

Electronic Supplementary Material (ESI) for Analysis of BMAA in the Environment
 This journal is © The Royal Society of Chemistry 2019

Year	Reference	BMAA Detected?	BMAA Detected in Cyanobacteria ¹ ?	BMAA Detected in Bioaccumulated Sources ² ?	BMAA Detected in Phytoplankton or Zooplankton ³ ?	BMAA Detected in Human Tissues/Waste ⁴ ?	Chromatography	Column ID	Elution Buffer(s)	Derivative	Detection Method	Detector
2019	100	Yes	N/A	Yes	N/A	N/A	RPLC	Hypersil Gold C18 (2.1 x 150 mm; 1.9 µm)	0.075% acetic acid in water/0.075% acetic acid in acetonitrile	FLEC	MS/MS	Waters Xevo TQ-S
2019	106	Yes	Yes	N/A	N/A	N/A	RPLC	Phenomenex Kinetex (1.7 µm; 100 Å pore size)	0.1% formic acid in 45% water/0.1% formic acid in 55% methanol	PCF	MS/MS	Agilent 6490 Triple Quadrupole

A Systematic Review of Analytical Methods for Detection and Quantification of β-N-methylamino-L-alanine

e (BMAA)

Stephanie L. Bishop¹ (ORCID iD 0000-0002-8134-7297) and Susan J. Murch¹ (ORCID iD 0000-0001-5803-9483)

¹Chemistry, University of British Columbia, Kelowna, British Columbia, Canada, V1V 1V7

Supplementary Tables

Table S1. Comparison of chromatography methods used for detection of BMAA. ¹Includes cyanobacterial cultures grown *in-vitro*, and cyanobacterial samples collected from natural habitats. ²Includes BMAA bioaccumulated in higher trophic level organisms such as traditional Chamorro dietary items such as cycad flour and flying foxes and seafoods such as mussels, lobster, fish and crab. ³Phytoplankton include diatoms and dinoflagellates. ⁴Includes human brain tissue and urine. Studies that did not analyze the specified sample type are labelled N/A. Derivatizing agents include 6-aminoquinoyl-*N*-hydroxysuccinimide (AQC), dansyl chloride (DC), ethyl chloroformate (ECF), (+)-1-(9-fluorenyl)-ethyl chloroformate (FLEC), fluorenylmethyloxycarbonyl (FMOC), isobaric tag for relative and absolute quantitation (iTRAQ), ninhydrin (post-column derivatization), and propyl chloroformate (PCF).

2019	35	Yes	N/A	Yes	N/A	N/A	RPLC	Waters C18 BEH (2.1 x 150 mm; 1.7 µm)	20 mM ammonium formate with 0.2% formic acid/0.1% formic acid in acetonitrile	AQC	MS/MS	Waters Xevo TQ-S
2019	35	Yes	N/A	Yes	N/A	N/A	RPLC	Waters Nova-Pak C18 (3.9 x 300 mm)	140 mM sodium acetate, 5.6 mM triethylamine (pH 5.7)/52% aqueous acetonitrile	AQC	FD	Waters 2475 Multi-k-Fluorescence Detector
2019	107	Yes	Yes	No	N/A	N/A	RPLC	Phenomenex AAA-MS (250 x 2.0 mm; 4 µm)	10 mM Ammonium Formate in water/10 mM ammonium formate in Methanol	PCF	MS/MS	Agilent 1200 Infinity Series
2019	108	Yes	N/A	Yes	N/A	N/A	RPLC	EclipseXDB C-18 (4.6 x 100 mm; 5 µm)	28% acetonitrile/0.1% ammonium formate in water	AQC	MS/MS	Agilent Technologies 1290/6460 MS
2018	25	Yes	No	Yes	N/A	N/A	HILIC	Tosoh TSKgel amide-80 (2.0 x 250 mm; 5 µm)	50 mM formic acid in water/50 mM formic acid in	none	DMS-MS/MS	AB Sciex SelectION (DMS)/AB Sciex Qtrap 5500 (MS)

