Supporting Information

Programmable One-pot Synthesis of Heparin Pentasaccharides Enabling Access to Regiodefined Sulfate Derivatives

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Table of Contents

A.	General procedures	S2-S4
B.	Experimental procedure and characterization data of monosaccharides	S4-S25
C.	Experimental procedure and characterization data of disaccharides	S25-S34
D.	Experimental procedure and characterization data of pentasaccharides	S34-S47
References		S47-S48
NMR spectra		S48-S174

A. General Procedures. All chemicals were purchased as reagent grade and used without further purification. The ACS grade solvents used for reactions were purchased from commercial source and further dried in accordance with standard procedures.¹ Molecular sieves (MS) AW-300 (for glycosylation) and MS 4 Å (for *O*-benzylation), were purchased from Aldrich, ground into powdered form and activated using standard procedure prior to use in the reaction. Reactions were performed under an inert atmosphere and strictly anhydrous conditions and monitored with analytical thin-layer chromatography (TLC) on silica gel 60 F254 plates and visualized under UV (254 nm) and/or by spraying with 20% anisaldehyde in ethanol or with a solution of (NH4)₆MorO24·4H2O 25 g/L. ¹H NMR spectra were recorded on a 600 MHz and 900 MHz NMR spectrometer. Chemical shifts (in ppm) were determined relative to tetramethylsilane in deuterated chloroform (δ 0 ppm). Coupling constant(s) in hertz (Hz) were measured from one-dimensional spectra. Mass spectra were obtained by the analytical services of the department.

General Procedure for Silyl Ether Cleavage. A pentasaccharide (0.025 mmol) was dissolved in pyridine (1.1 mL) followed by addition of HF·pyridine (30-40 equiv) at 0 °C. After stirring for 12 h, the solvent was evaporated in *vacuo* and the crude mixture was diluted with CH₂Cl₂ (50 mL) and washed with water (50 mL), saturated aqueous NaHCO₃(50 mL), and brine (50 mL). The organic phase was dried over MgSO₄, filtered, concentrated in *vacuo*. The residue was purified by silica gel column chromatography using a gradient of toluene and EtOAc to give the product with 6-OH group at the non-reducing end.

General Procedure for Lev Esters Cleavage. To a solution of pentasaccharide (0.03 mmol) in a mixture of THF and MeOH (1/1, v/v, 1.5 mL), hydrazine acetate (5 equiv per Lev group) was added at 0 °C. After stirring for 2 h, acetone (1 mL) was added and the reaction mixture was stirred for additional 30 min at the room temperature. The reaction mixture was diluted with EtOAc (30 mL) and washed with 25 mL of each of the water, saturated NaHCO₃ and then brine. The organic layer was dried over MgSO₄, filtered and concentrated in *vacuo*. The crude mixture was purified by silica gel column chromatography using a gradient of toluene and EtOAc to give the product.

General Procedure for Ag₂O-mediated *O***-benzylation:** To a solution of 6-hydroxyl-pentasaccharide (0.025 mmol) in a co-solvent (*n*-Hexane: CH₂Cl₂, 4/1, v/v, 2 mL) in a glass tube was added 4Å molecular sieves (200 mg). After stirring for 30 min under N₂-atmosphere, benzyl bromide (2.0 equiv per OH group) and, Ag₂O (3.0 equiv per OH group) were added, sequentially. The glass tube was sealed and the reaction mixture was heated to 70 °C in the dark for 12 h. After the complete conversion of the starting material, the mixture was filtered by a Celite pad and concentrated. The crude residue was purified by a silica gel column chromatography using a gradient of toluene and EtOAc to afford the corresponding *O*-benzyl pentasaccharide.

General Procedure for Saponification:

To a solution of the pentasaccharide (0.02 mmol) in THF (1 mL, for 20-30 mg), 30% solution of H₂O₂ (100 per ester) and 1 M LiOH (20 per ester) were added dropwise at -5 °C, sequentially. After stirring at room temperature for 8 h, a solution of NaOH (4N, 1.0 mL) was added until pH 14. The reaction mixture was stirred for 18 h at room temperature, then acidified by dropwise addition of AcOH to bring the pH to 8-8.5. It was diluted with CH₂Cl₂ (15 mL) and washed with 10 mL of each of the 10 % Na₂S₂O₃, H₂O and brine. The organic layer was dried over MgSO₄, filtered and concentrated in *vacuo* to give the product as a powder.

General Procedure for *O***-Sulfation.** Sulfur trioxide-pyridine complex (10 equiv per OH) was added to a solution of the pentasaccharide in DMF (1.0 mL for 0.02 mmol). The mixture was stirred at 55 °C for 12 h under N₂ atmosphere. The reaction flask was cooled down to room temperature, a solution of phosphate buffer (pH 7.5, 1 mL) was added and the mixture was kept stirring for an additional 1 h. The resulting reaction mixture was concentrated in *vacuo* and passed through a LH-20 gel using MeOH as solvent to remove the impurities. The resulting *O*-sultated pentasaccharide was used as such without further purification. **General Procedure for Hydrogenolysis.** Pd (OH) $_2$ /C (20%, 5 equi per OBn) was added to a solution of the starting material (0.02 mmol) in CH₃OH. The mixture was equipped with a H₂-ballon, and stirred for 36 h at room temperature, then filtered through a pad of Celite and concentrated in *vacuo* to give a white powder. The ¹H NMR of the crude mixture showed the absence of the signals of benzyl groups. The hydrogenolysis reaction cleaved all the *O*-Bn groups and reduced N₃ to NH₂. The crude powder was used as such for the next step without further purification.

General Procedure for Selective *N***-Sulfation**. The amino-alochol containing pentasaccharide (0.015 mmol) was dissolved in water (1-1.5 mL). The sulfur trioxide-pyridine complex (5 equi per NH₂ group) was added in five equal portions in half-hour intervals at room temperature. The pH value of the solution was adjusted to 9.5 by dropwise addition of 1N NaOH (aq). The pH was checked several times within 2-3 h and additional amount of 1N NaOH (aq) was added if pH drops below 9.5. After stirring for 38 h at room temperature, the reaction mixture was concentrated in *vacuo*. The residue was purified by column chromatography on Sephadex G-25 using water as an eluent. The desired fractions were collected, concentrated and passed through an ion exchange column of DOWEX 50WX8⁻Na⁺using water as eluent. The desired fractions were collected and lyophilized to give the *N*-sulfated pentasaccharide product.

B. Experimental procedure and characterization data of monosaccharides

Scheme S1. Synthesis of 2-azido thioglycoside donors



Reaction conditions:(a) p-CrSH, BF₃-Et₂O, CH₂Cl₂, 0 °C-rt, 12 h, 86%; (b) Zn-dust, AcOH : CH₂Cl₂ (1:2), rt, 12 h, 83% (c) TfN₃, EtOAc: MeOH (1:1), CuSO₄, K₂CO₃, rt, 12 h, 69%; (d) (i) PhH(OMe)₂, CSA, DMF, rt, 12 h, 81%; (ii) NaH, BnBr, DMF, 0 °C-rt, 2 h, 79%; (e) (i) BH₃-THF, ⁿBu₂BOTf, 0 °C, 2 h, 82%; (ii) TBDPSCl, Imidazole, rt, 12 h, 92%; (f) (i)TFA : CH₂Cl₂: H₂O (1:10:0.1), 0 °C-rt, 30 min, 81%; (ii) Ac₂O, Et₃N, CH₂Cl₂, 0 °C, 30 min, 88%

4-Methylphenyl 2-azido-2-deoxy-1-thio-β-D-glucopyranoside (S4)

Compound **S3** was prepared from D-glucosamine hydrochloride **16** using known procedures.² A solution of **S3** (10 g, 24.33 mmol) in MeOH: EtOAc (50:50 mL) was added CuSO4·5H₂O (0.6 g, 0.2433 mmol), and K₂CO₃ (3.35 g, 24.27 mmol). The mixture was cooled to 0 °C and treated with a freshly prepared TfN₃ solution. It was kept stirring at room temperature for overnight. After that the solvents were evaporated in *vacuo* and the crude mixture was purified by column chromatography on silica gel (PhCH₃: EtOAc 6:4) to furnish **S4** (5.25 g, 69%). R_f 0.25 (PhCH₃/EtOAc 6:4); ¹H NMR (600 MHz, CDCl₃): δ 7.42 (d, J = 7.8 Hz, 2 H), 7.12 (d, J = 7.8 Hz, 2 H), 4.45 (d, J = 10.2 Hz, 1 H), 3.89 (dd, J = 12.0, 3.0 Hz, 1 H), 3.82 (dd, J = 12.0, 4.2 Hz, 1 H), 3.54 (t, J = 9.6 Hz, 1 H), 3.48 (t, J = 9.0 Hz, 1 H), 3.2-3.30 (m, 1 H), 3.26 (t, J = 9.6 Hz, 1 H), 2.33 (s, 3 H); ¹³C NMR (150 MHz, CDCl₃): δ 138.9, 133.7, 130.0, 127.6, 86.8, 79.2, 69.6, 65.1, 62.1, 57.0, 21.2; m/z (HRMS) calcd for C₁₃H₁₇N₃O₄SNa [M+Na]⁺: 334.0837, found:.334.0844.

4-Methylphenyl 2-azido-3-benzyl-4,6-*O*-benzylidene-2-deoxy-1-thio-β-D-glucopyranoside (S5)



Compound **S4** (5.25 g, 16.88 mmol) in anhydrous DMF (40 mL) was treated with benzaldehyde dimethyl acetal (3.1 mL, 20.3 mmol) and camphor sulphonic acid (3.92 g, 16.88 mmol). The reaction mixture was stirred for 12 h at room temperature and then quenched with Et₃N. It was concentrated and diluted with EtOAc (2 × 100 mL), washed with water (2 ×50 mL), brine (50 mL), dried over MgSO₄ and filtered. The organic layer was concentrated in *vacuo* and the resulting residue was purified by silica gel column chromatography (Hexane: EtOAc, 1:1) to furnish **4-Methylphenyl-2-azido-4,6-***O***-benzylidene-2-deoxy-1-thio-β-D-glucopyranoside** (4.71 g, 81%). *R*_f 0.41 (Hexane/EtOAc 9:1); ¹H NMR (600 MHz, CDCl₃): δ 8.11-8.09 (m, 1 H), 7.62-7.60 (m, 1 H), 7.49-7.44 (m, 4 H), 7.38-7.35 (m, 2 H), 7.16 (d, *J* = 8.4 Hz, 2 H), 5.52 (s, 1 H), 4.49 (d, *J* = 10.2 Hz, 1 H), 4.36 (dd, *J* = 4.8, 10.8 Hz, 1 H), 3.78-3.75 (m, 2 H), 3.46-3.44 (m, 2 H), 3.32 (dd, *J* = 9.0, 10.2 Hz, 1 H), 2.37 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 139.4, 134.3, 133.7, 130.1, 129.9, 129.4-126.7, 101.9, 86.8, 80.2, 74.1, 70.2, 68.4, 65.0, 21.2; m/z (HRMS) calcd for C₂₀H₂₁N₃O₄S [M+H]⁺: 400.1326, found: 401.2591. The purified compound (4.71 g,

11.8 mmol) was dissolved in anhydrous DMF (40 mL) and NaH (0.944 g, 23.60 mmol of 60% dispersion in mineral oil) and benzyl bromide (1.69 mL, 14 mmol) were added at 0 °C. After 2 h, it was quenched with methanol and concentrated in *vacuo*, diluted with EtOAc (2 × 100 mL), washed with water (2 ×75 mL), brine (50 mL), then dried (MgSO4), filtered and concentrated in *vacuo*. The resulting residue was purified by flash column chromatography (Hexane: EtOAc, 8:2) to furnish **S5** (4.56 g, 79%) as yellow oil. *R*f 0.51 (Hexane/EtOAc 9:1); ¹H NMR (600 MHz, CDCl₃): δ 7.39-7.36 (m, 4 H), 7.29-7.19 (m, 8 H), 7.01 (d, *J* = 7.8 Hz, 2 H), 5.46 (s, 1 H), 4.81 (d, *J* = 10.8 Hz, 1 H), 4.69 (d, *J* = 11.4 Hz, 1 H), 4.32 (d, *J* = 10.2 Hz, 1 H), 4.28 (dd, *J* = 10.2, 4.8 Hz, 1 H), 3.67 (t, *J* = 10.2 Hz, 1 H), 3.56 (t, *J* = 9.0 Hz, 1 H), 3.51 (t, *J* = 9.0 Hz, 1 H), 3.35 (td, *J* = 4.8, 9.6 Hz, 1 H), 3.24 (dd, *J* = 9.0, 10.2 Hz, 1 H), 2.26 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 139.1, 137.5, 137.1, 134.5,129.9-128.0, 126.5, 125.9, 101.2, 86.5, 81.2, 81.0, 75.2, 70.4, 68.5, 64.5, 21.2; m/z (HRMS) calcd for C₂₇H₂₇N₃O₄SNa [M+Na]⁺: 512.1614, found:.512.1634.

4-Methylphenyl 6-O-acetyl-2-azido-3-O-benzyl-2-deoxy-1-thio-β-D-glucopyranoside (9)



A solution of a compound **S5** (4.56 g, 9.32 mmol) in a mixture of solvents (CH₂Cl₂: TFA: H₂O =10/1/0.1 v/v/v) was stirred at room temperature for 30 min. It was then concentrated in *vacuo*, and later co-evaporated with toluene to remove traces of water. The crude diol was further dissolved in anhydrous CH₂Cl₂(25 mL) and treated with Ac₂O (0.96 mL, 9.41 mmol) and Et₃N (11.68 mL, 115.68 mmol) at 0 °C under argon atmosphere for 30 min. After the complete conversion of starting material, it was diluted with CH₂Cl₂(100 mL), washed with saturated NaHCO₃(2 × 50 mL), brine (75 mL), then dried (MgSO₄), filtered and concentrated in *vacuo*. The resulting residue was purified by silica gel coulmn chromatography (Hexane: EtOAc, 6:4) to afford **9** as gum (3.63 g, 88%). *R*_f 0.38 (hexane/EtOAc, 6:4); ¹H NMR (600 MHz, CDCl₃): δ 7.49-7.47 (m, 2 H), 7.37-7.34 (m, 4 H), 7.32-7.30 (m, 1 H), 7.12 (d, *J* = 7.8 Hz, 2 H), 4.90 (d, *J* = 11.4 Hz, 1 H), 4.80 (d, *J* = 11.1 Hz, 1 H), 4.42 (dd, *J* = 12.0, 4.8 Hz, 1 H), 4.36 (d, *J* = 10.2 Hz, 1 H), 4.30 (dd, *J* = 12.0, 2.0 Hz, 1 H), 3.40-3.39 (m, 1 H), 3.36 (app dd, *J* = 10.2, 11.6 Hz, 2 H), 3.25 (t, *J* = 9.6 Hz, 1 H), 2.34 (s, 3H), 2.10 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 171.7, 138.9, 137.7, 134.4, 129.8-127.0, 86.2, 84.3, 77.8, 75.7, 69.8, 64.5, 63.1, 21.2, 20.9; m/z (HRMS) calcd for C₂₂H₂₅N₃O₅SNa [M+Na]⁺: 466.1407, found: 466.1431.

4-Methylphenyl glucopyranoside (4).

2-azido-3,4-di-O-benzyl-6-O-tert-butyldiphenylsilyl-2-deoxy-1-thio-β-D-



To a solution of compound S5 (2.0 gm, 4.08 mmol) in anhydrous THF (25 mL) was added BH₃·THF complex (15 mL of a 1M solution in THF) followed by Bu₂BOTf (0.500 g of a 1M solution in CH₂Cl₂) at 0 °C. After 2 h, the reaction mixture was quenched by a slow addition of MeOH at 0 °C. When no further evolution of hydrogen was found, it was concentrated in *vacuo* and residue was purified by flash column chromatography (hexane/EtOAc, 9:1) to afford the 6-hydroxy derivative (1.64 g, 82%) as a colourless gum. The 6-hydroxy derivative (3.36 mmol) was dissolved in anhydrous CH₂Cl₂(20 mL) and tert-butyl diphenyl silyl chloride (1.08 mL, 4.00 mmol) and imidazole (0.332 g, 4.88 mmol) were added under N₂ atmosphere. It was stirred over night at room temperature and quenched with MeOH. The solvent was evaporated in *vacuo* and the crude mixture was purified by silica gel column chromatography (Hexane: EtOAc, 20:1) to afford **4** as a gum (2.24 g, 92%). *R*_f0.58 (hexane/EtOAc, 20:1); ¹H NMR (600 MHz, CDCl₃): δ 7.80 (d, J = 7.2 Hz, 2H), 7.70 (d, J = 7.2 Hz, 2H), 7.54 (d, J = 7.8 Hz, 2H), 7.44-7.26 (m, 14 H), 7.16 (app d, 2H), 7.05 (d, J = 7.8 Hz, 2 H), 4.86 (app t, J = 6.0 Hz, 3H), 4.70 (d, J = 10.8 Hz, 1H), 4.38 (d, J = 10.2 Hz, 1H), 4.02 (d, J = 11.4 Hz, 1H), 3.94 (dd, J = 11.4, 3.0 Hz, 1 H), 3.78 (t, J = 9.6 Hz, 1 H), 3.54 (t, J = 9.6 Hz, 1H), 3.36 (app t, J = 9.6 Hz, 2H), 2.32 (s, 3H), 1.10 (s, 9H); 13 C NMR (150 MHz, CDCl₃): δ 138.0, 137.6, 135.9, 134.2, 133.4, 132.8, 129.8-127.7, 127.2, 86.1, 85.3, 80.0, 76.1, 75.1, 64.8, 62.3, 26.9, 21.2, 19.3; m/z (HRMS) calcd for C43H47N3O4SSiNa [M+Na]+: 752.2949, found:.752.2984.

Scheme S2. Synthesis of thioglycoside donors (Building Block B)





Reaction conditions: (a) (i) BF₃·OEt₂, CH₂Cl₂, 0 °C-rt, 12 h, 94.5%; (ii) NaOMe, MeOH : CH₂Cl₂(3:1), rt, 2 h; (b) PhH(OMe)₂,CSA DMF, rt, 12 h, 86%; (c) PhOMe(OMe)₂, CSA, DMF, rt, 12 h, 81%; (d) (i) Bu₂SnO, PhCH₃, 110 °C, 12 h; (ii) CSF, BnBr, DMF, 80 °C, 12 h, 82% (for **S7b**), 80 % (for **S7a**); (e) BzCl, Py, 0 °C-rt, 12 h, 80% (for **10a**), 81 % (for **S8**); (f) CH₃CO(CH₂)₂COOH, EDCI, DMAP, CH₂Cl₂, rt, 12 h, 81% (for **10b**), 83 % (for **17**), 81% (for **18**); (g) CH₂Cl₂: CF₃COOH : H₂O (10:1:0.1), rt, 1 h, 78%. (h) TBDPSCl, Py, rt, 12 h, 89%; (i) NaCNBH₃, TFA, DMF, 0 °C-rt, 12 h, 76 %.

4-Methylphenyl-4,6-*O*-benzylidine-1-thio-β-D-glucopyranoside (10)



To a solution of commercially available *per-O-acetyl*- β -D-glucopyranose **14** (20 g, 51.28 mmol) in CH₂Cl₂ (100 mL) was treated with *p*-toluenethiol (7.5 g, 60.38 mmol) followed by BF₃·OEt₂ (16.5 mL, 128 mmol) at 0 °C. After stirring for 12 h, the reaction mixture was washed with saturated NaHCO₃ (3 x 100 mL). The aqueous layer was further extracted with CH₂Cl₂ (3 x 150 mL). The combined organic layers were washed with a brine solution (200 mL), dried over MgSO₄, filtered and concentrated in *vacuo*. The residue was re-dissolved in a minimum amount of EtOAc and hexane was added until it solidified to give the desired derivative (22 g, 94.5%). The analytical data is well in agreement with the reported values. A solution of tri-acetate (10 g, 8.05 mmol) was dissolved in a mixture of MeOH: CH₂Cl₂(60:20 mL) and a catalytic amount of NaOMe (0.650 g) was added. The mixture was stirred for 2 h at room temperature , and after that it was neutralized with amberlite 120 H⁺ resin. The resin was filtered and the solvent was evaporated to furnish the tri-hydroxyl derivative **S6**. Compound **S6** (5.98 g, 20.90 mmol) in anhydrous DMF (40 mL) was treated with benzaldehyde dimethyl acetal (3.81 mL, 25.1 mmol) and camphor

sulphonic acid (4.85 g, 20.90 mmol). The reaction mixture was stirred for 12 h at room temperature and then quenched with Et₃N. It was concentrated and diluted with EtOAc (2 × 100 mL), washed with water (2 × 50 mL), brine (50 mL), then dried (MgSO₄), filtered and concentrated in *vacuo*. The resulting residue was purified by silica gel column chromatography (Hexane: EtOAc, 1:1) to furnish compound **10** (6.72 g, 86%). *R*_f 0.52 (hexane/EtOAc 1:1); ¹H NMR (600 MHz, CDCl₃): δ 7.48-7.44 (m, 2H), 7.42-7.36 (m, 5H), 7.35-7.14 (m, 2H), 5.52 (s, 1H), 4.55 (d, *J* = 9.6 Hz, 1H), 4.36 (dd, *J* = 10.8, 4.8 Hz, 1H), 3.83 (t, *J* = 8.4 Hz, 1H), 3.78 (td, *J* = 3.0, 7.2 Hz, 1H), 3.51-3.49 (m, 1H), 3.48 (br s, 1H), 3.42 (t, *J* = 9.0 Hz, 1H), 2.89 (s, 1H), 2.61 (s, 1H), 2.36 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 138.9, 136.9, 133.7, 129.8-127.2, 126.3, 102.0, 88.7, 80.3, 74.6, 72.5, 70.6, 68.6, 21.2; m/z (HRMS) calcd for C₂₀H₂₂O₅SNa [M+Na]⁺: 397.1080, found: 397.1097.

4-Methylphenyl-2-*O*-benzoyl-3-*O*-benzyl-4,6-*O*-benzylidine-1-thio-β-D-glucopyranoside (10a)



A mixture of compound 10 (6.72 g, 17.96 mmol) and dibutyltin oxide (6.71 g, 26.95 mmol) was stirred in toluene (60 mL) at 110 °C by using a dean-stark apparatus for 12 h. The reaction mixture was cooled down to room temperature and concentrated in vacuo. The residue was dissolved in anhydrous DMF (40 mL) and BnBr was added (2.79 mL, 23.34 mmol) followed by CsF (3.27 g, 21.3 mmol). The reaction mixture was heated at 80 °C for overnight. Solvents were removed in vacuo. It was diluted with EtOAc $(2 \times 150 \text{ mL})$, washed with water $(2 \times 70 \text{ mL})$, brine (80 mL), then dried (MgSO₄), filtered and concentrated in vacuo. The resulting residue was purified by silica gel column chromatography (Hexane: EtOAc, 8:2) to furnish the compound S7a (6.83 g, 82%) as an yellow oil. $R_f 0.52$ (hexane/EtOAc 8:2); ¹H NMR (600 MHz, CDCl₃): δ 7.48-7.44 (m, 2H), 7.43-7.26 (m, 10H), 7.12 (d, *J* = 7.8 Hz, 2H), 5.56 (s, 1H), 4.93 (d, J = 11.4 Hz, 1H), 4.80 (d, J = 11.4 Hz, 1H), 4.55 (d, J = 12.0 Hz, 1H), 3.86 (dd, J = 4.8, 10.2 Hz, 1H), 3.80 (t, J = 10.2 Hz, 1H), 3.70 (t, J = 9.0 Hz, 1H), 3.64 (t, J = 9.0 Hz, 1H), 3.51-3.48 (m, 2H), 2.34 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δδ138.9, 136.9, 133.8, 130.0, 129.4, 128.5, 127.3, 126.4, 102.0, 88.8, 80.3, 74.6, 72.6, 70.6, 68.7, 21.3; m/z (HRMS) calcd for C₂₇H₂₈O₅SNa [M+Na]⁺: 487.1555, found:.487.1578. A solution of copound S7a (6.83 g, 14.71 mmol) in pyridine (40 mL) was treated with BzCl (3.41 mL, 24.33 mmol) at 0 °C. The reaction mixture was allowed to warm up slowly to room temperature over a period of 12 h. It was diluted with EtOAc (2×100 mL), washed with water (2×50 mL), brine (80 mL), then dried (MgSO₄), filtered and concentrated in vacuo. The resulting residue was purified by silica gel column chromatography (Hexane: EtOAc, 8:2) to furnish the compound 10a (6.67

g, 80%). *R*f 0.57 (hexane/EtOAc 8:2); ¹H NMR (600 MHz, CDCl₃): δ 8.17 (app d, *J* = 2.0, 8.4 Hz, 2 H), 8.02 (d, *J* = 7.2 Hz, 2H), 7.68-7.59 (m, 2H), 7.53-7.50 (m, 2H), 7.40-7.38 (m, 3H), 7.32 (d, *J* = 7.8 Hz, 2H), 7.11-7.04 (m, 6H), 5.59 (s, 1 H), 5.25 (dd, *J* = 10.2, 9.0 Hz, 1H), 4.78 (d, *J* = 12.0 Hz, 2H), 4.65 (d, *J* = 12.0 Hz, 1H), 4.40 (dd, *J* = 8.4, 10.8 Hz, 1H), 3.89-3.85 (m, 1H), 3.80 (dd, *J* = 4.8, 9.6 Hz, 2H), 3.54 (td, *J* = 5.4, 10.2 Hz, 1H), 2.32 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 180.0, 137.7, 133.6, 133.1, 130.0, 129.8-128.0, 127.6, 126.0, 101.2, 87.1, 81.4, 79.3, 74.2, 72.0, 70.5, 68.6, 21.3;m/z (HRMS) calcd for C₃₄H₃₂O₆SNa [M+Na]⁺: 591.1812, found:591.1827.

