Tailored fluorescent solvatochromic test strips for quantitative on-site detection of gasoline fuel adulteration

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I. Materials, methods and instrumentation

Kerosene and absolute ethanol (max 5 ppm carbonyl compounds) were commercially available at Aldrich and gasoline E5 and E10 grade were obtained from HEM, Shell and Esso gas stations in Berlin, Germany. The pre-treatment to remove fluorescent interferences followed either a laboratory-based protocol in which 10 wt% of active charcoal was suspended in fresh mixtures, stirred for 1 h, centrifuged and filtered to remove the charcoal, or an embedded protocol using a stainless steel in-line filter holder (47 mm, PALL) with active carbon paper filters (typically 4).^[1]

Reagents for synthetic procedures were obtained from commercial suppliers and used without further purification, unless indicated. Air- and moisture-sensitive reactions were carried out with previously dried materials under N₂ atmosphere. TLC experiments were performed over Merck Silica Gel 60 F₂₅₄ TLC. Reactions were monitored using a 254 nm hand-held lamp. Column chromatography was carried out with Merck Silica gel 60 (0.040–0.063 mm). NMR spectra were recorded on a 400 MHz (101 MHz for ¹³C) Bruker AV 400 spectrometer at 300 K using residual protonated solvent signals as internal standard (¹H: δ (CDCl₃) = 7.26 ppm and ¹³C: δ (CDCl₃) = 77.16 ppm). Assignments are based on chemical shifts. Mass spectra were measured on a Waters LCT Premier XE.

UV-vis absorption spectra were recorded on a Specord 210-Plus spectrophotometer from Analytik Jena. Steady-state fluorescence measurements were carried out on a FluoroMax-4 spectrofluorometer from Horiba Jobin-Yvon, using standard 10 mm path length quartz cells. All the solvents employed for the spectroscopic measurements were of UV spectroscopic grade (Aldrich).

The smartphone application ("App") was developed using Android Studio 2.0 and Android SDK tools API 22 (Android 5.1). The phone camera was set with a shutter speed of 1/1000 s, a white balance at 5000 K, a Macro Focus and ISO value of 1600 (kerosene) or 64 (ethanol).

¹ Gotor, R.; Tiebe, C.; Schlischka, J.; Bell, J.; Rurack, K. Energy & Fuels 2017, 31 (11), 11594-11600.

II. Procedures

4-((4-(Diphenylamino)phenyl)ethynyl)benzonitrile 1

Compound 1 was synthesized according a reported protocol with a 63 % yield.^[2]

4-Bromo-N-(4-ethynylphenyl)-N-phenylaniline 3.

1 g (2.5 mmol) of 4.4'-Dibromotriphenvlamine and 346 µL (2.5 mmol) of ethynyltrimethylsilane were dissolved in a 20 mL mixture of dry THF and Et₃N 1:1 v/v. The solution was deoxygenated by sparkling Ar(g) for 10 min. After that time, 87 mg of bis(triphenylphosphine)palladium(II) dichloride (0.13 mmol) and 47 mg copper(I) iodide (0.25 mmol) were added to the mixture and the reaction was heated for 3 h at 50 °C, before increasing to 80°C and keeping it overnight. Afterwards, the mixture was allowed to cool down, saturated NH₄Cl solution was added, and the products were extracted three times using CH_2Cl_2 . The organic layer was washed with a saturated NaCl solution and dried with Na₂SO₄. 456 mg of an oily mixture were obtained. Consequently, 400 mg (0.98 mmol) of this product and 817 mg (5.9 mmol) of potassium carbonate were mixed in a 20 mL mixture of CH₂Cl₂ and MeOH 1:1 v/v. The reaction was kept for 1 h. Thereafter, saturated NH₄Cl solution was added, and the products were extracted three times using CH_2Cl_2 . The organic layer was washed with saturated NaCl solution and dried with Na₂SO₄. A column chromatography (PET:CH₂Cl₂, 95:5 v/v) yielded 232 mg (30 %) of the desired product **3** after two steps. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.37 – 7.31 (m, 4H, Ar-H), 7.30 – 7.23 (m, 3H, Ar-H), 7.10 – 7.04 (m, 3H, Ar-H), 6.95 (d, J = 8.7 Hz, 4H, Ar-H), 3.01 (s, 1H, =CH). ¹³C NMR (101 MHz, CDCl₃) δ 147.8, 146.7, 146.3, 133.2, 132.4, 129.6, 126.0, 125.1, 122.5, 115.9, 115.5, 83.7, 83.7, HR-MS (ESI+): m/z calculated for C₂₀H₁₅BrN [M+H]⁺: 348.0388, found: 348.0397.

