Supporting Information

Real-time Monitoring of Peroxynitrite (ONOO⁻) in the Rat Brain by Developing a Ratiometric Electrochemical Biosensor

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Contents

1. Developed ratiometric electrochemical for real-time monitoring of ONOO⁻

(Figure S1)

- 2. Synthetic route of HEMF (Figure S2)
- 3. ¹H NMR and ¹³C NMR of Compound 2 (Figure S3)
- 4. ¹H NMR, ¹³C NMR and Mass spectrum of Compound 3 (Figure S4)
- 5. ¹H NMR, ¹³C NMR, Mass spectrum and FTIR spectrum of HEMF (Figure S5)

6. Comparison of DPVs obtained at CFME/Au/MB+HEMF and Au/MB+HEMF electrodes (Figure S6)

7. XPS characterization of electrode modification processes (Figure S7)

8. Liquid chromatography-mass spectrum of ONOO⁻ reacted with HEMF (Figure S8)

9. The reaction rate of CFME/Au/MB+HEMF electrode toward different concentrations of ONOO⁻ (Figure S9)

10. DPVs obtained at CFME/Au/MB+HEMF electrode in aCSF (pH 7.4) with addition of different concentrations of ONOO⁻ (Figure S10)

11. Stability of CFME/Au/MB+HEMF electrode (Figure S11 - S13)

12. Reproducibility test for CFME/Au/MB+HEMF electrodes (Figure S14)

13. MTT assay (Figure S15)

14. Apoptosis assay (Figure S16)

1. Developed Ratiometric Electrochemical for Real-Time Monitoring of ONOO⁻



Figure S1. Developed Ratiometric Electrochemical for Real-Time Monitoring of ONOO⁻.

2. Synthetic route of HEMF

Synthesis of HEMF. 2-(4-Diethylamino-2-hydroxybenzoyl) benzoic acid (313.0 mg, 1.0 mmol) and 4'-piperazinoacetophenone (204.0 mg, 1.0 mmol) in methanesulfonic acid (8.0 mL) were stirred at 100 °C for 4 h at Ar atmosphere. After 4 h, the reaction product was cooled to room temperature. Then the mixture was extracted with dichloromethane (CH2Cl2) three times. The organic layers were collected, and then dried over Na₂SO₄. Finally, the mixture was evaporated under reduced pressure, and then purified by preparative TLC (CH₂Cl₂/EtOH=30:1) to offer compound 2 as a red solid (412.9 mg, 85.6%). A mixture of ferrocenecarboxylic (115.0 mg, 0.5 mmol), HATU (190.1 mg, 0.5 mmol) and triethylamine (20 µL) in 10.0 mL anhydrous CH₂Cl₂ was stirred for 2 h, then compound 2 (241.2 mg, 0.5 mmol) was added into the mixture under Ar atmosphere. The mixture was stirred at room temperature for 12 h. Then, the solvent was removed under vacuum condition, and the mixture was purified by preparative TLC (CH₂Cl₂/EtOH=40:1) to offer compound 3 as a blue solid (194.4 mg, 56.0%). A mixture of the corresponding compound 3 (173.6 mg, 0.25 mmol), 1,6-hexanedithiol (37.6 mg, 0.25 mmol), EDC (47.9 mg, 0.25 mmol), DMAP (12.2 mg,0.1 mmol) was added CH₂Cl₂ (20.0 ml) at room temperature. The mixture was stirred for 12 h. Then solvent was removed under reduced pressure and the mixture was purified by preparative TLC (CH₂Cl₂/EtOH=40:1, CH₂Cl₂/EA/EtOH=6:3:1) to offer HEMF as a blue solid (141.3 mg, 68.4%).



Figure S2. Synthetic route of HEMF.

3. ¹H NMR and ¹³C NMR of Compound 2



¹H NMR (500 MHz, CD₃OD) δ 8.30 (d, 3H), 7.85 (t, 1H), 7.80 – 7.76 (m, 2H), 7.51 (d, 1H), 7.33 (d, 1H), 7.27 - 7.22 (m, 4H), 3.83 (t, 4H), 3.73 (m, 4H), 3.43 (t, 4H), 1.35 (t, 6H), 1.30 (s, 1H).



¹³C NMR (126 MHz, CD₃OD) δ 173.1, 165.9, 163.6, 158.5, 154.9, 134.4, 130.2, 130.0, 129.7, 129.1, 128.6, 115.8, 115.3, 113.8, 109.0, 95.8, 45.2, 44.0, 38.1, 31.6, 29.3, 29.0, 22.3, 13.0, 11.4.

Figure S3. ¹H NMR and ¹³C NMR of Compound 2.

4. ¹H NMR, ¹³C NMR and Mass spectrum of Compound 3



¹H NMR (500 MHz, CD₃OD) δ 8.22 (d, 1H), 8.00 (m, 1H), 7.68 (s, 1H), 7.65 - 7.60 (m, 2H), 7.48 (d, 1H), 7.42 - 7.40 (m, 1H), 7.18 - 7.12 (m, 4H), 4.71 (m, 2H), 4.47 (m, 2H), 4.29 (d, 5H), 4.00 (s, 4H), 3.70 - 3.66 (dd, 8H), 8.22 (d, 1H), 3.33 (m, 4H), 1.30 (t, 6H).



¹³C NMR (126 MHz, CD₃OD) δ 171.3, 166.2, 163.5, 158.5, 154.9, 154.5, 134.4, 130.3, 129.7, 129.2, 128.6, 117.7, 115.7, 115.3, 113.4, 109.0, 95.7, 76.3, 70.3, 69.8, 69.5, 45.2, 11.3.



