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# **Supporting Information**

Antimicrobial Peptide based Magnetic Recognition
Elements and Au@Ag-GO SERS Tags with Stable Internal
Standards: A Three in One Biosensor for Isolation,
Discrimination and Killing of Multiple Bacteria in Whole
Blood<sup>†</sup>

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- 26 <sup>†</sup> Electronic supplementary information (ESI) available: Additional data and 13 supplementary figures.

1 ‡ K.Y. and Q.M. contributed equally.

### 2 1. Additional data

#### 3 1.1 Surface coverage of 4-MPBA on Au@AgNPs

4 The concentration of AuNPs could be calculated based on the Beer's law and the extinction 5 coefficient ( $\epsilon_{Au}$ =3×10<sup>9</sup> M<sup>-1</sup> cm<sup>-1</sup>). Thus, the concentrations of AuNPs is ~0.26 nM. As the Au@Ag 6 NPs are prepared through the coating of Au seed with Ag shell and 2.5 mL AgNO<sub>3</sub> was added to 7 form the 5 nm Ag shell, we can calculate that the concentration of Au@AgNPs is 0.208 nM.<sup>1</sup>

8 The total surface coverage ( $\theta$ ) of 4-MPBA on the Au@AgNPs surfaces can be calculated as

9 follows according to previous report:<sup>2</sup>

$$10 \quad \theta = \frac{0.25n}{S} = \frac{0.25nN_a}{C_{Au@Ag}V\pi \ d^2N_a} = \frac{0.25n}{C_{Au@Ag}V\pi \ d^2}$$

11 Where

12 a) S is the total surface of Au@AgNPs;

13 b) n is the total amounts of 4-MPBA;

14 c) N<sub>a</sub> is the Avogadro's number;

15 d)  $C_{Au@Ag}$  is the concentration of Au@AgNPs, which was calculated to be 0.208 nM;

16 e) V is the volume of Au@AgNPs colloidal solution;

17 f) d is the average diameter of Au@AgNPs, which is measured ~35 nm

18 In the calculation of surface coverage on Au@AgNPs, different amounts of MPBA were added into the Au@AgNPs to measure the max adsorb amount of MPBA on the Au@AgNPs 19 surfaces. As depicted in Figure S5A, while the final concentrations of MPBA in excess of 0.25 20 21  $\mu$ g/mL, the colloidal solution began to change its color. Thus, we set this value as the max adsorb amount of MPBA. While the total amounts of 4-MPBA (average molecular area of 0.25 nm<sup>2</sup>) 22 added were smaller than the max adsorb amount of 4-MPBA on the Au@AgNPs surfaces, it can 23 be speculated that the amounts of modified MPBA on the Au@AgNPs surfaces and the added 24 amounts of 4-MPBA were the same. In this experiment, we prepared the 4-MPBA modified 25 Au@AgNPs through the mixture of 4-MPBA (6 mL, final concentration: 10 µg/mL) and 26 Au@AgNPs. As a result, the surface coverage of MPBA on the surface of Au@AgNPs was 27 calculated to be 0.51. 28

1 In the calculation of surface coverage on Au@Ag-GO nanocomposites, a 10 µg/mL of 4-2 MPBA solution (6 mL) have been used to mixed with the Au@Ag-GO nanocomposites. After the 3 Au@AgNPs have been adhered to the GO nanosheets, the Au@Ag-GO nanocomposites will not be aggregation even in high concentration of 4-MPBA. UV-Vis results (Figure S5B) showed that 4 large amounts of 4-MPBA have been adsorbed on the Au@Ag-GO nanocomposites and it can be 5 6 calculated that 1.5 mg of 4-MPBA have been adsorbed on the Au@Ag-GO nanocomposites. Thus, 7 the total surface coverage of 4-MPBA on Au@Ag-GO nanocomposites is calculated to be 13.11. The surface coverage is over 1.00 due to the GO nanosheets will also adsorb the 4-MPBA. After 8 the Au@AgNPs on the GO nanosheets are full of 4-MPBA, the GO nanosheets will further adsorb 9 10 the 4-MPBA.

In conclusion, with the combination of GO nanosheets and Au@AgNPs, the adsorb amounts of 4-MPBA on SERS substrate will be significantly enhanced compare with the simple Au@AgNPs substrate.

