

Electronic Supplementary Information (ESI)

Synthesis of glycopolymers with specificity for bacterial strains via bacteria-guided polymerization

Yan Luo,^{a,b} Yan Gu,^{a,b} Ruyan Feng,^{a,b} John Brash,^{a,c} Ahmed M. Eissa,^d David M. Haddleton,^{*d} Gaojian Chen ^{*a,b} and
Hong Chen ^{*a}

- a. The Key Lab of Health Chemistry and Molecular Diagnosis of Suzhou, College of Chemistry, Chemical Engineering and Materials Science, Soochow University, 199 Ren-Ai Road, Suzhou, 215123, P. R. China
- b. Center for Soft Condensed Matter Physics and Interdisciplinary Research & School of Physical Science and Technology, Soochow University, Suzhou, 215006, P. R. China
- c. School of Biomedical Engineering and Department of Chemical Engineering, McMaster
- d. Department of Chemistry, University of Warwick, Gibbet Hill Road, Coventry, CV4 7AL (UK)University, Hamilton, Ontario, L8S4L7, Canada

E-mail: d.m.haddleton@warwick.ac.uk; gchen@suda.edu.cn;

chenh@suda.edu.cn

Characterization of polymers by NMR and GPC

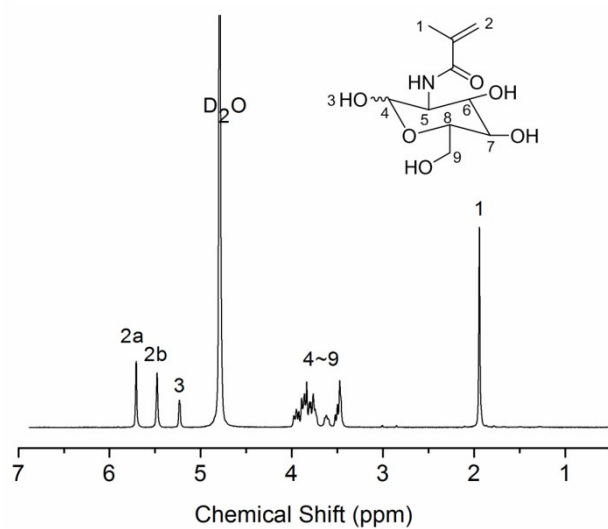


Figure S1 ¹H-NMR spectrum of MAG.

