

## Supporting Information

### One-step encapsulation, storage and controlled release of small molecular weight organic compounds *via* electroplated nanoparticles

Y.E. Silina<sup>1,2,3\*</sup>, D. Semenova<sup>4</sup>, B.A. Spiridonov<sup>5</sup>

#### Methods:

H<sub>2</sub>PdCl<sub>4</sub>, (NH<sub>4</sub>)<sub>2</sub>HPO<sub>4</sub> and Na<sub>2</sub>HPO<sub>4</sub>·12H<sub>2</sub>O used in nanoparticles synthesis were of 99 % purity (Sigma Aldrich), HCl (37%, ICP-MS quality) was obtained from Honeywell Fluka (Germany). The MALDI matrices, *viz.*  $\alpha$ -cyano-4-hydroxycinnamic acid (CHCA),  $\alpha$ -cyano-2,3,4,5,6-pentafluorocinnamic acid (FCCA) and polished steel target (MTP 384 type) were purchased from Bruker Daltonik GmbH (Germany). Reserpine was obtained from Sigma-Aldrich (Steinheim, Germany); acetonitrile and methanol were from Merck (Darmstadt, Germany). All other chemicals were purchased from Sigma-Aldrich (97-99% purity). All the solvents were of HPLC quality. Organic-free, deionized water was generated by an Elga PureLab (Celle, Germany) water purification system.

#### *MALDI matrices preparation*

Standard MALDI experiments were performed using the drying droplet method by mixing equal amounts of analyte and matrix solutions (5 mg/mL in 50:50 [v/v] in water: acetonitrile). Analyte solutions (1  $\mu$ L) were spotted directly onto the electroplated targets and spots dried under ambient conditions.

#### *Electroplating experiment*

The synthesis of Pd-NPs with standard MALDI matrices electrochemically bound to the steel target was conducted by electroplating from Pd-polyelectrolyte (pH=4.5-5) with CHCA or FCCA (2500 ppm) at room temperature 20 $\pm$ 2°C, directly over the steel plates. Electrolysis was performed using a VMP 3 (Biologic SAS, Claix, France) potentiostat. The following parameters were used:  $I_k$ , 10 mA/dm<sup>2</sup> to 100 mA/dm<sup>2</sup>,  $t_{el}$  from 30 sec to 3 min, Pd anode (99.9 %).

#### *Scanning Electron Microscopy (SEM)*

SEM images were captured on a Quanta (Hillsboro, OR, USA) 400 FEG system equipped with an EDAX (Mahwah, NJ, USA) Genesis V 6.04 X-ray spectral analysis system, at an accelerating voltage of 10 keV. The image size was 1024x884 pixels.

