

Electronic Supplementary Information

F₁₀BINOL-derived Chiral Phosphoric Acid-Catalyzed Enantioselective Carbonyl-Ene Reactions: Theoretical Elucidation of Stereochemical Outcomes

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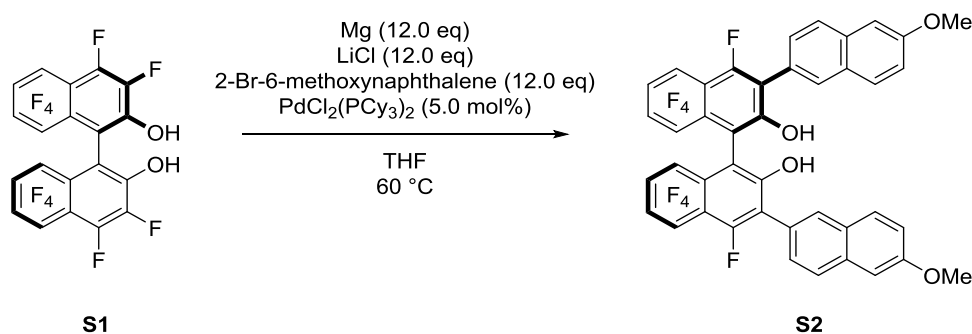
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1. General Information

All reactions were carried out under nitrogen atmosphere in flame-dried glassware. Toluene and tetrahydrofuran (THF) were supplied from KANTO Chemical Co., Inc. as “Dehydrated solvent system”. Other solvents and reagents were purchased from commercial suppliers and used without further purification. Purification of reaction products was carried out by flash column chromatography using silica gel 60 N ((Merck 40-63 μm). Analytical thin layer chromatography (TLC) was performed on Merck precoated TLC plates (silica gel 60 GF₂₅₄, 0.25 mm). ¹H NMR spectra were recorded on a JEOL ECA-600 (600 MHz) spectrometer. Chemical shifts are reported in ppm from the tetramethylsilane or solvent resonance as the internal standard (CDCl₃: 7.26 ppm, TMS: 0.00 ppm). ¹³C NMR spectra were recorded on a JEOL ECA600 (151 MHz) spectrometer with complete proton decoupling. Chemical shifts are reported in ppm from the solvent resonance as the internal standard (CDCl₃ 77.0 ppm). ³¹P NMR spectra were recorded on JEOL ECA-600 (243 MHz) spectrometer with complete proton decoupling. Chemical shifts are reported in ppm from the PPh₃ (-6 ppm) resonance as the external standard. ¹⁹F NMR spectra were recorded on JEOL ECA-600 (565 MHz) spectrometer with complete proton decoupling. Chemical shifts are reported in ppm from the C₆H₅CF₃ (-67.2 ppm) resonance as the external standard. Optical rotations were measured on a Jasco P-1020 digital polarimeter with a sodium lamp and reported as follows; [α]^{T °C}_D (c = g/100 mL, solvent, % ee). Infrared spectra were recorded on a Jasco FT/IR-4100 spectrometer. Chiral stationary phase HPLC analysis was performed on a Jasco LC-2000 Plus Series system with DACIEL chiral analytical column (4.6 mm Φ * 250 mm length). Mass spectra analysis using ESI ionization method was performed on a Bruker Daltonics solariX 9.4T spectrometer at the Research and Analytical Center for Giant Molecules, Graduate School of Science, Tohoku University.

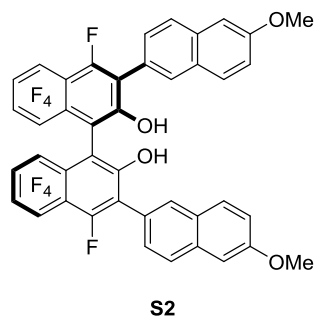
2. Preparation of Catalysts

2.1 Preparation of S2



In a flame dried, nitrogen-flushed three-necked round-bottom flask, equipped with a reflux condenser and two dropping funnels, magnesium (145.8 mg, 6.0 mmol) and LiCl (254.3 mg, 6.0 mmol) was suspended in THF (2.0 ml). A small amount of 2-bromo-6-methoxynaphthalene (1.42 g, 6.0 mmol) in THF (5.0 ml) was added to the mixture. The reaction was started by short heating to reflux. In order to keep the solution at reflux, further 2-Br-6-methoxynaphthalene solution was added slowly. After complete addition of the 2-Br-6-methoxynaphthalene, the mixture was stirred for 30 minutes at room temperature. To a mixture of PdCl₂(PCy₃)₂ (18.5 mg, 0.025 mmol) and **S1** (251 mg, 0.50 mmol) in THF (1.5 ml) was added the 6-methoxy-2-naphthylmagnesium bromide solution at -78 °C under argon atmosphere. After evacuated and refilled with argon twice, the reaction mixture was stirred at 60 °C for 36 h. After the resulting mixture was cooled to room temperature, 3.0 M HCl aq. (5.0 mL) was added. The resulting mixture was extracted with Et₂O. The combined

organic layer was dried over Na₂SO₄, and concentrated under reduced pressure after filtration. The residual crude was purified by silica gel column chromatography (Hexane/AcOEt = 5/1) to afford **S2** (317.2 mg, 0.40 mmol, 80%) as a white solid.



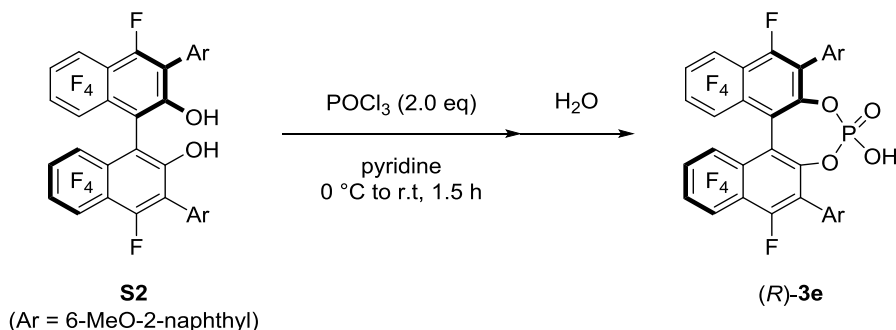
S2

(R)-4,4',5,5',6,6',7,7',8,8'-Decafluoro-3,3'-bis(6-methoxy-2-naphthyl)-1,1'-binaphthyl-2,2'-diol ((R)-3,3'-(6-MeO-2-naphthyl)₂-F₁₀BINOL) (S2**)**

S2: White solid; (317.2mg, 0.4 mmol, 80% yield); ¹H NMR (CDCl₃, 600 MHz) δ 7.93 (s, 2H), 7.90 (d, *J* = 8.4 Hz, 2H), 7.78 (d, *J* = 9.0 Hz, 2H), 7.55 (d, *J* = 7.8 Hz, 2H), 7.22–7.18 (m, 4H), 5.60 (s, 2H), 3.94 (s, 6H); ¹³C NMR (CDCl₃, 151 MHz) δ 158.7, 154.4 (d, *J* = 259 Hz), 150.9, 142.5 (dm, *J* = 253 Hz), 142.1 (dm, *J* = 246 Hz), 139.9 (dm, *J* = 254 Hz), 136.9 (dm, *J* = 251 Hz), 134.8, 129.7, 129.6, 128.8, 128.1, 127.9, 123.3, 120.2, 119.8, 118.2 (d, *J* = 17 Hz), 107.3, 106.8 (t, *J* = 8.8 Hz), 105.7, 55.4; ¹⁹F NMR (CDCl₃, 565 MHz) δ -115.1 (d, *J* = 74.0 Hz, 2F), -144.4 (dt, *J* = 74.0 Hz, 16.4 Hz, 2F), -146.6 (t, *J* = 15.5 Hz, 2F), -155.2 (t, *J* = 18.9 Hz, 2F), -160.7 (t, *J* = 19.2 Hz, 2F); IR (ATR) 3518, 3006, 2941, 2908, 2842, 1665, 1629, 1604, 1520, 1484, 1466, 1441, 1407, 1386, 1349, 1334, 1272, 1219, 1197, 1186, 1163, 1127, 1110, 1087, 1021, 973, 950, 900, 855 cm⁻¹; HRMS (ESI) calcd. for C₄₂H₁₉F₁₀O₄ [M-H]⁻: 777.1124, found: 777.1128; [α]_D²² = -76.9 (*c* = 1.00, CHCl₃, >99% ee).

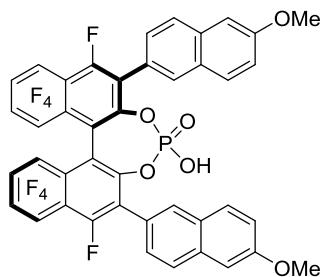
2.2 Preparation of 3d-3f

Representative procedure



To a solution of **S2** (185 mg, 0.24 mmol) in pyridine (2.4 ml) was added POCl₃ (73.6 mg, 0.48 mmol) dropwise at 0 °C. The reaction mixture was stirred at room temperature for 1.5 h. The resulting mixture was quenched with H₂O at 0 °C, and the product was extracted with Et₂O. The combined organic layer was washed with 6.0 M HCl aq., dried over Na₂SO₄ and evaporated. The residual crude was purified by silica gel column chromatography (hexane/AcOEt = 1:1 to AcOEt) to afford the desired product as a salt. Acidification in Et₂O with 3.0 M HCl aq. followed by drying under reduced pressure afforded the desired product as a free acid (**(R)-3e**) (168 mg, 0.20 mmol, 84% yield) as a white solid.

(R)-3,3'-(6-MeO-2-naphthyl)₂-F₁₀BINOL-derived phosphoric acid ((R)-3e)

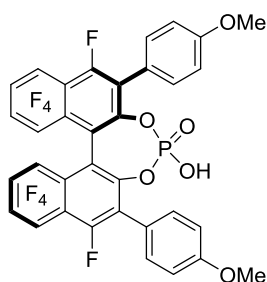


(R)-3e

(R)-3e: White solid; (168 mg, 0.20 mmol, 84% yield); ¹H NMR (CDCl₃, 600 MHz) δ 7.77 (brm, 2H), 7.53 (brm, 4H), 7.29 (brm, 2H), 6.91 (brm, 2H), 6.76 (brm, 2H), 3.77 (s, 6H); ¹³C NMR (CDCl₃, 151 MHz) δ 158.1, 154.9 (d, *J* = 262 Hz), 145.3, 142.8 (dm, *J* = 244 Hz), 142.1 (dm, *J* = 252 Hz), 140.2 (dm, *J* = 246 Hz), 138.5 (dm, *J* = 256 Hz), 134.3, 130.1, 129.7, 128.4, 127.9 (d, *J* = 15.9 Hz), 126.7, 123.1, 122.1 (d, *J* = 16 Hz), 119.6, 118.8, 114.6, 109.5 (m), 105.4 (m), 55.2; ¹⁹F NMR (CDCl₃, 565 MHz) δ -111.3 (m, 2F), -140.3 (s, 2F), -142.4 (d, *J* = 76.3 Hz, d), -152.7 (d, *J* = 14.1 Hz, 2F), -155.4 (d, *J* = 14.1 Hz, 2F); ³¹P NMR (CDCl₃, 243 MHz) δ 2.61 (s); IR (ATR); 3005, 2941, 2844, 1663, 1632, 1604, 1523, 1505, 1484, 1441, 1380, 1334, 1262, 1222, 1181, 1164, 1126, 1115, 1092, 1024, 973, 950, 854 cm⁻¹; HRMS (ESI) calcd. for C₄₂H₁₈F₁₀O₆P [M-H]⁻: 839.0681, found: 839.0686; [α]_D²¹ = -154.34 (*c* = 1.00, CHCl₃, >99% ee).

(R)-3,3'-(4-MeOC₆H₄)₂-F₁₀BINOL-derived phosphoric acid ((R)-3b)

(R)-3b was synthesized according to the same procedure of (R)-3e

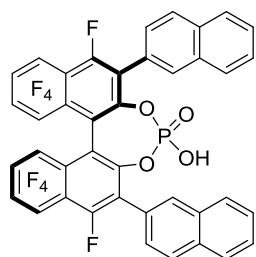


(R)-3b

White solid; (51.8 mg, 0.07 mmol, 92% yield); ¹H NMR (CDCl₃, 600 MHz) δ 8.90 (brs, 1H), 7.36 (d, *J* = 8.4 Hz, 4H), 6.82 (d, *J* = 8.4 Hz, 4H), 3.65 (s, 6H); ¹³C NMR (CDCl₃, 151 MHz) δ 159.9, 154.9 (d, *J* = 262 Hz), 145.4, 142.7 (dm, *J* = 253 Hz), 142.1 (dm, *J* = 253 Hz), 140.1 (dm, *J* = 253 Hz), 138.5 (dm, *J* = 254 Hz), 131.8, 121.7 (d, *J* = 17.4 Hz), 120.1, 119.5, 114.6, 113.9, 109.6 (m), 55.1; ¹⁹F NMR (CDCl₃, 565 MHz) δ -111.6 (d, *J* = 76.3 Hz, 2F), -140.3 (t, *J* = 17.0 Hz, 2F), -142.4 (dt, *J* = 79.1 Hz, 15.5 Hz, 2F), -152.9 (t, *J* = 16.4, 2F), -155.5 (t, *J* = 17.0, 2F); ³¹P NMR (CDCl₃, 243 MHz) δ 2.68 (s); IR (ATR) 2939, 2841, 1664, 1609, 1519, 1478, 1380, 1295, 1252, 1209, 1178, 1120, 1096, 1021, 1011, 944, 876 cm⁻¹; HRMS (ESI) calcd. for C₃₄H₁₄F₁₀O₆P [M-H]⁻: 739.0368, found: 739.0373; [α]_D²⁰ = -117.2 (*c* = 1.00, CHCl₃, >99% ee).

(R)-3,3'-(2-naphthyl)₂-F₁₀BINOL-derived phosphoric acid ((R)-3c)

(R)-3c was synthesized according to the same procedure of (R)-3e

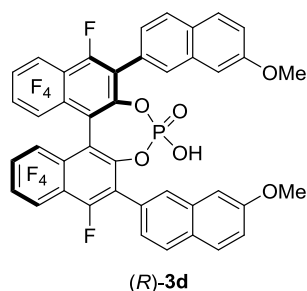


(R)-3c

White solid; (139 mg, 0.18 mmol, 89% yield); ¹H NMR (CDCl₃, 600 MHz) δ 7.81 (s, 2H), 7.67 (d, *J* = 8.4 Hz, 2H), 7.62 (d, *J* = 9.0 Hz, 2H), 7.52 (d, *J* = 7.8 Hz, 2H), 7.29 (d, *J* = 8.4 Hz, 2H), 7.22 (t, *J* = 7.2 Hz, 2H), 7.15 (t, *J* = 7.2 Hz, 2H), 5.46 (brs, 1H); ¹³C NMR (CDCl₃, 151 MHz) δ 155.0 (d, *J* = 265 Hz), 145.3, 142.8 (dm, *J* = 247 Hz), 142.2 (dm, *J* = 252 Hz), 140.3 (dm, *J* = 256 Hz), 138.5 (dm, *J* = 253 Hz), 133.0, 132.8, 130.3, 128.1, 127.8, 127.42, 127.38, 126.4, 126.0, 125.6, 122.0 (d, *J* = 17.4 Hz), 119.8, 114.7, 109.6 (m); ¹⁹F NMR (CDCl₃, 565 MHz) δ -111.8 (d, *J* = 76.3 Hz, 2F), -140.2 (t, *J* = 15.5 Hz, 2F), -141.8 (dt, *J* = 15.5, 78.5 Hz, 2F), -152.7 (t, *J* = 17.9 Hz, 2F), -155.5 (t, *J* = 17.8 Hz, 2F); ³¹P NMR (CDCl₃, 243 MHz) δ 1.23 (s); IR (ATR) 3058, 1664, 1618, 1600, 1523, 1482, 1456, 1381, 1337, 1277, 1208, 1179, 1117, 1092, 1025, 970, 934, 889 cm⁻¹; HRMS (ESI) calcd. for C₄₀H₁₄F₁₀O₄P [M-H]⁻: 779.0470, found: 779.0475; [α]_D²⁵ = -121.29 (*c* = 1.00, CHCl₃, >99% ee).

(R)-3,3'-(7-MeO-2-naphthyl)₂-F₁₀BINOL-derived phosphoric acid ((R)-3d)

(R)-3d was synthesized according to the same procedure of (R)-3e

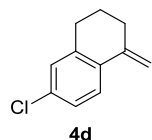


White solid; (84 mg, 0.10mmol, 79% yield); ¹H NMR (CDCl₃, 600 MHz) δ 10.75 (1H, bs), 7.75 (2H, bs), 7.50-7.40 (4H, m), 7.14 (2H, bs), 6.92 (2H, bs), 6.67 (2H, bs), 3.60 (6H, s); ¹³C NMR (CDCl₃, 151 MHz) δ 157.6, 155.0 (d, *J* = 263 Hz), 145.2, 142.7 (dm, *J* = 253 Hz), 142.2 (dm, *J* = 263 Hz), 140.3 (dm, *J* = 254 Hz), 138.6 (dm, *J* = 251 Hz), 134.1, 129.1, 128.9, 128.6, 127.7, 126.0, 125.3, 122.1 (d, *J* = 18.7 Hz), 119.7, 119.3, 114.5, 109.6 (m), 105.9, 55.0; ¹⁹F NMR (CDCl₃, 565 MHz) δ -111.2 (d, *J* = 76.8 Hz, 2F), -140.3 (s, 2F), -142.2 (dt, *J* = 76.3 Hz, 17.0 Hz, 2F), -152.6 (t, *J* = 19.2 Hz, 2F), -155.4 (t, *J* = 17.8 Hz, 2F); ³¹P NMR (CDCl₃, 243 MHz) δ 2.94 (s); IR (ATR) ; 3006, 2939, 2842, 1663, 1631, 1603, 1522, 1479, 1380, 1331, 1248, 1217, 1176, 1117, 1093, 1023, 976, 866 cm⁻¹; HRMS (ESI) calcd. for C₄₂H₁₈F₁₀O₆P [M-H]⁻: 839.0681, found: 839.0686; [α]_D²⁴ = -107.026 (*c* = 1.00, CHCl₃, >99% ee).

3. Preparation of substrates

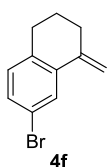
Novel 1,1-disubstituted olefins were prepared by following the literature procedure¹

6-chloro-1-methylene-1,2,3,4-tetrahydronaphthalene (4d)



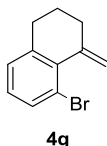
4d: ¹H NMR (CDCl₃, 600 MHz) δ 7.55 (d, *J* = 8.4 Hz, 1H), 7.12–7.06 (m, 2H), 5.44 (s, 1H), 4.96 (s, 1H), 2.81 (t, *J* = 6.0 Hz, 2H), 2.55–2.50 (m, 2H), 1.88–1.83 (m, 2H); ¹³C NMR (CDCl₃, 151 MHz) δ 142.4, 139.0, 133.2, 133.0, 128.8, 126.1, 125.6, 108.4, 32.9, 30.3, 23.4; IR (ATR) 3086, 3030, 2937, 2866, 2837, 1687, 1626, 1592, 1478, 1454, 1440, 1406, 1344, 1299, 1263, 1241, 1194, 1180, 1112, 1046, 972, 881 cm⁻¹; HRMS (APCI) calcd. for C₁₁H₁₁Cl [M]⁺: 178.0549, found: 178.0544.

7-bromo-1-methylene-1,2,3,4-tetrahydronaphthalene (4f)



4f: ¹H NMR (CDCl₃, 600 MHz) δ 7.75 (s, 1H), 7.26 (d, *J* = 8.4 Hz, 1H), 6.96 (d, *J* = 7.2 Hz, 1H), 5.45 (s, 1H), 4.99 (s, 1H), 2.77 (t, *J* = 6.2 Hz, 2H), 2.54–2.50 (m, 2H), 1.88–1.83 (m, 2H); ¹³C NMR (CDCl₃, 151 MHz) δ 142.2, 136.8, 136.1, 130.8, 130.3, 127.1, 119.6, 109.2, 32.7, 29.9, 23.4; IR (ATR) 3086, 2936, 2863, 2835, 1626, 1588, 1556, 1474, 1454, 1433, 1407, 1345, 1290, 1276, 1263, 1239, 1180, 1135, 1085, 968, 884, 867 cm⁻¹; HRMS (APCI) calcd. for C₁₁H₁₁Br [M]⁺: 222.0044, found: 222.0044.

8-bromo-1-methylene-1,2,3,4-tetrahydronaphthalene (4g)



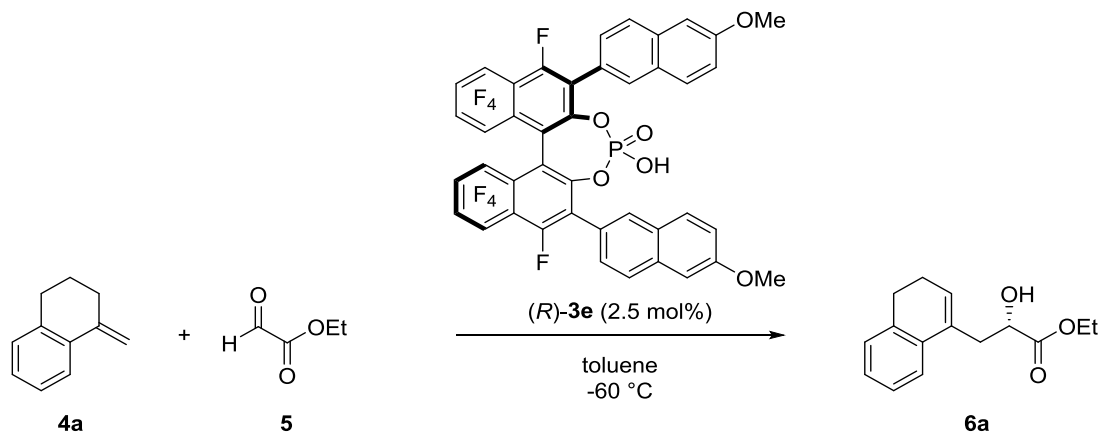
4g: ¹H NMR (CDCl₃, 600 MHz) δ 7.46 (d, *J* = 7.8 Hz, 1H), 7.04 (dd, *J* = 7.2 Hz, 1.2 Hz, 1H), 6.97 (t, *J* = 7.8 Hz, 1H), 5.65 (d, *J* = 0.6 Hz, 1H), 5.36 (q, *J* = 1.8 Hz, 1H), 2.71 (t, *J* = 6.6 Hz, 2H), 2.50 (tt, *J* = 7.2 Hz, 1.2 Hz, 2H), 1.88–1.82 (m, 2H); ¹³C NMR (CDCl₃, 151 MHz) δ 141.8, 141.6, 137.2, 131.8, 127.7, 127.1, 120.6, 116.3, 108.4, 32.4, 30.2, 23.2; IR (ATR) 3051, 2940, 2865, 2844, 1630, 1553, 1439, 1290,

¹ T. W. Liwosz, S. R. Chemler, *Chem. Eur. J.* **2013**, *19*, 12771.

1180, 1109, 968, 905 cm^{-1} ; HRMS (APCI) calcd. for $\text{C}_{11}\text{H}_{11}\text{Br} [\text{M}]^+$: 222.0044, found: 222.0045.

4. Enantioselective Carbonyl-Ene Reaction

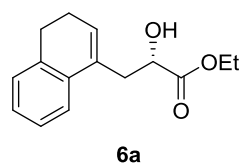
Representative procedure:



To a solution of (R) -**3e** (4.2 mg, 0.005 mmol) in toluene (1.0 mL) was added freshly distilled ethyl glyoxylate **5** (40.8 mg, 0.4 mmol) and α -methylene-tetraline **4a** (28.8 mg, 0.2 mmol) at $-60\text{ }^{\circ}\text{C}$, and the resulting solution was stirred for 3 h at this temperature. The mixture was directly passed through flash column chromatography (Hexane/AcOEt = 3/1) to give **6a** (41.4 mg, 0.17 mmol, 84% yield) as a colorless oil. The enantiomeric excess was determined by chiral stationary phase HPLC analysis.

ethyl (S)-3-(3,4-dihydronaphthalen-1-yl)-2-hydroxypropanoate (**6a**)

6a: Colorless oil: (41.4 mg, 84% yield); ^1H NMR (CDCl_3 , 600 MHz) δ 7.28 (d, $J = 7.8$ Hz, 1H), 7.22–7.18 (m, 1H),

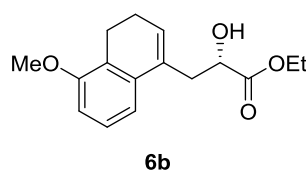


6a

7.16–7.14 (m, 2H), 5.99 (t, $J = 4.2$ Hz, 1H), 4.38–4.35 (m, 1H), 4.21–4.10 (m, 2H), 3.03 (dd, $J = 14.4$ Hz, 4.5 Hz, 1H), 2.80–2.71 (m, 3H), 2.69 (dd, $J = 6.3$ Hz, 0.9 Hz, 1H), 2.29–2.26 (m, 2H), 1.26 (dt, $J = 7.2$ Hz, 0.6 Hz, 3H); ^{13}C NMR (CDCl_3 , 151 MHz) δ 174.6, 136.7, 134.1, 131.7, 128.7, 127.7, 127.0, 126.3, 122.6, 69.5, 61.6, 38.0, 28.3, 23.2, 14.1; IR (ATR) 3474, 3058, 2980,

2934, 2885, 2830, 1732, 1488, 1447, 1369, 1268, 1208, 1096, 1025, 941, 864 cm^{-1} ; HRMS (ESI) calcd. for $\text{C}_{15}\text{H}_{18}\text{NaO}_3 [\text{M}+\text{Na}]^+$: 269.1154, found: 269.1149; HPLC analysis CHIRALPAK AS-H (Hexane/IPA = 90/10, 1.0 mL/min, 254 nm, $30\text{ }^{\circ}\text{C}$) 8.4 min (major), 11.7 min (minor), 87% ee; $[\alpha]_D^{23} = -12.56$ ($c = 1.00$, C_6H_6 , 87% ee), $[\alpha]_D^{22} = 0.979$ ($c = 1.00$, CHCl_3 , 87% ee). **Configuration Assignment**: The absolute configuration was assigned as (S) by comparison with (R) isomer.; $[\alpha]_D^{25} = 13.7$ ($c = 1.94$, C_6H_6 , 96% ee), Yuan, Y.; Zhang, X.; Ding, K. *Angew. Chem. Int. Ed.* **2003**, 42, 5478–5480.

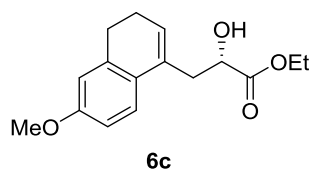
ethyl (S)-2-hydroxy-3-(5-methoxy-3,4-dihydronaphthalen-1-yl)propanoate (**6b**)



6b

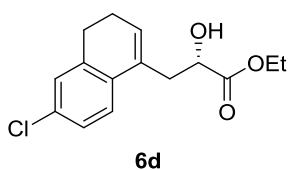
6b: Colorless oil: (42.6 mg, 77% yield); ^1H NMR (CDCl_3 , 600 MHz) δ 7.17 (t, $J = 7.8$ Hz, 1H), 6.95 (d, $J = 7.2$ Hz, 1H), 6.80 (d, $J = 8.4$ Hz, 1H), 5.99 (t, $J = 4.2$ Hz, 1H), 4.36–4.33 (m, 1H), 4.20–4.11 (m, 2H), 3.83 (s, 3H), 3.01 (dd, $J = 14.4$ Hz, 3.6 Hz, 1H), 2.83–2.77 (m, 1H), 2.74–2.67 (m, 3H), 2.24–2.22 (m, 2H), 1.26 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (CDCl_3 , 151 MHz) δ 174.6, 156.2, 135.1, 131.5, 128.9, 126.3, 124.6, 115.5, 109.7, 69.5, 61.5, 55.5,

38.5, 22.5, 19.8, 14.1; IR (ATR) 3471, 3072, 2937, 2886, 2835, 1733, 1597, 1573, 1473, 1461, 1440, 1368, 1345, 1261, 1208, 1141, 1091, 1046, 926 cm^{-1} ; HRMS (ESI) calcd. for $\text{C}_{16}\text{H}_{20}\text{NaO}_4$ $[\text{M}+\text{Na}]^+$: 299.1259, found: 299.1254; HPLC analysis CHIRALPAK AS-H (Hexane/IPA = 85/15, 1.0 mL/min, 254 nm, 30 °C) 10.2 min (major), 11.3 min (minor), 86% ee; $[\alpha]_{\text{D}}^{22} = -1.61$ ($c = 1.00$, CHCl_3 , 86% ee). **Configuration Assignment:** The absolute configuration was assigned as (S) by analogy.



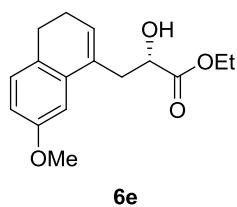
ethyl (S)-2-hydroxy-3-(6-methoxy-3,4-dihydronaphthalen-1-yl)propanoate(6c)

6c: Colorless oil: (48.1 mg, 87% yield); ^1H NMR (CDCl_3 , 600 MHz) δ 7.21 (d, $J = 9.0$ Hz, 1H), 6.74–6.70 (m, 2H), 5.86 (t, $J = 4.2$ Hz, 1H), 4.37–4.34 (m, 1H), 4.20–4.11 (m, 2H), 3.80 (s, 3H), 3.00 (dd, $J = 14.4$ Hz, 4.2 Hz, 1H), 2.76–2.65 (m, 4H), 2.28–2.24 (m, 2H), 1.27 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (CDCl_3 , 151 MHz) δ 174.5, 158.5, 138.6, 131.3, 127.2, 126.1, 123.8, 114.0, 110.8, 69.5, 61.5, 55.2, 38.2, 28.8, 23.1, 14.2; IR (ATR) 3481, 2979, 2935, 2833, 1733, 1607, 1569, 1499, 1444, 1429, 1372, 1303, 1281, 1252, 1205, 1165, 1142, 1096, 1080, 1031, 865 cm^{-1} ; HRMS (ESI) calcd. for $\text{C}_{16}\text{H}_{20}\text{NaO}_4$ $[\text{M}+\text{Na}]^+$: 299.1259, found: 299.1254; HPLC analysis CHIRALPAK AS-H (Hexane/IPA = 85/15, 1.0 mL/min, 254 nm, 30 °C) 9.4 min (major), 16.4 min (minor), 63% ee; $[\alpha]_{\text{D}}^{21} = 3.82$ ($c = 1.00$, CHCl_3 , 63% ee). **Configuration Assignment:** The absolute configuration was assigned as (S) by analogy.



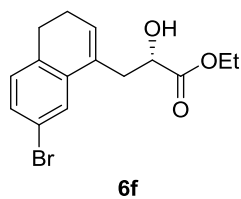
ethyl (S)-2-hydroxy-3-(6-chloro-3,4-dihydronaphthalen-1-yl)propanoate (6d)

6d: Colorless oil: (37.1 mg, 66% yield); ^1H NMR (CDCl_3 , 600 MHz) δ 7.20 (d, $J = 8.4$ Hz, 1H), 7.16 (dd, $J = 8.4$ Hz, 2.4 Hz, 1H), 7.13–7.12 (m, 1H), 5.99 (t, $J = 4.2$ Hz, 1H), 4.35–4.31 (m, 1H), 4.22–4.12 (m, 2H), 2.98 (dd, $J = 15.0$ Hz, 4.8 Hz, 1H), 2.77–2.68 (m, 4H), 2.28–2.25 (m, 2H), 1.27 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (CDCl_3 , 151 MHz) δ 174.5, 138.6, 132.7, 132.2, 131.1, 128.8, 127.7, 126.2, 123.9, 69.5, 61.7, 37.9, 28.1, 22.9, 14.1; IR (ATR) 3464, 2981, 2937, 2887, 2832, 1733, 1592, 1559, 1486, 1442, 1407, 1369, 1274, 1208, 1098, 1024 cm^{-1} ; HRMS (ESI) calcd. for $\text{C}_{15}\text{H}_{17}\text{ClNaO}_3$ $[\text{M}+\text{Na}]^+$: 303.0764, found: 303.0758; HPLC analysis CHIRALPAK AS-H (Hexane/IPA = 85/15, 1.0 mL/min, 254 nm, 30 °C) 6.6 min (major), 9.6 min (minor), 86% ee; $[\alpha]_{\text{D}}^{21} = 2.97$ ($c = 1.00$, CHCl_3 , 86% ee). **Configuration Assignment:** The absolute configuration was assigned as (S) by analogy.



ethyl (S)-2-hydroxy-3-(7-methoxy-3,4-dihydronaphthalen-1-yl)propanoate (6e)

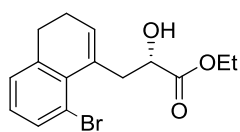
6e: Colorless oil: (47.5 mg, 86% yield); ^1H NMR (CDCl_3 , 600 MHz) δ 7.06 (d, $J = 7.8$ Hz, 1H), 6.87 (d, $J = 2.4$ Hz, 1H), 6.69 (dd, $J = 7.8$ Hz, 2.4 Hz, 1H), 6.01 (t, $J = 4.2$ Hz, 1H), 4.39–4.35 (m, 1H), 4.21–4.11 (m, 2H), 3.80 (s, 3H), 3.00 (dd, $J = 15.0$ Hz, 4.2 Hz, 1H), 2.74–2.65 (m, 4H), 2.27–2.24 (m, 2H), 1.26 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (CDCl_3 , 151 MHz) δ 174.6, 158.3, 135.1, 131.6, 129.3, 128.9, 128.3, 111.2, 109.6, 69.4, 61.6, 55.3, 38.1, 27.3, 23.5, 14.1; IR (ATR) 3469, 2980, 2934, 2884, 2832, 1733, 1604, 1572, 1492, 1466, 1444, 1422, 1369, 1300, 1279, 1252, 1203, 1096, 1045, 866 cm^{-1} ; HRMS (ESI) calcd. for $\text{C}_{16}\text{H}_{20}\text{NaO}_4$ $[\text{M}+\text{Na}]^+$: 299.1259, found: 299.1254; HPLC analysis CHIRALPAK AS-H (Hexane/IPA = 85/15, 1.0 mL/min, 254 nm, 30 °C) 8.4 min (major), 15.7 min (minor), 92% ee; $[\alpha]_{\text{D}}^{21} = 3.65$ ($c = 1.00$, CHCl_3 , 92% ee). **Configuration Assignment:** The absolute configuration was assigned as (S) by analogy.



6f

ethyl (S)-2-hydroxy-3-(7-bromo-3,4-dihydronaphthalen-1-yl)propanoate (6f)

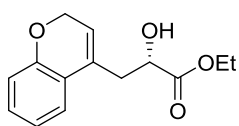
6f: Colorless oil: (41.6 mg, 64% yield); ^1H NMR (CDCl_3 , 600 MHz) δ 7.39 (d, J = 1.2 Hz, 1H), 7.28–7.24 (m, 1H), 7.01 (d, J = 8.4 Hz, 1H), 6.05 (t, J = 4.5 Hz, 1H), 4.37–4.34 (m, 1H), 4.24–4.10 (m, 2H), 2.94 (dd, J = 15.0 Hz, 4.8 Hz, 1H), 2.77–2.67 (m, 4H), 2.28–2.25 (m, 2H), 1.29 (t, J = 7.2 Hz, 3H); ^{13}C NMR (CDCl_3 , 151 MHz) δ 174.5, 136.3, 135.5, 130.8, 129.9, 129.6, 129.2, 125.6, 120.0, 69.3, 61.8, 37.6, 27.7, 23.0, 14.1; IR (ATR) 3462, 2980, 2935, 2886, 2832, 1733, 1590, 1555, 1479, 1442, 1397, 1369, 1270, 1233, 1207, 1097, 1024, 876 cm^{-1} ; HRMS (ESI) calcd. for $\text{C}_{15}\text{H}_{17}\text{BrNaO}_3$ $[\text{M}+\text{Na}]^+$: 347.0259, found: 347.0254; HPLC analysis CHIRALPAK AS-H (Hexane/IPA = 85/15, 1.0 mL/min, 254 nm, 30 °C) 6.9 min (major), 14.5 min (minor), 88% ee; $[\alpha]_{\text{D}}^{22}$ = 12.20 (c = 1.00, CHCl_3 , 88% ee). **Configuration Assignment:** The absolute configuration was assigned as (S) by analogy.



6g

ethyl (S)-3-(8-bromo-3,4-dihydronaphthalen-1-yl)-2-hydroxypropanoate (6g)

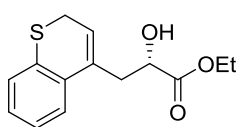
6g: Colorless oil: (24.1 mg, 37% yield); ^1H NMR (CDCl_3 , 600 MHz) δ 7.43 (dd, J = 7.8 Hz, 1.2 Hz, 1H), 7.13 (dd, J = 7.2 Hz, 1.2 Hz, 1H), 6.95 (t, J = 7.8 Hz, 1H), 6.28 (t, J = 5.4 Hz, 1H), 4.29–4.25 (m, 1H), 4.11–4.05 (m, 1H), 3.95–3.89 (m, 1H), 3.50 (ddd, J = 14.4 Hz, 4.8 Hz, 1.2 Hz, 1H), 3.28 (dd, J = 15.0 Hz, 7.8 Hz, 1H), 2.68–2.58 (m, 3H), 2.11–2.06 (m, 2H), 1.20 (t, J = 7.2 Hz, 3H); ^{13}C NMR (CDCl_3 , 151 MHz) δ 174.7, 142.5, 134.5, 133.6, 133.0, 132.7, 127.6, 126.6, 119.6, 69.7, 61.5, 39.6, 30.6, 22.7, 14.0; IR (ATR) 3479, 2980, 2938, 2893, 2834, 1731, 1439, 1271, 1208, 1097, 1028 cm^{-1} ; HRMS (ESI) calcd. for $\text{C}_{15}\text{H}_{17}\text{BrNaO}_3$ $[\text{M}+\text{Na}]^+$: 347.0259, found: 347.0254; HPLC analysis CHIRALPAK AS-H (Hexane/IPA = 75/25, 1.0 mL/min, 254 nm, 30 °C) 7.02 min (major), 17.4 min (minor), 60% ee; $[\alpha]_{\text{D}}^{20}$ = 5.46 (c = 0.95, CHCl_3 , 60% ee). **Configuration Assignment:** The absolute configuration was assigned as (S) by analogy.



