#### **Supporting Information**

# Copper-Catalyzed 1,4-Alkylarylation of 1,3-Enynes Masked Alkyl Electrophiles

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Materials and methods	3				
Synthesis of 1,3-enynes	4				
Characterization data for 1,3-enynes	5				
Synthesis of alkyl diacyl peroxides	9				
Characterization data for diacyl peroxides	10				
Optimization of the reaction conditions					
General procedure for Table S1:	12				
General procedure for 1,4-alkylarylation of 1,3-enynes	13				
General procedure A	13				
General procedure B	13				
Characterization data for products	14				
Synthetic applications					
a) Synthesis of indenyl iodide 7 with NIS					
b) Synthesis of 1 <i>H</i> -indene 8					
c) Synthesis of 2 <i>H</i> -pyran 9	31				
d) Synthesis of 2 <i>H</i> -pyran 10	31				
Preliminary mechanism study					
a) Radical trapping experiment					
b) Radical clock experiments					
c) Radical dimerization					
Single crystal data of 20					
Reference					
NMR spectra					

## Materials and methods

All reactions were carried out under an atmosphere of nitrogen in an flame-dried glassware with magnetic stirring unless otherwise indicated. Commercially obtained reagents were used as received. Solvents were dried by Innovative Technology Solvent Purification System. Liquids and solutions were transferred via syringe. All reactions were monitored by thin-layer chromatography. GC-MS data were recorded on Thermo ISQ QD. <sup>1</sup>H, <sup>19</sup>F, and <sup>13</sup>C NMR spectra were recorded on Bruker-BioSpin AVANCE III HD and JEOL ECZ600S. Data for <sup>1</sup>H NMR spectra are reported relative to TMS as an internal standard (0.00 ppm) and are reported as follows: chemical shift (ppm), multiplicity, coupling constant (Hz), and integration. Data for <sup>13</sup>C NMR spectra are reported relative to chloroform as an internal standard (77.16 ppm) and are reported in terms of chemical shift (ppm). HRMS data were recorded on Thermo Fisher Scientific LTQ FTICR-MS, Waters Micromass GCT Premier or Thermo Finnigan DECAX-30000 LCQ Deca XP.





To a 50 mL round bottomed flask was charged with terminal alkyne (5 mmol, 1 equiv) and 10 mL of THF. The solution was cooled to -78 °C and *n*-BuLi (2.5 M in THF, 2 mL, 5 mmol, 1 equiv) was added. The resulting solution was stirred for 20 minutes at room temperature and then cooled to -78 °C again. Ketone (5 mmol, 1 equiv) in THF solution was added dropwise. The reaction mixture was then allowed to warm to room temperature and was monitored by TLC for completion. On completion the reaction was quenched with saturated aqueous NH<sub>4</sub>Cl (40 mL). The aqueous layer was extracted with ethyl acetate and the combined organic layers were washed with brine (30 mL), dried over MgSO<sub>4</sub> and filtered. Then concentrated under reduced pressure to afford the crude propargyl alcohol<sup>1</sup>.

The resulting crude propargyl alcohol was dissolved in dry DCM (40 mL), and the mixture was cooled to 0 °C with a cooling bath. To this solution was added TEA (25 mmol, 5 equiv) and methylsulfonyl chloride (12.5 mmol, 2.5 equiv) sequentially. After 30 min the reaction was monitored by TLC for completion. Once completion the reaction was quenched with saturated aqueous NH<sub>4</sub>Cl (40 mL). The aqueous layer was extracted with ethyl acetate and the combined organic layers were washed with brine (30 mL), dried over MgSO<sub>4</sub>, filtered, and concentrated under reduced pressure. The crude material was purified by flash chromatography to yield the 1,3-enyne<sup>2</sup>.

## Characterization data for 1,3-enynes



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.49 (d, *J* = 8.0 Hz, 2H), 7.15 (d, *J* = 7.9 Hz, 2H), 5.82 (s, 1H), 5.54 (s, 1H), 3.70 (t, *J* = 6.4 Hz, 2H), 2.61 (t, *J* = 6.8 Hz, 2H), 2.35 (s, 3H), 2.05 (p, *J* = 6.6 Hz, 2H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  138.15, 134.78, 130.47, 129.05, 125.91, 119.14, 89.42, 80.90, 43.81, 31.43, 21.20, 16.88. HRMS (EI+) calcd for [C<sub>14</sub>H<sub>15</sub>Cl]<sup>+</sup> ([M]<sup>+</sup>): 218.0862, found: 218.0867.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.55 (d, J = 8.2 Hz, 2H), 7.14 (d, J = 7.9 Hz, 2H), 5.79 (s, 1H), 5.52 (s, 1H), 2.40 (t, J = 7.1 Hz, 2H), 2.35 (s, 3H), 1.66 – 1.54 (m, 2H), 1.51 – 1.39 (m, 2H), 1.37 – 1.27 (m, 4H), 0.94 – 0.86 (m, 3H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*) δ 137.98, 135.03, 130.77, 128.96, 125.97, 118.47, 91.88, 79.88, 31.39, 28.74, 28.68, 22.61, 21.18, 19.44, 14.10. HRMS (EI+) calcd for  $[C_{17}H_{22}]^+$  ([M]<sup>+</sup>): 226.1722, found: 226.1730.



<sup>1</sup>H NMR (600 MHz, Chloroform-*d*) δ 7.66 – 7.57 (m, 2H), 7.06 – 6.97 (m, 2H), 5.76 (s, 1H), 5.54 (s, 1H), 2.40 (t, J = 7.1 Hz, 2H), 1.64 – 1.54 (m, 2H), 1.48 – 1.41 (m, 2H), 1.35 – 1.29 (m, 4H), 0.91 – 0.88 (m, 3H). <sup>13</sup>C NMR (150 MHz, Chloroform-*d*) δ 162.81 (d, J = 247.5 Hz), 134.01 (d, J = 3.0 Hz), 130.01, 127.86 (d, J = 8.1 Hz), 119.10, 115.16 (d, J = 21.5 Hz), 92.42, 79.67, 31.42, 28.76, 28.73, 22.66, 19.47, 14.13. HRMS (EI+) calcd for  $[C_{16}H_{19}F]^+$  ([M]<sup>+</sup>): 230.1471, found: 230.1474.



<sup>1</sup>H NMR (600 MHz, Chloroform-*d*) δ 7.55 – 7.50 (m, 2H), 7.48 – 7.43 (m, 2H), 5.82 (s, 1H), 5.59 (s, 1H), 2.41 (t, J = 7.2 Hz, 2H), 1.64 – 1.56 (m, 2H), 1.48 – 1.41 (m, 2H), 1.36 – 1.30 (m, 4H), 0.91 (t, J = 6.9 Hz, 3H). <sup>13</sup>C NMR (150 MHz, Chloroform-*d*) δ 136.85, 131.44, 130.11, 127.80, 122.28, 119.80, 92.68, 79.39, 31.46, 28.77, 22.70, 19.50, 14.18. HRMS (EI+) calcd for  $[C_{16}H_{19}Br]^+$  ([M]<sup>+</sup>): 290.0670, found: 290.0675.

C<sub>6</sub>H<sub>13</sub> 1e Ph

<sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.76 – 7.69 (m, 2H), 7.63 – 7.54 (m, 4H), 7.46 – 7.39 (m, 2H), 7.37 – 7.30 (m, 1H), 5.88 (s, 1H), 5.60 (s, 1H), 2.42 (t, *J* = 7.1 Hz, 2H), 1.67 – 1.57 (m, 2H), 1.52 – 1.42 (m, 2H), 1.37 – 1.29 (m, 4H), 0.95 – 0.86 (m, 3H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)

 $\delta$  140.92, 140.68, 136.83, 130.59, 128.81, 127.41, 127.05, 126.99, 126.53, 119.25, 92.24, 79.75, 31.40, 28.76, 28.71, 22.63, 19.48, 14.11. The spectrum data matches previously reported values<sup>3</sup>.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.63 (s, 1H), 7.55 – 7.48 (m, 1H), 7.27 – 7.23 (m, 2H), 5.83 (s, 1H), 5.60 (s, 1H), 2.40 (t, J = 7.1 Hz, 2H), 1.63 – 1.56 (m, 2H), 1.48 – 1.41 (m, 2H), 1.35 – 1.30 (m, 4H), 0.90 (t, J = 6.8 Hz, 3H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*) δ 139.67, 134.31, 129.87, 129.47, 128.09, 126.37, 124.15, 120.31, 92.70, 79.19, 31.40, 28.70, 28.67, 22.63, 19.42, 14.10. HRMS (EI+) calcd for [C<sub>16</sub>H<sub>19</sub>CI]<sup>+</sup> ([M]<sup>+</sup>): 246.1175, found: 246.1185.

C<sub>6</sub>H<sub>13</sub> 1g

<sup>1</sup>H NMR (600 MHz, Chloroform-*d*) δ 7.57 – 7.39 (m, 2H), 7.26 – 7.21 (m, 1H), 7.14 – 7.09 (m, 1H), 5.83 (d, J = 1.3 Hz, 1H), 5.57 (d, J = 1.2 Hz, 1H), 2.42 (t, J = 7.1 Hz, 2H), 2.38 (s, 3H), 1.65 – 1.59 (m, 2H), 1.51 – 1.45 (m, 2H), 1.36 – 1.30 (m, 4H), 0.92 (t, J = 7.0 Hz, 3H). <sup>13</sup>C NMR (150 MHz, Chloroform-*d*) δ 137.91, 137.89, 131.15, 128.96, 128.26, 126.91, 123.30, 119.32, 92.04, 79.98, 31.48, 28.81, 28.76, 22.70, 21.57, 19.53, 14.17. HRMS (EI+) calcd for  $[C_{17}H_{22}]^+$  ([M]<sup>+</sup>): 226.1722, found: 226.1719.



<sup>1</sup>H NMR (600 MHz, Chloroform-*d*) δ 7.29 – 7.23 (m, 1H), 7.23 – 7.16 (m, 3H), 5.69 (d, J = 1.9 Hz, 1H), 5.39 (d, J = 2.0 Hz, 1H), 2.45 (s, 3H), 2.34 (t, J = 7.1 Hz, 2H), 1.58 – 1.53 (m, 2H), 1.44 – 1.38 (m, 2H), 1.33 – 1.28 (m, 4H), 0.90 (t, J = 6.9 Hz, 3H). <sup>13</sup>C NMR (150 MHz, Chloroform-*d*) δ 140.10, 135.52, 132.36, 130.39, 128.76, 127.75, 125.88, 123.91, 92.07, 80.71, 31.44, 28.74, 22.65, 20.28, 19.56, 14.13. HRMS (EI+) calcd for  $[C_{17}H_{22}]^+$  ([M]<sup>+</sup>): 226.1722, found: 226.1729.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 8.14 (s, 1H), 7.90 – 7.72 (m, 4H), 7.46 (m, 2H), 5.98 (s, 1H), 5.67 (s, 1H), 2.47 (t, J = 7.0 Hz, 2H), 1.72 – 1.59 (m, 2H), 1.55 – 1.47 (m, 2H), 1.40 – 1.31 (m, 4H), 0.91 (t, J = 7.0 Hz, 3H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*) δ 135.08, 133.28, 133.15, 130.91, 128.42, 127.82, 127.55, 126.22, 126.16, 125.91, 123.46, 119.69, 92.31, 79.78, 31.41, 28.76, 28.71, 22.63, 19.51, 14.10. The spectrum data matches previously reported values<sup>3</sup>.