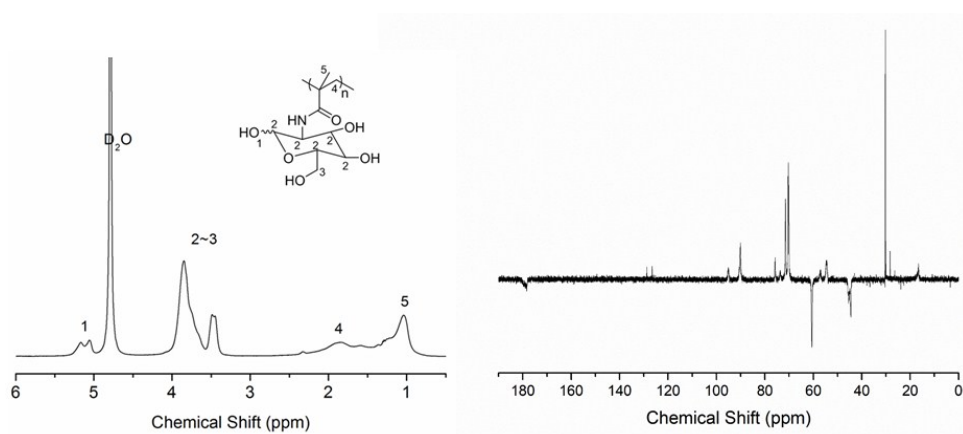


Figure S2 ¹H-NMR and ¹³C-NMR spectrum of pMAG

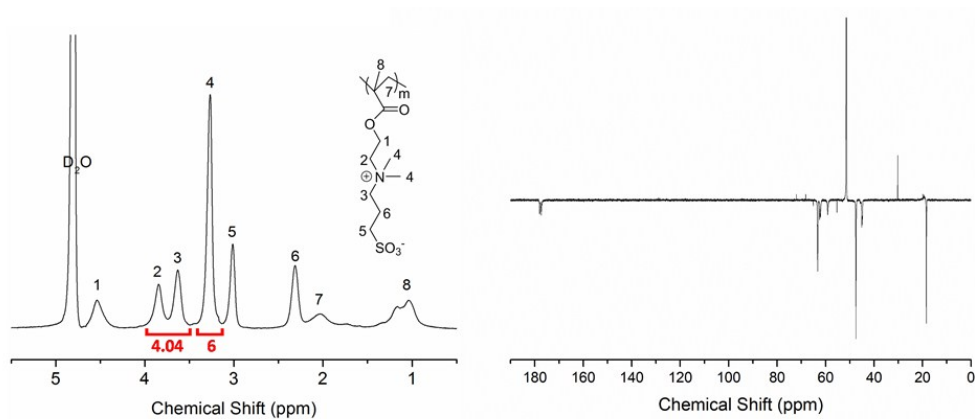


Figure S3 ¹H-NMR and ¹³C-NMR spectrum of pMEDSA

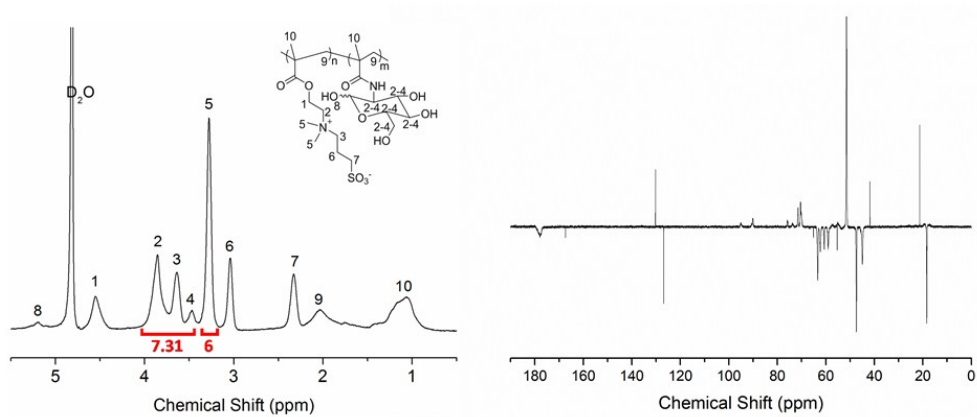


Figure S4 $^1\text{H-NMR}$ and $^{13}\text{C-NMR}$ spectrum of NP

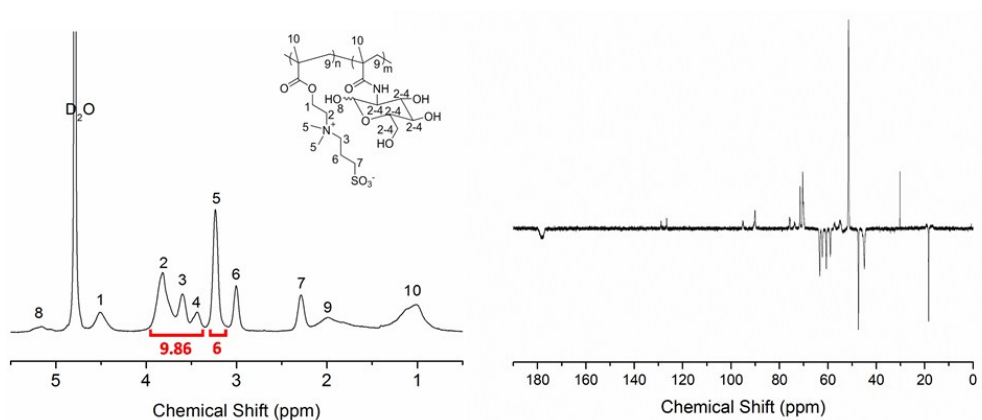