4-Methylphenyl-2-*O*-benzoyl-3-*O*-benzyl-1-thio-β-D-glucopyranoside (S9)

A solution of a compound **10a** (4.56 g, 9.32 mmol) in a mixture of solvents (CH₂Cl₂: TFA: H₂O = 10:1:0.1, v/v/v) was stirred at room temperature for 1 h. The reaction mixture was quenched with solid NaHCO₃, filtered and then concentrated in *vacuo*. The crude reaction mixture was purified by silica gel column chromatography (CH₂Cl₂: Hexane, 3:7) to afford **S9** (3.0 g, 78%) as white solid. *R*_f0.51 (CH₂Cl₂: Hexane, 3:7); ¹H NMR (600 MHz, CDCl₃): δ 8.11-8.00 (m, 3H), 7.62-7.59 (m, 2H), 7.49-7.40 (m, 3H), 7.33 (d, *J* = 8.4 Hz, 2H), 7.19-7.17 (m, 2H), 7.08 (d, *J* = 7.8 Hz, 2H), 5.23 (t, *J* = 9.6 Hz, 1H), 4.78 (t, *J* = 10.2 Hz, 1H), 4.71 (d, *J* = 11.4 Hz, 1H), 4.62 (d, *J* = 11.4 Hz, 1H), 3.92 (dd, *J* = 12.0, 3.0 Hz, 1H), 3.82 (dd, *J* = 4.8, 12.0 Hz, 1H), 3.72 (q, *J* = 7.8 Hz, 2H), 3.47 (q, *J* = 4.2 Hz, 1 H), 2.31 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 165.1, 138.5, 137.7, 133.5, 133.4, 133.2, 130.3, 130.0, 129.9, 129.8, 128.8, 128.6, 128.5, 128.1, 128.0, 86.7, 83.93, 79.5, 74.8, 72.5, 70.4, 62.6, 21.2; m/z (HRMS) calcd for C₂₇H₂₈O₆SNa [M+Na]⁺: 503.1504, found:503.1521.

$\label{eq:2-0-benzoyl-3-0-benzyl-6-0-tert-butyldiphenylsilyl-1-thio-\beta-D-glucopyranoside} (S11)$



Compound **S9** (3.0 g, 6.25 mmol) was dissolved in anhydrous pyridine (20 mL) and *tert*-butyl diphenyl silyl chloride (2.1 mL, 7.4 mmol) was added under N₂ atmosphere. It was stirred overnight at room temperature and quenched with MeOH. The solvents were evaporated in *vacuo* and crude mixture was purified by silica gel column chromatography (Hexane: EtOAc, 9:1) to afford **S11** as colourless gum (3.99 g, 89%). $R_{\rm f}$ 0.65 (hexane/EtOAc 9:1); ¹H NMR (600 MHz, CDCl₃): δ 8.21 – 8.13 (m, 1H), 8.13 –

8.08 (m, 1H), 7.79-7.77 (m, 4H), 7.66 – 7.59 (m, 2H), 7.52 – 7.44 (m, 12H), 7.44 – 7.37 (m, 2H), 7.02 (d, J = 8.4 Hz, 2H), 5.30 – 5.25 (m, 1H), 4.80 (d, J = 10.0 Hz, 1H), 4.76 – 4.70 (m, 2H), 4.03 (dd, J = 11.0, 3.7 Hz, 1H), 3.99 (dd, J = 11.0, 4.6 Hz, 1H), 3.93 – 3.88 (m, 1H), 3.77 (t, J = 9.0 Hz, 1H), 3.57 – 3.53 (m, 1H), 2.31 (s, 3H), 1.11 (s, 9H);¹³C NMR (150 MHz, CDCl₃): δ 165.2, 138.0, 137.9, 135.7, 133.2, 133.1, 132.9, 130.0, 129.8-127.8, 86.6, 84.0, 79.4, 74.7, 72.2, 71.3, 64.1, 31.6, 26.9, 22.7, 21.1, 19.3; m/z (HRMS) calcd for C₄₃H₄₆O₆SsiNa [M+Na]⁺: 741.2677, found:.741.2701.

4-Methylphenyl-2-*O*-benzoyl-3-*O*-benzyl-6-*O*-tert-butyldiphenylsilyl-4-*O*-levulinyl-1-thio-β-D-glucopyranoside (17).



To a solution of **S11** (3.99 g, 5.55 mmol) in CH₂Cl₂(35 mL) was added levulinic acid (0.892 mL, 11.02 mmol), EDCI (2.15 g, 13.89 mmol), DMAP (0.340 g, 2.77 mmol) and the mixture was stirred for overnight at room temperature. It was diluted with EtOAc (2×50 mL), washed with water (2×25 mL), brine (40 mL), then dried (MgSO₄), filtered and concentrated in *vacuo*. The resulting residue was purified by silica gel column chromatography (Hexane: EtOAc, 8:2) to furnish compound **17** (3.76 g, 83%). *R*_f 0.48 (hexane/EtOAc 8:2); ¹H NMR (600 MHz, CDCl₃): δ 8.08-8.06 (m, 2H), 7.77-7.72 (m, 4H), 7.62-7.60 (m, 2H), 7.49-7.45 (m, 2H), 7.47-7.37 (m, 8H), 7.16-7.11 (m, 4H), 7.01 (d, *J* = 7.8 Hz, 2H), 5.31 (t, *J* = 9.6 Hz, 1H), 5.20 (t, *J* = 9.6 Hz, 1H), 4.81 (d, *J* = 9.6 Hz, 1H), 4.53 (s, 2 H), 3.92 (t, *J* = 9.6 Hz, 1H), 3.82-3.76 (m, 2H), 3.64-3.61 (m, 1H), 2.56 (t, *J* = 7.2 Hz, 2H), 2.38-2.33 (m, 2H), 2.29 (s, 3H), 2.12 (s, 3H), 1.10 (s, 9H); ¹³C NMR (150 MHz, CDCl₃): δ 206.0, 171.2, 165.0, 138.0, 137.7, 135.8, 135.8, 135.6, 134.8, 133.3, 133.3, 133.0, 129.9, 129.7, 129.6, 129.2, 128.5, 128.2, 127.9, 127.8, 127.7, 127.7, 127.7, 127.6, 86.8, 81.7, 79.4, 74.1, 72.3, 70.0, 63.0, 37.8, 29.8, 27.8, 26.8, 21.2, 19.3; m/z (HRMS) calcd for C4₈H₅₂O₈SSiNa [M+Na]⁺: 839.3044, found: 839.3088.

4-Methylphenyl-4,6-*O*-*p*-methoxybenzylidine-1-thio-β-D-glucopyranoside (11).



Compound **S6** (6 g, 20.90 mmol) was treated with *p*-anisaldehyde dimethyl acetal (4.28 mL, 25.17 mmol) and camphor sulphonic acid (4.87 g, 20.90 mmol) in anhydrous DMF (40 mL). The reaction mixture was stirred for 12 h at room temperature and then quenched with Et₃N. The solvent was removed in *vacuo*. And it was diluted with EtOAc (2×100 mL), washed with water (2×50 mL), brine (50 mL), then dried

(MgSO₄), filtered and concentrated in *vacuo*. The resulting residue was purified by silica gel column chromatography (Hexane: EtOAc, 1:1) to furnish compound **11** (6.86 g, 81%). *R*_f 0.39 (hexane/EtOAc 1:1); ¹H NMR (600 MHz, CDCl₃): δ 7.41 (d, *J* = 7.8 Hz, 2H), 7.38 (d, *J* = 12.0 Hz, 2 H), 7.12 (d, *J* = 12.0 Hz, 2H), 6.86 (d, *J* = 9.0 Hz, 2H), 5.43 (s, 1H), 4.51 (d, *J* = 9.6 Hz, 1 H), 4.32 (dd, *J* = 10.8, 4.8 Hz, 1H), 3.82-3.77 (m, 1H), 3.76 (s, 3H), 3.74 (d, *J* = 8.4 Hz, 1H), 3.70 (t, *J* = 9.6 Hz, 1H), 3.44-3.41 (m, 2H), 3.39-3.36 (m, 1H), 3.16 (app d, *J* = 2.0 Hz, 1H), 2.36 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 160.2, 138.7, 133.5, 129.8-127.8, 113.7, 101.8, 88.6, 80.1, 74.5, 72.5, 70.4, 68.5, 55.3, 21.2;m/z (HRMS) calcd for C₂₁H₂₄O₆SNa [M+Na]⁺: 427.1186, found:.427.1209.

$\label{eq:2.2} 4-Methylphenyl-2-{\it O}-benzoyl-3-{\it O}-benzyl-4, 6-{\it p}-methoxybenzylidine-1-thio-\beta-D-glucopyranoside (S8).$



A mixture of compound 11 (6.86 g, 16.98 mmol) and dibutyltin oxide (5.07 g, 20.37 mmol) was stirred in toluene (60 mL) at 110 °C by using dean-stark apparatus for 12 h. The reaction mixture was cooled down to room temperature and solvent was removed in vacuo. The residue was dissolved in anhydrous DMF (40 mL) and BnBr was added (2.43 mL, 20.37 mmol) followed by CsF (3.10 g, 20.37 mmol). The reaction mixture was heated at 80 °C and stirred continuously for 12 h. The solvent was removed in vacuo. And it was diluted with EtOAc (2×150 mL), washed with water (2×70 mL), brine (80 mL), then dried (MgSO₄), filtered and concentrated in *vacuo*. The resulting residue was purified by silica gel column chromatography (Hexane: EtOAc, 7:3) to furnish compound S7b (6.87 g, 82%) as yellow oil. Rf 0.48 (hexane/EtOAc 7:3); ¹H NMR (600 MHz, CDCl₃): δ 7.42 (d, J = 7.8 Hz, 2H), 7.38 (d, J = 7.2 Hz, 2 H), 7.35 (d, J = 6.6 Hz, 2 H), 7.33-7.30 (m, 2 H), 7.28-7.27 (m, 2 H), 7.12 (d, J = 8.4 Hz, 2 H), 6.90 (app d, *J* = 6.6 Hz, 2 H), 5.51 (s, 1 H), 4.93 (d, *J* = 11.4 Hz, 1 H), 4.78 (d, *J* = 11.4 Hz, 1 H), 4.56 (d, *J* = 9.6 Hz, 1 H), 4.36 (dd, J = 4.8, 10.2 Hz, 1 H), 3.81 (s, 3 H), 3.76 (t, J = 10.8 Hz, 1 H), 3.66 (t, J = 4.8 Hz, 1 H), $3.60 (t, J = 9.6 Hz, 1 H), 3.50-3.48 (m, 1 H), 3.46 (dd, J = 9.6, 3.6 Hz, 1 H), 2.51 (s, 3 H); {}^{13}C NMR (150)$ MHz, CDCl₃): δ 160.0, 155.0, 138.7, 138.2, 133.8, 129.8-127.2, 113.6, 101.2, 88.5, 81.6, 81.1, 74.8, 72.1, 70.7, 68.6, 55.3, 21.1; m/z (HRMS) calcd for C₂₈H₃₀O₆SNa [M+Na]⁺: 517.1655, found:.517.1686. A solution of compound S7b (6.87 g, 14.07 mmol) in pyridine (40 mL) was treated with BzCl (3.41 mL, 24.33 mmol) at 0 °C. The reaction mixture was allowed to warm up slowly to room temperature over a period of 12 h. It was diluted with EtOAc ($2 \times 100 \text{ mL}$), washed with water ($2 \times 50 \text{ mL}$), brine (80 mL), then dried (MgSO₄), filtered and concentrated in vacuo. The resulting residue was purified by silica gel column chromatography (Hexane: EtOAc, 8:2) to furnish compound S8 (6.65 g, 81%). Rf 0.5

(hexane/EtOAc 8:2); ¹H NMR (600 MHz, CDCl₃): δ 8.00 (dd, *J* = 7.8, 1.2 Hz, 2 H), 7.63-7.59 (m, 2 H), 7.47 (t, *J* = 6.6 Hz, 2 H), 7.42 (d, *J* = 4.8 Hz, 2 H), 7.33 (d, *J* = 6.6 Hz, 2 H), 7.11-7.04 (m, 7 H), 6.91 (d, *J* = 7.2 Hz, 2 H), 5.55 (s, 1 H), 5.26 (app d, *J* = 9.6, 9.0 Hz, 1 H), 4.78 (app d, *J* = 12.0, 7.2 Hz, 2 H), 4.64 (d, *J* = 11.4 Hz, 1 H), 4.38 (dd, *J* = 10.2, 5.4 Hz, 1 H), 3.86 (q, *J* = 9.0 Hz, 1 H), 3.83 (s, 3 H), 3.80-3.76 (m, 1 H), 3.52 (td, *J* = 9.6, 4.8 Hz, 1 H), 2.32 (s, 3 H);¹³C NMR (150 MHz, CDCl₃): δ 165.3, 160.2, 138.6, 137.8, 133.7, 133.2, 130.0, 129.8-127.2, 113.7, 101.3, 87.2, 81.4, 79.4, 74.3, 72.1, 70.7, 68.6, 55.4, 21.2; m/z (HRMS) calcd for C₃₅H₃₄O₇SNa [M+Na]⁺: 621.1917, found: 621.1958.

4-Methylphenyl-2-*O*-benzoyl-3-*O*-benzyl-6-*O*-*p*-methoxybenzyl-1-thio-β-D-glucopyranoside (S10)



A solution of compound **S8** (6.65 g, 11.12 mmol) in anhydrous DMF (40 mL) was cooled to 0 °C. It was then treated with solid NaCNBH₃ (7.0 g, 111.25 mmol) and TFA (8.52 mL, 111.20 mmol), sequentially. The resulting suspension was stirred at room temperature for 12 h until consumption of starting material was observed. It was neutralized by solid NaHCO₃, and the solution was filtered and diluted with EtOAc (2 × 100 mL), washed with water (2 ×50 mL), brine (50 mL), then dried (MgSO₄), filtered and concentrated in *vacuo*. The resulting residue was purified by silica gel column chromatography (hexane: EtOAc, 7:3) to furnish compound **S10** (5.07 g, 76%). The ring opening of the acetal was stereoselective and gave only 4-OH derivative. *R*f0.4 (hexane/EtOAc 7:3); ¹H NMR (600 MHz, CDCl₃): δ 8.16-8.14 (m, 2 H), 7.69-7.67 (m, 1 H), 7.56 (t, *J* = 8.4 Hz, 2 H), 7.45 (d, *J* = 7.8 Hz, 2 H), 7.39-7.34 (m, 5 H), 7.12 (d, *J* = 7.8 Hz, 2 H), 6.99-6.97 (m, 4 H), 5.31 (t, *J* = 9.6 Hz, 1 H), 4.82 (d, *J* = 9.6 Hz, 1 H), 4.80 (d, *J* = 11.4 Hz, 1 H), 4.71 (s, 2 H), 4.60 (d, *J* = 9.6 Hz, 1 H), 3.90 (s, 3 H), 3.87 (d, *J* = 4.8 Hz, 2 H), 3.77 (t, *J* = 9.0 Hz, 1 H), 3.64 (q, *J* = 4.8 Hz, 2 H), 2.42 (s, 3 H); ¹³C NMR (150 MHz, CDCl₃): δ 165.3, 159.4, 159.3, 138.2, 137.9, 133.6, 133.3, 133.1, 133.0, 130.0, 130.0, 129.8-127.9, 114.0, 113.9, 86.7, 83.7, 78.3, 74.9, 73.5, 72.2, 72.1, 70.2, 65.1, 55.4, 21.2; m/z (HRMS) calcd for C₃₅H₃₆O₇SNa [M+Na]⁺: 623.2074, found: 623.2102.

 $\label{eq:2.2.2} 4-Methylphenyl-2-{\it O}-benzoyl-3-{\it O}-benzyl-4-{\it O}-levulinyl-6-{\it O}-p-methoxybenzyl-1-thio-\beta-D-glucopyranoside (18).$



Compound **S10** (5.07 g) was treated as described for preparation of **17**. Compound **18** (4.78 g) was obtained in 81% yield. $R_f 0.5$ (hexane/EtOAc 6:4); ¹H NMR (600 MHz, CDCl₃): δ 8.04-8.02 (m, 2 H),

7.59-7.58 (m, 2 H), 7.46 (t, J = 7.8 Hz, 2 H), 7.35 (d, J = 6.8 Hz, 2 H), 7.28-7.26 (m, 2 H), 7.13-7.08 (m, 4 H), 7.00 (d, J = 7.8 Hz, 2 H), 6.86 (d, J = 8.4 Hz, 2 H), 5.26 (t, J = 9.6 Hz, 1 H), 5.09 (t, J = 9.6 Hz, 1 H), 4.74 (d, J = 10.2 Hz, 1 H), 4.58 (s, 2 H), 4.46 (s, 2 H), 3.86 (t, J = 9.0 Hz, 1 H), 3.81 (s, 3 H), 3.69 (td, J = 4.2, 5.4 Hz, 1 H), 3.60-3.59 (m, 2 H), 2.64-2.54 (m, 2 H), 2.44-2.39 (m, 2 H), 2.28 (s, 3 H), 2.12 (s, 3 H);^{13}C NMR (150 MHz, CDCl_3): \delta 206.2, 177.5, 165.0, 159.2, 138.0, 137.6, 133.7, 133.2, 133.0, 130.2, 130.1, 129.8-127.9, 113.7, 86.5, 81.4, 77.9, 74.1, 73.2, 72.1, 71.0, 69.4, 55.2, 37.7, 29.7, 27.8, 21.2; m/z (HRMS) calcd for C₄₀H₄₂O₉SNa [M+Na]⁺: 721.2442, found:721.2482.

4-Methylphenyl-3-*O*-benzyl-4,6-*O*-benzylidine-2-*O*-levulinyl-1-thio-β-D-glucopyranoside (10b)



To a solution of compound **S7b** (4.0 g, 8.59 mmol) in CH₂Cl₂ (35 mL) was added levulinic acid (0.895 mL, 11.1 mmol), EDCI (3.33 g, 21.4 mmol), DMAP (0.526 g, 4.30 mmol) and the mixture was stirred for overnight at room temperature. It was diluted with EtOAc (2×100 mL), washed with water (2×50 mL), brine (40 mL), then dried (MgSO₄), filtered and concentrated in *vacuo*. The resulting residue was purified by silica gel column chromatography (hexane: EtOAc, 7:3) to furnish compound **10b** (3.92 g, 81%). *R*_f 0.51 (hexane/EtOAc 7:3); ¹H NMR (600 MHz, CDCl₃): δ 7.49 – 7.46 (m, 2H), 7.39 – 7.36 (m, 5H), 7.31 – 7.28 (m, 4H), 7.28 – 7.27 (m, 1H), 7.13 – 7.11 (m, 2H), 5.57 (s, 1H), 4.99 (dd, *J* = 10.1, 8.4 Hz, 1H), 4.85 (d, *J* = 11.9 Hz, 1H), 4.69 (d, *J* = 11.9 Hz, 1H), 4.63 (d, *J* = 10.1 Hz, 1H), 4.38 (dd, *J* = 10.5, 5.0 Hz, 1H), 3.82 – 3.73 (m, 2H), 3.71 (t, *J* = 9.2 Hz, 2H), 3.48 (ddd, *J* = 10.1, 9.0, 5.0 Hz, 1H), 2.75 (td, *J* = 6.8, 2.3 Hz, 2H), 2.66 – 2.58 (m, 1H), 2.58 – 2.51 (m, 1H), 2.34 (s, 3H), 2.19 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 206.2, 171.3, 138.6, 138.2, 137.2, 133.6, 129.7-126.0, 101.3, 87.1, 81.3, 79.8, 76.9, 74.4, 71.8, 70.5, 68.6, 37.9, 29.9, 28.1, 21.2; m/z (HRMS) calcd for C₃₂H₃₅O₇S [M+H]⁺: 563.2098, found: 563.2077.





Reaction conditions:(a) (i) *p*-CrSH, BF₃-Et₂O, CH₂Cl₂, 0 °C-rt, 12 h, 90%; (b) (i) NaOMe, MeOH : CH₂Cl₂(3:1), rt, 5 h; (ii) PhH(OMe)₂, CSA, DMF, rt, 12 h, 81%; (c) BzCl, Py, 0 °C-rt, 12 h, 80%; (d) TFA: CH₂Cl₂:H₂O (1:10:0.1), rt, 1 h, 79%; (e) (i) BAIB, TEMPO, CH₂Cl₂:H₂O (2:1), rt, 2 h; (ii) CH₃I, KHCO₃, DMF, 0 °C-rt, 4 h, 59%; (f) FmocCl, Py, CH₂Cl₂, 0 °C-rt, 4 h, 80%.

4-Methylphenyl 3-O-benzyl-4,6-O-benzylidine-1-thio-α-L-idopyranoside (S14)



1,2,4,6-Tetra-O-acetyl-3-O-benzyl- α/β -D-idopyranoside **S12** was prepared from diacetone α -D-glucose 15 using known procedures.³ It was further treated with *p*-toluenethiol in the presence of BF₃·OEt₂ to furnish **S13** in 90% yield using a reported procedure.^{2c} Compound **S13** (3.0 g, 5.97 mmol) was dissolved in a mixture of MeOH : CH₂Cl₂(30:10 mL) and a catalytic amount of NaOMe (100 mg) was added. The mixture was stirred for 5 h at room temperature and it was neutralized with amberlite 120 H⁺ resin. The resin was filtered and the solvents were evaporated in *vacuo*. The crude mixture dissolved in anhydrous DMF (30 mL) was treated with benzaldehyde dimethyl acetal (0.96 mL, 6.4 mmol) and camphor sulphonic acid (1.23 g, 5.31 mmol). The reaction mixture was stirred for 12 h at room temperature and then quenched with Et₃N. Solvents were removed in *vacuo* and diluted with Et₃OAC (2×100 mL), washed with water $(2 \times 50 \text{ mL})$, brine (50 mL), then dried (MgSO₄), filtered and concentrated in *vacuo*. The resulting residue was purified by silica gel column chromatography (hexane: EtOAc, 1:1) to furnish S14 (1.99 g, 81%). Rf 0.41 (hexane/EtOAc 1:1); ¹H NMR (600 MHz, CDCl₃): δ 7.47-7.43 (m, 4 H), 7.42-7.32 (m, 8 H), 7.10 (d, J = 7.8 Hz, 2 H), 5.59 (s, 1H), 5.55 (s, 1H), 4.85 (d, J = 12.0 Hz, 1 H), 4.60 (d, J = 12.0 Hz, 1 Hz, 1 Hz), 4.60 (d, J = 12.0 Hz), 4.60 (d, JHz, 1 H), 4.47 (s, 1 H), 4.34 (d, J = 12.0 Hz, 1 H), 4.15-4.10 (m, 3 H), 3.80 (app d, J = 2.4 Hz, 2 H), 2.32 (s, 3 H);¹³C NMR (150 MHz, CDCl₃): δ 137.4, 137.3, 136.9, 133.5, 130.8, 129.7-127.8, 126.0, 101.6, 89.5, 74.5, 73.9, 72.4, 70.2, 67.7, 60.6, 21.1; m/z (HRMS) calcd for C₂₇H₂₈O₅SNa [M+Na]⁺: 487.1555, found: 487.1550.