6h

ethyl (S)-3-(2H-chromen-4-yl)-2-hydroxypropanoate (6h)

6h: Colorless oil: (37.2 mg, 75% yield); ^1H NMR (CDCl_3 , 600 MHz) δ 7.20 (dd, J = 7.2 Hz, 1.2 Hz, 1H), 7.13 (dt, J = 7.2 Hz, 1H), 6.80 (d, J = 7.8 Hz, 1.8 Hz, 1H), 6.91 (dt, J = 7.2 Hz, 1.2 Hz, 1H), 6.82 (d, J = 7.8 Hz, 1H), 5.75 (t, J = 3.9 Hz, 1H), 4.76 (d, J = 3.0 Hz, 2H), 4.40–4.36 (m, 1H), 4.25–4.15 (m, 2H), 2.95 (dd, J = 14.4 Hz, 4.2 Hz, 1H), 2.80 (d, J = 6.6 Hz, 1H), 2.70 (dd, J = 15.5 Hz, 7.8 Hz, 1H), 1.28 (t, J = 6.6 Hz, 3H); ^{13}C NMR (CDCl_3 , 151 MHz) δ 174.3, 154.4, 129.6, 129.2, 123.2, 123.0, 121.1, 121.0, 116.2, 69.2, 65.1, 61.8, 36.4, 14.1; IR (ATR) 3470, 2980, 2937, 2905, 2837, 1731, 1651, 1605, 1577, 1487, 1449, 1369, 1299, 1219, 1128, 1097, 1028, 937, 862 cm^{-1} ; HRMS (ESI) calcd. for $\text{C}_{14}\text{H}_{16}\text{NaO}_4$ $[\text{M}+\text{Na}]^+$: 271.2678, found: 271.2673; HPLC analysis CHIRALPAK AS-H (Hexane/IPA = 90/10, 1.0 mL/min, 254 nm, 30 °C) 10.6 min (major), 12.2 min (minor), 91% ee; $[\alpha]_{\text{D}}^{22}$ = -5.50 (c = 1.00, CHCl_3 , 91% ee). **Configuration Assignment:** The absolute configuration was assigned as (S) by analogy.



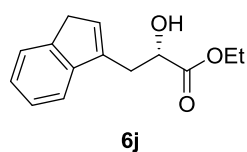
6i

ethyl (S)-2-hydroxy-3-(2H-thiophene-4-yl)propanoate (6i)

6i: Colorless oil: (31.2 mg, 59% yield); ^1H NMR (CDCl_3 , 600 MHz) δ 7.36 (d, J = 7.8 Hz, 1H), 7.32 (d, J = 7.2 Hz, 1H), 7.17–7.10 (m, 2H), 6.01 (t, J = 5.4 Hz, 1H), 4.32–4.27 (m, 1H), 4.19–4.09 (m, 2H), 3.33 (dd, J = 15.6 Hz, 6.0 Hz, 1H), 3.29 (dd, J = 15.0 Hz, 6.0 Hz, 1H), 3.01 (dd, J = 14.4 Hz, 4.8 Hz, 1H), 2.79 (dd, J = 14.4 Hz, 7.8 Hz, 1H), 2.71 (d, J = 6.6 Hz, 1H), 1.27 (t, J = 7.2 Hz, 3H); ^{13}C NMR

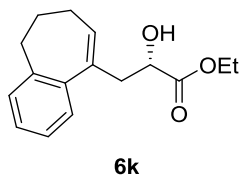
(CDCl₃, 151 MHz) δ 174.3, 133.8, 133.6, 133.1, 127.9, 127.6, 125.6, 124.8, 122.5, 69.5, 61.7, 39.1, 24.8, 14.1; IR (ATR) 3448, 2978, 2934, 1734, 1633, 1588, 1471, 1446, 1368, 1219, 1152, 1099, 1024 cm⁻¹; HRMS (ESI) calcd. for C₁₄H₁₆NaO₃S [M+Na]⁺: 287.0718, found: 287.0714; HPLC analysis CHIRALPAK AS-H (Hexane/IPA = 90/10, 1.0 mL/min, 254 nm, 30 °C) 15.8 min (major), 17.5 min (minor), 83% ee; $[\alpha]^{22}_D = +6.94$ ($c = 1.00$, CHCl₃, 83% ee).

Configuration Assignment: The absolute configuration was assigned as (S) by analogy.



ethyl (S)-2-hydroxy-3-(1H-inden-3-yl)propanoate (6j)

6j: Colorless oil: (22.3 mg, 48% yield); ¹H NMR (CDCl₃, 600 MHz) δ 7.46 (d, $J = 7.2$ Hz, 1H), 7.40 (d, $J = 7.2$ Hz, 1H), 7.30 (t, $J = 7.5$ Hz, 1H), 7.21 (t, $J = 7.5$ Hz, 1H), 6.39 (brs, 1H), 4.55–4.51 (m, 1H), 4.25–4.16 (m, 2H), 3.37 (brs, 2H), 3.10 (dd, $J = 14.4$ Hz, 2.1 Hz, 1H), 2.95 (dd, $J = 15.0$ Hz, 7.2 Hz, 1H), 2.84 (d, $J = 6.0$ Hz, 1H), 1.26 (t, $J = 7.2$ Hz, 3H); ¹³C NMR (CDCl₃, 151 MHz) δ 174.4, 145.0, 144.2, 139.2, 131.2, 126.0, 124.7, 123.8, 119.0, 69.7, 61.8, 38.0, 32.7, 14.1; IR (ATR) 3486, 3065, 2981, 2904, 1732, 1462, 1395, 1259, 1206, 1096, 1055, 1028 cm⁻¹; HRMS (ESI) calcd. for C₁₄H₁₆NaO₃ [M+Na]⁺: 255.0997, found: 255.0992; HPLC analysis CHIRALPAK AS-H (Hexane/IPA = 85/15, 1.0 mL/min, 254 nm, 30 °C) 6.5 min (major), 8.0 min (minor), 62% ee; $[\alpha]^{20}_D = 1.21$ ($c = 1.00$, CHCl₃, 62% ee). **Configuration Assignment:** The absolute configuration was assigned as (S) by analogy.

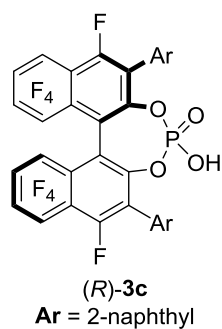
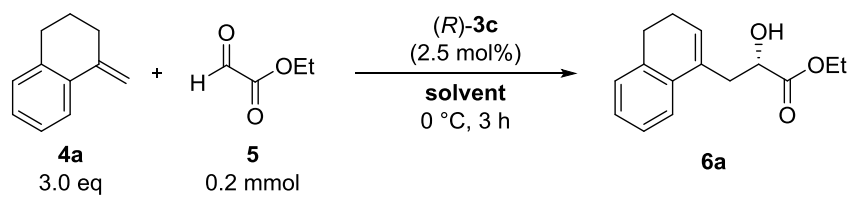


ethyl (S)-3-(6,7-dihydro-5H-benzo[7]annulen-9-yl)-2-hydroxypropanoate (6k)

6k: Colorless oil: (39.0 mg, 75% yield); ¹H NMR (CDCl₃, 600 MHz) δ 7.26–7.15 (m, 4H), 6.08 (t, $J = 7.2$ Hz, 1H), 4.17–4.14 (m, 1H), 4.03–3.98 (m, 1H), 3.86–3.80 (m, 1H), 3.01 (dd, $J = 13.8$ Hz, 4.2 Hz, 1H), 2.86 (dd, $J = 14.4$ Hz, 6.6 Hz, 1H), 2.65 (d, $J = 6.0$ Hz, 1H), 2.62–2.57 (m, 2H), 2.12–2.07 (m, 2H), 1.86–1.77 (m, 2H), 1.18 (t, $J = 7.2$ Hz, 3H); ¹³C NMR (CDCl₃, 151 MHz) δ 174.6, 141.8, 139.6, 136.0, 130.2, 129.0, 126.8, 126.2, 125.8, 69.4, 61.4, 41.4, 34.7, 32.1, 24.6, 14.0; IR (ATR) 3482, 3015, 2980, 2932, 2856, 1731, 1486, 1448, 1368, 1259, 1208, 1097, 1037 cm⁻¹; HRMS (ESI) calcd. for C₁₆H₂₀NaO₃ [M+Na]⁺: 283.1310, found: 283.1305; HPLC analysis CHIRALPAK AS-H (Hexane/IPA = 85/15, 1.0 mL/min, 254 nm, 30 °C) 5.9 min (major), 7.0 min (minor), 93% ee; $[\alpha]^{24}_D = 14.47$ ($c = 1.04$, CHCl₃, 93% ee). **Configuration Assignment:** The absolute configuration was assigned as (S) by analogy.

5. Additional Screening

Screening of solvents

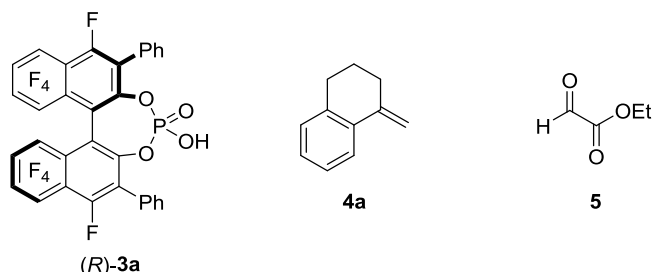


entry	solvent	yield (%) ^a	ee (%)
1 ^b	benzene	73	62
2	toluene	97	71
3	<i>o</i> -xylene	92	72
4	<i>m</i> -xylene	59	74
5 ^b	<i>p</i> -xylene	90	69
6	CF ₃ -benzene	54	59
7 ^b	cyclohexane	67	74
8	CH ₂ Cl ₂	76	43

^a NMR yield (isolated yield in parentheses), ^b 15 °C

6. Computational Studies

6-1. Model system: As the preliminary computational study, the mechanism of the carbonyl-ene reaction using the model system of F₁₀BINOL-derived phosphoric acid catalyst (*R*)-**3a** and the standard substrates (**4a** and **5**) were investigated. Geometries were optimized and characterized using frequency calculations at the M06-2X/6-31G(d,p) level. Energies in the solution phase were calculated using single-point energy calculations at the same level according to the CPCM solvation model (toluene: $\epsilon=2.379$) for the optimized structures.



A variety of initial structures were thoroughly explored to identify the transition state of carbonyl ene reaction, and it was found that the reaction proceeds through a stepwise mechanism via **INT-1** (Figure S1). Initially, catalyst forms hydrogen bonds with **4a** and **5** (association or association'). C-C bond formation through transition state **TS-1** yields the **INT-1** respectively. The following abstraction of proton proceeds via transition state **TS-2** or **TS-2'** (**TS-2**: Phosphorous oxygen of catalyst abstracts the proton., **TS-2'**: Alcohol oxygen abstracts the proton.). The energy profile indicates that **TS-2** is more energetically favored pathway than **TS-2'**.

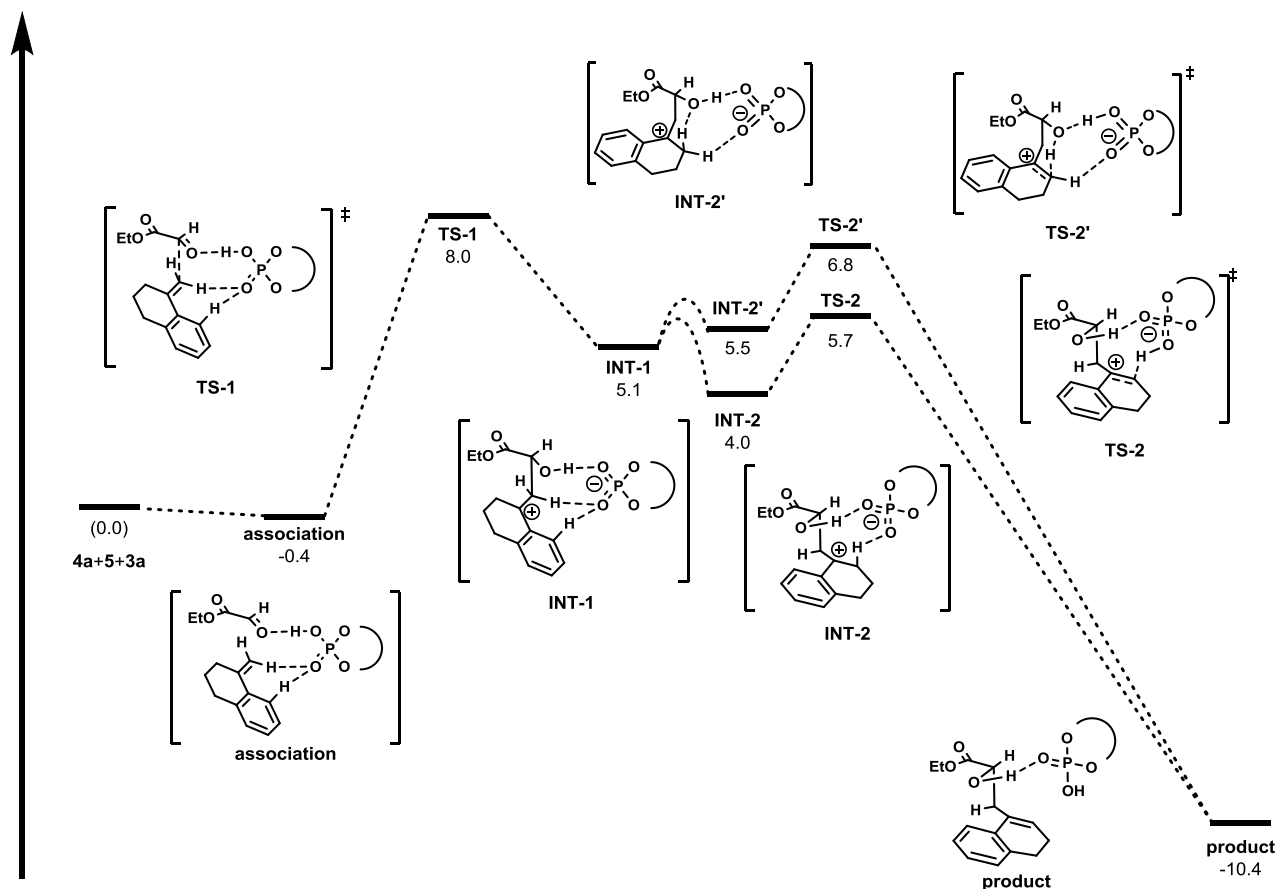


Figure S1. Energy profile of the carbonyl-ene reaction for the model reaction. The potential energy for the sum of **4a**, **5** and **3** was set to zero. Geometries were optimized and characterized using frequency calculations at the M06-2X/6-31G(d,p) level. Gibbs free energies (kcal/mol) in solution phase were calculated using single-point energy calculations at the same level as those for the optimized structures according to the SCRF method based on CPCM (toluene).

A summary of calculated values including solvent corrected single point energy values, as well as enthalpic and entropic corrections at 298.15 K are provided (**Table S1**).

	G_{CCPM} (kcal/mol)	E_{CCPM} (kcal/mol)	Enthalpic correction(kcal/mol)	S (cal/mol K)	ΔG (kcal/mol)
4a+5+3a	-2304858.6	-	-	-	0.0
association	-2304859.0	-2305223.0	458.1	315.5	-0.4
TS-1	-2304850.6	-2305214.9	456.8	310.5	8.0
INT-1	-2304853.5	-2305220.4	459.3	310.3	5.1
INT-2	-2304854.6	-2305221.7	459.3	309.3	4.0
INT-2'	-2304853.1	-2305244.2	458.2	306.7	5.5
TS-2	-2304852.9	-2305218.4	456.7	306.0	5.7
TS-2'	-2304851.8	-2305217.2	455.7	303.1	6.8
product	-2304869.0	-2305235.4	459.6	312.4	-10.4

Table S1. Summary of calculated values in model system

6-2. Real system: In order to conduct rigorous conformational analysis of these transition states, we systematically generated initial structures of the transition states by changing the dihedral angles of $C^2-C^1-C^a-C^b$ and $C^{2'}-C^{1'}-C^{a'}-C^{b'}$ (for each dihedral angle: -135° , -45° , 45° , and 135°) as well as the orientation of the methoxy group (dihedral angle: 0° and 180° to the naphthyl ring) (Figure S2a). This procedure generates eight conformers for each naphthyl ring and hence totally 64 conformers were considered for the initial structures of the catalyst. In addition, on the basis of the model studies of the transition states, conformationally different transition states of the real system were considered by changing the relative orientation of ene-component **4a** and the conformation of the six-membered ring of **4a** for each enantiomeric pathway. Thus, nearly 200 initial structures were calculated to identify the energetically favored transition states accurately. Figure S2b presents the relative Gibbs free energy distribution of 10 TSs for (S)-**6a** and 7 TSs for (R)-**6a** within 3.0 kcal/mol. Although there are many energetically similar TSs, a comparison between the lowest TSs for each enantiomer clearly shows that the (S)-**6a** is the main product in this reaction in agreement with the experiment. The lowest TS for the (R)-**6a** (TS-r (e)) is 1.1 kcal/mol higher in energy than the lowest TS for the (S)-**6a** (TS-s (a)).

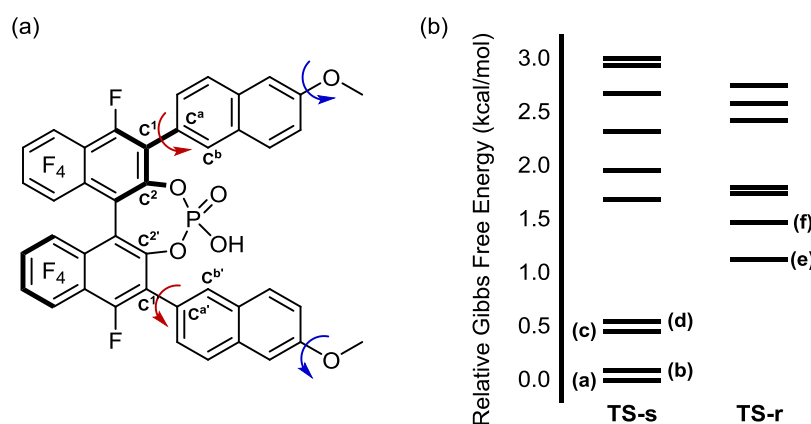


Figure S2. (a) Bond rotation of catalyst substituents. (b) Gibbs free-energy distribution of TSs for (S)-**6a** and TSs for (R)-**6a** within 3.0 kcal/mol.

The 6 lowest TS structures were listed (Figure S3) and the following key geometric parameters in each structures of $(R)\text{-3PA}_{\text{TS}}$ were extracted (Table 2): dihedral angle ($\text{C}^2\text{-C}^1\text{-C}^a\text{-C}^b$), dihedral angles ($\text{C}^2\text{-C}^1\text{-C}^a\text{-C}^b$, $\text{C}^{2'}\text{-C}^{1'}\text{-C}^{a'}\text{-C}^{b'}$), interior bond angles (A and A'), and exterior bond angles (B and B').

In all TS structures, similar structural distortion of the catalyst was observed.

*In the main text, **TS-s (a)** and **TS-r (e)** are shown in **TS-s** and **TS-r**, respectively.

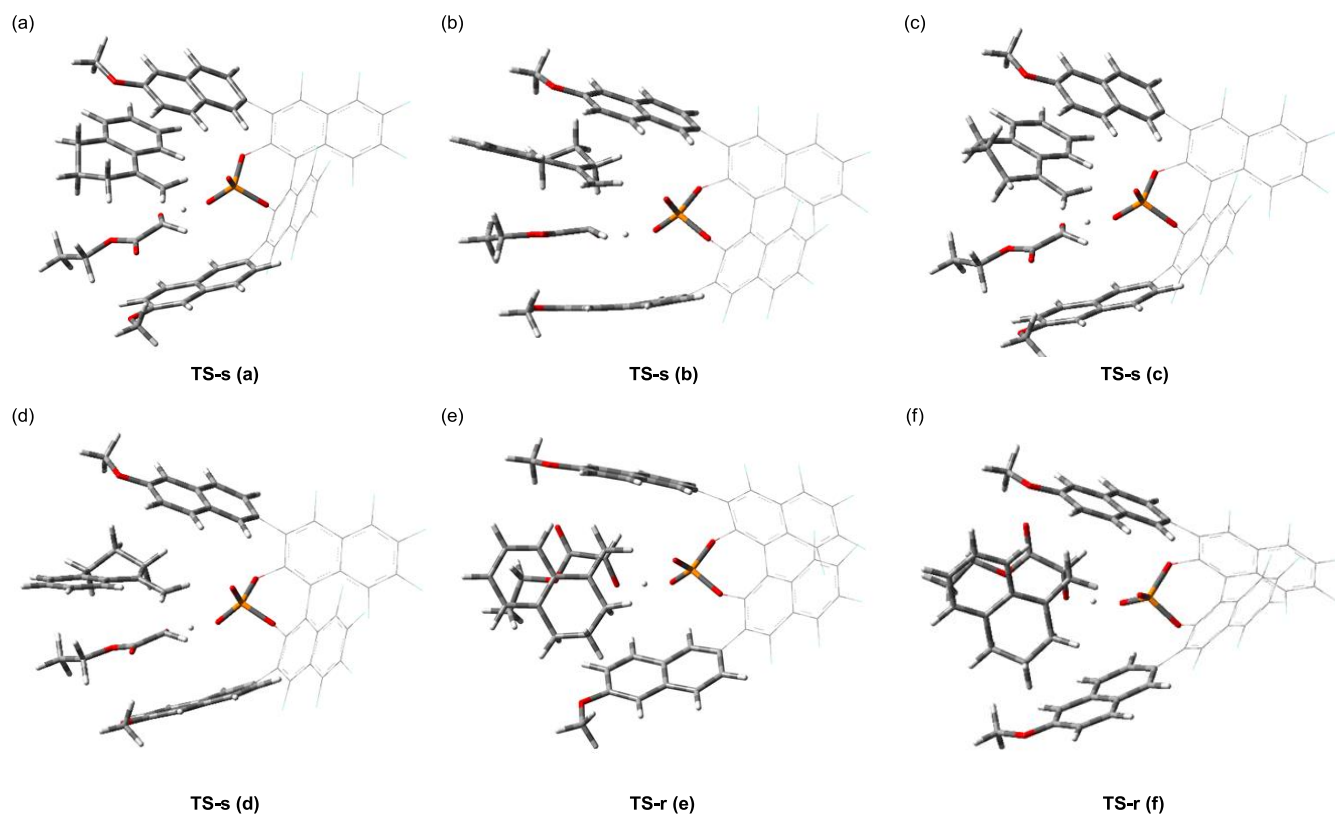
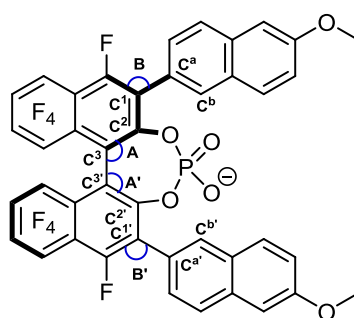


Figure S3. The 6 lowest TS structures



	$\text{C}^2\text{-C}^1\text{-C}^a\text{-C}^b$	$\text{C}^{2'}\text{-C}^{1'}\text{-C}^{a'}\text{-C}^{b'}$	$\text{C}^2\text{-C}^3\text{-C}^{3'}\text{-C}^{2'}$	A	A'	B	B'
$(R)\text{-3PA}_{\text{TS-s (a)}}^*$	76.4°	115.7°	-52.2°	118.0°	117.4°	122.8°	123.6°
$(R)\text{-3PA}_{\text{TS-s (b)}}$	96.4°	116.2°	-51.5°	117.4°	116.9°	123.7°	124.2°
$(R)\text{-3PA}_{\text{TS-s (c)}}$	75.1°	117.1°	-52.2°	118.0°	117.4°	122.8°	123.5°
$(R)\text{-3PA}_{\text{TS-s (d)}}$	76.5°	115.8°	-52.1°	117.8°	117.3°	123.0°	123.5°
$(R)\text{-3PA}_{\text{TS-r (e)}}$	77.8°	121.3°	-51.9°	118.0°	117.4°	124.3°	123.3°
$(R)\text{-3PA}_{\text{TS-r (f)}}$	121.9°	124.8°	-51.8°	116.8°	116.9°	123.2°	122.9°

Table S2. Selected geometric parameters for structures of $(R)\text{-3PA}_{\text{TS}}$. *In the main text, $(R)\text{-3PA}_{\text{TS-s (a)}}$ is shown in $(R)\text{-3PA}_{\text{TS-s}}$.

A summary of calculated values in the lowest TS for the (*S*)-**6a** (**TS-s (a)**) and the lowest TS for the (*R*)-**6a** (**TS-r (e)**) including solvent corrected single point energy values, as well as enthalpic and entropic corrections associated with vibrational energy at 298.15 K are provided (**Table S3**). These result indicates that the enantiorecognition of this carbonyl-ene reaction contains a large enthoropic contribution.

	G_{CPCM} (kcal/mol)	E_{CPCM} (kcal/mol)	Enthalpy correction(kcal/mol)	S (cal/mol K)	ΔG (kcal/mol)
TS-s (a)	-2641191.3	-2641652.5	563.9	344.6	0.0
TS-r (e)	-2641190.2	-2641649.7	563.0	347.1	1.1

Table S3. Summary of calculated values in real system

6-3. Transition state in the carbonyl-ene reaction catalyzed by BINOL-derived phosphoric acid (*R*)-1

The transition state in the major pathway of the carbonyl-ene reaction catalyzed by (*R*)-**1** (Ar = 6-methoxy-2-naphthyl) was identified by performing calculation after replacing the fluorine atoms of **TS-s (a)** with hydrogen atoms (Figure S4). In obtained **TS-s ((R)-1)**, it was confirmed that multi-point C–H···O hydrogen bonds and the π interactions were formed between the catalyst and the substrates, as likely shown in transition state **TS-s (a)**.*

*In the main text, **TS-s (a)** is shown in **TS-s**.

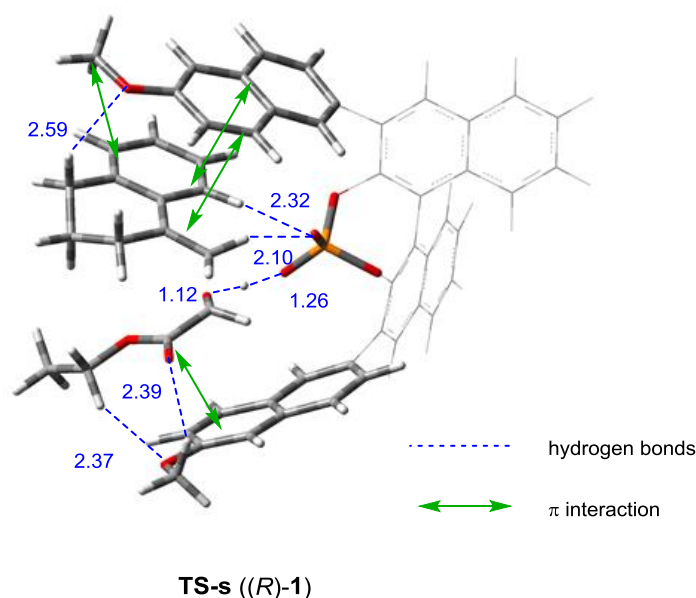


Figure S4. Transition state in the carbonyl-ene reaction of **4a** with **5** catalyzed by (*R*)-**1**. Geometries were optimized and characterized using frequency calculations at the M06-2X/6-31G(d,p) level. Relative Gibbs free energies (kcal/mol) obtained by single-point energy calculations at the same level for the optimized structures with the SCRF method based on CPCM (toluene) are shown.

6-4. Second-Perturbation Energy (kcal/mol) of Donor-acceptor Interaction

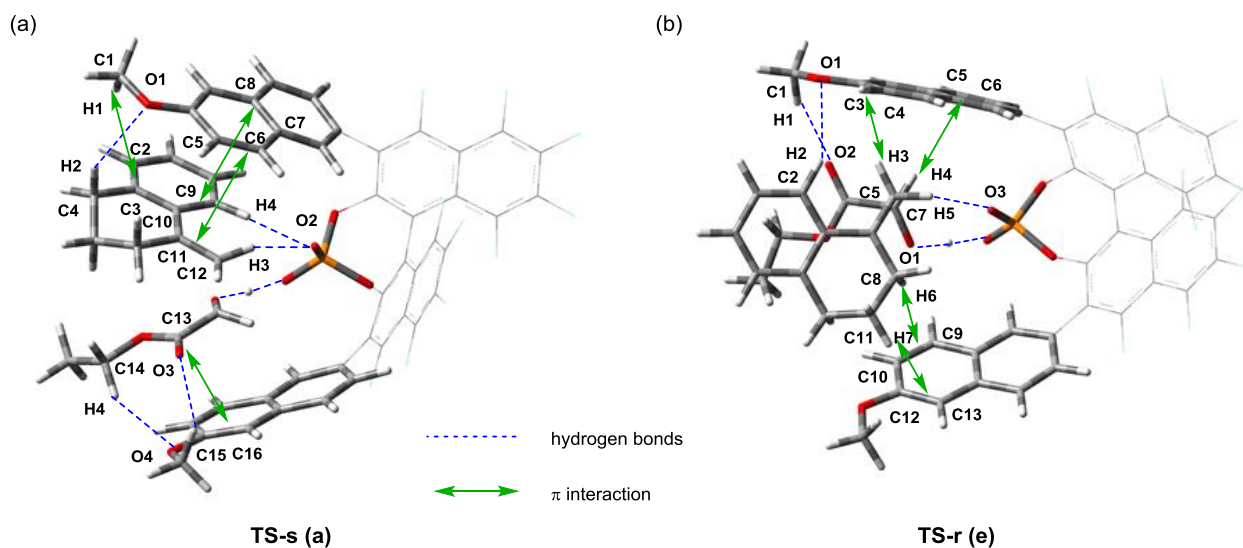


Figure S5. Donor-acceptor Interaction with Respect to (a) **TS-s (a)** and (b) **TS-r (e)**.

*In the main text, **TS-s (a)** and **TS-r (e)** are shown in **TS-s** and **TS-r**, respectively.

TS-s (a)				TS-r (e)			
donor	acceptor	Interaction	E (kcal/mol)	donor	acceptor	Interaction	E (kcal/mol)
BD C2-C3	BD* C1-H1	$\pi \rightarrow \sigma^*$	0.56	LP O1	BD* C2-H2	$n \rightarrow \sigma^*$	2.73
LP O1	BD* C4-H2	$n \rightarrow \sigma^*$	0.60	LP(2) O2	BD* C1-H1	$n \rightarrow \sigma^*$	0.61
BD C11-C12	BD* C5-C6	$\pi \rightarrow \pi^*$	0.53	BD C3-C4	BD* C5-H3	$\pi \rightarrow \sigma^*$	1.96
BD C7-C8	BD* C9-C10	$\pi \rightarrow \pi^*$	0.95	BD C5-C6	BD* C7-H4	$\pi \rightarrow \sigma^*$	0.55
LP O2	BD* C12-H3	$n \rightarrow \sigma^*$	3.47	LP O3	BD* C5-H5	$n \rightarrow \sigma^*$	5.75
LP O2	BD* C12-H3	$n \rightarrow \sigma^*$	3.22	LP(2) O3	BD* C5-H5	$n \rightarrow \sigma^*$	4.09
LP O2	BD* C9-H4	$n \rightarrow \sigma^*$	1.14	BD C9-C10	BD* C8-H6	$\pi \rightarrow \sigma^*$	0.69
LP O2	BD* C9-H4	$n \rightarrow \sigma^*$	1.24	BD C12-C13	BD* C11-H7	$\pi \rightarrow \sigma^*$	0.91
BD C15-C16	BD* C13-O3	$\pi \rightarrow \pi^*$	0.74				
LP O4	BD* C14-H4	$n \rightarrow \sigma^*$	1.17				
LP(2) O4	BD* C14-H4	$n \rightarrow \sigma^*$	1.38				

Table S4. Second-Perturbation Energy (kcal/mol)

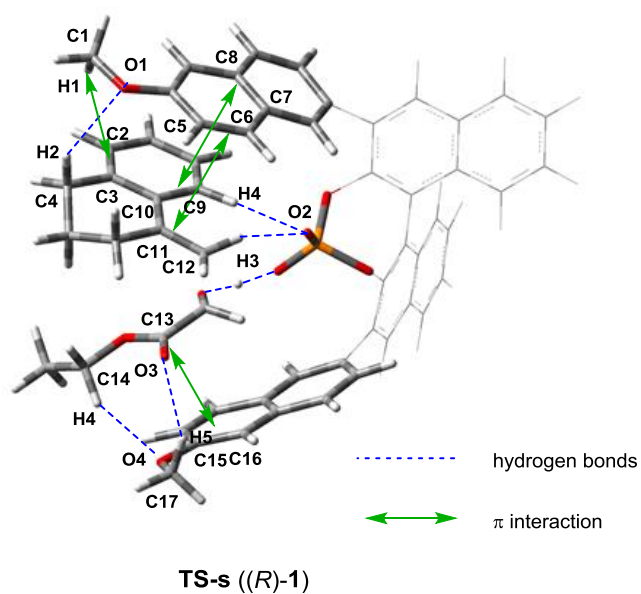


Figure S6. Donor-acceptor Interaction with Respect to TS-s ((R)-1)

TS-s ((R)-1)							
donor	acceptor	Interaction	E (kcal/mol)	donor	acceptor	Interaction	E (kcal/mol)
BD C2-C3	BD* C1-H1	$\pi \rightarrow \sigma^*$	0.53	LP(3) O2	BD* C9-H4	$n \rightarrow \sigma^*$	1.66
LP O1	BD* C4-H2	$n \rightarrow \sigma^*$	0.53	BD C13-O3	BD* C15-H16	$\pi \rightarrow \sigma^*$	0.69
BD C12-C13	BD* C5-C6	$\pi \rightarrow \pi^*$	0.99	LP O3	BD* C17-H5	$n \rightarrow \sigma^*$	1.15
BD C7-C8	BD* C9-C10	$\pi \rightarrow \pi^*$	0.89	LP O4	BD* C14-H4	$n \rightarrow \sigma^*$	1.14
LP O2	BD* C12-H3	$n \rightarrow \sigma^*$	3.65				
LP(2) O2	BD* C12-H3	$n \rightarrow \sigma^*$	3.26				
LP(2) O2	BD* C9-H4	$n \rightarrow \sigma^*$	0.99				

Table S5. Second-Perturbation Energy (kcal/mol)

6-5. Structural analysis of BINOL-derived phosphate anion

The structural analysis of BINOL-derived phosphate anions (**PA**s) was conducted as follows: (i) The catalyst structure of BINOL-derived phosphate anion (*R*)-**1PA**_{TS-s} was extracted from **TS-s** ((*R*)-**1**) (Figure S7b). (ii) the structure of BINOL-derived phosphate anion (*R*)-**1PA**_{opt} was optimized from (*R*)-**1PA**_{TS-s} as the initial structure (Figure S7d) (The obtained structure of (*R*)-**1PA**_{opt} was identical to the structure which was optimized after replacing the fluorine atoms of (*R*)-**3PA**_{opt} with hydrogen atoms.)

As shown in Table S6, the dihedral angles $C^2-C^1-C^a-C^b$ and $C^{2'}-C^{1'}-C^{a'}-C^{b'}$ in (*R*)-**1PA**_{TS-s} are markedly different from those in (*R*)-**1PA**_{opt}. These angle differences indicate that 6-methoxy-2-naphthyl substituents rotate to meet the requirements for the noncovalent bond interactions and the steric exclusion with the substrates in the transition state, as observed in (*R*)-**3PA** similarly. In both (*R*)-**1PA** and (*R*)-**3PA**, the dihedral angle $C^2-C^3-C^{3'}-C^{2'}$ in transition structures is smaller than that in the optimized structures. These angle differences imply that the binaphthyl (or perfluoro-binaphthyl) axis twists to interact with the substrates and lead to structural distortion of the catalyst framework.

The interior bond angles A and A' of (*R*)-**1PA**_{TS-s} are almost identical to those of (*R*)-**1PA**_{opt} and larger than those of (*R*)-**3PA**. In contrast, the exterior bond angles B and B' of (*R*)-**1PA**_{TS-s} are larger than those of (*R*)-**3PA**_{TS-s} (a), although these angles of (*R*)-**1PA**_{opt} are smaller than those of (*R*)-**3PA**. The total bond angle differences of B and B' ($\Delta B + \Delta B'$) between (*R*)-**1PA**_{TS-s} and (*R*)-**1PA**_{opt} is 7.2° (Between (*R*)-**3PA**_{TS-s} (a) and (*R*)-**3PA**_{opt}: 4.5°). In the case of BINOL-derived phosphoric acid, the more significant distortion of the catalyst fragment is enforced than that of F₁₀BINOL-derived one, because BINOL-derived phosphoric acid has the wider reaction space than that of F₁₀BINOL-derived one.