Figure S5 $^1\text{H-NMR}$ and $^{13}\text{C-NMR}$ spectrum of RP

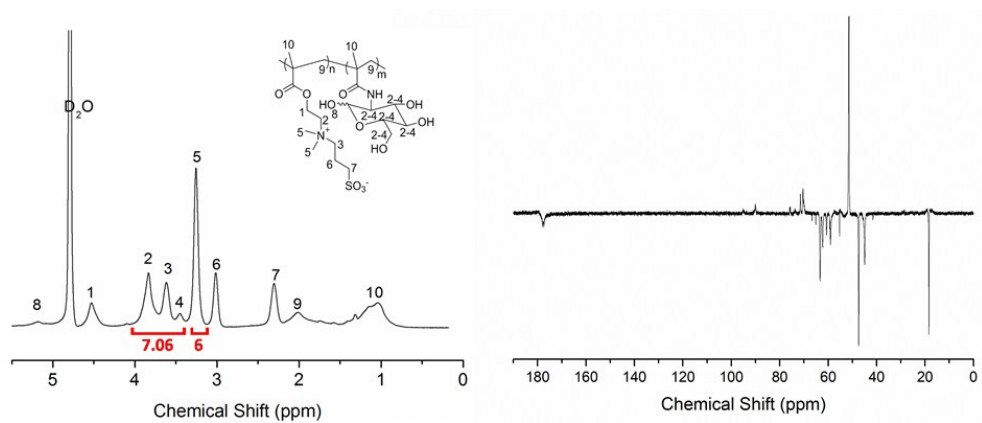


Figure S6 $^1\text{H-NMR}$ and $^{13}\text{C-NMR}$ spectrum of SP

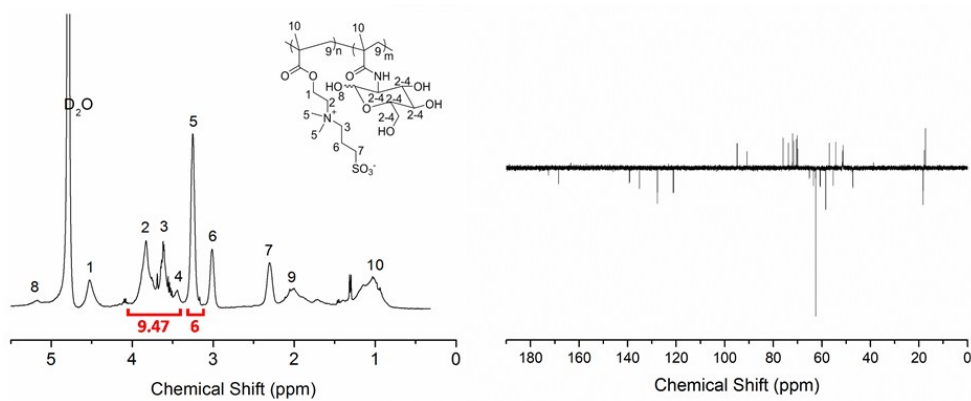


Figure S7 $^1\text{H-NMR}$ and $^{13}\text{C-NMR}$ spectrum of BP

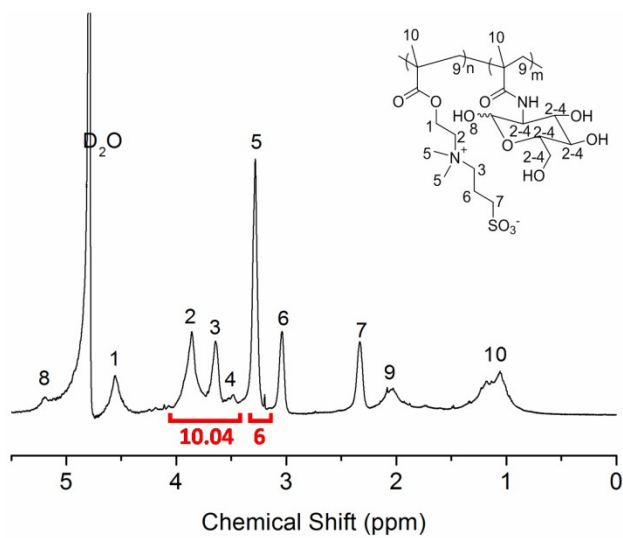


Figure S8 $^1\text{H-NMR}$ spectrum of SP2.