4-Methylphenyl 2-*O*-benzoyl-3-*O*-benzyl-4,6-*O*-benzylidene-1-thio-α-L-idopyranoside (S15)



A solution of **S14** (1.99 g, 4.30 mmol) in pyridine (25 mL) was treated with BzCl (0.99 mL, 8.57 mmol) at 0 °C. The reaction mixture was allowed to warm up slowly to room temperature over a period of 12 h. It was diluted with EtOAc (2 × 100 mL), washed with water (2 × 50 mL), brine (80 mL), then dried (MgSO4), filtered and concentrated in *vacuo*. The resulting residue was purified by silica gel column chromatography (hexane/EtOAc 9:1) to furnish compound **S15** (1.94 g, 80%). *R*f 0.57 (hexane/EtOAc 8:2); ¹H NMR (600 MHz, CDCl₃): δ 7.98 (d, *J* = 7.2 Hz, 2 H), 7.53-7.48 (m, 5 H), 7.46 (d, *J* = 8.4 Hz, 2 H), 7.40 (t, *J* = 8.4 Hz, 2 H), 7.35 (t, *J* = 6.0 Hz, 2 H), 7.30 (t, *J* = 7.8 Hz, 2 H), 7.24 (t, *J* = 7.2 Hz, 2 H), 7.12 (d, *J* = 7.8 Hz, 2 H), 5.77 (s, 1 H), 5.61 (s, 1 H), 5.56 (app d, *J* = 1.2 Hz, 1 H), 5.01 (d, *J* = 11.4 Hz, 1 H), 4.74 (d, *J* = 12.0 Hz, 1 H), 4.55 (s, 1 H), 4.42 (d, *J* = 12.6 Hz, 1 H), 4.24 (d, *J* = 12.6 Hz, 1 H), 4.14 (s, 1 H), 3.94 (s, 1 H), 2.34 (s, 3 H);¹³C NMR (150 MHz, CDCl₃): δ 165.7, 137.9, 137.3, 137.2, 133.1, 132.8, 131.0, 130.1,129.7-127.9, 126.4, 101.1, 86.4, 73.3, 72.5, 70.0, 67.9, 60.2, 21.1; m/z (HRMS) calcd for C₃₄H₃₂O₆SNa [M+Na]⁺: 591.1812, found, 591.1827.

4-Methylphenyl 2-*O*-benzoyl-3-*O*-benzyl-1-thio-α-L-idopyranoside (S16)



A solution of a compound **S15** (1.94 g, 3.41 mmol) in a mixture of solvents (CH₂Cl₂: TFA: H₂O = 10/1/0.1, v/v/v) was stirred at room temperature for 1 h. It was neutralized by solid NaHCO₃, filtered and then concentrated in *vacuo*, and later co-evaporated with toluene to remove traces of water. The crude mixture was purified by silica gel column chromatography (hexane/EtOAc 7:3) to furnish **S16** (1.58 g, 79%). *R*_f 0.28 (hexane/EtOAc 7:3); ¹H NMR (600 MHz, CDCl₃): δ 8.04 (d, *J* = 8.4 Hz, 2 H), 7.58 (t, *J* = 7.8 Hz, 1 H), 7.46-7.43 (m, 6 H), 7.38 (t, *J* = 7.8 Hz, 2 H), 7.33-7.31 (m, 1 H), 7.13 (d, *J* = 7.8 Hz, 2 H), 5.56 (s, 1 H), 5.52 (app s, 1 H), 4.91 (d, *J* = 11.4 Hz, 1 H), 4.81 (d, *J* = 5.4 Hz, 1 H), 4.66 (d, *J* = 10.2 Hz, 1 H), 3.98 (dd, *J* = 12.0, 4.2 Hz, 1 H), 3.85 (app s, 2 H), 2.32 (s, 3 H);¹³C NMR (150 MHz, CDCl₃): δ 165.1, 138.1, 137.3, 133.7, 131.9, 129.8-128.1, 127.9, 87.2, 74.1, 72.4, 69.9, 68.4, 68.2, 63.4, 21.2; m/z (HRMS) calcd for C₂₇H₂₈O₆SNa [M+Na]⁺: 503.1504, found, 503.1532.

Methyl *p*-tolyl-2-*O*-benzoyl-3-*O*-benzyl-1-thio-α-L-idopyranosyl uronate (S17)



To a vigorously stirred solution of the diol derivative **S16** (1.58 g, 3.29 mmol) in a mixture of CH₂Cl₂: H₂O (2:1) was added 2,2,6,6-tetramethyl-1-piperidinyloxy (TEMPO, 0.154 g, 0.987 mmol) in the presence of iodobenzene diacetate (BAIB, 2.65 g, 8.21 mmol) as the cooxidant at 0 °C. The reaction mixture was allowed to warm up to room temperature and continued to stir until the complete conversion of the starting material was observed. After 2 h, it was diluted with CH_2Cl_2 (2 × 50 mL), washed with 10% Na₂S₂O₃(2×30 mL), brine (40 mL), then dried (MgSO₄), filtered and concentrated in *vacuo*. It was co-evaporated with toluene to remove traces of water. The resulting residue was dissolved in anhydrous DMF (15 mL) and cooled to 0 °C. Methyl iodide (CH₃I, 0.47 mL, 7.54 mmol) and KHCO₃ (0.345 g, 3.42 mmol) were added to the mixture under N_2 atmosphere. It was stirred for a period of 4 h allowing the reaction mixture to warm up to room temperature, slowly. After 4 h, it was diluted with EtOAc (2×50 mL), washed with water $(2 \times 30 \text{ mL})$, brine (40 mL), then dried (MgSO₄), filtered and concentrated in vacuo. The crude mixture was purified by silica gel column chromatography (hexane/EtOAc 7:3) to furnish **S17** (0.986 g, 59%) as colorless gum. *R*_f 0.46 (hexane/EtOAc 7:3); ¹H NMR (600 MHz, CDCl₃): δ 7.97-7.95 (m, 2 H), 7.58 (t, J = 7.2 Hz, 1 H), 7.46-7.37 (m, 8 H), 7.33-7.31 (m, 1 H), 7.12 (d, J = 7.8 Hz, 2 H), 5.67 (s, 1 H), 5.50 (app s, 1 H), 5.42 (app s, 1 H), 4.92 (d, J = 12.0 Hz, 1 H), 4.70 (d, J = 12.0 Hz, 1 Hz, 1 Hz), 4.70 (d, J = 12.0 Hz, 1 Hz), 4.70 (d, J = 12.0 Hz, 1 Hz), 4.70 (d, J = 12.0 Hz), 4.70 (Hz, 1 H), 4.15 (d, J = 11.4 Hz, 1 H), 3.98 (s, 1 H), 3.84 (s, 3 H), 2.83 (d, J = 12.0 Hz, 1 H), 2.32 (s, 3 H); ¹³C NMR (150 MHz, CDCl₃): δ 169.7, 165.0, 137.9, 137.1, 133.8, 132.0, 131.8, 129.5-127.9, 87.3, 73.6, 72.5, 69.7, 69.0, 68.4, 52.5, 21.1;m/z (HRMS) calcd for C₂₈H₂₈O₇SNa [M+Na]⁺: 531.1448, found, 531.1479.

Methyl*p*-tolyl-2-*O*-benzoyl-3-*O*-benzyl-4-*O*-(9-fluorenylmethoxycarbonyl)-1-thio-α-L-idopyranosyl uronate (12)



To a solution of the **S17** (0.986 g, 1.94 mmol) in anhydrous CH₂Cl₂ was added 9-fluroenylmethoxycarbonyl chloride (0.754 g, 2.91 mmol) in the presence of pyridine (1.56 mL, 19.45 mmol) at 0 °C. It was stirred for a period of 4 h allowing the reaction mixture to warm up to room temperature, slowly. After 4 h, the reaction mixture was diluted with EtOAc (2×50 mL), washed with

water (2 × 30 mL), brine (40 mL), then dried (MgSO₄), filtered and concentrated in *vacuo*. The crude mixture was purified by silica gel column chromatography (hexane/EtOAc 8:2) to furnish **12** (1.13 g, 80%) as solid. *R*_f 0.56 (hexane/EtOAc 8:2); ¹H NMR (600 MHz, CDCl₃): δ 8.12 (d, *J* = 7.2 Hz, 2 H), 7.75 (d, *J* = 7.8 Hz, 2 H), 7.58 (d, *J* = 7.8 Hz, 2 H), 7.47-7.39 (m, 12 H), 7.34 (t, *J* = 7.2 Hz, 1 H), 7.28-7.21 (m, 2 H), 7.19 (t, *J* = 7.2 Hz, 1 H), 5.71 (s, 1 H), 5.53 (s, 1 H), 5.49 (s, 1 H), 5.26 (s, 1 H), 4.92 (d, *J* = 12.0 Hz, 1 H), 4.81 (d, *J* = 12.6 Hz, 1 H), 4.33 (t, *J* = 9.6 Hz, 1 H), 4.23 (t, *J* = 7.2 Hz, 1 H), 4.11-4.08 (m, 2 H), 3.82 (s, 3 H), 2.32 (s, 3 H);¹³C NMR (150 MHz, CDCl₃): δ 168.7, 165.4, 154.4, 143.3, 142.9, 141.3, 141.2, 137.8, 136.9, 133.5, 131.8, 130.2, 129.6-127.3, 125.3, 125.2, 120.1, 86.2, 72.9, 71.8, 71.2, 70.3, 68.5, 66.9, 52.7, 46.6, 21.1; m/z (HRMS) calcd for C₄₃H₃₈O₉SNa [M+Na]⁺: 753.2129, found: 753.2166.





Reaction conditions: (a) TfN₃, MeOH, CuSO₄, Et₃N, rt, 12 h, 69 %; (b) 10 % HCl/MeOH, 90 °C, 7 h, 82%; (c) PhH(OMe)₂, CSA, DMF, rt, 12 h, 73%; (d) Ac₂O, Py, 0 °C-rt, 5 h; (e) (i) NaOMe, MeOH : $CH_2Cl_2(3:1)$, rt, 2 h; (ii) NaH, BnBr, THF, 0 °C-rt, 12 h, 92%; (f) 80% AcOH, 60 °C, 5h, 86%; (g) Ac₂O (1.1), Et₃N (9), CH₂Cl₂, 0 °C, 1 h, 86%; (h) CMPI, CH₃CO(CH₂)₂COOH, CH₂Cl₂, rt, 15 min; then DABCO, -20 °C -rt, 81%

$Methyl-2-azido-4, 6-{\it O}-benzylidene-2-deoxy-\alpha/\beta-D-glucopyranoside~(S20)$



A solution of sodium azide (20 g, 307.7 mmol) and pyridine (25 mL, 316.45 mmol) in MeCN (100 mL) at 0 °C was treated with trifluoromethanesulfonic anhydride (Tf₂O, 40 mL, 235 mmol) slowly from an addition funnel. It was stirred at the same temperature for 2 h and the mixture was filtered through Celite pad with the temperature of the filtrate maintained at 0 °C. In a separate flask, D-glucosamine hydrochloride (16, 25 g, 121.95 mmol), CuSO4·5H2O (0.195 g, 1.21 mmol), and Et3N (34.0 mL, 243 mmol) were dissolved in MeOH (80 mL) and it was cooled to 0 °C. A freshly prepared cold TfN3solution was sequentially added and the reaction was stirred at room temperature for 12 h.² After the complete conversion of the starting material, the reaction mixture was concentrated in vacuo and the crude S18 was obtained in 69% yield. The crude mixture was treated with 10% HCl in MeOH (100 mL) and heated at 90 °C for a period of 7 h.^{3a} After 7 h, the solvents were evaporated in vacuo and the mixture was redissolved in MeOH and treated with solid NaHCO₃ till the pH of the solution is 7. The crude mixture was filtered and concentrated to give crude methyl glycosides S19 (8 g, 82%). The crude reaction mixture was co-evaporated with toluene to remove traces of water and then dried for 2-3 h in vaccum. To a solution of compound **S19** (8 g, 36.52 mmol) in anhydrous DMF (40 mL) was added benzaldehyde dimethyl acetal (6.7 mL, 43.82 mmol) and camphor sulphonic acid (8.48 gm, 36.52 mmol). The reaction was continued with stirring for 12 h at room temperature and then quenched with Et₃N. The solvents were removed in *vacuo* and the crude product was diluted with EtOAc (2×200 mL), washed with water (2×100 mL), brine (50 mL), dried (MgSO₄), filtered. The organic layer was concentrated in vacuo and the resulting residue was purified by silica gel column chromatography (Hexane: EtOAc, 8:2) to afford S20 (8.18 g, 73%) as anomeric mixture (α : β = 1:1). $R_f 0.58$ (hexane/EtOAc 8:2); ¹H NMR (600 MHz, CDCl₃): δ 7.49-7.47 (m, 4H), 7.39-7.37 (m, 6H), 5.53 (s, 1H), 4.79 (d, *J* = 3.6 Hz, 1H), 4.34 (dd, *J* = 10.2, 4.8 Hz, 1H), 4.30 (d, J = 8.4 Hz, 1H), 4.27 (dd, J = 12.0, 4.8 Hz, 1H), 4.16 (t, J = 9.0 Hz, 1H), 4.11 (d, J = 7.2 Hz, 1H),3.81 (td, *J* = 10.2, 4.8 Hz, 1H), 3.75 (td, *J* = 10.2, 3.0 Hz, 2H), 3.65 (t, *J* = 9.0 Hz, 1 H), 3.59 (s, 3 H), 3.51 (td, J = 9.0, 2.4 Hz, 1H), 3.44 (s, 3H), 3.41-3.35 (m, 3H), 3.30 (dd, J = 10.2, 3.6 Hz, 1H);¹³C NMR (150 MHz, CDCl₃): § 136.9, 136.8, 129.5, 128.5, 126.3, 126.3, 103.5, 102.2, 102.0, 99.5, 81.8, 80.7, 72.1, 69.1, 68.9, 68.5, 66.4, 66.2, 63.3, 62.3, 57.6, 55.6; m/z (HRMS) calcd for C14H18N3O5[M+H]⁺: 308.1241, found:308.1269.

Methyl 3-*O*-acetyl-2-azido-4,6-*O*-benzylidene-2-deoxy-α-D-glucopyranoside (S21) Methyl 3-*O*-acetyl-2-azido-4,6-*O*-benzylidene-2-deoxy-β-D-glucopyranoside (S22)



A solution of **S20** (4.0 g, 13.01 mmol) in pyridine (30 mL) was treated with Ac₂O (15 mL) at 0 °C. The reaction mixture was stirred at room temperature for 5 h. The reaction mixture was concentrated in *vacuo* and diluted with EtOAc (2 × 100 mL), washed with water (2 × 50 mL), brine (50 mL), then dried (MgSO₄), filtered and concentrated in *vacuo*. The resulting residue was purified by silica gel column chromatography (Hexane: EtOAc, 7:3) to furnish crystalline α -anomer **S21** in (2.81 g, 62%) and β -anomer **S22** as colorless gum (1.63 g, 36%). *R*_f 0.62 (hexane/EtOAc 7:3);

S21:¹H NMR (600 MHz, CDCl₃): δ 7.44-7.43 (m, 2H), 7.36-7.34 (m, 3H), 5.59 (t, *J* = 10.2 Hz, 1H), 5.50 (s, 1H), 4.86 (d, *J* = 3.0 Hz, 1H), 4.30 (dd, *J* = 10.2, 4.8 Hz, 1H), 3.95 (td, *J* = 10.2, 4.8 Hz, 1H), 3.76 (d, *J* = 10.2 Hz, 1H), 3.62 (d, *J* = 9.6 Hz, 1H), 3.48 (s, 3H), 3.27 (dd, *J* = 10.2, 3.6 Hz, 1H), 2.16 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 169.8, 136.9, 129.2, 128.3, 126.2, 101.7, 99.9, 79.5, 69.1, 68.9, 62.7, 61.8, 55.5, 20.9; m/z (HRMS) calcd for C₁₆H₁₉N₃O₆Na [M+Na]⁺: 372.1166, found:.372.1187.

S22:¹H NMR (600 MHz, CDCl₃): δ 7.43-7.36 (m, 2H), 7.36-7.26 (m, 3H), 5.50 (s, 1H), 5.17 (t, *J* = 9.6 Hz, 1H), 4.38 (d, *J* = 7.8 Hz, 1H), 4.36 (dd, *J* = 10.8, 5.4 Hz, 1H), 3.80 (t, *J* = 10.2 Hz, 1H), 3.62 (d, *J* = 12.0 Hz, 1H), 3.60 (s, 3H), 3.50-3.47 (m, 1H), 3.46-3.45 (m, 1H), 2.13 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 169.7, 136.8, 129.1, 128.3, 126.1, 103.6, 101.5, 78.7, 71.2, 68.5, 66.4, 64.7, 57.6, 20.9; m/z (HRMS) calcd for C₁₆H₁₉N₃O₆Na [M+Na]⁺: 372.1166, found: 372.1187.

Methyl 2-azido-3-*O*-benzyl-4,6-*O*-benzylidene-2-deoxy-α-D-glucopyranoside (S23).



Compound **S21** (2.81 g, 8.05 mmol) was dissolved in a mixture of MeOH : CH₂Cl₂ (30:10 mL) and a catalytic amount of NaOMe (100 mg) was added. The mixture was stirred for 2 h at room temperature after that it was neutralized with amberlite 120 H⁺ resin. The resin was filtered and the solvents were evaporated to give the crude mixture which was co-evaporated with toluene and dissolved in THF (20 mL). It was cooled to 0 °C and NaH (0.651 g, 17.5 mmol) was added. After the mixture was stirred for 1h, BnBr (1.17 mL, 9.77 mmol) was added to it and the reaction mixture was further stirred for a period of 12 h at room temperature. The resulting mixture was filtered through a Celite pad and the filtrate was concentrated in *vacuo*. It was diluted with EtOAc (100 mL), washed with water (2 × 50 mL), brine (50

mL), then dried (MgSO₄), filtered and concentrated in *vacuo*. The resulting residue was purified by silica gel column chromatography (hexane: EtOAc, 8:2) to furnish **S23** as gum (2.94 g, 92%). *R*f 0.46 (hexane/EtOAc 8:2); ¹H NMR (600 MHz, CDCl₃): δ 7.50-7.48 (m, 2 H), 7.41-7.37 (m, 5 H), 7.33-7.24 (m, 3 H), 5.59 (s, 1 H), 4.94 (d, *J* = 10.8 Hz, 1 H), 4.81 (s, 1 H), 4.78 (dd, *J* = 10.8, 4.2 Hz, 1 H), 4.30 (dd, *J* = 10.8, 4.2 Hz, 1 H), 4.06 (t, *J* = 9.6 Hz, 1 H), 3.88 (td, *J* = 10.2, 4.8 Hz, 1 H), 3.77 (t, *J* = 10.2 Hz, 1 H), 3.72 (t, *J* = 6.0 Hz, 1 H), 3.45 (app d, *J* = 3.6 Hz, 1 H), 3.44 (app s, 3 H); ¹³C NMR (150 MHz, CDCl₃): δ 137.9, 137.3, 129.1-126.1, 101.5, 99.5, 82.8, 76.4, 75.1, 69.0, 63.2, 62.7, 55.5; m/z (HRMS) calcd for C₂₁H₂₃N₃O₅Na [M+Na]⁺: 420.1530, found: 420.1547.

Methyl 6-O-acetyl-2-azido-3-O-benzyl-2-deoxy-α-D-glucopyranoside (13a).



Compound **S23** (2.94 g, 7.40 mmol) was dissolved in 80% AcOH-H₂O and the mixture was stirred at 60 °C for a period of 5 h, then concentrated and the residue was co-evaporated with toluene to remove traces of water to generate the 4,6-diol derivative **S24**. Then, it was dissolved in anhydrous CH₂Cl₂ (25 mL) and treated with Ac₂O (0.85 mL, 8.88 mmol) and Et₃N (9.28 mL, 91.96 mmol) at 0 °C under argon atmosphere for 1 h. After complete conversion of the starting material, it was diluted with CH₂Cl₂ (100 mL), washed with saturated NaHCO₃ (2 × 50 mL), brine (50 mL), then dried (MgSO₄), filtered and concentrated in *vacuo*. The resulting residue was purified by silica gel column chromatography (hexane: EtOAc, 6:4) to furnish **13a** as gum (2.23 g, 86%). *R*f 0.38 (hexane/EtOAc 6:4); ¹H NMR (600 MHz, CDCl₃): δ 7.42-7.32 (m, 5 H), 4.92 (d, *J* = 10.8 Hz, 1 H), 4.81 (s, 1H), 4.80 (app d, *J* = 3.0 Hz, 1 H), 4.51 (dd, *J* = 12.0, 4.2 Hz, 1 H), 4.19 (dd, *J* = 11.4, 2.4 Hz, 1 H), 3.82 (app dd, *J* = 10.2, 9.0 Hz, 1 H), 3.76-3.74 (m, 1 H), 3.48 (td, *J* = 10.8, 3.0 Hz, 1 H), 3.43 (s, 3 H), 3.36 (dd, *J* = 10.2, 3.6 Hz, 1 H), 2.63 (s, 1 H), 2.09 (s, 3 H); ¹³C NMR (150 MHz, CDCl₃): δ 171.9, 137.9, 128.8, 128.3, 128.2, 98.9, 79.7, 75.4, 70.6, 69.9, 63.2, 55.4, 20.9; m/z (HRMS) calcd for C₁₆H₂₁N₃O₆Na [M+Na]⁺: 374.1323, found: .374.1345.

Methyl 2-azido-3-O-benzyl-6-O-levulinyl-2-deoxy-α-D-glucopyranoside (13b).



To a solution of **S24** (1.5 g, 4.85 mmol) in CH₂Cl₂(20 mL) was added levulinic acid (0.741, 7.27 mmol) and 2-chloromethyl pyridinium chloride (CMPI) (3.10 g, 12.13 mmol). The reaction mixture was stirred for 15 min at room temperature and then cooled to -20 °C. And 1,4-diazabicyclo[2,2,2] octane (1.90 g, 16.97 mmol) was added to the mixture at the same temperature. The reaction mixture was allowed to warm up to room temperature slowly with a period of 90 min. It was filtered through celite, diluted with EtOAc (2 × 50 mL), washed with brine (2 × 50 mL), then dried (MgSO4), filtered and concentrated in *vacuo*. The resulting residue was purified by silica gel column chromatography (Hexane: EtOAc, 6:4) to furnish **13b** as colorless oil (1.60 g, 81%). *R*_f 0.42 (hexane/EtOAc 6:4); ¹H NMR (600 MHz, CDCl₃): δ 7.43 – 7.39 (m, 2H), 7.37 (ddd, *J* = 7.5, 6.7, 1.2 Hz, 1H), 7.34 – 7.30 (m, 1H), 7.21 – 7.15 (m, 1H), 4.92 (d, *J* = 11.1 Hz, 1H), 4.83 (d, *J* = 11.1 Hz, 1H), 4.79 (d, *J* = 3.6 Hz, 1H), 4.56 (dd, *J* = 12.2, 4.1 Hz, 1H), 4.19 (dd, *J* = 12.3, 2.2 Hz, 1H), 3.43 (s, 3H), 3.37 (dd, *J* = 10.2, 3.6 Hz, 1H), 2.82 – 2.77 (m, 1H), 2.79 – 2.75 (m, 2H), 2.61 (dd, *J* = 6.8, 6.0 Hz, 1H), 2.19 (s, 3H);¹³C NMR (150 MHz, CDCl₃): δ 206.7, 173.5, 138.1, 129.1, 128.2, 98.9, 79.7, 75.3, 70.6, 70.0, 63.2, 63.1, 55.4, 38.0, 29.9, 27.9;m/z (HRMS) calcd for C₁₉H₂₅N₃O₇Na [M+Na]⁺: 430.1585, found: 430.1594.