4-((4-((4-Bromophenyl)(phenyl)amino)phenyl)ethynyl)benzonitrile 4.

127 mg (0.55 mmol) of 4-Iodobenzonitrile was dissolved in a 5 mL mixture of dry THF and Et₃N 1:1 v/v. The mixture was flushed with Ar(g) for 10 min and afterwards 20 mg of bis(triphenylphosphine)-palladium(II) dichloride (28 μmol) and 10 mg copper(I) iodide (56 μmol) were added to the mixture, followed by 200 mg (0.57 mmol) of **3.** The reaction was kept at room temperature overnight. Saturated NH₄Cl solution was then added, and the products were extracted three times using CH₂Cl₂. The organic layer was washed with saturated NaCl solution and dried with Na₂SO₄. A column chromatography (PET:CH₂Cl₂, 8:2 v/v) yielded 167 mg (65 %) of the desired product **4.** ¹H NMR (400 MHz, Chloroform-*d*) δ 7.62 (d, *J* = 8.7 Hz, 2H, Ar-H), 7.57 (d, *J* = 8.8 Hz, 2H, Ar-H), 7.41 – 7.35 (m, 4H, Ar-H), 7.30 (dd, *J* = 8.7, 7.3 Hz, 2H, Ar-H), 7.14 – 7.08 (m, 3H, Ar-H), 7.03 – 6.97 (m, 4H, Ar-H). ¹³C NMR (101 MHz, CDCl₃) δ 148.2, 146.5, 146.1, 132.9, 132.5, 132.0, 131.8, 129.6, 128.5, 126.3, 125.3, 124.3, 122.2, 118.6, 116.3, 115.3, 111.1, 94.2, 87.4. HR-MS (ESI+): *m/z* calculated for C₂₇H₁₈BrN₂ [M+H]⁺: 449.0653, found: 449.0643.

² Piao, M. J.; Chajara, K.; Yoon, S. J.; Kim, H. M.; Jeon, S.-J.; Kim, T.-H.; Song, K.; Asselberghs, I.; Persoons, A.; Clays, K.; Cho, B. R. *Journal of Materials Chemistry* **2006**, *16* (23), 2273-2281.

4-((4-((4-(5-Hydroxypent-1-yn-1-yl)phenyl)(phenyl)amino)phenyl)ethynyl)benzonitrile 5.