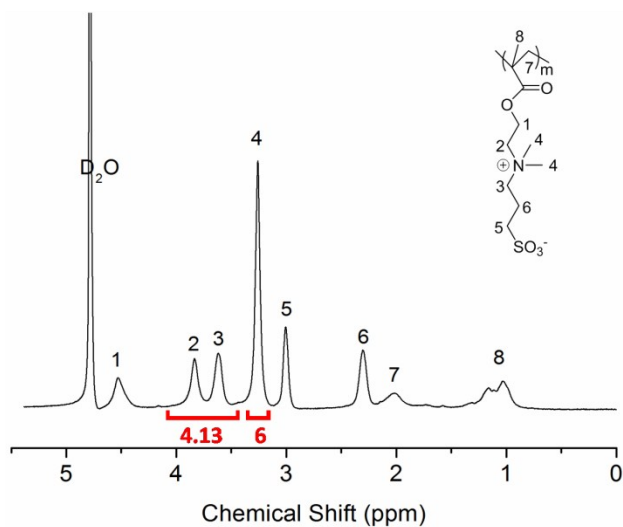


Figure S9 $^1\text{H-NMR}$ spectrum of BP2.

GPC measurements

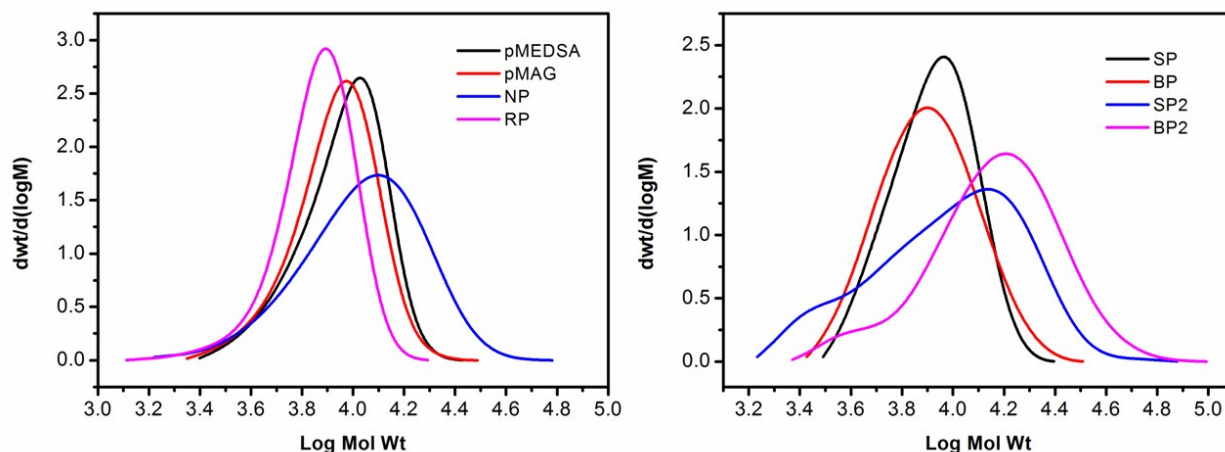


Figure S10 The GPC elution profiles of polymers.

Calculation of monomer composition of copolymers

The composition ratio was calculated by comparing the CH_3 (peak 5, Figure S3-S9) of quaternary amine which belong to MEDSA and protons (peak 2-4, Figure S3-S9) belonging to MAG and MEDSA (2~4, Figure S5). The results of calculation are shown in the Table S1.

Bacteria aggregation

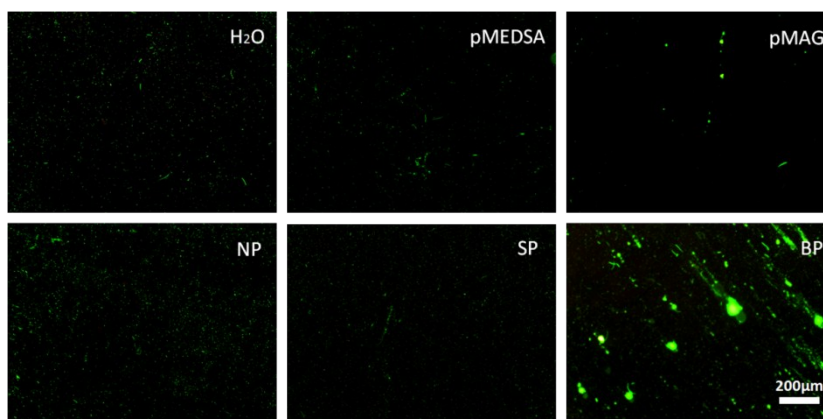


Figure S11 Template bacteria (*E. coli* MG1655) clusters by different treatment.

