

# Supplementary Information

## A Label-Free Aptamer Based Cytosensor for Specific Cervical Cancer HeLa Cell Recognition through g- C<sub>3</sub>N<sub>4</sub>-AgI/ITO Photoelectrode

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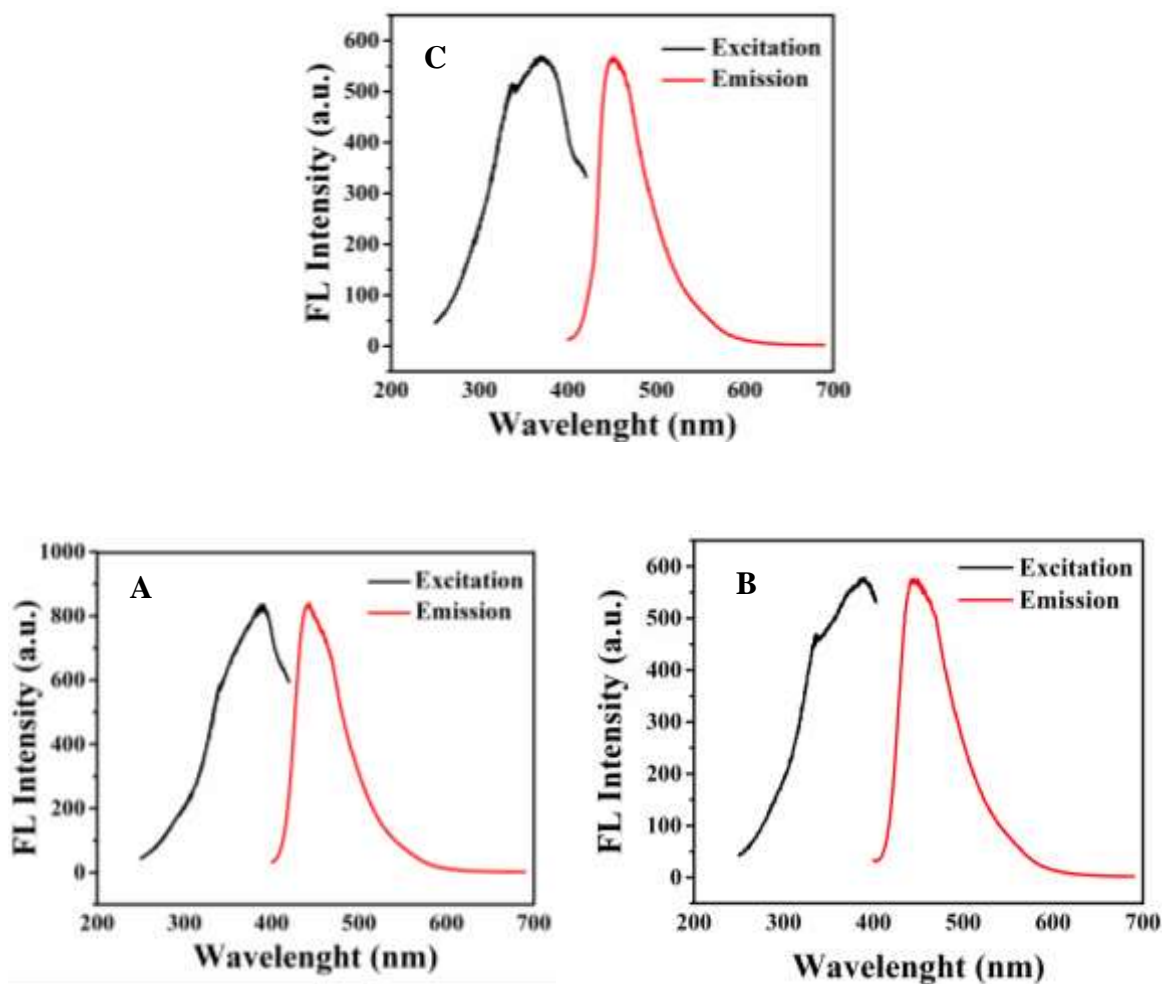
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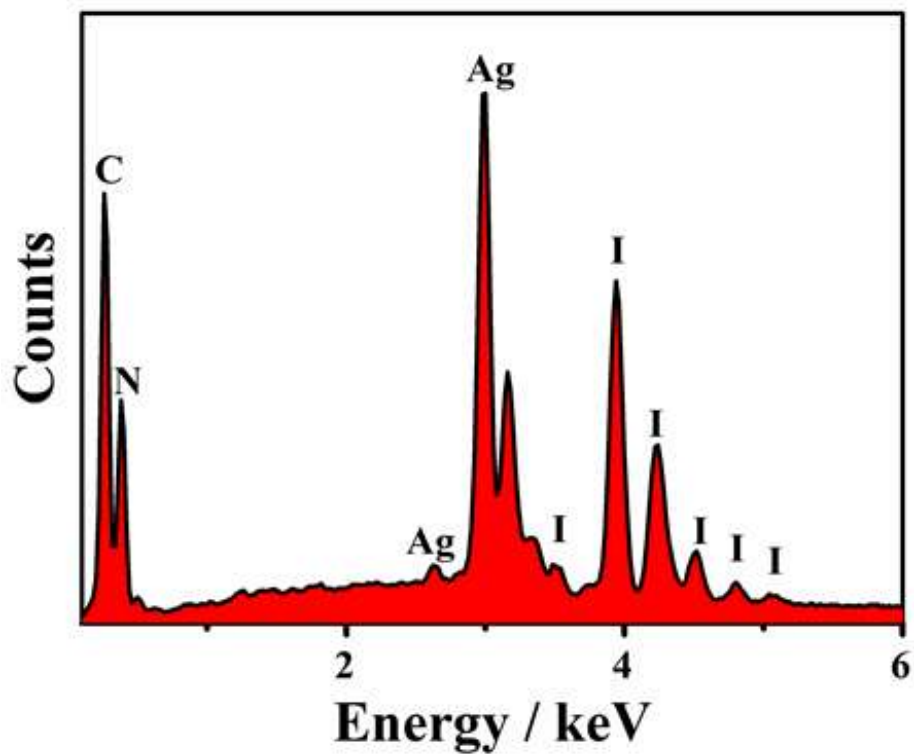
**KEYWORDS:** g-C<sub>3</sub>N<sub>4</sub>-AgI nanocomposites; photoelectrochemistry; aptamer; HeLa cancer cells; cytosensor

