## **Supporting Information for**

The supramolecular polymer complexes with oppositely charged calixresorcinarene: the hydrophobic domains formation and the synergistic binding mode

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macrocycles in $D_2O$ solutions (1 mM) from NMR FT-PGSE experiments						
Macrocycle	тт	D 1010 2/ -	$R_{H}$ , Å		17	
	рН	$D_s \times 10^{-10},  {\rm m^{2}/s}$	exp	teor <sup>[a]</sup>	N <sub>ag</sub>	
C <sub>1</sub> OPh-CR	8.04	2.55 <sup>[b,c]</sup>	10.9	8.4	2.2 <sup>d</sup>	
C <sub>5</sub> OPh-CR	8.08	2.06	13.5	9.7	2.7	
C <sub>5</sub> -CR	7.86	2.49 <sup>[b]</sup>	11.2	8.6	2.2	
C <sub>8</sub> -CR	7.97	1.85 <sup>[b]</sup>	15.0	9.1	4.5	
C <sub>11</sub> -CR	8.10	1.04 <sup>[b]</sup>	26.7	9.5	22	

**Table S1**. pH-values of 1 mM solutions of carboxycalixresorcinarenes in an aqueous solution; self-diffusion coefficients ( $D_s$ ), hydrodynamic radius ( $R_H$ ), and aggregation numbers ( $N_{ag}$ ) of macrocycles in D<sub>2</sub>O solutions (1 mM) from NMR FT-PGSE experiments

a - the value of  $R_{H}^{\text{teor}}$  was estimated with the help of HYDRONMR program [1]; b - the data from ref [2]; c -  $C(C_1\text{OPh-CR}) = 0.5 \text{ mM}$ ; d - The aggregation numbers of calixresorcinarenes Nag were estimated as described in ref [3]:  $Nag = (R_{H}^{\text{agr}}/R_{H}^{\text{mon}})^3$ , where  $R_{H}^{\text{agr}}$  and  $R_{H}^{\text{mon}}$  are hydrodynamic radii of the molecules in aggregated and monomeric states,  $R_{H}^{\text{mon}} = R_{H}^{\text{teor}}$ .

**Table S2**. pH-values, the values of averaged hydrodynamic diameters of particles (d, nm), their intensities of scattering (I, %), and polydispersity index (PDI) in individual PEI solutions and in the presence of  $C_n$ -CR (1 mM).

Macrocycle	C(PEI), mM	d <sub>1</sub> , nm (I <sub>1</sub> , %)	d <sub>2</sub> , nm (I <sub>2</sub> , %)	PDI	ξ, mV	pH
	0.5	16 (2.6)	106 (17.5)	0.284	+33.0	9.65
	1	18 (3.7)	91 (22.2)	0.312	+22.2	9.53
-	5	24 (2.8)	106 (16.8)	0.32	+15.1	10.59
	10	28 (3.0)	106 (13.8)	0.399	+14.9	10.35
	20	21 (2.4)	106 (12.8)	0.461	+13.2	10.83
	0.5	-	220 (39.8)	0.007	-23.0	10.44
	1	-	295 (31.1)	0.159	-17.0	10.72
C₁OPh-CR C₅OPh-CR	5	28 (5.9)	106 (16)	0.288	-12.7	10.49
	10	21 (3.4)	91 (15.1)	0.286	-10.3	11.09
	20	28 (5.7)	106 (14)	0.313	-7.6	11.26
	0.5	28 (7.5)	68 (12.9)	0.203	-44.1	10.78
	1	-	37 (15)	0.151	-29.7	11.21
C5OPh-CR	5	24 (2.7)	78 (16.7)	0.165	-19.1	10.87
	10	16 (1.8)	79 (11.9)	0.278	-15.0	11.65
	20	14 (2.1)	79 (10.3)	0.302	-12.6	11.59
	0.5	-	58 (13.7)	0.15	-55.6	10.78
	1	-	58 (20.4)	0.089	-34.5	11.11
C <sub>5</sub> -CR	5	21 (3.9)	91 (18)	0.283	-21.7	10.77
	10	12 (3.1)	79 (14.1)	0.288	-15.3	11.52
	20	24 (3.3)	91 (14.5)	0.376	-10.2	11.55
	0.5	18 (2.1)	58 (14.3)	0.186	-55.5	11.39
	1	-	68 (14.6)	0.182	-46.7	11.39
C <sub>8</sub> -CR	5	24 (3.6)	91 (12.7)	0.295	-27.3	11.00
	10	18 (2.6)	91 (14.5)	0.300	-22.5	11.86
	20	16 (2.2)	91 (14.4)	0.310	-13.7	11.65
	0.5	-	78 (14.3)	0.192	0	10.92
C <sub>1</sub> OPh-CR C <sub>5</sub> OPh-CR C <sub>5</sub> -CR C <sub>8</sub> -CR C <sub>11</sub> -CR	1	-	68 (14.1)	0.228	0	11.37
	5	18 (3.6)	78 (14.3)	0.277	0	10.94
	10	14 (2.4)	68 (13.8)	0.251	0	11.13
	20	18 (4.3)	91 (14.6)	0.311	0	11.59