100 mg (0.22 mmol) of **4** and was dissolved in a 5 mL mixture of dry THF and Et₃N 1:1 v/v. The solution was deoxygenated by sparkling Ar(g) for 10 min. Thereafter, 15 mg of bis(triphenylphosphine)-palladium(II) dichloride (0.02 mmol) and 8 mg copper(I) iodide (0.04 mmol) were added to the mixture and allowed to react. Then, 30 μ L (0.33 mmol) of 4-pentyn-1-ol was added and the mixture was heated overnight at 70 °C. Afterwards, the mixture was cooled down and the mixture was filtered through Celite and then saturated NH₄Cl solution was added. The products were extracted three times using CH₂Cl₂. The organic layer was washed with saturated NaCl solution and dried with Na₂SO₄. An isocratic column chromatography (cyclohexane:EtOAc, 8:2 to 7:3 v/v) yielded 78 mg (78 %) of the desired product **5**. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.55 (d, *J* = 8.4 Hz, 2H, Ar-H), 7.50 (d, *J* = 8.3 Hz, 2H, Ar-H), 7.31 (d, *J* = 8.7 Hz, 2H, Ar-H), 7.26 – 7.19 (m, 4H, Ar-H), 7.04 (dd, *J* = 6.2, 2.7 Hz, 3H, Ar-H), 6.94 (dd, *J* = 8.7, 2.8 Hz, 4H, Ar-H), 3.76 (t, *J* = 6.1 Hz, 2H, ≡C-CH₂-CH₂-OH), 2.47 (t, *J* = 7.0 Hz, 2H, ≡C-CH₂-CH₂-OH), 1.79 (p, *J* = 6.5 Hz, 2H, ≡C-CH₂-CH₂-OH). ¹³C NMR (101 MHz, CDCl₃) δ 148.2, 146.6, 146.4, 132.9, 132.7, 132.0, 131.9, 129.6, 128.6, 125.5, 124.3, 124.1, 122.5, 118.6, 118.4, 115.3, 111.1, 94.3, 88.3, 87.4, 80.9, 61.9, 31.5, 16.1. HR-MS (ESI+): *m/z* calculated for C₃₂H₂₅N₂O [M+H]⁺: 453.1967, found: 453.1975.

4-((5-(4-((4-((4-Cyanophenyl)ethynyl)phenyl)(phenyl)amino)phenyl)pent-4-yn-1-yl)oxy)-4-oxobutanoic acid 2.

20 mg of **5** (0.04 mmol), 0.53 mg of 4-dimethylaminopyridine (4.4 µmol) and 5 mg of succinic anhydride (0.05 mmol) were dissolved in 1.6 mL of CH₂Cl₂. To this mixture, 3.2 µL of Et₃N (0.05 mmol) were added, and the reaction was kept overnight at room temperature. Then, 1 mL of water was added, and the pH was adjusted to 2 with AcOH. The mixture was washed two times with water and a saturated NaCl solution. Next, it was purified using column chromatography (PET:EtOAc 3:7 v/v) yielding 21 mg (86 %) of the desired dye **2**. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.54 (d, *J* = 8.3 Hz, 2H, Ar-H), 7.49 (d, *J* = 8.4 Hz, 2H, Ar-H), 7.31 (d, *J* = 8.7 Hz, 2H, Ar-H), 7.24 – 7.19 (m, 4H, Ar-H), 7.06 – 7.00 (m, 3H, Ar-H), 6.93 (dd, *J* = 8.7, 4.2 Hz, 4H, Ar-H), 4.19 (t, *J* = 6.3 Hz, 2H, ≡C-CH₂-CH₂-O-), 2.64 – 2.55 (m, 4H, -CO-CH₂-CH₂-COOH), 2.43 (t, *J* = 7.0 Hz, 2H, ≡C-CH₂-CH₂-CH₂-O-), 1.86 (p, *J* = 6.7 Hz, 2H, ≡C-CH₂-C

III. Photophysical properties

Dyes	1				2			
Solvent	λ _{abs} / nm	λ_{em} / nm	Stokes shift / cm ⁻¹	$\Phi_{ m f}$	$\begin{array}{ccc} \lambda_{abs} & \lambda_{em} & Stokes shift \\ / nm & / nm & / cm^{-1} \end{array} \Phi_{f} \end{array}$			
n-Hexane	380	401	1378	0.66	382 406 1547 0.45			
Toluene	381	432	3099	0.95	385 436 3038 0.790			
CHCl ₃	384	472	4855	0.99	386 475 4854 0.99			
EtOH	375	515	7249	0.78	379 521 7191 0.57			
MeCN	372	543	8466	0.58	375 549 8452 0.37			
Gasoline	379	430	3129	0.77	387 435 2851 0.69			
Kerosene	384	406	1411	0.706	388 411 1442 0.714			

Tab. S1 Photophysical properties of dyes 1 and 2 in various solvents and gasoline.