									acetonitrile			
2018	48	No	N/A	No	N/A	N/A	HILIC	SeQuant ZIC-HILIC (dimensions unspecified)	0.1% formic acid in water/0.1% formic acid in acetonitrile	none	MS/MS	Thermo LTQ XL Linear Ion Trap MS
2018	26	Yes	Yes	No	N/A	No	HILIC	Tosoh TSKgel amide-80 (2.0 x 250 mm; 5 µm)	50 mM formic acid/5% 50 mM formic acid in 95% acetonitrile	none	MS/MS	Waters Xevo TQ-S
2018	47	Yes	N/A	Yes	N/A	N/A	HILIC	Tosoh TSKgel amide-80 (2.0 x 250 mm; 5 µm)	50 mM formic acid in water/50 mM formic acid in 95% acetonitrile	none	MS/MS	Agilent 6430 MS
2018	109	Yes	Yes	Yes	N/A	N/A	RPLC	Waters Acquity BEH C18 (2.1 x 150 mm; 1.7 µm)	20 mM ammonium formate in water (pH 5.0)/methanol	AQC	MS/MS	Waters Xevo TQ-S/Thermo Scientific TSQ Quantiva
2018	26	Yes	Yes	Yes	N/A	No	RPLC	Phenomene x AAA-MS (2.0 x 250 mm; 4 µm)	10 mM ammonium formate in water/10 mM ammonium formate in methanol	PCF	MS/MS	Waters Xevo TQ-S

2018	26	Yes	Yes	Yes	N/A	No	RPLC	Waters Acquity BEH (2.1 x 150 mm; 1.7 µm)	20 mM ammonium formate in water (pH 5.0)/methanol	AQC	MS/MS	Waters Xevo TQ-S
2018	47	Yes	N/A	Yes	N/A	N/A	RPLC	Phenomenex Kinetex (2.1 x 100 mm; 1.7 µm)	20 mM ammonium formate in water (pH 5.0)/methanol	AQC	MS/MS	Agilent 6430 MS
2018	44	Yes	Yes	N/A	N/A	N/A	RPLC	Waters Acquity BEH C18 (2.1 x 100 mm; 1.7 µm)	20 mM ammonium formate in water (pH 5.0)/methanol	AQC	MS/MS	Waters Xevo TQ-S
2018	81	Yes	N/A	N/A	N/A	Yes	RPLC	Waters UltraTag C18 (2.1 x 100 mm; 1.7 µm)	5% acetonitrile in water with 0.3% acetic acid/acetonitrile with 0.3% acetic acid	AQC	MS/MS	Thermo Fisher TQS Vantage Triple Quad
2018	110	Yes	Yes	N/A	N/A	N/A	RPLC	Waters UltraTag C18 (2.1 x 100 mm; 1.7 µm)	0.1% (v/v) formic acid/0.1% (v/v) formic acid in acetonitrile	AQC	MS/MS	Thermo Scientific Finnigan TSQ Quantum UltraAM
2018	111	Yes	Yes	No	N/A	N/A	RPLC	Phenomenex Kinetex C18, 100A (2.1 x 100 mm; 1.7 µm)	20 mM ammonium acetate (pH 5.0)/methanol	AQC	MS	Waters EMD 1000 Single Quad MS