<sup>1</sup>H NMR (600 MHz, Chloroform-*d*) δ 7.53 – 7.43 (m, 2H), 7.36 – 7.32 (m, 2H), 7.30 – 7.24 (m, 3H), 7.16 – 7.11 (m, 2H), 5.82 (d, J = 1.2 Hz, 1H), 5.54 (d, J = 1.2 Hz, 1H), 2.95 (t, J = 7.5 Hz, 2H), 2.74 (t, J = 7.5 Hz, 2H), 2.37 (s, 3H). <sup>13</sup>C NMR (150 MHz, Chloroform-*d*) δ 140.76, 138.09, 134.89, 130.73, 129.05, 128.69, 128.53, 126.44, 126.07, 118.79, 90.88, 80.77, 35.18, 21.69, 21.27. HRMS (EI+) calcd for [C<sub>19</sub>H<sub>18</sub>]<sup>+</sup> ([M]<sup>+</sup>): 246.1409, found: 246.1413.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.71 – 7.59 (m, 2H), 7.36 – 7.26 (m, 3H), 5.83 (d, J = 1.2 Hz, 1H), 5.58 (d, J = 1.1 Hz, 1H), 2.31 (d, J = 6.6 Hz, 2H), 1.90 – 1.84 (m, 2H), 1.77 – 1.64 (m, 4H), 1.31 – 1.10 (m, 5H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*) δ 137.87, 131.04, 128.26, 128.08, 126.08, 119.32, 90.97, 80.61, 37.52, 32.84, 27.26, 26.31, 26.18. HRMS (EI+) calcd for  $[C_{17}H_{20}]^+$  ([M]<sup>+</sup>): 224.1565, found: 224.1574.



<sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.66 – 7.61 (m, 2H), 7.35 – 7.32 (m, 2H), 7.30 – 7.27 (m, 1H), 5.85 (d, *J* = 1.1 Hz, 1H), 5.59 (d, *J* = 1.1 Hz, 1H), 3.67 (s, 3H), 2.51 – 2.47 (m, 4H), 1.95 – 1.90 (m, 2H).<sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  173.69, 137.68, 130.83, 128.41, 128.28, 126.11, 119.93, 90.51, 80.71, 51.70, 33.01, 24.00, 18.96. HRMS (ESI) calcd for [C<sub>15</sub>H<sub>26</sub>O<sub>2</sub>Na]<sup>+</sup> ([M+Na]<sup>+</sup>): 251.1043, found: 251.1042.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.70 – 7.55 (m, 2H), 7.36 – 7.24 (m, 3H), 5.85 (s, 1H), 5.61 (s, 1H), 3.82 – 3.69 (m, 2H), 2.83 – 2.73 (m, 1H), 2.63 (t, J = 6.5 Hz, 2H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*) δ 137.49, 130.60, 128.43, 128.34, 126.08, 120.34, 88.30, 81.46, 61.13, 23.75. HRMS (EI+) calcd for  $[C_{12}H_{12}O]^+$  ([M]<sup>+</sup>): 172.0888, found: 172.0894.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.60 – 7.45 (m, 4H), 7.38 – 7.23 (m, 5H), 6.46 (q, J = 6.9 Hz, 1H), 2.12 (d, J = 7.2 Hz, 3H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*) δ 136.81, 133.70, 133.23, 131.60, 128.55, 128.48, 128.44, 127.24, 123.51, 123.37, 96.02, 86.26, 17.19. HRMS (EI+) calcd for  $[C_{17}H_{13}CI]^+$  ([M]<sup>+</sup>): 252.0706, found: 252.0698.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.66 – 7.49 (m, 2H), 7.35 – 7.27 (m, 2H), 7.26 – 7.19 (m, 1H), 6.39 (q, J = 6.9 Hz, 1H), 2.46 (t, J = 7.0 Hz, 2H), 2.04 (d, J = 6.9 Hz, 3H), 1.66 – 1.57 (m, 2H), 1.53 – 1.46 (m, 2H), 1.35 – 1.29 (m, 4H), 0.93 – 0.88 (m, 3H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*) δ 138.94, 131.52, 128.21, 127.14, 125.87, 124.77, 96.85, 77.71, 31.38, 28.95, 28.64, 22.61, 19.64, 16.78, 14.07. The spectrum data matches previously reported values<sup>3</sup>.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.76 – 7.69 (m, 2H), 7.58 – 7.50 (m, 2H), 7.41 – 7.31 (m, 6H), 5.98 (s, 1H), 5.76 (s, 1H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*) δ 137.28, 131.68, 130.62,

128.42, 128.36, 126.11, 123.11, 120.68, 90.78, 88.56. The spectrum data matches previously reported values<sup>3</sup>.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.70 – 7.55 (m, 2H), 7.39 – 7.25 (m, 3H), 5.81 (d, *J* = 1.1 Hz, 1H), 5.55 (d, *J* = 1.2 Hz, 1H), 1.53 – 1.33 (m, 1H), 0.87 – 0.77 (m, 4H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  137.89, 130.96, 128.36, 128.21, 126.14, 119.55, 95.20, 75.10, 8.77, 0.30. The spectrum data matches previously reported values<sup>4</sup>.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.86 – 7.47 (m, 2H), 7.39 – 7.23 (m, 7H), 7.22 – 7.15 (m, 1H), 6.43 (d, J = 16.0 Hz, 1H), 6.22 (dt, J = 15.9, 7.0 Hz, 1H), 5.85 (s, 1H), 5.60 (s, 1H), 2.46 (t, J = 7.1 Hz, 2H), 2.41 – 2.33 (m, 2H), 1.84 – 1.73 (m, 2H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*) δ 137.79, 137.70, 130.94, 130.80, 129.75, 128.58, 128.37, 128.22, 127.04, 126.13, 126.05, 119.64, 91.58, 80.26, 32.23, 28.41, 18.96. MS (EI+) calcd for  $[C_{21}H_{20}CI]^+$  ([M]<sup>+</sup>): 272.1, found: 272.3.



<sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.64 – 7.61 (m, 2H), 7.45 – 7.41 (m, 2H), 7.19 (d, *J* = 7.9 Hz, 2H), 7.15 (d, *J* = 7.8 Hz, 2H), 5.92 (d, *J* = 1.1 Hz, 1H), 5.69 (d, *J* = 1.3 Hz, 1H), 2.37 (s, 3H), 2.36 (s, 3H). <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  138.62, 138.32, 134.70, 131.68, 130.62, 129.22, 129.18, 126.12, 120.19, 119.57, 90.89, 88.20, 21.63, 21.30. The spectrum data matches previously reported values<sup>5</sup>.

## Synthesis of alkyl diacyl peroxides

Alkyl diacyl peroxides have potentials to explode. Any alkyl diacyl peroxides involved reaction (as product or substrate) should be carried out with precautions!

Lauroyl peroxide (LPO) 2a was purchased from Admas. Other peroxides were prepared according to our previous work<sup>6</sup>.

A solution of DMAP (0.6 mmol), 30% hydrogen peroxide (8 mmol), and acid (6 mmol) in DCM (8 mL) was cooled to -15 °C for about 10 min, then DCC (6.72 mmol) was added. After stirring at -15~-10 °C for 1.5 h, DCM (15 mL) was added into the reaction solution and the solution was filtered through a short pad of silica gel. Then washed the pad of silica gel by additional 40 mL of DCM. The combined solution was concentrated on a rotary evaporator under vacuum at 10~15 °C and then purified by flash column chromatography on silica gel to give the alkyl diacyl peroxide.

## Characterization data for diacyl peroxides



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 2.42 (t, J = 7.5 Hz, 4H), 1.76 – 1.66 (m, 4H), 1.41 – 1.24 (m, 16H), 0.93 – 0.83 (m, 6H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*) δ 169.25, 31.54, 30.00, 28.87, 28.75, 24.81, 22.55, 14.00. The spectrum data matches previously reported values<sup>7</sup>.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 2.49 – 2.40 (m, 4H), 1.87 – 1.69 (m, 10H), 1.65 – 1.48 (m, 8H), 1.19 – 1.02 (m, 4H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*) δ 169.40, 39.47, 32.29, 30.94, 29.36, 25.10. The spectrum data matches previously reported values<sup>6</sup>.

<sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.36 – 7.26 (m, 4H), 7.26 – 7.17 (m, 6H), 3.02 (t, *J* = 7.8 Hz, 4H), 2.80 – 2.68 (m, 4H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  168.42, 139.33, 128.71, 128.29, 126.71, 31.70, 30.70. The spectrum data matches previously reported values<sup>6</sup>.

<sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  3.67 – 3.47 (m, 4H), 2.57 – 2.45 (m, 4H), 1.95 – 1.81 (m, 8H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  168.77, 44.23, 31.42, 29.20, 22.15. The spectrum data matches previously reported values<sup>6</sup>.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  3.68 – 3.30 (m, 4H), 2.73 – 2.50 (m, 4H), 2.36 – 2.11 (m, 4H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  168.19, 31.80, 28.39, 27.60. The spectrum data matches previously reported values<sup>6</sup>.

<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 2.60 (t, J = 7.0 Hz, 4H), 2.49 (t, J = 7.0 Hz, 4H), 2.16 (s, 6H), 1.97 (p, J = 6.9 Hz, 4H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*) δ 207.44, 168.76, 41.60, 29.95, 28.88, 18.64. The spectrum data matches previously reported values<sup>6</sup>.

$$\begin{array}{c} 0 \\ -0 \\ 0 \\ 0 \\ 2h \\ 0 \end{array} \begin{array}{c} 0 \\ 0 \\ 0 \\ 0 \end{array} \right)$$

<sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  3.72 (s, 6H), 2.81 – 2.69 (m, 8H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  171.80, 168.21, 52.25, 28.72, 25.34. The spectrum data matches previously reported values<sup>6</sup>.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 5.89 – 5.72 (m, 2H), 5.06 – 4.88 (m, 4H), 2.42 (t, J = 7.4 Hz, 4H), 2.07 – 2.00 (m, 4H), 1.75 – 1.67 (m, 4H), 1.40 – 1.29 (m, 16H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*) δ 169.30, 139.09, 114.33, 33.81, 30.07, 29.02, 28.94, 28.90, 28.88, 24.88. The spectrum data matches previously reported values<sup>6</sup>.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 2.19 (s, 4H), 2.00 (s, 6H), 1.74 - 1.63 (m, 24H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*) δ 166.72, 44.46, 42.11, 36.53, 32.99, 28.55. The spectrum data matches previously reported values<sup>7</sup>.

## Optimization of the reaction conditions

#### General procedure for Table S1: In a flame-dried Schlenk tube, Cat. (0.01

mmol, 5 mol%) and Py-Box ligand (±)-L16 (0.014 mmol, 7 mol%) were dissolved in THF (1 mL)

under a nitrogen atmosphere, and the mixture was stirred at room temperature for 30 minutes. Then 1,3-enyne **1a** (0.2 mmol, 1.0 equiv), peroxide **2a** (0.3 mmol, 1.5 equiv), PhB(OH)<sub>2</sub> (0.6 mmol, 3 equiv) and base (0.6 mmol, 3 equiv) were sequentially added. The reaction mixture was stirred at rt for 5 h. Upon completion of the reaction as monitored by TLC, the solvent was concentrated under vacuum. The crude residue was purified by flash column chromatography on silica gel to give the product.