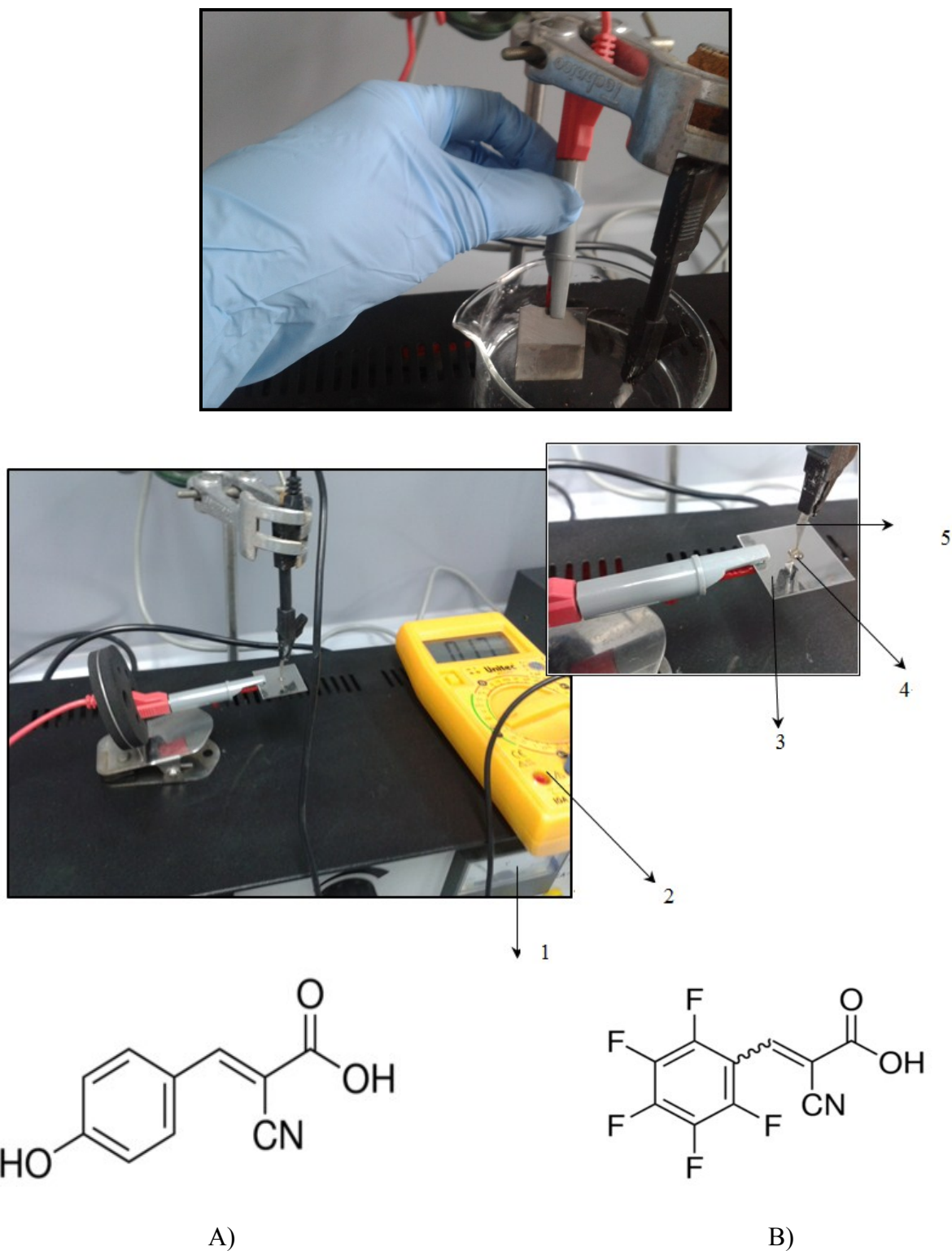
### ***HPLC analysis and Mass Spectrometry***

The HPLC analysis was performed on an Agilent Infinity 1260 series instrument (Agilent Technologies, CA, USA) coupled with a quadrupole time-of-flight mass spectrometer Q-TOF LC/MS 6545 (Agilent Technologies, CA, USA) equipped with Jet Stream Thermal Focusing Technology ESI source. For elution experiment, a droplet of 10  $\mu\text{l}$  of ACN:H<sub>2</sub>O (50:50, v/v) preliminary spotted for 5 min onto the hybrid plates, was taken for subsequent LC-MS analysis. Chromatographic separation was achieved on the ZORBAX Eclipse Plus C18 column (2.1 $\times$ 50 mm, particles size 1.8  $\mu\text{m}$ ) (Agilent, CA, USA). The mobile phase consisting of ACN (Solvent A) and 0.3% formic acid (FA) in water (Solvent B) and was used in the following gradient elution step: 10% Solvent A, was held for 2 min, then increased to 90% in 3 min and held for 2 min, and returned back to the starting conditions in 2 min. The column operation temperature was fixed at 30  $^{\circ}\text{C}$ , the mobile phase flow rate was 300  $\mu\text{l}/\text{min}$  and the injection volume was 1  $\mu\text{L}$ . MS scans were recorded in a negative ion mode, operating under capillary voltage at 4500 V; the fragmentor voltage was set at 30 eV; dry gas temperature at 350  $^{\circ}\text{C}$ ; gas flow at 9 L/min; nebulizing gas pressure at 45 psi. To monitor a leakage of CHCA and FCCA after elution experiment from the hybrid plates, mass spectra at  $m/z$  100-500 were recorded. The data acquisition was controlled by MassHunter Software Tool.