Scheme S5. Synthesis of 2-azido acceptors (Building Block E)





Reaction conditions: (a) (i) Ac₂O, DMAF, 0 °C-rt, 16 h, 84%; (b) NH₂(CH₂)₂NH₂, AcOH, THF, rt, 16 h, 87 %; (c) CCl₃CN, DBU, CH₂Cl₂, 0 °C-rt, 90 min, 74%; (d) (i) benzaldehyde (1.0 eq.), EtOH, 50 °C, reduced pressure; (ii) NaBH₄(1.2 eq.), MeOH, 0 °C to r.t.; (iii) Benzyl chloroformate (1.0 eq.), Et₂O/H₂O, NaHCO₃, 0° C to r.t., 89% over three steps. (e) TMSOTf, CH₂Cl₂:Et₂O (1:5), MS, -20 °C, 1 h, 84%; (f) (i) NaOMe/ MeOH, rt, 12 h; (ii) PhH(OMe)₂, CSA, DMF, rt, 12 h, 81 %; (iii) NaH, BnBr, DMF, 12 h, 95%; (g) 80% AcOH, 5 h, 60 °C; (h) CMPI, CH₃CO(CH₂)₂COOH, CH₂Cl₂, rt, 15 min, DABCO, -20 °C -rt, 81%; (i) Ac₂O (1.1), Et₃N (9), CH₂Cl₂, 0 °C, 1 h, 82%.

1,3,4,6-Tetra-O-acetyl -2-azido-2-deoxy-D-glucopyranose (S26).

Compound **S25** can be easily prepared from the readily available D-glucosamine hydrochloride **16** based on previously reported method.² After the complete conversion of the starting material, the reaction mixture was concentrated in *vacuo* and the crude **S25** (4.0 g, 19.55 mmol) was dissolved in acetic anhydride (30 mL). The reaction mixture was cooled to 0 °C and N,N-dimethyl amino pyridine (DMAP, 0.235 g, 1.95 mmol) was added. The mixture was stirred for 16 h allowing to warm up to room temperature. The solvents were evaporated in *vacuo* and the crude mixture was diluted with EtOAc (200 mL), washed with water (2 × 50 mL), brine (75 mL), then dried (MgSO4), filtered and concentrated in *vacuo*. The resulting residue was purified by silica gel column chromatography (hexane: EtOAc, 7:3) to furnish **S26** as gum (α : β =4:1, 6.12 g, 84%). *R*f0.46 (hexane/EtOAc 7:3); ¹H NMR (600 MHz, CDCl₃): δ 6.24 (d, *J* = 3.6 Hz, 0.25 H), 5.52 (d, *J* = 8.6 Hz, 1H), 5.40 (dd, *J* = 10.5, 9.4 Hz, 0.3H), 5.26 (s, 0.4H), 5.05 (dd, *J* = 10.1, 9.3 Hz, 1H), 4.99 (t, *J* = 9.7 Hz, 1H), 4.24 (ddd, *J* = 12.4, 5.8, 4.2 Hz, 1H), 4.08 – 4.04 (m, 1H), 4.05 – 4.00 (m, 2H), 3.78 (ddd, *J* = 10.0, 4.5, 2.2 Hz, 1H), 3.64 – 3.59 (m, 1H), 2.14 (app d, *J* = 1.7 Hz, 4H), 2.05 (s, 1H), 2.04 (s, 3H), 2.02 (s, 4H), 1.99 (s, 1H), 1.98 (s, 1H), 1.97 (s, 3H).¹³C NMR (150 MHz, CDCl₃): δ 171.1, 170.5, 170.0, 169.7, 169.6, 169.5, 168.5, 168.4, 92.5, 89.9, 77.3, 76.9, 72.6, 72.6, 70.7, 69.7, 67.9, 67.8, 62.5, 61.4, 61.4, 60.3, 60.2, 53.5, 21.0, 20.8, 20.8, 20.6, 20.6, 20.5, 20.4; m/z (HRMS) calcd for C₂₁H₂₃N₃O₅Na [M+Na]⁺: 420.1530, found: 420.1547.

3,4,6-Tri-O-acetyl-2-azido-2-deoxy-D-glucopyranose (S27)



In a round bottom flask, ethylene diamine (1.31 mL, 19.65 mmol) and AcOH (1.37 mL, 22.4 mmol) were added in THF (100 mL) under argon atmosphere. To this suspension, compound **S26** was added (6.12 g, 16.1 mmol) and the reaction mixture was stirred for 16 h at room temperature. The crude mixture was diluted with CH₂Cl₂ (200 mL), washed with water (2×50 mL), brine (75 mL), then dried (MgSO₄), filtered and concentrated in *vacuo*. The resulting residue was purified by silica gel column chromatography (Hexane: EtOAc, 6:4) to furnish **S27** as white gum (4.71 g, 87%). The spectroscopic data was in agreement with that in the literature.⁴

3,4,6-Tri-O-acetyl-2-azido-2-deoxy-α-D-glucopyranosyl trichloroacetimidate (S28)



To a solution of compound **S27** (4.71 g, 14.22 mmol) in anhydrous $CH_2Cl_2(30 \text{ mL})$ at 0 °C was added trichloroacetonitrile (14.27 mL, 142.2 mmol) and 1,8-diazabicyclo-[5.4.0]undec-7-ene (0.324 mL, 2.13 mmol) under argon atmosphere. The reaction mixture was stirred for 90 min at 0 °C. Then the solvent was evaporated in *vacuo* and crude residue was purified by silica gel chromatography (hexane: EtOAc, 8:2) to furnish **S28** (4.99 gm, 74%) as a colorless foam. The spectroscopic data was in agreement with that in the literature.⁵

 $N-(Benzyl)-benzyloxycarbonyl-5-aminopentyl-3,4,6-tri-{\it O}-acetyl-2-azido-2-deoxy-\alpha-D-glucopyranoside~(S30)$



Glycosyl donor **S28** (2.5 g, 5.26 mmol), acceptor **S29** (2.06 g, 6.31 mmol) and flame activated AW-300 MS (6.0 g) were suspended in CH₂Cl₂:Et₂O (1:5, 40 mL) for 1 h at room temperature under argon atmosphere. The mixture was cooled to -25 °C. After then, TMSOTf (95 μ L, 0.526 mmol) was added and the mixture was stirred at the same temperature for 1h. It was warmed to room temperature slowly. Triethylamine (1 mL) was added and the mixture was filtered using Celite pad and the filtrate was diluted with CH₂Cl₂ (2 × 30 mL), washed with saturated NaHCO₃(40 mL), brine (30 mL), then dried (MgSO₄) and filtered. Then, the solvent was evaporated in *vacuo* and the crude residue was purified by silica gel column chromatography (hexane: EtOAc, 8:2) to furnish **S30** (2.70 gm, 84%) as an oil. The spectroscopic data was in agreement with that in the literature.⁵

$N-(Benzyl)-benzyloxy carbonyl-5-aminopentyl-2-azido-4-{\it O}-benzyl-4, 6-{\it O}-benzylidene-2-deoxy- \alpha-D-glucopyranoside~(S31)$



A solution of S30 (2.70 g, 4.31 mmol) was dissolved in MeOH (25 mL) and a catalytic amount of NaOMe (0.100 g) was added. The mixture was stirred for 12 h at room temperature, and after that it was neutralized with amberlite 120 H⁺ resin. The resin was filtered and the solvents were evaporated to furnish the triol derivative which was dissolved in anhydrous DMF (40 mL) and benzaldehyde dimethyl acetal (0.9 mL, 5.55 mmol) and camphor sulphonic acid (1.06 g, 4.6 mmol) were added. The reaction was continued with stirring for 12 h at room temperature and then quenched with Et₃N. The solvents were removed in *vacuo* and the mixture was diluted with EtOAc ($2 \times 100 \text{ mL}$), washed with water ($2 \times 50 \text{ mL}$), brine (50 mL), then dried (MgSO₄), filtered and concentrated in *vacuo*. The resulting residue was purified by silica gel column chromatography (hexane: EtOAc, 1:1) to furnish the 4,6-O-benzylidene derivative (2.20 g, 81%). The spectroscopic data was in agreement with that in the literature. The 4,6-O-benzylidene derivative (2.20 g, 3.74 mmol) was dissolved in DMF (25 mL) and cooled to 0 °C. Then, NaH (0.3 g, 7.48 mmol) and BnBr (1.17 mL, 9.77 mmol) were added. The reaction mixture was further stirred for a period of 12 h at room temperature. The resulting mixture was filtered through a Celite and was concentrated in *vacuo*. It was diluted with EtOAc (100 mL), washed with water (2×50 mL), brine (50 mL), then dried (MgSO₄), and the filtrate was concentrated in *vacuo*. The resulting residue was purified by silica gel chromatography (hexane: EtOAc, 8:2) to give S31 as gum (2.40 g, 95%). ¹H NMR (600 MHz, CDCl₃):δ 7.51 (dd, J = 7.5, 2.1 Hz, 2H), 7.41-7.36 (m, 8H), 7.34-7.29 (m, 5H), 7.28-7.25 (m, 2H), 7.20-7.16 (m, 2H), 5.60 (s, 1H), 5.19-5.17 (m, 2H), 4.96 (d, J = 11.0 Hz, 1H), 4.85-4.83 (m, 1H), 4.81 (d, J = 11.0 Hz,

1H), 4.51 (d, J = 11.8 Hz, 2H), 4.28 (app d, J = 9.6 Hz, 1H), 4.07 (app t, J = 10.1 Hz, 1H), 3.88 (br s, 1H), 3.76 (t, J = 10.3 Hz, 1H), 3.71 (t, J = 9.3 Hz, 1H), 3.47 – 3.36 (m, 2H), 3.34 (dd, J = 10.0, 3.6 Hz, 1H), 3.29-3.22 (m, 1H), 1.60-1.52 (m, 4H), 1.43 – 1.28 (m, 2H); ¹³C NMR (150 MHz, CDCl₃): δ 156.8, 156.3, 138.0, 137.2, 136.9, 129.9-126.0, 101.5, 98.6, 82.9, 76.1, 75.1, 69.0, 67.2, 63.0, 62.8, 50.6, 50.3, 47.1, 46.2, 29.8, 29.1, 27.9, 27.5, 23.4; (HRMS) calcd for C₃₉H₄₂N₄O₇Na [M+Na]⁺: 678.3053, found, 678.3094.

$N-(Benzyl)-benzyloxycarbonyl-5-aminopentyl-6-{\it O}-acetyl-2-azido-3-{\it O}-benzyl-2-deoxy-\alpha-D-glucopyranoside~(24a)$



Compound **S32** (1.20 g, 2.03 mmol) was dissolved in 80 % AcOH-H₂O and was stirred at 60 °C for a period of 5 h. It was concentrated and the residue was co-evaporated with toluene to remove the last traces of water. It was then dissolved in anhydrous CH₂Cl₂(15 mL) and treated with Ac₂O (0.24 mL, 2.23 mmol) and Et₃N (2.6 mL, 18.3 mmol) at 0 °C under argon atmosphere for 30 min. The reaction mixture was diluted with CH₂Cl₂(50 mL), washed with saturated NaHCO₃(2×30 mL), brine (50 mL), then dried (MgSO₄), filtered and concentrated in *vacuo*. The resulting residue was purified by silica gel chromatography (Hexane: EtOAc, 6:4) to generate **24a** as gum (1.06 g, 82%). The spectroscopic data was the same as that in the literature.⁶

N-(Benzyl)-benzyloxycarbonyl-5-aminopentyl-2-azido-3-*O*-benzyl-6-*O*-levulinyl-2-deoxy-α-D-glucopyranoside (24b)



To a solution of **S32** (1.20 g, 2.03 mmol) in CH₂Cl₂(15 mL) was added levulinic acid (0.353 g, 3.05 mmol) and 2-chloromethyl pyridinium chloride (CMPI) (1.29 g, 5.07 mmol). It was stirred for 15 min at room temperature and then cooled to -20 °C. 1,4-diazabicyclo[2,2,2] octane (0.797 g, 7.1 mmol) was added to The mixture at the same temperature. The reaction mixture was allowed to warm up to room temperature slowly in a period of 90 min and then filtered through Celite pad, diluted with EtOAc ($2 \times 50 \text{ mL}$), washed with brine ($2 \times 50 \text{ mL}$), dried (MgSO₄), filtered and concentrated in *vacuo*. The resulting residue was purified by silica gel column chromatography (hexane: EtOAc, 6:4) to generate **24b** as colourless oil (1.13 g, 81%). The spectroscopic data was in agreement with that in the literature.⁶

C. Experimental procedure and characterization data of disaccharides

4-Methylphenyl 6-*O*-acetyl-2-azido-3-*O*-benzyl-2-deoxy-1-thio-4-*O*-(2-*O*-benzoyl-3-*O*-benzyl-6-*O*-*tert*-butyldiphenylsilyl-4-*O*-levulinyl-β-D-glucopyranosyl)-β-D-glucopyranoside (19)



Glucosyl donor 17 (0.6 g, 0.74 mmol), azidoglucosyl acceptor 90325 g, 0.74 mmol) and flame activated AW-300 MS (2.0 g) were suspended in anhydrous CH₂Cl₂(8 mL) for 1 h at room temperature under argon atmosphere. The mixture was cooled to -45 °C and then N-iodosuccinimide (NIS) (0.198 g, 0.88 mmol) and trifluoromethanesulfonic acid (TfOH) ($21 \,\mu$ L, 0.22 mmol) were added. After 1 h, the reaction mixture was warmed up to -30 °C and stirred for another 1 h. The mixture was quenched with Et₃N and filtered, diluted with $CH_2Cl_2(2 \times 30 \text{ mL})$, washed with saturated NaHCO₃(25 mL), Na₂S₂O₃(25 mL) and brine (20 mL). It was dried over MgSO4 and concentrated in *vacuo*. The crude residue was purified by silica gel column chromatography (PhCH₃: EtOAc, 20:1) to furnish **19** (0.634 g, 76%) as yellow oil. Rf 0.48 (PhCH₃/EtOAc 9:1); ¹H NMR (600 MHz, CDCl₃): δ 8.01 (d, *J* = 1.2 Hz, 2 H), 7.69 (d, *J* = 7.2 Hz, 2 H), 7.66-7.58 (m, 4 H), 7.46 (t, J = 7.8 Hz, 2 H), 7.43-7.37 (m, 6 H), 7.35-7.33 (m, 4 H), 7.27-7.24 (m, 4 H), 7.22-7.18 (m, 3 H), 7.12 (d, J = 7.2 Hz, 2 H), 5.32 (t, J = 9.0 Hz, 1 H), 5.10 (t, J = 9.0 Hz, 1 H), 4.93 (d, J = 12.0 Hz, 1 H), 4.76 (d, J = 11.4 Hz, 1 H), 4.64 (t, J = 7.8 Hz, 1 H), 4.55 (q, J = 11.4 Hz, 2 H),4.32 (d, J = 11.4 Hz, 1 H), 4.20 (d, J = 10.8 Hz, 1 H), 4.10 (dd, J = 11.4, 4.8 Hz, 1 H), 3.82 (t, J = 9.0 Hz, 1 H), 3.77 (t, *J* = 9.6 Hz, 1 H), 3.64 (app q, *J* = 6.6 Hz, 2 H), 3.46-3.45 (m, 1 H), 3.43 (t, *J* = 9.6 Hz, 1 H), 3.30 (app dd, J = 10.2, 4.2 Hz, 1 H), 3.16 (t, J = 9.0 Hz, 1 H), 2.60-2.50 (m, 2 H), 2.34-2.31 (m, 1 H), 2.29 (s, 3 H), 2.25- 2.23 (m, 1 H), 2.10 (s, 3 H), 1.96 (s, 3 H), 0.99 (s, 9 H);¹³C NMR (150 MHz, CDCl₃): 8 205.9, 171.3, 170.5, 164.7, 138.7, 138.0, 137.9, 137.6, 135.8(3), 135.7(2), 134.3, 133.5, 133.4, 133.1, 129.6-127.5, 125.3, 100.6, 85.8, 81.8, 80.2, 76.2, 75.8, 75.0, 74.0, 73.9, 70.6, 64.6, 63.0, 62.4, 37.8, 29.8, 27.8, 26.8, 21.2, 20.8, 19.2; m/z (HRMS) calcd for C₆₃H₆₉N₃O₁₃SSiNa [M+Na]⁺: 1158.4218, found, 1158.4211.

 $\label{eq:2.2} 4-Methylphenyl-6-O-acetyl-2-azido-3-O-benzyl-2-deoxy-1-thio-4-O-(2-O-benzoyl-3-O-benzyl-4-O-levulinyl-6-O-p-methoxybenzyl-\beta-D-glucopyranosyl)-\beta-D-glucopyranoside (20)$



Glucosyl donor 18 (0.5 g, 0.716 mmol), azidoglucosyl acceptor 9 (0.317 g, 0.716 mmol) and flame activated AW-300 MS (2.0 g) were suspended in anhydrous CH₂Cl₂(8 mL) for 1 h at room temperature under argon atmosphere. It was cooled to -45 °C and then N-iodosuccinimide (NIS) (0.193 g, 0.86 mmol) and trifluoromethanesulfonic acid (TfOH) (20 µL, 0.21 mmol) were added. After 1 h, the reaction mixture was warmed up to -30 °C and stirred for another 1 h. It was guenched with Et₃N. It was filtered, diluted with $CH_2Cl_2(2 \times 30 \text{ mL})$, washed with saturated NaHCO₃(25 mL), Na₂S₂O₃(25 mL), brine (20 mL) and then dried (MgSO₄). The solvent was removed in vacuo, and the residue was purified by silica gel column chromatography (PhCH₃: EtOAc, 9:1) to afford **20** (0.531 g, 73 %) as yellow oil. Rf 0.42 (PhCH₃/EtOAc 9:1); ¹H NMR (600 MHz, CDCl₃): δ 8.01-7.99 (m, 2 H), 7.60 (d, J = 7.2 Hz, 2 H), 7.45 (t, J = 7.8 Hz, 2 H), 7.41-7.36 (m, 4 H), 7.31 (t, J = 7.2 Hz, 2 H), 7.15-7.08 (m, 5H), 7.06 (d, J = 8.4 Hz, 4 H), 6.80 (d, J = 9.0 Hz, 2 H), 5.31 (dd, J = 9.6, 7.2 Hz, 1 H), 5.12 (t, J = 9.6 Hz, 1 H), 5.00 (d, J = 10.8 Hz, 1 H), 4.72 (d, J = 10.8 Hz, 1 H), 4.56 (dd, J = 2.4, 6.0 Hz, 2 H), 4.53 (d, J = 11.4 Hz, 1 H), 4.24-4.22 (m, 1 H), 4.17 (t, J=12.0 Hz, 2 H), 4.10 (dd, J=12.0, 4.8 Hz, 1 H), 3.80 (t, J=9.6 Hz, 1 H), 3.72 (s, 3 H), 3.68 (t, J=9.6 Hz, 1 H), 3.72 (s, 3 H), 3.68 (t, J=9.6 Hz, 1 H), 3.72 (s, 3 H), 3.68 (t, J=9.6 Hz, 1 H), 3.72 (s, 3 H), 3.68 (t, J=9.6 Hz, 1 H), 3.72 (s, 3 H), 3.68 (t, J=9.6 Hz, 1 H), 3.72 (s, 3 H), 3.68 (t, J=9.6 Hz, 1 H), 3.72 (s, 3 H), 3.68 (t, J=9.6 Hz, 1 H), 3.72 (s, 3 H), 3.68 (t, J=9.6 Hz, 1 H), 3.72 (s, 3 H), 3.68 (t, J=9.6 Hz, 1 H), 3.72 (s, 3 H), 3.68 (t, J=9.6 Hz, 1 H), 3.72 (s, 3 H), 3.68 (t, J=9.6 Hz, 1 H), 3.72 (s, 3 H), 3.68 (t, J=9.6 Hz, 1 H), 3.72 (s, 3 H), 3.68 (t, J=9.6 Hz, 1 H), 3.72 (s, 3 Hz, 1 Hz, 1 Hz), 3.72 (s, 3 Hz), Hz, 1 H), 3.52 (dt, J = 9.6, 4.2 Hz, 1 H), 3.44 (dd, J = 10.8, 4.2 Hz, 1 H), 3.40 (d, J = 9.0 Hz, 1 H), 3.30 (dd, J =10.8, 5.4 Hz, 1 H), 3.29-3.26 (m, 1 H), 3.18 (t, J =10.2 Hz, 1 H), 2.79 (app d, J =1.2 Hz, 1 H), 2.55 (t, J = 6.6 Hz, 2 H), 2.35-2.33 (m, 2 H), 2.30 (s, 3 H), 2.11 (s, 3 H), 1.95 (s, 3 H); ¹³C NMR (150 MHz, CDCl₃): § 206.2, 171.6, 170.5, 164.7, 159.2, 138.8, 138.1, 137.6, 134.4, 133.5, 130.5, 130.1, 129.6-127.0, 113.7, 101.0, 85.8, 82.8, 80.0, 75.8, 74.2, 73.8, 73.7, 73.2, 71.6, 69.7, 64.7, 62.3, 55.3, 37.7, 29.8, 27.9, 21.2, 20.8; m/z (HRMS) calcd for C55H59N3O14SNa [M+Na]⁺: 1040.3615, found:1040.3632.

4-Methylphenyl 6-*O*-acetyl-2-azido-3-*O*-benzyl-2-deoxy-1-thio-4-*O*-(2-*O*-benzyl-3-*O*-benzyl-β-D-glucopyranoside (22a)



To a solution of **19** (0.634 mg, 0.55 mmol) in pyridine/AcOH (3:2) (2.5 mL) was added 1M hydrazine hydrate (NH₂NH₂ H₂O, 0.8 mL). The reaction mixture was stirred at room temperature for 2 h and the solvent was removed; the residue was diluted with H₂O and the mixture was extracted by EtOAc (2 x 25 mL). The combined organic layers were washed with saturated NaHCO₃ (25 mL), brine (25 mL), then dried (MgSO₄) and concentrated. It was further dried under vacuum for 2-3 h. The crude compound **21** was dissolved in pyridine (6 mL), and HF-Pyridine (0.7 mL) was added at 0 °C. It was stirred for overnight at room temperature under nitrogen atmosphere. The solvent was evaporated under *vacuum*. The residue was dissolved in CH₂Cl₂(50 mL) and washed with saturated NaHCO₃(25 mL), brine (25 mL), then dried (MgSO₄). The residue was purified by flash column chromatography on silica gel (toluene/EtOAc = 8:2)

to furnish diol **22a** (0.32 g, 81%) as colorless oil. $R_f 0.42$ (PhCH₃/EtOAc 8:2); ¹H NMR (600 MHz, CDCl₃): δ 8.06 (d, J = 7.8 Hz, 2 H), 7.60 (t, J = 7.8 Hz, 2 H), 7.48 (t, J = 7.8 Hz, 1 H), 7.40-7.36 (m, 6 H), 7.21-7.16 (m, 6 H), 7.10 (d, J = 7.8 Hz, 2 H), 5.26 (t, J = 8.4 Hz, 1 H), 4.96 (d, J = 10.8 Hz, 1 H), 4.80 (d, J = 10.8 Hz, 1 H), 4.69 (d, J = 11.4 Hz, 1 H), 4.58 (d, J = 12.0 Hz, 1 H), 4.48 (d, J = 8.4 Hz, 1 H), 4.42 (d, J = 10.2 Hz, 2 H), 4.12 (dd, J = 12.0, 6.0 Hz, 1 H), 3.64-3.59 (m, 4 H), 3.44 (t, J = 9.0 Hz, 1 H), 3.32 (dd, J = 12.0, 5.4 Hz, 1 H), 3.29-3.25 (m, 2 H), 3.22 (d, J = 10.2 Hz, 1 H), 2.33 (s, 3 H), 1.99 (s, 3 H);¹³C NMR (150 MHz, CDCl₃): δ 170.5, 164.9, 138.9, 138.1, 137.7, 134.4, 133.6, 129.9-127.4, 101.2, 85.7, 82.9, 82.7, 75.7, 75.6, 74.9, 73.8, 70.8, 64.5, 62.2, 21.2, 20.8; m/z (HRMS) calcd for C₄₂H₄₅N₃O₁₁SNa [M+Na]⁺: 822.2672, found, 822.2678.