1a	CI	+ LPO + + P 2a	hB(OH) <sub>2</sub> — <b>3a</b>	Cat, 5 mol % (±)-L16, 7 mol % DIPEA, 3 equiv Solvent, rt, N <sub>2</sub>		$\begin{array}{c} C_{11}H_{23} \\ Ph \\ C = C_{6}H_{13} \\ 4a \end{array}$
	entry	cat.	solvent	base	yield (%)	-
	1	CuTc	THF	DIPEA	69%	-
	2	Cu(OAc) <sub>2</sub>	THF	DIPEA	67%	
	3	Cu(OTf) <sub>2</sub>	THF	DIPEA	51%	
	4	Cu(TFA) <sub>2</sub>	THF	DIPEA	66%	
	5	Pd(OAc) <sub>2</sub>	THF	DIPEA	trace	
	6	NiCl <sub>2</sub>	THF	DIPEA	trace	
	7	$CoCl_2$	THF	DIPEA	trace	
	8	Cu(CH <sub>3</sub> CN) <sub>4</sub> BF <sub>4</sub>	THF	DIPEA	70%	
	9	Cu(CH <sub>3</sub> CN) <sub>4</sub> BF <sub>4</sub>	DCM	DIPEA	50%	
	10	Cu(CH <sub>3</sub> CN) <sub>4</sub> BF <sub>4</sub>	TBME	DIPEA	21%	
	11	Cu(CH <sub>3</sub> CN) <sub>4</sub> BF <sub>4</sub>	1,4-dioxan	e DIPEA	18%	
	12	Cu(CH <sub>3</sub> CN) <sub>4</sub> BF <sub>4</sub>	DMF	DIPEA	Trace	
	13	Cu(CH <sub>3</sub> CN) <sub>4</sub> BF <sub>4</sub>	DME	DIPEA	47%	
	14	Cu(CH <sub>3</sub> CN) <sub>4</sub> BF <sub>4</sub>	THF	KF	18%	
	15	Cu(CH <sub>3</sub> CN) <sub>4</sub> BF <sub>4</sub>	THF	DMAP	trace	
	16	Cu(CH <sub>3</sub> CN) <sub>4</sub> BF <sub>4</sub>	THF	$K_2CO_3$	trace	
	17	Cu(CH <sub>3</sub> CN) <sub>4</sub> BF <sub>4</sub>	THF	Et <sub>3</sub> N	65%	
	$18^b$	Cu(CH <sub>3</sub> CN) <sub>4</sub> BF <sub>4</sub>	THF	DIEPA	53%	
	$19^{c}$	$Cu(CH_3CN)_4BF_4$	THF	DIEPA	trace	

#### Table S1. Optimized reaction conditions<sup>a</sup>

<sup>*a*</sup> Reaction conditions: **1a** (0.2 mmol, 1 equiv), **2a** (0.3 mmol, 1.5 equiv), **3a** (0.6 mmol, 3 equiv), Cat. (5 mol%), ( $\pm$ )-L16 (7 mol%), base (0.6 mmol, 3 equiv), solvent (1 mL), rt. <sup>*b*</sup>2,4,6-Triphenyloroxin instead of PhB(OH)<sub>2</sub>. <sup>*c*</sup>PhBpin instead of PhB(OH)<sub>2</sub>. <sup>*d*</sup> Isolated yield.

# General procedure for 1,4-alkylarylation of 1,3-enynes

General procedure A: In a flame-dried Schlenk tube, Cu(CH<sub>3</sub>CN)<sub>4</sub>BF<sub>4</sub> (0.01 mmol,

5 mol%) and Py-Box ligand (±)-L16 (0.014 mmol, 7 mol%) were dissolved in THF (1 mL) under

a nitrogen atmosphere, and the mixture was stirred at room temperature for 30 minutes. Then 1,3-enyne (0.2 mmol, 1.0 equiv), peroxide (0.3 mmol, 1.5 equiv),  $PhB(OH)_2$  (0.6 mmol, 3 equiv) and DIPEA (0.6 mmol, 3 equiv) were sequentially added. The reaction mixture was stirred at rt for 5 h. Upon completion of the reaction as monitored by TLC, the solvent was concentrated under vacuum. The crude residue was purified by flash column chromatography on silica gel to give the product.

General procedure B: In a flame-dried Schlenk tube, Cu(CH<sub>3</sub>CN)<sub>4</sub>BF<sub>4</sub> (0.01 mmol,

5 mol%) and Py-Box ligand (±)-L16 (0.014 mmol, 7 mol%) were dissolved in THF (1 mL) under

a nitrogen atmosphere, and the mixture was stirred at room temperature for 30 minutes. Then 1,3-enyne (0.2 mmol, 1.0 equiv), peroxide (0.3 mmol, 1.5 equiv),  $PhB(OH)_2$  (0.6 mmol, 3 equiv) and DMF (1 mL) were sequentially added. The reaction mixture was stirred at rt for 5 h. Upon completion of the reaction as monitored by TLC, the solvent was concentrated under vacuum. The crude residue was purified by flash column chromatography on silica gel to give the product.

## Characterization data for products



Following the general procedure **A**, **4a** was obtained as a liquid (63 mg, 70% yield). <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.47 (d, *J* = 8.2 Hz, 2H), 7.38 – 7.30 (m, 4H), 7.28 – 7.21 (m, 1H), 7.15 (d, *J* = 8.0 Hz, 2H), 3.63 (t, *J* = 6.4 Hz, 2H), 2.74 (t, *J* = 7.5 Hz, 2H), 2.56 (t, *J* = 7.6 Hz, 2H), 2.36 (s, 3H), 2.15 – 2.01 (m, 2H), 1.66 – 1.54 (m, 2H), 1.40 (p, *J* = 7.6, 7.1 Hz, 2H), 1.37 – 1.22 (m, 16H), 0.91 (t, *J* = 6.9 Hz, 3H). <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  204.80, 136.79, 133.71, 129.38, 128.63, 126.95, 125.95, 110.04, 107.78, 45.06, 32.07, 31.10, 30.58, 29.88, 29.82, 29.78, 29.68, 29.51, 28.30, 27.65, 22.85, 21.22, 14.28. HRMS (DART) calcd for  $[C_{31}H_{44}Cl]^+([M+H]^+)$ : 451.3126, found: 451.3123.



Following the general procedure A, **4b** was obtained as a liquid (58 mg, 63% yield). <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.46 (d, J = 8.1 Hz, 2H), 7.36 (d, J = 8.1 Hz, 2H), 7.32 (t, J = 7.6 Hz, 2H), 7.21 (d, J = 7.8 Hz, 1H), 7.14 (d, J = 8.3 Hz, 2H), 2.56 (q, J = 7.2 Hz, 4H), 2.35 (s, 3H), 1.67 – 1.57 (m, 4H), 1.45 – 1.39 (m, 4H), 1.32 – 1.25 (m, 20H), 0.92 – 0.87 (m, 6H). <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  205.15, 137.36, 136.42, 134.18, 129.25, 128.49, 126.62, 126.03, 125.95, 109.00, 32.07, 31.93, 30.53, 29.88, 29.82, 29.79, 29.71, 29.58, 29.51, 28.28, 28.26, 22.84, 21.20, 14.27, 14.22. HRMS (DART) calcd for [C<sub>34</sub>H<sub>51</sub>]<sup>+</sup> ([M+H]<sup>+</sup>): 459.3985, found: 459.3981.



Following the general procedure A, **4c** was obtained as a liquid (43 mg, 46% yield). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.42 (d, *J* = 7.7 Hz, 2H), 7.40 – 7.34 (m, 2H), 7.31 (t, *J* = 7.6 Hz, 2H), 7.20 (t, *J* = 7.3 Hz, 1H), 6.99 (t, *J* = 8.7 Hz, 2H), 2.72 – 2.38 (m, 4H), 1.63 – 1.51 (m, 4H), 1.42 – 1.34 (m, 4H), 1.30 – 1.21 (m, 20H), 0.90 – 0.83 (m, 6H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  204.98 (d, *J* = 1.9 Hz), 161.78 (d, *J* = 245.7 Hz), 133.04 (d, *J* = 3.2 Hz), 128.45, 127.38 (d, *J* = 7.8 Hz), 126.71, 125.91, 115.23 (d, *J* = 21.3 Hz), 109.23, 108.26, 31.96, 31.80, 30.59, 30.40, 29.73, 29.71, 29.70, 29.67, 29.58, 29.44, 29.40, 28.13, 28.08, 22.73, 14.16, 14.11. HRMS (DART) calcd for  $[C_{33}H_{48}F]^+$  ([M+H]<sup>+</sup>): 463.3735, found: 463.3731.



Following the general procedure A, **4d** was obtained as a liquid (68 mg, 65% yield). <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.45 – 7.41 (m, 4H), 7.34 – 7.29 (m, 4H), 7.24 – 7.20 (m, 1H), 2.56 (t, J = 7.6 Hz, 2H), 2.52 (t, J = 7.6 Hz, 2H), 1.64 – 1.53 (m, 4H), 1.43 – 1.36 (m, 4H), 1.32 – 1.24 (m, 20H), 0.92 – 0.87 (m, 6H). <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  205.39, 136.80, 136.23, 131.54, 128.57, 127.62, 126.90, 126.02, 120.46, 109.63, 108.45, 32.03, 31.87, 30.43, 30.38, 29.78, 29.75, 29.64, 29.51, 29.48, 28.19, 28.14, 22.80, 14.23, 14.18. HRMS (DART) calcd for  $[C_{33}H_{48}Br]^+$  ( $[M+H]^+$ ): 523.2934, found: 523.2931.



Following the general procedure A, **4e** was obtained as a liquid (76 mg, 73% yield). <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.64 (d, *J* = 7.6 Hz, 2H), 7.60 (d, *J* = 8.1 Hz, 2H), 7.56 (d, *J* = 8.5 Hz, 2H), 7.51 (d, *J* = 7.8 Hz, 2H), 7.47 (t, *J* = 7.6 Hz, 2H), 7.37 (t, *J* = 7.6 Hz, 3H), 7.26 (t, *J* = 7.5 Hz, 1H), 2.74 – 2.37 (m, 4H), 1.72 – 1.62 (m, 4H), 1.50 – 1.42 (m, 4H), 1.36 – 1.28 (m, 20H), 0.95 – 0.90 (m, 6H). <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  205.71, 141.03, 139.56, 137.16, 136.22, 128.91, 128.58, 127.28, 127.15, 127.08, 126.81, 126.47, 126.10, 109.36, 108.95, 32.10, 31.96, 30.57,

30.53, 29.93, 29.87, 29.83, 29.75, 29.63, 29.55, 28.34, 28.30, 22.88, 14.31, 14.26. HRMS (DART) calcd for  $[C_{39}H_{53}]^+$  ([M+H]<sup>+</sup>): 521.4142, found: 521.4136.



Following the general procedure A, **4f** was obtained as a liquid (53 mg, 55% yield). <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.43 – 7.39 (m, 2H), 7.39 – 7.36 (m, 1H), 7.34 – 7.28 (m, 3H), 7.24 – 7.18 (m, 2H), 7.18 – 7.14 (m, 1H), 2.54 (t, *J* = 8.4 Hz, 2H), 2.50 (t, *J* = 7.8 Hz, 2H), 1.60 – 1.52 (m, 4H), 1.42 – 1.36 (m, 4H), 1.27 – 1.22 (m, 20H), 0.88 – 0.85 (m, 6H). <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  205.51, 139.37, 136.72, 134.51, 129.63, 128.57, 126.93, 126.66, 126.06, 125.95, 124.19, 109.73, 108.35, 32.03, 31.87, 30.44, 30.38, 29.79, 29.76, 29.74, 29.63, 29.51, 29.47, 28.16, 28.10, 22.80, 14.24, 14.18. HRMS (DART) calcd for [C<sub>33</sub>H<sub>48</sub>Cl]<sup>+</sup> ([M+H]<sup>+</sup>): 479.3439, found: 479.3435.



Following the general procedure A, **4g** was obtained as a liquid (65 mg, 71% yield). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.46 – 7.41 (m, 2H), 7.30 (t, *J* = 7.6 Hz, 2H), 7.26 – 7.16 (m, 4H), 7.01 (d, *J* = 7.3 Hz, 1H), 2.65 – 2.42 (m, 4H), 2.33 (s, 3H), 1.66 – 1.53 (m, 4H), 1.45 – 1.34 (m, 4H), 1.30 – 1.19 (m, 20H), 0.91 – 0.83 (m, 6H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  205.30, 137.96, 137.20, 137.05, 128.42, 128.34, 127.47, 126.63, 126.58, 125.97, 123.11, 109.09, 108.89, 32.00, 31.86, 30.48, 30.42, 29.80, 29.76, 29.72, 29.63, 29.51, 29.44, 28.19, 28.17, 22.78, 21.65, 14.21, 14.16. HRMS (DART) calcd for [C<sub>34</sub>H<sub>51</sub>]<sup>+</sup> ([M+H]<sup>+</sup>): 459.3985, found: 459.3981.