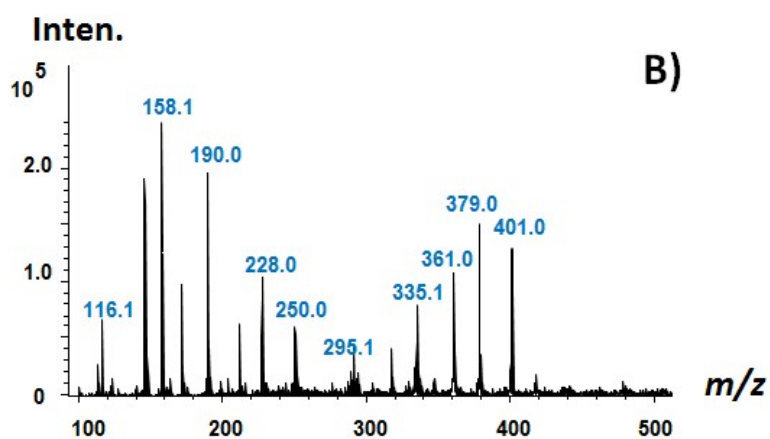
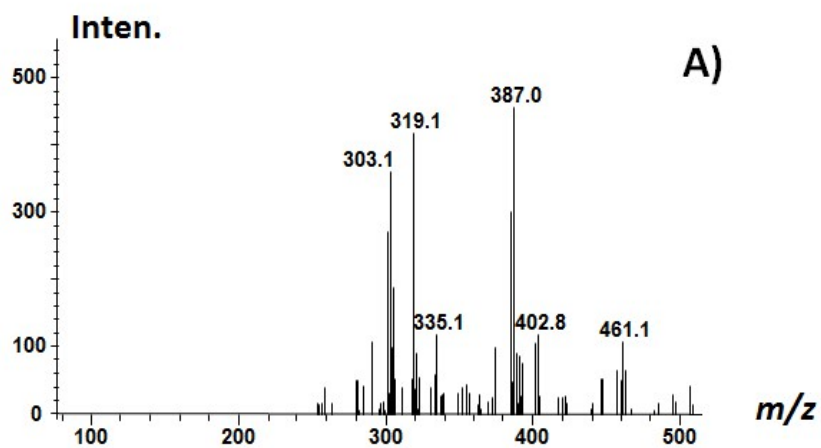
MALDI/LDI experiments were performed on a Bruker Esquire 3000+ESI-ion trap MS (Bruker Daltonics, Bremen, Germany) operated by Bruker esquire control 5.3 software equipped with atmospheric pressure (AP)-MALDI ion source and Nd:YAG solid-state laser (355 nm). Mass spectra were recorded in positive ion full-scan mode from  $m/z$  100-700.

### ***Monitoring co-deposition of small molecular weight organic compounds using QCM system***

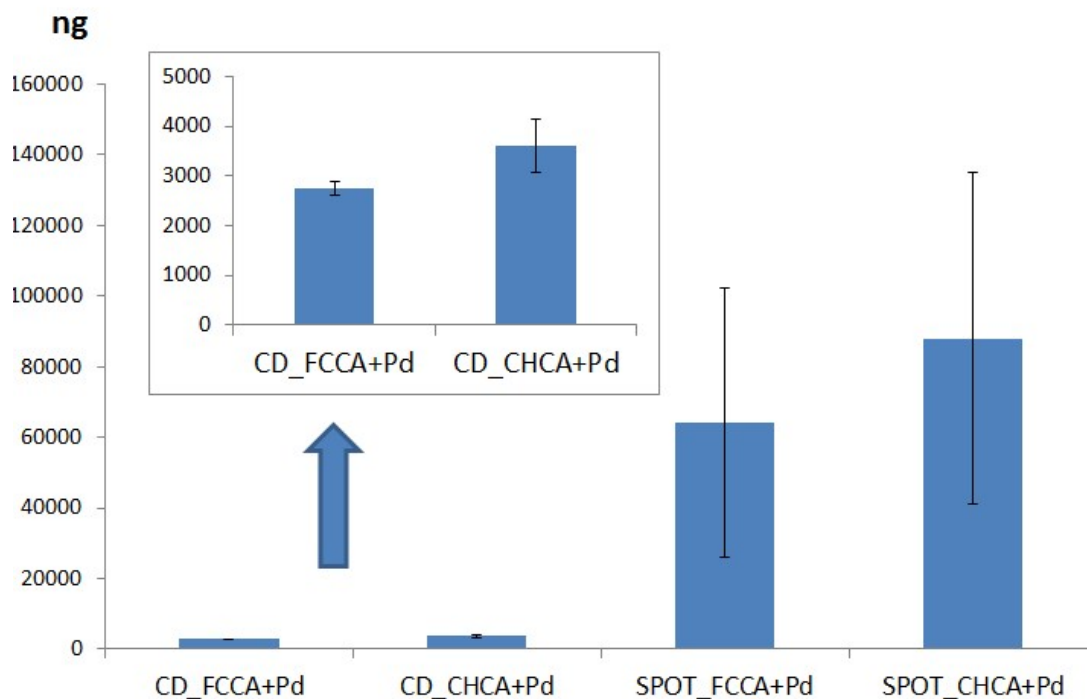
The synthesis reproducibility of hybrid targets were estimated in a droplet by means of quartz crystal microbalance (QCM) measurement system. QCM quartz crystals (AT-cut, diameter 14 mm, surface area 1.54  $\text{cm}^2$ ) with a basic frequency of 10 MHz and software “NanoSens Explorer” were provided by Ltd “Sensors and New technology” (SNT, Voronezh, Russia). The temperature within the experiments was maintained at 20 $\pm$ 2 C. Frequency shifts represent weight of co-deposited organic-inorganic hybrid structures with nanogram precision. The mass of the co-deposited hybrid structures was estimated using *Sauerbrey* equation [22].