#### 14 1.2 Enhancement Calculation (EF)

15 The EF value is calculated through the following well-established equation:<sup>3</sup>

$$16 \quad EF = \frac{I_{SERS} \times N_{bulk}}{I_{bulk} \times N_{SERS}} \tag{1}$$

I7 I<sub>bulk</sub> and I<sub>SERS</sub> are the intensity of analyte in solution for SERS and bulk Raman spectra,
 18 respectively. N<sub>bulk</sub> and N<sub>SERS</sub> means the number of molecules within the laser spot excited by a
 19 laser beam in SERS and Raman scattering.

$$20 N_{SERS} = N_A \times CV \frac{S_{Laser}}{S_{Sub}} (2)$$

21  $N_A$  is Avogadro constant; C means the molar concentration; V is the volume;  $S_{Laser}$  is the size 22 of the laser spot and  $S_{Sub}$  is the size of the substrate. Hence, for SERS detection, a  $V_{SERS}$  volume 23 of R6G is dispersed on an area of  $S_{SER}S$  at a concentration of  $C_{SERS}$  on the clean Si substrate.

$$24 N_{\text{bulk}} = N_A \times \rho_v S_{\text{Laser}} (3)$$

 $\rho_{v}$  [mol/µm<sup>3</sup>] means the volume density of R6G powder on a glass slide. In this experiment, mass density of R6G powder is 1.26 g/cm<sup>3</sup>, while molecular weight of R6G is 479 g/mol, thus it 1 can be calculated as  $\rho_v [mol/\mu m^3] = (1.26/479) \times 10^{-12} = 2.63 \times 10^{-15} mol/\mu m^3$ 

$$2 \quad EF = \frac{I_{SERS} \times \rho_v S_{Sub}}{I_{bulk} \times CV}$$
(4)

In our experiment, a 25  $\mu$ L of R6G (10<sup>-9</sup>M) was mixed with 25  $\mu$ L of Au@Ag-GO nanocomposites, then the mixture was drop on the glass slide and dry in the air to form a circle with a diameter of 5195  $\mu$ m. As depicted in Figure S4, SERS signals of R6G was obviously enhanced compared with Raman signals of R6G powder. Therefore, for the 613 cm<sup>-1</sup> Raman peak, I<sub>bulk</sub> is 2054.0 counts from Raman spectrum of R6G powder and I<sub>SERS</sub> is 25410.8 counts from SERS spectrum of R6G. The EF can be calculated as:

9 EF =  $(25410.8 \text{ counts} \times 2.63 \times 10^{-15} \text{ mol/}\mu\text{m}^3 \times (5195 \ \mu\text{m})^2 \times 3.14)/(2054.0 \text{ counts} \times 10^{-9} \text{ M} \times 25 \ \mu\text{L} \times 10^{-6})$ 10 =1.1×10<sup>8</sup>

#### 11 1.3 XRD and FTIR of 4-MPBA modified Au@Ag-GO SERS Tags

12 The powder XRD patterns of GO and Au@Ag-GO are shown in Figure S2C. After the adsorbtion 13 of Au@AgNPs, the presence of intense peaks of (111), (200), and (220) could be indexed to face 14 centered cubic (fcc) structure of Au@AgNPs.<sup>4</sup> These confirm that Au@AgNPs have been adhered 15 to the GO nanosheets successfully.

16 The FTIR spectrum of the GO, and 4-MPBA modified Au@Ag-GO have been measured and results are showed in Figure S2D. The characteristic vibrations of GO are a broad and intense 17 18 peak of O-H group at 3250 cm<sup>-1</sup>, a C=O peak at 1723 cm<sup>-1</sup>, a C-OH stretching peak at 1254 cm<sup>-1</sup>, a C-O stretching peak at 1060 cm<sup>-1</sup>, and a peak attributed to the vibration of graphitic skeletal 19 domains at 1605 cm<sup>-1</sup>. Such fact revealed that the GO surface is functionalized with different 20 kinds of oxygen-containing groups.<sup>5</sup> The absorption bands of 4-MPBA modified Au@Ag-GO at 21 22 1594 cm<sup>-1</sup> was attributed to the C=C stretching vibration of phenyl ring, while the new absorption 23 band at ~1360 cm<sup>-1</sup> could be associated with B-O bond and confirm the presence of the boronic acid derivative.6 24

#### 25 1.4 FTIR of AMP modified Fe<sub>3</sub>O<sub>4</sub>NPs

The FTIR spectrum of the Fe<sub>3</sub>O<sub>4</sub>, SiO<sub>2</sub>@Fe<sub>3</sub>O<sub>4</sub> and AMP@SiO<sub>2</sub>@Fe<sub>3</sub>O<sub>4</sub> have been measured and results are showed in Figure S6. For all the nanomaterials, the Fe-O stretching vibration can be observed at 586 cm<sup>-1</sup>. As well as peaks at 3367 cm<sup>-1</sup> and 1635 cm<sup>-1</sup> are assigned to the -OH 1 stretching vibration due to the existence of surface carboxyl. Compared with the absorption bands 2 of pure Fe<sub>3</sub>O<sub>4</sub>, the characteristic absorption peaks of Si-O-Si at 1063 cm<sup>-1</sup> and 1628 cm<sup>-1</sup> 3 confirmed the formation of silica on the surface of Fe<sub>3</sub>O<sub>4</sub> after the modification with TEOS. For 4 the AMP@SiO<sub>2</sub>@Fe<sub>3</sub>O<sub>4</sub>, the appearance of peaks at 1087 cm<sup>-1</sup>, 1043 cm<sup>-1</sup> indicated C-N aliphatic 5 amines, which confirmed the successful modification of AMP.<sup>7</sup>

> sugar on the bacteria wall

OF



- 11 Figure S2 (A) TEM image of AgNPs; (B) TEM image of Au@AgNPs in low magnification; (C)
- 12 TEM image of Au@AgNPs in high magnification.