4-Methylphenyl 6-*O*-acetyl-2-azido-3-*O*-benzyl-2-deoxy-1-thio-4-*O*-(methyl 2-*O*-benzoyl-3-*O*-benzyl-β-D-glucopyranosyluronate)-β-D-glucopyranoside (5a)



To a vigorously stirred solution of the diol derivative 22a (0.32 g, 0.5 mmol) in a mixture of CH₂Cl₂:H₂O (2:1, 9 mL) was added 2,2,6,6-tetramethyl-1-piperidinyloxy (TEMPO, 0.018 g, 0.15 mmol) in the presence of iodobenzene diacetate (BAIB, 0.32 g, 1.25 mmol) as the cooxidant at 0 °C. The reaction mixture was allowed to warm up to room temperature and continued to stir until a complete conversion of the starting material was observed. After 2 h, it was diluted with CH_2Cl_2 (2 × 25 mL), washed with 10% Na₂S₂O₃(2×15 mL), brine (30 mL), then dried (MgSO₄), filtered and concentrated in *vacuo*. It was co-evaporated with toluene to remove trace of waters. The resulting residue was dissolved in anhydrous DMF (6 mL) and cooled to 0 °C. Methyl iodide (CH₃I, 0.1 mL) and KHCO₃ (0.05 g, 0.48 mmol) were added to the reaction mixture under N₂ atmosphere. It was stirred for a period of 4 h allowing the reaction mixture to warm up to room temperature, slowly. After 4 h, it was diluted with EtOAc (2×25 mL), washed with water $(2 \times 20 \text{ mL})$, brine (30 mL), then dried (MgSO₄), filtered and concentrated in *vacuo*. The crude mixture was purified by silica gel column chromatography (toluene/EtOAc 8:2) to furnish 5 (0.21 g, 63 %) as colorless gum. $R_f 0.41$ (toluene/EtOAc 8:2); ¹H NMR (600 MHz, CDCl₃): δ 8.12 (d, J = 7.8 Hz, 2 H), 7.78 (t, J = 7.2 Hz, 1 H), 7.58 (t, J = 7.8 Hz, 2 H), 7.50-7.44 (m, 7 H), 7.40-7.38 (m, 2 H), 7.28-7.26 (m, 3 H), 7.18 (d, J = 7.8 Hz, 2 H), 5.40 (t, J = 8.4 Hz, 1 H), 5.20 (d, J = 11.4 Hz, 1 H), 4.88 (d, J = 12.0 Hz, 2 H), 4.82 (d, J = 11.4 Hz, 1 H), 4.77 (d, J = 7.8 Hz, 1 H), 4.38-4.36 (m, 1 H), 4.32 (d, J = 1.4 Hz, 1 H), 4.77 (d, J = 7.8 Hz, 1 H), 4.38-4.36 (m, 1 H), 4.32 (d, J = 1.4 Hz, 1 H), 4.77 (d, J = 7.8 Hz, 1 H), 4.38-4.36 (m, 1 H), 4.32 (d, J = 1.4 Hz, 1 H), 4.77 (d, J = 7.8 Hz, 1 H), 4.38-4.36 (m, 1 H), 4.32 (d, J = 1.4 Hz, 1 H), 4.77 (d, J = 7.8 Hz, 1 H), 4.38-4.36 (m, 1 H), 4.32 (d, J = 1.4 Hz, 1 H), 4.77 (d, J = 7.8 Hz, 1 H), 4.38-4.36 (m, 1 H), 4.32 (d, J = 1.4 Hz, 1 H), 4.38 (d, J = 1.4 Hz, 1 Hz, 10.2 Hz, 1 H), 4.22 (dd, J = 12.0, 4.8 Hz, 1 H), 4.14 (t, J = 9.0 Hz, 1 H), 3.90 (d, J = 9.6 Hz, 1 H), 3.81-3.77 (m, 2 H), 3.65 (s, 3 H), 3.55 (t, J = 9.0 Hz, 1 H), 3.40-3.34 (m, 2 H), 3.16 (s, 1H), 2.44 (s, 3 H), 2.05 (s, 3 H); ¹³C NMR (150 MHz, CDCl₃): δ 170.4, 169.3, 164.8, 138.9, 138.3, 137.7, 134.6, 133.5, 129.9126.6, 101.4, 85.7, 83.0, 80.9, 77.6, 76.8, 75.7, 74.7, 74.4, 73.1, 72.3, 64.6, 62.1, 52.8, 21.2, 20.8; m/z (HRMS) calcd for C₄₃H₄₅N₃O₁₂SNa [M+Na]⁺: 850.2622, found, 850.2627.



A solution of 20 (0.531 mg, 0.55 mmol) in pyridine/AcOH (3:2, 2.5 mL), 1M hydrazine hydrate (NH2NH2 H₂O, 0.8 mL) was added. The reaction mixture was stirred at room temperature for 2 h and the solvent was removed; the residue was diluted with H₂O and the mixture was extracted by EtOAc (2 x 20 mL). The combined organic layers were washed with saturated NaHCO₃(30 mL), brine (30 mL) and dried over (MgSO₄). The solvents were removed in *vacuo* and the residue was purified by flash column chromatography on silica gel (toluene/EtOAc 9:1) to furnish disaccharide 6 (0.43 g, 90%) as colorless oil. $R_{\rm f}$ 0.36 (toluene/EtOAc 9:1); ¹H NMR (600 MHz, CDCl₃): δ 8.13 (d, J = 7.8 Hz, 2 H), 7.70 (t, J = 7.8 Hz, 2 H), 7.58 (t, J = 7.8 Hz, 2 H), 7.50 (t, J = 8.4 Hz, 2 H), 7.47 (d, J = 7.8 Hz, 2 H), 7.41 (t, J = 6.6 Hz, 2 H), 7.38 (t, J = 7.2 Hz, 2 H), 7.28-7.24 (m, 5 H), 7.18 (d, J = 7.8 Hz, 2 H), 6.97 (d, J = 8.4 Hz, 2 H), 5.36 (t, J = 8.4 Hz, 1 H), 5.12 (t, J = 10.8 Hz, 1 H), 4.85 (dd, J = 10.8, 5.4 Hz, 1 H), 4.79 (d, J = 12.0 Hz, 1 H),4.66 (d, J = 7.8 Hz, 1 H), 4.36 (s, 2 H), 4.35 (app dd, J = 11.4, 1.2 Hz, 1 H), 4.32 (d, J = 10.2 Hz, 1 H),4.22 (dd, J = 12.0, 5.4 Hz, 1 H), 3.92 (s, 3 H), 3.89 (d, J = 9.0 Hz, 1 H), 3.76 (q, J = 9.0 Hz, 2 H), 3.62 (dd, *J* = 9.6, 4.8 Hz, 1 H), 3.56 (app dd, *J* = 9.6, 6.0 Hz, 1 H), 3.35-3.52 (m, 2 H), 3.39-3.36 (m, 2 H), 3.32 (t, J = 9.6 Hz, 1 H), 2.43 (s, 3H), 1.99 (s, 3H); 13 C NMR (150 MHz, CDCl₃): δ 170.5, 164.9, 159.4, 138.8, 138.2, 137.9, 134.4, 133.4, 129.6-127.6, 126.8, 113.9, 101.1, 85.6, 82.8, 81.8, 75.6, 74.6, 73.8, 73.5, 73.4, 70.7, 64.5, 62.2, 55.3, 21.2, 20.8; m/z (HRMS) calcd for C₅₀H₅₃N₃O₁₂SNa [M+Na]⁺: 942.3248, found, 942.3244.

4-Methylphenyl 6-*O*-acetyl-2-azido-3-*O*-benzyl-2-deoxy-1-thio-4-*O*-(3-*O*-benzyl-4,6-*O*-benzylidene-2-*O*-levulinyl-β-D-glucopyranosyl)-β-D-glucopyranoside (23b)



Glucosyl donor **10b** (0.5 g, 0.89 mmol), azidoglucosyl acceptor **9** (0.33 g, 0.746 mmol) and flame activated AW-300 MS (2.0 g) were suspended in anhydrous CH₂Cl₂(8 mL) for 1 h at room temperature under argon atmosphere. The mixture was cooled to -20 °C and then *N*-iodosuccinimide (NIS) (0.240 g,

1.06 mmol) and tri methylsilyl trifluoromethanesulfonic acid (TMSOTf) (16μ L, 0.09 mmol) were added. After 5 min, the reaction mixture was warmed up to 0 °C and stirred for another 10 min. It was quenched with Et₃N. The mixture was filtered, diluted with CH₂Cl₂ (2 × 30 mL), washed with saturated. NaHCO₃ (25 mL), Na₂S₂O₃(25 mL) and brine (20 mL). It was dried over MgSO4. Solvents were removed in *vacuo* and the residue was purified by silica gel column chromatography (PhCH₃: EtOAc, 9:1) to furnish **23b** (0.697 g, 89%) as yellow oil. ¹H NMR (600 MHz, CDCl₃): 7.38-7.37 (m, 4 H), 7.30-7.27 (m, 9 H), 7.23-7.21 (m, 3 H), 7.00 (d, *J* = 7.8 Hz, 3 H), 5.41 (s, 1 H), 5.00 (s, 1 H), 4.96-4.93 (m, 1 H), 4.84 (d, *J* = 8.4 Hz, 1 H), 4.78 (d, *J* = 10.2 Hz, 1 H), 4.66 (d, *J* = 10.2 Hz, 1 H), 4.56 (d, *J* = 12.0 Hz, 1 H), 4.52 (dd, *J* = 12.0, 1.8 Hz, 1 H), 4.42 (d, *J* = 7.8 Hz, 1 H), 4.26 (d, *J* = 10.2 Hz, 1 H), 4.14 (dd, *J* = 12.0, 4.2 Hz, 1 H), 4.04 (dd, *J* = 10.2, 4.8 Hz, 1 H), 3.63-3.57 (m, 3 H), 3.40 (t, *J* = 9.0 Hz, 1 H), 3.32 (t, *J* = 10.2 Hz, 1 H), 3.25-3.22 (m, 1 H), 3.18 (t, *J* = 9.6 Hz, 1 H), 2.78-2.74 (m, 1 H), 2.56 (dd, *J* = 13.2, 6.6 Hz, 1 H), 2.53-2.48 (m, 2 H), 2.27 (s, 3 H), 2.09 (s, 3 H), 2.02 (s, 3 H);¹³C NMR (150 MHz, CDCl₃): δ 206.1, 171.4, 170.5, 139.0, 138.2, 138.1, 137.0, 134.8, 129.6-126.0, 101.5, 101.2, 85.2, 82.8, 81.5, 78.8, 75.3, 74.4, 73.4, 68.4, 66.4, 64.3, 62.3, 38.0, 37.6, 29.8, 27.7, 21.2, 21.0; m/z (HRMS) calcd for C₄₇H₅₁N₃O₁₂SNa [M+Na]⁺: 904.3091, found, 904.3099.



A solution of a compound **23b** (0.672 g, 0.762 mmol) was treated with 80% AcOH at 70 °C for 3 h. It was neutralized by solid NaHCO₃, filtered and then concentrated in *vacuo*, and later co-evaporated with toluene to remove traces of water. The crude mixture was purified by silica gel column chromatography (PhCH₃: EtOAc, 7:3) to furnish **22b** (0.447 g, 79%) as colorless gum. R_f 0.3 (PhCH₃/EtOAc 7:3); The spectroscopic data was in agreement with that in the literature.^{3c}



To a vigorously stirred solution of the diol derivative **22b** (0.477 g, 0.60 mmol) in a mixture of CH₂Cl₂:H₂O (2:1, 9 mL) was added 2,2,6,6-tetramethyl-1-piperidinyloxy (TEMPO)(0.037 g, 0.24 mmol)

in the presence of iodobenzene diacetate (BAIB) (0.484 g, 1.50 mmol) as the cooxidant at 0 °C. The reaction mixture was allowed to warm up to room temperature and continued to stir until complete conversion of the starting material was observed. After 2 h, it was diluted with CH_2Cl_2 (2 × 25 mL), washed with 10 % Na₂S₂O₃ (2×15 mL), brine (30 mL), then dried (MgSO₄), filtered and concentrated in vacuo. It was co-evaporated with toluene to remove traces of water. The resulting residue was dissolved in anhydrous DMF (6 mL) and cooled to 0 °C. Methyl iodide (CH₃I, 0.1 mL) and KHCO₃ (0.07 g, 0.65 mmol) were added under N₂ atmosphere. It was stirred for a period of 2 h, allowing the reaction mixture to warm up to room temperature slowly. After 2 h, it was diluted with EtOAc (2×25 mL), washed with water (2 ×20 mL), brine (30 mL), then dried (MgSO₄), filtered and concentrated in vacuo. The crude mixture was purified by silica gel column chromatography (PhCH₃/EtOAc 9:1) to furnish 5b (0.307 g, 61%) as colorless gum. *R*_f0.37 (PhCH₃/EtOAc 9:1); ¹H NMR (600 MHz, CDCl₃): δ 7.47 – 7.42 (m, 2H), 7.36 – 7.31 (m, 5H), 7.31 – 7.23 (m, 5H), 7.13 – 7.10 (m, 2H), 5.05 (d, J = 11.2 Hz, 1H), 4.99 (dd, J = 9.5, 8.0 Hz, 1H), 4.82 (d, J = 11.8 Hz, 1H), 4.72 (d, J = 11.9 Hz, 1H), 4.69 (d, J = 11.1 Hz, 1H), 4.57 (dd, J = 11.1 Hz, 1H), 4.57 (dd, J = 11.1 Hz, 1H), 4.57 (dd, J = 11.1 Hz, 1H), 4.58 (dd, J = 11.1 Hz, 1H), 4.59 (dd, J = 1 J = 12.1, 2.1 Hz, 1H), 4.52 (d, J = 8.0 Hz, 1H), 4.33 (d, J = 10.2 Hz, 1H), 4.22 (dd, J = 12.1, 4.5 Hz, 1H), 3.93 (ddd, *J* = 9.7, 8.7, 2.4 Hz, 1H), 3.77 – 3.71 (m, 1H), 3.74-3.71 (m, 2H), 3.64 (ddd, *J* = 9.9, 4.6, 2.1 Hz, 1H), 3.56 – 3.51 (m, 1H), 3.50 (s, 3H), 3.47 (dd, J = 9.4, 8.6 Hz, 1H), 3.27 (dd, J = 10.2, 9.4 Hz, 1H), 2.83 – 2.74 (m, 1H), 2.64 – 2.53 (m, 2H), 2.35 (s, 3H), 2.34 – 2.29 (m, 1H), 2.16 (s, 3H), 2.11 (s, 3H);¹³C NMR (150 MHz, CDCl₃): δ 206.1, 171.3, 170.5, 169.2, 138.9, 138.3, 138.2, 134.6, 129.9-126.5, 101.2, 85.5, 82.9, 81.4, 77.6, 76.7, 75.6, 74.7, 74.3, 72.7, 72.0, 64.5, 62.3, 52.7, 37.6, 29.7, 27.7, 21.2, 20.9; m/z (HRMS) calcd for C₄₁H₄₇N₃O₁₃SNa [M+Na]⁺: 844.2727, found, 844.2738.

Methyl 6-O-acetyl-2-azido-3-O-benzyl-2-deoxy-4-O-(methyl 2-O-benzoyl-3-O-benzyl-α-L-idopyranosyluronate)-α-D-glucopyranoside (7a)



Iodopyranosyl donor **12** (0.748 g, 1.02 mmol), azidoglucosyl acceptor **13a** (0.3 g, 0.85 mmol) and flame activated AW-300 MS (2.0 g) were suspended in anhydrous CH₂Cl₂(10 mL) for 1 h at room temperature under argon atmosphere. The mixture was cooled to -60 °C and then *N*-iodosuccinimide (NIS, 0.345 g, 1.53 mmol), trifluoromethanesulfonic acid (TfOH, 28 μ L, 0.3 mmol) were added. After 30 min, the reaction mixture was warmed up to -30 °C and stirred for another 30 min. It was quenched with excess of Et₃N and stirred for another 1 h at room temperature. The mixture was filtered, diluted with CH₂Cl₂(2 × 40 mL), washed with saturated NaHCO₃(35 mL), Na₂S₂O₃(40 mL) and brine (40 mL). It was dried over MgSO₄ and concentrated in *vacuo*. The resulting mixture was purified by silica gel column

chromatography (PhCH₃: EtOAc, 8:2) to furnish **7a** (0.655 g, 87%) as colorless gum. R_f 0.38 (PhCH₃/EtOAc 8:2); ¹H NMR (600 MHz, CDCl₃): δ 7.97 (m, 2 H), 7.59 (t, J = 7.2 Hz, 1 H), 7.44 (t, J = 7.8 Hz, 2 H), 7.40-7.22 (m, 9 H), 7.28-7.15 (m, 1 H), 5.27 (s, 1 H), 5.16 (br s, 1 H), 4.94 (d, J = 2.4 Hz, 1 H), 4.81 (d, J = 4.8 Hz, 1 H), 4.80-4.78 (m, 2 H), 4.71-4.67 (m, 2 H), 4.43 (dd, J = 12.6, 1.8 Hz, 1 H), 4.32 (dd, J = 12.0, 4.2 Hz, 1 H), 4.00 (app dd, J = 10.2, 2.4 Hz, 1 H), 3.94-3.92 (m, 1 H), 3.90 (app t, J = 3.0 Hz, 1 H), 3.86-3.83 (m, 2 H), 3.53-3.51 (m, 1 H), 3.50 (s, 3 H), 3.43 (app s, 3 H), 2.66 (app d, J = 10.8 Hz, 1 H), 2.07 (s, 3 H);¹³C NMR (150 MHz, CDCl₃): δ 170.7, 169.7, 165.1, 137.9, 137.8, 137.5, 133.9, 129.9-125.4, 98.7, 98.2, 78.8, 75.4, 75.1, 74.7, 72.7, 69.0, 68.9, 68.1, 68.0, 63.8, 62.3, 55.5, 52.1, 20.9;m/z (HRMS) calcd for C₃₇H₄₁N₃O₁₃Na [M+Na]⁺: 758.2537, found 758.2548.

 $Methyl \ 2-azido- 3-{\it O}-benzyl- 2-deoxy- 6-{\it O}-levulinyl- 4-{\it O}-(methyl \ 2-{\it O}-benzoyl- 3-{\it O}-benzyl- \alpha-l-idopyranosyluronate)- \alpha-D-glucopyranoside (7b)$



Iodopyranosyl donor 12 (0.310 g, 0.424 mmol), azidoglucosyl acceptor 13b (0.15 g, 0.354 mmol) and flame activated AW-300 MS (1.5 g) were suspended in anhydrous CH₂Cl₂ (5 mL) for 1 h at room temperature under argon atmosphere. The mixture was cooled to -60 °C and then N-iodosuccinimide (NIS, 0.115 g, 0.5 mmol) and trifluoromethanesulfonic acid (TfOH, 16 µL, 0.17 mmol) were added. After 30 min, the reaction mixture was warmed up to -30 °C and stirred for another 30 min. It was guenched with excess of Et₃N and stirred for another 1 h in room temperature. The mixture was filtered, diluted with $CH_2Cl_2(2 \times 40 \text{ mL})$, washed with saturated NaHCO₃(35 mL), Na₂S₂O₃(40 mL) and brine (40 mL). It was dried over MgSO4 and concentrated in vacuo. The resulting mixture was purified by silica gel column chromatography (PhCH₃: EtOAc, 8:2) to furnish **7b** (0.268 g, 80%) as light-yellow gum. Rf 0.32 (PhCH₃/EtOAc 8:2); ¹H NMR (600 MHz, CDCl₃): δ 7.95 – 7.90 (m, 2H), 7.55 – 7.49 (m, 1H), 7.39 – 7.36 (m, 2H), 7.34 - 7.30 (m, 4H), 7.30 - 7.27 (m, 2H), 7.24 (dt, J = 7.5, 1.8 Hz, 1H), 7.23 - 7.20 (m, 2H), 7.36 (m, 2H), 7.34 - 7.30 (m, 2H), 7.34 - 7.34 (m, 2H), 7.34 - 7.34 (m, 2H), 7.32H), 7.18 – 7.14 (m, 1H), 5.18 (d, J = 1.5 Hz, 1H), 5.10 (s, 1H), 4.94 (s, 1H), 4.87 (d, J = 2.3 Hz, 1H), 4.75 (d, J = 2.1 Hz, 1H), 4.73 (dd, J = 5.5, 3.2 Hz, 3H), 4.62 (dd, J = 11.1, 8.1 Hz, 2H), 4.36 (dd, J = 12.4, 1.1), 4.73 (dd, J = 12.4), 4.36 (dd, J = 122.2 Hz, 1H), 4.27 (dd, J = 12.4, 3.8 Hz, 1H), 3.88 (t, J = 9.6 Hz, 1H), 3.83 (t, J = 3.0 Hz, 1H), 3.79–3.76 (m, 2H), 3.44 (s, 3H), 3.38 (d, J = 3.6 Hz, 1H), 3.37 (s, 3H), 2.65 (m, 2H), 2.60 – 2.53 (m, 1H), 2.50 (m, 1H), 2.02 (s, 3H);¹³C NMR (150 MHz, CDCl₃): δ 206.5, 177.0, 172.4, 169.7, 165.1, 137.9, 137.3, 133.7, 129.9-127.4, 98.6, 98.0, 79.3, 78.7, 75.1, 75.0, 74.8, 72.6, 69.1, 68.9, 68.1, 68.0, 63.8, 62.5, 55.5, 52.1, 37.9, 29.7, 29.6, 27.6; m/z (HRMS) calcd for C40H45N3O14Na [M+Na]⁺: 814.2799, found. 814.2792.

N-(Benzyl)-benzyloxycarbonyl-5-aminopentyl-2-azido-3-*O*-benzyl-2-deoxy-6-*O*-levulinyl-4-*O*-(methyl 2-*O*-benzoyl-3-*O*-benzyl-α-L-idopyranosyluronate)-α-D-glucopyranoside (8b)



Iodopyranosyl donor 12 (0.436 g, 0.597 mmol), azidoglucosyl acceptor 24b (0.35 g, 0.498 mmol) and flame activated AW-300 MS (1.5 g) were suspended in anhydrous CH₂Cl₂ (8 mL) for 1 h at room temperature under argon atmosphere. The mixture was cooled to -60 °C and then N-iodosuccinimide (NIS, 0.161 g, 0.72 mmol) and trifluoromethanesulfonic acid (TfOH, 20 µL, 0.238 mmol) were added. After 30 min, the reaction mixture was warmed up to -30 °C and stirred for another 30 min. It was quenched with excess of Et₃N and stirred for another 1 h in room temperature. The mixture was filtered, diluted with $CH_2Cl_2(2 \times 40 \text{ mL})$, washed with saturated NaHCO₃(35 mL), Na₂S₂O₃(35 mL) and brine (40 mL). It was dried over MgSO4 and concentrated in vacuo. The resulting mixture was purified by silica gel column chromatography (PhCH₃: EtOAc, 8:2) to furnish **8b** (0.44 g, 82%) as yellow oil. Rf 0.38(PhCH₃/EtOAc 7:3); ¹H NMR (600 MHz, CDCl₃): δ 7.91 (d, *J* = 7.5 Hz, 2H), 7.51 (t, *J* = 7.4 Hz, 2H), 7.36 (d, *J* = 15.5 Hz, 2H), 7.32-7.14 (m, 14H), 7.10 (d, J = 7.1 Hz, 4H), 5.18 (s, 1H), 5.14-5.07 (m, 4 H), 4.92-4.86 (m, 1 H), 4.77 - 4.70 (m, 2H), 4.60 (dd, J = 11.4, 2.6 Hz, 2H), 4.43 (d, J = 9.8 Hz, 2H), 4.36-4.30 (m, 1H), 4.29–4.19 (m, 1H), 3.99–3.94 (m, 2H), 3.87 (t, J = 9.4 Hz, 1H), 3.82 (t, J = 3.4 Hz, 1H), 3.78 (d, J = 10.5 Hz, 3H), 3.61-3.50 (m, 1 H), 3.42 (s, 3H), 3.36-3.30 (m, 1 H), 3.26 (dd, *J* = 10.2, 3.4 Hz, 1H), 3.21-3.14 (m, 1H), 2.73 (d, J = 10.6 Hz, 1H), 2.65 - 2.61 (m, 2H), 2.58 - 2.52 (m, 1H), 2.50 - 2.45 (m, 1H), 2.00 (s, 2H)3H), 1.53–1.45 (m, 4H), 1.29-1.17 (m, 2H); ¹³C NMR (150 MHz, CDCl₃); ¹³C NMR (151 MHz, CDCl₃) δ 206.5, 172.5, 169.7, 165.2, 156.8, 156.3, 138.0, 137.9, 137.4, 133.8, 130.0, 129.1-127.3, 125.4, 98.1, 97.7, 78.5, 75.1, 74.5, 72.6, 69.2, 68.9, 68.3, 68.1, 67.3, 63.5, 62.6, 52.2, 50.7, 50.4, 47.2, 46.3, 38.0, 29.8, 29.7, 29.1, 28.0, 28.0, 27.5, 23.4, 21.6;m/z (HRMS) calcd for C₅₉H₆₆N₄O₁₆Na [M+Na]⁺: 1109.4372, found. 1109.4398.