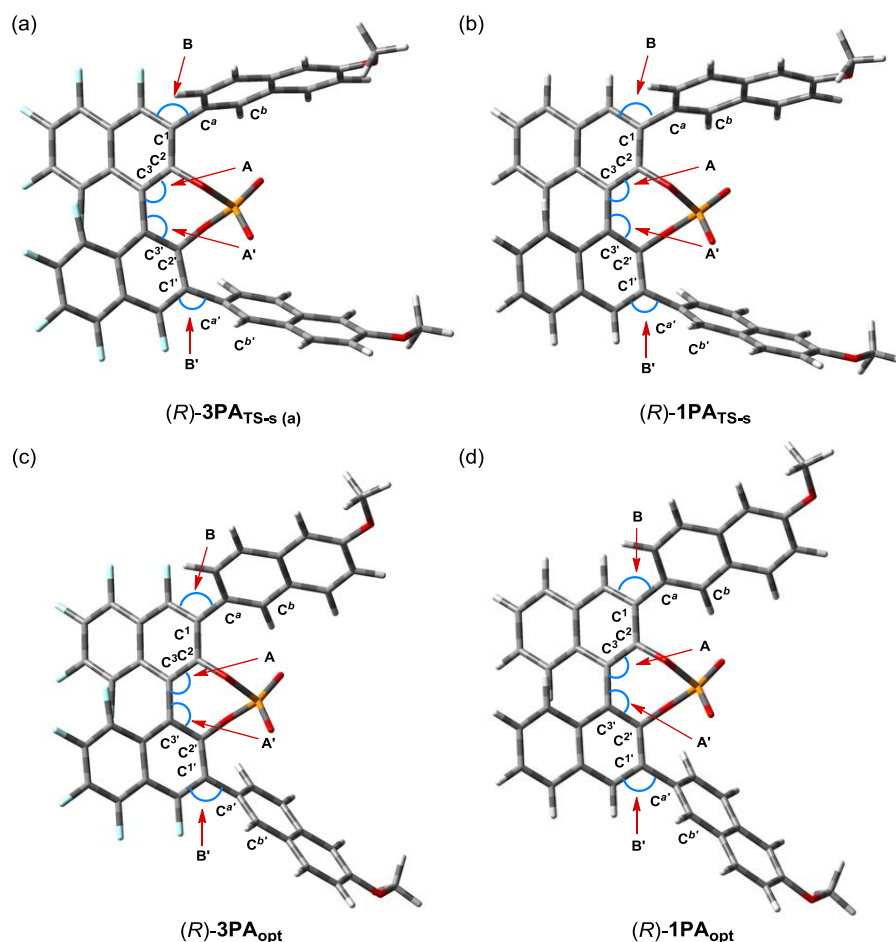


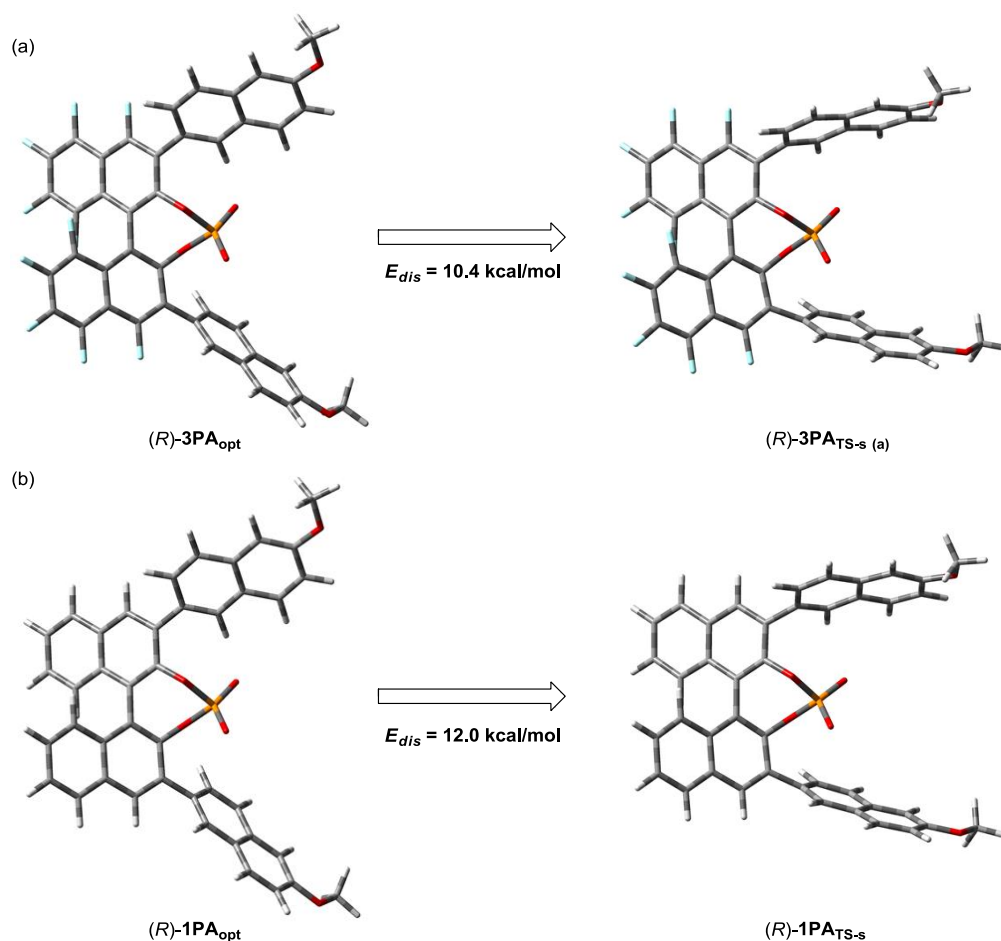
Figure S7. Structural analysis of the catalysts. (a) F₁₀BINOL-derived phosphate anion extracted from **TS-s** (a). (b) BINOL-derived phosphate anion extracted from **TS-s** ((*R*)-**1**). (c) F₁₀BINOL-derived phosphate anion optimized from (*R*)-**3PA**_{TS-s}. (d) BINOL-derived phosphate anion optimized from (*R*)-**1PA**_{TS-s}.

Table S6. Selected geometric parameters for catalyst structures.

	$\begin{matrix} 2 & 1 & a & b \\ C-C-C-C \end{matrix}$	$\begin{matrix} 2' & 1' & a' & b' \\ C-C-C-C \end{matrix}$	$\begin{matrix} 2 & 3 & 3' & 2' \\ C-C-C-C \end{matrix}$	A	A'	B	B'
(<i>R</i>)-3PA _{TS-s} (a)	76.4°	115.7°	-52.2°	118.0°	117.4°	122.8°	123.6°
(<i>R</i>)-1PA _{TS-s}	69.4°	114.8°	-50.3°	119.5°	118.8°	123.4°	123.8°
(<i>R</i>)-3PA _{opt}	50.3°	134.4°	-55.6°	117.8°	117.2°	120.2°	121.7°
(<i>R</i>)-1PA _{opt}	46.0°	139.7°	-53.3°	119.1°	118.6°	119.5°	120.5°

6-6. The distortion energy of catalysts

The analysis of the distortion energy in catalyst frameworks was conducted (Figure S8). The E_{dis} is the energy required to distort the catalyst framework from their optimized structures to their transition structures. As a result, it was found that the distortion energy difference between (*R*)-1PA_{opt} and (*R*)-1PA_{TS-s} ($E_{dis} = 12.0$ kcal/mol) is larger than the difference between (*R*)-3PA_{opt} and (*R*)-3PA_{TS-s} (a) ($E_{dis} = 10.4$ kcal/mol). These results clearly indicate that the energetic destabilization of the BINOL-derived phosphoric acid framework is more significant than that of the F₁₀BINOL-derived one in the transition state.

Figure S8. Energy for the catalyst distortion

	E_{TS-S} (a.u.)	E_{opt} (a.u.)	E_{dis} (a.u.)	E_{dis} (kcal/mol)
(<i>R</i>)-3PA	-3401.451913	-3401.468532	0.016619	10.42858869
(<i>R</i>)-1PA	-2409.466063	-2409.485134	0.019071	11.96724321

6-7. The optimization of the catalyst structure as phosphoric acid

DFT calculations for the structural analysis of chiral phosphoric acids were carried out in the following order: (i) the structure of F₁₀BINOL-derived phosphoric acid (*R*)-**3**_{TS-s} (**a**) was extracted from **TS-s** (**a**) (Figure S9a); (ii) the structure of F₁₀BINOL-derived phosphoric acid (*R*)-**3**_{opt} was optimized based on the (*R*)-**3**_{TS-s} (**a**) (Figure S9b); and (iii) the structure of BINOL-derived phosphoric acid (*R*)-**1**_{opt} was optimized by replacing the fluorine atoms of (*R*)-**3**_{opt} with hydrogen atoms (Figure S9c).

Figure S9. Structural analysis for Catalyst. (a) F₁₀BINOL-derived phosphoric acid from **TS-s** (**a**). (b) Optimized structure of F₁₀BINOL-derived phosphoric acid (*R*)-**3**_{opt}. (c) Optimized structure of BINOL-derived phosphoric acid (*R*)-**1**_{opt}.

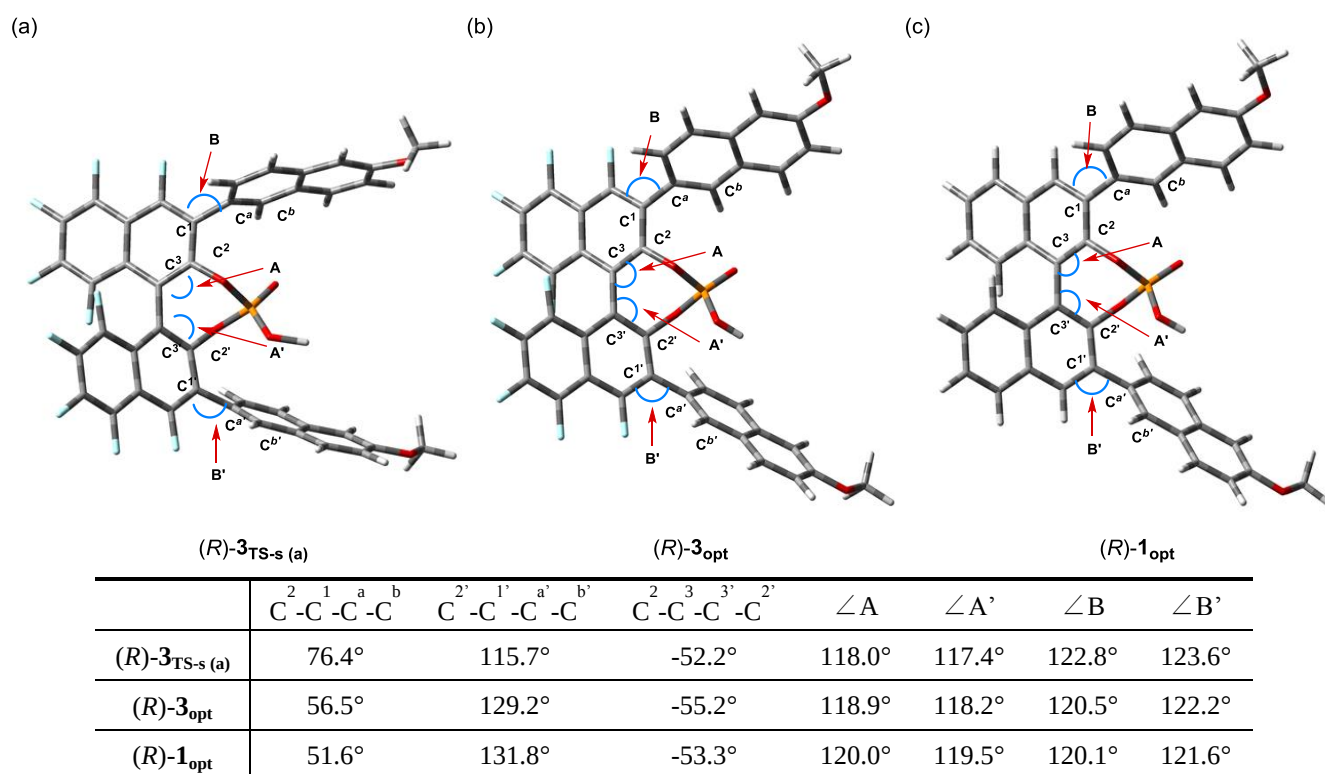
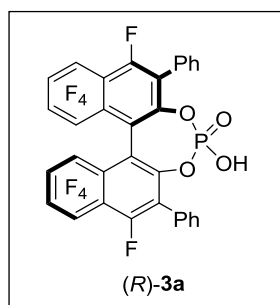


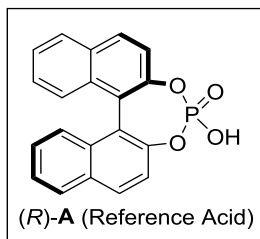
Table S7. Selected geometric parameters for catalyst structures.

6-8. Theoretical Study on the acidity of (R)-3a



The pK_a value of (R)-3a in DMSO was calculated with the SMD/M06-2x/6-311++G(2df,2p)//B3LYP/6-31+G(d) method². The geometries were fully optimized using the B3LYP/6-31+G(d) methods in gas-phase. The nature of the stationary points and thermal corrections to Gibbs free energy (**ZPG**) were obtained by frequency calculations at the same level of theory. The energies in solution phase (**E**) were obtained with thermal corrections to the Gibbs free energies (**G**) by performing SMD calculation (M06-2x/6-311++G(2df,2p)) with the gas-phase optimized geometries. In the proton exchange method, (R)-A (pK_a exp 3.37) was chosen as the reference acid.

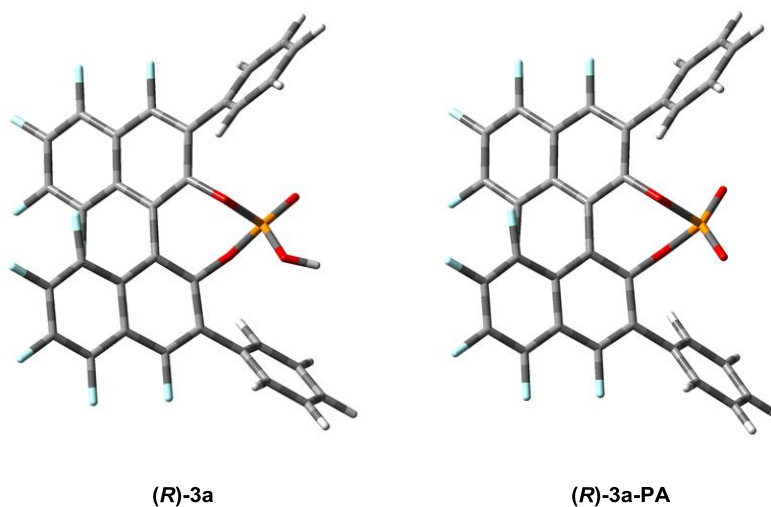
Reference Acid A



	G
(R)-A (Phosphoric Acid)	-1412.07198721
(R)-A-PA (Phosphate Anion)	-1411.63352682

² a) C. Yang, X.-S. Xue, X. Li, J.-P. Cheng, *J. Org. Chem.* 2013, **78**, 7076-7085; b) C. Yang, X.-S. Xue, X. Li, J.-P. Cheng, *J. Org. Chem.* 2014, **79**, 4340-4351.

Figure S10. Structures of 3,3'-Ph-F₁₀BINOL-derived Phosphoric Acid ((*R*)-**3a**) and Phosphate Anion ((*R*)-**3a-PA**)



	ZPG (hartree)	E (hartree)	G (hartree)
(<i>R</i>)-3a	0.290558	-2866.756485	-2866.465927
(<i>R</i>)-3a-PA	0.278222	-2866.310306	-2866.032084

• Direct method

$$\begin{aligned}
 \Delta G_{\text{soln}}^* &= \Delta G_{\text{gas}}^* + \Delta G_{\text{solv}}^*(\text{A}^-) + \Delta G_{\text{solv}}^*(\text{H}^+) - \Delta G_{\text{solv}}^*(\text{HA}) \\
 &= G_{\text{gas}}^*(\text{A}^-) + G_{\text{gas}}^*(\text{H}^+) - G_{\text{gas}}^*(\text{HA}) + \Delta G_{\text{solv}}^*(\text{A}^-) + \Delta G_{\text{solv}}^*(\text{H}^+) - \Delta G_{\text{solv}}^*(\text{HA}) \\
 &= [G_{\text{gas}}^*(\text{A}^-) + \Delta G_{\text{solv}}^*(\text{A}^-)] + [G_{\text{gas}}^*(\text{H}^+) + \Delta G_{\text{solv}}^*(\text{H}^+)] - [G_{\text{gas}}^*(\text{HA}) + \Delta G_{\text{solv}}^*(\text{HA})] \\
 &= G_{\text{soln}}^*(\text{A}^-) + G_{\text{soln}}^*(\text{H}^+) - G_{\text{soln}}^*(\text{HA})
 \end{aligned}$$

$$G_{\text{soln}}^*(\text{H}^+) = G_{\text{gas}}^*(\text{H}^+) + G_{\text{solv}}^*(\text{H}^+) = -4.39 + (-268.34 + 1.89) = -270.84 \text{ (kcal/mol)}$$

$$G_{\text{soln}}^*(\text{HA}) = \text{ZPG} + E_{\text{solv}}^*(\text{HA}) = 0.290558 + (-2866.756485) = -2866.465927 \text{ (hartree)}$$

$$G_{\text{soln}}^*(\text{A}^-) = \text{ZPG} + E_{\text{solv}}^*(\text{A}^-) = 0.278222 + (-2866.310306) = -2866.032084 \text{ (hartree)}$$

$$\Delta G_{\text{soln}}^* = 627.5095 \times \{-2866.032084 - (-2866.465927)\} - 270.84 = (1.40 \text{ kcal/mol})$$

$$\text{pKa} = \Delta G_{\text{soln}}^* \div 2.303RT = \mathbf{1.03}$$

• Proton Exchange Method

$$\begin{aligned}
 \Delta G_{\text{exchange}}^* &= \Delta G_{\text{gas,exchange}}^* + \Delta G_{\text{solv}}^*(\text{A}^-) + \Delta G_{\text{solv}}^*(\text{HRef}) - \Delta G_{\text{solv}}^*(\text{HA}) - \Delta G_{\text{solv}}^*(\text{Ref}) \\
 &= 627.51 \times \{-2866.032084 - (-2866.465927) + 1411.63352682 - 1412.07198721\} \\
 &= (-2.90 \text{ kcal/mol})
 \end{aligned}$$

$$\text{pKa} = \Delta G_{\text{exchange}}^* \div 2.303RT + \text{pKa}(\text{HRef}) = \mathbf{1.24}$$

30	9	0	-3.613241	4.338669	2.698734
31	9	0	-4.940336	2.387299	1.422877
32	9	0	-4.893427	0.248914	0.092374
33	15	0	-0.037229	-2.865463	-0.094837
34	8	0	0.751351	-3.839202	-0.863178
35	8	0	-1.149922	-3.450766	0.905153
36	1	0	-1.412577	-4.345229	0.629735
37	6	0	-3.629408	-1.948004	-1.092838
38	6	0	-3.362482	-2.369656	-2.404962
39	6	0	-4.613518	-2.622410	-0.352570
40	6	0	-4.063380	-3.438904	-2.961026
41	1	0	-2.606003	-1.857996	-2.990717
42	6	0	-5.310428	-3.693443	-0.910001
43	1	0	-4.829114	-2.307994	0.663630
44	6	0	-5.038205	-4.104894	-2.215979
45	1	0	-3.846498	-3.750356	-3.979011
46	1	0	-6.067672	-4.205433	-0.322445
47	1	0	-5.582783	-4.938612	-2.650748
48	6	0	3.531083	-2.118457	1.094442
49	6	0	4.498200	-2.847791	0.386916
50	6	0	3.228413	-2.492842	2.412707
51	6	0	5.145380	-3.928279	0.983565
52	1	0	4.735785	-2.573074	-0.635735
53	6	0	3.880815	-3.570650	3.008515
54	1	0	2.484585	-1.935056	2.972864
55	6	0	4.840302	-4.292042	2.295914
56	1	0	5.886954	-4.487471	0.419898
57	1	0	3.639164	-3.845546	4.031606
58	1	0	5.346336	-5.133932	2.760581

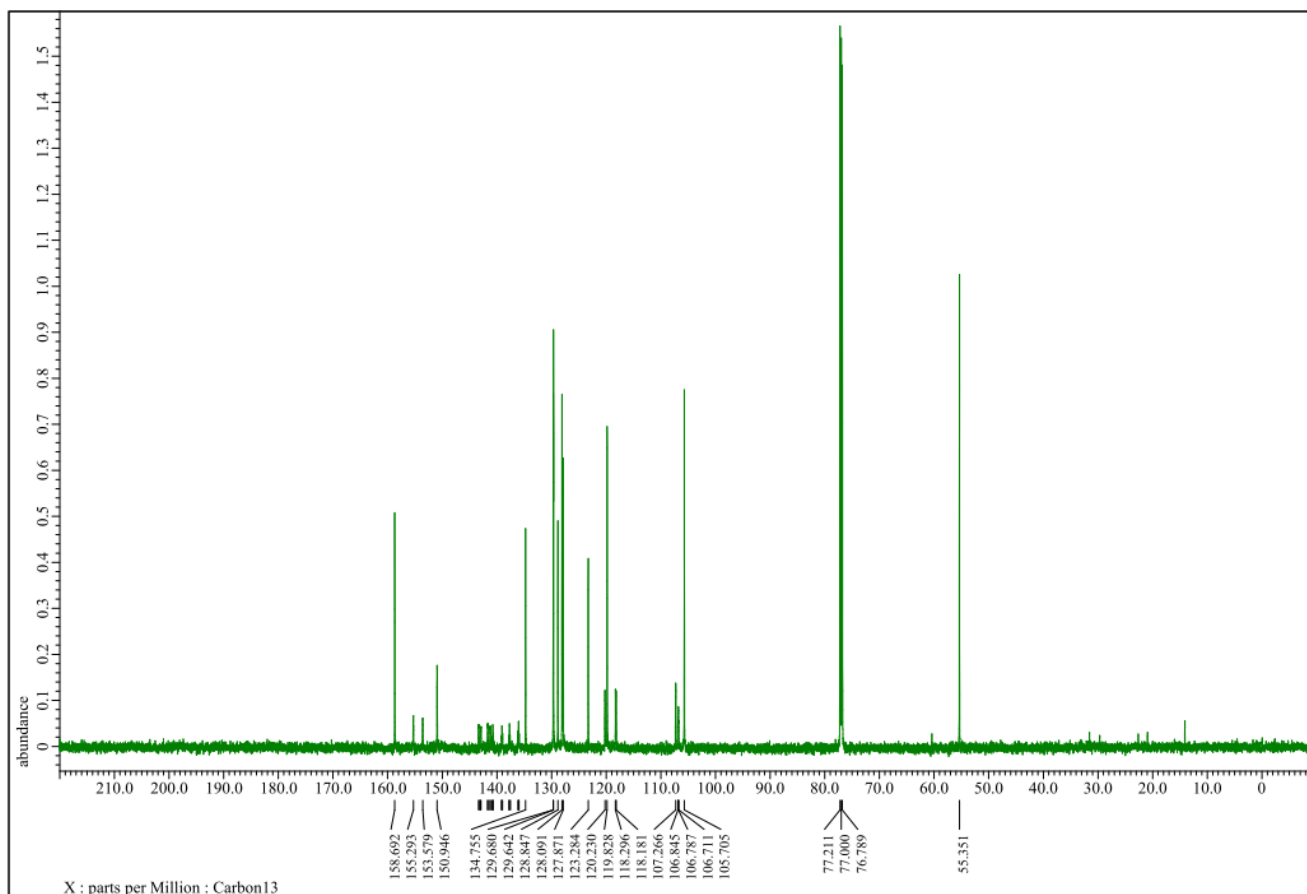
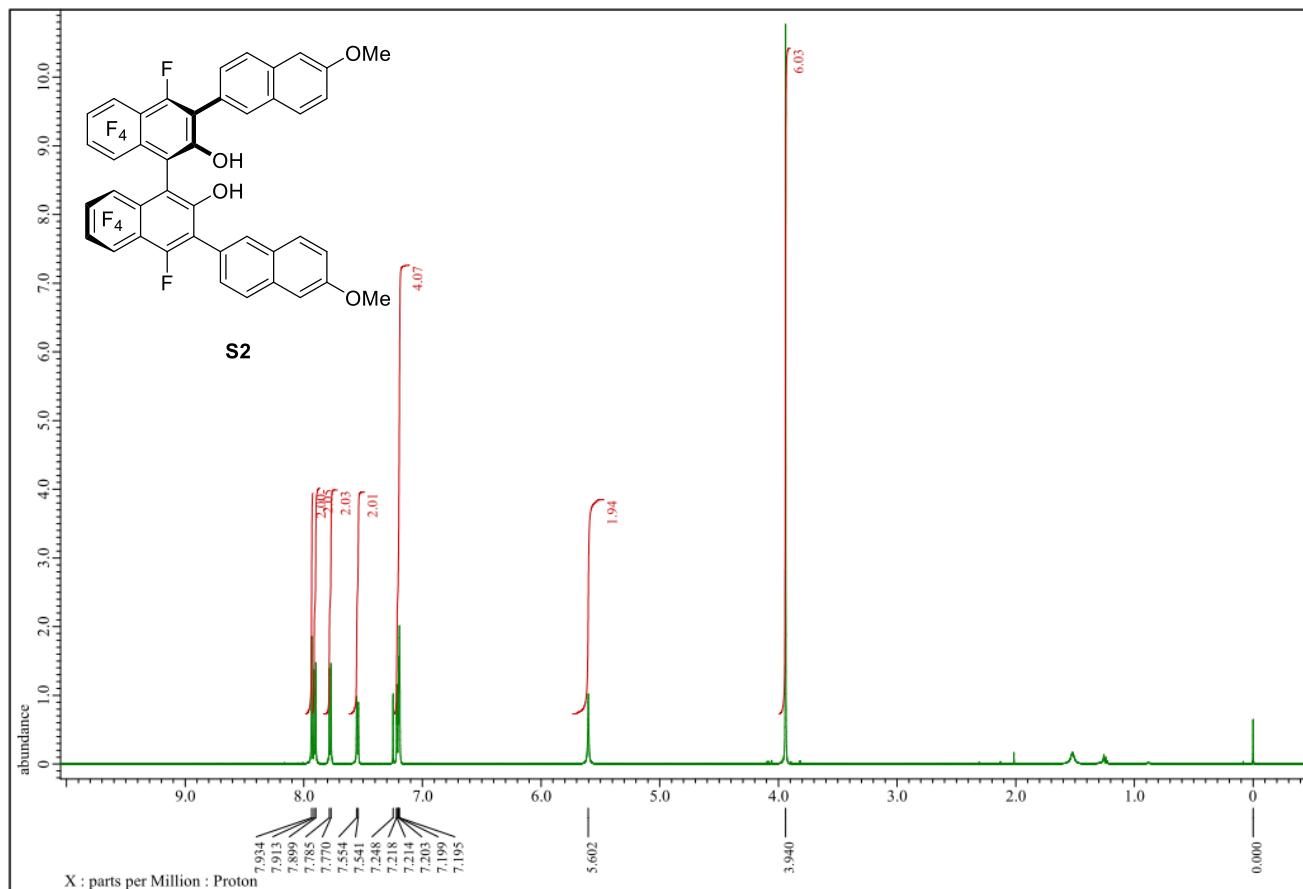
12	6	0	-1.489267	1.289644	0.759907
13	6	0	-1.433703	-0.813975	-0.460352
14	6	0	-0.894573	2.365754	1.472566
15	6	0	-2.930141	1.225521	0.783335
16	6	0	-2.862519	-0.900981	-0.462058
17	6	0	-1.636750	3.326482	2.117271
18	6	0	-3.668754	2.236908	1.452696
19	6	0	-3.552695	0.128510	0.134715
20	6	0	-3.039913	3.268107	2.105453
21	8	0	-0.733598	-1.814896	-1.036199
22	8	0	0.733579	-1.814867	1.036202
23	9	0	-4.904503	0.101902	0.116690
24	9	0	-5.013138	2.216084	1.486066
25	9	0	-3.744467	4.217865	2.744088
26	9	0	-1.032676	4.325488	2.780933
27	9	0	0.443566	2.490193	1.560242
28	9	0	-0.443577	2.490228	-1.560238
29	9	0	1.032667	4.325534	-2.780902
30	9	0	3.744453	4.217908	-2.744056
31	9	0	5.013125	2.216109	-1.486061
32	9	0	4.904494	0.101905	-0.116738
33	15	0	0.000054	-2.985848	0.000063
34	8	0	-1.043352	-3.631994	0.836329
35	8	0	1.043462	-3.631954	-0.836237
36	6	0	3.577304	-2.034516	1.107925
37	6	0	3.252511	-2.434868	2.413324
38	6	0	4.606155	-2.710204	0.434930
39	6	0	3.946002	-3.474656	3.029925
40	1	0	2.445369	-1.936614	2.938337
41	6	0	5.295355	-3.752877	1.051111
42	1	0	4.854852	-2.425236	-0.581734
43	6	0	4.971069	-4.137195	2.352853
44	1	0	3.677178	-3.772329	4.040417
45	1	0	6.082076	-4.270961	0.508064
46	1	0	5.506283	-4.953350	2.832522
47	6	0	-3.577312	-2.034521	-1.107960
48	6	0	-4.606192	-2.710185	-0.434985
49	6	0	-3.252494	-2.434895	-2.413347
50	6	0	-5.295393	-3.752856	-1.051171
51	1	0	-4.854914	-2.425200	0.581668
52	6	0	-3.945986	-3.474677	-3.029953
53	1	0	-2.445328	-1.936661	-2.938343
54	6	0	-4.971080	-4.137195	-2.352899
55	1	0	-6.082136	-4.270921	-0.508137
56	1	0	-3.677142	-3.772367	-4.040435
57	1	0	-5.506295	-4.953346	-2.832573

(A)-3a-PA

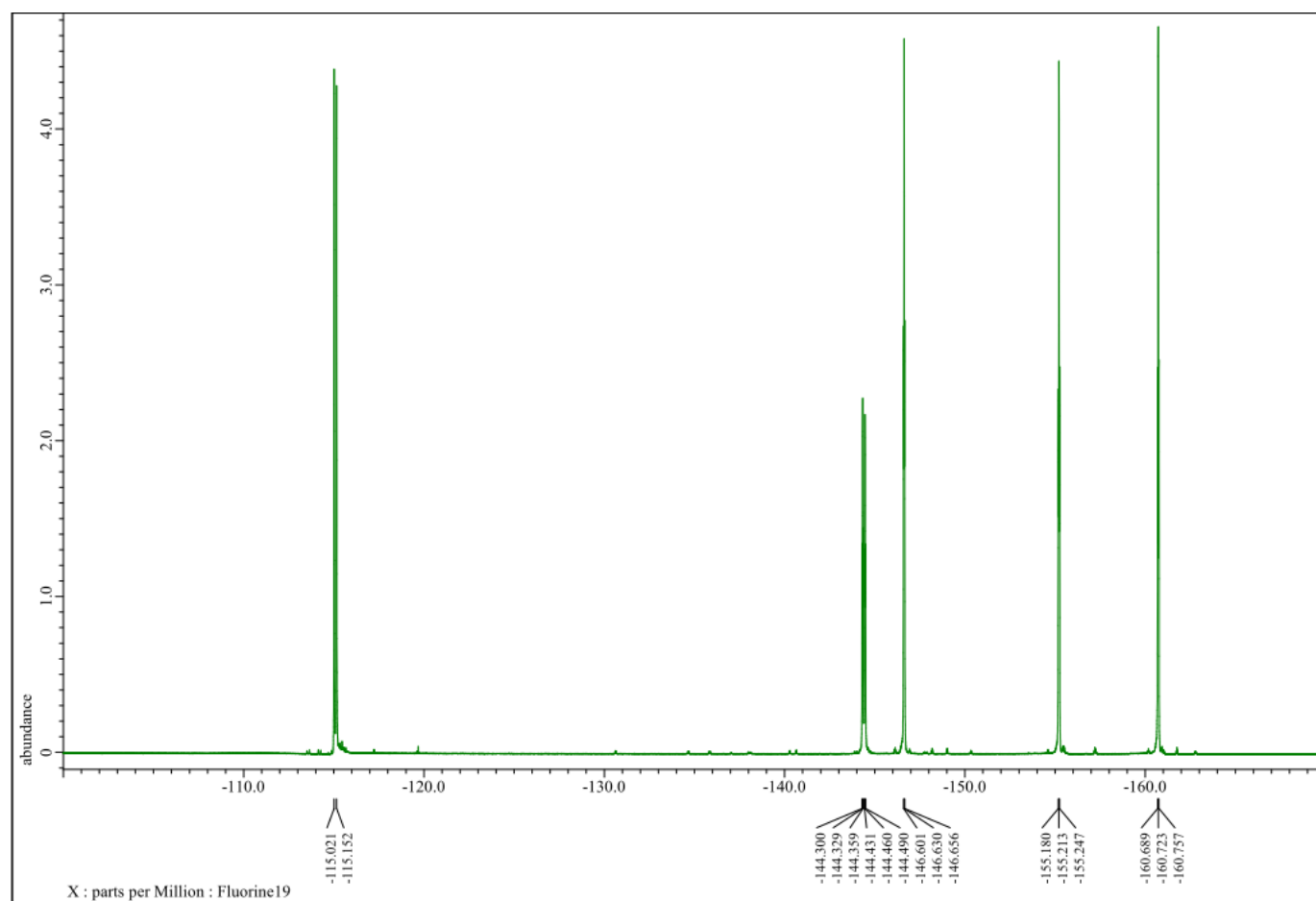
Center Number	Atomic Number	Atomic Type	Coordinates (Angstroms)		
			X	Y	Z
1	6	0	1.636738	3.326520	-2.117250
2	6	0	0.894558	2.365787	-1.472557
3	6	0	1.489253	1.289669	-0.759909
4	6	0	2.930126	1.225539	-0.783342
5	6	0	3.668743	2.236936	-1.452691
6	6	0	3.039901	3.268143	-2.105432
7	6	0	0.745461	0.278699	-0.068998
8	6	0	3.552687	0.128521	-0.134745
9	6	0	2.862512	-0.900974	0.462029
10	6	0	1.433711	-0.813938	0.460333
11	6	0	-0.745477	0.278685	0.068991

7. NMR Spectra Data

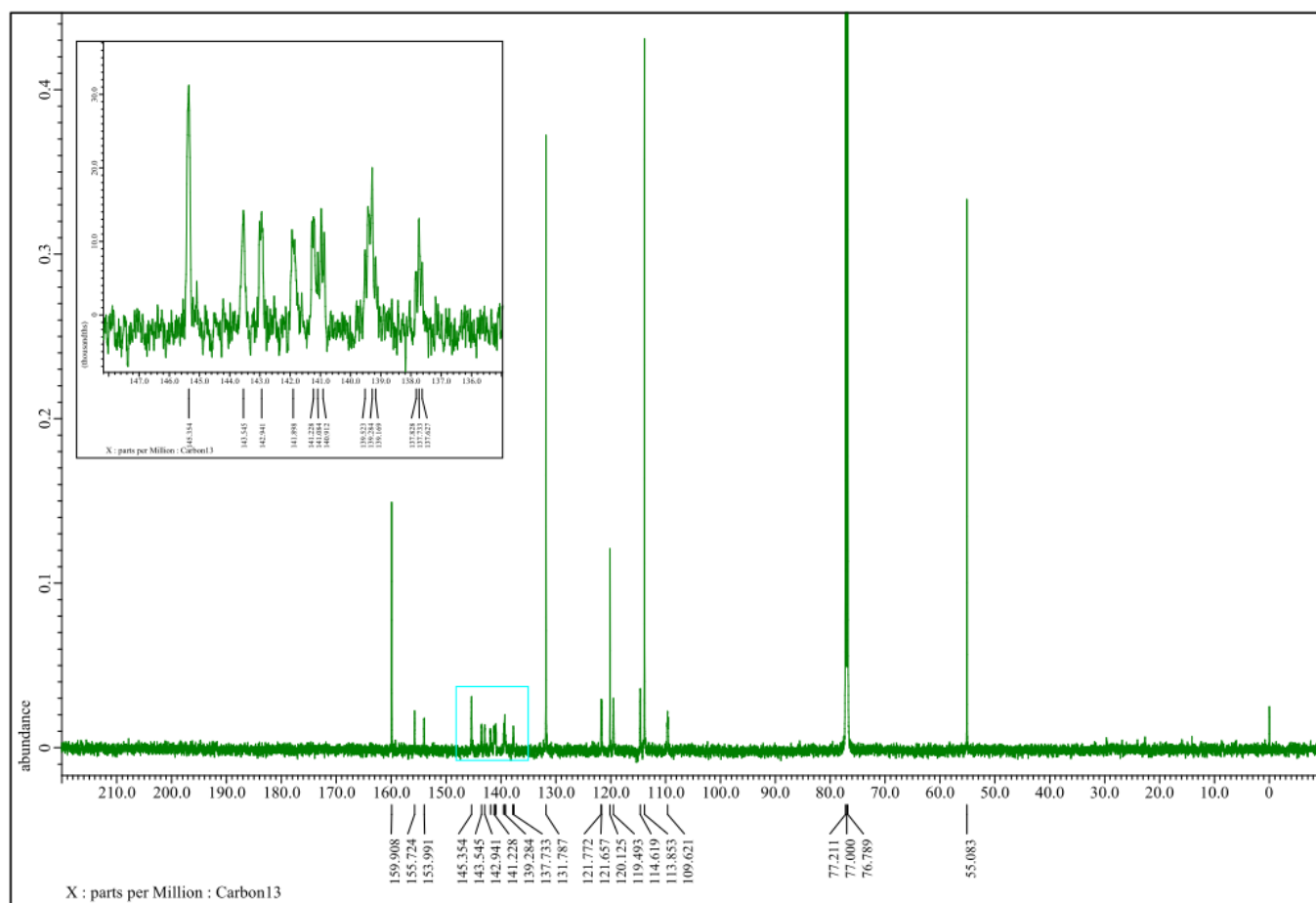
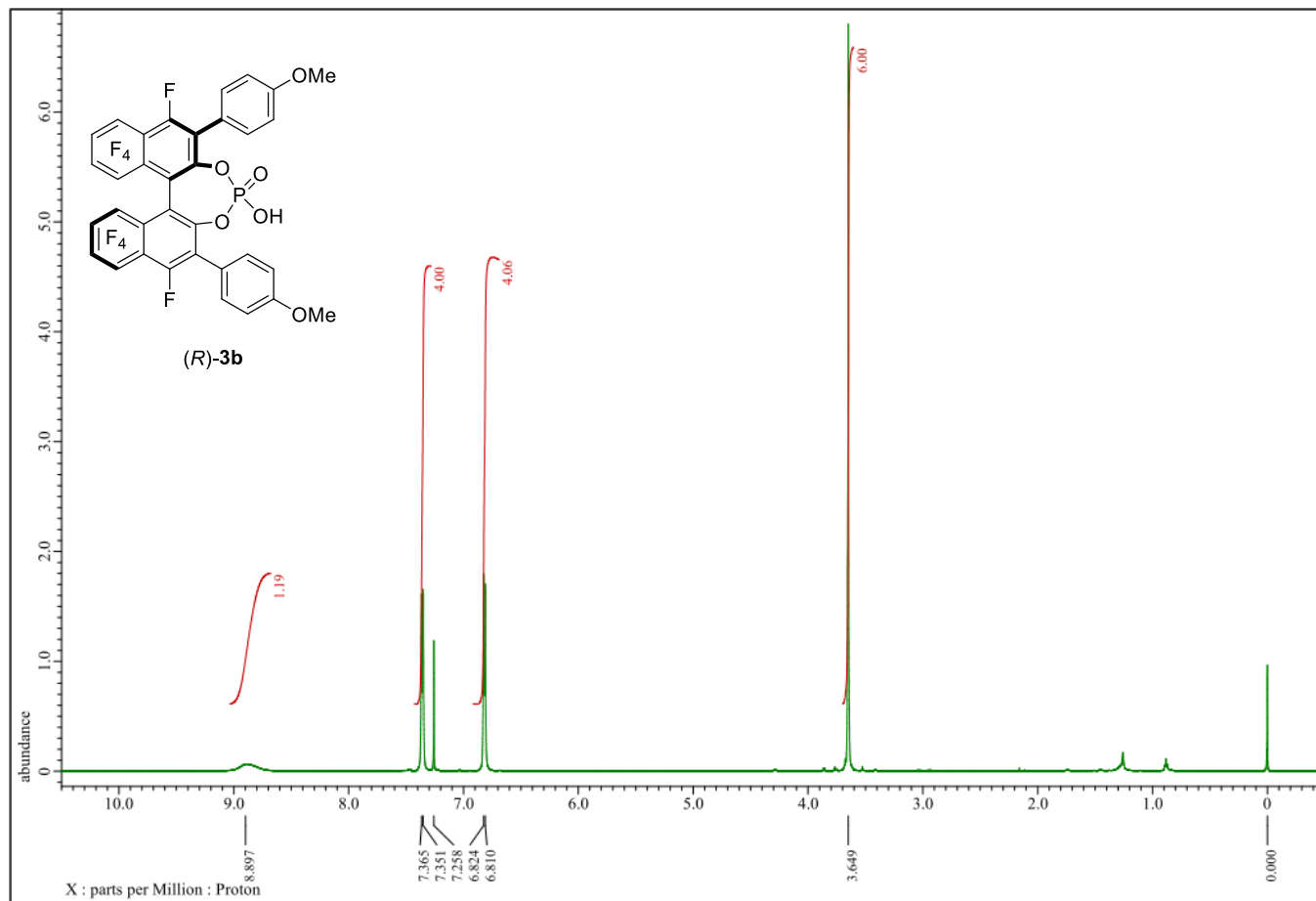
^1H NMR (600 MHz) and ^{13}C NMR (150 MHz) spectra of **S2**