2018	79	No	N/A	N/A	N/A	No	RPLC	Mac-Mod Analytical ACE Excel SuperC18 (2.1 x 100 mm; 2 µm)	0.1% (v/v) heptafluorobutyric acid/0.1% (v/v) heptafluorobutyric acid in methanol	none	MS/MS	Sciex 5500 Triple Quad MS
2018	112	Yes	Yes	N/A	N/A	N/A	RPLC	Phenomenex Kinetex C18 (2.1 x 150 mm; 1.7 µm)	0.1% (v/v) formic acid/0.1% (v/v) formic acid in methanol	PCF	MS/MS	Agilent Technologies 6490 Triple Quad MS
2018	113	Yes	N/A	Yes	Yes	N/A	RPLC	Phenomenex EZ:Faast™ AAA-MS (2 x 250 mm)	10 mM ammonium formate in water/10 mM ammonium formate in methanol	PCF	MS	Shimadzu 2010EV
2017	80	Yes	N/A	N/A	N/A	Yes	HILIC	Waters Atlantis HILIC SILICA (2.1 x 100 mm; 3 µm)	50 mM formic acid in water/50 mM formic acid in 95% acetonitrile	none	MS/MS	Waters Xevo TQ-S
2017	90	Yes	Yes	N/A	N/A	N/A	HILIC	SeQuant ZIC-HILIC (2.1 x 100 mm; 5 µm); SeQuant (1 x 14 mm; 5 µm) guard column	0.1% formic acid in acetonitrile/0.1% formic acid in water	none	MS	Thermo LCQ Fleet Ion Trap
2017	114	No	No	N/A	N/A	N/A	HILIC	SeQuant ZIC-HILIC (2.1 x 150 mm; 5 µm)	0.1% formic acid in acetonitrile/0.1% formic acid in water	none	MS	Agilent Technologies 6490 Triple Quad MS

2017	115	Yes	N/A	Yes	N/A	N/A	RPLC	Waters UltraTag C18 (2.1 mm x 100 mm; 1.7 µm)	0.01% formic acid in 0.05% ammonia in water/0.01 % formic acid in methanol	AQC	MS/MS	Waters Xevo TQ-S
2017	116	Yes	N/A	N/A	N/A	N/A	RPLC	Phenomene x Chirex 3126 chiral column (4.6 x 250mm); Waters AccQ Tag Ultra (2.1 x 100 mm)	2 mM copper sulfate in water/2 mM copper sulfate in 15% aq. acetonitrile; 0.1% formic acid in water/0.1% formic acid in acetonitrile	AQC	MS/MS	Thermo Scientific Finnigan TSQ Quantum UltraAM
2017	117	Yes	Yes	N/A	N/A	N/A	RPLC	Thermo Hypesil GOLD C18 (2.1 x 100 mm, 1.9 µm); Thermo guard column (2.1 x 5 mm; 3 µm)	0.1% formic acid in water/0.1% formic acid in acetonitrile	DC7	HRMS	Thermo Fisher Q-Exactive MS
2016	94	Yes	No	Yes	N/A	No	HILIC	SeQuant ZIC-HILIC (2.1 x 150 mm; 5 µm)	0.1% formic acid in acetonitrile/ 0.1% formic acid in water	none	MS	Agilent Technologies 6490 Triple Quad MS

2016	105	Yes	N/A	Yes	N/A	N/A	HILIC	Waters Acquity BEH amide (2.1 x 100 mm; 1.7 µm); Waters Acquity BEH amide (2.1 x 5 mm) pre-column	acetonitrile/ 0.1% formic acid in water	none	MS/MS	Waters Xevo TQ-S
2016	118	Yes	N/A	Yes	N/A	N/A	HILIC	Waters Acquity BEH amide (2.1 x 100 mm; 1.7 µm); pre-column Waters Acquity BEH amide (2.1 x 5 mm)	acetonitrile/ 0.1% formic acid in water	none	MS/MS	Waters Xevo TQ-S
2016	49	Yes	No	Yes	Yes	N/A	HILIC	Merck SeQuant ZIC-HILIC (2.1 x 150 mm; 5 µm); TSKgel amide-80 guard column (2 x 10 mm; 5 µm)	0.1% formic acid in water / 0.1% formic acid in acetonitrile	none	MS/MS	AB Sciex AP 5500 Qtrap Triple Quad MS
2016	5	Yes	N/A	N/A	Yes	N/A	HILIC	SeQuant ZIC-HILIC (2.1 x 150 mm; 5 µm); TSKgel amide-80 guard column (2 x 10 mm; 5 µm)	0.1% formic acid in water / 0.1% formic acid in acetonitrile	none	MS/MS	Sciex Qtrap 5500 AB