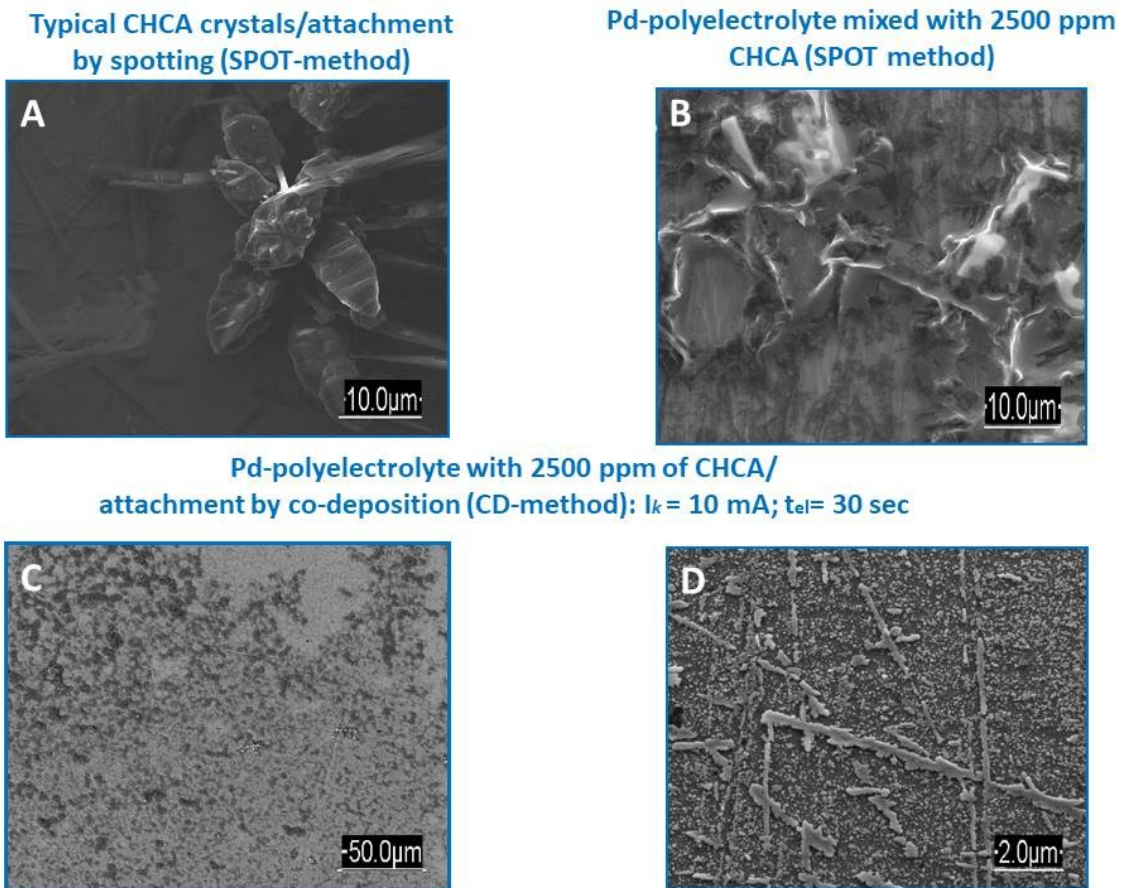
**Figure S1.** (*top*) – laboratory settings for the hybrid MALDI/LDI co-deposition in bulk; (*middle*) – co-deposition in droplet: 1 – potentiostat; 2 – digital multimeter (DMM); 3 – polished steel plate; 4 – 10  $\mu$ l of Pd-electrolyte doped with MALDI matrices; 5 – Pd-wires/electrode; (*bottom*) – chemical structure of the tested MALDI-matrices: A)  $\alpha$ -Cyano-4-hydroxycinnamic acid (CHCA), 189.17 g/mol; B)  $\alpha$ -Cyano-2,3,4,5,6-pentafluorocinnamic acid (FCCA), 263.12 g/mol.