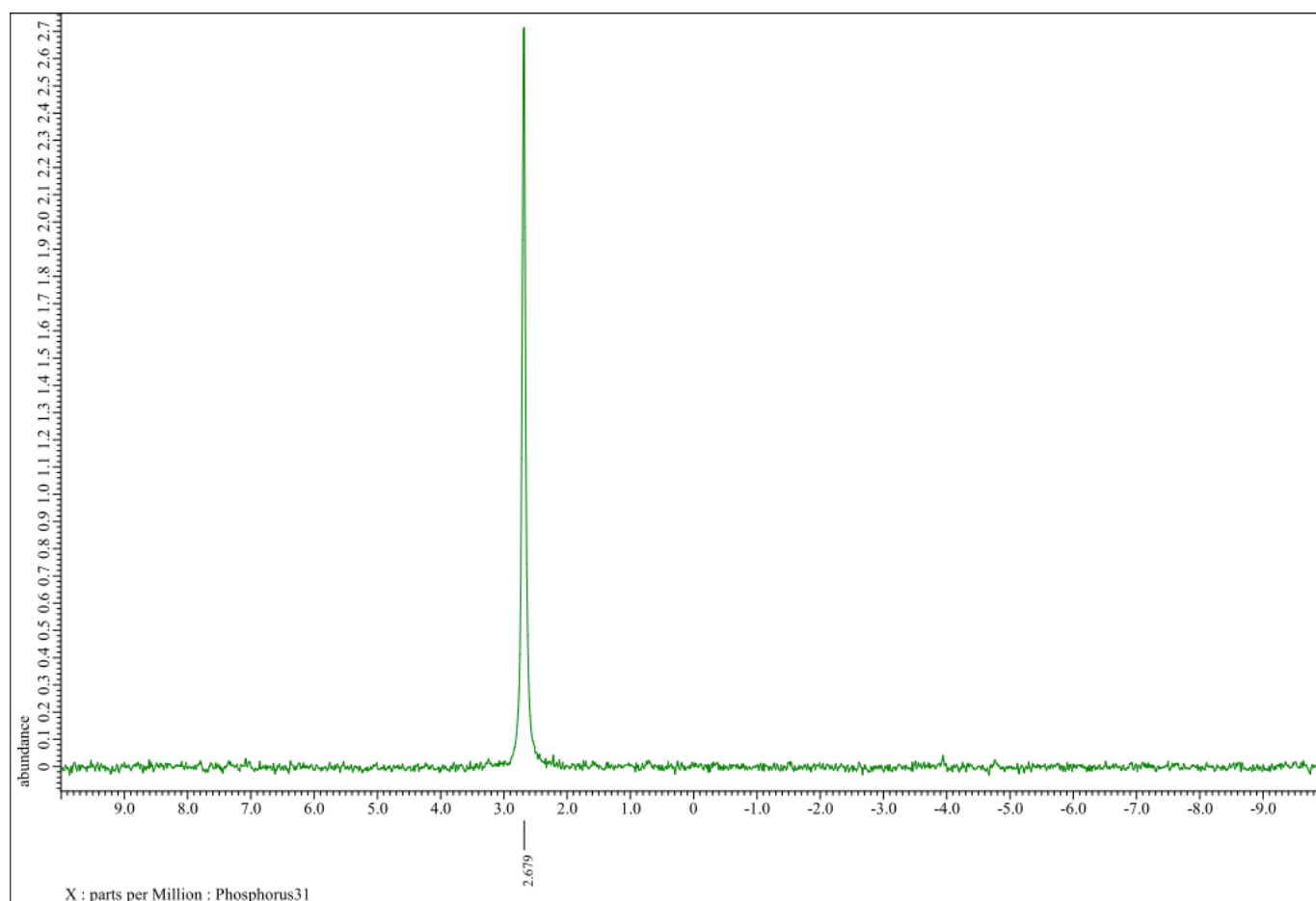
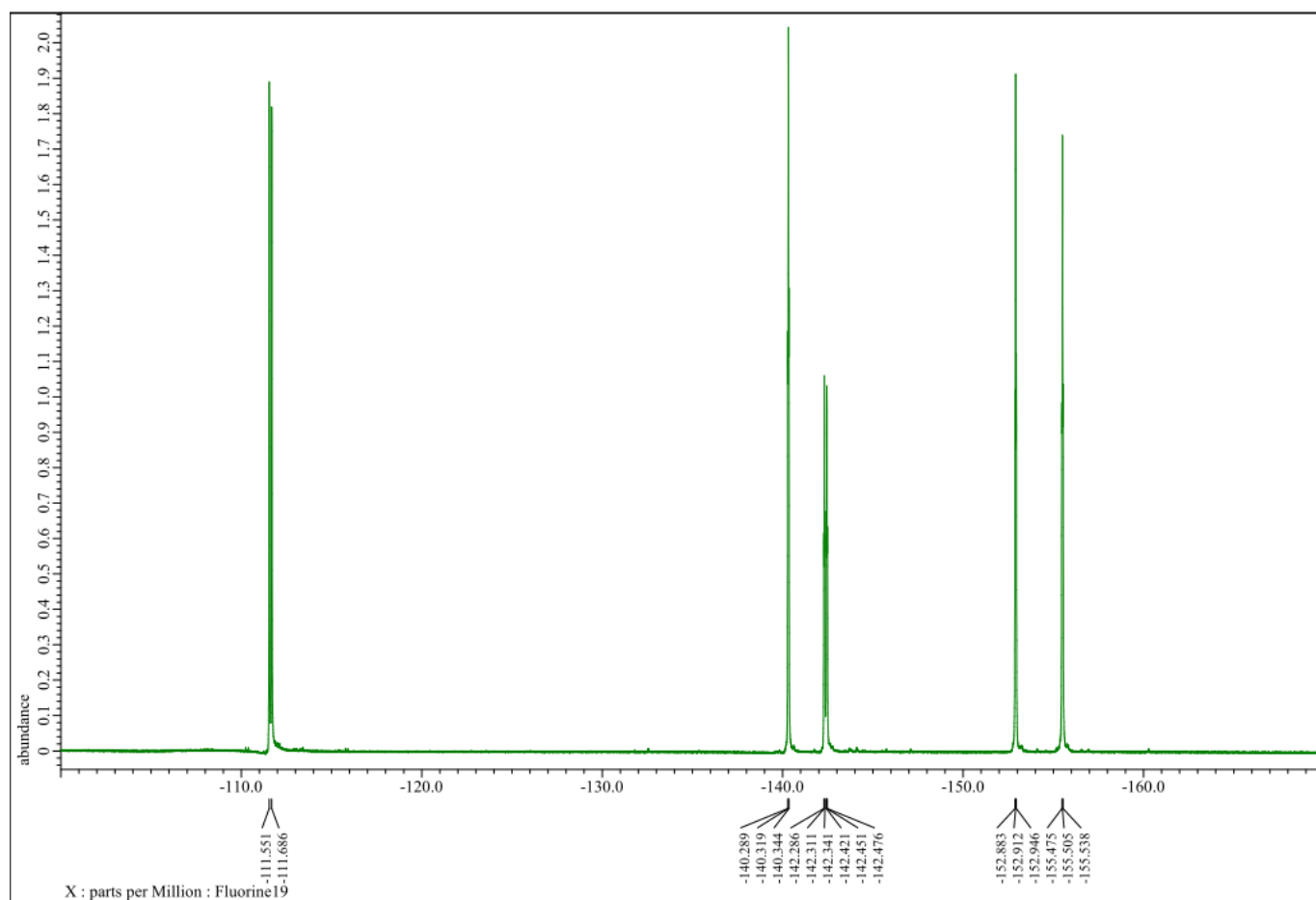
^{19}F NMR (565 MHz) spectra of **S2**



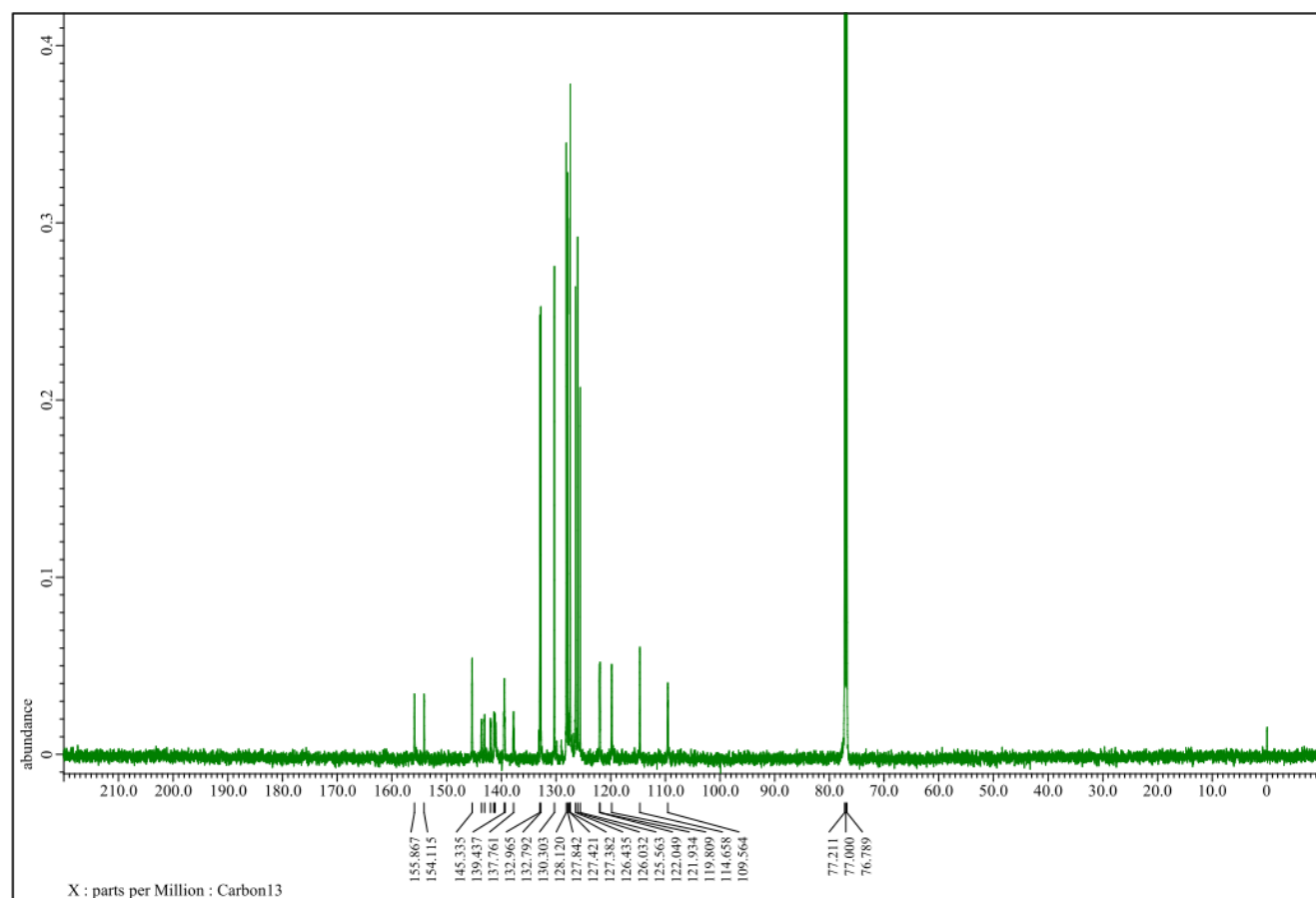
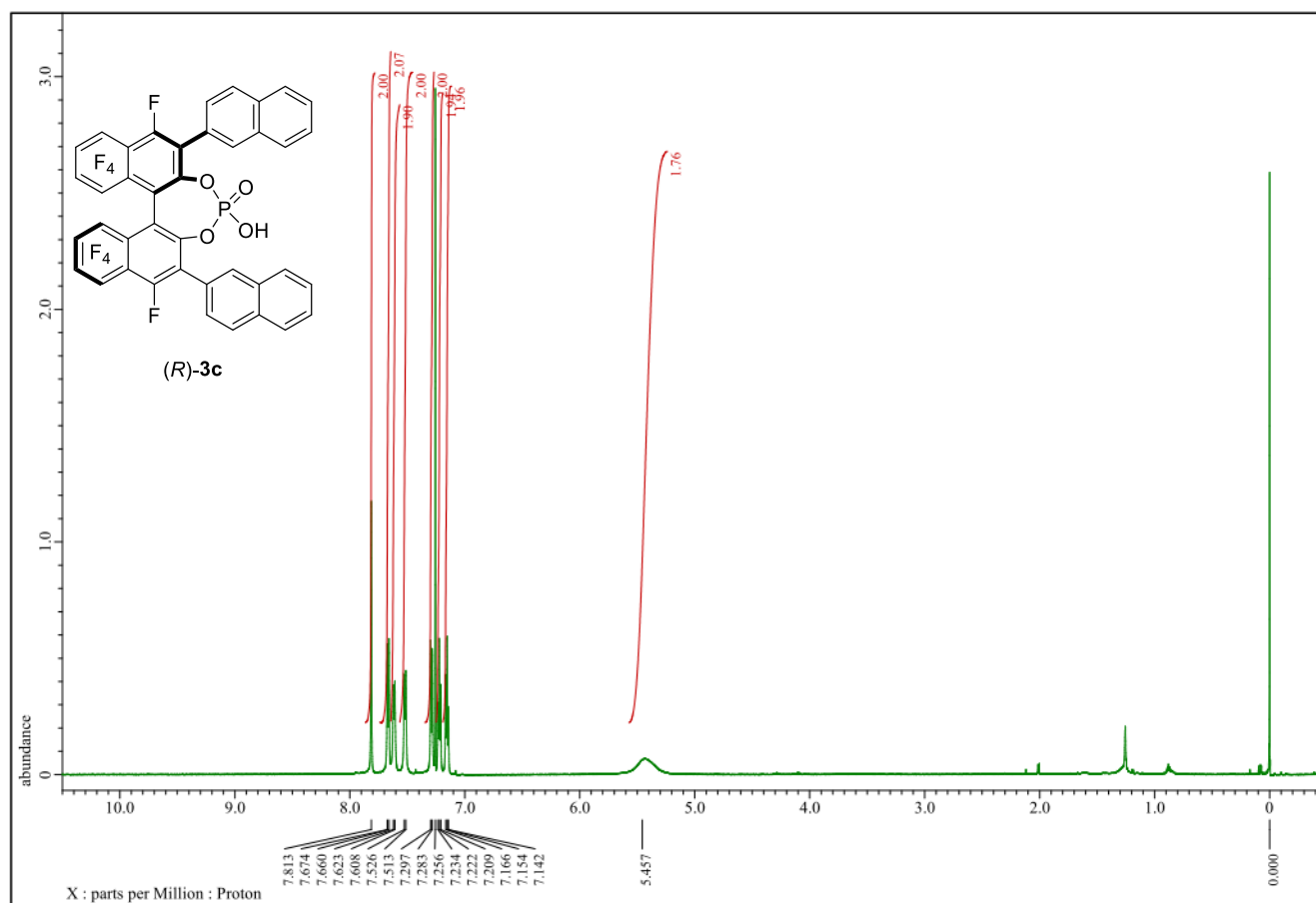
^1H NMR (600 MHz) and ^{13}C NMR (150 MHz) spectra of (*R*)-**3b**



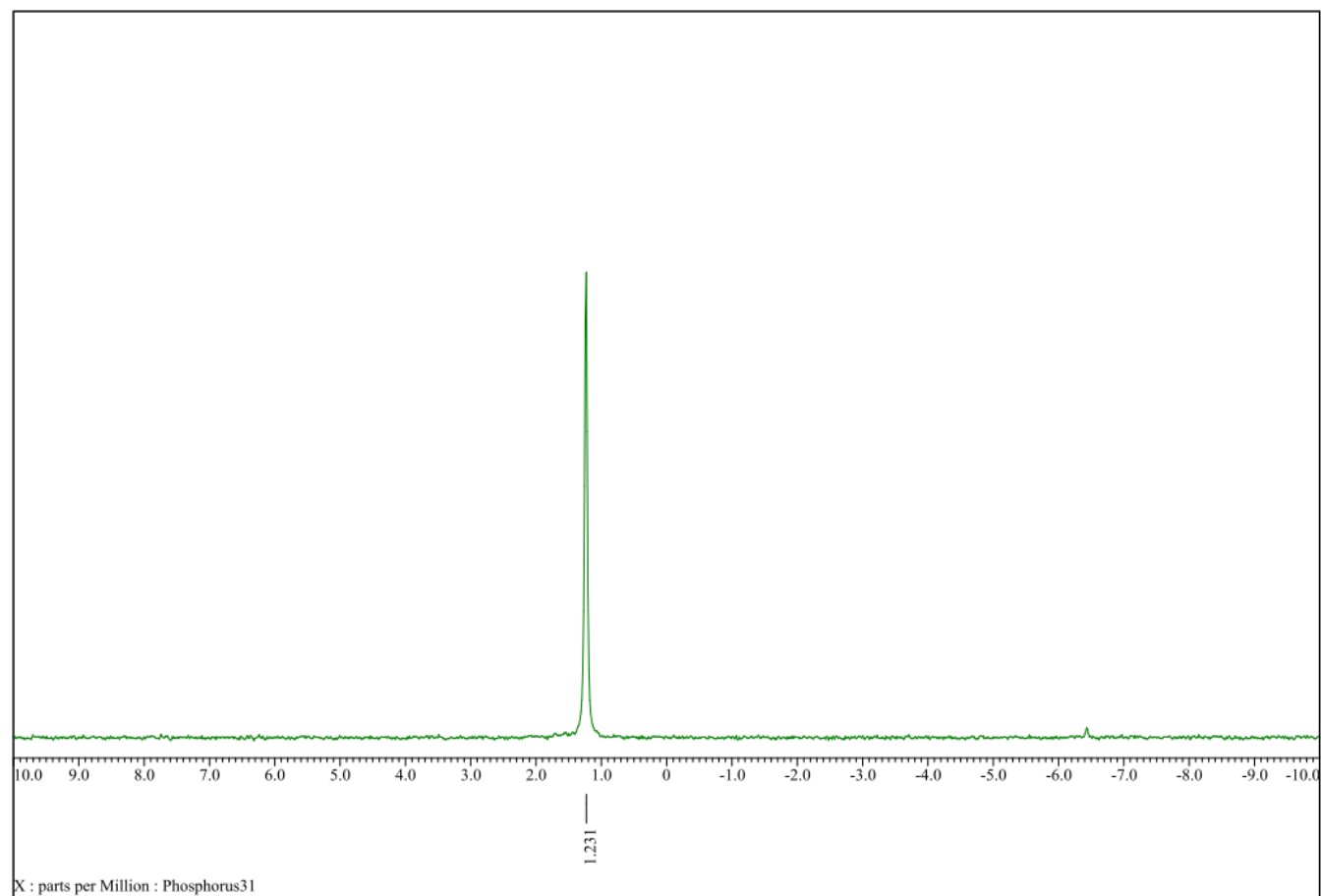
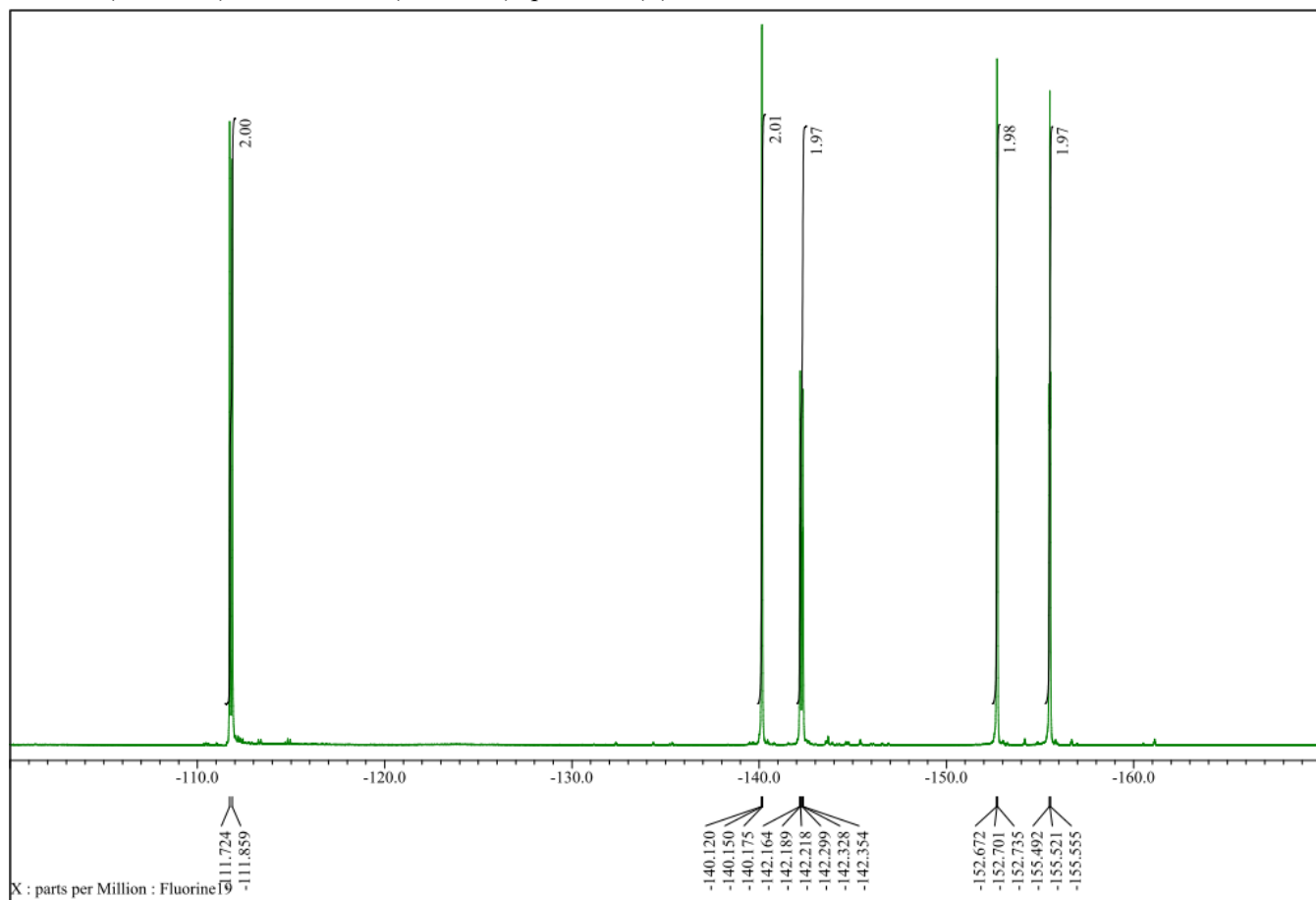
^{19}F NMR (565 MHz) and ^{31}P NMR (243 MHz) spectra of (*R*)-**3b**



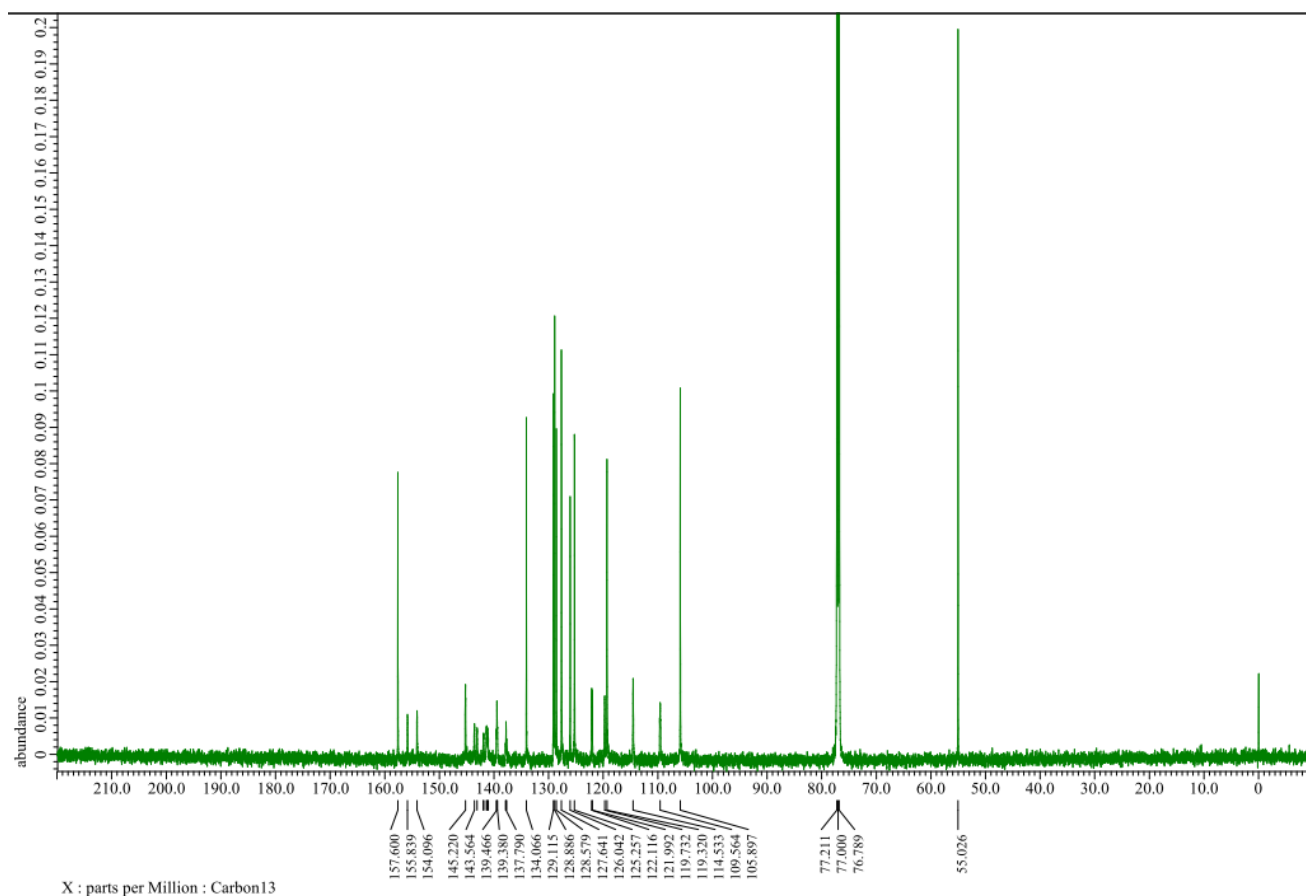
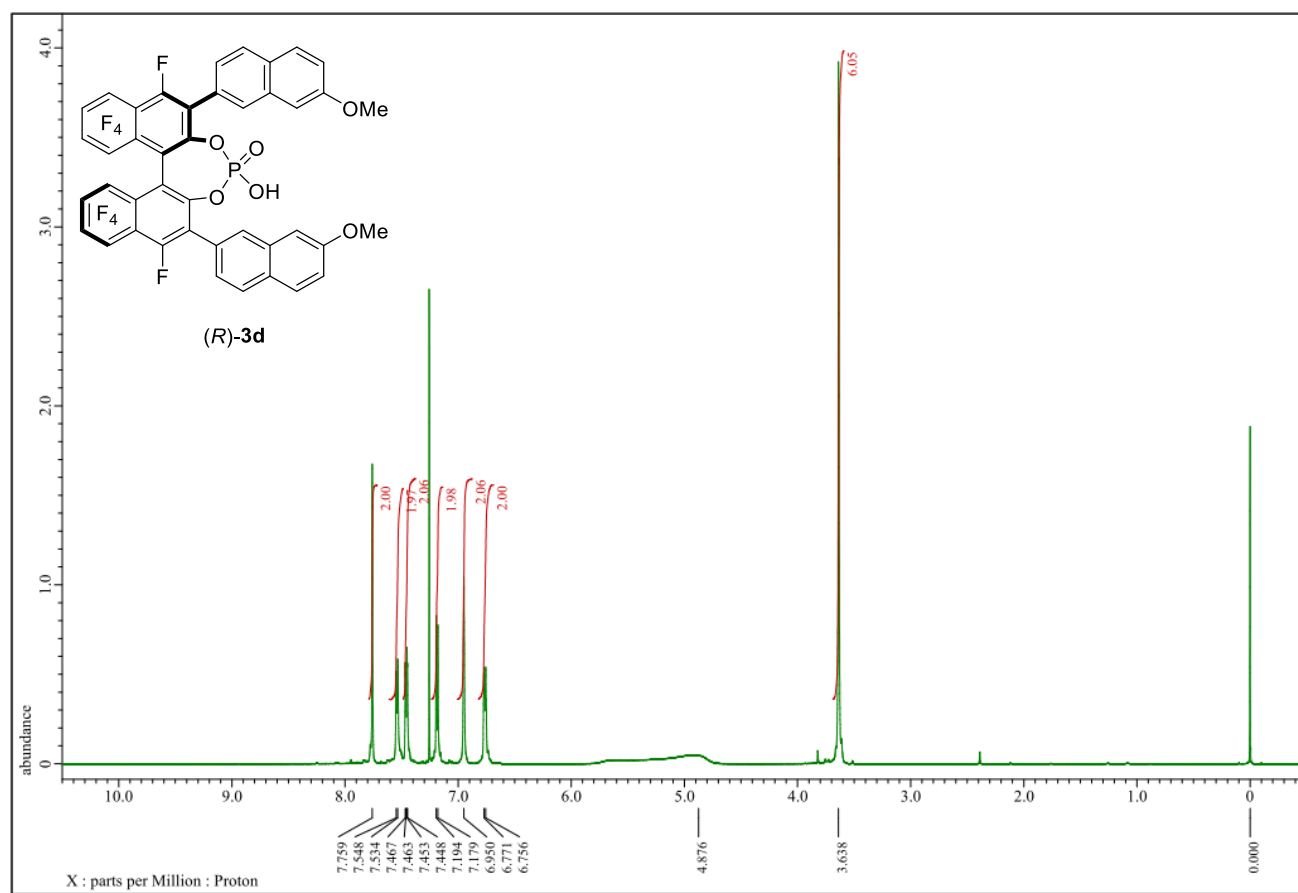
^1H NMR (600 MHz) and ^{13}C NMR (150 MHz) spectra of (*R*)-**3c**



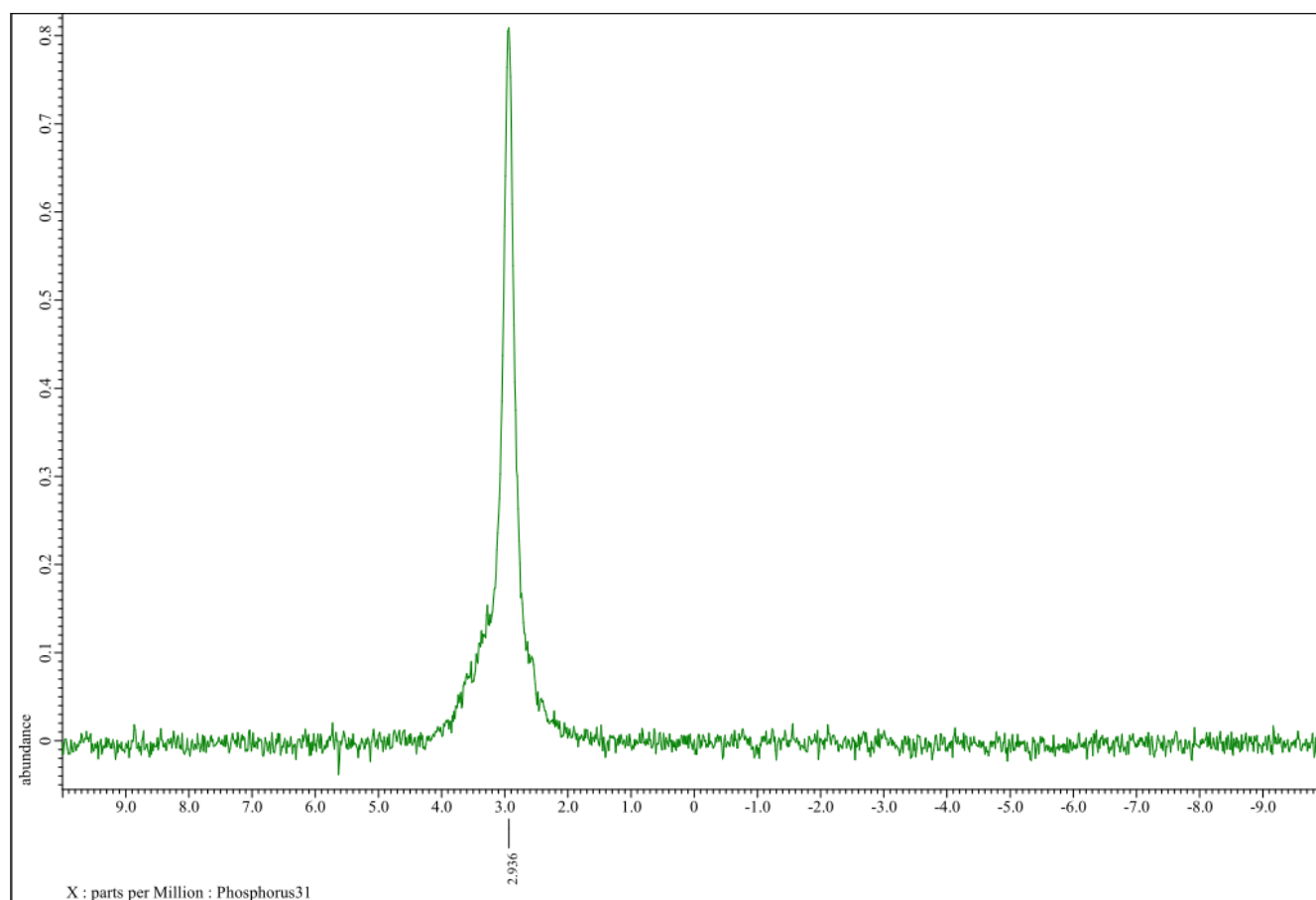
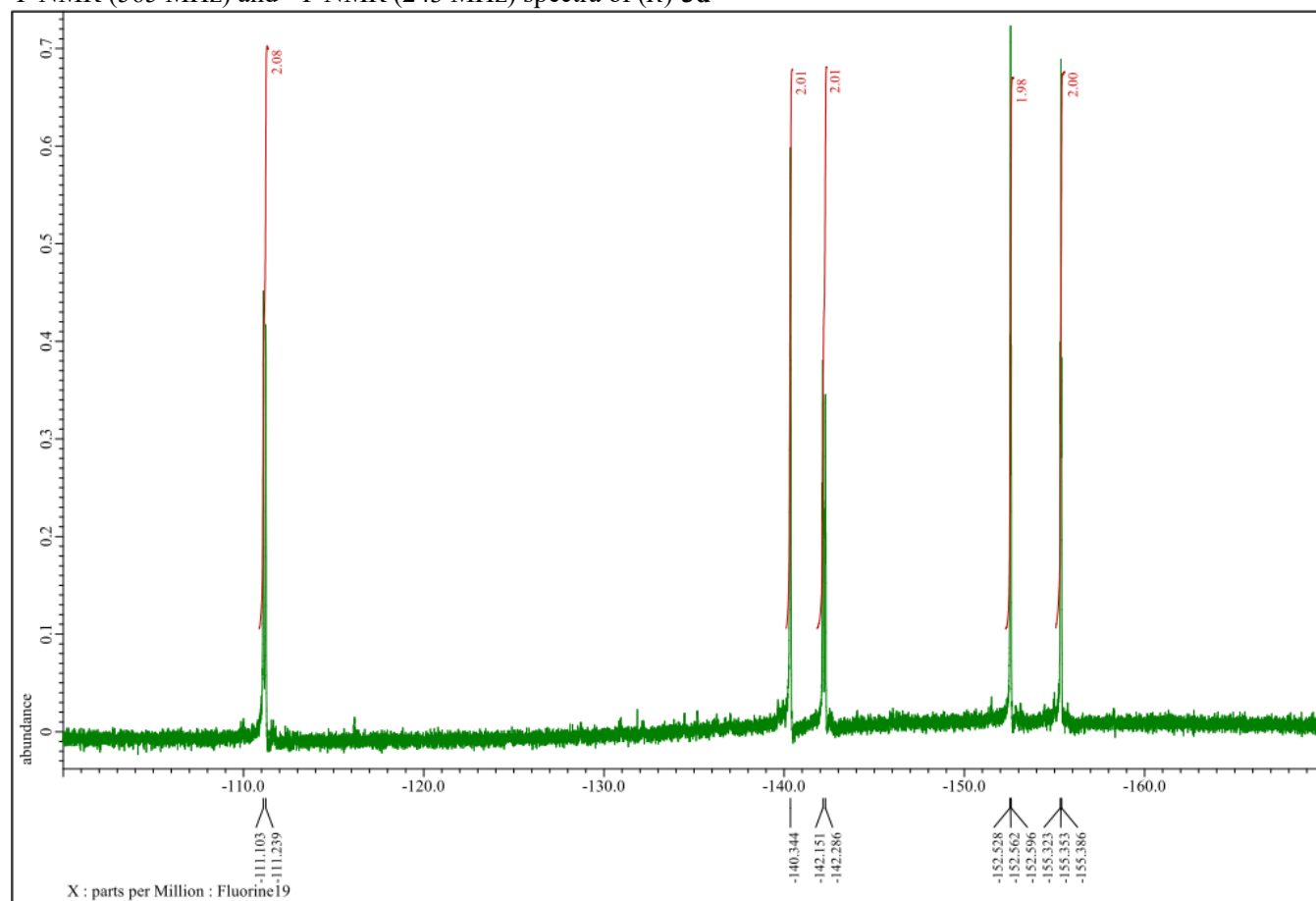
^{19}F NMR (565 MHz) and ^{31}P NMR (243 MHz) spectra of (*R*)-**3c**