2016	119	Yes	N/A	Yes	N/A	N/A	HILIC	Tosoh TSKgel Amide-80 (2.0 x 250 mm; 5 µm)	50 mM formic acid in water/50 mM formic acid in 95% acetonitrile	none	MS/MS	Agilent 6430 MS
2016	120	Yes	N/A	Yes	N/A	N/A	RPLC	Waters Nova-Pak C18 (3.9 x 300 mm)	140 mM sodium acetate, 5.6 mM triethylamine (pH 5.2)/52% acetonitrile in water	AQC	FD	Waters 2475 Multi λ-Fluorescence Detector
2016	120	Yes	N/A	Yes	N/A	N/A	RPLC	Waters Acquity BEH C18 (2.1 x 150 mm; 1.7 µm)	20 mM ammonium formate in 0.2% formic acid/0.1% formic acid in acetonitrile	AQC	MS/MS	Waters Xevo TQ-S
2016	7	Yes	Yes	N/A	Yes	N/A	RPLC	Waters AccQ-Tag Ultra C18 (2.1 x 100 mm; 1.7 µm)	0.01% formic acid in 0.05% ammonia in water/0.01% formic acid in methanol	AQC	MS/MS	Waters Xevo TQ-S
2016	121	No	N/A	N/A	N/A	N/A	RPLC	Waters UltraTag C18 (2.1 mm x 100 mm; 1.7 µm)	0.1% formic acid in water / 0.1% formic acid in acetonitrile	AQC	MS/MS	Thermo Scientific Finnigan TSQ Quantum UltraAM
2016	105	Yes	N/A	Yes	N/A	N/A	RPLC	Waters Acquity BEH, 1.7 µm (2.1 x 100 mm); Waters C18 guard	0.1% formic acid in water / 0.1% formic acid in acetonitrile	DC	MS/MS	Waters Xevo MS

								column				
2016	85	No	N/A	N/A	N/A	No	RPLC	Waters Acquity BEH C18 (2.1 x 50 mm; 1.7 µm)	0.1% formic acid in water / 0.1% formic acid in methanol	DC	MS/MS	Waters Xevo TQ-S
2015	45	Yes	No	Yes	Yes	N/A	HILIC	Tosoh TSKgel amide-80 (2 x 250 mm; 5 µm)	50 mM formic acid in water/50 mM formic acid in acetonitrile	none	DMS-MS/MS	AB Sciex Qtrap 5500
2015	6	Yes	Yes	Yes	Yes	N/A	HILIC	SeQuant ZIC-HILIC (2.1 x 150 mm; 5µm); TSKgel amide-80 guard column (2 x 10 mm; 5 µm)	0.1% formic acid in water/0.1% formic acid in acetonitrile	none	MS/MS	AB Sciex Qtrap 5500
2015	122	No	No	N/A	Yes	N/A	HILIC	Tosoh TSKgel amide-80 (2 x 250 mm; 5 µm)	50 mM ammonium formate in water/50 mM ammonium formate in acetonitrile	none	MS/MS	AB Sciex API 4000 Qtrap/Agilent 6430
2015	123	Yes	N/A	Yes	N/A	N/A	RPLC	Phenomenex Kinetex C18 (2.1 x 100 mm; 1.7 µm)	0.1% formic acid in water/0.1% formic acid in acetonitrile	AQC	MS/MS	Thermo Scientific Finnigan TSQ Quantum UltraAM

2015	71	Yes	Yes	N/A	N/A	N/A	RPLC	Waters Acquity BEH C18 (2.1 x 150 mm; 1.7 µm)	20 mM ammonium formate in water (pH 5.0)/methanol	AQC	MS/MS	Waters Xevo TQ-S
2015	124	No	N/A	N/A	N/A	N/A	RPLC	Waters AccQ-Tag Ultra C18 (2.1 x 100 mm; 1.7 µm)	0.1% formic acid in water/0.1% formic acid in acetonitrile	AQC	MS/MS	Thermo Scientific Finnigan TSQ Quantum UltraAM
2015	7	Yes	N/A	Yes	N/A	N/A	RPLC	Waters AccQ-Tag Ultra C18 (2.1 x 100 mm; 1.7 µm)	0.01% formic acid and 0.05% ammonia in water/0.01 % formic acid in methanol	AQC	MS/MS	Waters Xevo TQ-S
2015	125	Yes	N/A	Yes	N/A	N/A	RPLC	Thermo Scientific Hypersil GOLD C18 (2.1 x 100 mm; 3 µm)	0.3% formic acid in water/0.3% formic acid in acetonitrile	AQC	MS/MS	Agilent G6410B Triple Quad
2015	78	Yes	N/A	N/A	N/A	Yes	RPLC	Waters Acquity BEH C18 (2.1 x 100 mm; 1.7µm)	0.01% formic acid and 0.05% ammonia in water/0.01 % formic acid in methanol	AQC	MS/MS	Waters Xevo TQ-S
2015	126	Yes	N/A	Yes	N/A	N/A	RPLC	Waters Acquity BEH (2.1 x 100 mm; 1.7 µm); Waters C18 guard column	0.1% formic acid in water / 0.1% formic acid in acetonitrile	DC	MS/MS	Waters Xevo MS