**Figure S1.** The intensity-averaged size distribution for PEI and PEI -  $C_n$ -CR solutions (DLS method, 25 °C, C( $C_n$ -CR) 1 mM, C(PEI) 0.5, 5 and 10 mM).



Figure S2. The conformational structures of Cn-CR (according to H<sup>1</sup> NMR data [2, 4]).



Figure S3. The I/III dependence of values of pyrene on concentration of Cn-CR in the absence and presence of 5 and 20 mM PEI:  $a - for C_1OPh-CR$ ,  $b - for C_5OPh-CR$ ,  $c - for C_5-CR$ .



**Figure S4.** UV VIS spectra of Orange OT after solubilization by individual and mixed solutions of PEI (a), C1OPh-CR (b), C5-CR (c), C5OPh-CR (d), C8-CR (e), C11-CR (f)



**Figure S5.** a – The photo of  $C_8$ -CR - PEI (left) and  $C_8$ -CR - PEI – Orange OT (on right) solutions (C(PEI) 5 mM). b, c – The intensity-averaged size distribution for  $C_8$ -CR – PEI – Orange OT (b) and  $C_5$ OPh-CR - PEI – Orange OT (c) solutions (C(PEI) 5 mM).



**Figure S6.** Emission spectra of CV (0.001 mM) in individual solution and in the individual and mixed solutions of Cn-CR (1 mM) and PEI (5 mM);  $\lambda_{ex}$  580 nm, voltage 800V, (a) and (b) - ex.slit 10 nm, em.slit 10 nm; (c) - ex.slit 5 nm, em.slit 5 nm, the spectrum of C5OPh-CR - PEI solution is added as a blank because a weakly fluorescent "background" from PEI was observed. The emission maximum of CV (633 nm) is shifted to 628 nm in the presence of tetra-alkyl macrocycles C<sub>5</sub>-CR, C<sub>8</sub>-CR, and C<sub>11</sub>-CR and to 635 nm in the presence of tetra-phenylenoxyalkyl macrocycles C<sub>1</sub>OPh-CR and C<sub>5</sub>OPh-CR due to the binding. Probably, this difference between observed shifts occurs due to the binding of the dye near the phenylenoxy groups of macrocycles in the last case.

**Table S3.** The NMR FT-PGSE data for PEI<sup>a</sup> and C<sub>5</sub>-CR and C<sub>5</sub>OPh-CR (D<sub>2</sub>O, 303 K):  $D_s$  - the self-diffusion coefficient of macrocycle,  $P_b$  - fraction of macrocycle bound by PEI,  $D_s^{\text{fast}}$ ,  $D_s^{\text{slow}}$ ,  $P^{\text{fast}}$ , and  $P^{\text{slow}}$  - the self-diffusion coefficient and the population for fast and slow components in two exponential analyses of PEI diffusion slope, respectively;  $D_s^P$  - the population weighted average of the diffusion coefficients of PEI;  $R_H \text{fast/slow}$  - experimental value of hydrodynamic radius of fast and slow components of PEI, respectively;  $C_{\text{Cn-CR}}$  1 mM.

