SUPPORTING INFORMATION

Highly Selective, Red Emitting BODIPY-Based Fluorescent Indicators for Intracellular Mg²⁺ Imaging

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1. Supporting figures and schemes



Figure S1. Absorption spectrum of **MagQ1** in 50 mM PIPES, 100 mM KCl, pH 7.0 buffer, 25 °C before (black line) and after (red line) treatment with Mg^{2+} .



Figure S2. Fluorescence response of **MagQ1** to increasing concentrations of Mg^{2+} in 50 mM aqueous PIPES buffer, 100 mM KCl, pH 7.0, 25 °C. (A) Non-linear fit (red line) of the integrated fluorescence emission (black circles) as a function of total magnesium concentration, $[Mg^{2+}]_t$, using a 1:1 metal-to-indicator binding model. (B) Integrated fluorescence emission as a function of total magnesium concentration, $[Mg^{2+}]_t$, at low magnesium concentrations. (C) Benesi-Hildebrand plot.



Scheme S1. Synthesis of MagQ1-AM and MagQ2-AM.



Figure S3. Viability of HeLa cells treated with MagQ1 (in the membrane permeable acetoxymethyl ester form, MagQ1-AM) under typical sensor loading and imaging conditions, as determined by CellTiter-Glo® Luminescent Cell Viability Assay. Error bars correspond to standard deviations on eight replicas.



Figure S4. Normalized average fluorescence of **MagQ1** per cell in samples imaged in normal growth medium (black triangles) or under supplementation with exogenous Ca^{2+} and ionophore (violet triangles). Error bars represent the SD, N = 20 cells.



Figure S5. Fluorescence imaging of Mg^{2+} in live HeLa cells loaded with **MagQ1-AM** over a period of 30 min. Cells incubated in HHBSS, containing 0.8 mM Mg^{2+} , and 1.8 mM Ca^{2+} . Scale bar: 50 μ m.



Figure S6. (A) Absorption and (B) fluorescence emission spectra of a 1.0 μ M solution of **MagQ2** treated with increasing concentrations of Mg²⁺. Titration conducted in 50 mM aqueous PIPES buffer, 100 mM KCl, pH 7.0, 25 °C. Excitation wavelength $\lambda_{ex} = 600$ nm. * = scattered light from excitation beam.



Figure S7. Fluorescence response of **MagQ2** to increasing concentrations of Mg^{2+} in 50 mM aqueous PIPES buffer, 100 mM KCl, pH 7.0, 25 °C. (A) Non-linear fit (red line) of the integrated fluorescence emission (black circles) as a function of total magnesium concentration, $[Mg^{2+}]_t$, using a 1:1 metal-to-indicator binding model. (B) Integrated fluorescence emission as a function of total magnesium concentration, $[Mg^{2+}]_t$, at low magnesium concentrations. (C) Benesi-Hildebrand plot.



Figure S8. (A) Fluorescence response of 1 μ M MagQ2 to physiological concentration of Mg²⁺ (1 mM) or to other divalent metal ions (10 μ M) in aqueous buffer at 25 °C. (B) Fluorescence response of 1.0 μ M MagQ2 to 1 mM or 100 mM of Mg²⁺ in the presence of competing divalent cations, showing the selectivity of the detection in 50 mM PIPES, 100 mM KCl, pH 7.0 buffer. Error bars correspond to standard deviations on measurements conducted in triplicate.



Figure S9. Fluorescence emission of a 1 μ M solution of **MagQ2** in aqueous buffer at pH ranging from 5.5 to 8.0, 25 °C, in the absence (black squares) or presence (red circles) of 100 mM Mg²⁺. Error bars correspond to standard deviations on measurements conducted in triplicate.



Figure S10. Fluorescence imaging of Mg^{2+} in live HeLa cells loaded with **MagQ2-AM** over a period of 30 min. Cells incubated in HHBSS, containing 0.8 mM Mg^{2+} and 1.8 mM Ca^{2+} . Scale bar: 50 μ m.



Figure S11. Viability of HeLa cells treated with **MagQ2** (in the membrane permeable acetoxymethyl ester form, **MagQ2-AM**) under typical sensor loading and imaging conditions, as determined by CellTiter-Glo® Luminescent Cell Viability Assay. Error bars correspond to standard deviations on eight replicas.

2. Additional experimental details

2.1. General procedure for the preparation of quantitative solutions of MagQ1 and MagQ2 A quantitative sample of **MagQ1** was weighed and dissolved in DMSO to prepare a 1.0 mM

stock solution in a volumetric flask. The solution was divided into small aliquots, flash frozen in liquid nitrogen and stored at -20 °C until use.

An aliquot of the stock solution was diluted in aqueous buffer (50 mM PIPES, 100 mM KCl, pH 7.0) to obtain a series of solutions with concentrations ranging from 0.2 to 1.0 μ M that were used to determine the molar absorptivity at 600 nm, 25 °C ($\epsilon = 84000 \pm 1000 \text{ M}^{-1} \cdot \text{cm}^{-1}$). This molar absorptivity was used in subsequent experiments to quantify the concentrations of **MagQ1** and **MagQ2** in various samples and stock solutions.







-20 -40 -60 -80 -100 -120 -140 -160 -180 ppm *Figure S14.* ¹⁹F NMR spectrum of **3** in CDCl₃









Figure S21. ¹H NMR spectrum of **7** in acetone-d₆





Sig=254 Ref=800 mAU Sig=600 Ref=800 mAU

200 · 0 ·

Figure S25. Reversed phase HPLC chromatogram of MagQ1, eluted with an acetonitrile/water (+0.1% TFA) gradient.

min

min



Figure S26. Reversed phase HPLC chromatogram of MagQ2, eluted with an acetonitrile/water (+0.1% TFA) gradient.



Figure S27. Reversed phase HPLC chromatogram of MagQ1-AM, eluted with an acetonitrile/water (+0.1% TFA) gradient.



Figure S28. Reversed phase HPLC chromatogram of MagQ2-AM, eluted with an acetonitrile/water (+0.1% TFA) gradient.