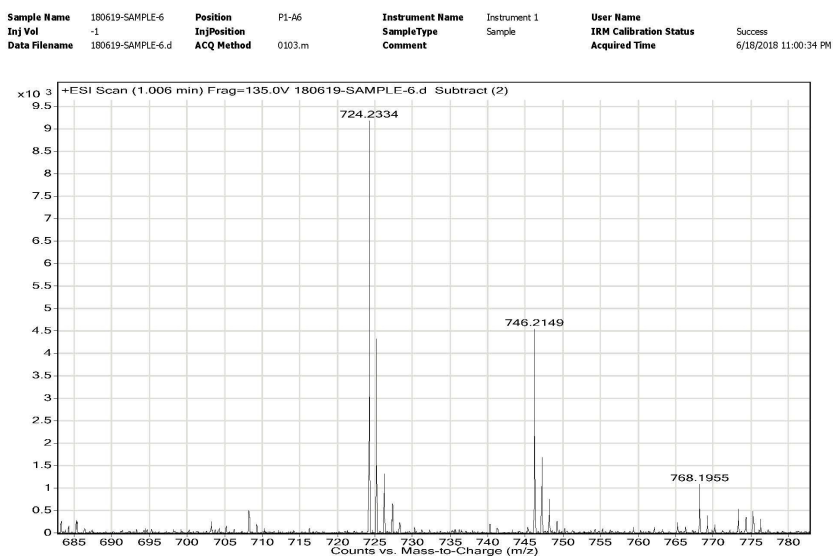


Fig. S10 HRMS of probe **PBI-S** after addition of NaHSO_3 .

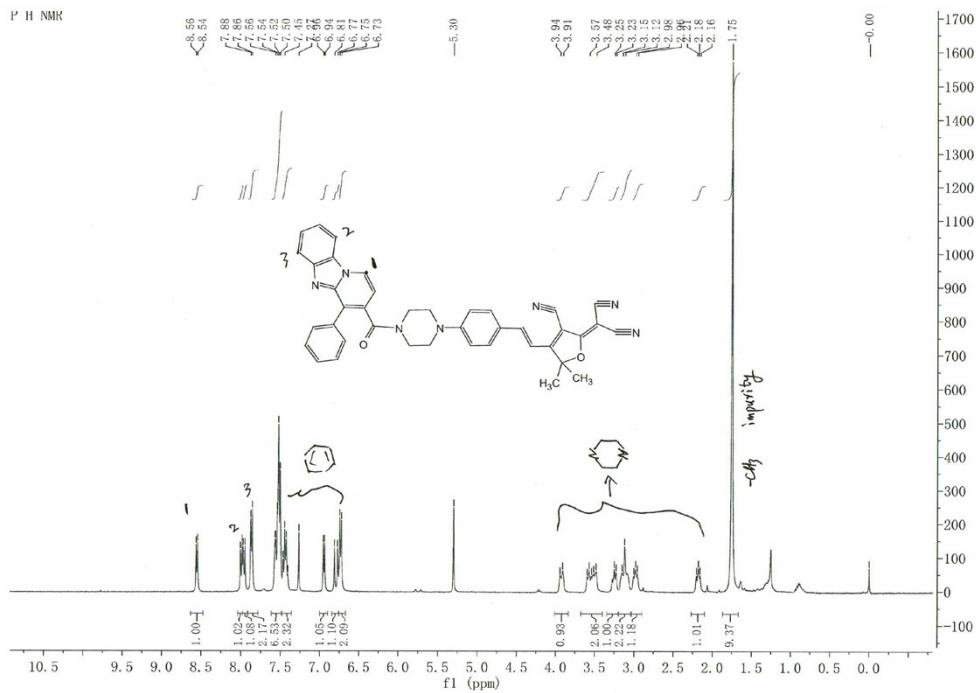


Fig. S11 ¹H NMR spectrum of probe **PBI-S**.

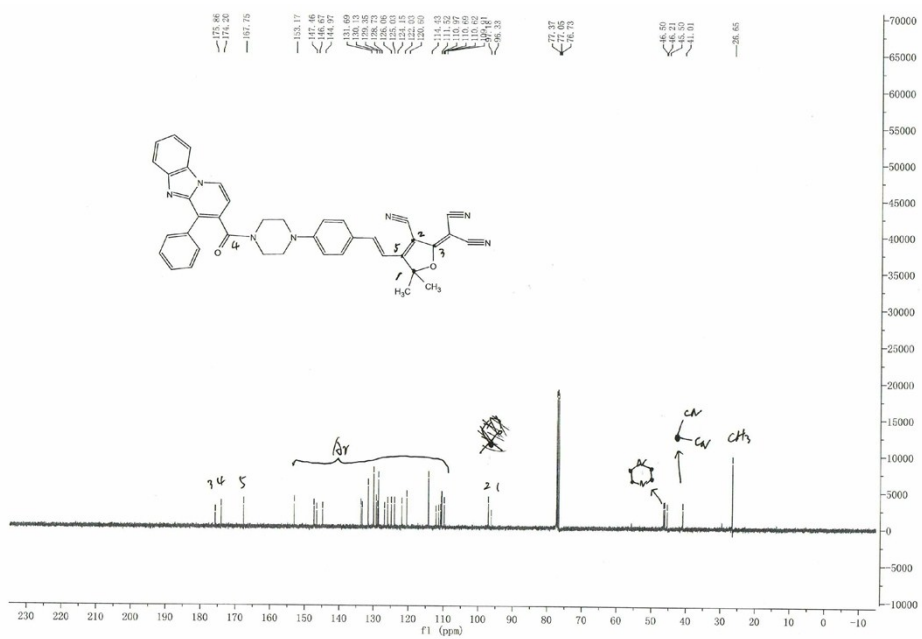
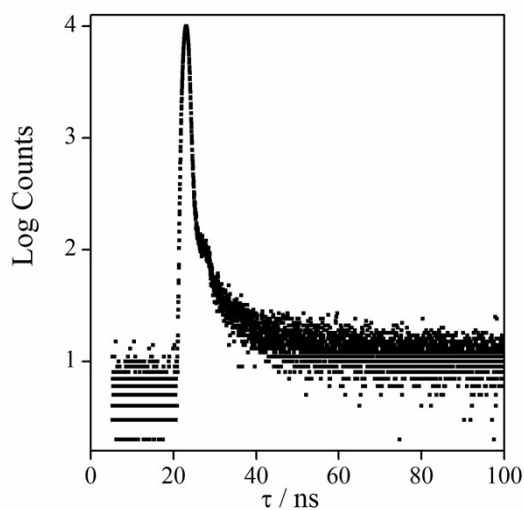


Fig. S12 ¹³C NMR spectrum of probe **PBI-S**.

Table S1. The detection of sugar samples by titration method

Sample	This method(mg/kg)	Titration method(mg/kg)
Crytal sugar	0.79	0.82
Granulated sugar	1.52	1.65
Soft sugar	0.96	1.02

Fluorescence quantum yields in C₂H₅OH were determined ($\phi = 0.41$) by a comparative method, using Rhodamine B (purchased from Sigma-Aldrich) as a standard sample with $\phi = 0.97$ as the reference.



The fluorescence lifetime of the probe

$\tau_1 = 0.34$ ns(68.8%), $\tau_2 = 3.58$ ns(31.2%). CHISQ= 1.20. The luminescence lifetime studies were conducted with a HORIBA Jobin Yvon Fluorolog-3 spectro-fluorometer fitted with a time-correlated single photon counting detector and a NanoLED pulsed laser diode excitation source (370 nm).