2015	127	Yes	Yes	N/A	N/A	N/A	RPLC	Thermo Hypesil GOLD C18 (2.1 x 100 mm; 1.9 µm); Thermo guard column (2.1 x 5 mm; 3 µm)	0.1% formic acid in water / 0.1% formic acid in acetonitrile	DC	HRMS	Thermo Fisher Q-Exactive MS
2015	128	Yes	Yes	N/A	N/A	N/A	RPLC	Waters AccQ-Tag Ultra C18 (2.1 x 100 mm; 1.7 µm)	0.1% formic acid in water/0.1% formic acid in acetonitrile	AQC	MS/MS	Thermo Scientific Finnigan TSQ Quantum UltraAM
2014	101	Yes	No	Yes	N/A	N/A	HILIC	SeQuant ZIC-HILIC (2.1 x 150 mm; 5 µm); TSKgel amide-80 guard column (2 x 10 mm; 5 µm)	0.1% formic acid in water / 0.1% formic acid in acetonitrile	none	MS/MS	Sciex Qtrap 5500 AB
2014	129	No	No	N/A	N/A	N/A	HILIC	Tosoh TSKgel Amide-80 (2.0 x 250 mm; 5 µm)	50 mM formic acid in water/50 mM formic acid in acetonitrile: water (95:5)	none	MS/MS	AB Sciex API4000 Qtrap
2014	130	No	N/A	N/A	N/A	No	HILIC	Merck Sequant ZIC-HILIC (2.1 x 150 mm; 3.5 µm)	0.1% formic acid in water / 0.1% formic acid in acetonitrile	None	MS	Thermo TSQ Quantum Discovery Max

2014	131	Yes	Yes	Yes	N/A	N/A	RPLC	Kromasil-Thermohypersil C8 (4.6 x 250 mm)	not specified	AQC	FD	Waters 2475 Multi λ -Fluorescence Detector
2014	131	Yes	Yes	Yes	N/A	N/A	RPLC	Thermo Hypurity C18 (2.5 x 250 mm; 5 μ m)	0.5 g/L ammonium formate in water/0.5 g/L ammonium formate in methanol	AQC	MS/MS	Finnigan LCQ Classic Ion Trap MS
2014	132	Yes	N/A	Yes	N/A	N/A	RPLC	Waters AccQ Tag Ultra (2.1 x 100 mm)	0.1% formic acid in water/0.1% formic acid in acetonitrile	AQC	MS/MS	Thermo Scientific Finnigan TSQ Quantum UltraAM
2014	133	Yes	N/A	Yes	N/A	N/A	RPLC	Waters AccQTag Ultra column (2.1 x 100 mm)	0.01% formic acid in 0.05% ammonia / 0.01% formic acid in methanol	AQC	MS/MS	Waters Xevo TQ-MS
2014	76	Yes	N/A	Yes	N/A	N/A	RPLC	Waters Nova-Pak C18 (3.9 x 300 mm)	140 mM sodium acetate, 5.6 mM triethylamine (pH 5.7)/52% acetonitrile in water	AQC	FD	Waters 2475 Multi λ -Fluorescence Detector