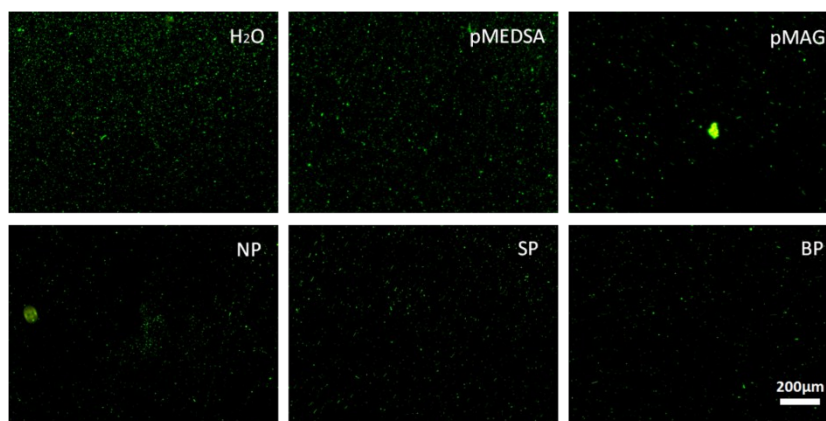


Figure S12 Non-template bacteria (*E. coli* DH5α) clusters by different treatment.

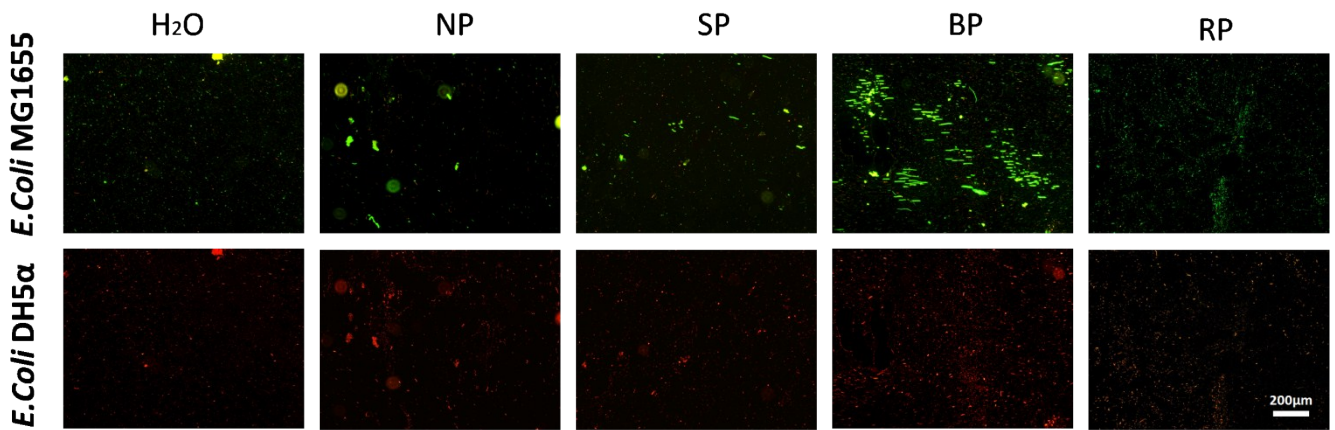


Figure S13 Both the two strains of bacteria clusters by different treatment in the mixed state.