Following the general procedure A, **4h** was obtained as a liquid (50 mg, 54% yield). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.47 – 7.39 (m, 2H), 7.31 (t, *J* = 7.6 Hz, 2H), 7.27 – 7.24 (m, 1H), 7.21 – 7.11 (m, 4H), 2.50 – 2.38 (m, 4H), 2.33 (s, 3H), 1.61 – 1.47 (m, 4H), 1.42 – 1.33 (m, 4H), 1.31 – 1.23 (m, 20H), 0.91 – 0.86 (m, 6H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  202.77, 138.50, 137.67,

135.78, 130.39, 128.35, 128.25, 126.73, 126.34, 126.15, 125.68, 108.21, 106.00, 34.50, 31.97, 31.88, 30.55, 29.73, 29.69, 29.67, 29.59, 29.41, 29.39, 28.30, 28.15, 22.75, 20.71, 14.18, 14.16. HRMS (DART) calcd for  $[C_{34}H_{51}]^+$  ([M+H]<sup>+</sup>): 459.3985, found: 459.3982.



Following the general procedure A, **4i** was obtained as a liquid (53 mg, 54% yield). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.83 – 7.76 (m, 3H), 7.71 (d, *J* = 8.7 Hz, 1H), 7.59 (dd, *J* = 8.6, 1.8 Hz, 1H), 7.48 – 7.39 (m, 4H), 7.32 (t, *J* = 7.6 Hz, 2H), 7.21 (t, *J* = 7.3 Hz, 1H), 2.67 (t, *J* = 7.6 Hz, 2H), 2.58 (t, *J* = 7.6 Hz, 2H), 1.71 – 1.57 (m, 4H), 1.46 – 1.38 (m, 4H), 1.32 – 1.23 (m, 20H), 0.89 – 0.82 (m, 6H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  206.04, 137.05, 134.51, 133.73, 132.46, 128.46, 128.01, 127.78, 127.56, 126.69, 126.05, 125.99, 125.55, 125.43, 123.39, 109.38, 31.96, 31.81, 30.45, 30.35, 29.82, 29.73, 29.69, 29.62, 29.48, 29.41, 28.17, 22.74, 22.72, 14.17, 14.11. HRMS (DART) calcd for [C<sub>37</sub>H<sub>51</sub>]<sup>+</sup> ([M+H]<sup>+</sup>): 495.3985, found: 495.3980.



Following the general procedure A, **4j** was obtained as a liquid (43 mg, 45% yield). <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.49 – 7.44 (m, 2H), 7.35 – 7.31 (m, 2H), 7.31 – 7.27 (m, 4H), 7.24 – 7.19 (m, 4H), 7.13 (d, *J* = 8.4 Hz, 2H), 2.96 – 2.86 (m, 4H), 2.56 – 2.45 (m, 2H), 2.35 (s, 3H), 1.55 – 1.50 (m, 2H), 1.41 – 1.24 (m, 18H), 0.92 – 0.89 (m, 3H). <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  205.11, 142.19, 137.05, 136.56, 133.93, 129.28, 128.58, 128.46, 126.79, 126.01, 125.98, 109.68, 108.41, 34.42, 32.33, 32.05, 30.54, 29.87, 29.82, 29.77, 29.68, 29.50, 28.22, 22.83, 21.21, 14.27. HRMS (DART) calcd for [C<sub>36</sub>H<sub>47</sub>]<sup>+</sup> ([M+H]<sup>+</sup>): 479.3672, found: 479.3669.



Following the general procedure **A**, **4k** was obtained as a liquid (61 mg, 67% yield). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.46 – 7.40 (m, 4H), 7.30 (t, *J* = 7.6 Hz, 4H), 7.22 – 7.15 (m, 2H),

2.54 (t, J = 7.6 Hz, 2H), 2.44 (d, J = 6.9 Hz, 2H), 1.90 – 1.74 (m, 3H), 1.71 – 1.56 (m, 6H), 1.31 – 1.19 (m, 22H), 0.89 (t, J = 6.4 Hz, 3H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  205.96, 137.22, 137.10, 128.41, 126.58, 126.17, 126.07, 107.97, 107.18, 38.54, 36.41, 33.73, 33.68, 31.98, 30.63, 29.80, 29.74, 29.72, 29.70, 29.59, 29.42, 28.30, 26.57, 26.32, 26.27, 22.76, 14.20. HRMS (DART) calcd for [C<sub>34</sub>H<sub>49</sub>]<sup>+</sup> ([M+H]<sup>+</sup>): 457.3829, found: 457.3826.



Following the general procedure A, **41** was obtained as a liquid (64 mg, 69% yield). <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.49 – 7.43 (m, 4H), 7.36 – 7.30 (m, 4H), 7.23 – 7.18 (m, 2H), 3.66 (s, 3H), 2.61 (t, *J* = 7.7 Hz, 2H), 2.57 (t, *J* = 7.6 Hz, 2H), 2.43 (t, *J* = 7.4 Hz, 2H), 1.99 – 1.89 (m, 2H), 1.64 – 1.54 (m, 2H), 1.42 – 1.36 (m, 2H), 1.31 – 1.24 (m, 16H), 0.90 (t, *J* = 7.0 Hz, 3H). <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  205.21, 174.04, 136.90, 136.74, 128.60, 128.58, 126.90, 126.06, 125.98, 109.85, 108.37, 51.61, 33.91, 32.05, 30.53, 29.87, 29.80, 29.77, 29.67, 29.49, 28.28, 23.48, 22.83, 14.26. HRMS (DART) calcd for [C<sub>32</sub>H<sub>45</sub>O<sub>2</sub>]<sup>+</sup> ([M+H]<sup>+</sup>): 461.3414, found: 461.3411.



Following the general procedure **A**, **4m** was obtained as a liquid (26 mg, 32% yield). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.44 (d, J = 7.7 Hz, 4H), 7.34 – 7.26 (m, 4H), 7.23 – 7.17 (m, 2H), 3.87 (t, J = 6.5 Hz, 2H), 2.94 – 2.74 (m, 2H), 2.55 (t, J = 7.6 Hz, 2H), 2.31 (t, J = 7.5 Hz, 1H), 1.64 – 1.53 (m, 2H), 1.43 – 1.34 (m, 2H), 1.32 – 1.24 (m, 16H), 0.88 (t, J = 6.8 Hz, 3H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  204.82, 136.34, 136.28, 128.67, 128.60, 127.10, 127.06, 125.90, 125.84, 110.19, 105.80, 61.39, 33.47, 31.95, 30.41, 29.75, 29.69, 29.66, 29.63, 29.55, 29.39, 28.18, 22.73, 14.17. HRMS (DART) calcd for [C<sub>29</sub>H<sub>41</sub>O]<sup>+</sup> ([M+H]<sup>+</sup>): 405.3152, found: 405.3149.



Following the general procedure **A**, **4n** was obtained as a liquid (33 mg, 42% yield). <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.47 – 7.42 (m, 4H), 7.36 – 7.31 (m, 4H), 7.27 – 7.22 (m, 2H), 4.72 – 4.61 (m, 2H), 2.64 – 2.55 (m, 2H), 1.62 – 1.55 (m, 2H), 1.42 – 1.36 (m, 2H), 1.32 – 1.21 (m, 17H), 0.88 (t, J = 7.1 Hz, 3H). <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  203.57, 136.15, 134.54, 128.82, 128.70, 127.43, 127.40, 126.23, 126.12, 112.40, 110.07, 61.86, 32.03, 30.53, 29.77, 29.74, 29.72, 29.62, 29.47, 28.20, 22.81, 14.25. HRMS (EI+) calcd for [C<sub>28</sub>H<sub>38</sub>O]<sup>+</sup> ([M]<sup>+</sup>): 390.2923, found: 390.2931.



Following the general procedure A, **40** was obtained as a liquid (38 mg, 49% yield). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.34 – 7.27 (m, 4H), 7.25 – 7.20 (m, 3H), 7.13 (d, *J* = 7.9 Hz, 2H), 2.35 (s, 3H), 2.12 (t, *J* = 7.5 Hz, 2H), 1.85 (s, 3H), 1.55 – 1.47 (m, 2H), 1.29 – 1.18 (m, 18H), 0.88 (t, *J* = 6.8 Hz, 3H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  203.08, 138.38, 136.39, 135.23, 128.96, 128.37, 128.33, 128.19, 126.62, 108.57, 102.87, 34.42, 31.98, 29.75, 29.72, 29.70, 29.68, 29.58, 29.48, 29.43, 27.66, 22.76, 21.19, 19.08, 14.19. HRMS (DART) calcd for [C<sub>29</sub>H<sub>41</sub>]<sup>+</sup> ([M+H]<sup>+</sup>): 389.3203, found: 389.3200.



Following the general procedure A, **4p** was obtained as a liquid (13 mg, 27% yield). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.40 – 7.35 (m, 2H), 7.32 – 7.27 (m, 2H), 7.19 – 7.15 (m, 1H), 3.47 (t, J = 6.8 Hz, 2H), 2.41 (q, J = 7.3 Hz, 2H), 2.14 – 2.04 (m, 2H), 1.80 (s, 3H), 1.77 – 1.71 (m, 2H), 1.52 – 1.43 (m, 4H), 1.10 (t, J = 7.3 Hz, 3H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  200.98, 138.38, 128.23, 126.11, 125.78, 106.70, 103.35, 45.15, 34.17, 32.54, 26.97, 26.74, 23.18, 19.12, 12.77. HRMS (EI+) calcd for [C<sub>17</sub>H<sub>23</sub>Cl]<sup>+</sup> ([M]<sup>+</sup>): 262.1488, found: 262.1485.



Following the general procedure A, **4q** was obtained as a liquid (27 mg, 32% yield). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.43 – 7.26 (m, 14H), 3.37 – 3.22 (m, 2H), 2.84 – 2.74 (m, 1H), 1.71 – 1.54 (m, 4H), 1.52 – 1.38 (m, 2H), 1.36 – 1.22 (m, 3H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  206.11, 136.68, 136.49, 134.78, 132.76, 128.81, 128.54, 128.53, 128.25, 128.15, 127.77, 127.49, 127.45, 114.30, 114.14, 44.93, 35.59, 34.45, 32.81, 25.00, 20.61. HRMS (DART) calcd for  $[C_{27}H_{27}Cl_2]^+$  ([M+H]<sup>+</sup>): 421.1484, found: 421.1481.



Following the general procedure A, **4r** was obtained as a liquid (16 mg, 20% yield). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.48 – 7.38 (m, 4H), 7.35 – 7.26 (m, 4H), 7.24 – 7.16 (m, 2H), 3.51 (t, *J* = 6.7 Hz, 1.5 H), 3.35 (t, *J* = 6.7, Hz, 0.5H), 2.88 – 2.74 (m, 1H), 2.54 (t, *J* = 7.6 Hz, 2H), 1.77 (m, 1.5H), 1.61 – 1.54 (m, 3.5H), 1.43 – 1.36 (m, 3H), 1.29 – 1.24 (m, 6H), 1.19 (dd, *J* = 6.8, 1.8 Hz, 3H), 0.87 – 0.84 (m, 3H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  204.53, 136.98, 136.83, 128.51, 128.45, 126.70, 126.66, 126.40, 125.77, 45.11, 35.79, 34.13, 32.89, 31.80, 30.50, 29.55, 28.41, 24.96, 22.72, 20.25, 14.12. HRMS (DART) calcd for [C<sub>27</sub>H<sub>36</sub>Cl]<sup>+</sup> ([M+H]<sup>+</sup>): 395.2500, found: 395.2497.