Cn-CR C(	C(PEI),	$D_s \times 10^{-10}$ , $m^{2/s}$	$D_s \times 10^{-10},  \mathrm{m}^{2/s}$	s (P) PEI	$D_s^P \times 10^{-10}$ , m <sup>2</sup> /s PEI	$R_H$ fast/slow,	P <sub>b</sub> Cn-CR
	mM	Cn-CR	fast slow	slow		PEI	
PEI	5	-	1.59 (0.40)	0.21 (0.60)	0.75	18 / ~135	-
C <sub>5</sub> -CR	0	2.49	-	-	-	-	-
	0.25	2.47	b	b	-	-	-
	2.5	1.96	1.53 (0.73)	0.25 (0.27)	1.19	18 / 110	0.49
	5	1.81	1.43 (0.63)	0.25 (0.37)	0.99	20 / 110	0.52
	10	1.39	1.72 (0.45)	0.25 (0.55)	0.91	16 / 110	0.73
	20	1.03	1.65 (0.43)	0.26 (0.57)	0.85	17 / 110	0.90
	0	2.06	-	-	-	-	-
Cn-CR PEI C₅-CR C₅OPh-CR	0.25	2.03	b	b	-	-	-
	0.5	2.08	b	b	-	-	-
	0.75	2.08	b	b	-	-	-
	1	2.06	b	b	-	-	-
	2.5	1.96	1.65 (0.53)	0.25 (0.47)	0.99	17 / 110	0.10
	5	1.87	1.65 (0.74)	0.28 (0.26)	1.29	17 / 100	0.24
	10	1.26	1.90 (0.47)	0.29 (0.53)	1.04	15 / 100	0.78
	20	0.89	1.73 (0.53)	0.26 (0.47)	1.05	16 / 105	1.16 <sup>c</sup>

a - The concentration of PEI is per monomer unit in all case; b - the short times of transverse relaxation T2\* for polymer signals are observed that don't allow to estimate PEI self-diffusion coefficient; c - the macrocycle is bound by slow component of PEI, that leads to the overestimation of its  $P_b$  value.



**Figure S7.** 2D NOESY spectrum of  $C_5$ -CR – PEI solution (C( $C_5$ -CR) 1 mM, C(PEI) 5 mM) in D<sub>2</sub>O, the red circle indicates the cross peaks between signals of macrocycles groups, the blue circle indicates the cross peaks between signals of groups of macrocycle and PEI.

In the 2D NOESY spectrum the cross peaks between signals of ethylene groups of PEI and hydrophilic and aliphatic groups of the macrocycle are observed in the 2D NOESY spectrum (indicated by blue circles). The closeness of CH<sub>2</sub>O groups of the macrocycle and NCH<sub>2</sub>CH<sub>2</sub>N groups of PEI are occured as a result of electrostatic interaction of charged groups of components. The cross peaks between NCH<sub>2</sub>CH<sub>2</sub>N groups of PEI and pentyl groups of the macrocycle can mean the inclusion of PEI fragments in macrocycle dimer or enveloping of macrocycle aliphatic groups by polymer chain. The cross peaks between signals of macrocycles groups indicated by red circles can be intra- or intermolecular.



**Figure S8.** 2D NOESY spectrum of  $C_5OPh-CR - PEI$  solution (C( $C_5OPh-CR$ ) 1 mM, C(PEI) 5 mM) in D<sub>2</sub>O, the red circle indicates the cross peaks between signals of macrocycles groups, the blue circle indicates the cross peaks between signals of groups of macrocycle and PEI.

In the 2D NOESY spectrum the cross peaks between signals of PEI fragments and aromatic, aliphatic and hydrophilic groups of the macrocycle indicated by blue circles as well as the cross peaks between signals of aromatic and aliphatic groups and hydrophilic and aliphatic groups of macrocycle molecules are observed in the 2D NOESY spectrum.



**Figure S9.** 2D NOESY spectrum of  $C_5$ OPh-CR – PEI solution (C( $C_5$ OPh-CR) 1 mM, C(PEI) 20 mM) in D<sub>2</sub>O, the blue circle indicates the cross peaks between signals of groups of macrocycle and PEI.

The cross peaks between signals of PEI fragments and aromatic, aliphatic and hydrophilic groups of the macrocycle indicated by blue circles are observed in the 2D NOESY spectrum. The cross peaks between signals of PEI fragments and aliphatic groups of the macrocycle are very strong.



**Figure S10.** The proposed scheme of  $C_8OPh-CR + PEI$  (up) and  $C_8OPh-CR + CV + PEI$  (down) complexes.

In the scheme the electrostatic and hydrophobic interactions between the macrocycle and PEI as well as the increasing of CV emission as a result of partial participation of PEI in the bounding of CV are took in to the account. The inner part of complex comprises alkyl groups of macrocycle and some fragments of PEI, and the upper part is mainly composed by hydrophilic groups of macrocycle molecules.



**Figure S11.** The proposed scheme of  $C_1OPh-CR + PEI$  (up) and  $C_1OPh-CR + CV + PEI$  (down) complexes. The molecules of  $C_1OPh-CR$  are in chair conformation.

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