QCM results

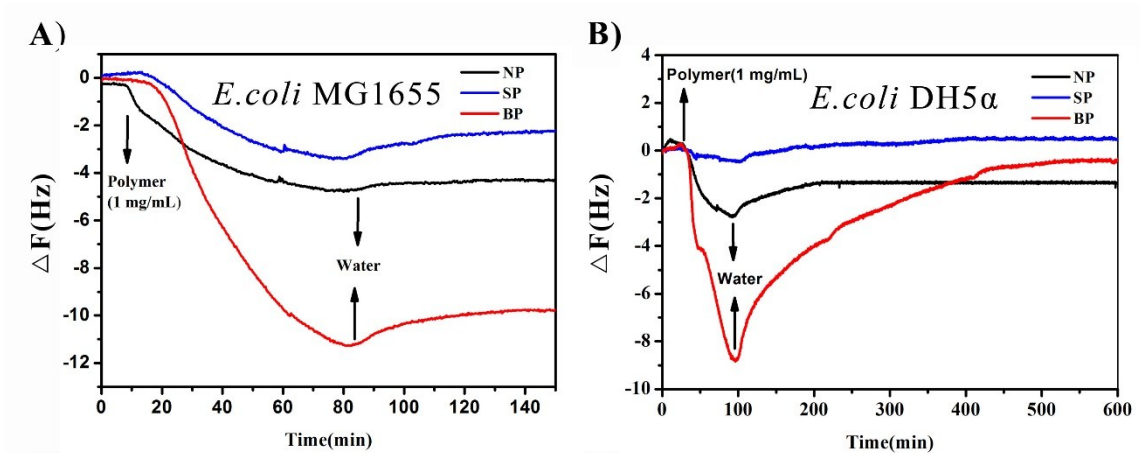


Figure S14 A) Frequency change with time of template bacteria layers on contact with the different polymer solutions (NP, SP, BP). B) Frequency change with time of non-template bacteria layers on contact with the different polymer solutions (NP, SP, BP)

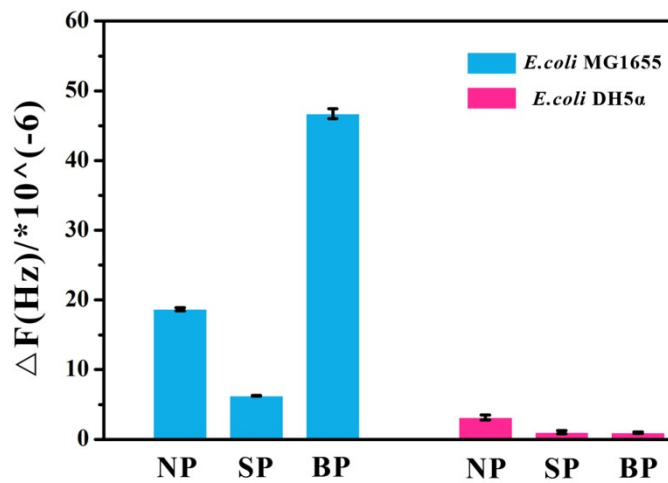


Figure S15 Final frequency change following the interaction between bacteria and polymers.

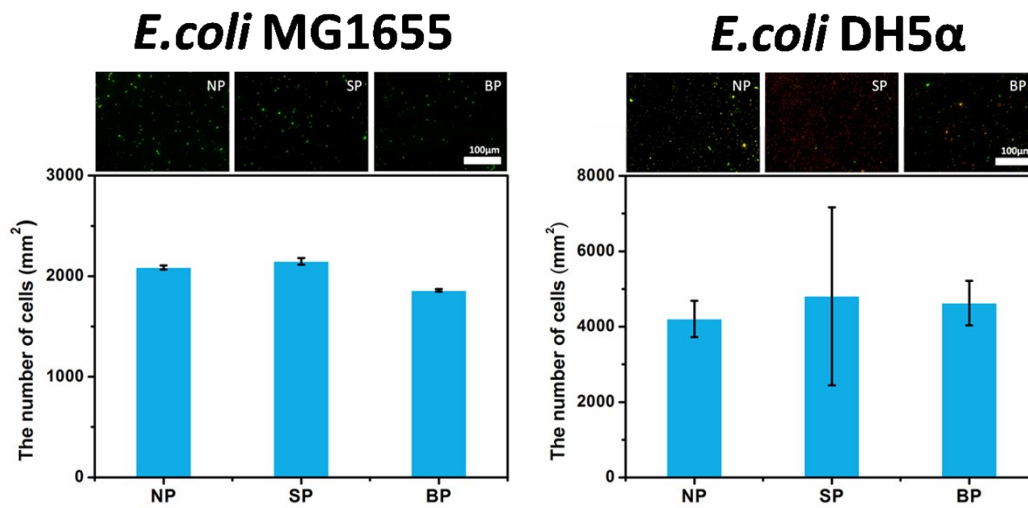


Figure S16 Number of bacteria adhered to the chips.