Following the general procedure A, **4s** was obtained as a liquid (34 mg, 46% yield). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.53 – 7.46 (m, 2H), 7.44 – 7.37 (m, 4H), 7.37 – 7.30 (m, 6H), 7.30 – 7.20 (m, 3H), 3.36 (t, J = 6.8 Hz, 2H), 2.64 (t, J = 7.4 Hz, 2H), 1.82 – 1.60 (m, 4H), 1.52 – 1.41 (m, 2H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  206.99, 136.85, 136.11, 128.65, 128.51, 128.32, 127.38, 127.17, 126.05, 113.12, 108.77, 45.05, 32.57, 30.18, 27.23, 26.87. HRMS (DART) calcd for [C<sub>26</sub>H<sub>26</sub>Cl]<sup>+</sup> ([M+H]<sup>+</sup>): 373.1718, found: 373.1714.



Following the general procedure **A**, **4t** was obtained as a liquid (19 mg, 32% yield). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.48 – 7.41 (m, 2H), 7.36 – 7.29 (m, 6H), 7.24 – 7.18 (m, 2H), 6.54 (t, *J* = 3.2 Hz, 1H), 3.48 (t, *J* = 6.7 Hz, 2H), 2.65 – 2.52 (m, 2H), 1.84 – 1.72 (m, 2H), 1.69 – 1.51 (m, 4H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  206.42, 136.06, 134.54, 128.78, 128.57, 127.15, 127.12, 126.79, 126.10, 109.65, 98.15, 45.11, 32.49, 29.93, 27.17, 26.81. HRMS (DART) calcd for  $[C_{20}H_{22}Cl]^+$  ([M+H]<sup>+</sup>): 297.1405, found: 297.1403.



Following the general procedure A, **5a** was obtained as a liquid (50 mg, 63% yield). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.48 – 7.40 (m, 2H), 7.37 – 7.25 (m, 4H), 7.20 (d, *J* = 7.2 Hz, 1H), 7.12 (d, *J* = 8.0 Hz, 2H), 3.60 (t, *J* = 6.4 Hz, 2H), 2.71 (t, *J* = 7.5 Hz, 2H), 2.52 (d, *J* = 8.0 Hz, 2H), 2.33 (s, 3H), 2.11 – 1.97 (m, 2H), 1.63 – 1.49 (m, 2H), 1.43 – 1.33 (m, 2H), 1.32 – 1.19 (m, 8H), 0.86 (t, *J* = 6.8 Hz, 3H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  204.71, 136.70, 133.62, 129.29, 128.54, 126.86, 125.86, 109.94, 107.69, 44.99, 31.89, 31.02, 30.50, 29.79, 29.55, 29.39, 28.22, 27.56, 22.72, 21.14, 14.17. HRMS (DART) calcd for [C<sub>27</sub>H<sub>36</sub>Cl]<sup>+</sup> ([M+H]<sup>+</sup>): 395.2500, found: 395.2498.



Following the general procedure **A**, **5b** was obtained as a liquid (48 mg, 61% yield). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.50 – 7.40 (m, 2H), 7.36 – 7.26 (m, 4H), 7.21 (d, *J* = 7.5 Hz, 1H), 7.12 (d, *J* = 7.9 Hz, 2H), 3.60 (t, *J* = 6.5 Hz, 2H), 2.71 (t, *J* = 7.5 Hz, 2H), 2.53 (t, *J* = 7.6 Hz, 2H), 2.32 (s, 3H), 2.10 – 1.98 (m, 2H), 1.79 – 1.63 (m, 3H), 1.62 – 1.43 (m, 6H), 1.41 – 1.35 (m, 2H), 1.09 – 0.94 (m, 2H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  204.74, 136.70, 133.63, 129.30, 128.55, 126.87, 125.88, 125.87, 109.92, 107.71, 44.99, 39.98, 36.27, 32.75, 32.71, 31.03, 30.72, 27.58, 27.32, 25.22, 21.15. HRMS (DART) calcd for [C<sub>27</sub>H<sub>34</sub>Cl]<sup>+</sup> ([M+H]<sup>+</sup>): 393.2344, found: 393.2341.



Following the general procedure A, **5c** was obtained as a liquid (41 mg, 51% yield). <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.49 – 7.44 (m, 2H), 7.36 – 7.29 (m, 4H), 7.29 – 7.25 (m, 2H), 7.25 – 7.21 (m, 1H), 7.18 (t, J = 6.8 Hz, 1H), 7.16 – 7.10 (m, 4H), 3.60 (t, J = 6.4 Hz, 2H), 2.79 – 2.67 (m, 4H), 2.60 (t, J = 7.6 Hz, 2H), 2.34 (s, 3H), 2.11 – 2.01 (m, 2H), 1.98 – 1.87 (m, 2H). <sup>13</sup>C NMR

128.43, 127.06, 125.96, 125.90, 109.70, 108.15, 45.02, 35.92, 31.08, 30.04, 29.88, 27.66, 21.20. HRMS (DART) calcd for  $[C_{28}H_{30}Cl]^+$  ([M+H]<sup>+</sup>): 401.2031, found: 401.2027.



Following the general procedure A, **5d** was obtained as a liquid (56 mg, 72% yield). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.48 – 7.38 (m, 2H), 7.37 – 7.26 (m, 4H), 7.21 (t, *J* = 7.5 Hz, 1H), 7.13 (d, *J* = 8.0 Hz, 2H), 3.61 (t, *J* = 6.4 Hz, 2H), 3.46 (t, *J* = 6.7 Hz, 2H), 2.70 (t, *J* = 7.6 Hz, 2H), 2.56 (t, *J* = 7.2 Hz, 2H), 2.33 (s, 3H), 2.13 – 1.98 (m, 2H), 1.80 – 1.70 (m, 2H), 1.65 – 1.47 (m, 5H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  204.62, 136.86, 136.55, 133.39, 129.35, 128.60, 126.98, 125.85, 125.81, 109.56, 107.99, 45.13, 44.96, 32.55, 31.01, 30.30, 27.57, 27.42, 27.00, 21.15. HRMS (DART) calcd for [C<sub>24</sub>H<sub>29</sub>Cl<sub>2</sub>]<sup>+</sup> ([M+H]<sup>+</sup>): 387.1641, found: 387.1638.



Following the general procedure **A**, **5e** was obtained as a liquid (51 mg, 54% yield). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.47 – 7.39 (m, 2H), 7.37 – 7.27 (m, 4H), 7.22 (t, *J* = 7.3 Hz, 1H), 7.13 (d, *J* = 8.0 Hz, 2H), 3.61 (t, *J* = 6.4 Hz, 2H), 3.37 (t, *J* = 6.8 Hz, 2H), 2.72 (t, *J* = 7.5 Hz, 2H), 2.57 (t, *J* = 7.6 Hz, 2H), 2.33 (s, 3H), 2.11 – 2.00 (m, 2H), 1.99 – 1.90 (m, 2H), 1.79 – 1.65 (m, 2H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  204.60, 136.94, 136.44, 133.24, 129.39, 128.64, 127.06, 125.88, 125.81, 109.25, 108.22, 44.94, 33.57, 32.64, 31.02, 29.57, 27.63, 26.65, 21.16. HRMS (EI+) calcd for [C<sub>23</sub>H<sub>26</sub>ClBr]<sup>+</sup> ([M]<sup>+</sup>): 416.0906, found: 416.0904.



Following the general procedure A, **5f** was obtained as a liquid (40 mg, 52% yield). <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.45 – 7.42 (m, 2H), 7.34 – 7.29 (m, 4H), 7.24 – 7.20 (m, 1H), 7.13 (d, *J* = 8.1 Hz, 2H), 3.61 (t, *J* = 6.4 Hz, 2H), 2.72 (t, *J* = 7.5 Hz, 2H), 2.56 (t, *J* = 7.5 Hz, 2H), 2.41 (t, *J* = 7.5 Hz, 2H), 2.34 (s, 3H), 2.08 – 2.01 (m, 5H), 1.71 – 1.65 (m, 2H), 1.62 – 1.51 (m, 2H). <sup>13</sup>C NMR

(150 MHz, Chloroform-*d*)  $\delta$  209.10, 204.67, 136.93, 136.61, 133.41, 129.42, 128.68, 127.06, 125.91, 125.88, 109.58, 108.06, 45.02, 43.69, 31.06, 30.36, 29.92, 27.69, 27.63, 23.89, 21.20. HRMS (DART) calcd for  $[C_{25}H_{30}OC1]^+$  ([M+H]<sup>+</sup>): 381.1980, found: 381.1977.



CI/

Following the general procedure **A**, **5g** was obtained as a liquid (46 mg, 60% yield). <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.46 – 7.43 (m, 2H), 7.35 – 7.31 (m, 4H), 7.25 – 7.21 (m, 1H), 7.14 (d, *J* = 8.2 Hz, 2H), 3.65 (s, 3H), 3.62 (t, *J* = 6.5 Hz, 2H), 2.73 (t, *J* = 7.5 Hz, 2H), 2.60 (t, *J* = 7.7 Hz, 2H), 2.42 (t, *J* = 7.3 Hz, 2H), 2.34 (s, 3H), 2.11 – 2.02 (m, 2H), 1.96 – 1.87 (m, 2H). <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  204.62, 173.99, 137.01, 136.47, 133.25, 129.44, 128.69, 127.12, 125.96, 125.88, 109.21, 108.44, 51.67, 45.01, 33.88, 31.06, 29.94, 27.68, 23.49, 21.21. HRMS (DART) calcd for [C<sub>24</sub>H<sub>28</sub>O<sub>2</sub>Cl]<sup>+</sup> ([M+H]<sup>+</sup>): 383.1772, found: 383.1769.



Following the general procedure **A**, **5h** was obtained as a liquid (53 mg, 63% yield). <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.48 – 7.44 (m, 2H), 7.35 – 7.31 (m, 4H), 7.25 – 7.21 (m, 1H), 7.17 – 7.11 (m, 2H), 5.86 – 5.77 (m, 1H), 5.04 – 4.97 (m, 1H), 4.97 – 4.90 (m, 1H), 3.63 (t, *J* = 6.5 Hz, 2H), 2.73 (t, *J* = 7.5 Hz, 2H), 2.55 (d, *J* = 7.8 Hz, 2H), 2.35 (s, 3H), 2.14 – 1.99 (m, 4H), 1.65 – 1.50 (m, 2H), 1.42 – 1.34 (m, 4H), 1.31 – 1.24 (m, 6H). <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  204.79, 139.39, 139.37, 136.80, 136.77, 133.68, 129.38, 128.63, 126.95, 125.94, 114.24, 110.01, 107.79, 45.06, 33.92, 31.09, 30.56, 29.83, 29.60, 29.21, 29.03, 28.27, 27.63, 21.21. HRMS (DART) calcd for [C<sub>29</sub>H<sub>38</sub>Cl]<sup>+</sup> ([M+H]<sup>+</sup>): 421.2657, found: 421.2654.



Following the general procedure **A**, **5i** was obtained as a liquid (44 mg, 49% yield). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.49 – 7.40 (m, 2H), 7.36 – 7.26 (m, 4H), 7.23 – 7.18 (m, 1H), 7.12 (d, *J* =

7.9 Hz, 2H), 3.61 (t, J = 6.4 Hz, 2H), 2.72 (t, J = 7.2 Hz, 2H), 2.55 – 2.47 (m, 2H), 2.33 (s, 3H), 2.12 – 2.00 (m, 2H), 1.96 (s, 3H), 1.76 – 1.60 (m, 6H), 1.58 -1.48 (m, 6H), 1.39 – 1.33 (m, 2H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  204.59, 136.72, 136.68, 133.58, 129.28, 128.55, 126.83, 125.88, 125.85, 110.41, 107.54, 44.98, 42.89, 42.42, 37.27, 32.43, 30.96, 28.77, 27.46, 23.77, 21.14. HRMS (DART) calcd for  $[C_{31}H_{38}Cl]^+$  ([M+H]<sup>+</sup>): 445.2657, found: 445.2654.