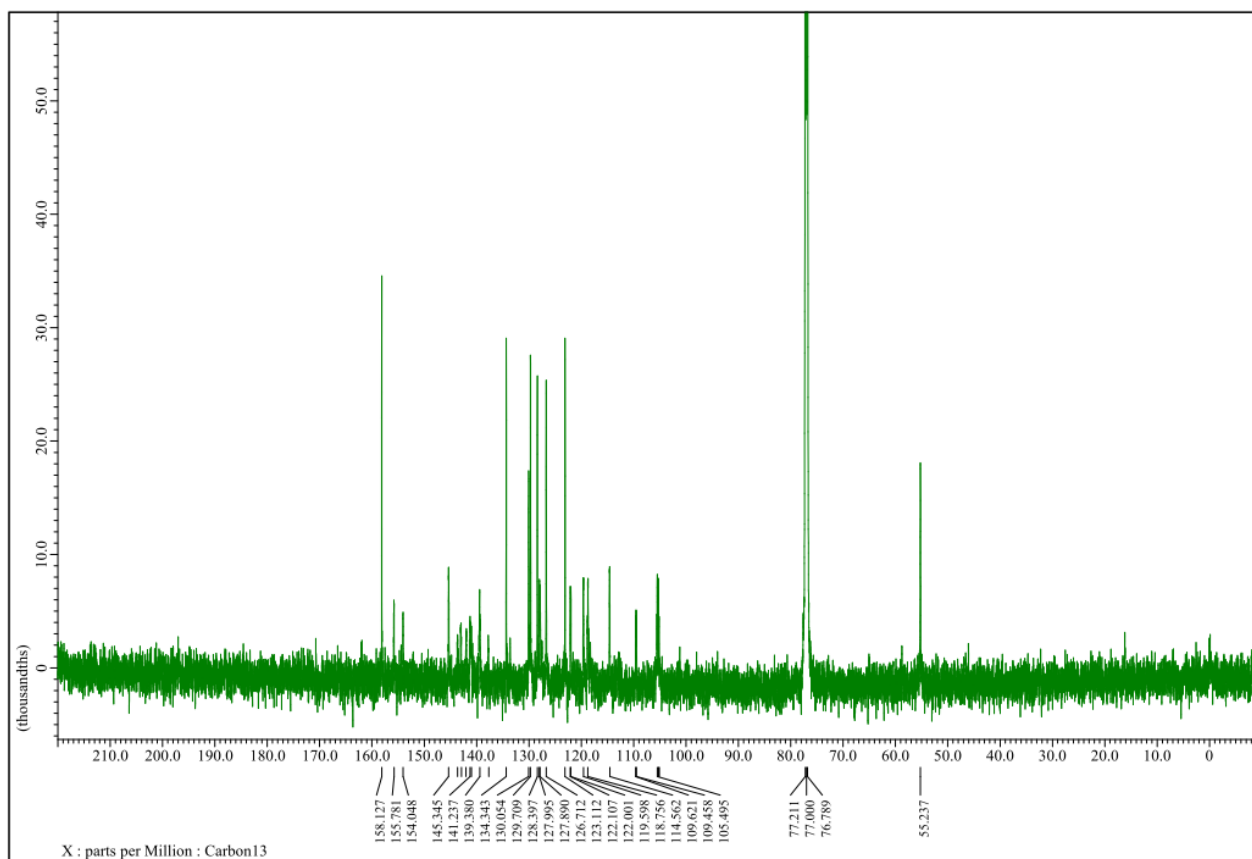
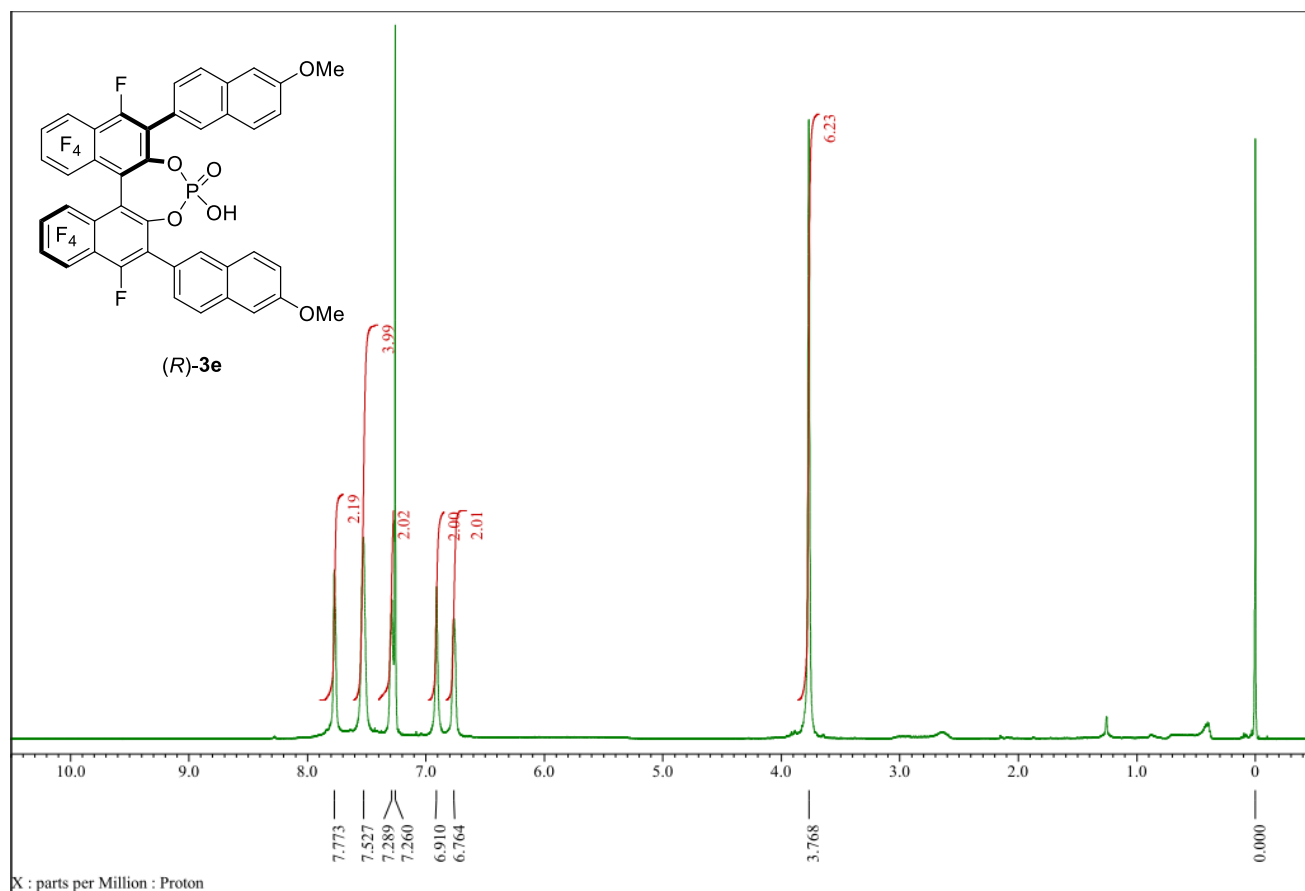
^1H NMR (600 MHz) and ^{13}C NMR (150 MHz) spectra of (*R*)-**3d**



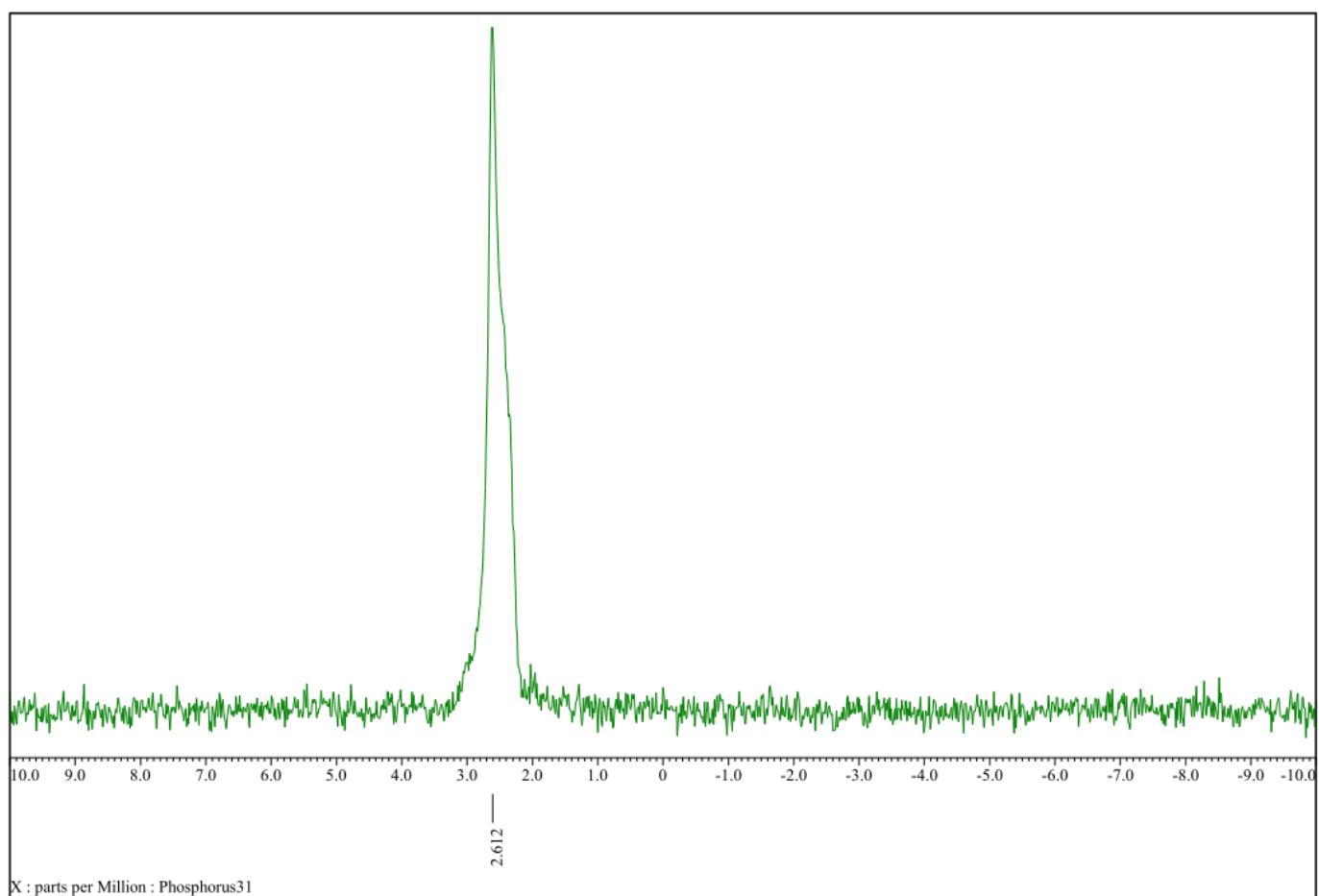
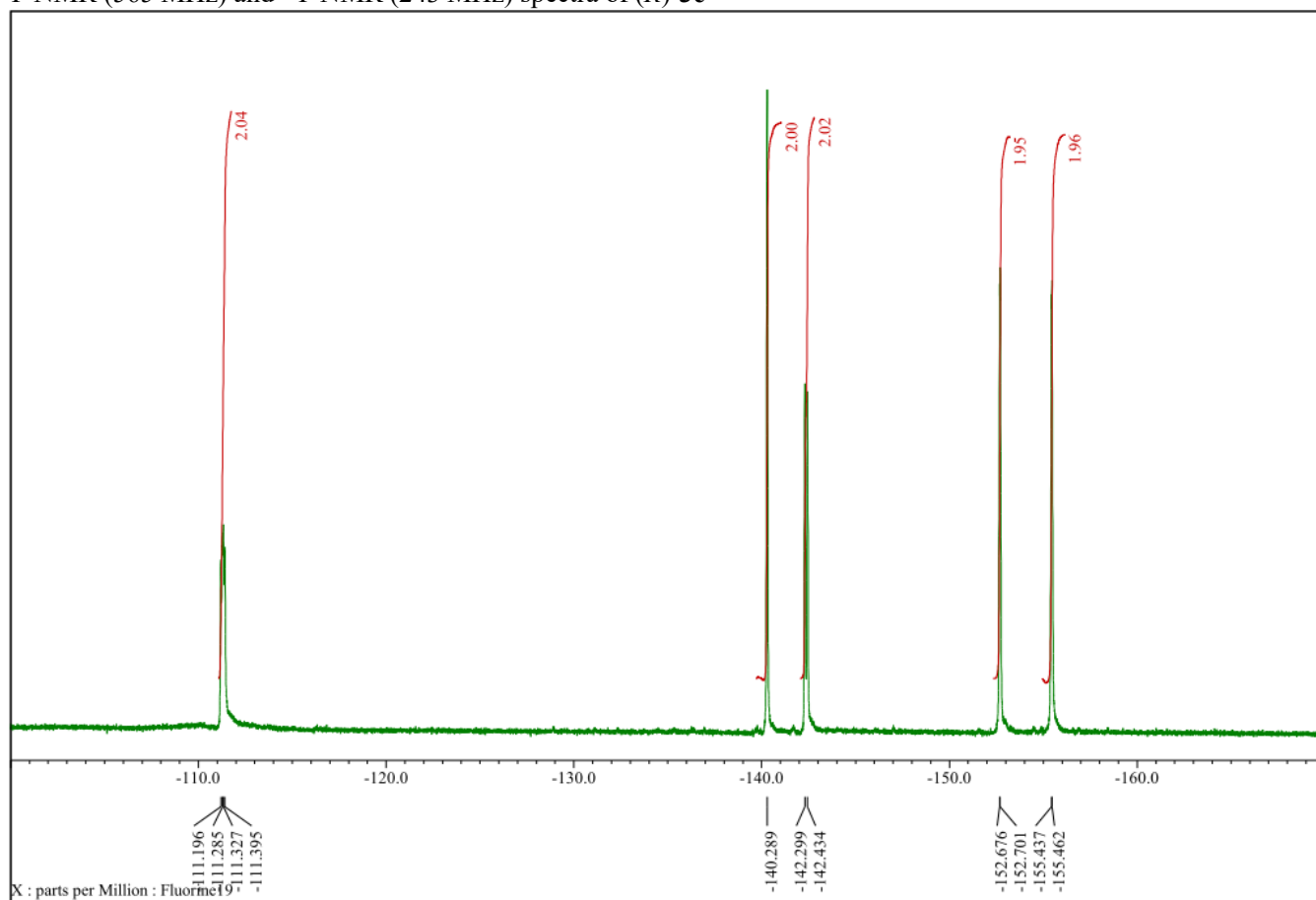
^{19}F NMR (565 MHz) and ^{31}P NMR (243 MHz) spectra of (*R*)-**3d**



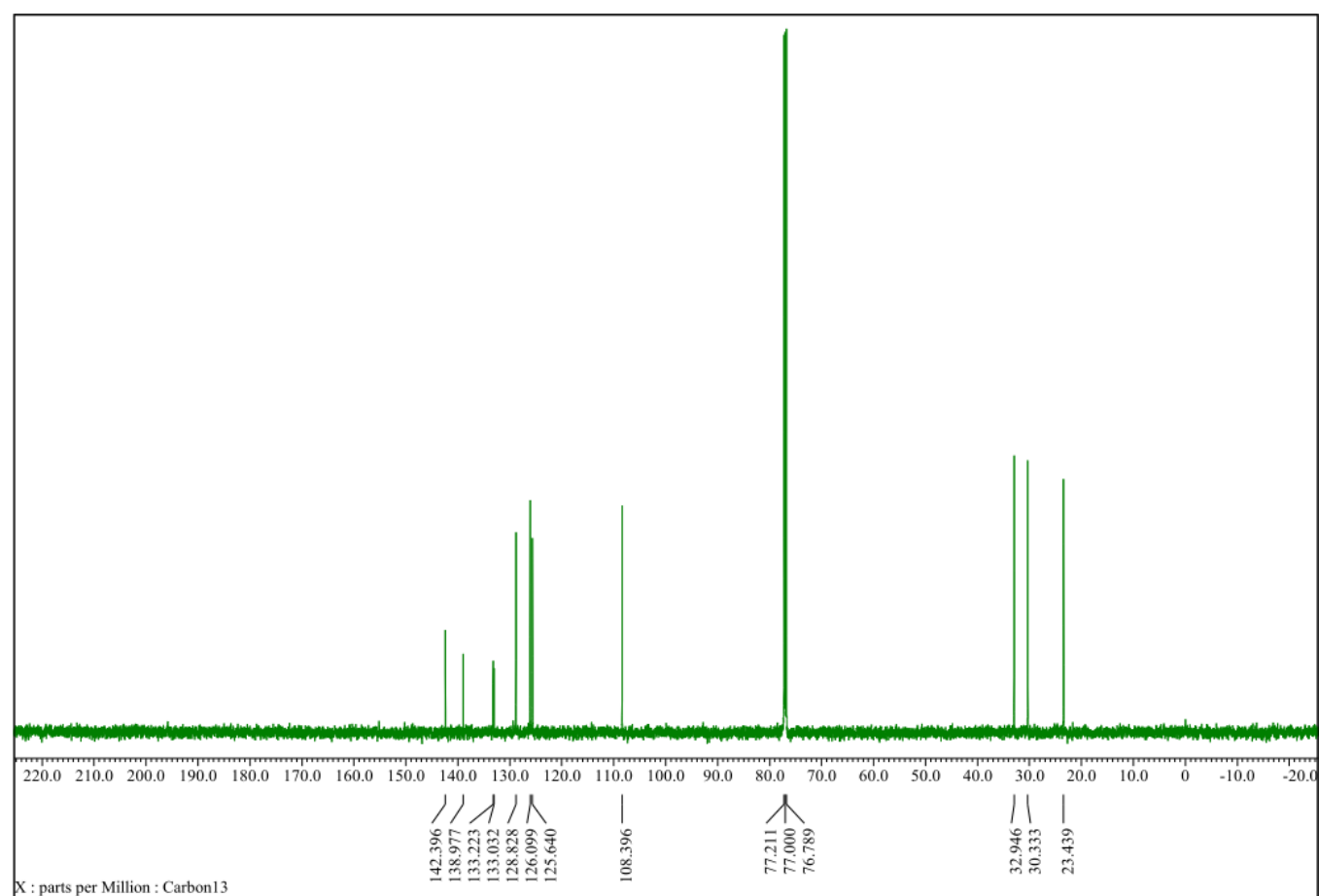
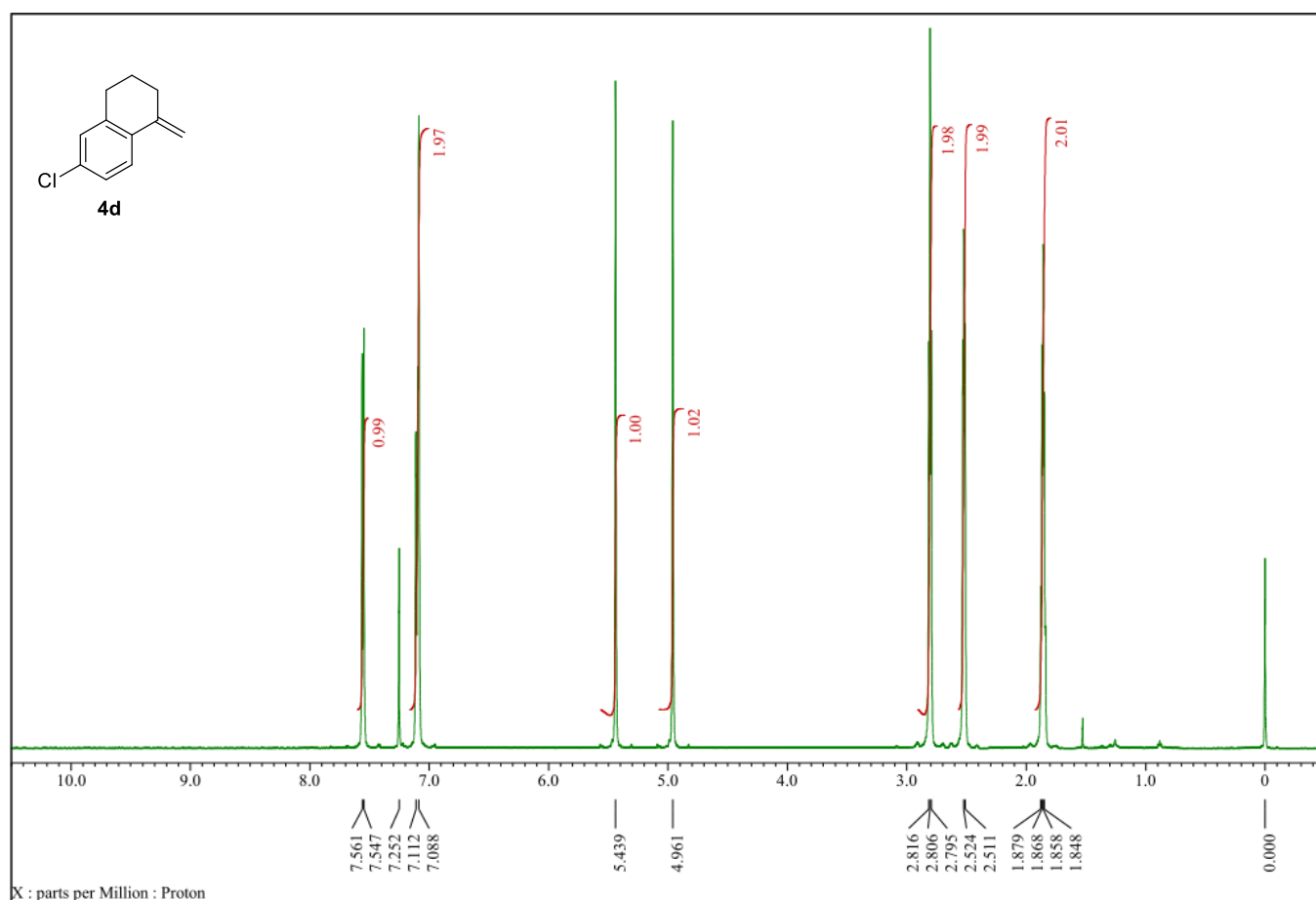
^1H NMR (600 MHz) and ^{13}C NMR (150 MHz) spectra of (*R*)-**3e**



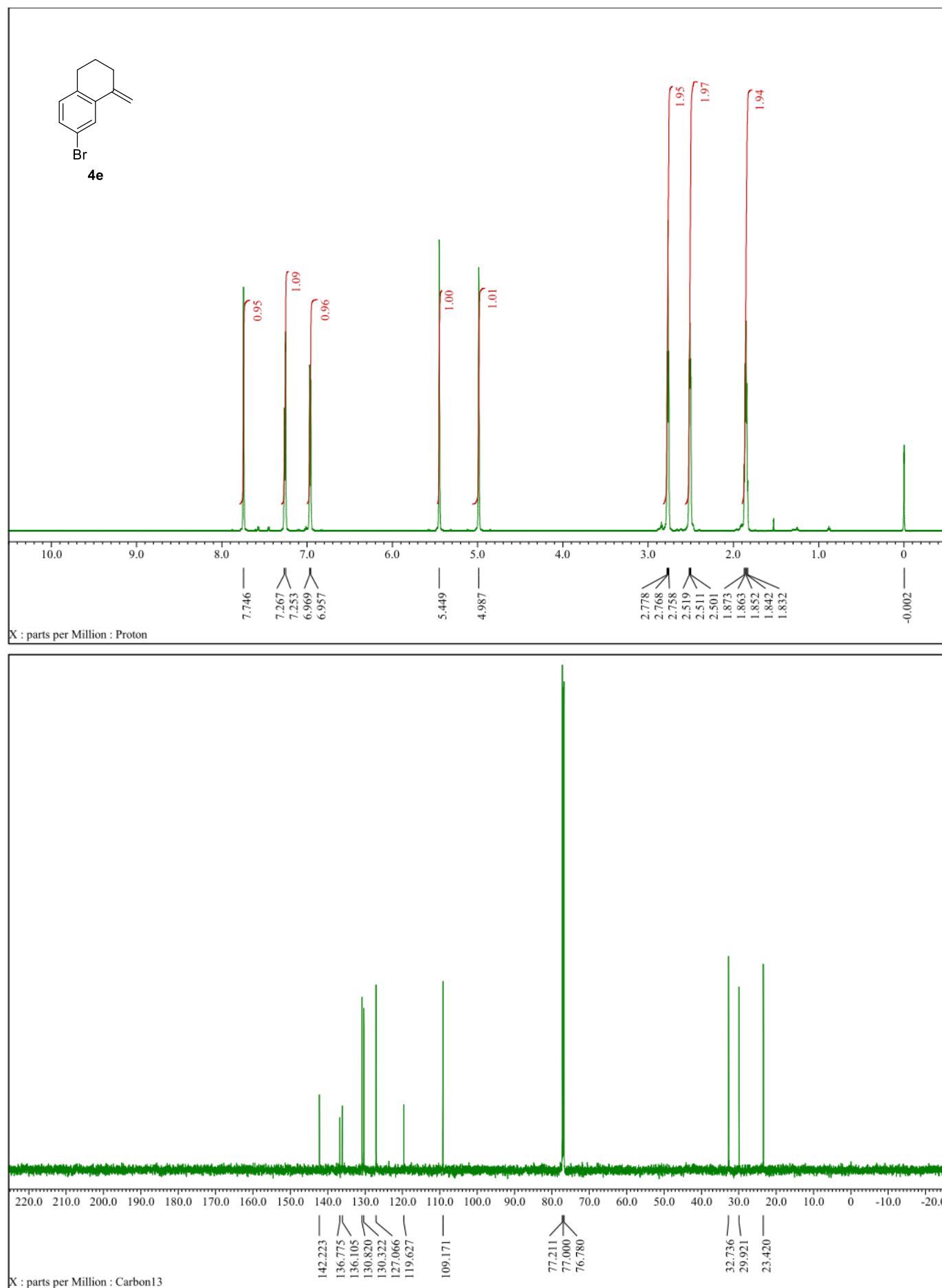
^{19}F NMR (565 MHz) and ^{31}P NMR (243 MHz) spectra of (*R*)-**3e**



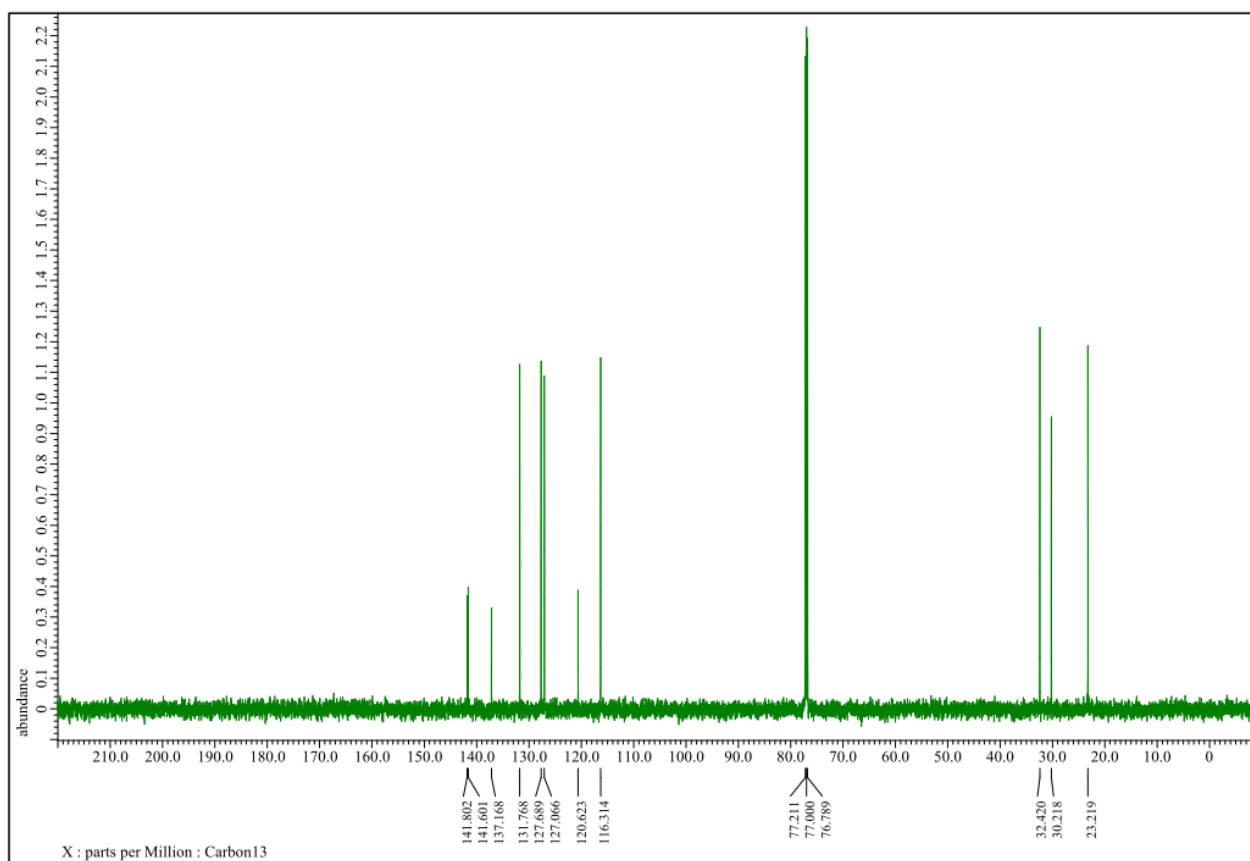
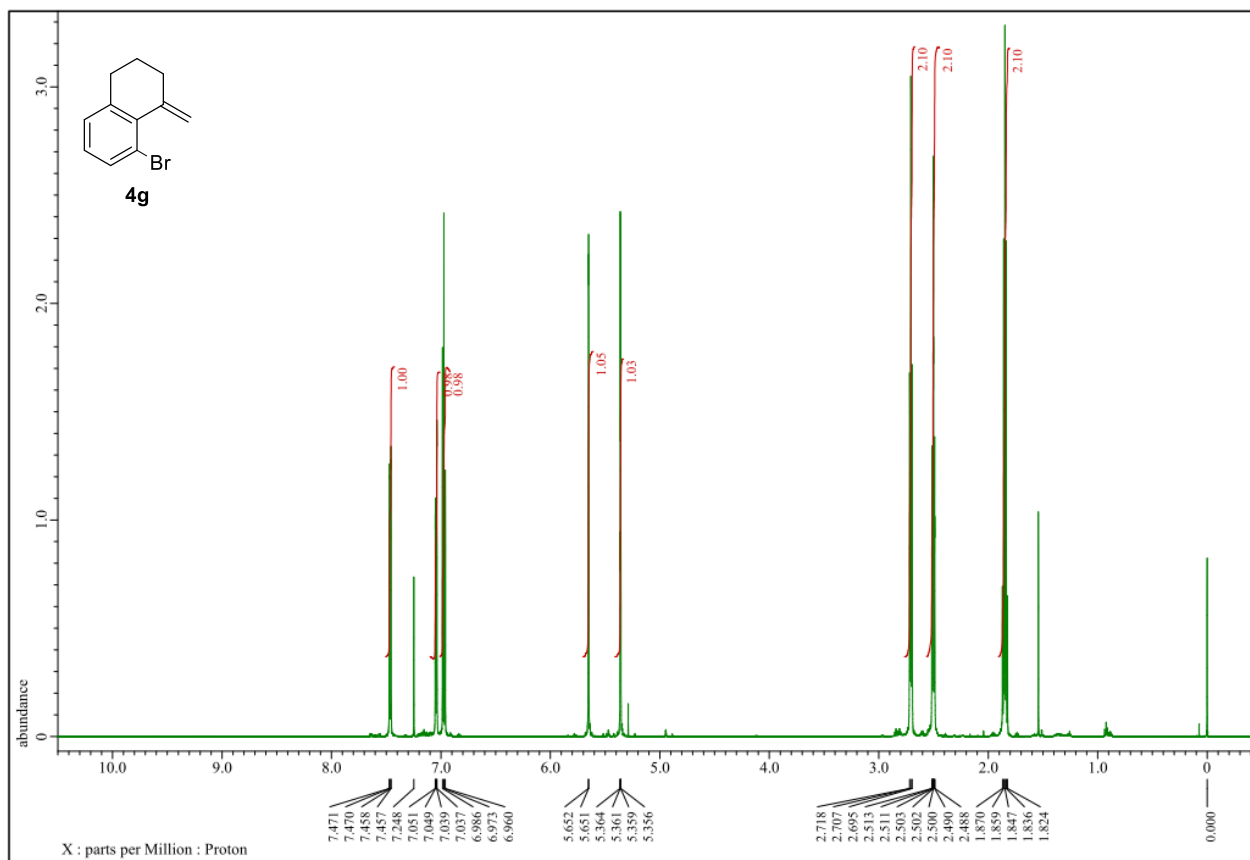
^1H NMR (600 MHz) and ^{13}C NMR (150 MHz) spectra of **4d**