Bacterial aggregation by turbidimetry

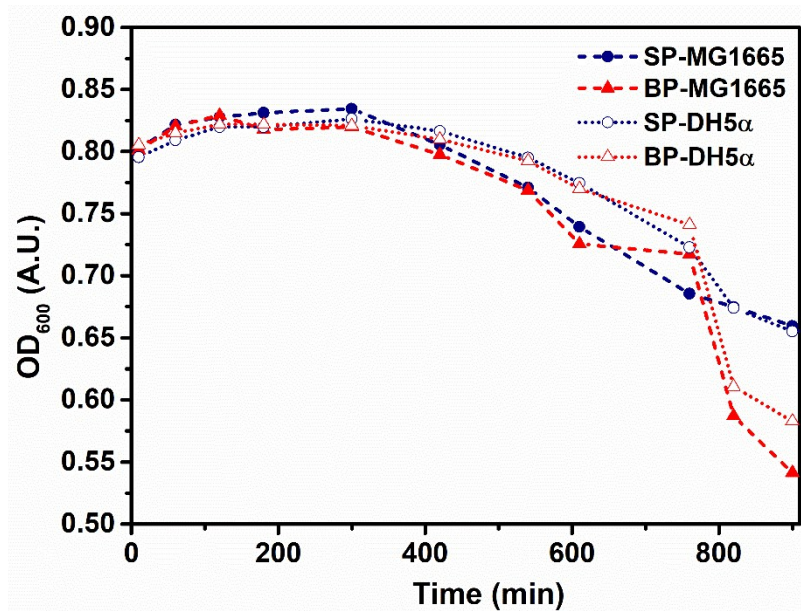


Figure S17 The change of optical density (600nm) for both template (MG1655) and non-template (DH5α) bacteria with time after addition of glycopolymers (SP, BP).

Characterization of Golden nano-particles (GNPs) decorated with polymers

TEM measurements

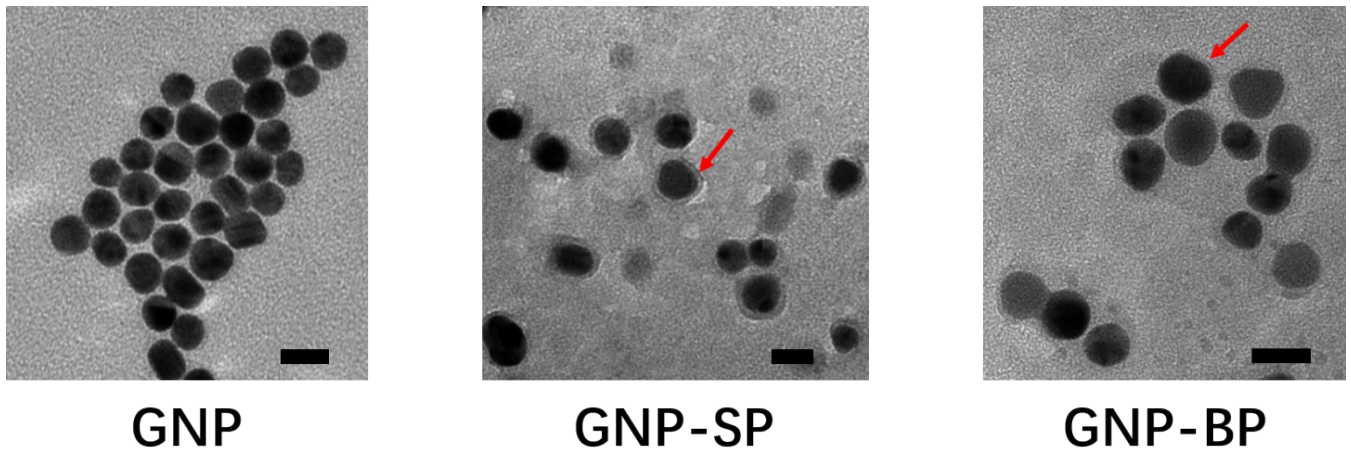


Figure S18 TEM images of GNPs before and after modification with glycopolymers (scale bar =15 nm).