Iodopyranosyl donor 12 (0.406 g, 0.557 mmol), azidoglucosyl acceptor 24a (0.30 g, 0.464 mmol) and flame activated AW-300 MS (1.5 g) were suspended in anhydrous CH₂Cl₂ (8 mL) for 1 h at room temperature under argon atmosphere. The mixture was cooled to -60 °C and then N-iodosuccinimide (NIS, 0.150 g, 0.667 mmol) and trifluoromethanesulfonic acid (TfOH, 20 µL, 0.228 mmol) were added. After 30 min, the reaction mixture was warmed up to -30 °C and stirred for another 30 min. It was quenched with excess of Et₃N and stirred for another 1 h in room temperature. The mixture was filtered, diluted with $CH_2Cl_2(2 \times 40 \text{ mL})$, washed with saturated NaHCO₃(35 mL), Na₂S₂O₃(35 mL), and brine (40 mL). It was dried over MgSO₄ and concentrated in *vacuo*. The resulting mixture was purified by silica gel column chromatography (PhCH₃: EtOAc, 8:2) to furnish **8a** (0.378 g, 86%) as an oil. ¹H NMR (600 MHz, CDCl₃): δ 7.97 (d, J = 7.8 Hz, 2H), 7.58 (t, J = 7.8 Hz, 1H), 7.44 (t, J = 7.8 Hz, 3H), 7.43 – 7.25 (m, 17H), 7.18 - 7.16 (m, 2H), 5.27 (s, 1H), 5.19 - 5.17 (m, 4 H), 4.95 (s, 1 H), 4.87 - 4.83 (m, 1 H), 4.80 (d, J = 10.8Hz, 2H), 4.70–4.66 (m, 2H), 4.50 (d, J = 9.0 Hz, 2H), 4.43 (dd, J = 12.6, 1.8 Hz, 1H), 4.33 (d, J = 12.0 Hz, 1H), 4.05 (dt, J = 10.8, 3.2 Hz, 1H), 3.92 (t, J = 9.0 Hz, 1H), 3.90–3.86 (m, 2H), 3.70–3.61 (m, 1H), 3.49 (s, 3H), 3.47-3.38 (m, 1H), 3.32 (dd, J = 10.2, 3.6 Hz, 1H), 3.30-3.21 (m, 1H), 2.65 (t, J = 10.2 Hz, 1H), 2.35 (d, J = 2.4 Hz, 1H), 2.06 (s, 3H), 1.65–1.54 (m, 4H), 1.40–1.29 (m, 2H);¹³C NMR (150 MHz, CDCl₃): δ 170.7, 169.9, 165.2, 156.8, 156.3, 138.0-137.3, 133.8-133.5, 130.0, 129.9-127.9, 98.2, 97.7, 78.4, 75.4, 75.0, 74.5, 72.6, 69.1, 69.0, 68.7, 68.4, 68.3, 68.1, 67.8, 67.2, 63.4, 62.3, 52.1, 50.6, 50.3, 47.1, 46.2, 29.0, 28.2, 27.9, 27.4, 23.2, 20.8; m/z (HRMS) calcd for C₅₆H₆₂N₄O₁₅Na [M+Na]⁺: 1053.4109, found. 1053.4106.

D. Experimental procedure and characterization data of pentasaccharides

 $N-(Benzyl)-benzyloxycarbonyl-5-aminopentyl 6-O-acetyl-2-azido-3-O-benzyl-2-deoxy-4-O-{methyl 2-O-benzyl-3-O-benzyl-4-O-[6-O-acetyl-2-azido-3-O-benzyl-2-deoxy-4-O-(methyl 2-O-benzyl-3-O-benzyl-4-O-{2-azido-3,4-di-O-benzyl-6-O-tert-butyldiphenylsilyl-2-deoxy-a-D-glucopyranoside}- \beta-D-glucopyranosyluronate}-\alpha-D-glucopyranoside(25a)$



Glucosyl donor 4 (0.040 g, 0.054 mmol), azidoglucosyl acceptor 5a (0.037 g, 0.045 mmol) and flame activated AW-300 MS (0.5 g) were suspended in anhydrous CH₂Cl₂(1 mL) for 1 h at room temperature under N_2 atmosphere. The mixture was then cooled to to -45°C. NIS (0.015 g, 0.064 mmol) and TfOH (2 μ L, 0.021 mmol) were added and the reaction mixture was allowed to warm to -30°C. The disaccharide acceptor 8a (0.046 g, 0.045 mmol) in anhydrous CH₂Cl₂(1 mL) and MS AW-300 (0.1 g) were stirred for 30 min at room temperature. After consumption of donor 4 and acceptor 5a (TLC, PhCH₃/EtOAc, 20:1), the mixture was again cooled to -45° C and acceptor 8a was added to it. Then, an additional amount of NIS (0.015 g, 0.066 mmol) and TfOH (3 μ L, 0.040 mmol) was added and the reaction mixture was allowed to warm up to -25 °C. After complete consumption of the starting materials, it was quenched with saturated NaHCO3 and solid Na2S2O3. The mixture was filtered and washed with 25 mL each of the saturated NaHCO₃, saturated Na₂S₂O₃, H₂O, brine, then dried (MgSO₄) and concentrated in *vacuo*. The residue was purified by silica gel column chromatography (PhCH₃/EtOAc, 20:1) to give pentasaccharide (0.051 g, 50%) as yellow gum. R_f 0.43 (PhCH₃/EtOAc 20:1); ¹H NMR (600 MHz, CDCl₃): δ 8.04 – 7.96 (m, 6H), 7.57 (ddd, J = 18.9, 8.0, 1.4 Hz, 6 H), 7.52–7.44 (m, 4 H), 7.41–7.35 (m, 7 H), 7.33–7.29 (m, 5H), 7.28–7.25 (m, 4H), 7.24–7.19 (m, 12H), 7.14–7.06 (m, 16H), 5.47 (d, *J* = 6.0 Hz, 1H), 5.46 – 5.44 (m, 1H), 5.43 - 5.41 (m, 2H), 5.23 - 5.17 (m, 4H), 4.92 (dd, J = 10.9, 2.1 Hz, 3H), 4.88 (dd, J = 10.5, 4.5)Hz, 4H), 4.79 (d, J = 11.1 Hz, 2H), 4.74 (dd, J = 14.9, 2.7 Hz, 2H), 4.72 - 4.66 (m, 4H), 4.55 - 4.49 (m, 4H), 4.36 - 4.28 (m, 2H), 4.26 - 4.21 (m, 2H), 4.21 - 4.15 (m, 2H), 4.06 (td, J = 5.8, 5.3, 2.9 Hz, 2H), 4.01 - 3.99 (m, 1H), 3.98 (dd, J = 8.7, 2.4 Hz, 2H), 3.96 - 3.94 (m, 1H), 3.94 - 3.90 (m, 3H), 3.86 (ddd, J = 11.6, 9.9, 2.6 Hz, 2H), 3.80 - 3.74 (m, 1H), 3.73 - 3.69 (m, 1H), 3.58 - 3.51 (m, 2H), 3.45 - 3.42 (m, 1H), 3.73 - 3.69 (m, 1H), 3.58 - 3.51 (m, 2H), 3.45 - 3.42 (m, 1H), 3.58 - 3.51 (m, 2H), 3.45 - 3.42 (m, 1H), 3.58 - 3.51 (m, 2H), 3.45 - 3.42 (m, 1H), 3.58 - 3.51 (m, 2H), 3.45 - 3.42 (m, 1H), 3.58 - 3.51 (m, 2H), 3.45 - 3.42 (m, 2H), 3.58 - 3.51 (m, 2H), 3.45 - 3.42 2H), 3.37 - 3.34 (m, 1H), 3.33 - 3.29 (m, 2H), 3.26 (app s, 3H), 3.22 (dt, J = 10.4, 3.5 Hz, 1H), 2.10 (s, 3H), 2.04 (s, 3H), 1.60 – 1.37 (m, 2H), 1.29 – 1.09 (m, 4H), 0.96 (s, 9H);¹³C NMR (150 MHz, CDCl₃): δ 170.8, 170.5, 169.6, 167.6, 165.3, 164.8, 138.3-137.3, 136.0-135.7, 133.7, 133.1, 130.0-129.7, 128.9-127.5, 101.1, 98.6, 98.1, 97.9, 97.6, 82.6, 80.1, 77.9, 77.8, 76.3, 75.7, 75.3, 75.2, 75.0, 74.9, 74.7, 73.6, 72.5, 69.5, 68.9, 68.8, 68.3, 67.2, 63.6, 63.1, 62.1, 61.8, 61.6, 52.4, 51.9, 50.6, 50.3, 47.1, 46.2, 32.0, 31.5, 30.4, 29.8, 29.7, 29.4, 29.0, 27.8, 27.5, 27.0, 26.8, 23.3, 22.8, 20.9, 20.9 (2), 19.4; m/z (HRMS) calcd for C₁₂₈H₁₃₈N₁₀O₃₁Si [M+H]⁺: 2339.9377, found 2339.9319.

 $\label{eq:stable} N-(Benzyl)-benzyloxycarbonyl-5-aminopentyl 2-azido-3-O-benzyl-2-deoxy-6-O-levulinyl-4-O- \{methyl 2-O-benzyl-3-O-benzyl-4-O-[6-O-acetyl-2-azido-3-O-benzyl-2-deoxy-4-O-(methyl 2-O-benzyl-3-O-benzyl-4-O-\{2-azido-3,4-di-O-benzyl-6-O-tert-butyldiphenylsilyl-2-deoxy-\alpha-D-glucopyranoside\}-\beta-D-glucopyranosyluronate)-\alpha-D-glucopyranoside]-\alpha-L-idopyranosyluronate}-\alpha-D-glucopyranoside (25b)$


Glucosyl donor 4 (0.040 g, 0.054 mmol), azidoglucosyl acceptor 5a (0.037 g, 0.045 mmol) and flame activated AW-300 MS (0.5 g) were suspended in anhydrous CH₂Cl₂(1 mL) for 1 h at room temperature under N₂ atmosphere. The mixture was then cooled to -45° C. NIS (0.015 g, 0.064 mmol) and TfOH (2 μ L, 0.021 mmol) were added and the reaction mixture was allowed to warm to -30°C. The disaccharide acceptor **8b** (0.048 g, 0.045 mmol) in anhydrous CH₂Cl₂(1 mL) and MS AW-300 (0.1 g) were stirred for 30 min at room temperature. After consumption of donor 4 and acceptor 5a (TLC, PhCH₃/EtOAc, 20:1), the mixture was again cooled to -45°C and acceptor 8b was added to it. Then, an additional amount of NIS (0.015 g, 0.066 mmol) and TfOH (3 µL, 0.040 mmol) was added and the reaction mixture was allowed to warm up to -25 °C. After complete consumption of the starting materials, it was quenched with solid NaHCO₃ and solid Na₂S₂O₃. The mixture was filtered and washed with 25 mL of each of the saturated NaHCO3, saturated Na2S2O3, H2O, brine, then dried (MgSO4) and concentrated in vacuo. The resulting residue was purified by silica gel column chromatography (PhCH₃/EtOAc, 20:1) to give pentasaccharide (0.044 g, 42%) as colourless oil. Rf 0.37 (PhCH₃/EtOAc 20:1); ¹H NMR (600 MHz, CDCl₃): δ 8.04 - 7.98 (m, 5H), 7.61 - 7.53 (m, 7H), 7.53 - 7.45 (m, 4H), 7.43 - 7.35 (m, 4H), 7.34 - 7.19 (m, 20H), 7.18 - 7.05 (m, 20H), 5.35 (d, J = 8.6 Hz, 1H), 5.34 - 5.30 (m, 4H), 5.08 (td, J = 11.6, 10.5, 10.5)6.7 Hz, 5H), 4.87 - 4.83 (m, 1H), 4.82 (dd, J = 10.9, 2.0 Hz, 3H), 4.79 - 4.75 (m, 3H), 4.75 - 4.71 (m, 1H), 4.71 – 4.66 (m, 2H), 4.66 – 4.61 (m, 2H), 4.59 (d, J = 10.7 Hz, 2H), 4.56 (d, J = 7.9 Hz, 1H), 4.41 (d, J = 10.7 Hz, 3H), 4.15 (d, J = 14.0 Hz, 1H), 4.11 (td, J = 9.1, 2.8 Hz, 1H), 4.07 - 4.04 (m, 1H), 3.97 (t, J = 5.9 Hz, 1H), 3.90 - 3.85 (m, 2H), 3.84 (d, J = 7.7 Hz, 1H), 3.82 - 3.78 (m, 3H), 3.74 (dt, J = 11.7, 10.98 (m, 3.98 (m,3.1 Hz, 2H), 3.67 – 3.63 (m, 1H), 3.60 – 3.55 (m, 2H), 3.45 – 3.39 (m, 1H), 3.34 – 3.31 (m, 1H), 3.28 (d, J = 8.7 Hz, 3H), 3.24 (tt, J = 7.6, 3.3 Hz, 1H), 3.19 (dd, J = 9.8, 4.2 Hz, 1H), 3.14 (s, 3H), 3.10 (dd, J = 9.8, 4.2 Hz, 1H), 3.14 (s, 3H), 3.14 (s, 10.3, 3.4 Hz, 2H), 2.70 – 2.57 (m, 2H), 2.57 – 2.47 (m, 2H), 2.06 (s, 3H), 2.06 – 1.99 (m, 2H), 1.93 (s, 3H), 1.55-1.50 (m, 4H), 1.35-1.27 (m, 2H), 1.04 (s, 9H);¹³C NMR (150 MHz, CDCl₃): δ 206.5, 172.4, 170.5, 169.5, 167.6, 165.4, 164.8, 156.8, 156.2, 138.3-137.3, 136.0-135.7, 133.6-133.0, 130.0-129.7, 129.1-127.6, 125.4, 101.1, 98.5, 98.1, 97.8, 97.6, 82.5, 80.1, 77.8, 77.5, 75.7, 75.3, 75.2, 75.0, 74.9, 74.7, 73.6, 72.6, 69.4, 68.9, 68.2, 67.2, 63.6, 63.2, 62.9, 62.3, 61.8, 61.5, 52.4, 51.8, 50.6, 50.3, 47.1, 46.2, 38.0,

32.6, 29.9, 29.8, 29.0, 28.3, 27.9, 27.0, 26.8, 23.3, 22.8, 21.5, 20.9, 19.4; m/z (HRMS) calcd for $C_{131}H_{142}N_{10}O_{32}Si$ [M]⁺: 2396.9717, found 2396.9635.



Method A: Glucosyl donor 4 (0.038 g, 0.052 mmol), azidoglucosyl acceptor 6 (0.04 g, 0.043 mmol) and flame activated AW-300 MS (0.3 g) were suspended in anhydrous CH₂Cl₂(1 mL) for 1 h at room temperature under N₂ atmosphere. The mixture was then cooled to to -45°C. NIS (0.014 g, 0.062 mmol) and TfOH (3 μ L, 0.016 mmol) were added and the reaction mixture was allowed to warm to -30°C. The disaccharide acceptor 7a (0.031 g, 0.043 mmol) in anhydrous CH₂Cl₂(1 mL) and MS AW-300 (0.1 g) were stirred for 30 min at room temperature. After consumption of donor 4 (TLC, PhCH₃/EtOAc, 20:1), the mixture was again cooled to -45° C followed acceptor **7a** was added to it. Then, an additional amount of NIS (0.013 g, 0.06 mmol) and TfOH (3 µL, 0.032 mmol) was added and the reaction mixture was allowed to warm up to -25 °C. After the complete consumption of the starting materials, it was quenched with solid NaHCO₃ and solid Na₂S₂O₃. The mixture was filtered and washed with 25 mL of each of the saturated NaHCO₃, saturated Na₂S₂O₃, H₂O, brine, then dried (MgSO₄) and concentrated in *vacuo*. The resulting mixture was purified by silica gel column chromatography (PhCH₃/EtOAc, 20:1) to give pentasaccharide. It was dissolved in a mixture of CH₂Cl₂: H₂O (1:0.1 mL) and DDQ (4 mg, 0.018 mmol) was added. It was stirred for 1h at room temperature and diluted with CH₂Cl₂(20 mL), washed with H₂O (15 mL) to make the organic phase colourless. The crude 6-hydroxy derivative was re dissolved in CH₂Cl₂:H₂O (2:1, 2 mL) and was added 2,2,6,6-tetramethyl-1-piperidinyloxy (TEMPO, 1.5 mg) in the presence of iodobenzene diacetate (BAIB, 0.013 g, 0.043 mmol) as the cooxidant at 0 °C. It was stirred at room temperature for 2 h. The reaction mixture was diluted with CH₂Cl₂ (20 mL), washed with 10% Na₂S₂O₃ (15 mL) and brine, then dried (MgSO₄) and concentrated in *vacuo*. It was also co-evaporated with toluene to remove traces of water. The crude carboxyl acid was dissolved in anhydrous DMF and cooled to 0 °C. After then, methyl iodide (CH₃I, 5 µL) and KHCO₃ (2 mg) were added under N₂ atmosphere. It was stirred for a period of 4 h allowing the reaction mixture to warm up to room

temperature, slowly. After 4 h, it was diluted with EtOAc (2×15 mL), washed with water (10 mL), brine (10 mL), then dried (MgSO₄), filtered and concentrated in *vacuo*. The crude mixture was purified by silica gel column chromatography (PhCH₃/EtOAc 20:1) to furnish **3a** (0.035 g) as colourless gum in 42% yield over three steps.

Method B: Glucosyl donor 4 (0.042 g, 0.058 mmol), azidoglucosyl acceptor 5a (0.04 g, 0.048 mmol) and flame activated AW-300 MS (0.3 g) were suspended in anhydrous CH₂Cl₂(1 mL) for 1 h at room temperature under argon atmosphere. The mixture was then cooled to -45°C. NIS (0.019 g, 0.087 mmol) and 1 M TfOH in CH₂Cl₂(12 µL, 0.018 mmol) were added and the reaction was allowed to warm up to -30°C. The disaccharide acceptor 7a (0.035 g, 0.043 mmol) in anhydrous CH₂Cl₂(1 mL) and MS AW-300 (0.1 g) were stirred for 30 min at room temperature. After consumption of donor 4 and acceptor 5a (TLC, PhCH₃/EtOAc, 20:1), the mixture was again cooled to -45°C and acceptor 7a was added. Then, an additional amounts of NIS (0.017 g, 0.072 mmol) and TfOH (3 µL, 0.040 mmol) were added and the mixture was allowed to warm up to -25 °C. After complete consumption of the starting materials, it was quenched with Et₃N. The mixture was filtered and washed with 25 mL each of saturated NaHCO₃. saturated Na₂S₂O₃, H₂O, brine, then dried (MgSO₄) and concentrated in vacuo. The resulting mixture was purified by silica gel column chromatography (PhCH₃/EtOAc, 20:1) to give pentasaccharide **3a** (0.054 g, 54%) as colourless gum. R_f0.41 (PhCH₃/EtOAc 20:1); ¹H NMR (600 MHz, CDCl₃): δ 8.09-8.07 (m, 4 H), 7.65 (dd, *J* = 7.8, 1.2 Hz, 2 H), 7.62 (dd, *J* = 7.8, 1.2 Hz, 2 H), 7.57 (d, *J* = 7.2 Hz, 1 H), 7.54 (d, *J* = 7.2 Hz, 1 H), 7.46 (td, J = 7.8, 3.6 Hz, 4 H), 7.39-7.35 (m, 5 H), 7.34 (d, J = 5.4 Hz, 2 H), 7.29-7.26 (m, 10 H), 7.25-7.22 (m, 6 H), 7.21-7.17 (m, 9 H), 7.15-7.13 (m, 4 H), 5.42 (app t, J = 4.8 Hz, 1 H), 5.40 (d, J = 9.0 Hz, 1 H), 5.38 (d, J = 3.6 Hz, 1 H), 5.14 (t, J = 5.4 Hz, 1 H), 4.94 (d, J = 10.8 Hz, 1 H), 4.88 (dd, J = 10. *J* = 10.8, 2.4 Hz, 2 H), 4.86-4.83 (m, 4 H), 4.76-4.72 (m, 2 H), 4.70 (dd, *J* = 10.2, 3.0 Hz, 3 H), 4.68-4.67 (m, 2 H), 4.63 (t, J = 7.2 Hz, 2 H), 4.40 (d, J = 5.4 Hz, 1 H), 4.27 (dd, J = 10.8, 3.6 Hz, 2 H), 4.21-4.18(m, 2 H), 4.16 (app d, J = 3.6 Hz, 1 H), 4.12 (app dd, J = 12.0, 1.8 Hz, 2 H), 4.02 (t, J = 6.6 Hz, 1 H), 3.97-3.96 (m, 1 H), 3.94-3.93 (m, 1 H), 3.91 (app d, *J* = 5.4 Hz, 1 H), 3.90-3.85 (m, 3 H), 3.80 (ddd, *J* = 3.6, 6.6, 10.2 Hz, 2 H), 3.72 (q, J = 8.4 Hz, 1 H), 3.69-3.67 (m, 2 H), 3.53-3.49 (m, 2 H), 3.43 (dd, J = 9.6, 3.6 Hz, 2 H), 3.34 (s, 3 H), 3.15-3.12 (m, 1 H), 3.23 (s, 3 H), 3.18 (dd, *J* = 3.6, 10.2 Hz, 1 H), 2.35 (s, 3 H), 2.05 (s, 3 H), 1.03 (s, 9 H);¹³C NMR (150 MHz, CDCl₃): δ 170.8, 170.5, 169.6, 167.8, 165.4, 164.8, 138.2, 138.1, 138.0, 138.0, 137.9, 137.2, 134.0, 133.7, 130.0, 129.9-127.5, 125.4, 101.3, 98.6, 98.6, 98.1, 97.9, 82.6, 80.2, 78.5, 77.9, 77.8, 77.7, 76.3, 75.9, 75.5, 75.4, 75.2, 75.2 (2), 75.1, 75.1 (2), 74.9, 74.7, 74.2, 73.7, 71.5, 71.2, 70.8, 69.5, 68.9, 67.7, 63.4, 63.0, 62.1, 61.6, 55.5, 52.6, 51.9, 31.0, 29.8, 20.9, 20.8.; m/z (HRMS) calcd for C₁₀₉H₁₁₇N₉O₂₉SiNa [M+Na]⁺: 2066.7618, found 2066.7584.