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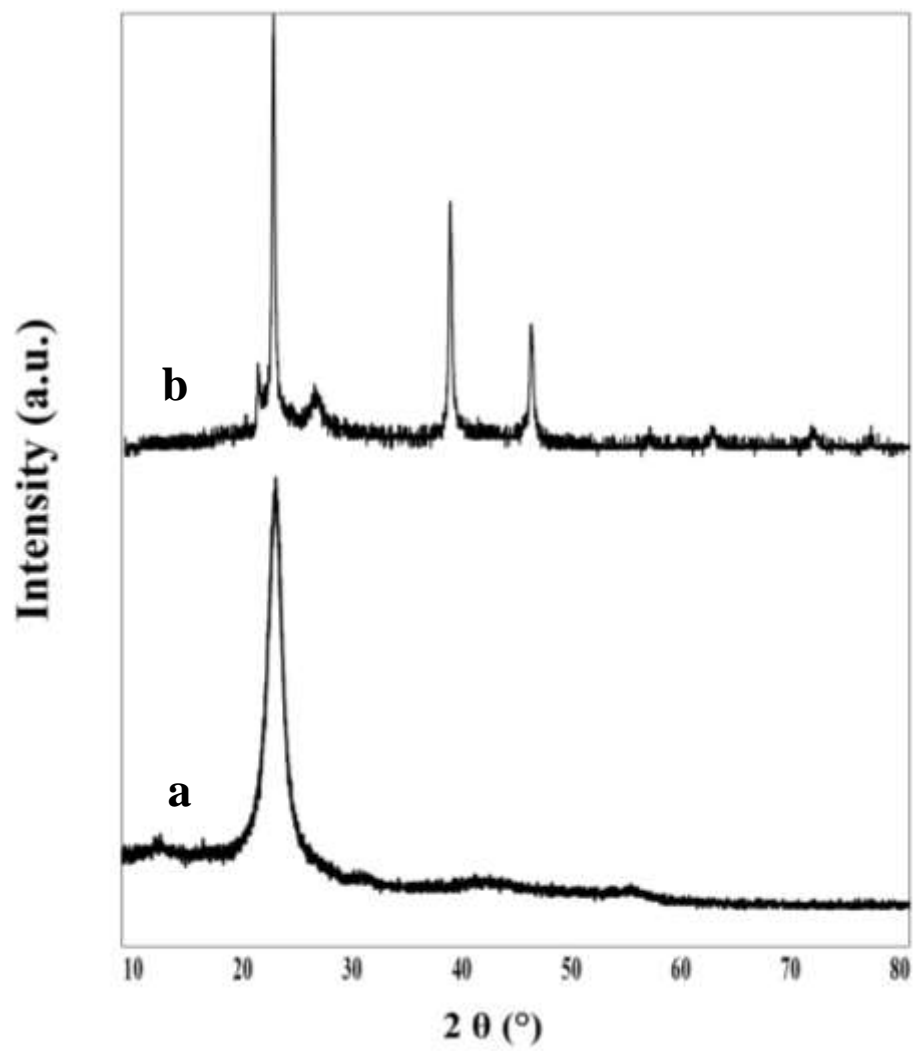
**Figure S1.** Fluorescence characterization; (A) excitation (388 nm) and emission (442 nm) spectra of the  $g\text{-C}_3\text{N}_4$ , (B) excitation (378 nm) and emission (446 nm) spectra of the  $g\text{-C}_3\text{N}_4\text{-AgBr}$  and (C) excitation (365 nm) and emission (446 nm) spectra of the  $g\text{-C}_3\text{N}_4\text{-AgI}$ .