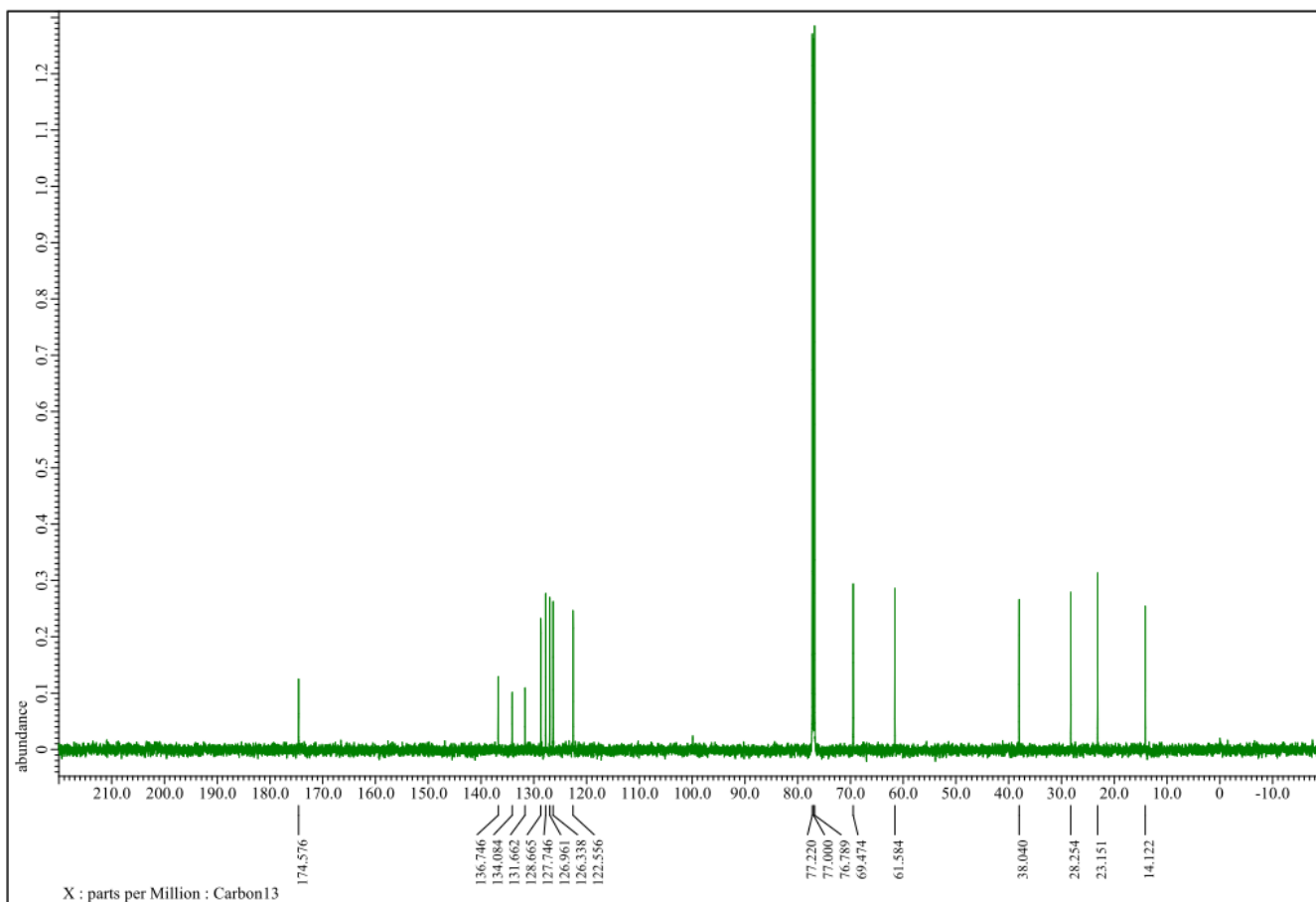
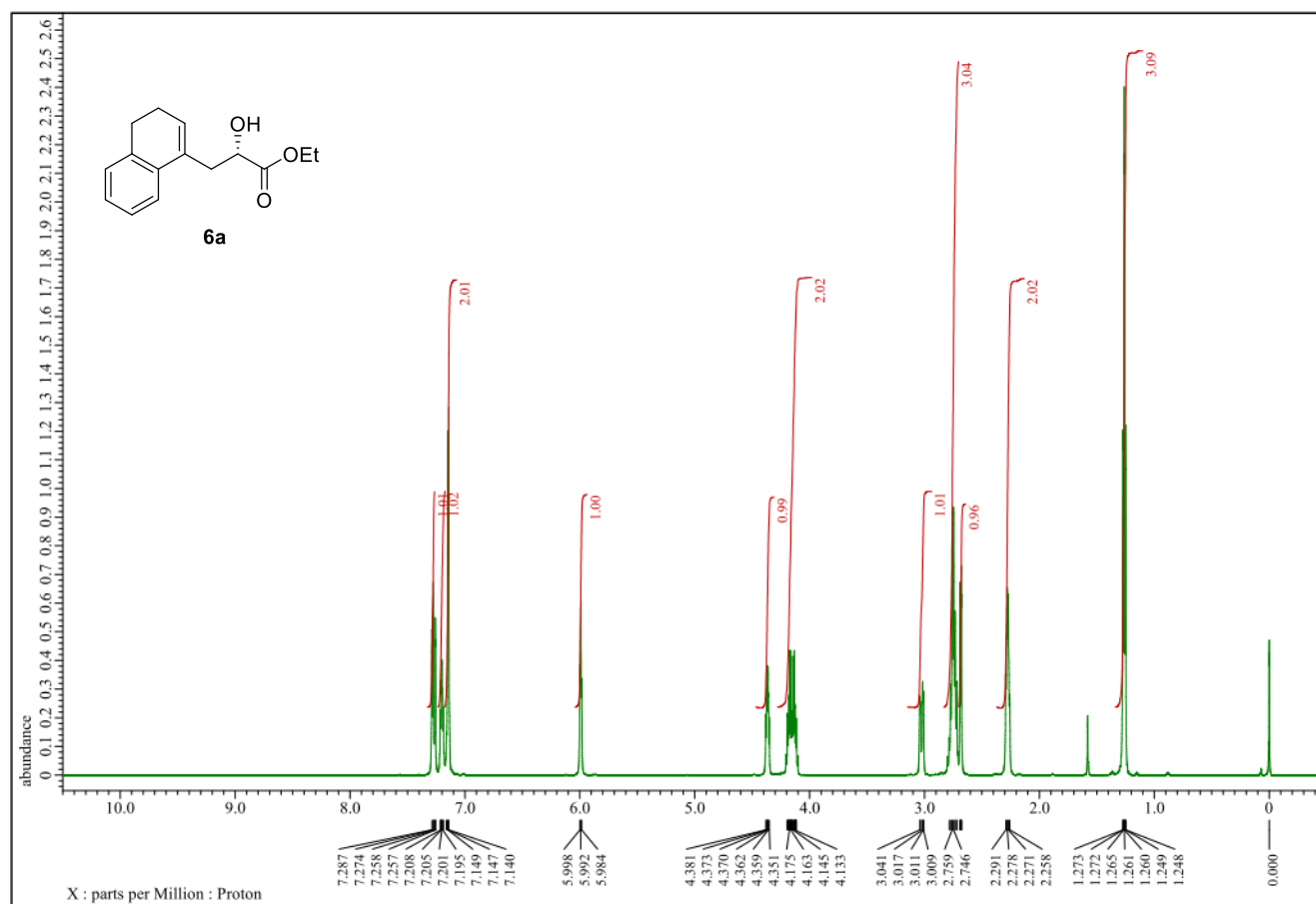
^1H NMR (600 MHz) and ^{13}C NMR (150 MHz) spectra of **4e**



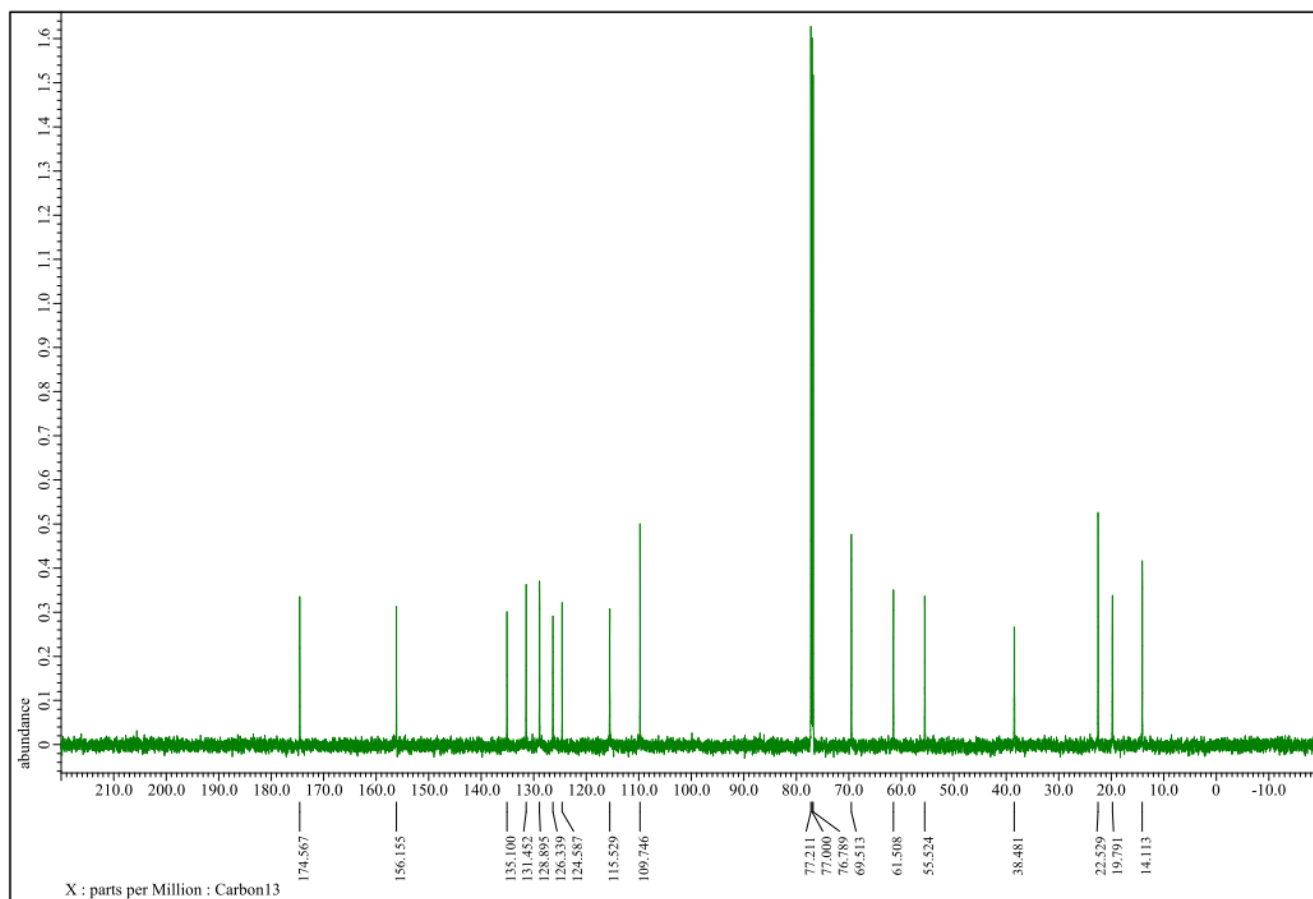
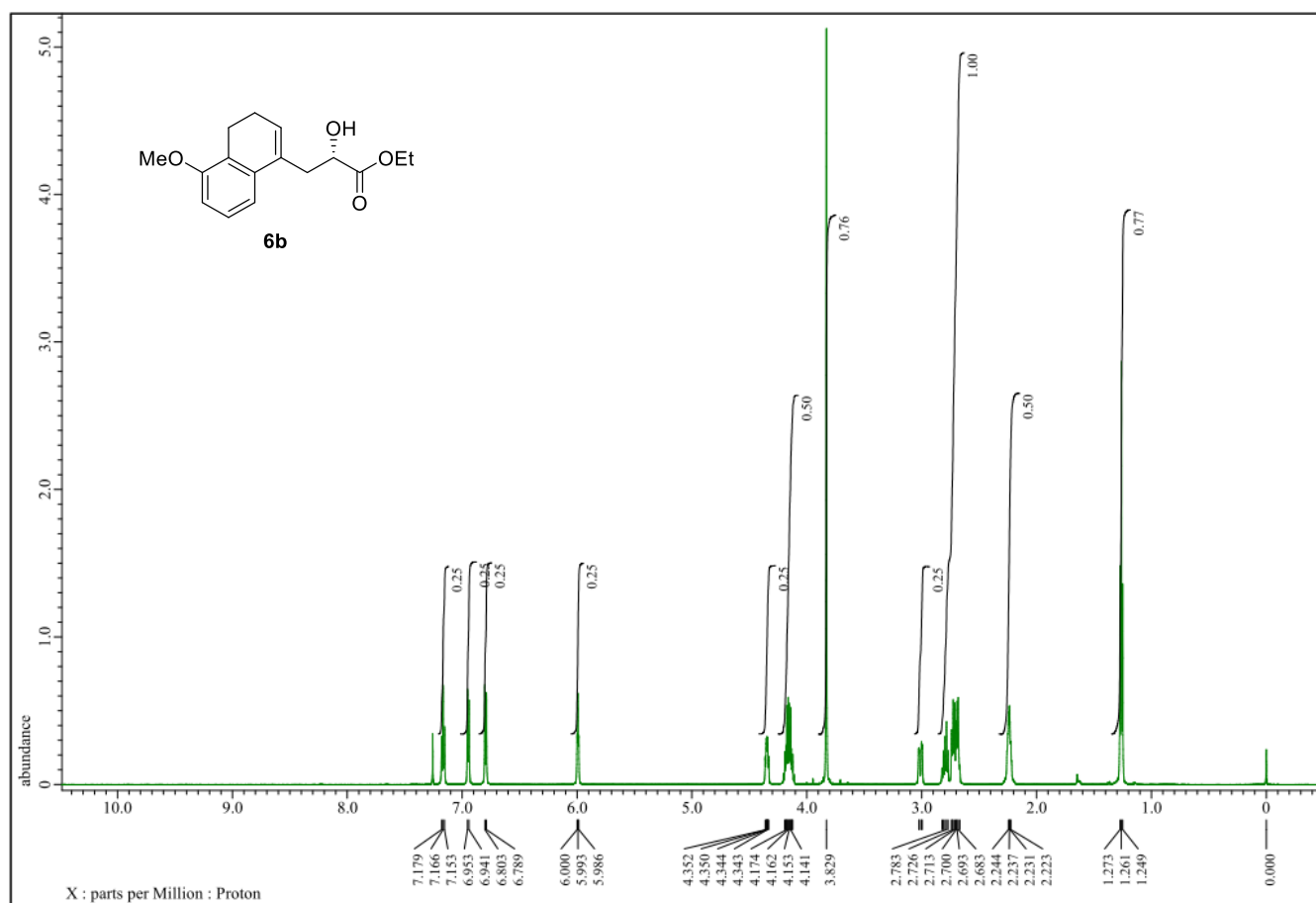
^1H NMR (600 MHz) and ^{13}C NMR (150 MHz) spectra of **4g**



^1H NMR (600 MHz) and ^{13}C NMR (150 MHz) spectra of **6a**

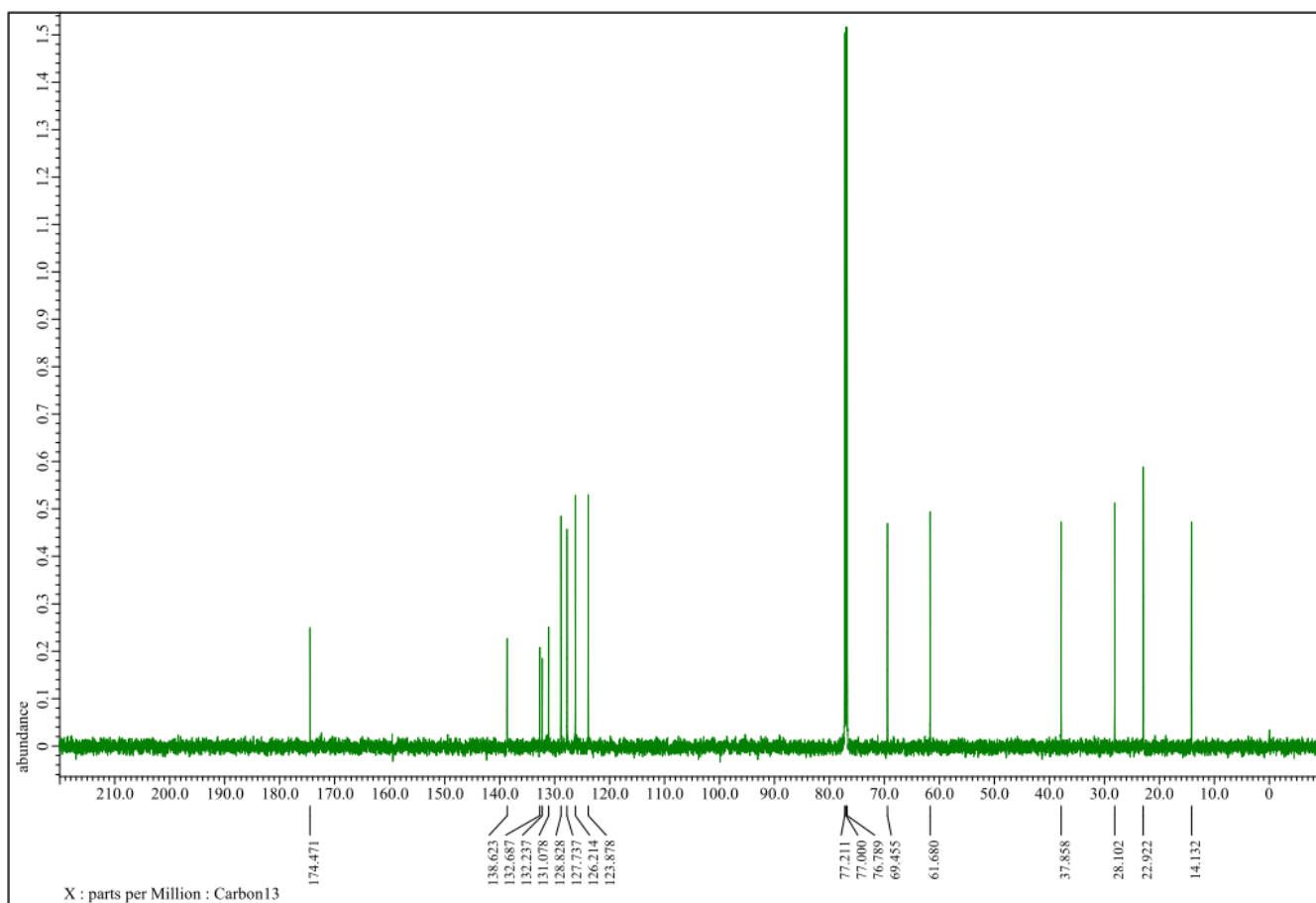
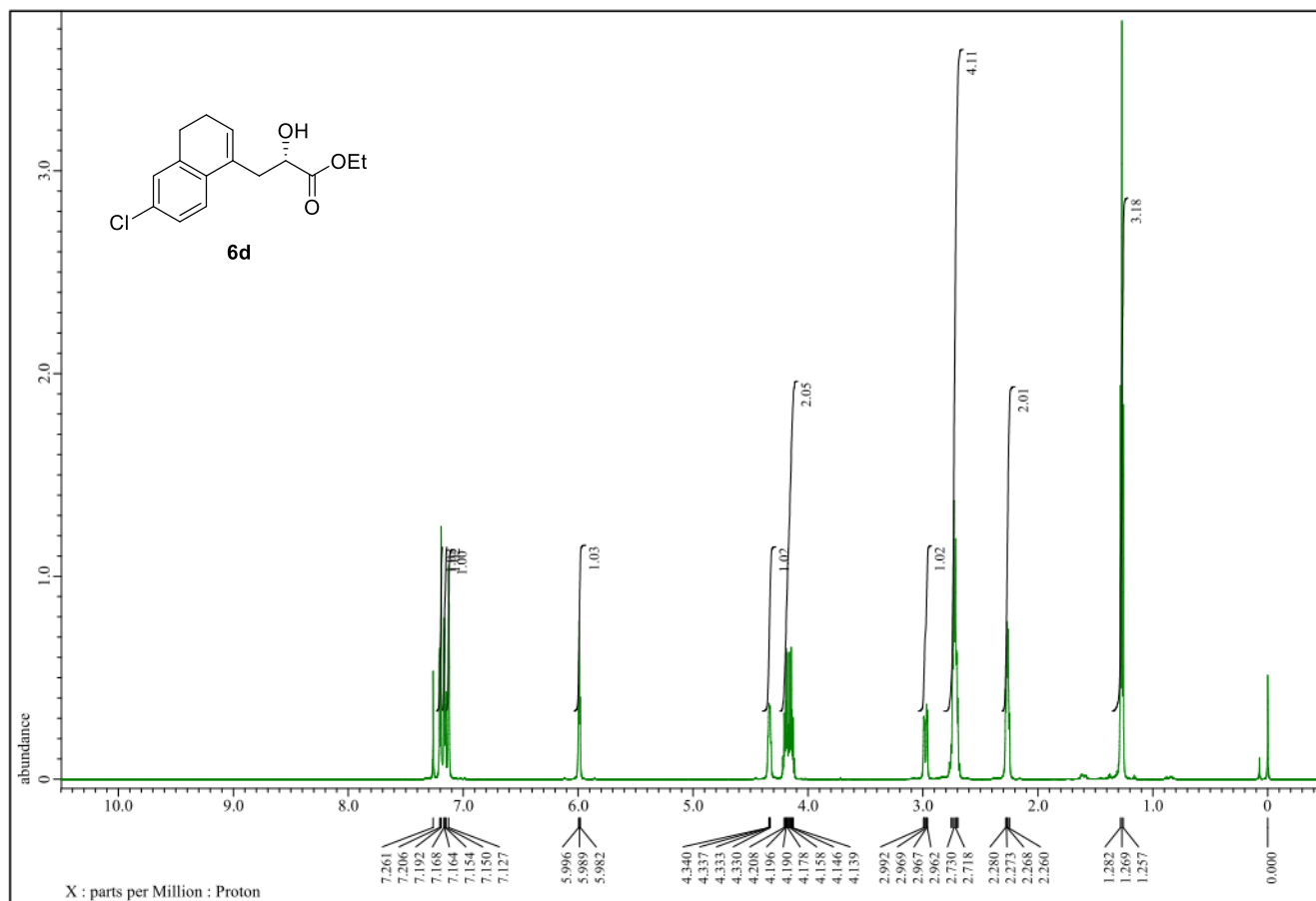


^1H NMR (600 MHz) and ^{13}C NMR (150 MHz) spectra of **6b**

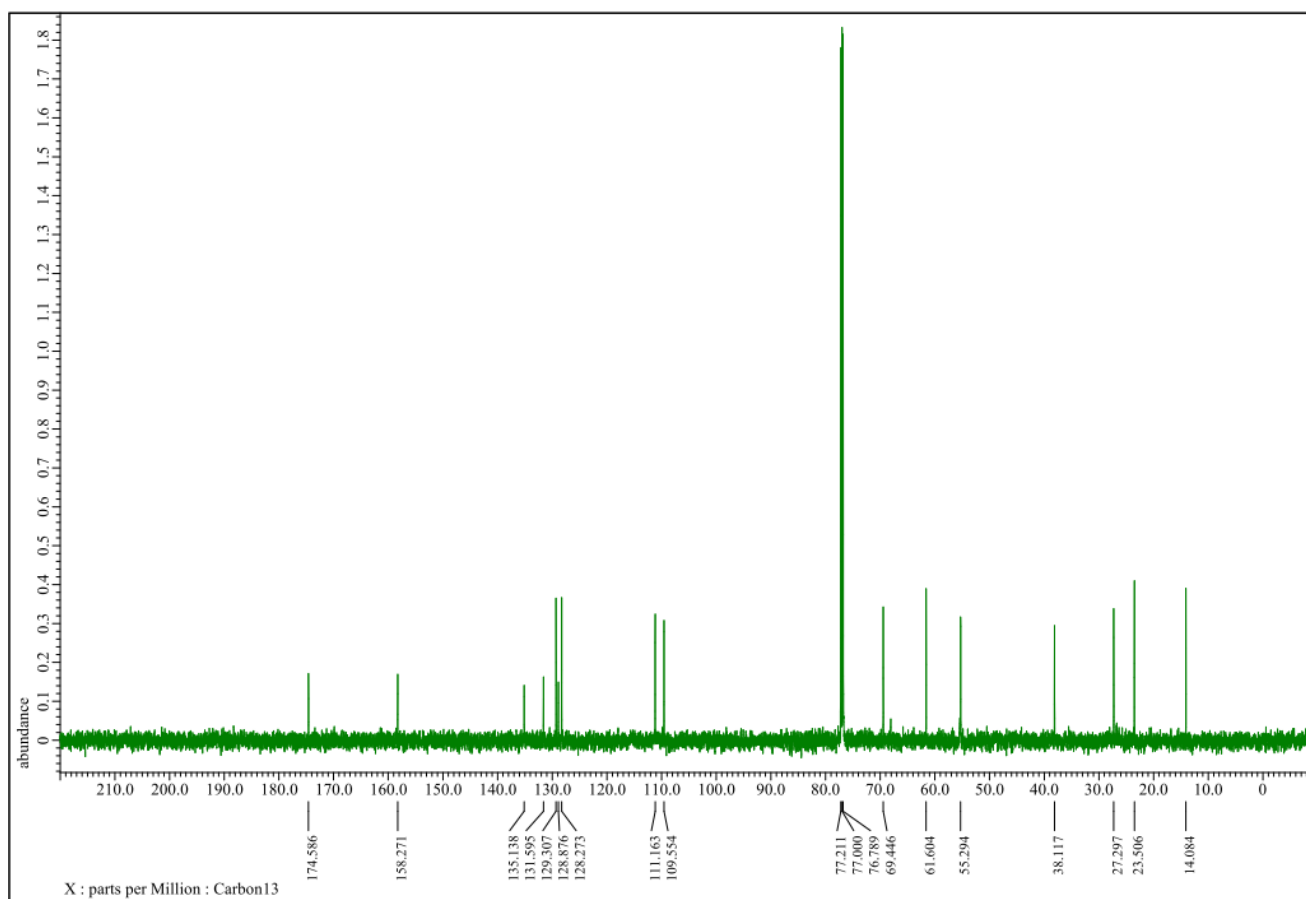
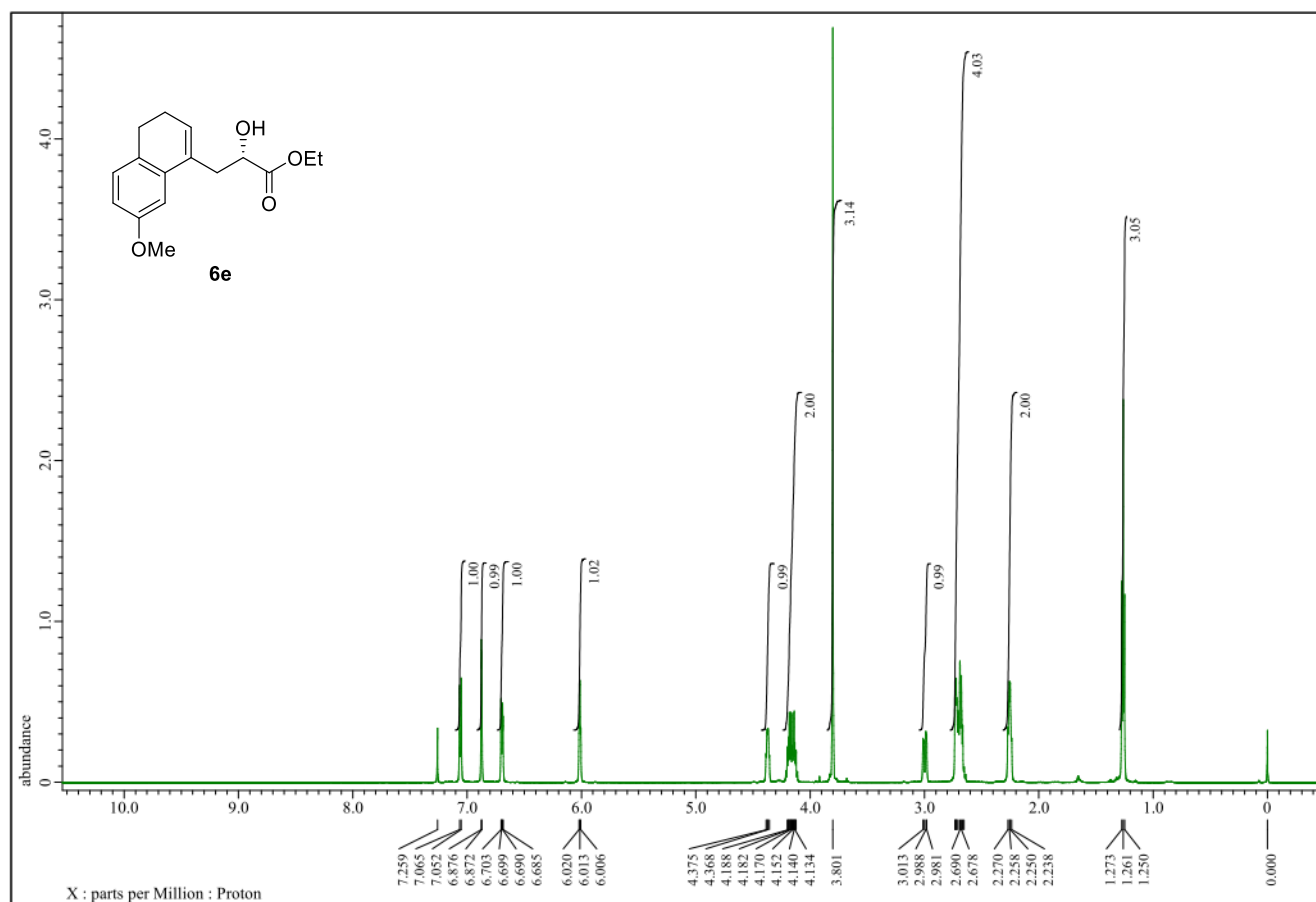


^1H NMR (600 MHz) and ^{13}C NMR (150 MHz) spectra of **6c**

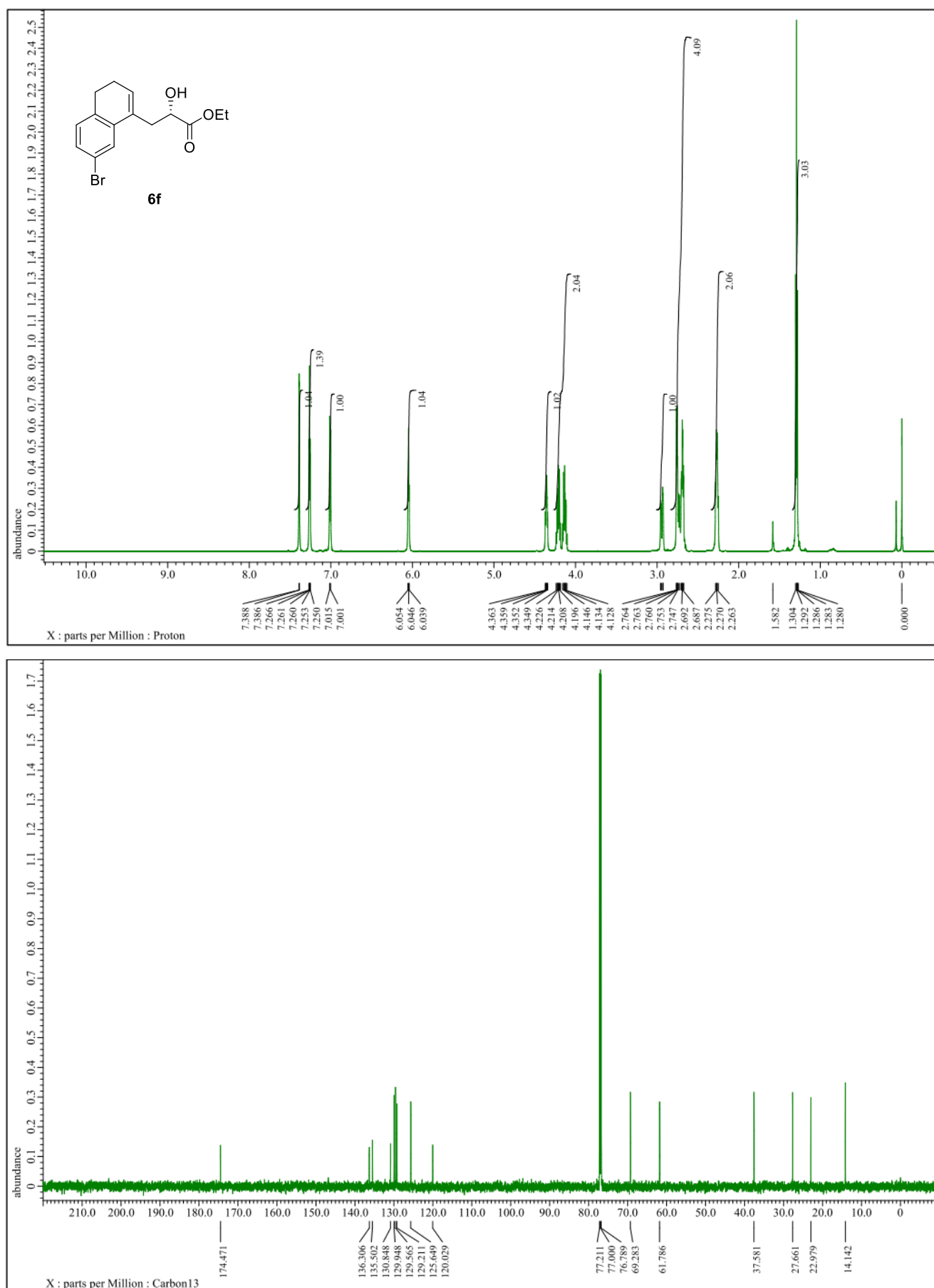
^1H NMR (600 MHz) and ^{13}C NMR (150 MHz) spectra of **6d**



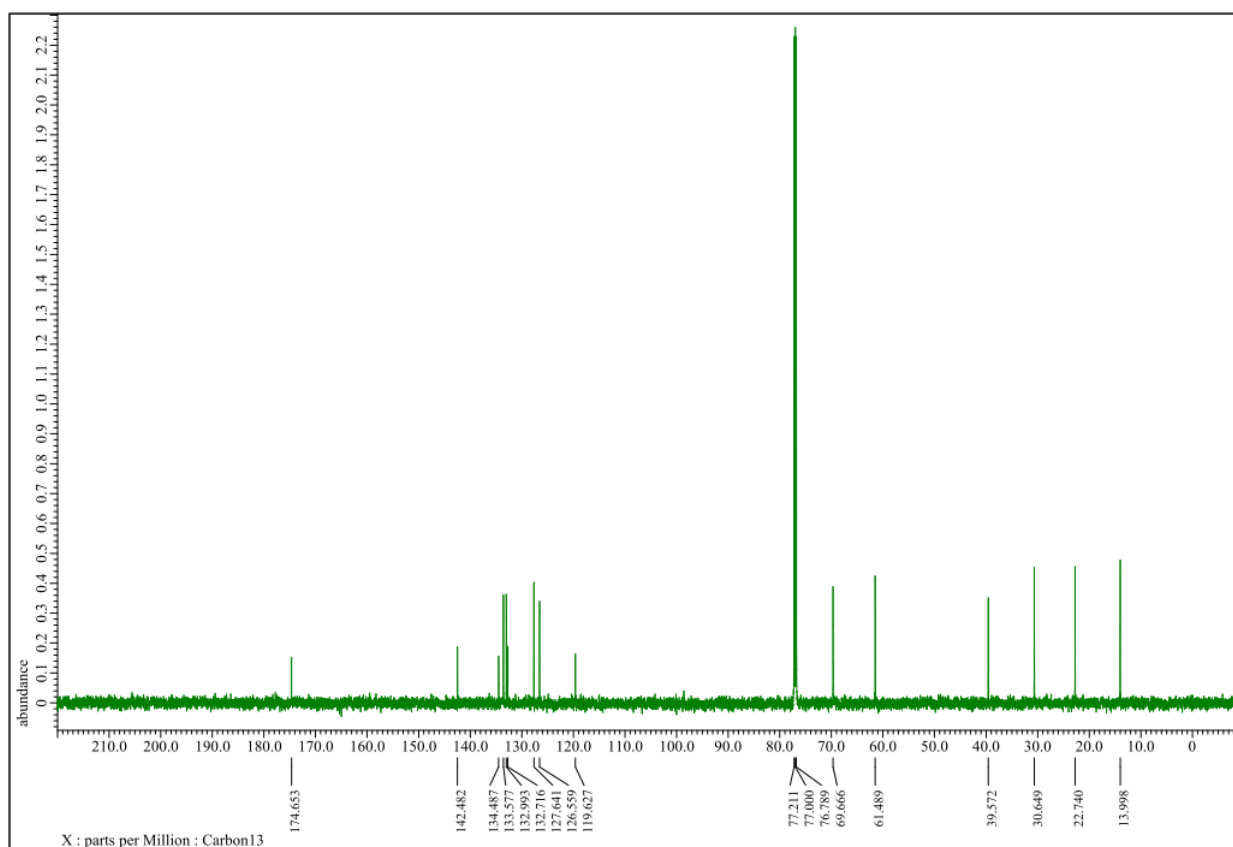
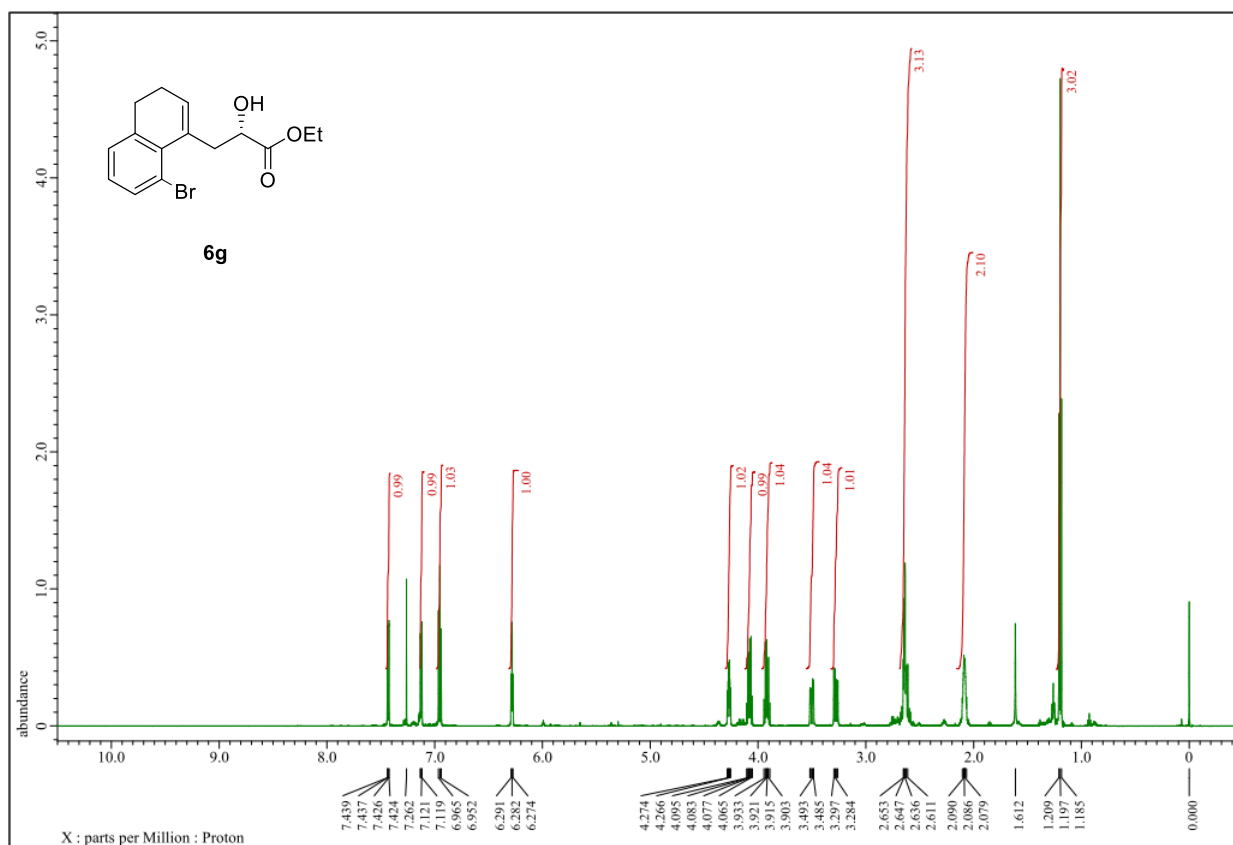
^1H NMR (600 MHz) and ^{13}C NMR (150 MHz) spectra of **6e**