Fig. S2 Normalized absorption of (a) 2 and (b) 1 and normalized emission of (c) 1 in various solvents ([1] = [2] = 4 μ M, λ_{exc} = 340 nm)

IV. Test strip preparation and analyses

Preparation of M1 test strips

Whatman 1 paper filter (1×4 cm, 20 pieces) were added to a 100 mL toluene solution containing 2.5 mL of APTES and the mixture was agitated at 90 °C for 24 h. After that time the strips were washed thoroughly with EtOH and dried in air. Then, 10 mL of a $4 \cdot 10^{-4}$ M solution of **2** in CH₂Cl₂ and 0.8 mg (4 µmol) of DCC were mixed for 30 min. After that time, 280 mg of aminated Whatman paper 1 and 0.5 µL of Et₃N (4 µmol) were added to the mixture, and the reaction was kept for additional 2 h. The paper was washed several times with CH₂Cl₂ and then with EtOH and left to dry in air. Differential absorption analysis of the initial solution and final solutions, together with the washing fractions, revealed that the dye was grafted to the paper at a concentration of 3.3 µmol·g⁻¹_{paper}.

Preparation of M2' and M2 test strips

A mixture of 15 mL toluene and 1 mL APTES was added to 15 glass fiber test strips (Whatman glass fiber paper, 0.5×2.5 cm, 0.3 g). The reaction proceeded over 24 h at room temperature with constant stirring. The strips were then removed and washed 3 times with ethanol (3×15 mL) and dried in air for 15 min. Then, 10 mL of a 4.10^{-4} M solution of **2** in CH₂Cl₂ and 0.8 mg (4 µmol) of DCC were mixed for 30 min at room temperature. The test strips and 0.25 µL of triethylamine were added and stirred for 2 h at room temperature. The strips were then removed and washed 2 times with dichloromethane (2×10 mL) and 2 times with ethanol (2×10 mL) and dried in air for 15 min, yielding **M2'**. For the passivated strips **M2**, a mixture of 10 mL hexane and 1.5 mL HMDS was added to strips **M2'**. The reaction proceeded over 24 h at room temperature with constant stirring. Strips **M2** were then removed and washed 2 times with acetone (2×10 mL) and 2 times with ethanol (2×10 mL) and dried in air for 15 mI, yielding **M2'**. The reaction proceeded over 24 h at room temperature with constant stirring. Strips **M2** were then removed and washed 2 times with acetone (2×10 mL) and 2 times with ethanol (2×10 mL) and dried in air for 15 mI.)

Preparation of silica coated cellulose strips^[3,4]

A mixture (7 mL) with pre-determined proportion of TEOS, distilled water, alcohol and catalyst (see table S2) was added to 3 cellulose test strips (Whatman 1 paper filter, 0.5×5 cm, 0.09 g). The reaction proceeded for 24 h at room temperature with constant stirring. The final hybrid material was removed and washed 3 times with ethanol (3×5 mL) and dried under reduced pressure. The percentage of silica incorporation was determined by weighing the hybrid material.

³ Sequeira, S.; Evtuguin, D. V.; Portugal, I.; Esculcas, A. P. Materials Science and Engineering: C 2007, 27, 172.

⁴ Cai, J.; Liu, S.; Feng, J.; Kimura, S.; Wada, M.; Kuga, S.; Zhang, L. *Angewandte Chemie International Edition* **2012**, *51*, 2076.