2014	76	Yes	N/A	Yes	N/A	N/A	RPLC	Waters BEH C18 (2.1 x 150 mm; 1.7 µm)	20 mM ammonium formate with 0.2% formic acid/0.1% formic acid in acetonitrile	AQC	MS/MS	Waters Xevo TQ-S
2014	93	Yes	Yes	Yes	N/A	N/A	RPLC	Agilent Eclipse XDB-C18 (4.6 x 100 mm; 5 µm)	28% acetonitrile in water with 0.1% ammonium acetate	AQC	MS/MS	Agilent
2014	24	Yes	Yes	N/A	Yes	N/A	RPLC	Waters AccQ-Tag Ultra C18 (2.1 x 100 mm; 1.7 µm)	acetonitrile /formic acid	AQC	MS/MS	Thermo TSQ Vantage
2014	134	Yes	Yes	N/A	N/A	N/A	RPLC	Phenomenex AAA-MS (2.0 x 250 mm; 4 µm)	10 mM ammonium formate in water/10 mM ammonium formate in methanol	PCF	MS/MS	Waters Micromass Quattro Micro
2014	135	Yes	N/A	Yes	N/A	N/A	RPLC	Phenomenex AAA-MS (2.0 x 250 mm; 4 µm)	10 mM ammonium formate in methanol	PCF	MS/MS	Waters Micromass Quattro Micro
2013	136	Yes	Yes	N/A	N/A	N/A	HILIC	Merck Sequant ZIC-HILIC (2.1 x 150 mm; 3.5 µm)	0.1% formic acid in water / 0.1% formic acid in acetonitrile	None	MS	Thermo TSQ Quantum Discovery Max

2013	137	Yes	Yes	Yes	N/A	N/A	RPLC	Kromasil-Thermohypersil C8 (4.6 x 250 mm)	140 mM ammonium acetate, 5.6 mM triethylamine (pH 5.7) in water/52% acetonitrile in water	AQC	FD	Waters 2475 Multi-Fluorescence Detector
2013	137	Yes	Yes	Yes	N/A	N/A	RPLC	Thermo Hypurity C18 (2.5 x 250 mm; 5 µm)	0.5 g/L ammonium formate in water/0.5 g/L ammonium formate in methanol	AQC	MS/MS	Finnigan LCQ Classic Ion Trap MS
2013	138	Yes	N/A	Yes	N/A	N/A	RPLC	Waters AccQTag Ultra column (2.1 x 100 mm)	0.1% formic acid in water / 0.1% formic acid in acetonitrile	AQC	MS/MS	Thermo Scientific Finnigan TSQ Quantum UltraAM
2013	139	Yes	N/A	N/A	N/A	N/A	RPLC	Waters C18 AccQ-Tag Ultra (2.1 x 100 mm)	0.1% formic acid in water/0.1% formic acid in acetonitrile	AQC	MS/MS	Finnigan TSQ Quantum Ultra AM
2013	140	Yes	Yes	N/A	N/A	N/A	RPLC	Agilent Bonus RP Rapid Resolution High Throughput (2.1 x 100 mm, 1.8 µm)	acetonitrile/ formic acid	AQC	MS/MS	Thermo TSQ Vantage

2013	53	Yes	N/A	Yes	N/A	N/A	RPLC	Waters Acquity BEH (2.1 x 100 mm; 1.7 µm); Waters C18 guard column	0.1% formic acid in water / 0.1% formic acid in acetonitrile	DC	MS/MS	Waters Xevo MS
2013	141	Yes	N/A	Yes	N/A	N/A	RPLC	Phenomenex AAA-MS (2.0 x 250 mm; 4 µm)	10 mM ammonium formate in methanol	PCF	MS/MS	Thermo Scientific Finnigan TSQ Quantum UltraAM
2012	60	Yes	Yes	N/A	N/A	N/A	RPLC	Waters NovaPak C18 (3.9 x 300 mm; 4 µm)	140 mM sodium acetate and 5.6 mM triethylamine in water (pH 5.2)/acetonitrile/water	AQC	FLD	Agilent 1100
2012	60	No	No	N/A	N/A	N/A	RPLC	Agilent Zorbax Eclipse AAA (4.6 x 75 mm; 3.5 µm)	0.1% formic acid in acetonitrile/ 0.1% formic acid in water	AQC	MS/MS	Agilent 6401A QQQ
2012	60	No	No	N/A	N/A	N/A	HILIC	Sequant ZIC-HILIC (2.1 x 150 mm; 5 µm)	0.1% formic acid in acetonitrile/ 0.1% formic acid in water	none	MS/MS	Agilent 6401A QQQ
2012	95	No	No	N/A	N/A	N/A	HILIC	Tosoh TSK-gel Amide-80 (2.0 x 250 mm; 5 µm)	acetonitrile /formic acid	None	MS/MS	API 4000 Qtrap MS