Following the general procedure A, **5j** was obtained as a liquid (45 mg, 60% yield). <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.47 – 7.41 (m, 2H), 7.37 – 7.28 (m, 4H), 7.24 – 7.20 (m, 1H), 7.18 – 7.09 (m, 2H), 3.61 (t, J = 6.5 Hz, 2H), 2.80 – 2.66 (m, 2H), 2.52 – 2.39 (m, 2H), 2.35 (s, 3H), 2.15 – 2.02 (m, 2H), 1.88 – 1.78 (m, 2H), 1.72 – 1.65 (m, 2H), 1.64 – 1.53 (m, 2H), 1.23 – 1.10 (m, 3H), 1.03 – 0.89 (m, 2H). <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  205.50, 136.80, 136.75, 133.83, 129.36, 128.60, 126.89, 126.16, 126.05, 108.12, 106.59, 45.05, 38.73, 36.49, 33.78, 33.73, 31.20, 27.86, 26.60, 26.35, 26.33, 21.21. HRMS (DART) calcd for  $[C_{26}H_{32}Cl]^+$  ( $[M+H]^+$ ): 379.2187, found: 379.2184.



Following the general procedure A, **5k** was obtained as a liquid (14 mg, 19% yield). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.49 – 7.41 (m, 2H), 7.37 – 7.29 (m, 4H), 7.24 – 7.19 (m, 1H), 7.13 (d, *J* = 8.0 Hz, 2H), 5.69 – 5.56 (m, 2H), 3.61 (t, *J* = 6.5 Hz, 2H), 2.72 (t, *J* = 7.6 Hz, 2H), 2.67 – 2.60 (m, 2H), 2.59 – 2.41 (m, 3H), 2.33 (s, 3H), 2.17 – 1.99 (m, 4H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  205.29, 136.80, 136.46, 133.69, 129.90, 129.70, 129.30, 128.54, 126.89, 125.88, 125.81, 109.15, 107.65, 45.01, 39.59, 39.56, 37.66, 35.61, 31.02, 27.68, 21.12. HRMS (DART) calcd for [C<sub>25</sub>H<sub>28</sub>Cl]<sup>+</sup> ([M+H]<sup>+</sup>): 363.1874, found: 363.1871.



Following the general procedure **B**, **6a** was obtained as a liquid (63 mg, 60% yield). <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.64 – 7.61 (m, 2H), 7.60 – 7.57 (m, 2H), 7.57 – 7.52 (m, 2H), 7.47 – 7.43 (m, 2H), 7.39 – 7.33 (m, 3H), 7.17 (d, *J* = 7.9 Hz, 2H), 3.66 (t, *J* = 6.4 Hz, 2H), 2.78 (t, *J* = 7.5 Hz, 2H), 2.62 – 2.55 (m, 2H), 2.37 (s, 3H), 2.17 – 2.07 (m, 2H), 1.68 – 1.58 (m, 2H), 1.46 – 1.40 (m, 2H), 1.34 – 1.26 (m, 16H), 0.91 (t, *J* = 7.1 Hz, 3H). <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  205.09, 140.90, 139.78, 136.86, 135.78, 133.68, 129.44, 128.91, 127.35, 127.06, 126.36, 126.01, 110.20, 107.55, 45.05, 32.06, 31.15, 30.63, 29.90, 29.84, 29.80, 29.70, 29.52, 28.34, 27.69, 22.85, 21.23, 14.27. HRMS (DART) calcd for [C<sub>37</sub>H<sub>48</sub>Cl]<sup>+</sup> ([M+H]<sup>+</sup>): 527.3439, found: 727.3434.



Following the general procedure A, **6b** was obtained as a liquid (64 mg, 69% yield). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.31 (d, J = 8.0 Hz, 2H), 7.27 – 7.17 (m, 3H), 7.12 (d, J = 7.9 Hz, 2H), 7.03 (d, J = 7.2 Hz, 1H), 3.60 (t, J = 6.5 Hz, 2H), 2.70 (t, J = 7.5 Hz, 2H), 2.59 – 2.47 (m, 2H), 2.33 (s, 6H), 2.09 – 1.98 (m, 2H), 1.62 – 1.50 (m, 2H), 1.43 – 1.34 (m, 2H), 1.30 – 1.21 (m, 16H), 0.88 (t, J = 6.8 Hz, 3H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  204.70, 138.09, 136.64, 136.63, 133.71, 129.28, 128.45, 127.69, 126.52, 125.88, 123.02, 109.74, 107.69, 45.01, 31.99, 31.04, 30.49, 29.81, 29.75, 29.71, 29.62, 29.44, 28.21, 27.64, 22.77, 21.65, 21.14, 14.21. HRMS (DART) calcd for [C<sub>32</sub>H<sub>46</sub>Cl]<sup>+</sup> ([M+H]<sup>+</sup>): 465.3283, found: 465.3279.



Following the general procedure **A**, **6c** was obtained as a liquid (53 mg, 55% yield). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.30 (d, *J* = 8.1 Hz, 2H), 7.15 (d, *J* = 7.6 Hz, 1H), 7.12 (d, *J* = 8.0 Hz, 2H), 7.04 – 6.92 (m, 2H), 3.58 (t, *J* = 6.5 Hz, 2H), 2.56 (d, *J* = 7.6 Hz, 2H), 2.43 (d, *J* = 7.6 Hz, 2H), 2.33 (s, 3H), 2.29 (s, 6H), 2.04 – 1.94 (m, 2H), 1.59 – 1.49 (m, 2H), 1.38 – 1.23 (m, 18H), 0.88 (t, *J* = 6.5 Hz, 2H), 2.56 (d, *J* = 7.6 Hz, 2H), 1.59 – 1.49 (m, 2H), 1.38 – 1.23 (m, 18H), 0.88 (t, *J* = 6.5 Hz, 2H), 2.56 (d, *J* = 7.6 Hz, 2H), 1.59 – 1.49 (m, 2H), 1.59 – 1.49 (m, 2H), 1.58 – 1.23 (m, 18H), 0.88 (t, *J* = 6.5 Hz, 2H), 1.59 – 1.49 (m, 2H), 1.59 – 1.49 (m, 2H), 1.58 – 1.23 (m, 18H), 0.88 (t, J) = 0.5 Hz, 2H), 0.58 (t, J) = 0.5 Hz, 0.5 Hz,

J = 6.8 Hz, 3H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  202.53, 136.57, 136.27, 135.64, 135.01, 134.30, 131.37, 129.09, 128.16, 126.54, 126.08, 106.58, 106.53, 44.86, 32.00, 31.62, 31.02, 30.68, 29.77, 29.73, 29.72, 29.65, 29.44, 28.34, 22.77, 21.13, 21.04, 20.69, 14.21. HRMS (DART) calcd for  $[C_{33}H_{48}Cl]^+$  ([M+H]<sup>+</sup>): 479.3439, found: 479.3434.



Following the general procedure **B**, **6d** was obtained as a liquid (35 mg, 35% yield). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.34 – 7.21 (m, 6H), 7.03 (d, *J* = 7.7 Hz, 2H), 3.52 (t, *J* = 6.4 Hz, 2H), 2.62 (t, *J* = 7.4 Hz, 2H), 2.49 – 2.40 (m, 2H), 2.24 (s, 3H), 2.02 – 1.91 (m, 2H), 1.53 – 1.45 (m, 2H), 1.36 – 1.26 (m, 2H), 1.23 (s, 9H), 1.21 – 1.14 (m, 16H), 0.80 (t, *J* = 6.7 Hz, 3H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  204.57, 149.85, 136.59, 133.76, 133.66, 129.25, 125.86, 125.54, 125.48, 109.85, 107.37, 45.03, 34.52, 31.99, 31.36, 31.05, 30.54, 29.84, 29.75, 29.71, 29.61, 29.43, 28.26, 27.53, 22.77, 21.14, 14.20. HRMS (DART) calcd for [C<sub>35</sub>H<sub>52</sub>Cl]<sup>+</sup> ([M+H]<sup>+</sup>): 507.3752, found: 507.3748.



Following the general procedure **B**, **6e** was obtained as a liquid (34 mg, 36% yield). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.34 – 7.28 (m, 2H), 7.22 – 7.15 (m, 2H), 7.11 (d, *J* = 7.9 Hz, 2H), 7.08 (d, *J* = 7.9 Hz, 1H), 3.60 (t, *J* = 6.5 Hz, 2H), 2.68 (t, *J* = 7.5 Hz, 2H), 2.57 – 2.48 (m, 2H), 2.33 (s, 3H), 2.24 (s, 6H), 2.09 – 1.98 (m, 2H), 1.60 – 1.51 (m, 3H), 1.41 – 1.34 (m, 2H), 1.30 – 1.21 (m, 16H), 0.88 (t, *J* = 6.8 Hz, 3H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  204.41, 136.65, 136.53, 135.34, 133.86, 129.83, 129.23, 127.08, 125.85, 123.29, 109.57, 107.50, 45.03, 31.98, 31.05, 30.49, 29.80, 29.74, 29.70, 29.61, 29.42, 28.20, 27.64, 22.75, 21.12, 20.01, 19.47, 14.19. HRMS (DART) calcd for [C<sub>33</sub>H<sub>48</sub>Cl]<sup>+</sup> ([M+H]<sup>+</sup>): 479.3439, found: 479.3434.



Following the general procedure A, **6f** was obtained as a liquid (58 mg, 62% yield). <sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.44 – 7.35 (m, 2H), 7.30 (d, *J* = 8.0 Hz, 2H), 7.13 (d, *J* = 8.0 Hz, 2H), 6.99 (t, *J* = 8.7 Hz, 2H), 3.60 (t, *J* = 6.4 Hz, 2H), 2.68 (t, *J* = 7.5 Hz, 2H), 2.53 (t, *J* = 7.6 Hz, 2H), 2.33 (s, 3H), 2.09 – 1.97 (m, 2H), 1.61 – 1.48 (m, 2H), 1.41 – 1.33 (m, 2H), 1.31 – 1.21 (m, 16H), 0.88 (t, *J* = 6.7 Hz, 3H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  204.39 (d, *J* = 2.0 Hz), 161.92 (d, *J* = 246.1 Hz), 136.82, 133.47, 132.69, 132.65, 129.32, 127.32 (d, *J* = 7.9 Hz), 125.83, 115.38 (d, *J* = 21.4 Hz), 110.15, 106.93, 44.91, 31.97, 30.93, 30.48, 29.76, 29.73, 29.70, 29.68, 29.58, 29.42, 28.20, 27.73, 22.75, 21.13, 14.19. HRMS (DART) calcd for [C<sub>31</sub>H<sub>43</sub>ClF]<sup>+</sup> ([M+H]<sup>+</sup>): 469.3032, found: 469.3028.



Following the general procedure **B**, **6g** was obtained as a liquid (42 mg, 43% yield). <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.38 – 7.33 (m, 2H), 7.32 – 7.25 (m, 4H), 7.16 – 7.11 (m, 2H), 3.61 (t, J = 6.4 Hz, 2H), 2.68 (t, J = 7.5 Hz, 2H), 2.53 (t, J = 7.6 Hz, 2H), 2.34 (s, 3H), 2.10 – 1.97 (m, 2H), 1.62 – 1.49 (m, 2H), 1.40 – 1.34 (m, 2H), 1.31 – 1.22 (m, 16H), 0.88 (t, J = 7.1 Hz, 3H). <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  204.79, 137.00, 135.33, 133.35, 132.61, 129.42, 128.71, 127.15, 125.92, 110.47, 107.04, 44.92, 32.04, 30.97, 30.52, 29.81, 29.79, 29.77, 29.75, 29.63, 29.48, 28.24, 27.59, 22.82, 21.20, 14.25. HRMS (DART) calcd for  $[C_{31}H_{43}Cl_2]^+$  ([M+H]<sup>+</sup>): 485.2736, found: 485.2732.