Preparation of M3' and M3 test strips

A mixture of 15 mL toluene and 1 mL APTES was added to 7 silica coated cellulose test strips (0.5×2.5 cm, 0.09 g) prepared following the entries #6 and #10 from Table S2. The reaction proceeded over 24 h at room temperature with constant stirring. The strips were then removed and washed 3 times with ethanol (3×15 mL) and dried in air for 15 min. Then, 10 mL of a 4.10^{-4} M solution of **2** in CH₂Cl₂ and 0.82 mg (4 µmol) of DCC were mixed for 30 min at room temperature. The test strips and 0.3 µL of triethylamine were added and stirred for 2 h at room temperature. The strips were then removed and washed 2 times with dichloromethane (2×10 mL) and 2 times with ethanol (2×10 mL) and dried in air for 15 min, yielding M3'. For the passivated strips M3, a mixture of 10 mL hexane and 1.5 mL HMDS was added to M3' strips. The reaction proceeded over 24 h at room temperature with constant stirring. Strips M3 were then removed and washed 2 times with acetone (2×10 mL) and 2 times with ethanol (2×10 mL) and dried in air for 15 min. Differential absorption analysis of the initial solutions and final solutions after grafting of **2** revealed that the dye was grafted to M3' at concentrations of 9 µmol·g⁻¹_{paper} for strips prepared following entry #6 and 21 µmol·g⁻¹_{paper} for strips prepared following entry #10.



Fig. S3. Infrared absorption spectra acquired on a BRUKER-Equinox 55, IR-Scope with ATR-Objective of the cellulose without coating (a) and silica-coated cellulose strip obtained from entries #4 (b), #6 (c) and #10 (d) of Table S2. The bands at 800 and 1080 cm⁻¹ are associated with rocking, bending (or symmetric stretching) and asymmetric stretching of SiO₂ inter-tetrahedral oxygen atoms. The band at 1080 cm⁻¹ overlaps with a broad band between 1000 and 1150 cm⁻¹ attributed to O–H bending of primary and secondary alcohol groups of cellulose. The presence of cyclic silica (SiO)₆ structures was confirmed as revealed from the characteristic band at 1220 cm⁻¹.



Fig. S4 Pictures upon UV excitation of the dried M3 test strips obtained from silica-coated cellulose strips prepared following entries #6 (a) and #10 (b) of table S2 ($\lambda_{exc} = 365$ nm).

Tab. S2 Preparation and analyses of the silica coated cellulose strips and of the sensor	y test strips M1, M2',
M2, M3' and M3.	

#	Water	Ethanol	TEOS	HNO ₃	NH ₃	Weight	Strain	$\lambda_{em}(max)$ for		
				0.1111	32 /0	merease		Kerosene	Gasoline	Ethanol
	mL	mL	mL	μL	μL	%	%	nm	nm	nm
1	1.8	4.3	0.7	210	-	-10.0	4.5	-	-	-
2	2.0	4.3	0.7	21	-	9.8	3.6	-	-	-
3	2.0	4.3	0.7	2	-	-1.5	3.1	-	-	-
4	2.0	4.5	0.3	210	-	-12.4	3.6	-	-	-
5	1.6	4.1	1.3	210	-	-1.6	4.1	-	-	-
6	1.8	3.9	1.1	-	226	23.8	3.0	-	-	-
7	1.9	3.9	1.1	-	90	13.4	3.6	-	-	-
8	2.0	3.9	1.1	-	9	-0.7	3.2	-	-	-
9	2.1	4.2	0.5	-	90	5.3	3.7	-	-	-
10	1.7	3.7	1.5	-	90	26.6	2.7	-	-	-
11	1,4	3,9	1,1	-	630	13.9	4.0	-	-	-
12	1,6	3,9	1,1	-	420	15.8	3.4	-	-	-
13	2,0	2,7	2,5	-	90	8.8	3.9	-	-	-
14	2,0	3,2	2,0	-	90	19.7	3.3	-	-	-
Cellulose	-	-	-	-	-	-	3.9	-	-	-
Glass fiber	-	-	-	-	-	-	3.1	-	-	-
M1	-	-	-	-	-	-	7.1	472	475	515
M2′	-	-	-	-	-	-	1.6	455	468	506
M2	-	-	-	-	-	-	2.6	431	468	513
M3′	-	-	-	-	-	-	4.6	470	470	515
M3	-	-	-	-	-	-	7.5	431	470	515