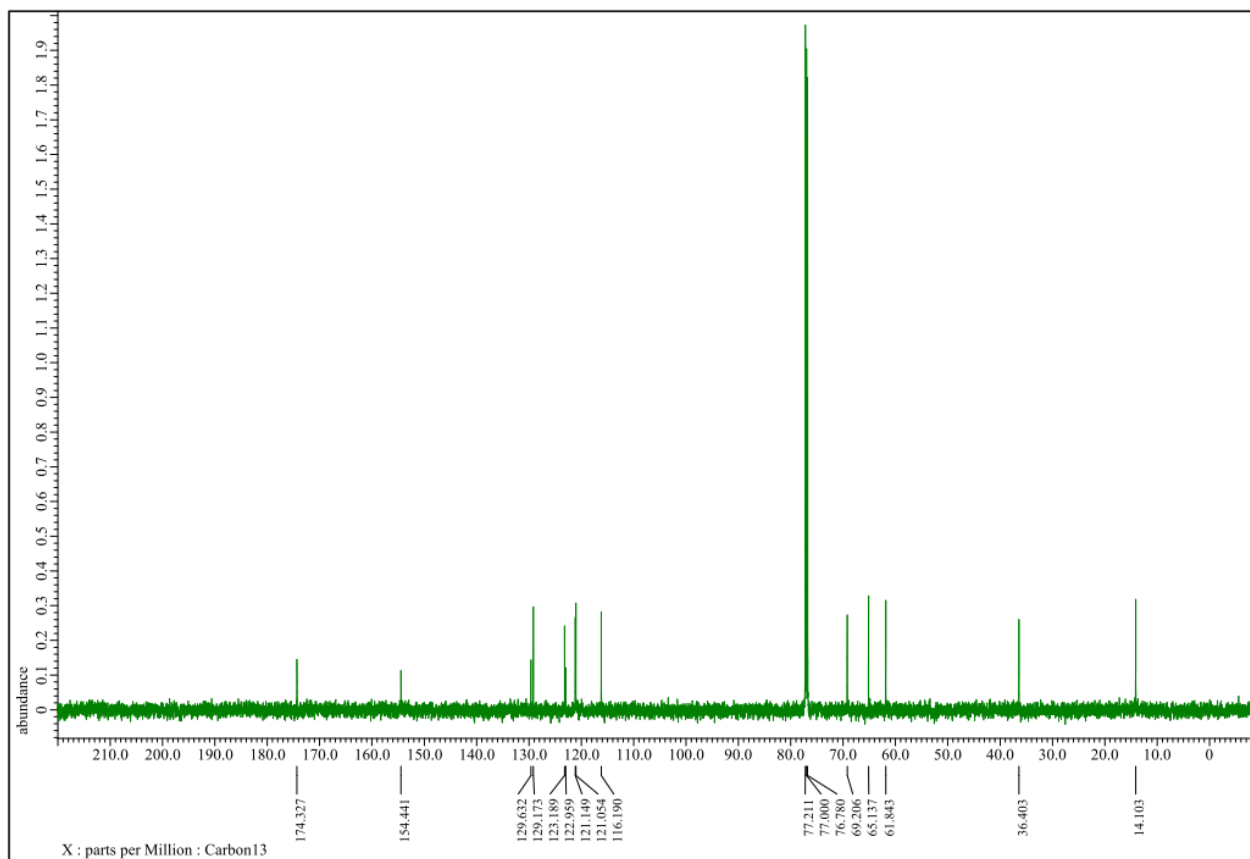
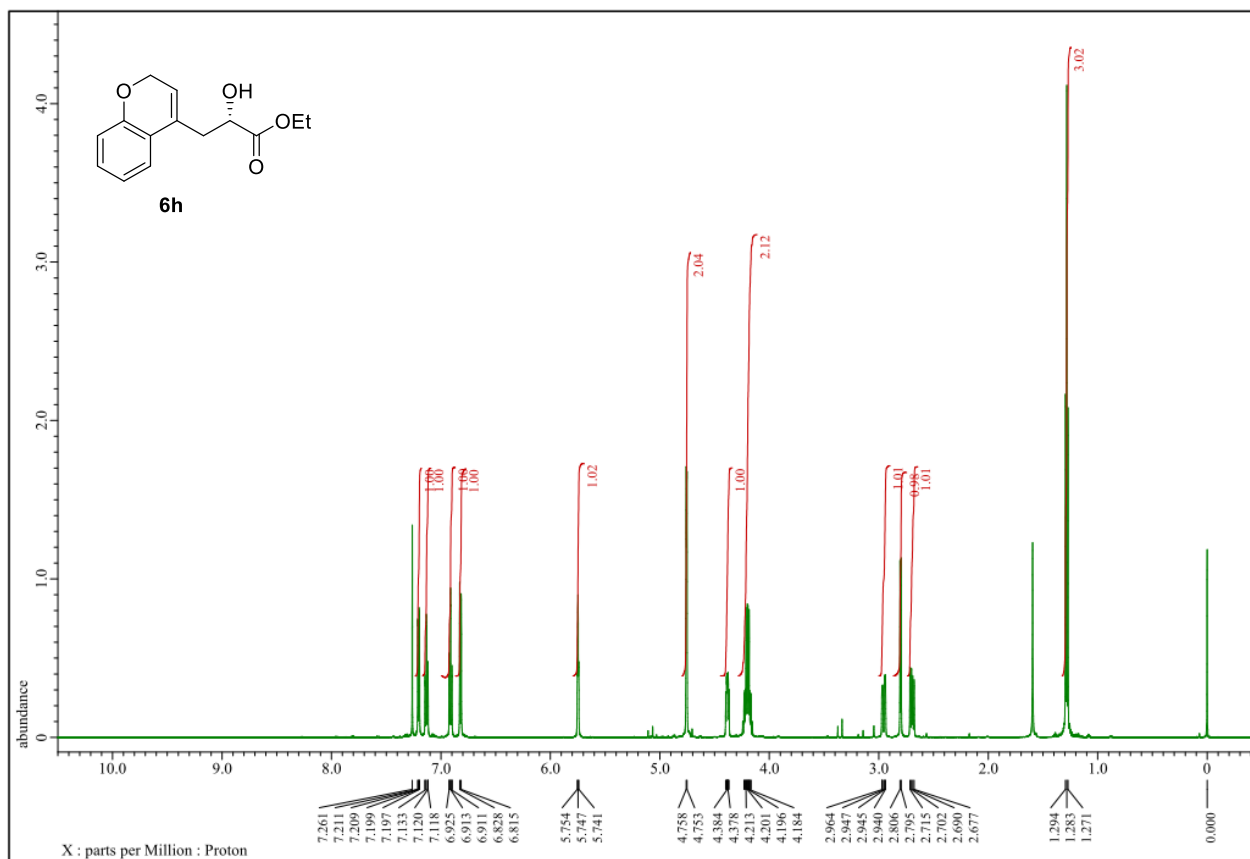
^1H NMR (600 MHz) and ^{13}C NMR (150 MHz) spectra of **6f**



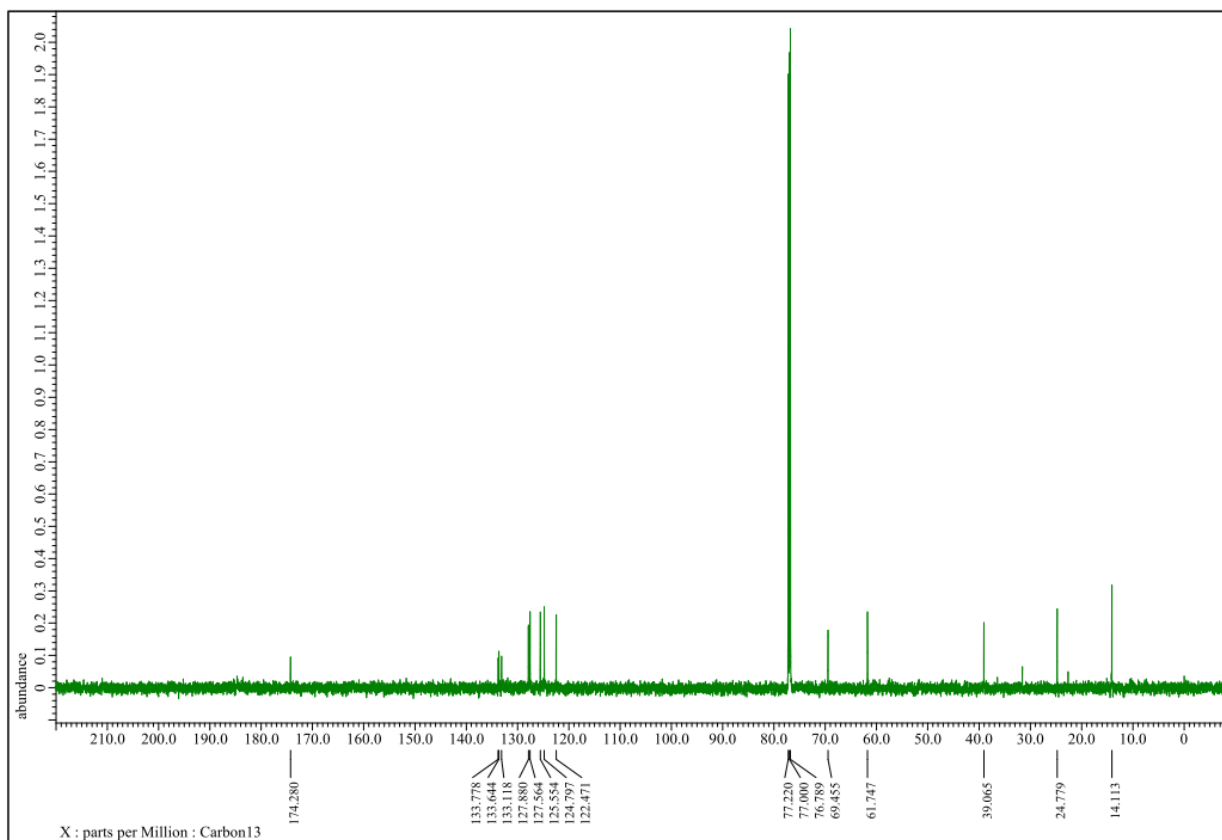
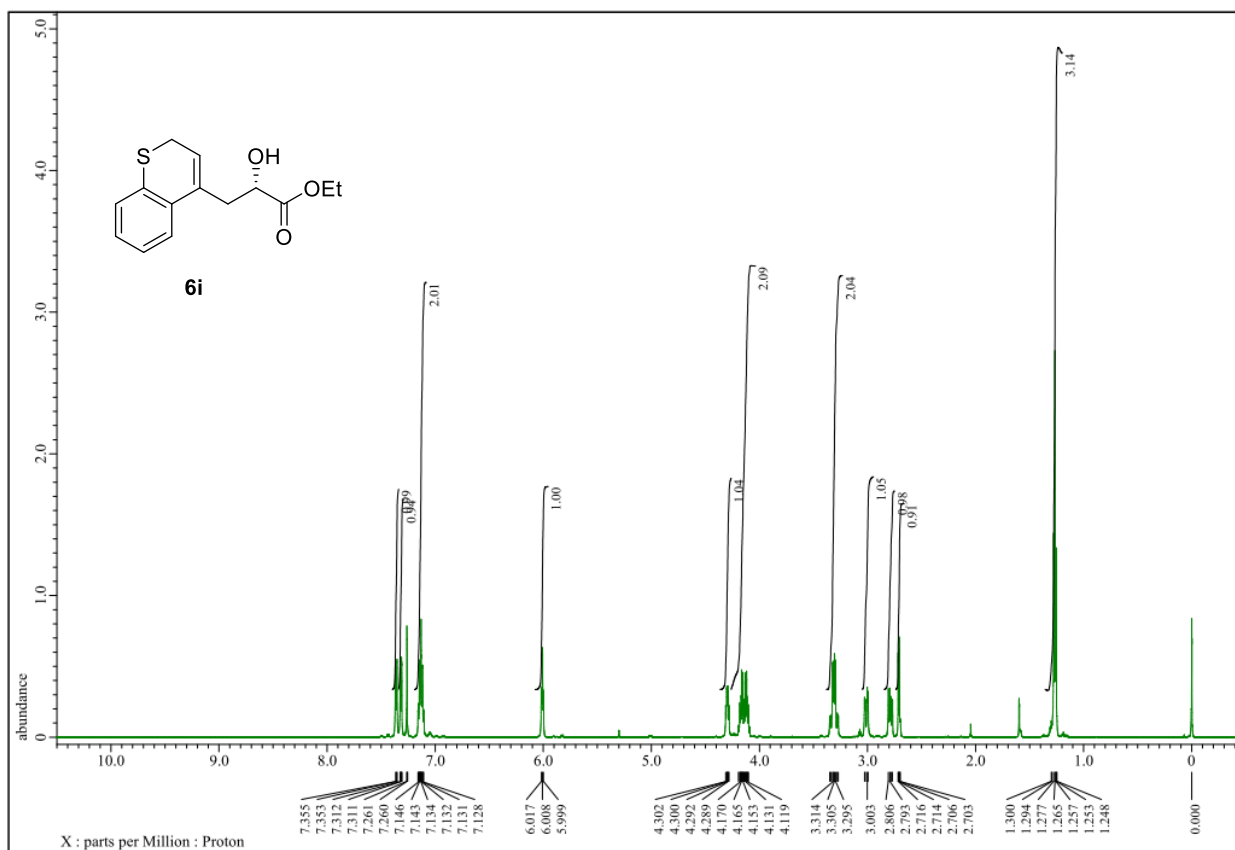
^1H NMR (600 MHz) and ^{13}C NMR (150 MHz) spectra of **6g**



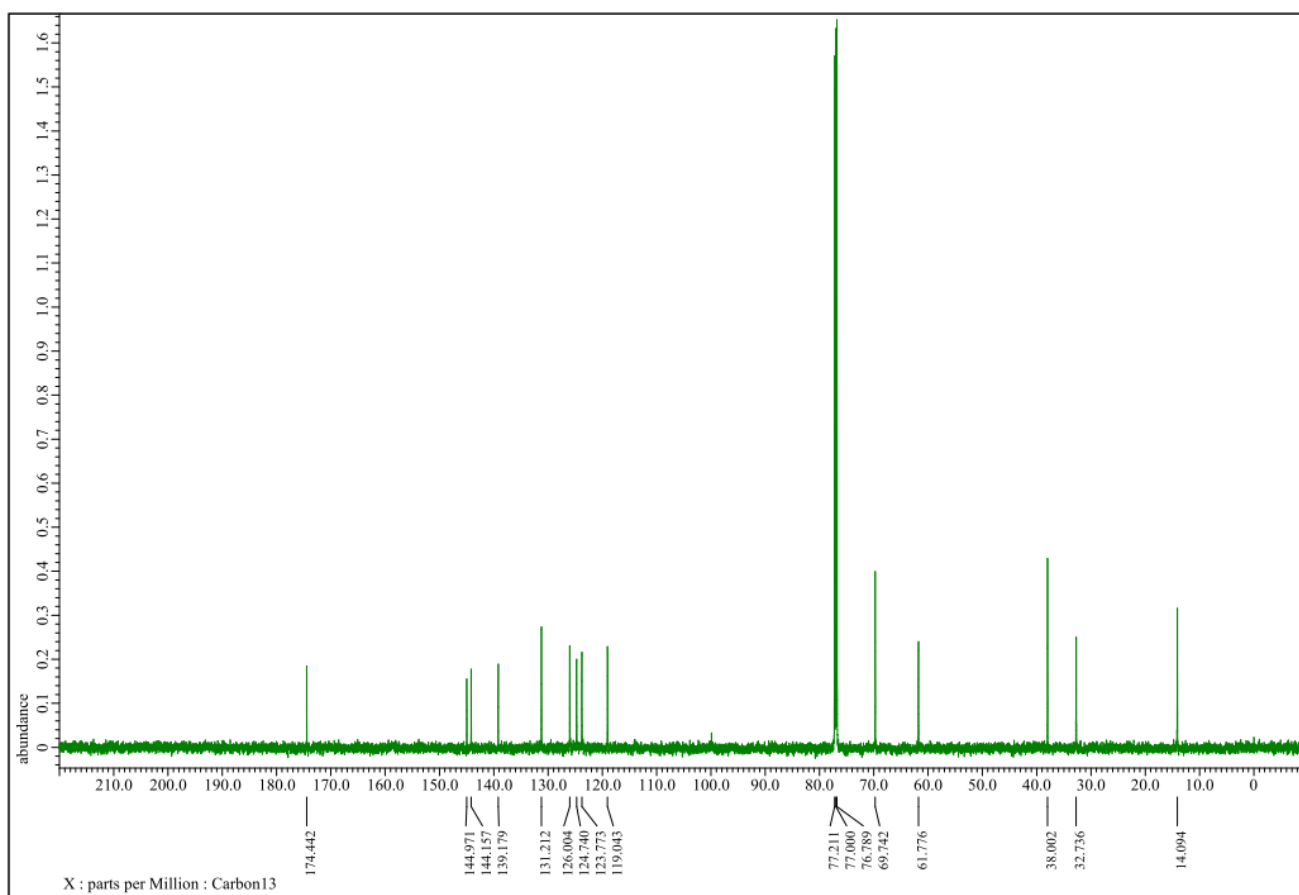
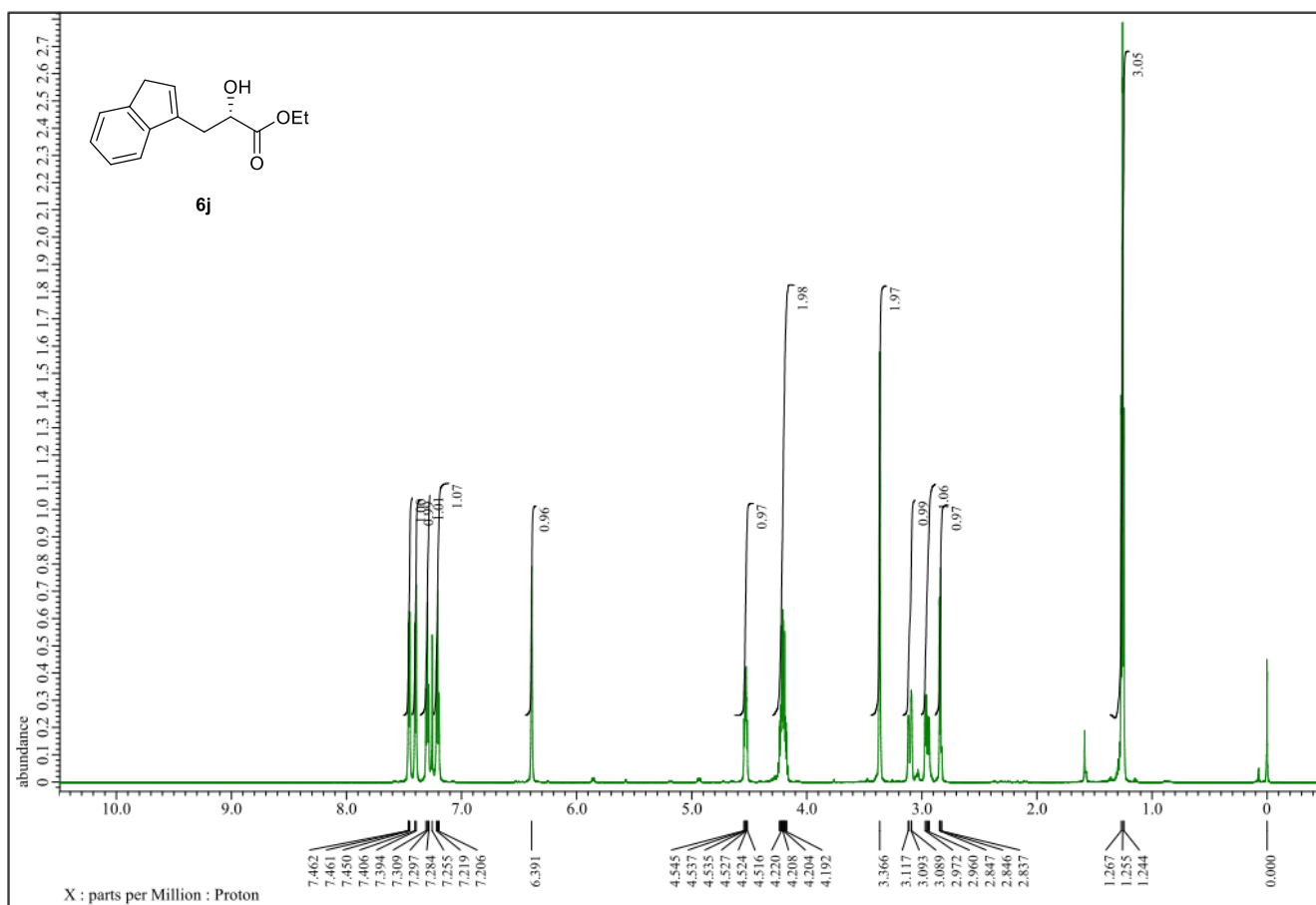
^1H NMR (600 MHz) and ^{13}C NMR (150 MHz) spectra of **6h**



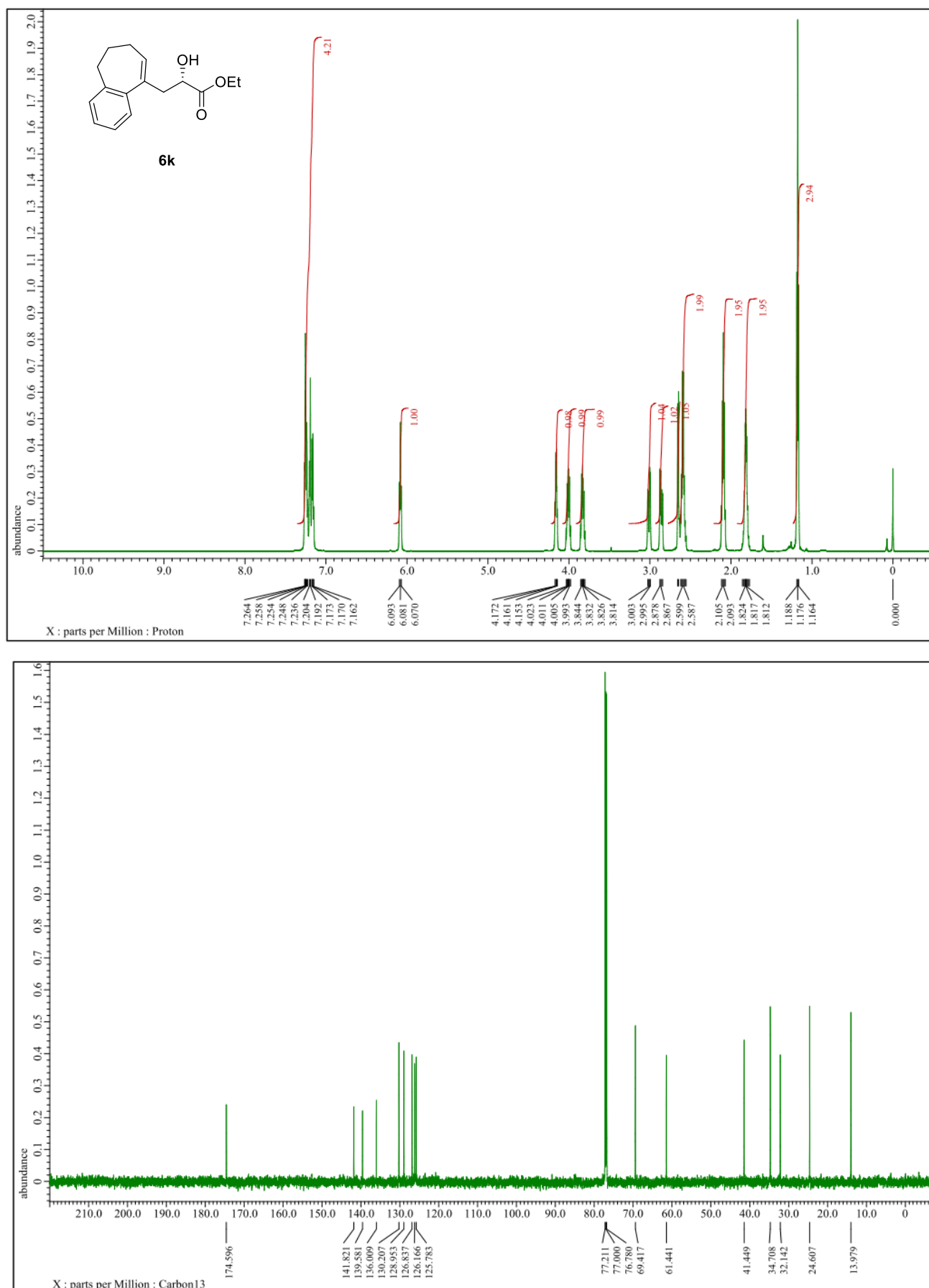
H NMR (600 MHz) and ^{13}C NMR (150 MHz) spectra of **6i**



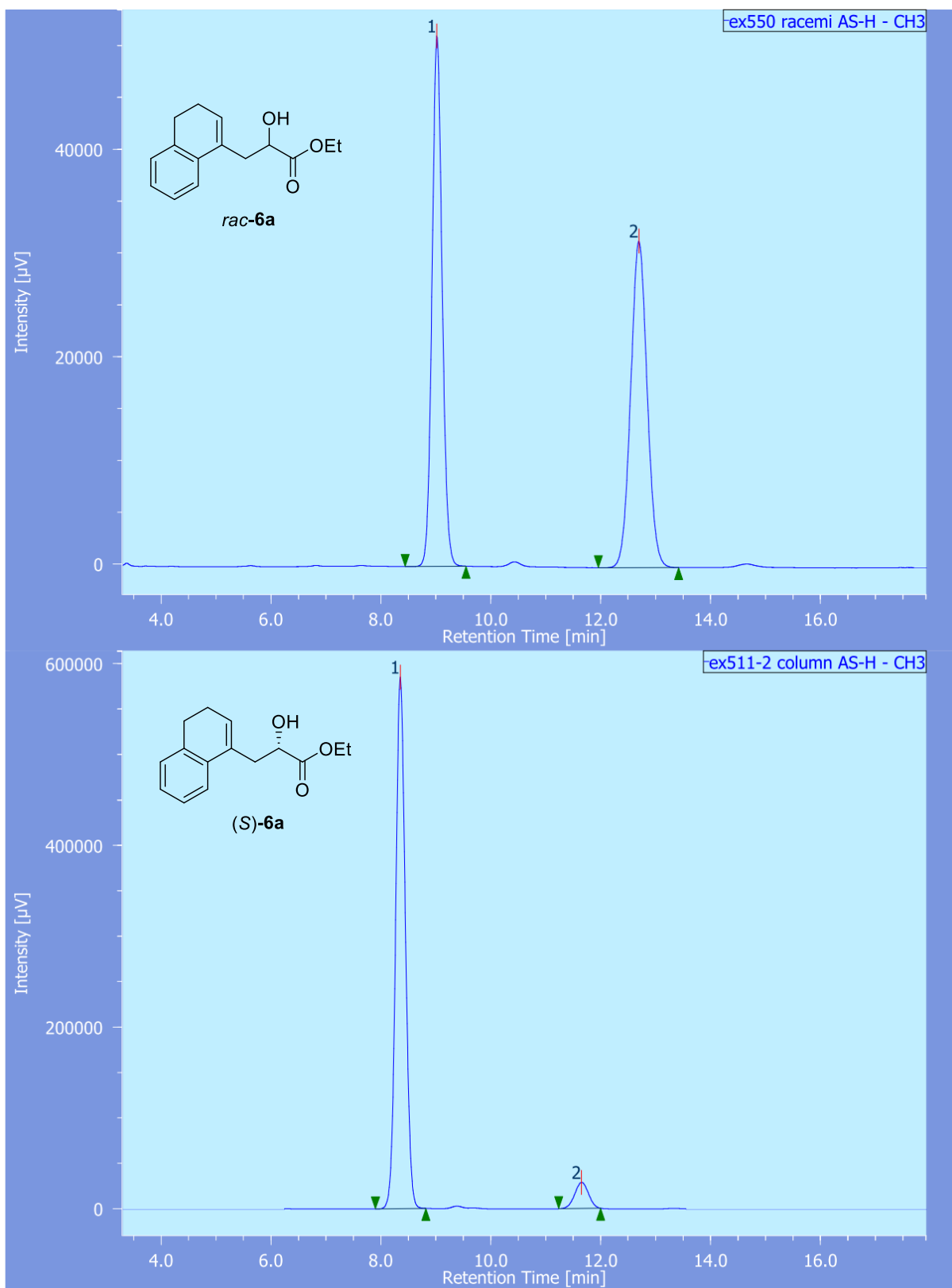
^1H NMR (600 MHz) and ^{13}C NMR (150 MHz) spectra of **6j**

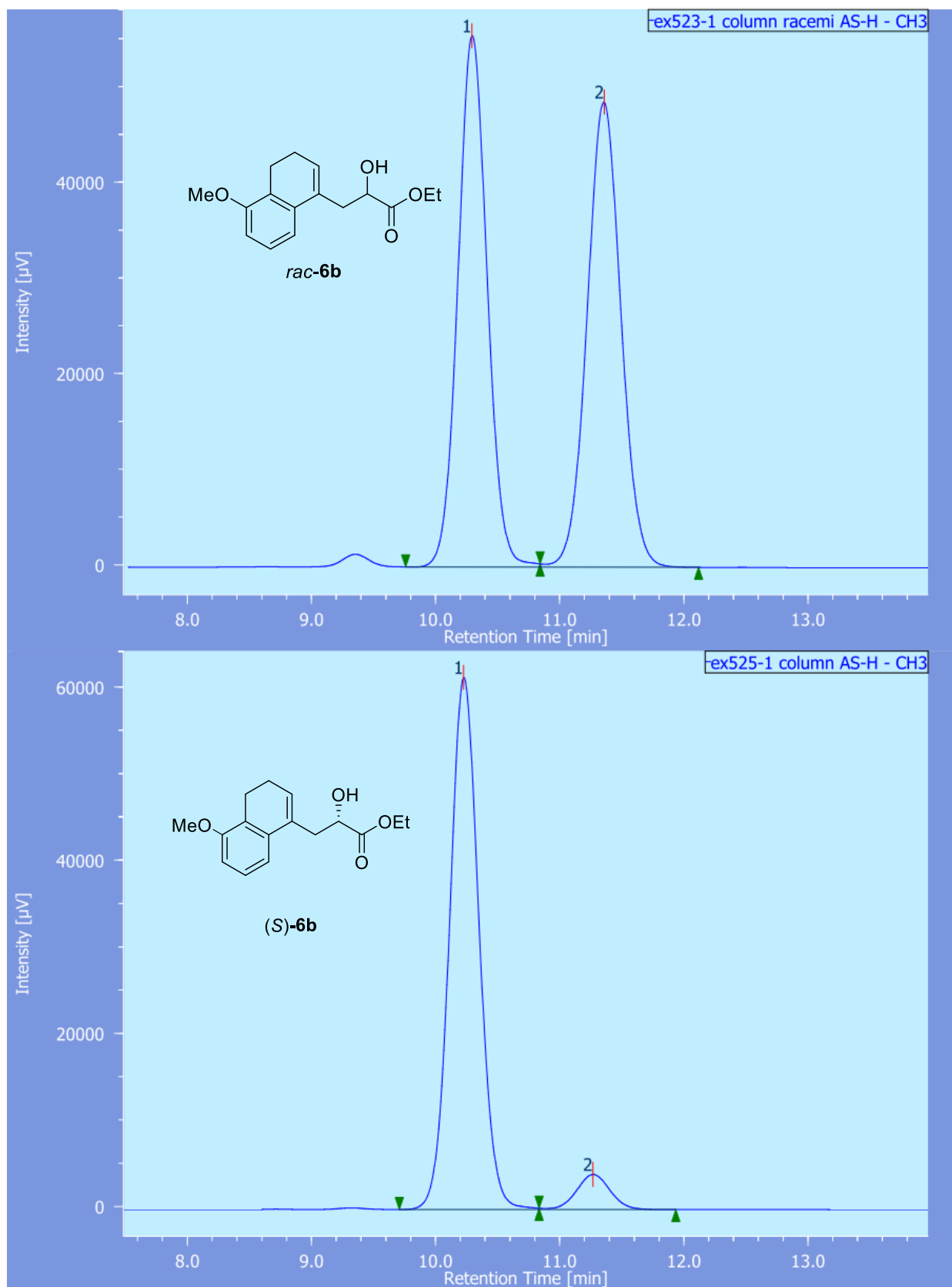


^1H NMR (600 MHz) and ^{13}C NMR (150 MHz) spectra of **6k**

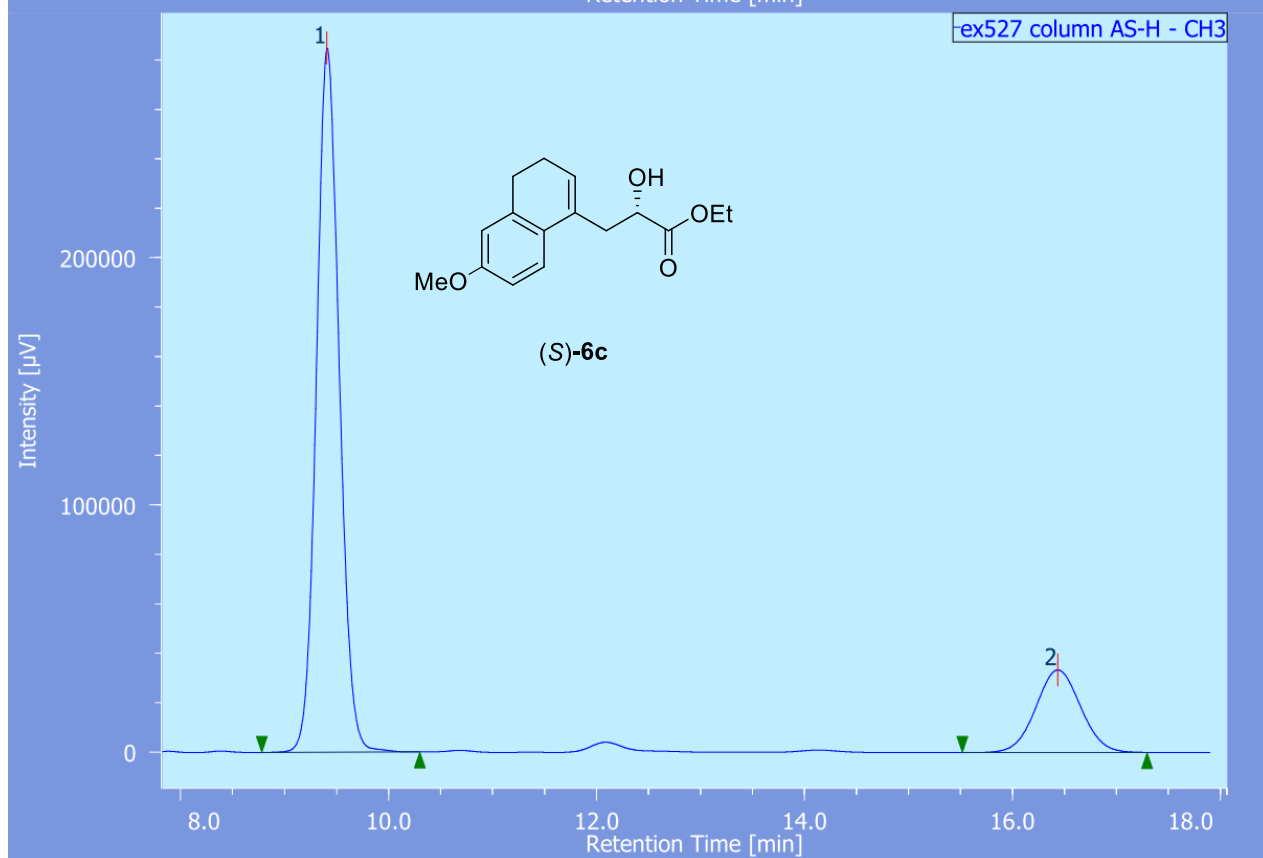
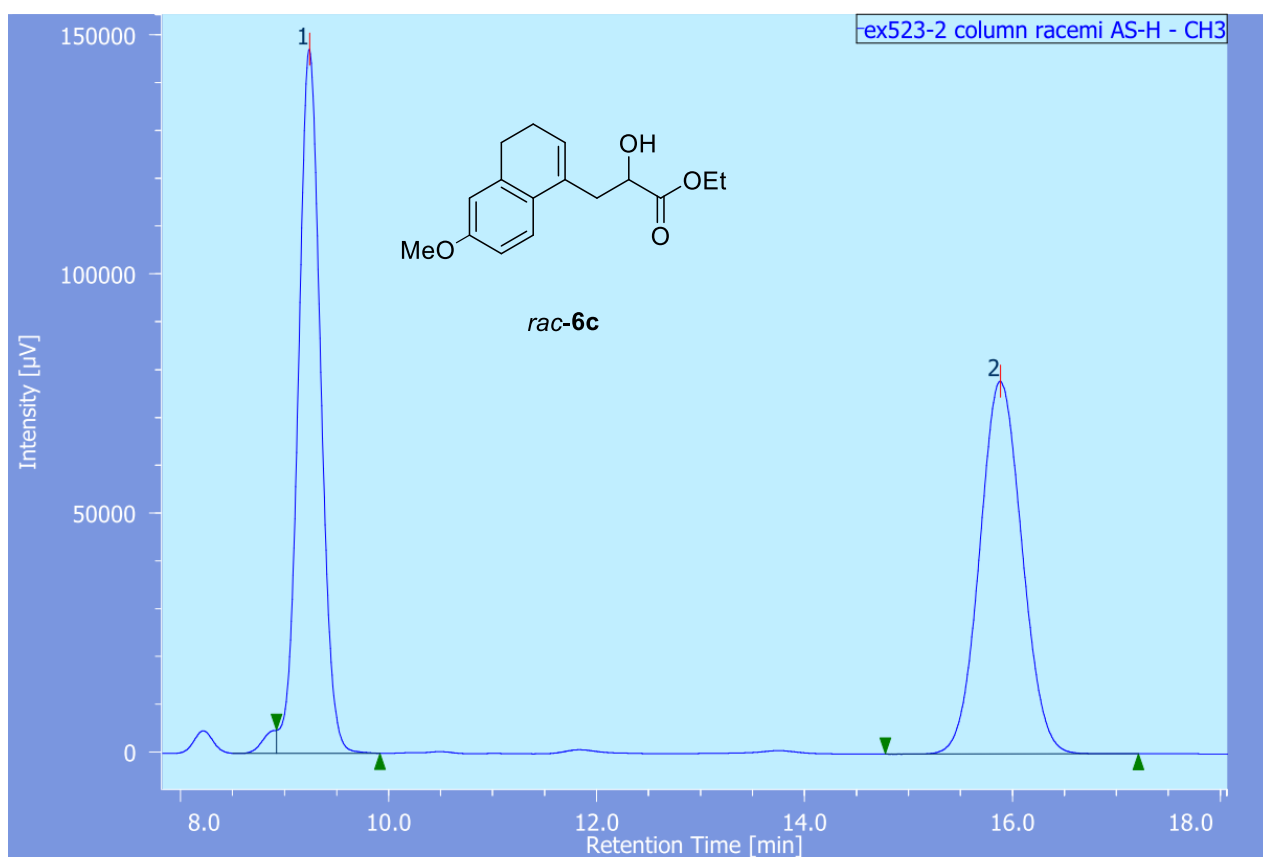