Figure S3 (A) SERS spectrum of R6G solution enhanced with Au@AgNPs (red line), AgNPs
(blue line) and AuNPs (brown line); (B) XRD of GO nanosheets (brown line) and Au@Ag-GO
nanocomposites (blue line); (C) FTIR of GO (brown line) and 4-MPBA modified Au@Ag-GO
(blue line); (D) Raman spectrum of 4-MPBA adsorbed on Au@Ag-GO nanocomposites for
different storage times.



Figure S4 Raman spectrum of R6G powder on a glass slide and SERS spectra of R6G solution

(10<sup>-9</sup> M).



Figure S5 (A) Different amounts of MPBA mixed with Au@AgNPs, final concentrations of
MPBA (from right to left) were 0, 0.032, 0.063, 0.125, 0.25, 0.5 μg/mL, respectively; (B) UV
spectra of the 10 μg/mL MPBA solution (6 mL, green line), and the supernatant (red line) after 10
μg/mL of MPBA solution were mixed with Au@Ag-GO nanocomposites. The Au@Ag-GO
nanocomposites are synthesis from 6mL of Au@AgNPs (Au@AgNPs/GO ratio: 10:1), and all the
Au@Ag-GO nanocomposites are collected and resolved in 6 mL of pure water.







Figure S6 FTIR spectra of (a)  $Fe_3O_4$ , (b)  $SiO_2@Fe_3O_4$  and (c)  $AMP@SiO_2@Fe_3O_4$ .



2 Figure S7 Sequestration of the pyrophosphate group by AMP. (A) A semitransparent surface
3 representation is shown to highlight the almost complete burial of the target's pyrophosphate
4 group by AMP; (B) The AMP still keep burial of the target's pyrophosphate group even after
5 peptide modification.





8 Figure S8 (A) SERS spectra of *P. auruginosa*, *S. aureus*, and *E. coli* (with 4-MPBA), with
9 concentrations of 1×10<sup>4</sup> CFU/mL respectively; (B) Label free detection of *P. auruginosa*, *S. aureus*, and *E. coli* (without 4-MPBA), with concentrations of 1×10<sup>8</sup> CFU/mL respectively. In this
11 situation, AgNPs were simply mixed with bacteria for SERS detection.



2 Figure S9 (A) Peak intensities of 15 batches (*E.coli*) with (I<sub>1586 cm<sup>-1</sup></sub> / I<sub>1188 cm<sup>-1</sup></sub>) and (B) without
3 (I<sub>1188 cm<sup>-1</sup></sub>) 4-MPBA internal standard normalization; (C) Peak intensities of 15 batches (*S.aureus*)
4 with (I<sub>1586 cm<sup>-1</sup></sub> / I<sub>1188 cm<sup>-1</sup></sub>) and (D) without (I<sub>1188 cm<sup>-1</sup></sub>) 4-MPBA internal standard normalization.



7 Figure S10 SERS mapping in the detection of *E. coli* at different concentrations of 1×10<sup>1</sup>(A),
8 1×10<sup>2</sup>(B), 1×10<sup>3</sup>(C), 1×10<sup>4</sup>(D), 1×10<sup>5</sup>(E), 1×10<sup>6</sup>(F) CFU/mL.



Figure S11 Cell morphology microscopic pictures showing the cytotoxicity of "AMP modified
Fe<sub>3</sub>O<sub>4</sub>NPs" against RAW264.7 cells. (A) control group (untreated cell lines); (B-D) Treated cells
with 800 µg/mL AMP-Fe<sub>3</sub>O<sub>4</sub>NPs (B), 400 µg/mL AMP-Fe<sub>3</sub>O<sub>4</sub>NPs (C), 200 µg/mL AMPFe<sub>3</sub>O<sub>4</sub>NPs (D); (E) Positive control group (cell lines treated with DOX).



8 Figure S12 SERS spectra of whole blood from 39 patients infected with P. aeruginosa, S. aureus





- 1 Figure S13 SERS spectra of blood spiked with P. aeruginosa, S. aureus and E. coli. Blood
- 2 without any bacteria is used as a control.

## 3 **Reference**

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