Glucosyl donor 4 (0.134 g, 0.183 mmol), azidoglucosyl acceptor 5a (0.105 g, 0.127 mmol) and flame activated AW-300 MS (1.5 g) were suspended in anhydrous CH₂Cl₂(4 mL) for 1 h at room temperature under argon atmosphere. The mixture was then cooled to -45°C. NIS (0.052 g, 0.227 mmol) and TfOH (8 μ L, 0.073 mmol) were added and the reaction mixture was allowed to warm up to -30°C. The disaccharide acceptor 7b (0.100 g, 0.127 mmol) in anhydrous CH₂Cl₂(2 mL) and MS AW-300 (0.5 g) were stirred for 30 min at room temperature. After consumption of donor 4 and acceptor 5a (TLC, PhCH₃/EtOAc, 20:1), the mixture was again cooled to -45°C and acceptor **7b** was added. Then, an additional amounts of NIS (0.052 g, 0.227 mmol) and TfOH (8 µL, 0.073 mmol) were added and the reaction mixture was allowed to warm up to -25 °C. After complete consumption of the starting materials, it was quenched with Et₃N. The mixture was filtered and washed with 25 mL of each of saturated NaHCO₃, saturated Na₂S₂O₃, H₂O₃, brine, then dried (MgSO₄) and concentrated in vacuo. The resulting mixture was purified by silica gel column chromatography (PhCH₃/EtOAc, 20:1) to give pentasaccharide **3b** (0.133 g, 48%) as yellow oil. *R*_f0.36 (PhCH₃/EtOAc 9:1); ¹H NMR (600 MHz, CDCl₃): δ 8.14 (d, *J* = 7.7 Hz, 4H), 7.71 (d, *J* = 7.4 Hz, 2H), 7.68 (d, J = 7.2 Hz, 2H), 7.65 – 7.58 (m, 2H), 7.52 (q, J = 8.1 Hz, 4H), 7.43 (m, 4H), 7.41 – 7.33 (m, 10H), 7.33 – 7.28 (m, 6H), 7.27 – 7.18 (m, 16H), 5.47 – 5.45 (m, 2H), 5.44 (d, J = 3.7 Hz, 1H), 5.19 (t, J = 5.4 Hz, 1H), 4.98 (d, J = 11.0 Hz, 1H), 4.96 - 4.92 (m, 2H), 4.90 (dd, J = 6.7, 3.9 Hz, 2H), 4.88 (t, J = 6.7, 3.9 (t, A = 6.7, 3.9 (t, A = 6.7, 3.9 (t, A = 6.7, 3.9 3.1 Hz, 2H), 4.81 (d, J = 11.1 Hz, 1H), 4.78 – 4.76 (m, 2H), 4.73 (d, J = 10.1 Hz, 2H), 4.71 – 4.67 (m, 2H), 4.50 (d, J = 4.8 Hz, 1H), 4.31 - 4.26 (m, 4H), 4.25 - 4.17 (m, 3H), 4.10 (t, J = 5.8 Hz, 1H), 4.03 - 4.023.95 (m, 3H), 3.95 – 3.91 (m, 3H), 3.91 – 3.84 (m, 3H), 3.80 – 3.72 (m, 2H), 3.70 (dt, *J* = 10.0, 2.6 Hz, 1H), 3.57-3.52 (m, 1H), 3.46 – 3.43 (m, 1H), 3.43 (s, 3H), 3.41 (s, 3H), 3.39-3.35 (m, 1H), 3.27 (s, 3H), 3.22 (dd, J = 10.2, 3.8 Hz, 1H), 2.85-2.73 (m, 2H), 2.71-2.61 (m, 2H), 2.20 (s, 3H), 2.06 (s, 3H), 1.09 (s, 9H);¹³C NMR (150 MHz, CDCl₃): δ 206.4, 172.4, 170.4, 169.5, 167.6, 165.3, 164.8, 138.9-137.2, 135.9-135.6, 133.9, 133.6, 133.5, 132.9, 129.8-127.4, 125.3, 101.0, 98.5, 98.5, 98.0, 97.7, 82.5, 80.1, 78.4, 77.8, 77.7, 77.5, 75.9, 75.7, 75.2, 75.1, 75.0, 74.9, 74.8, 74.7, 74.6, 73.9, 73.5, 72.5, 70.6, 70.3, 69.4, 68.8, 63.5,

63.4, 62.8, 62.3, 61.8, 61.3, 55.4, 52.4, 51.8, 37.9, 29.9, 27.9, 26.8, 21.5, 20.8, 19.4;m/z (HRMS) calcd for C₁₁₂H₁₂₁N₉O₃₀SiNa [M+Na]⁺: 2122.7886, found 2122.7909.



Pentasaccharide 26a was prepared from 3a (0.05 g, 0.024 mmol) using the general procedure for silvl ether cleavage, followed by purification by silica gel column chromatography (PhCH₃: EtOAc = 75:25) to furnish the product (0.036 g) in 83% yield.¹H NMR (600 MHz, CDCl₃) δ 8.12 – 8.10 (m, 2H), 8.10 – 8.07 (m, 3H), 7.62 - 7.56 (m, 2H), 7.48 (td, J = 7.9, 7.5, 1.6 Hz, 5H), 7.38 - 7.36 (m, 1H), 7.35 (dd, J = 7.9, 7.5, 1.6 Hz, 5H)3.2, 1.2 Hz, 2H), 7.34 – 7.33 (m, 2H), 7.32 (d, J = 2.3 Hz, 1H), 7.31 (t, J = 1.6 Hz, 3H), 7.29 (d, J = 1.9 Hz, 2H), 7.27 (dd, J = 2.9, 1.7 Hz, 1H), 7.26 - 7.24 (m, 1H), 7.23 - 7.21 (m, 4H), 7.21 - 7.19 (m, 2H), 7.19 (d, *J* = 1.9 Hz, 3H), 7.18 (d, *J* = 1.2 Hz, 2H), 7.17 – 7.15 (m, 2H), 7.12 (dd, *J* = 7.5, 2.0 Hz, 2H), 5.45 (d, J = 5.1 Hz, 1H), 5.44 - 5.41 (m, 2H), 5.16 (t, J = 5.6 Hz, 1H), 4.95 (dd, J = 10.8, 5.8 Hz, 1H), 4.89 (d, J = 10.7 Hz, 1H), 4.88 - 4.84 (m, 2H), 4.82 (d, J = 10.4 Hz, 2H), 4.74 - 4.71 (m, 3H), 4.71 - 4.69 (m, 1H), 4.67 (dd, J = 5.4, 2.5 Hz, 2H), 4.65 (d, J = 3.7 Hz, 1H), 4.43 (d, J = 5.1 Hz, 1H), 4.31 - 4.26 (m, 1H), 4.51 - 4.26 (m, 2H), 4.51 - 4.26 (m, 2H)2H), 4.23 (dd, *J* = 12.5, 3.2 Hz, 2H), 4.18 (dd, *J* = 12.4, 2.4 Hz, 1H), 4.16 – 4.12 (m, 1H), 4.05 (td, *J* = 6.1, 2.2 Hz, 1H), 3.96 (d, J = 9.1 Hz, 2H), 3.95 - 3.90 (m, 1H), 3.90 - 3.88 (m, 1H), 3.89 - 3.87 (m, 1H), 3.86 (d, *J* = 2.2 Hz, 1H), 3.82 (dd, *J* = 10.0, 8.9 Hz, 1H), 3.77 (ddd, *J* = 11.0, 9.1, 2.2 Hz, 2H), 3.70 (dt, *J* = 10.0, 2.8 Hz, 2H), 3.65 (dd, J = 11.9, 3.9 Hz, 1H), 3.55 (d, J = 1.6 Hz, 1H), 3.54 - 3.51 (m, 1H), 3.51 (s, 3H), 3.45 (ddd, *J* = 10.2, 3.9, 2.8 Hz, 1H), 3.42 (s, 3H), 3.41 (d, *J* = 4.6 Hz, 1H), 3.39 (dd, *J* = 10.0, 3.6 Hz, 1H), 3.36 (s, 3H), 3.31 (dd, J = 10.4, 3.8 Hz, 1H), 3.22 (ddd, J = 10.1, 3.8, 2.1 Hz, 1H), 2.08 (s, 3.6 Hz, 1H), 3.6 Hz, 3.6 Hz, 3.6 Hz, 3.6 Hz, 1Hz, 3.6 Hz, 1Hz, 3.6 Hz, 1Hz, 1 3H), 2.04 (s, 3H);¹³C NMR (150 MHz, CDCl₃): δ 170.8, 170.4, 169.6, 167.9, 165.4, 164.8, 138.1, 138.0, 137.9, 137.8, 137.5, 137.2, 134.5, 134.0, 133.7, 133.7, 130.1, 129.9, 129.2-127.6, 127.1, 125.4, 101.2, 98.6, 98.5, 98.1, 97.8, 82.6, 80.1, 78.4, 77.9, 77.8, 77.6, 76.2, 75.6, 75.5, 75.4, 75.2, 75.1, 75.1, 75.0, 74.7, 74.1, 73.7, 72.3, 71.1, 70.7, 69.5, 68.9, 63.5, 63.4, 62.9, 62.1, 61.6, 61.4, 55.5, 52.9, 52.0, 21.0, 20.9; m/z (HRMS) calcd for C₉₃H₉₉N₉O₂₉SiNa [M+Na]⁺: 1828.6441, found 1828.6441.



Pentasaccharide **26b** was prepared from **3b** (0.05 g, 0.024 mmol) using the general procedure for silvl ether cleavage, and then purification by silica gel column chromatography (PhCH₃: EtOAc = 8:2) to give the product (0.037 g) in 85% yield.¹H NMR (600 MHz, CDCl₃) δ 8.11 – 8.07 (m, 4H), 7.62 – 7.53 (m, 3H), 7.48 (app td, *J* = 7.9, 4.0 Hz, 4H), 7.38 – 7.29 (m, 8H), 7.29 – 7.24 (m, 9H), 7.24 – 7.19 (m, 4H), 7.19 - 7.13 (m, 6H), 7.11 (app dd, J = 7.3, 2.1 Hz, 2H), 5.45 - 5.37 (m, 4H), 5.13 (dt, J = 12.8, 5.2 Hz, 1H), 4.95 - 4.89 (m, 1H), 4.85 - 4.79 (m, 4H), 4.73 (d, J = 3.0 Hz, 3H), 4.70 - 4.63 (m, 4H), 4.47 (d, J = 3.0 Hz, 4.0 Hz, 44.8 Hz, 1H), 4.28 (d, J = 9.0 Hz, 1H), 4.27 - 4.20 (m, 4H), 4.16 - 4.11 (m, 1H), 4.10 - 4.04 (m, 1H), 3.94(dd, *J* = 9.1, 7.6 Hz, 2H), 3.90 – 3.85 (m, 3H), 3.84 – 3.78 (m, 1H), 3.78 – 3.72 (m, 2H), 3.71 (dt, *J* = 9.8, 3.1 Hz, 1H), 3.69 - 3.59 (m, 2H), 3.53 (td, J = 10.1, 2.4 Hz, 2H), 3.50 (app s, 4H), 3.45 (ddt, J = 10.0, 3.45 (ddt, J = 106.3, 3.2 Hz, 1H), 3.38 (app s, 4H), 3.36 (s, 3H), 3.33 – 3.27 (m, 1H), 3.20 (dt, J = 10.4, 2.6 Hz, 1H), 2.81 -2.67 (m, 2H), 2.61-2.54 (m, 2H), 2.15 (d, J = 3.9 Hz, 3H), 2.04 (d, J = 1.5 Hz, 3H); ${}^{13}C$ NMR (150 MHz, CDCl₃) δ 206.5, 172.5, 170.6, 169.6, 168.0, 164.8, 138.1, 138.0, 138.0 (2), 137.3, 137.7, 137.4, 137.2, 134.0, 133.7, 130.1, 129.9, 129.3, 129.1, 129.0, 129.0 (2), 128.8, 128.7, 128.6 (2), 128.6, 128.5, 128.4, 128.3, 128.2, 128.2, 128.1, 128.1, 128.1, 128.0, 127.9, 127.9, 127.9 (2), 127.9 (3), 127.8, 127.6, 127.5, 125.4, 101.2, 98.6, 98.5, 98.1, 97.8, 82.6, 80.1, 78.5, 77.9, 77.8, 77.6, 76.0, 75.6, 75.5, 75.3, 75.2, 75.1, 75.0, 74.8, 74.0, 73.7, 72.3, 70.6, 70.30, 69.5, 68.9, 63.5, 63.5, (2), 63.0, 62.3, 61.6, 61.4, 55.5, 52.9, 51.9, 38.0, 29.9, 27.7, 21.6, 20.1; m/z (HRMS) calcd for C₉₆H₁₀₃N₉O₃₀Na [M+Na]⁺: 1884.6703, found 1884.6763.



Pentasaccharide 27a was prepared from 26a (0.025 g, 0.014 mmol) using the general procedure for Ag₂Omediated O-benzylation, and then purification by silica gel column chromatography (PhCH₃: EtOAc = 8:2) to furnish the product (0.021 g) in 82% yield. ¹H NMR (600 MHz, CDCl₃): δ 8.14 – 8.06 (m, 7H), 7.61 – 7.56 (m, 6H), 7.50 – 7.44 (m, 7H), 7.36 – 7.33 (m, 7H), 7.31 - (m, 4 H), 7.29 – 7.28 (m, 4H), 7.22 -7.19 (m, 4H), 7.20 - 7.17 (m, 4H), 7.12 - 7.10 (m, 2H), 5.47 (dd, J = 8.3, 4.5 Hz, 2H), 5.45 - 5.40 (m, 2H), 5.45 (m, 2H), 5.45 (m,1H), 5.38 (s, 1H), 5.17 (dd, J = 11.4, 5.7 Hz, 1H), 4.99 (dd, J = 10.9, 5.2 Hz, 1H), 4.87-4.84 (m, 4H), 4.82-4.77 (m, 2H), 4.76 (d, J = 6.2 Hz, 1H), 4.74 - 4.68 (m, 4H), 4.66 (t, J = 9.3 Hz, 2H), 4.60 (d, J = 12.2Hz, 1H), 4.54 - 4.50 (m, 1H), 4.47 - 4.43 (m, 1H), 4.42 (d, J = 5.2 Hz, 1H), 4.33 - 4.30 (m, 2H), 4.28 (d, J = 9.1 Hz, 1H), 4.24 (dd, J = 5.4, 3.1 Hz, 1H), 4.20 (dd, J = 14.2, 2.8 Hz, 1H), 4.18 – 4.12 (m, 1H), 4.06 -4.03 (m, 1H), 3.96 - 3.92 (m, 2H), 3.92 - 3.89 (m, 1H), 3.89 - 3.84 (m, 1H), 3.84 - 3.81 (m, 1H), 3.77 -3.74 (m, 2H), 3.73 - 3.72 (m, 1H), 3.72 - 3.68 (m, 1H), 3.62 - 3.58 (m, 1H), 3.54 (ddd, J = 10.8, 8.4, 3.54 (ddd, J = 10.8, 8.54 (ddd, J = 10.8, 8.54 (d 2.9 Hz, 1H), 3.51 – 3.47 (m, 2H), 3.45 (s, 3H), 3.43 (s, 3H), 3.39 (dd, J = 10.0, 3.6 Hz, 1H), 3.35 (s, 3H), 3.24-3.18 (m, 1H), 2.09 (s, 3H), 2.02 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 170.8, 170.5, 169.6, 167.9, 165.4, 164.8, 138.2, 138.1, 138.0, 137.9, 137.5, 133.7, 133.1, 130.1, 129.9, 129.8, 129.1, 128.9-127.7, 101.3, 98.6, 98.6, 98.1, 97.9, 82.6, 80.2, 78.4, 77.9, 77.9 (2), 77.7, 76.3, 75.6, 75.4, 75.2, 75.2, 75.1 (2), 75.1, 74.9, 74.7, 74.2, 73.7, 73.7, 71.5, 71.2, 70.7, 69.8, 69.5, 68.9, 67.7, 66.8, 63.4, 63.0, 62.2, 55.5, 52.6, 51.9, 20.9, 20.9(2); m/z (HRMS) calcd for C100H105N9O29Na [M+Na]+: 1918.6912, found 1918.6849.

$$\label{eq:linear} \begin{split} Methyl \ 2-azido-3-O-benzyl-2-deoxy-6-O-levulinyl-4-O-\{methyl \ 2-O-benzyl-3-O-benzyl-4-O-[6-O-acetyl-2-azido-3-O-benzyl-2-deoxy-4-O-(methyl \ 2-O-benzoyl-3-O-benzyl-4-O-\{2-azido-3,4,6-tri-O-benzyl-2-deoxy-\alpha-D-glucopyranoside\}-\beta-D-glucopyranosyluronate)-\alpha-D-glucopyranoside]-\alpha-L-idopyranosyluronate}-\alpha-D-glucopyranoside (27b) \end{split}$$



Pentasaccharide 27b was prepared from 26b (0.030 g, 0.016 mmol) using the general procedure for Ag₂Omediated O-benzylation, and then purification by silica gel column chromatography (PhCH₃: EtOAc = 75:25) to give the product (0.024 g) in 78% yield. ¹H NMR (600 MHz, CDCl₃): δ 8.09 (tt, J = 8.4, 1.5Hz, 5H), 7.61–7.55 (m, 3H), 7.50–7.46 (m, 5H), 7.38–7.32 (m, 6H), 7.31–7.27 (m, 12H), 7.24–7.17 (m, 9H), 7.14 (dd, J = 7.4, 2.1 Hz, 2H), 7.10 (dd, J = 7.4, 2.3 Hz, 3H), 5.47 (d, J = 3.7 Hz, 1H), 5.43–5.39 (m, 2H), 5.16–5.11 (m, 2H), 4.96 (d, *J* = 10.7 Hz, 1H), 4.88–4.83 (m, 2H), 4.84–4.81 (m, 1H), 4.80–4.75 (m, 1H), 4.73 (d, J = 2.6 Hz, 2H), 4.70 (dd, J = 10.1, 3.7 Hz, 1H), 4.68–4.63 (m, 2H), 4.61–4.57 (m, 2H), 4.52 (d, J = 10.9 Hz, 2H), 4.47–4.43 (m, 2H), 4.30–4.27 (m, 1H), 4.27–4.23 (m, 2H), 4.23–4.19 (m, 1H), 4.14 (d, J = 12.8 Hz, 1H), 4.10-4.05 (m, 2H), 3.94 (ddd, J = 8.8, 5.0, 3.3 Hz, 2H), 3.90-3.87 (m, 1H),3.86–3.83 (m, 1H), 3.83–3.80 (m, 1H), 3.80–3.75 (m, 1H), 3.75–3.73 (m, 1H), 3.72 (dd, *J* = 5.6, 3.0 Hz, 1H), 3.67 (d, J = 10.0 Hz, 1H), 3.60 (dd, J = 10.8, 2.0 Hz, 1H), 3.54 - 3.49 (m, 2H), 3.50 - 3.46 (m, 1H), 3.44 (s, 3H), 3.39 (s, 3H), 3.38 (d, *J* = 6.7 Hz, 1H), 3.36 (s, 3H), 3.35 – 3.32 (m, 1H), 3.19 (ddd, *J* = 10.2, 3.7, 1.6 Hz, 1H), 3.19 (ddd, J = 10.2, 3.7, 1.6 Hz, 1H), 2.80 - 2.69 (m, 2H), 2.67 - 2.56 (m, 2H), 2.16 (app s, 3H), 2.02 (app s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 206.5, 172.5, 170.5, 169.6, 165.4, 164.8, 138.2, 138.2 (2), 138.0, 138.0, 137.9, 137.9 (2), 137.5, 137.5 (2), 137.2, 134.0, 133.7, 130.1, 129.9-127.7, 101.3, 98.6, 98.6, 98.1, 97.9, 82.6, 80.2, 78.5, 78.0, 77.9, 77.7, 76.1, 75.6, 75.3, 75.2, 75.1, 75.1, (2), 75.0, 74.9, 74.7, 74.0, 73.8, 73.7, 71.5, 69.5, 68.9, 67.7, 63.5, 63.4, 63.0, 62.35, 61.61, 55.5, 52.6, 51.9, 38.0, 29.9, 29.8, 28.0, 22.8, 20.9; m/z (HRMS) calcd for C103H109N9O30Na [M+Na]+: 1974.7172, found 1974.7160.



Pentasaccharide **28** was prepared from **27b** (0.024 g, 0.012 mmol) using the general procedure cleavage of Lev esters, followed by purification using silica gel column chromatography (PhCH₃: EtOAc = 7:3) to furnish the product (0.019 g) in 85% yield.¹H NMR (600 MHz, CDCl₃): δ 8.08 (dd, *J* = 12.8, 7.7 Hz, 5H), 7.62–7.55 (m, 4H), 7.52–7.38 (m, 6H), 7.33 (d, *J* = 6.9 Hz, 6H), 7.31–7.26 (m, 12H), 7.25–7.22 (m, 2H), 7.21–7.15 (m, 5H), 7.15–7.07 (m, 5H), 5.45 (dd, *J* = 7.1, 4.0 Hz, 2H), 5.41 (t, *J* = 8.5 Hz, 1H), 5.14 (t, *J*

= 4.1 Hz, 1H), 4.88 (d, J = 10.6 Hz, 1H), 4.85 – 4.83 (m, 2H), 4.80 (dd, J = 10.5, 5.0 Hz, 2H), 4.77–4.70 (m, 4H), 4.69–4.62 (m, 3H), 4.62–4.58 (m, 2H), 4.51 (d, J = 11.0 Hz, 1H), 4.45 (d, J = 12.1 Hz, 1H), 4.27 (t, J = 8.9 Hz, 1H), 4.21 (app d, J = 12.3 Hz, 1H), 4.17 (app d, J = 11.8 Hz, 1H), 4.10–4.04 (m, 3H), 3.98–3.89 (m, 4H), 3.82 (app dt, J = 15.3, 9.5 Hz, 2H), 3.76–3.70 (m, 5H), 3.67 (d, J = 10.2 Hz, 1H), 3.62–3.54 (m, 3H), 3.54–3.45 (m, 3H), 3.42 (s, 3H), 3.38 (app s, 3H), 3.33 (s, 3H), 3.20 (dd, J = 10.2, 3.8 Hz, 1H), 2.03 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 170.5, 169.3, 167.9, 165.6, 164.9, 138.2, 137.9, 137.4, 137.2, 133.9, 133.8, 129.9–127.5, 101.2, 98.7, 97.9, 97.9 (2), 82.7, 80.2, 78.5, 78.0, 77.9, 77.7, 75.6, 75.3, 75.2, 75.1, 75.1, 74.9, 74.7, 73.7, 73.7 (2), 73.3, 71.5, 71.4, 70.3, 69.9, 69.6, 63.7, 63.4, 63.2, 61.6, 61.2, 55.4, 52.6, 52.0, 21.0; m/z (HRMS) calcd for C₉₈H₁₀₃N₉O₂₈Na [M+Na]⁺: 1876.6810, found 1877.6826.