V. Sensing of gasoline/kerosene and gasoline/ethanol mixtures

Fig. S5 Emission spectra of (a) 1 in decane/ethanol mixtures and (b) 2 in gasoline/kerosene and gasoline/ethanol mixtures with arrow signaling gasoline spectra and steps of 10 vol% ($[1] = [2] = 4 \mu M$, $\lambda_{exc} = 340$ nm). (c) Scheme representing the sensing mechanism that occurs inside the fluorometric test strips (i.e. M3) upon solvation in various type of fuels. The solvation of the charge transfer dyes 1 or 2, which have only moderate dipole moments in the ground state but very pronounced dipole moments in the excited state, leads to solvent relaxation upon Franck-Condon excitation.^[5] The more polar the solvent is the higher the stabilization of the excited and the higher the destabilization of the ground state of the two species involved in the vertical emission process, hence the larger the red-shift of the emission spectrum.

⁵ B. Valeur, *Molecular Fluorescence: Principles and Applications*, **2001** Wiley-VCH Verlag GmbH.



Fig. S6 Successive steps to analyze the purity of a gasoline fuel: dip in sample, place in the holder, start the LED, place the holder and press the shoot button to obtain directly the gasoline purity on screen.

Tab. S3 Hue values of **M3** after dipping in kerosene, gasoline E5 or ethanol upon various ISO parameters with phone camera settings of 1/1000 s for the shutter speed, 5000 K for white balance and a Macro Focus.

Fuel	ISO 64	ISO 1600
Kerosene	239.2 ± 1.0	224.2 ± 0.8
Gasoline E5	239.0 ± 1.0	220.8 ± 0.4
Ethanol	199.4 ± 0.8	186.0 ± 0.9



Fig. S7 (a) 520/480 nm fluorescence intensity ratio vs time of **M1** test-strip wetted with a drop (approx. 0.2 mL) of 2:8 gasoline/ethanol mixtures ($\lambda_{exc} = 340$ nm). Fluorescence signals of the test strips upon UV excitation ($\lambda_{exc} = 365$ nm) were monitored through an optical fiber placed below the strip coupled to an Ocean Optics USB2000+ spectrometer. (b) Hue variation over time after dipping of **M3** test-strip in gasoline, kerosene or ethanol ($\lambda_{exc} = 380$ nm).



Fig. S8 (a) Emission maxima of **M1** test-strip after addition of 30 μ L of gasoline (blue) and ethanol (green) for ten cycles ($\lambda_{exc} = 340$ nm). (b) Hue value upon successive dipping of **M3** test-strip in gasoline (G), kerosene (K) and ethanol (E) ($\lambda_{exc} = 380$ nm). (c) Stability of the test strips upon long-term storage at room-temperature in a dark and dry place.



Fig. S9 Respective Green and Blue channels signals measured from M3 test-strips when dipped in (a) gasoline E5/kerosene and (b) gasoline E5/ethanol mixtures ($\lambda_{exc} = 380$ nm).



Fig. S10 Emission spectra of various dry strips (cellulose, glass fiber paper (GF), TEOS coated cellulose type #6 and #10, $M3_{#6}$ and $M3_{#10}$) obtained from replacing the smartphone detection by an optical fibre to a USB spectrometer (STS-UV Ocean Optics), ($\lambda_{exc} = 380$ nm).

Tab. S4 Hue values of **M3** after dipping in kerosene/gasoline E5/ethanol mixtures recorded with two different ISO parameters and phone camera settings of 1/1000 s for the shutter speed, 5000 K for white balance and a Macro Focus.

Fu	els proportion	i / %	Acmost	Hue / °		
Kerosene	Gasoline	Ethanol	Aspect	ISO 64	ISO 1600	
33	33	33	Biphasic	n.m.	n.m.	
10	50	40	Clear	233.6	194.5	
40	50	10	Turbid	235.9	204.5	
n.m.: not measured						