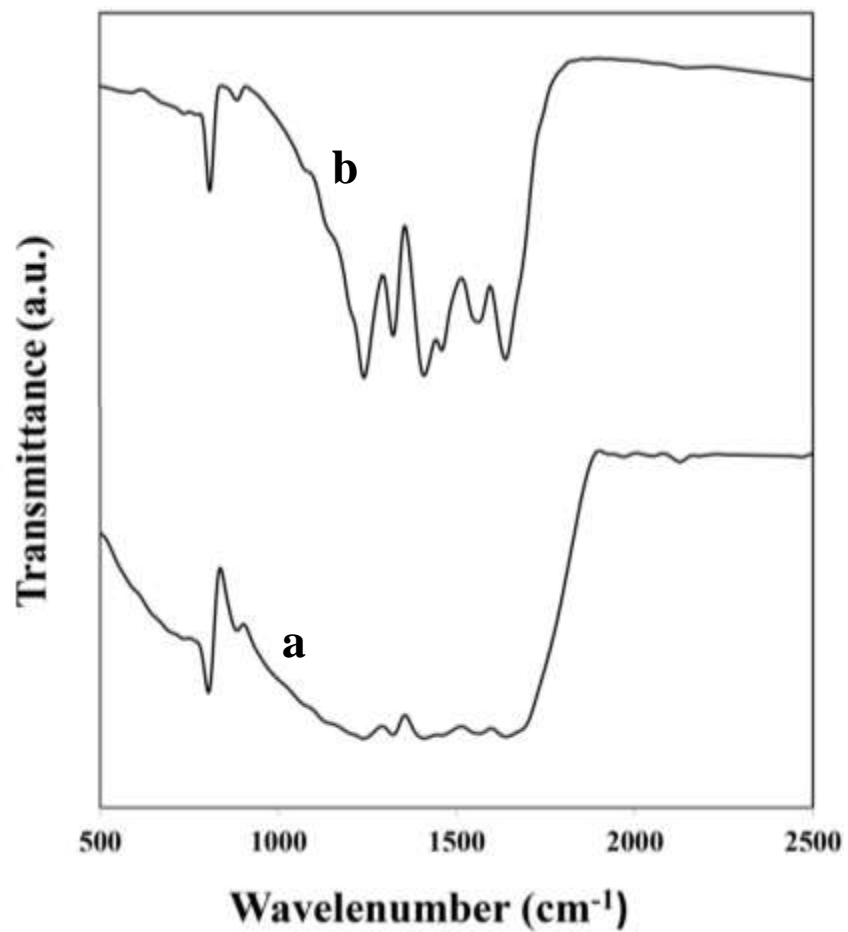
**Figure S2.** EDS analysis of g-C<sub>3</sub>N<sub>4</sub>-AgI