**Figure S2.** TIC-mass spectra obtained in positive detection mode (before pipetting of analytes) from the hybrid MALDI/LDI targets (*shown for encapsulated CHCA as a case study*) prepared at the same deposition time of 30 sec and different currents: **A)** 100 mA: **B)** 10 mA.

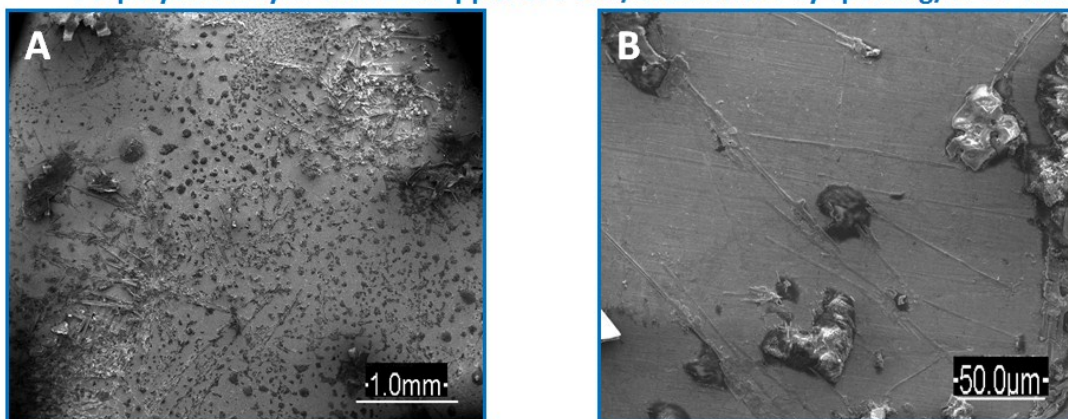


**Figure S3.** Estimation of the deposited rates ( $t = 20 \pm 2$  °C) of MALDI/LDI hybrids in CD and SPOT methods by QCM measurements using *Sauerbrey* model [22]. The bars represent the mean  $\pm$ SD of experiments ( $n = 7$ ). NOTE: 10  $\mu$ l of polyelectrolyte was spotted onto QCM in CD and 1  $\mu$ l in SPOT method to avoid overloading of microbalances.

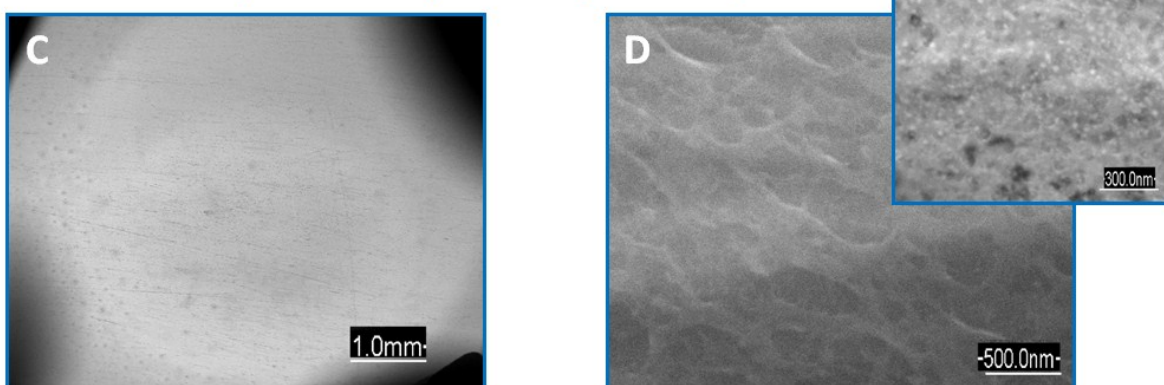


**Figure S4.** SEM images illustrating evolution of CHCA matrix from macro-stage in SPOT-method to nano-stage in CD-method: (A) – pure/spotted CHCA matrix onto the polished steel plate; (B) – spotted mixture of CHCA and Pd-electrolyte onto the steel plate; (C) – hybrid MALDI/LDI target (Pd-NPs with encapsulated CHCA produced by electroplating; CD-method); (D) – the same template as it shown in (C) at different magnification.

Pd-polyelectrolyte with 2500 ppm of FCCA/attachment by spotting/SPOT method



Pd-NPs with encapsulated FCCA (CD method):  $I_k = 10 \text{ mA}$ ;  $t_{el} = 30 \text{ sec}$



**Figure S5.** SEM images of the FCCA spotted with Pd-electrolyte and dried at the ambient conditions (**A**, **B**) and FCCA after encapsulation with Pd-NPs, washed and dried (**C**, **D**) at different magnification (hybrid MALDI/LDI target).