8. HPLC charts

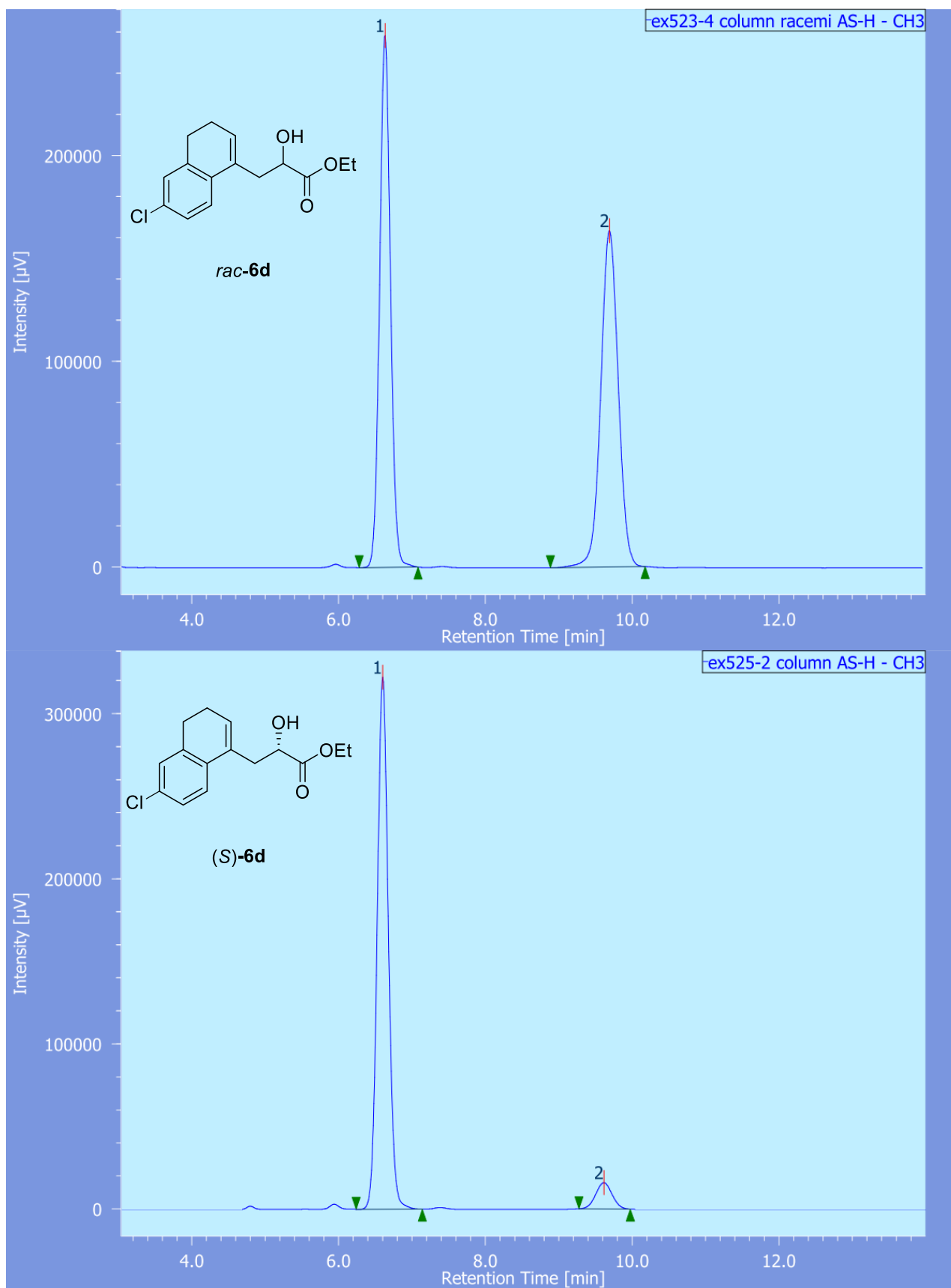




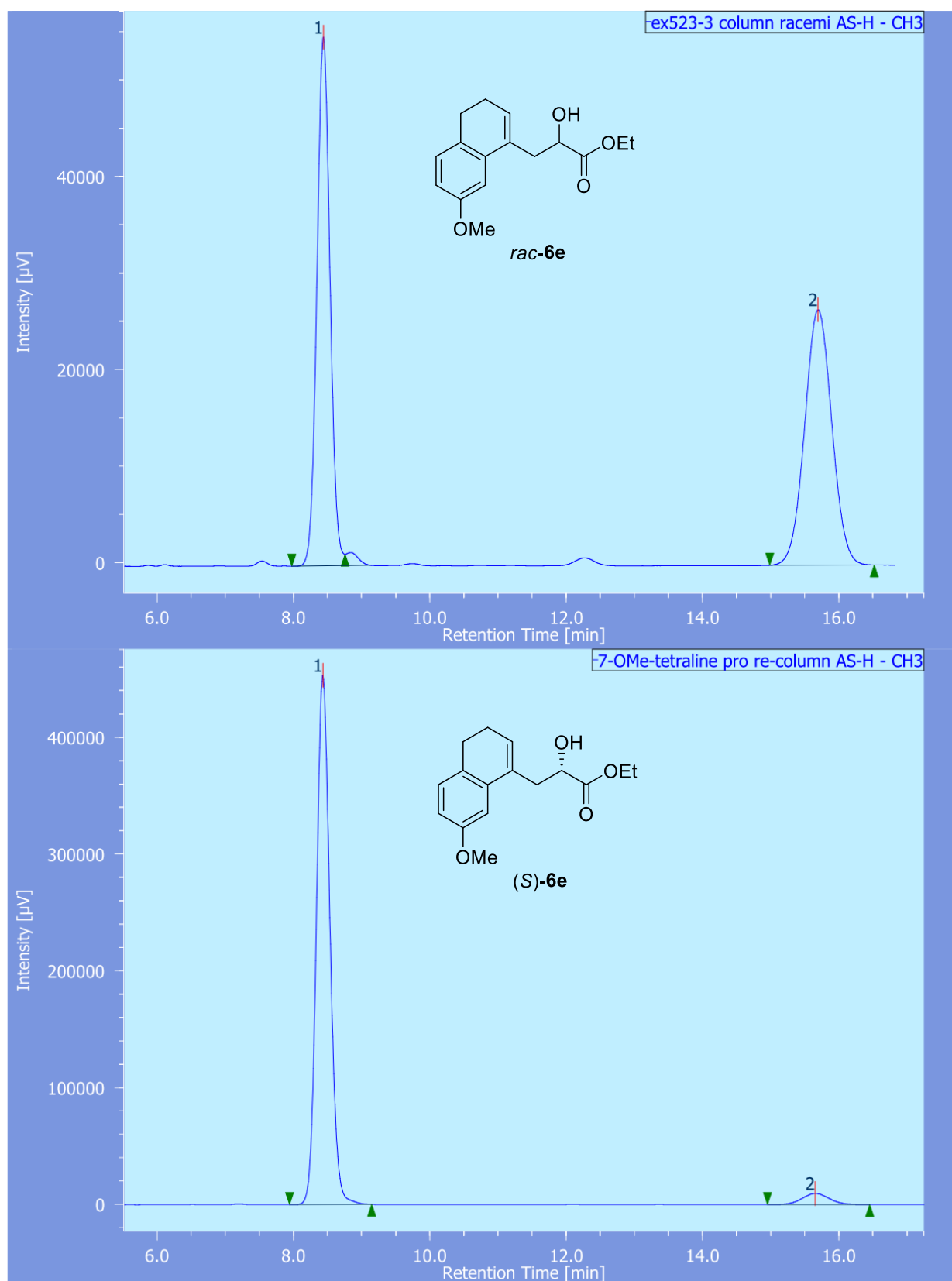
	Retention Time (1)	Retention Time (2)	Area (1)	Area (2)	%Area (1)	%Area (2)
<i>rac</i> -6b	10.29	11.36	910634	922917	49.67	50.33
<i>(S)</i> -6b	10.23	11.27	1009897	76729	92.94	7.06



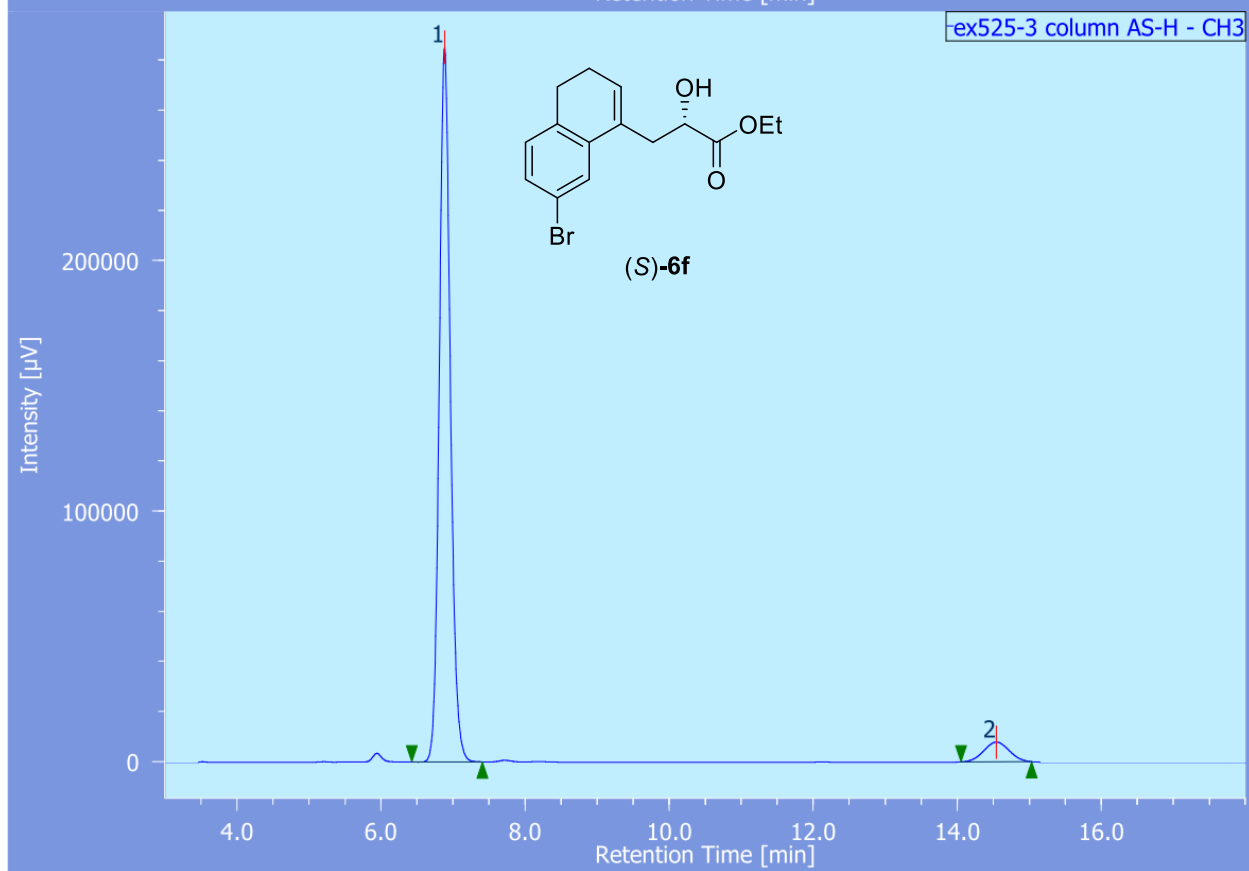
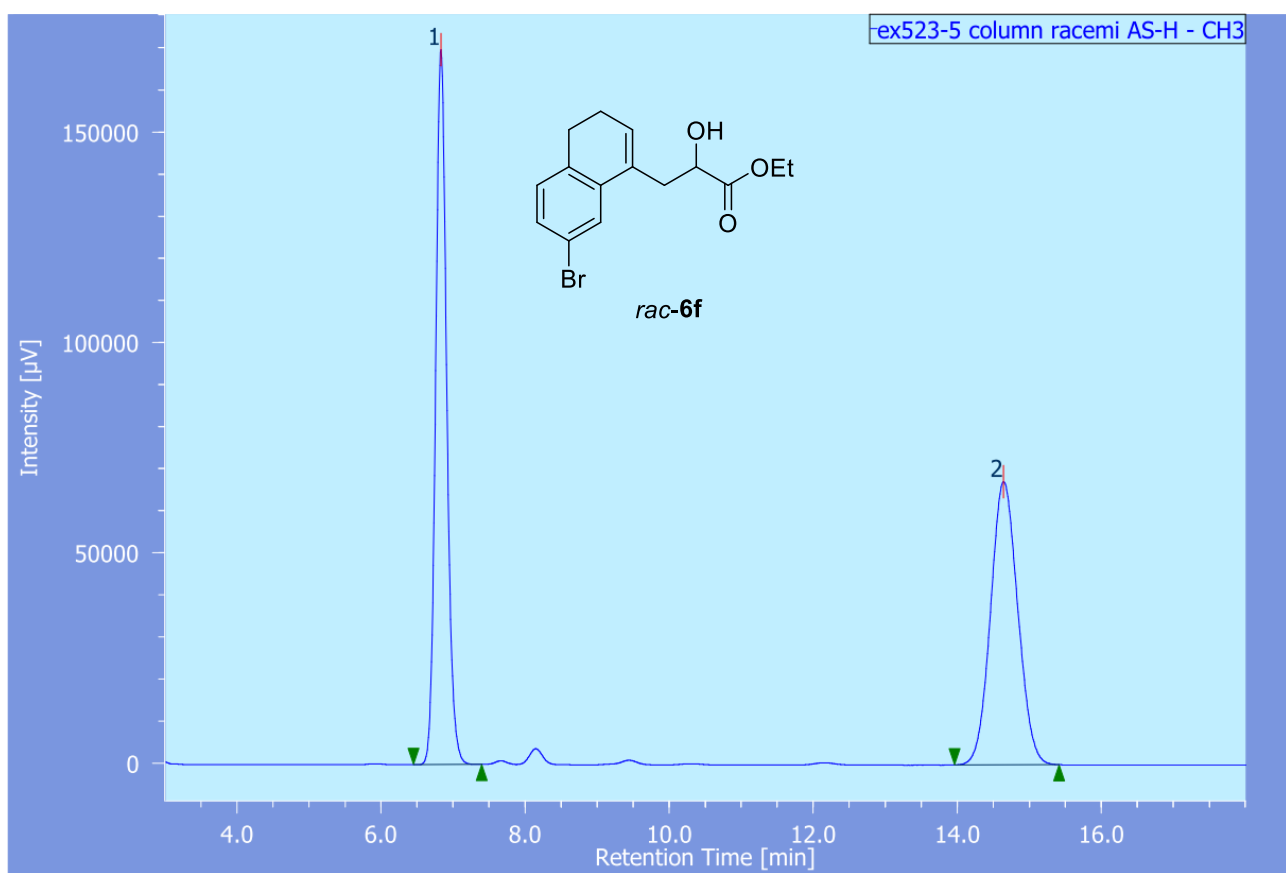
	Retention Time (1)	Retention Time (2)	Area (1)	Area (2)	%Area (1)	%Area (2)
<i>rac</i> - 6c	9.24	15.88	2199884	2198497	50.02	49.98
(<i>S</i>)- 6c	9.41	16.43	4316582	999712	81.20	18.80



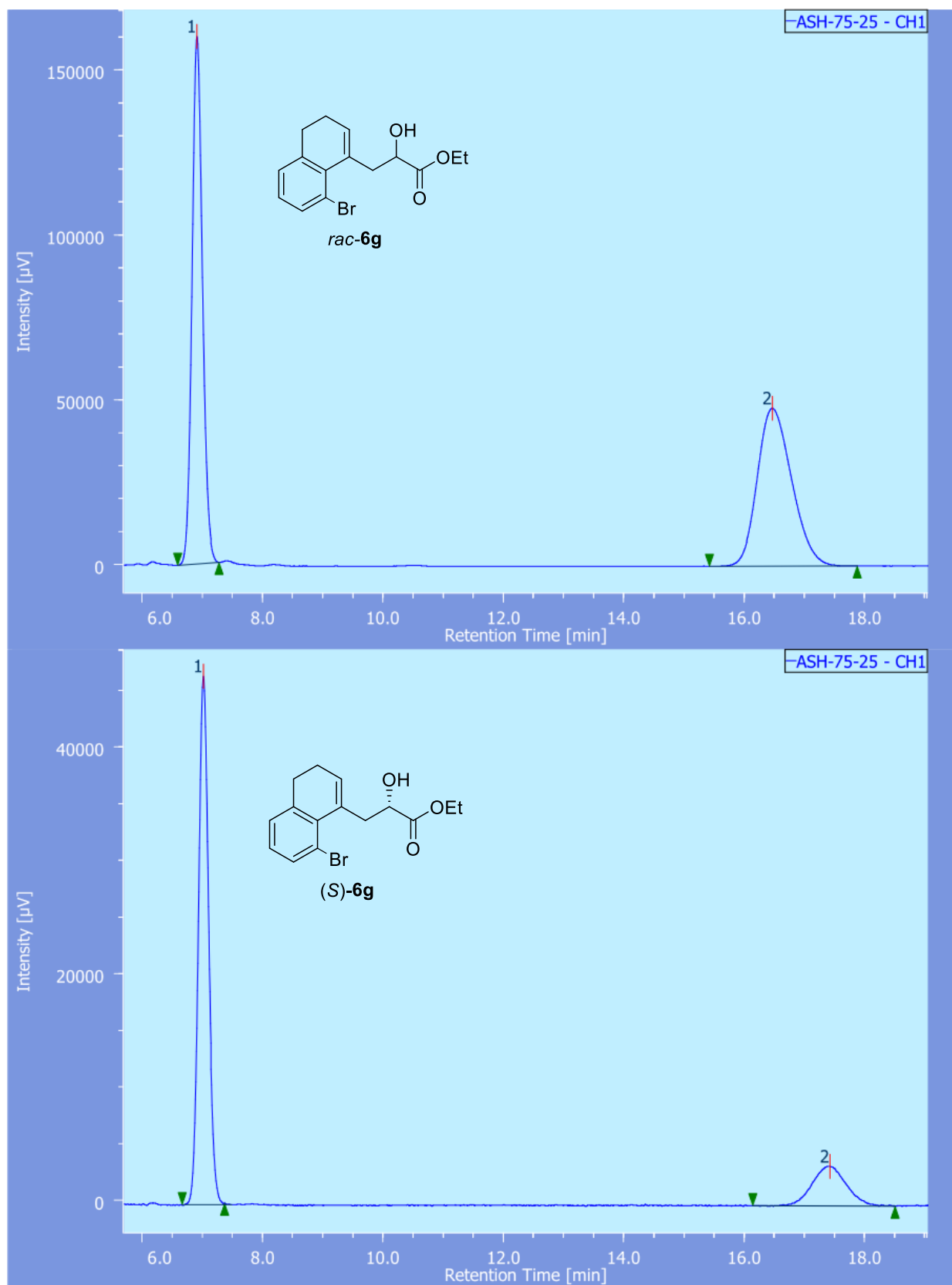
	Retention Time (1)	Retention Time (2)	Area (1)	Area (2)	%Area (1)	%Area (2)
<i>rac</i> - 6d	6.63	9.69	2624371	2696846	49.32	50.68
(<i>S</i>)- 6d	6.60	9.62	3331082	247448	93.09	6.91



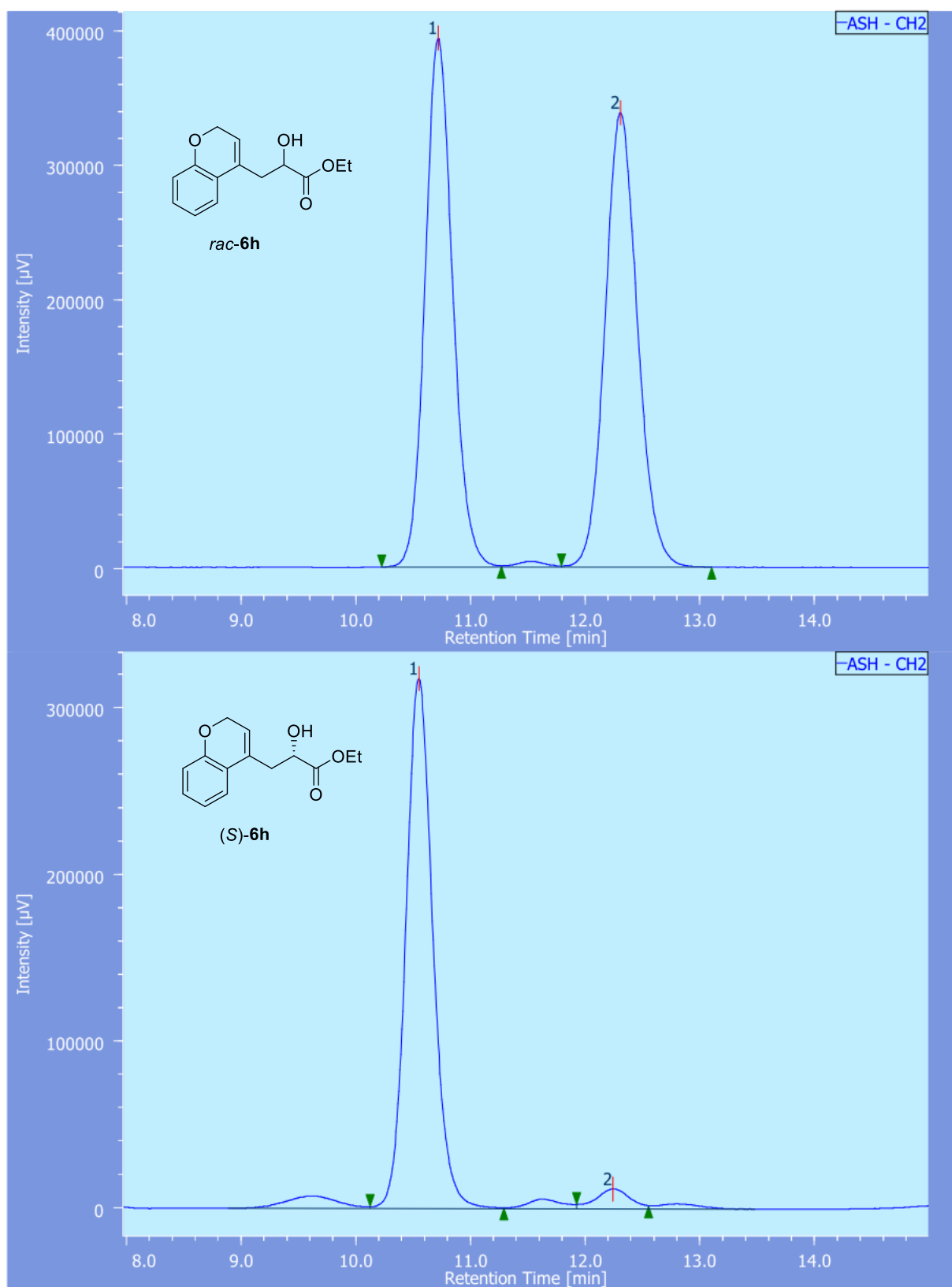
	Retention Time (1)	Retention Time (2)	Area (1)	Area (2)	%Area (1)	%Area (2)
<i>rac</i> - 6e	8.44	15.69	730716	732529	49.94	50.06
(<i>S</i>)- 6e	8.43	15.65	6222514	264688	95.92	4.08



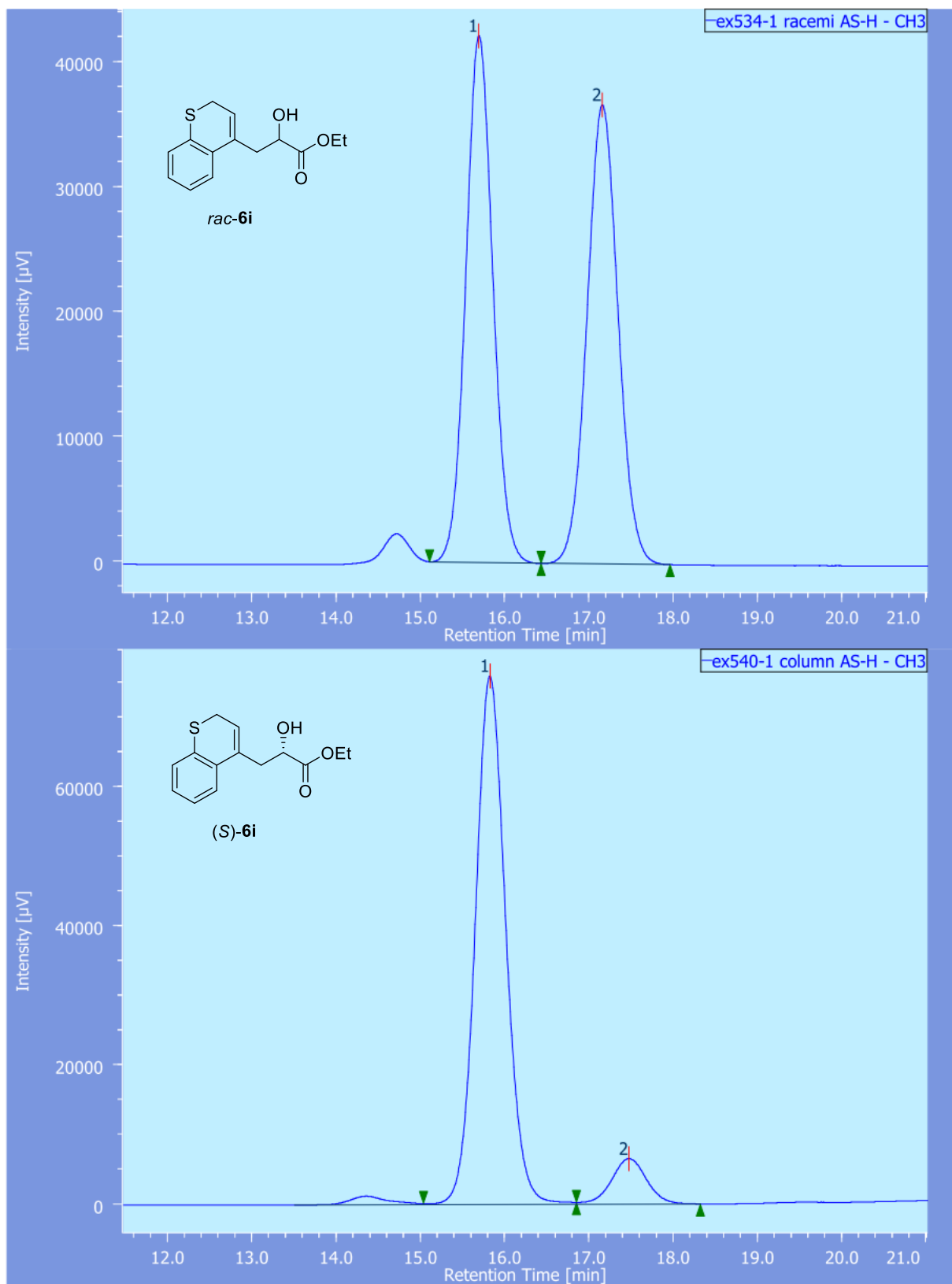
	Retention Time (1)	Retention Time (2)	Area (1)	Area (2)	%Area (1)	%Area (2)
<i>rac</i> - 6f	6.83	14.64	1827455	1759401	50.95	49.05
(<i>S</i>)- 6f	6.88	14.5	3077119	193503	94.08	5.92



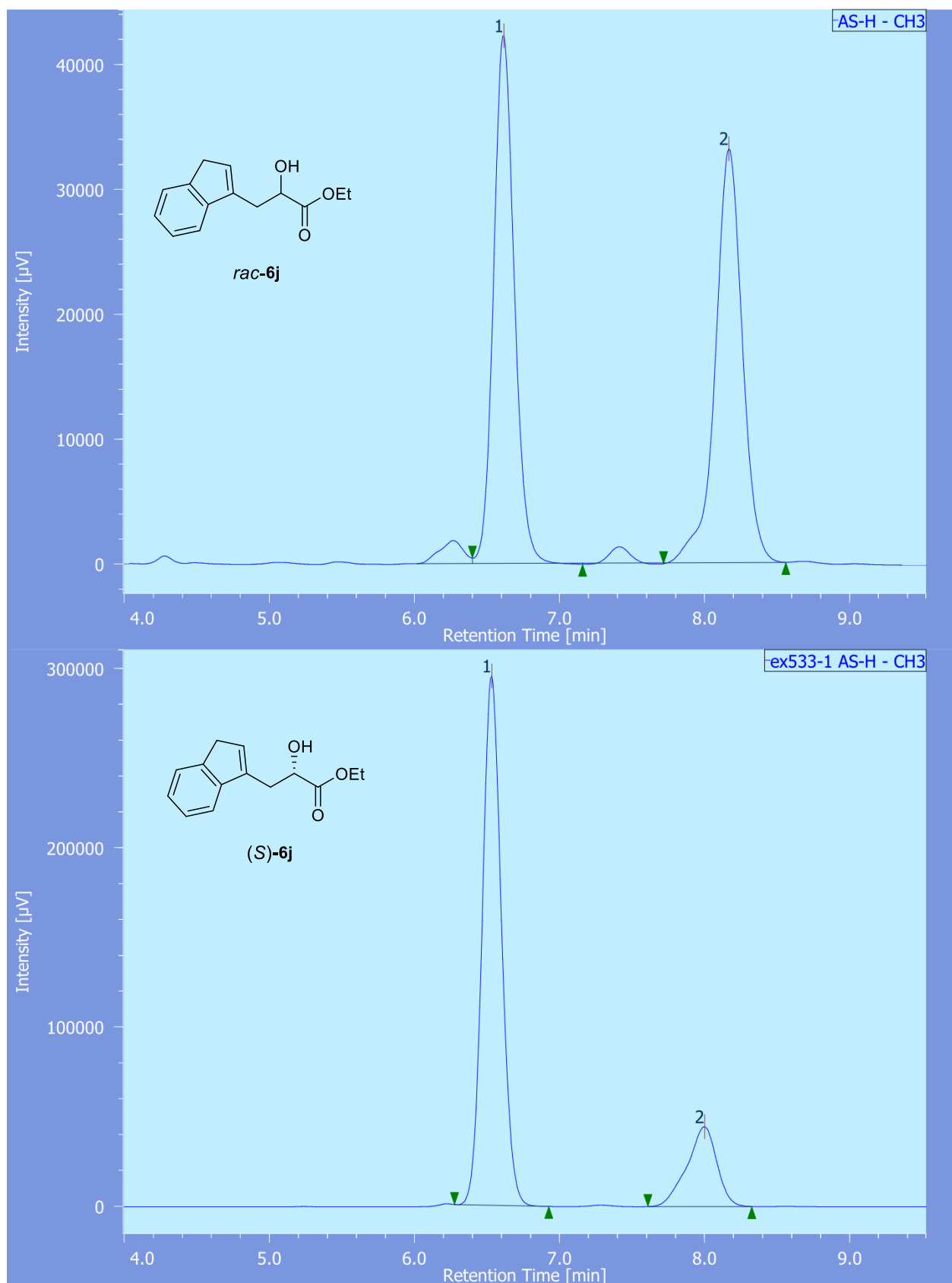
	Retention Time (1)	Retention Time (2)	Area (1)	Area (2)	%Area (1)	%Area (2)
<i>rac</i> -6g	6.91	16.48	1918922	1892939	50.34	49.66
(<i>S</i>)-6g	7.02	17.42	545778	137692	79.85	20.15



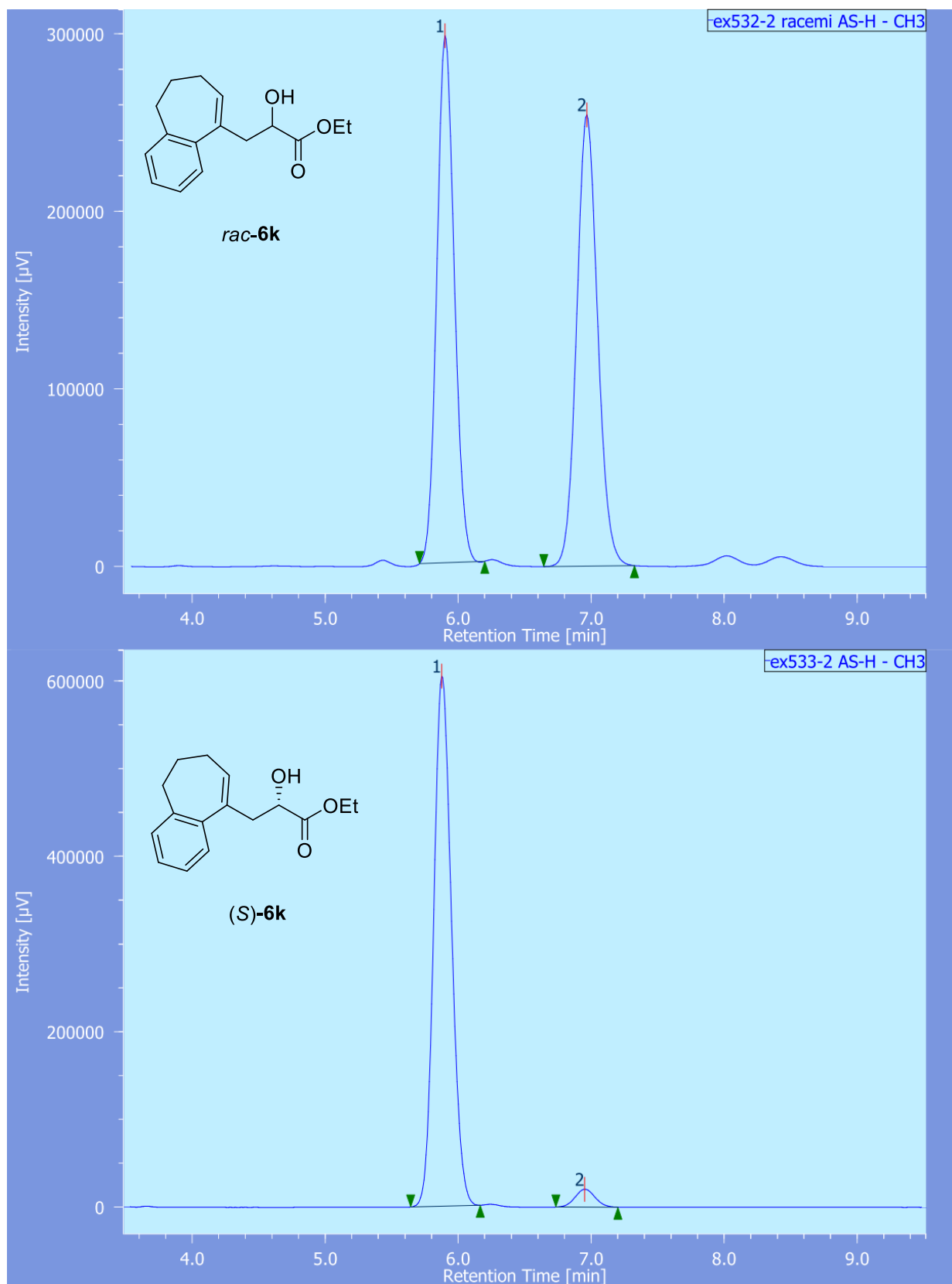
	Retention Time (1)	Retention Time (2)	Area (1)	Area (2)	%Area (1)	%Area (2)
<i>rac</i> -6h	10.72	12.31	6571418	6616825	49.83	50.17
(<i>S</i>)-6h	10.55	12.24	5172358	246307	95.45	4.55



	Retention Time (1)	Retention Time (2)	Area (1)	Area (2)	%Area (1)	%Area (2)
<i>rac</i> - 6i	15.69	17.16	940063	940252	49.99	50.01
(<i>S</i>)- 6i	15.83	17.48	1895098	187383	91.00	9.00



	Retention Time (1)	Retention Time (2)	Area (1)	Area (2)	%Area (1)	%Area (2)
<i>rac</i> -6j	6.62	8.17	415013	430149	49.11	50.89
(<i>S</i>)-6j	6.53	8.00	2708200	638423	80.92	19.08



	Retention Time (1)	Retention Time (2)	Area (1)	Area (2)	%Area (1)	%Area (2)
<i>rac</i> - 6k	5.90	6.97	2707995	2797543	49.19	50.81
(<i>S</i>)- 6k	5.88	6.95	5578569	212968	96.32	3.68