**Table S1.** Elements weight and atomic ratio of the synthesized g-C<sub>3</sub>N<sub>4</sub>-AgI.

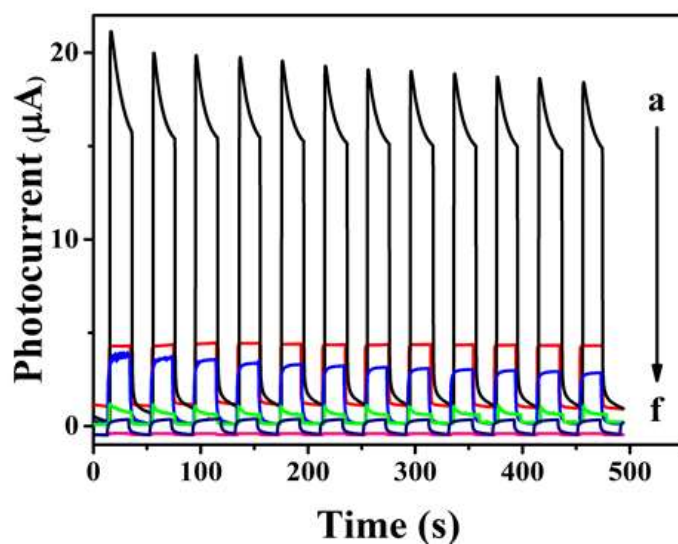
Element	Weight (%)	Atom (%)
C	16.34	29.86
N	39.48	61.86
Ag	20.96	4.26
I	23.23	4.02
<b>Total</b>	<b>100.00</b>	<b>100.00</b>



**Figure S3.** X-ray diffraction patterns of (a) g-C<sub>3</sub>N<sub>4</sub> and (b) g-C<sub>3</sub>N<sub>4</sub>-AgI nanocomposites



**Figure S4.** FTIR spectra of (a) g-C<sub>3</sub>N<sub>4</sub> and (b) g-C<sub>3</sub>N<sub>4</sub>-AgI samples

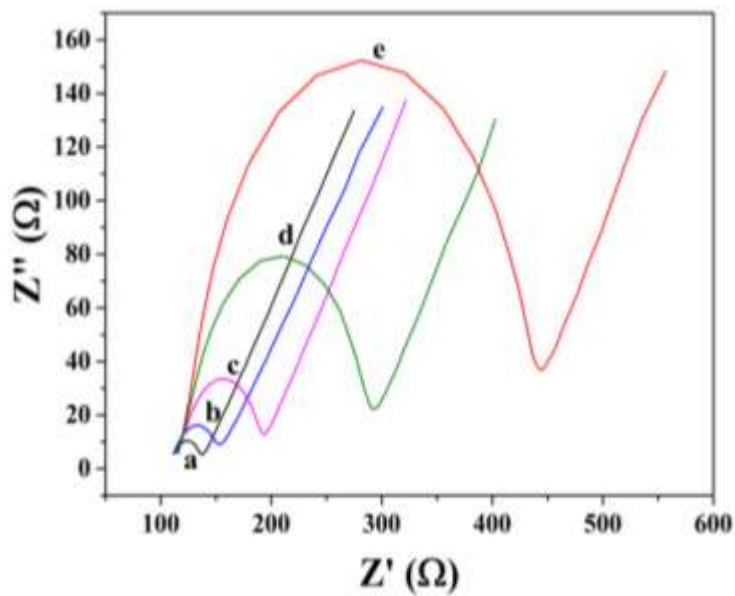


**Figure S5.** Time based photocurrent comparison of different photoelectrodes ITO/C<sub>3</sub>N<sub>4</sub>-AgI (**a**, **b**), ITO/C<sub>3</sub>N<sub>4</sub>-AgBr (**c**, **d**) and ITO/C<sub>3</sub>N<sub>4</sub> (**e**, **f**) in buffer solution in the presence (**a**, **c**, **e**) and absence (**b**, **d**, **f**) of K<sub>4</sub>[Fe(CN)<sub>6</sub>].