Following the general procedure **B**, **6h** was obtained as a liquid (75 mg, 77% yield). <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.36 – 7.27 (m, 3H), 7.15 (d, *J* = 7.9 Hz, 2H), 6.88 – 6.76 (m, 2H), 3.59 (t,

J = 6.5 Hz, 2H), 2.67 (d, J = 7.2 Hz, 2H), 2.49 (t, J = 7.8 Hz, 2H), 2.35 (s, 3H), 2.06 – 1.93 (m, 2H), 1.63 – 1.51 (m, 2H), 1.39 – 1.24 (m, 18H), 0.90 (t, J = 7.0 Hz, 3H). <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  204.91, 161.93 (dd, J = 267.5, 11.5 Hz), 160.28 (dd, J = 270.9, 12.2 Hz), 136.82, 133.56, 130.26 (dd, J = 8.8, 5.3 Hz), 129.33, 126.07, 121.82 (dd, J = 12.9, 2.9 Hz), 121.82 (dd, J = 12.9, 2.9 Hz), 108.11, 104.47 (t, J = 25.9 Hz), 102.58, 44.71, 32.05, 31.11, 30.45, 29.91, 29.80, 29.77, 29.72, 29.66, 29.49, 28.12, 22.82, 21.19, 14.24. HRMS (DART) calcd for  $[C_{31}H_{42}CIF_{2}]^{+}$  ( $[M+H]^{+}$ ): 487.2938, found: 487.2932.



Following the general procedure **B**, **6i** was obtained as a liquid (61 mg, 61% yield). <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.86 – 7.83 (m, 2H), 7.80 (d, *J* = 7.9 Hz, 1H), 7.74 (d, *J* = 8.6 Hz, 1H), 7.62 (dd, *J* = 8.6, 1.9 Hz, 1H), 7.49 – 7.43 (m, 2H), 7.38 – 7.36 (m, 2H), 7.16 (d, *J* = 8.0 Hz, 2H), 3.67 (t, *J* = 6.4 Hz, 2H), 2.87 (t, *J* = 7.5 Hz, 2H), 2.61 (t, *J* = 7.2 Hz, 2H), 2.36 (s, 3H), 2.19 – 2.11 (m, 2H), 1.67 – 1.57 (m, 2H), 1.45 – 1.39 (m, 2H), 1.31 – 1.23 (m, 16H), 0.90 (t, *J* = 7.1 Hz, 3H). <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  205.54, 136.88, 134.20, 133.83, 133.67, 132.64, 129.45, 129.40, 128.14, 128.03, 127.67, 126.26, 126.03, 125.98, 125.81, 125.33, 123.47, 110.33, 108.12, 45.11, 32.05, 31.14, 30.62, 29.87, 29.79, 29.76, 29.68, 29.49, 28.31, 27.63, 22.83, 21.22, 14.26. HRMS (DART) calcd for [C<sub>35</sub>H<sub>46</sub>Cl]<sup>+</sup> ([M+H]<sup>+</sup>): 501.3283, found: 501.3278.



Following the general procedure **B**, **6j** was obtained as a liquid (69 mg, 69% yield). <sup>1</sup>H NMR (600 MHz, Chloroform-*d*)  $\delta$  7.57 (d, J = 4.0 Hz, 1H), 7.31 – 7.25 (m, 2H), 7.12 (d, J = 8.0 Hz, 2H), 7.00 (d, J = 4.0 Hz, 1H), 3.59 (t, J = 6.4 Hz, 2H), 2.70 – 2.64 (m, 2H), 2.56 – 2.52 (m, 2H), 2.50 (s, 3H), 2.33 (s, 3H), 2.09 – 2.01 (m, 2H), 1.58 – 1.52 (m, 2H), 1.41 – 1.36 (m, 2H), 1.29 – 1.23 (m, 16H), 0.87 (t, J = 7.0 Hz, 3H). <sup>13</sup>C NMR (150 MHz, Chloroform-*d*)  $\delta$  205.25, 190.63, 151.59, 142.28, 137.49, 133.38, 132.60, 129.46, 126.27, 123.62, 111.90, 103.73, 44.60, 32.03, 30.92, 30.76, 29.78, 29.76, 29.61, 29.48, 28.43, 28.13, 26.56, 22.81, 21.23, 14.24. HRMS (DART) calcd for [C<sub>31</sub>H<sub>44</sub>OCIS]<sup>+</sup> ([M+H]<sup>+</sup>): 499.2796, found: 499.2792.

## Synthetic applications

### a) Synthesis of indenyl iodide 7 with NIS



In a flame-dried Schlenk tube was charged with allene (**1a**, 0.2 mmol), NIS (0.6 mmol) and CH<sub>3</sub>CN (1 mL). The resulting suspension was stirred at 80  $^{\circ}$ C for 10 h. Upon completion of the reaction as monitored by TLC, the solvent was concentrated under vacuum. The crude residue was purified by flash column chromatography on silica gel to give the products **7a** and **7b** as a mixture in 75% yield.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.37 – 7.18 (m, 3H), 7.14 – 6.78 (m, 5H), 3.70 – 3.34 (m, 2H), 2.88 – 2.53 (m, 2H), 2.51 – 2.00 (m, 6H), 1.77 – 1.55 (m, 1H), 1.46 – 1.05 (m, 20H), 0.89 – 0.85 (m, 3H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  152.40, 151.95, 148.43, 146.03, 143.01, 141.73, 140.75, 138.66, 136.45, 135.74, 129.20, 128.47, 127.59, 126.90, 126.69, 126.65, 126.55, 125.82, 124.13, 123.33, 118.79, 118.74, 116.58, 112.66, 63.58, 62.88, 45.58, 44.62, 34.55, 32.01, 31.99, 31.96, 31.15, 29.92, 29.90, 29.76, 29.72, 29.71, 29.68, 29.66, 29.62, 29.60, 29.44, 29.42, 29.39, 28.16, 27.20, 26.30, 22.78, 22.70, 21.56, 21.11, 14.23. HRMS (DART) calcd for  $[C_{31}H_{42}CII]^+$  ([M+H]<sup>+</sup>): 576.2014, found: 576.2008.

## b) Synthesis of 1 H-indene 8



In a flame-dried Schlenk tube was charged with allene (**6i**, 0.1 mmol),  $H_2SO_4$  (1 mL) and  $CH_3CH_2OH$  (0.2 mL). The resulting suspension was stirred at room temperature for 1 h. Upon completion of the reaction as monitored by TLC, the solvent was concentrated under vacuum. The crude residue was purified by flash column chromatography on silica gel to give the product **8** in 60% yield.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.22 (s, 1H), 7.22 – 7.15 (m, 2H), 7.12 (d, *J* = 8.0 Hz, 1H), 6.85 – 6.75 (m, 1H), 6.72 – 6.60 (m, 1H), 6.25 (s, 1H), 3.40 (t, *J* = 6.7 Hz, 2H), 2.58 – 2.43 (m, 3H), 2.41 (s, 3H), 2.22 – 2.07 (m, 1H), 1.70 – 1.56 (m, 3H), 1.42 – 1.23 (m, 19H), 0.88 (t, *J* = 6.7 Hz, 3H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  162.17 (dd, *J* = 250.2, 11.5 Hz), 161.80 (dd, *J* = 247.8, 12.3 Hz), 149.40, 143.33, 142.02, 134.14 (d, *J* = 3.0 Hz), 134.12, 129.00 (dd, *J* = 9.1, 6.7 Hz), 127.90, 125.81 (dd, *J* = 12.3, 3.8 Hz), 124.59 (d, *J* = 2.8 Hz), 119.60, 110.61 (dd, *J* = 20.3, 3.4 Hz), 104.68 (dd, *J* = 28.0, 25.0 Hz), 57.28 (d, *J* = 3.2 Hz), 45.50, 33.87, 33.81, 31.96, 29.72, 29.69, 29.67, 29.53, 29.40, 28.17, 27.90, 27.52, 22.74, 21.72, 14.17. HRMS (DART) calcd for  $[C_{31}H_{42}ClF_2]^+$  ([M+H]<sup>+</sup>): 487.2938, found: 487.2934.

#### c) Synthesis of 2H-pyran 9



In a flame-dried Schlenk tube was charged with allene (**4m**, 0.1 mmol),  $CF_3COOH$  (2 equiv) and DCM (0.2 mL). The resulting suspension was stirred at room temperature for 1 h. Upon completion of the reaction as monitored by TLC, the solvent was concentrated under vacuum. The crude residue was purified by flash column chromatography on silica gel to give the product **9** in 51% yield.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.39 (d, J = 7.3 Hz, 4H), 7.33 – 7.24 (m, 4H), 7.24 – 7.20 (m, 1H), 7.20 – 7.15 (m, 1H), 6.44 (s, 1H), 3.99 – 3.86 (m, 1H), 3.57 (td, J = 11.1, 3.6 Hz, 1H), 2.75 – 2.56 (m, 1H), 2.23 – 2.14 (m, 1H), 1.87 – 1.66 (m, 2H), 1.24 – 1.12 (m, 20H), 0.80 (t, J = 6.8 Hz, 3H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*) δ 145.95, 140.62, 134.90, 128.48, 128.21, 127.46, 126.74, 126.19, 125.10, 78.34, 60.47, 43.98, 31.95, 30.03, 29.67, 29.60, 29.39, 27.25, 23.90, 22.73, 14.17. HRMS (DART) calcd for [C<sub>29</sub>H<sub>41</sub>O]<sup>+</sup> ([M+H]<sup>+</sup>): 405.3152, found: 405.3149.

#### d) Synthesis of 2H-pyran 10



In a flame-dried Schlenk tube was charged with allene (**4m**, 0.1 mmol), NIS (2 equiv) and CH<sub>3</sub>CN (0.2 mL). The resulting suspension was stirred at room temperature for 10 h. Upon completion of the reaction as monitored by TLC, the solvent was concentrated under vacuum. The crude residue was purified by flash column chromatography on silica gel to give the product **10** in 56% yield.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.67 – 7.60 (m, 2H), 7.44 – 7.31 (m, 6H), 7.25 – 7.20 (m, 2H), 3.78 – 3.68 (m, 1H), 3.67 – 3.58 (m, 1H), 2.88 – 2.74 (m, 1H), 2.43 – 2.33 (m, 2H), 2.27 – 2.18 (m, 1H), 1.76 – 1.63 (m, 1H), 1.61 – 1.50 (m, 1H), 1.42 – 1.26 (m, 18H), 0.88 (t, *J* = 6.7 Hz, 3H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  146.25, 146.24, 142.31, 128.50, 128.33, 128.00, 127.85, 127.80, 127.56, 105.63, 81.80, 58.73, 40.83, 35.03, 31.98, 30.00, 29.79, 29.76, 29.74, 29.72, 29.69, 29.42, 23.36, 22.75, 14.19. HRMS (DART) calcd for  $[C_{29}H_{40}OI]^+$  ([M+H]<sup>+</sup>): 531.2118, found: 531.2114.

## Preliminary mechanism study

#### a) Radical trapping experiment



In a flame-dried Schlenk tube,  $Cu(CH_3CN)_4BF_4$  (0.01 mmol, 5 mol %) and Py-Box ligand (±)-L16 (0.014 mmol, 7 mol %) were dissolved in THF (1 mL) under a nitrogen atmosphere, and the mixture was stirred at room temperature for 30 minutes. Then 1,3-enyne (0.2 mmol, 1.0 equiv), peroxide (0.3 mmol, 1.5 equiv), PhB(OH)<sub>2</sub> (0.6 mmol, 3 equiv), DIPEA (0.6 mmol, 3 equiv) and TEMPO (3 mmol) were sequentially added. The reaction mixture was stirred at rt for 5 hours. The solution was filtrated with ethyl acetate on silica gel, and then detected by GC-MS. No desired coupling product was detected, but compound **11** was detected by GC-MS analysis.