Dynamic Light scattering (DLS) measurements

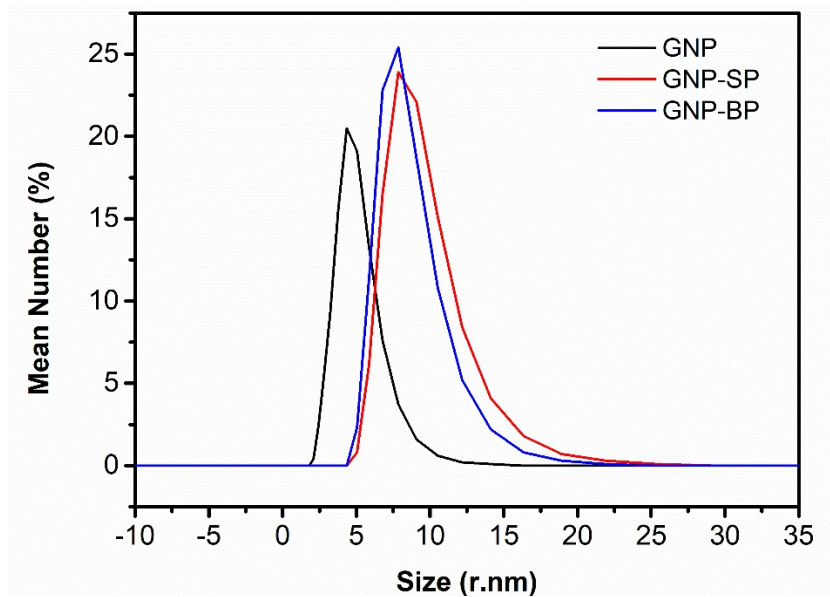


Figure S19 DLS characterization of GNP-SP, GNP-BP and GNP.

Zeta potential measurements

Table S1 The zeta potential of GNP-SP, GNP-BP, and GNP

Samples	GNP	GNP-SP	GNP-BP
Zeta Potential (mV)	-36.2 ± 1.7	-30.6 ± 2.6	-32.2 ± 1.2

Inhibition experiments

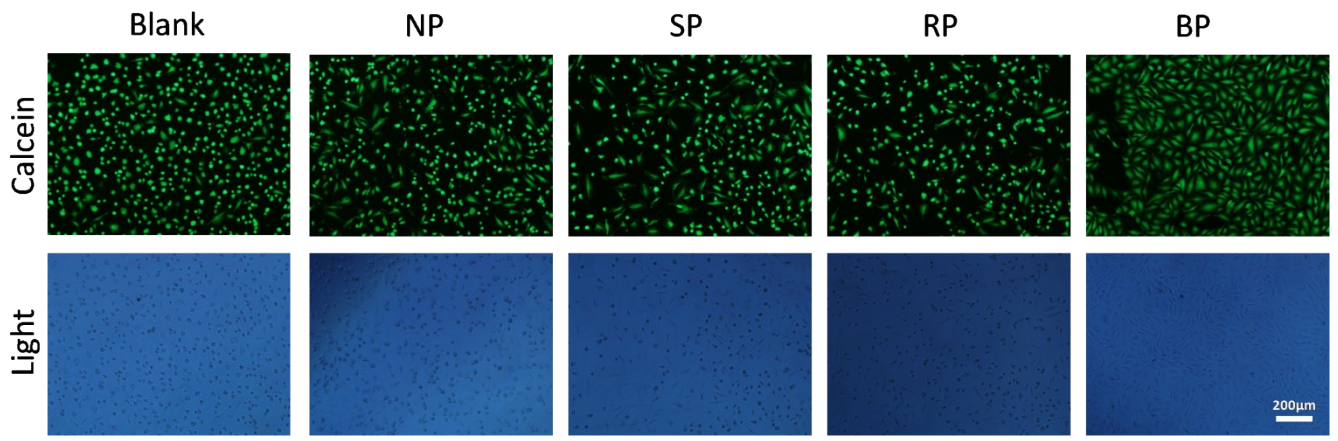


Figure S20 Inhibiting effects of glycopolymers in the anti-infection experiment.