**Table S2.** Comparison of analytical properties of different methods towards Hela cells

<b>Method</b>	<b>Linearity Range (cell/ml)</b>	<b>LOD (cell/ml)</b>	<b>Reference</b>
Differential pulse voltammetry	$3.0 \times 10^2 - 1.0 \times 10^7$	300	1
Differential pulse voltammetry	$10 - 10^6$	10	2
Differential pulse voltammetry	$10 - 10^6$	10	3
Electrochemical impedance spectroscopy	$30 - 10^6$	10	4
Differential pulse voltammetry	$10 - 10^6$	10	5
Cyclic voltammetry	$250 - 5 \times 10^3$	250	6
Electrochemical impedance spectroscopy	$2.4 \times 10^2 - 2.4 \times 10^5$	90	7
Photoelectrochemistry	$10 - 10^6$	5	This work





**Figure S6.** Characterization of different modification steps of the ITO surface by Nyquist plots of EIS in  $5 \times 10^{-4}$  M  $[\text{Fe}(\text{CN})_6]^{3-}/[\text{Fe}(\text{CN})_6]^{4-}$  (1:1) and 0.1M KCl solution of layer-by-layer assembled electrode (a-d), ITO (a), ITO/ $\text{C}_3\text{N}_4$ -AgI (b), ITO/ $\text{C}_3\text{N}_4$ -AgI /Aptamer, (c) ITO/ $\text{C}_3\text{N}_4$ -AgI/ Aptamer/BSA (d) / ITO/ $\text{C}_3\text{N}_4$ -AgI/ Aptamer / BSA/ Hella (e).

**Table S3.** Influence of interferences for the designed biosensor

<b>Interferences</b>	<b>Tolerance Level</b>
Galactose	1000
Glucose	1000
L- cysteine	50
Tryptophan	50
Ribose	1000
Dioxoribose	1000
Cholesterin	50
Ascorbic Acid	0
Fe <sup>2+</sup>	0
Fe <sup>3+</sup>	0
K <sup>+</sup>	1000
NO <sub>3</sub> <sup>-</sup>	1000
Cl <sup>-</sup>	1000
Na <sup>+</sup>	1000
SO <sub>4</sub> <sup>2-</sup>	1000
NO <sub>3</sub> <sup>-</sup>	1000
ClO <sub>3</sub> <sup>-</sup>	0

## References

1. X. Wang, J. Ju, J. Li, J. Li, Q. Qian, C. Mao and J. Shen, *Electrochim. Acta*, 2014, **123**, 511-517.
2. J. Liu, Y. Qin, D. Li, T. Wang, Y. Liu, J. Wang and E. Wang, *Biosens. Bioelectron.*, 2013, **41**, 436-441.
3. S. Xu, J. Liu, T. Wang, H. Li, Y. Miao, Y. Liu, J. Wang and E. Wang, *Talanta*, 2013, **104**, 122-127.
4. H. Shen, J. Yang, Z. Chen, X. Chen, L. Wang, J. Hu, F. Ji, G. Xie and W. Feng, *Biosens. Bioelectron.*, 2016, **81**, 495-502.
5. T. Wang, J. Liu, X. Gu, D. Li, J. Wang and E. Wang, *Anal. Chim. Acta*, 2015, **882**, 32-37.
6. J. J. Castillo, W. E. Svendsen, N. Rozlosnik, P. Escobar, F. Martinez and J. Castillo-Leon, *Analyst*, 2013, **138**, 1026-1031.
7. Z. Wang, S. Chen, C. Hu, D. Cui and N. Jia, *Electrochem. Commun.*, 2013, **29**, 4-7.