#### b) Radical clock experiments

**Ring-opening reaction:** 



In a flame-dried Schlenk tube,  $Cu(CH_3CN)_4BF_4$  (0.01 mmol, 5 mol %) and Py-Box ligand (±)-L16 (0.014 mmol, 7 mol %) were dissolved in THF (1 mL) under a nitrogen atmosphere, and the mixture was stirred at room temperature for 30 minutes. Then 1,3-enyne 12 (0.2 mmol, 1.0 equiv), peroxide 2e (0.3 mmol, 1.5 equiv), PhB(OH)<sub>2</sub> (0.6 mmol, 3 equiv) and DIPEA (0.6 mmol, 3 equiv) were sequentially added. The reaction mixture was stirred at rt for 5 hours. Upon completion of the reaction as monitored by TLC, the solvent was concentrated under vacuum. The crude residue was purified by flash column chromatography on silica gel to give the product 13 in

52% yield.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.60 (d, J = 7.7 Hz, 2H), 7.46 – 7.26 (m, 6H), 7.26 – 7.17 (m, 2H), 3.46 (t, J = 6.7 Hz, 2H), 2.55 (t, J = 6.8 Hz, 2H), 1.82 – 1.67 (m, 3H), 1.59 – 1.49 (m, 4H), 0.89 – 0.85 (m, 2H), 0.64 – 0.40 (m, 2H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*) δ 204.22, 137.23, 136.54, 128.54, 128.44, 126.95, 126.93, 126.17, 125.80, 112.60, 110.03, 45.10, 32.56, 30.09, 27.28, 26.90, 11.33, 6.96, 6.90. HRMS (DART) calcd for  $[C_{23}H_{26}Cl]^+$  ([M+H]<sup>+</sup>): 337.1718, found: 337.1715.

#### **Ring closing reaction**



In a flame-dried Schlenk tube,  $Cu(CH_3CN)_4BF_4$  (0.01 mmol, 5 mol %) and Py-Box ligand (±)-L16 (0.014 mmol, 7 mol %) were dissolved in THF (1 mL) under a nitrogen atmosphere, and

the mixture was stirred at room temperature for 30 minutes. Then 1,3-enyne **14** (0.2 mmol, 1.0 equiv), peroxide **2e** (0.3 mmol, 1.5 equiv), PhB(OH)<sub>2</sub> (0.6 mmol, 3 equiv) and DIPEA (0.6 mmol, 3 equiv) were sequentially added. The reaction mixture was stirred at rt for 5 hours. Upon completion of the reaction as monitored by TLC, the solvent was concentrated under vacuum. The crude residue was purified by flash column chromatography on silica gel to give the product **15** in 58% yield.





<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.52 – 7.41 (m, 4H), 7.34 – 7.15 (m, 11H), 6.32 (d, J = 15.9 Hz, 1H), 6.20 (dt, J = 15.8, 6.8 Hz, 1H), 3.43 (t, J = 6.7 Hz, 2H), 2.67 – 2.53 (m, 4H), 2.30 (q, J = 6.9 Hz, 2H), 1.82 – 1.69 (m, 4H), 1.65 – 1.47 (m, 4H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*) δ 205.21, 137.78, 136.86, 136.82, 130.50, 130.33, 128.61, 128.58, 128.56, 126.95, 126.92, 126.01, 125.97, 109.21, 109.12, 45.16, 33.07, 32.61, 30.26, 29.90, 27.92, 27.45, 27.03. HRMS (DART) calcd for  $[C_{31}H_{34}CI]^+$  ([M+H]<sup>+</sup>): 441.2344, found: 441.2340.



In a flame-dried Schlenk tube,  $Cu(CH_3CN)_4BF_4$  (0.01 mmol, 5 mol %) and Py-Box ligand (±)-L16 (0.014 mmol, 7 mol %) were dissolved in THF (1 mL) under a nitrogen atmosphere, and the mixture was stirred at room temperature for 30 minutes. Then 1,3-enyne 1a (0.2 mmol, 1.0 equiv), peroxide 16 (0.3 mmol, 1.5 equiv), PhB(OH)<sub>2</sub> (0.6 mmol, 3 equiv) and DIPEA (0.6 mmol, 3 equiv) were sequentially added. The reaction mixture was stirred at rt for 5 hours. Upon completion of the reaction as monitored by TLC, the solvent was concentrated under vacuum. The crude residue was purified by flash column chromatography on silica gel to give the products 17a and 17b in 28% and 15% yields.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.46 – 7.41 (m, 2H), 7.35 – 7.28 (m, 4H), 7.24 – 7.19 (m, 1H), 7.13 (d, *J* = 8.0 Hz, 2H), 5.85 – 5.68 (m, 1H), 5.02 – 4.85 (m, 2H), 3.61 (t, *J* = 6.4 Hz, 2H), 2.70 (t, *J* = 7.4 Hz, 2H), 2.53 (t, *J* = 7.6 Hz, 2H), 2.33 (s, 3H), 2.11 – 1.96 (m, 4H), 1.62 – 1.56 (m, 2H), 1.45 – 1.36 (m, 4H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  204.66, 139.07, 136.73, 136.64, 133.53, 129.28, 128.54, 126.87, 125.83, 114.29, 109.84, 107.74, 44.98, 33.82, 30.98, 30.42, 29.23, 28.81, 28.01, 27.53, 21.13. HRMS (EI+) calcd for [C<sub>26</sub>H<sub>31</sub>Cl]<sup>+</sup> ([M]<sup>+</sup>): 378.2114, found: 378.2108.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.48 – 7.39 (m, 2H), 7.37 – 7.26 (m, 4H), 7.21 (t, *J* = 7.2 Hz, 1H), 7.13 (d, *J* = 8.0 Hz, 2H), 3.61 (t, *J* = 6.4 Hz, 2H), 2.71 (t, *J* = 7.5 Hz, 2H), 2.60 – 2.44 (m, 2H), 2.33 (s, 3H), 2.11 – 1.99 (m, 2H), 1.90 – 1.70 (m, 3H), 1.58 (td, *J* = 11.4, 10.3, 3.3 Hz, 4H),

1.53 - 1.40 (m, 2H), 1.17 - 1.03 (m, 2H).<sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  204.65, 136.70, 133.56, 129.27, 128.52, 126.83, 125.84, 125.82, 110.02, 107.60, 44.99, 40.21, 34.58, 32.73, 32.67, 30.97, 29.74, 27.51, 25.20, 21.13. HRMS (EI+) calcd for  $[C_{26}H_{31}CI]^+$  ([M]<sup>+</sup>): 378.2114, found: 378.2110.

#### c) Radical dimerization



In a flame-dried Schlenk tube,  $Cu(CH_3CN)_4BF_4$  (0.01 mmol, 5 mol%) and Py-Box ligand L10 (0.014 mmol, 7 mol%) were dissolved in THF (1 mL) under a nitrogen atmosphere, and the mixture was stirred at room temperature for 30 minutes. Then 1,3-enyne 18 (0.2 mmol, 1.0 equiv), peroxide 2h (0.3 mmol, 1.5 equiv), PhB(OH)<sub>2</sub> (0.6 mmol, 3 equiv) and DIPEA (0.6 mmol, 3 equiv) were sequentially added. The reaction mixture was stirred at rt for 5 h. Upon completion of the reaction as monitored by TLC, the solvent was concentrated under vacuum. The crude residue was purified by flash column chromatography on silica gel to give the products 19 in 28% yields, and a mixture of homocoupling dimers in 60% yield.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.35 – 7.27 (m, 4H), 7.29 – 7.15 (m, 6H), 7.06 (dd, J = 8.1, 4.1 Hz, 4H), 3.55 (s, 3H), 2.57 (t, J = 7.6 Hz, 2H), 2.34 – 2.21 (m, 8H), 2.02 – 1.83 (m, 2H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*) δ 206.46, 173.96, 137.10, 137.03, 136.93, 133.84, 132.97, 129.35, 129.19, 128.43, 128.32, 128.22, 127.25, 125.92, 113.07, 108.06, 51.55, 33.70, 29.83, 23.27, 21.23, 21.16. HRMS (ESI) calcd for  $[C_{28}H_{28}O_2Na]^+$  ([M+Na]<sup>+</sup>): 419.1982, found: 419.1981.


<sup>1</sup>H NMR (400 MHz, Chloroform-*d*)  $\delta$  7.62 (d, *J* = 7.8 Hz, 4H), 7.36 (d, *J* = 7.9 Hz, 4H), 7.14 (t, *J* = 8.9 Hz, 8H), 3.54 (s, 6H), 2.60 – 2.44 (m, 2H), 2.37 (s, 6H), 2.36 (s, 6H), 2.24 – 2.14 (m, 4H), 1.65 – 1.57 (m, 4H), 1.43 – 1.32 (m, 2H). <sup>13</sup>C NMR (100 MHz, Chloroform-*d*)  $\delta$  173.99, 137.97, 136.52, 136.00, 131.32, 129.08, 127.80, 120.65, 91.79, 87.88, 54.69, 51.36, 35.65, 34.31, 21.52, 21.12, 21.07. HRMS (ESI) calcd for [C<sub>44</sub>H<sub>46</sub>O<sub>4</sub>Na]<sup>+</sup> ([M+Na]<sup>+</sup>): 661.3288, found: 661.3289.

## Single crystal data of 20



Table S2. Crystal data and structure refinement for data.

Identification code	Compound-20	Compound-20	
Empirical formula	C44 H46 O4	C44 H46 O4	
Formula weight	638.81	638.81	
Temperature	99.99(10) K	99.99(10) K	
Wavelength	1.3405 Å		
Crystal system	Triclinic		
Space group	P -1		
Unit cell dimensions	a = 10.8484(4)  Å	α= 76.980(4)°.	
	b = 11.0591(5) Å	$\beta = 84.293(3)^{\circ}.$	
	c = 15.2489(6) Å	$\gamma = 86.909(3)^{\circ}.$	
Volume	1772.66(13) Å <sup>3</sup>		
Z	2		
Density (calculated)	1.197 Mg/m <sup>3</sup>	1.197 Mg/m <sup>3</sup>	
Absorption coefficient	0.374 mm <sup>-1</sup>	0.374 mm <sup>-1</sup>	
F(000)	684	684	
Crystal size	$0.15 \ge 0.08 \ge 0.07 \text{ mm}^3$	0.15 x 0.08 x 0.07 mm <sup>3</sup>	
Theta range for data collection	2.596 to 60.549°.	2.596 to 60.549°.	
Index ranges	-13<=h<=14, -14<=k<=	-13<=h<=14, -14<=k<=14, -18<=l<=19	
Reflections collected	24617	24617	
Independent reflections	7932 [R(int) = 0.0281]	7932 [R(int) = 0.0281]	
Completeness to theta = $53.543^{\circ}$	99.9 %	99.9 %	
Absorption correction	Semi-empirical from eq	Semi-empirical from equivalents	
Max. and min. transmission	1.00000 and 0.81422	1.00000 and 0.81422	
Refinement method	Full-matrix least-square	Full-matrix least-squares on F <sup>2</sup>	
Data / restraints / parameters	7932 / 0 / 439	7932 / 0 / 439	
Goodness-of-fit on F <sup>2</sup>	1.085		
	37		

Final R indices [I>2sigma(I)]	R1 = 0.0449, wR2 = 0.1234
R indices (all data)	R1 = 0.0516, wR2 = 0.1283
Extinction coefficient	n/a
Largest diff. peak and hole	0.401 and -0.291 e.Å-3

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## NMR spectra





























































































































