 $\label{eq:constraint} \begin{array}{l} Methyl \ 2-azido-3, 6-di-{\it O}-benzyl-2-deoxy-4-{\it O}-\{methyl \ 2-{\it O}-benzyl-3-{\it O}-benzyl-4-{\it O}-[6-{\it O}-acetyl-2-azido-3-{\it O}-benzyl-2-deoxy-4-{\it O}-(methyl \ 2-{\it O}-benzyl-3-{\it O}-benzyl-4-{\it O}-\{2-azido-3,4,6-tri-{\it O}-benzyl-2-deoxy-\alpha-D-glucopyranoside\}-\beta-D-glucopyranosyluronate)-\alpha-D-glucopyranoside]-\alpha-L-idopyranosyluronate}-\alpha-D-glucopyranoside (29) \end{array}$



Pentasaccharide **29** was prepared from **28** (0.019 g, 0.010 mmol) using the general procedure for Ag₂O-mediated *O*-benzylation, followed by purification using silica gel column chromatography (PhCH₃: EtOAc = 9:1) to give the product (0.016 g) in 81% yield. ¹H NMR (600 MHz, CDCl₃): δ 8.11–8.08 (m, 2H), 8.00–7.95 (m, 3H), 7.62–7.54 (m, 3H), 7.50–7.42 (m, 5H), 7.36–7.32 (m, 5H), 7.32–7.27 (m, 10H), 7.26–7.16 (m, 18H), 7.15 (dd, *J* = 7.5, 2.0 Hz, 2H), 7.11 (dd, *J* = 7.4, 2.1 Hz, 2H), 5.47 (d, *J* = 3.9 Hz, 2H), 5.42 (dd, *J* = 9.1, 7.9 Hz, 1H), 5.15 – 5.13 (m, 1H), 4.94 – 4.87 (m, 1H), 4.86 (d, *J* = 6.8 Hz, 2H), 4.65 (d, *J* = 3.9 Hz, 1H), 4.78 (a, *J* = 6.3 Hz, 1H), 4.75 (app dd, *J* = 12.6, 2.4 Hz, 2H), 4.70 – 4.66 (m, 2H), 4.65 (d, *J* = 3.9 Hz, 1H), 4.62–4.58 (m, 1H), 4.55–4.47 (m, 2H), 4.46 (d, *J* = 12.1 Hz, 1H), 4.28 (t, *J* = 8.9 Hz, 1H), 4.22 (dd, *J* = 12.4, 3.1 Hz, 1H), 4.18–4.13 (m, 2H), 4.13–4.07 (m, 2H), 4.00–3.95 (m, 1H), 3.93 (dd, *J* = 9.2, 3.5 Hz, 2H), 3.94 (s, 1H), 3.88 – 3.86 (m, 1H), 3.86–3.77 (m, 2H), 3.77–3.72 (m, 3H), 3.69–3.62 (m, 3H), 3.62–3.57 (m, 2H), 3.54–3.51 (m, 1H), 3.28 (s, 3H), 3.27 (d, *J* = 2.8 Hz, 1H), 3.20 (dd, *J* = 10.3, 3.8 Hz, 1H), 2.01 (s, 3H);¹³C NMR (150 MHz, CDCl₃) δ 170.5, 169.2, 167.9, 165.4, 164.8,

138.1, 138.1 (2), 138.0, 137.9, 137.9 (2), 137.8, 137.5, 137.2, 133.9, 133.6, 129.9, 129.9 (2), 128.5-127.6, 101.2, 98.9, 98.7, 98.2, 97.9, 82.7, 80.2, 78.6, 77.9, 77.7, 75.9, 75.6, 75.4, 75.3, 75.2, 75.1, 74.9, 74.7, 73.7, 73.7, 73.7, 73.7, 71.5, 70.5 (2), 70.1, 69.5, 67.7, 63.5, 63.4, 63.1, 55.4, 52.6, 51.9, 20.95; m/z (HRMS) calcd for C105H109N9O28Na [M+Na]⁺: 1966.7274, found 1966.7273.



Pentasaccharide 27a and 27b were converted to 2a via saponification and O-sulfation. The resulting Osultated 2a was used as such without further purification. Pentasaccharide 1a was prepared from 2a (0.012 g) via a two-step sequences: (i) hydrogenolysis (ii) selective N-sulfation. The crude O-and N-sulfated pentasaccharide was purified by a column of Sephadex G-25 using water as eluent. The appropriate fractions were concentrated and passed through a column of Dowex 50WX8-Na+ with water. The product protion was collected and lyophilized to furnish the protecting group free pentasaccharide 1a (0.0036 g, 40%). ¹H NMR (900 MHz, D₂O) δ 5.64 (d, J = 4.0 Hz, 1H), 5.45 – 5.39 (m, 1H), 5.27 – 5.18 (m, 1H), 5.05 – 4.99 (m, 1H), 4.56 (d, J = 11.4 Hz, 2H), 4.40 – 4.35 (m, 1H), 4.34 – 4.28 (m, 2H), 4.25 – 4.20 (m, 1H), 4.20 - 4.12 (m, 1H), 4.08 (m, 1H), 4.05 - 3.95 (m, 1H), 3.86 (d, J = 3.1 Hz, 2H), 3.84 - 3.76 (m, 3H), 3.76 – 3.69 (m, 6H), 3.65 – 3.57 (m, 1H), 3.49 – 3.45 (m, 2H), 3.42 (s, 3H), 3.40 (d, J = 3.1 Hz, 1H), 3.31 - 3.24 (m, 2H), 3.21 (dt, J = 10.4, 3.3 Hz, 1H), 2.92 - 2.86 (m, 1H), 2.83 - 2.77 (m, 2H); 13 C NMR $(151 \text{ MHz}, D_2 O) \delta 174.3, 174.0, 99.5, 97.8, 97.6, 97.2, 96.6, 79.3, 77.4, 76.6, 76.2, 75.9, 75.8, 74.6, 71.2, 75.9, 75.8, 75.$ 70.9, 69.5, 69.4, 69.2, 68.8, 68.6, 68.5, 68.4, 68.0, 67.1, 66.4, 65.4, 59.7, 57.6, 57.3, 57.2, 57.1, 54.9, 54.9 (2); m/z (HRMS) calcd for C₃₁H₅₂N₃O₄₆S₇ [M-Na-H]²⁻: calcd for 701.4983, found 701.4982; C31H51N3O46S7[M-2H]²⁻ calcd for 712.4892, found 712.4907; C31H49N3O46S7[M+2Na-4H]²⁻ : calcd for 734.4710, found 734.4729; C₃₁H₅₃N₃O₄₆S₇ [M-3Na]³⁻:calcd for 452.6750, found 452.6770; C₃₁H₅₂N₃O₄₆S₇[M-2Na-H]³⁻: calcd for 460.0023, found 460.0049; C₃₁H₅₁N₃O₄₆S₇[M-Na-2H]³⁻: calcd for 467.3295, found 467.3321; C₃₁H₅₁N₃O₄₆S₇ [M-3H]³⁻: calcd for 474.6568, found 474.6586.



Pentasaccharide **29** was converted to **2b** *via* saponification and *O*-sulfation. The resulting *O*-sultated **2b** was used as such without further purification. Pentasaccharide **1b** was prepared from **2b** (0.014 g) *via* a two-step sequences: (i) hydrogenolysis (ii) selective *N*-sulfation. The crude *O*-and *N*-sulfated pentasaccharide was purified by a column of Sephadex G-25 using water as eluent. The appropriate fractions were collected, concentrated and passed through a column of Dowex 50WX8⁻Na⁺ with water as eluent. The product protion was lyophilized to furnish the protecting group free pentasaccharide **1a** (0.0048 g, 47%) ¹H NMR (900 MHz, D₂O) δ 5.65 – 5.60 (m, 1H), 5.44 – 5.33 (m, 1H), 5.26 – 5.18 (m, 1H), 5.03 (d, *J* = 4.5 Hz, 1H), 5.01 – 4.92 (m, 1H), 4.57 (d, *J* = 10.3 Hz, 1H), 4.34 – 4.25 (m, 1H), 4.22 – 4.12 (m, 3H), 4.03 – 3.94 (m, 1H), 3.93 – 3.85 (m, 3H), 3.83 – 3.78 (m, 4H), 3.79 – 3.72 (m, 2H), 3.72 – 3.64 (m, 2H), 3.63 – 3.58 (m, 1H), 3.49 – 3.47 (m, 3H), 3.46 (app d, *J* = 9.3 Hz, 2H), 3.41 (app d, *J* = 10.3 Hz, 3H), 3.33 – 3.23 (m, 2H), 3.21 (dd, *J* = 10.4, 3.5 Hz, 1H), 3.05 – 2.96 (m, 2H), 2.92 – 2.87 (m, 2H), 2.79 (d, *J* = 7.3 Hz, 2H); ¹³C NMR (151 MHz, D₂O) δ 174.2, 174.0, 101.3, 99.5, 97.8, 97.2, 95.6 79.2, 77.2, 75.8, 75.6, 75.3, 74.4, 71.3, 71.2, 70.8, 70.7, 70.2, 70.0, 69.1, 68.6, 66.3, 65.2, 64.9, 60.55, 59.6, 59.5, 59.2, 57.55, 57.5(2), 57.0, 56.8, 54.8, 54.5, 54.2, 53.6; m/z (HRMS) calcd for C₃₁H₄₈N₃O₄₃S6 [M-H]⁻: calcd for 1340.9933, found 1340.9834.

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Figure 1.¹H NMR spectrum of 4-Methylphenyl 2-azido-2-deoxy-1-thio-β-D-glucopyranoside (600 MHz, CDCl₃)



Figure 2.¹³C NMR spectrum of 4-Methylphenyl 2-azido-2-deoxy-1-thio-β-D-glucopyranoside (150 MHz, CDCl₃).



S50



Figure 4.¹³C NMR spectrum of 4-Methylphenyl-2-azido-4,6-O-benzylidene-2-deoxy-1-thio-β-D-glucopyranoside (150 MHz, CDCl₃).



Figure 5.⁻¹H NMR spectrum of 4-Methylphenyl 2-azido-3-benzyl-4,6-O-benzylidene-2-deoxy-1-thio-β-D-glucopyranoside (600 MHz, CDCl₃).



Figure 6.¹³C NMR spectrum of 4-Methylphenyl 2-azido-3-benzyl-4,6-O-benzylidene-2-deoxy-1-thio-β-D-glucopyranoside (150 MHz, CDCl₃).





Figure 8.⁻¹³C NMR spectrum of 4-Methylphenyl 2-azido-3,4-di-O-benzyl-2-deoxy-6-O-tert-butyldiphenylsilyl-1-thio-β-D-glucopyranoside (150 MHz, CDCl₃). (150 MHz, CDCl₃).



Figure 9.¹H NMR spectrum of 4-Methylphenyl-2-azido-6-O-acetyl-3-O-benzyl-2-deoxy-1-thio-β-D-glucopyranoside (600 MHz, CDCl₃).



Figure 10.⁻¹³C NMR spectrum of 4-Methylphenyl-2-azido-6-O-acetyl-3-O-benzyl-2-deoxy-1-thio-β-D-glucopyranoside (150 MHz, CDCl₃).



Figure 11.⁻¹H NMR spectrum of 4-Methylphenyl-4,6-O-benzylidine-1-thio-β-D-glucopyranoside (600 MHz, CDCl₃).



Figure 12.⁻¹³C NMR spectrum of 4-Methylphenyl-4,6-O-benzylidine-1-thio-β-D-glucopyranoside (150 MHz, CDCl₃).



Figure 13.⁻¹H NMR spectrum of 4-methylphenyl 3-O-benzyl-4,6-O-benzylidine-1-thio-β-D-glucopyranoside (600 MHz, CDCl₃).



S61







Figure 17.⁻¹H NMR spectrum of 4-Methylphenyl-2-O-benzoyl-3-O-benzyl-1-thio-β-D-glucopyranoside (600 MHz, CDCl₃).







Figure 20.⁻¹H NMR spectrum of 4-Methylphenyl-2-O-benzoyl-3-O-benzyl-6-O-tert-butyldiphenylsilyl-1-thio-β-D-glucopyranoside (150 MHz, CDCl₃).







Figure 23.¹H NMR spectrum of 4-Methylphenyl-4,6-O-p-methoxybenzylidine-1-thio-β-D-glucopyranoside (600 MHz, CDCl₃).



Figure 24.¹³C NMR spectrum of 4-Methylphenyl-4,6-O-p-methoxybenzylidine-1-thio-β-D-glucopyranoside (150 MHz, CDCl₃).


Figure 25.⁻¹H NMR spectrum of 4-methylphenyl-3-O-benzyl-4,6-O-p-methoxybenzylidine-1-thio-β-D-gluco-pyranoside (600 MHz, CDCl₃).



Figure 26.¹³C NMR spectrum of 4-methylphenyl-3-O-benzyl-4,6-O-p-methoxybenzylidine-1-thio-β-D-gluco-pyranoside (150 MHz, CDCl₃).





Figure 28.¹³C NMR spectrum of 4-Methylphenyl-2-O-benzoyl-3-O-benzyl-4,6-p-methoxybenzylidine-1-thio-β-D-glucopyranoside (150 MHz, CDCl₃).



 $Figure \ 29.^{-1}H \ NMR \ spectrum \ of \ 4-Methyl phenyl-2-O-benzoyl-3-O-benzyl-6-O-p-methoxybenzyl-1-thio-\beta-D-glucopyranoside \ (600 \ MHz, \ CDCl_3).$



Figure 30.¹³C NMR spectrum of 4-Methylphenyl-2-O-benzoyl-3-O-benzyl-6-O-p-methoxybenzyl1-thio-β-D-glucopyranoside (150 MHz, CDCl₃).



Figure 31. ¹H NMR spectrum of 4-Methylphenyl-2-O-benzoyl-3-O-benzyl-4-O-levulinyl-6-O-p-methoxybenzyl-1-thio- β -D-glucopyranoside (600 MHz, CDCl₃).



Figure 32.⁻¹³C NMR spectrum of 4-Methylphenyl-2-O-benzoyl-3-O-benzyl-4-O-levulinyl-6-O-p-methoxybenzyl-1-thio-β-D-glucopyranoside (150 MHz, CDCl₃).



CDCl₃).



Figure 34.⁻¹³C NMR spectrum of 4-Methylphenyl-2-O-levulinyl-3-O-benzyl-4-O-levulinyl-6-O-p-methoxybenzyl-1-thio-β-D-glucopyranoside (150 MHz, CDCl₃).



Figure 35.⁻¹H NMR spectrum of 4-Methylphenyl 3-O-benzyl-4,6-O-benzylidine-1-thio-α-L-idopyranoside (600 MHz, CDCl₃).



Figure 36.¹³C NMR spectrum of 4-Methylphenyl 3-O-benzyl-4,6-O-benzylidine-1-thio-α-L-idopyranoside (150 MHz, CDCl₃).



Figure 37.⁻¹H NMR spectrum of 4-Methylphenyl 2-O-benzoyl-3-O-benzyl-4,6-O-benzylidene-1-thio-α-L-idopyranoside (600 MHz, CDCl₃).



Figure 38.⁻¹³C NMR spectrum of 4-Methylphenyl 2-O-benzoyl-3-O-benzyl-4,6-O-benzylidene-1-thio-α-L-idopyranoside (150 MHz, CDCl₃).



Figure 39.¹H NMR spectrum of 4-Methylphenyl 2-O-benzoyl-3-O-benzyl-1-thio-α-L-idopyranoside (600 MHz, CDCl₃).



Figure 40.⁻¹³C NMR spectrum of 4-Methylphenyl 2-O-benzoyl-3-O-benzyl-1-thio-α-L-idopyranoside (150 MHz, CDCl₃).



Figure 41.⁻¹H NMR spectrum of Methyl p-methylphenyl-2-O-benzoyl-3-O-benzyl-1-thio-α-L-idopyranosyl uronate (600 MHz, CDCl₃).



Figure 42.¹³C NMR spectrum of Methyl p-methylphenyl-2-O-benzoyl-3-O-benzyl-1-thio-α-L-idopyranosyl uronate (600 MHz, CDCl₃).



sd-IV-13



Figure 43.⁻¹H NMR spectrum of Methyl p-methylphenyl-2-O-benzoyl-3-O-benzyl-4-O-(9-fluorenylmethoxycarbonyl)-1-thio-α-L-idopyranosyl uronate (600 MHz, CDCl₃).



 $\label{eq:Figure 44.} Figure 44.^{\cdot 13}C \, \text{NMR spectrum of Methyl} \quad p-methylphenyl-2-O-benzoyl-3-O-benzyl-4-O-(9-fluorenylmethoxycarbonyl)-1-thio-α-L-idopyranosyl uronate} (150 \, \text{MHz}, \text{CDCl}_3).$



Figure 45.⁻¹H NMR spectrum of Methyl-2-azido-4,6-O-benzylidene-2-deoxy-D-glucopyranoside (600 MHz, CDCl₃).



Figure 46.¹³C NMR spectrum of Methyl-2-azido-4,6-O-benzylidene-2-deoxy-D-glucopyranoside (150 MHz, CDCl₃).





Figure 48.⁻¹³C NMR spectrum of Methyl-2-azido-3-O-acetyl-4,6-O-benzylidene-2-deoxy-α-D-glucopyranoside (150 MHz, CDCl₃).





Figure 49.⁻¹H NMR spectrum of Methyl-2-azido-3-O-acetyl-4,6-O-benzylidene-2-deoxy-β-D-glucopyranoside (600 MHz, CDCl₃).



S97



Figure 51.⁻¹H NMR spectrum of Methyl-2-azido-3-O-benzyl-4,6-O-benzylidene-2-deoxy-α-D-glucopyranoside (600 MHz, CDCl₃).



 $Figure \ 52.^{\cdot 13}C \ NMR \ spectrum \ of \ Methyl-2-azido-3-O-benzyl-4, 6-O-benzylidene-2-deoxy-\alpha-D-glucopyranoside \ (150 \ MHz, \ CDCl_3).$



Figure 53. ¹H NMR spectrum of Methyl-2-azido-6-O-acetyl-3-O-benzyl-2-deoxy-α-D-glucopyranoside (600 MHz, CDCl₃).



 $Figure \ 54. \ ^{13}C \ NMR \ spectrum \ of \ Methyl-2-azido-6-O-acetyl-3-O-benzyl-2-deoxy-\alpha-D-glucopyranoside \ (150 \ MHz, \ CDCl_3).$







Figure 57.¹H NMR spectrum of S26 (600 MHz, CDCl₃).



Figure 58.¹³C NMR spectrum of S26 (150 MHz, CDCl₃).



Figure 59. 1 H NMR spectrum of S31 (600 MHz, CDCl₃).



S107




Figure 62.⁻¹³C NMR spectrum of 4-Methylphenyl 6-O-acetyl-2-azido-3-O-benzyl-2-deoxy-1-thio-4-O-(2-O-benzoyl-3-O-benzyl-6-O-tert-butyldiphenylsilyl-4-O-levunlinyl-β-D-glucopyranosyl)-β-D-glucopyranoside (150 MHz, CDCl₃).



Figure 63. ¹H NMR spectrum of 4-Methylphenyl-6-O-acetyl-2-azido-3-O-benzyl-2-deoxy-1-thio-4-O-(2-O-benzoyl-3-O-benzyl-4-O-levunlinyl-6-O-p-methoxybenzyl-β-D-glucopyranosyl)-β-D-glucopyranoside (600 MHz, CDCl₃).



Figure 64. ¹³C NMR spectrum of 4-Methylphenyl-6-O-acetyl-2-azido-3-O-benzyl-2-deoxy-1-thio-4-O-(2-O-benzoyl-3-O-benzyl-4-O-levunlinyl-6-O-p-methoxybenzyl-β-D-glucopyranosyl)-β-D-glucopyranoside (150 MHz, CDCl₃).



sd-IV-47 rredo





 $\label{eq:Figure 65.1} Figure \ 65.^{1} H \ NMR \ spectrum \ of \ 4-Methylphenyl \ 6-O-acetyl-2-azido-3-O-benzyl-2-deoxy-1-thio-4-O-(2-O-benzyl-3-O-benzyl-\beta-D-glucopyranosyl)-\beta-D-glucopyranoside \ (600 \ MHz, \ CDCl_3).$



Figure 66.⁻¹³C NMR spectrum of 4-Methylphenyl 6-O-acetyl-2-azido-3-O-benzyl-2-deoxy-1-thio-4-O-(2-O-benzyl-3-O-benzyl-β-D-glucopyranosyl)-β-D-glucopyranoside (150 MHz, CDCl₃).







 $\label{eq:sector} Figure \ 69.^{1} H \ NMR \ spectrum \ of \ 4-Methylphenyl-6-O-acetyl-2-azido-3-O-benzyl-2-deoxy-1-thio-4-O-(2-O-benzyl-3-O-benzyl-6-O-p-methoxybenzyl-\beta-D-glucopyranosyl)-\beta-D-glucopyranoside \ (600 \ MHz, \ CDCl_3)$



 $\label{eq:Figure 70.13} Figure ~ 70.^{13}C NMR spectrum of 4-Methylphenyl-6-O-acetyl-2-azido-3-O-benzyl-2-deoxy-1-thio-4-O-(2-O-benzoyl-3-O-benzyl-6-O-p-methoxybenzyl-\beta-D-glucopyranosyl)-\beta-D-glucopyranoside (150 MHz, CDCl_3).$





 $\label{eq:Figure 72.} \ ^{13}C \ NMR \ spectrum \ of \ 4-Methylphenyl-6-O-acetyl-2-azido-3-O-benzyl-2-deoxy-1-thio-4-O-(2-O-levunlinyl-3-O-benzyl-4, 6-O-benzylidene-\beta-D-glucopyranosyl)-\beta-D-glucopyranoside \ (150 \ MHz, \ CDCl_3).$



glucopyranosyl)-β-D-glucopyranoside (600 MHz, CDCl₃).



glucopyranosyl)-β-D-glucopyranoside (150 MHz, CDCl₃).



Figure 75.¹H NMR spectrum of Methyl 6-O-acetyl-2-azido-3-O-benzyl-2-deoxy-4-O-(methyl-2-O-benzoyl-3-O-benzyl-α-L-idopyranosyluronate)-β-D-glucopyranoside (600 MHz, CDCl₃).



Figure 76.⁻¹³C NMR spectrum of Methyl 6-O-acetyl-2-azido-3-O-benzyl-2-deoxy-4-O-(methyl 2-O-benzoyl-3-O-benzyl-α-L-idopyranosyluronate)-β-D-glucopyranoside (150 MHz, CDCl₃).



Figure 77. ¹H NMR spectrum of Methyl 6-O-levunlinyl-2-azido-3-O-benzyl-2-deoxy-4-O-(methyl 2-O-benzoyl-3-O-benzyl-α-L-idopyranosyluronate)-β-D-glucopyranoside (600 MHz, CDCl₃).











Figure 82. 13 C NMR spectrum of N-(Benzyl)-benzyloxycarbonyl-5-aminopentyl-6-O-acetyl-2-azido-3-O-benzyl-2-deoxy-4-O-(methyl 2-O-benzoyl-3-O-benzyl-a-L-idopyranosyluronate)- α -D-glucopyranoside (150 MHz, CDCl₃).



Figure 83.¹H NMR spectrum of 25a (600 MHz, CDCl₃).



Figure 84. ¹³C NMR spectrum of 25a (150 MHz, CDCl₃).



Figure 85.¹H NMR spectrum of 25b (600 MHz, CDCl₃).



Figure 86.¹³C NMR spectrum of 25b (150 MHz, CDCl₃).





Figure 88.¹³C NMR spectrum of 3a (150 MHz, CDCl₃).









S139



Figure 93. ¹H NMR spectrum of 26b (600 MHz, CDCl₃).





S142



Figure 96. ¹³C NMR spectrum of 27a (150 MHz, CDCl₃).




S145



Figure 99.¹H NMR spectrum of 28 (600 MHz, CDCl₃).



Figure 100.¹³C NMR spectrum of 28 (150 MHz, CDCl₃).



Figure 101.⁻¹H NMR spectrum of 29 (600 MHz, CDCl₃).





Figure 103. ¹H NMR spectrum of 1a (900 MHz, D₂O).



Figure 104. ¹³C NMR spectrum of 1a (150 MHz, D₂O).



Figure 105. ¹H NMR spectrum of 1b (900 MHz, D₂O).







Figure 106. ¹³C NMR spectrum of 1b (150 MHz, D₂O).



Figure 107. COSY spectrum of 3b.







Figure 109. Expanded HSQC spectrum of 3b.



109.¹³C coupled HSQC spectrum of 3b.

Figure



f1 (ppm)



Figure 111. HSQC spectrum of 27b.









Figure 114. Expanded ¹³C coupled HSQC spectrum of 27b.



Figure 115. COSY spectrum of 25b.





Figure 116. ¹³C coupled HSQC spectrum of 25b.



Figure 117. Expanded ¹³C coupled HSQC spectrum of 25b.



Figure 118. COSY spectrum of 25a.



Figure 119. HSQC spectrum of 25a.





Figure 121. Expanded ¹³C coupled HSQC spectrum of 25a.





Figure 123. HSQC spectrum of 26a.



S173



Figure 125. Expanded ¹³C Coupled HSQC spectrum of 26a.