ESI-MS C₄₁H₄₀FeN₃O₄⁺ [M⁺], found 694.2353.

Figure S4. ¹H NMR, ¹³C NMR and Mass spectrum of Compound 3.

5. ¹H NMR, ¹³C NMR, mass spectra and FTIR spectrum of HEMF



¹H NMR (500 MHz, DMSO) δ 8.33 (d, 2H), 8.09 (d, 1H), 7.94 - 7.81(m, 3H), 7.61 (d, 1H), 7.35 (s, 1H), 7.19 - 7.12 (m, 4H), 4.61 (d, 2H), 4.42 (s, 2H), 4.28 (s, 5H), 3.91 (s, 4H), 3.74 - 3.63 (d, 8H), 2.83 (s, 2H), 2.63 - 2.57 (m, 2H), 2.43 - 2.35 (m, 2H), 2.12 (m, 1H), 1.55 (s, 2H), 1.31 (t, 6H), 1.14 (s, 4H).



¹³C NMR (126 MHz, DMSO) δ 192.5, 169.1, 166.2, 160.2, 158.0, 154.8, 154.8, 137.4, 131.2, 129.0, 116.7, 114.8, 113.8, 109.6, 96.8, 77.7, 72.3, 70.7, 70.6, 69.9, 68.9, 46.2, 45.5, 38.1, 33.6, 33.5, 29.3, 28.8, 28.3, 27.9, 27.8, 27.7, 27.6, 27.6, 27.5, 24.1, 24.1, 12.9.



ESI-MS C₄₇H₅₂FeN₃O₃S₂⁺ [M⁺], found 826.2796.



FTIR spectrum of HEMF.

Figure S5. ¹H NMR, ¹³C NMR, Mass spectrum and FTIR spectrum of HEMF.

6. Comparison of DPVs obtained at CFME/Au/MB+HEMF and Au/MB+HEMF electrode



Figure S6. DPVs obtained at (a) CFME/Au/MB+HEMF, (b) Au/MB+HEMF electrode in 0.1 M PBS (pH 7.4) at scan rate of 100 mV s⁻¹.

7. XPS characterization of electrode modification processes



Figure S7. XPS spectra of (A) Au $4f_{7/2}$ and Au $4f_{5/2}$, (B) P 2p, (C) S 2p and (D) Fe $2p_{3/2}$ and Fe $2p_{1/2}$ for (a) CFME, (b) CFME/Au, (c) CFME/Au/MB and (d) CFME/Au/MB+HEMF electrode.

8. Liquid chromatography-mass spectrum of ONOO⁻ reacted with HEMF



Figure S8. Liquid chromatography-mass spectrum of ONOO⁻ reacted with HEMF.

9. The response time of CFME/Au/MB+HEMF electrode toward different concentrations of ONOO⁻



Figure S9. The relationship between $j_p/j_{p(R)}$ value and time obtained at CFME/Au/MB+HEMF electrode with addition of different concentrations of ONOO⁻ (a: 1.0 μ M; b: 1.5 μ M, c: 2.0 μ M) in 0.1 M PBS (pH 7.4).

10. DPVs obtained at CFME/Au/MB+HEMF electrode in aCSF (pH

7.4) with addition of different concentrations of ONOO-



Figure S10. (A) DPVs obtained at CFME/Au/MB+HEMF electrode in aCSF (pH 7.4) with addition of different concentrations of ONOO⁻ (From a to i: 0, 50, 100, 200, 400, 400, 200, 100, 50 nM). (B) The linear relationship of the peak current difference (Δj) obtained at 364 mV vs Ag/AgCl with different concentration differences of ONOO⁻.

(Δj versus $\Delta C_{(ONOO^-)}$), (a, black) the concentration difference of ONOO⁻ from low to high (50 nM to 400 nM), (b, red) the concentration difference of ONOO⁻ from high to low (400 nM to 50 nM).

11. Stability of CFME/Au/MB+HEMF electrode (Figure S11-S13)



Figure S11. DPVs obtained at CFME/Au/MB+HEMF electrode in 0.1 M PBS (pH 7.4) by scanning after (a) the first cycle, (b) 50 cycles, (c) 100 cycles, (d) 250 cycles, and (e) 500 cycles, respectively.



Figure S12. Stability test for CFME/Au/MB+HEMF electrode in 0.1 M PBS (pH 7.4) over 6 days.



Figure S13. The $j_p/j_{p(R)}$ value of 1.0 μ M ONOO⁻ under different pH values (6.6, 7.0, 7.4, 7.8, 8.2).

12. Reproducibility test for CFME/Au/MB+HEMF electrodes



Figure S14. Reproducibility test for different CFME/Au/MB+HEMF electrodes in aCSF (pH 7.4).

13. MTT assay



Figure S15. Viabilities of neurons after incubation for 24 h (black bars) and 48 h (red bars) with different concentrations of (A) HEMF, SH-DNA-MB (B) and (C) ferrocene which may fall into the rat brain after reacted with ONOO⁻.

14. Apoptosis assay



Figure S16. Apoptosis assay of neurons incubated with the HEMF (A, B, C, D), SH-DNA-MB (E, F, G, H) and ferrocene which may fall into the rat brain after reacted with ONOO⁻(I, J, K, L) at concentrations of 0 μ M (A, E, I), 0.01 μ M (B, F, J), 0.1 μ M (C, G, K) and 1.0 μ M (D, H, L). I, II, III, and IV represents the region of normal cells, early apoptotic cells, late apoptotic cells, and dead cells, respectively.