Electronic Supplementary Information

## pH-based immunoassay: explosive generation of hydrogen ions through an

## immuno-triggered nucleic acid exponential amplification reaction

Dongsheng Mao,<sup>a</sup> Tianshu Chen,<sup>a</sup> Huinan Chen,<sup>a</sup> Mengru Zhou,<sup>a</sup> Xingwei Zhai,<sup>a, b</sup>

Guifang Chen,<sup>\* a, c</sup> Xiaoli Zhu<sup>\* a, b</sup>

<sup>a</sup>Center for Molecular Recognition and Biosensing, School of Life Sciences, Shanghai University, Shanghai 200444, China. E-mail: xiaolizhu@shu.edu.cn

<sup>b</sup>CAS Key Lab of Bio-Medical Diagnostics, Suzhou Institute of Biomedical Engineering and Technology, Chinese Academy of Sciences, Suzhou 215163, P.R. China.

<sup>b</sup>Department of Bioengineering, University of Washington, Seattle, WA 98195, USA.

 Table S1. Sequences of oligonucleotides.

Sequences (5'-3')	Modification
GCATCTACCTCAGCATCTACCTCA	3'-C3 Spacer
TGAGGTAGAT <mark>GC<sup>▽</sup>TGAGG</mark> TAGATGC	
TGAGGTAGAT <mark>GC<sup>▽</sup>TGAGG</mark> TAGATGC	5'-biotin
GAAGCTGTTGTAATATCACTGAAA	5'-biotin
	Sequences (5'-3') GCATCTACCTCAGCATCTACCTCA TGAGGTAGATGC <sup>\U22</sup> TGAGGTAGATGC TGAGGTAGATGC <sup>\U22</sup> TGAGGTAGATGC GAAGCTGTTGTAATATCACTGAAA

Note: The red marked bases are the recognition sequence of nicking enzyme and " $\bigtriangledown$ " represents the nicking site.



**Fig. S1** The influence of some factors on pH changes. (a) Formula for calculating the changes of pH. (b) The influence of cycle numbers on the changes of pH (nucleobase = 500 bp, template = 1, volume = 100  $\mu$ L, initial pH = 7). (c) The influence of nucleobase numbers on the changes of pH (template = 1, cycles = 30, volume = 100  $\mu$ L, initial pH = 7). (d) The influence of the concentration of Tris-HCl on the changes of pH (template = 1, cycles = 37, nucleobase = 500 bp, volume = 100  $\mu$ L, initial pH = 8.2, (Effective buffering range of Tris-HCl: 7.0 - 9.2, Ka = 10<sup>-8.1</sup>)).



**Fig. S2** Polyacrylamide gel electrophoretic patterns of EXPAR products with different amplification times.



**Fig. S3** The influence of the purification of C3-Spacer blocked template on unspecific amplification.



**Fig. S4** Log-linear relationship between the concentration of primer ranging from 10 fM to 100 pM and the point of inflection (POI) obtained from the quantification real-time EXPAR (Fig. 1f). (linear regression equation: y=14.402-1.852x,  $R^2=0.980$ ).



**Fig. S5** Polyacrylamide gel electrophoretic patterns of EXPAR products with different primer concentrations.

Sample	Added PDGF-BB	Found PDGF-BB	Recovery (%)	R.S.D.
	(ng/L)	(ng/L)		(%, n=3)
1	20	18.5	93	3
2	4	3.89	97	3
3	0.3	0.311	104	3

**Table S2.** Determination of spiked human PDGF-BB in 10% serum samples.

Sample	Added CAP (nM)	Found CAP (nM)	Recovery (%)	R.S.D.
				(%, n=3)
1	3.71	3.34	90	5
2	0.464	0.432	93	2
3	0.0309	0.0315	102	1

**Table S3.** Determination of spiked CAP in 10% serum samples.