2012	142	Yes	N/A	Yes	N/A	N/A	RPLC	Waters Nova-Pak C18 (3.9 x 300 mm)	140 mM sodium acetate, 5.6 mM triethylamine (pH 5.2)/60% acetonitrile	AQC	FLD	Waters 2487 Dual-I FLD
2012	142	Yes	N/A	Yes	N/A	N/A	RPLC	Waters AccQTag Ultra (2.1 x 100 mm)	0.1% formic acid in water / 0.1% formic acid in acetonitrile	AQC	MS/MS	Thermo Scientific Finnigan TSQ Quantum UltraAM
2012	143	Yes	Yes	N/A	N/A	N/A	RPLC	Waters Nova-Pak C18 (3.9 x 300 mm)	140 mM sodium acetate, 5.6 mM triethylamine (pH 5.2)/60% acetonitrile	AQC	FLD	Merck Hitachi L-7480 FLD
2012	46	Yes	Yes	Yes	N/A	N/A	RPLC	Agilent Bonus RP Rapid Resolution High Throughput (2.1 x 100 mm; 1.8 µm)	acetonitrile/ formic acid	AQC	MS/MS	Thermo TSQ Vantage
2012	144	Yes	Yes	N/A	N/A	N/A	RPLC	Waters Nova-Pak C18 (3.9 x 300 mm)	140 mM sodium acetate, 5.6 mM triethylamine/65% methanol in water	AQC	FLD	Merck Hitachi L-7480 FLD
2012	145	Yes	N/A	Yes	N/A	N/A	RPLC	Waters Xbridge C18 (4.6 x 150 mm; 3.5 µm)	Waters AccQ-Tag Ultra Eluents A& B	AQC	MS/MS	Agilent G410A Triple Quad

2011	146	No	N/A	N/A	N/A	N/A	ESI	NA	0.1% formic acid/acetone nitrile	none	MS/MS	Sciex API 5000 Triple Quad
2011	146	No	N/A	N/A	N/A	N/A	GC	Fused silica (0.25 x 300 mm; 0.25mm) coated with DB-5	Helium Gas	ECF	MS	Shimadzu CLASS17A-QP5000
2011	51	Yes	Yes	N/A	N/A	N/A	RPLC	Phenomene x AAA-MS (2.0 x 250 mm; 4 µm)	10 mM ammonium formate in water/10 mM ammonium formate in methanol	PCF	MS	Shimadzu 2010EV
2011	68	Yes	N/A	N/A	N/A	N/A	RPLC	Phenomene x AAA-MS (2.0 x 250 mm; 4 µm)	10 mM ammonium formate in water/10 mM ammonium formate in methanol	PCF	MS	Shimadzu 2010EV
2011	147	Yes	Yes	N/A	N/A	N/A	RPLC	Phenomene x AAA-MS (2.0 x 250 mm; 4 µm)	10 mM ammonium formate in methanol	PCF	MS/MS	Thermo Scientific Finnigan TSQ Quantum UltraAM
2010	148	No	N/A	N/A	N/A	No	GCxGC	Restek RTX- 5MS (60 m x 0.25 mm; 0.25 µm)	helium gas	ECF	MS	HP 5971A GC/MS

2010	149	No	No	N/A	N/A	N/A	HILIC	Phenomene x Luna C18(2) (4.60 x 250 mm; 5 mm)	2 mM ammonium formate (pH 3) in water/2 mM ammonium formate (pH 3) in acetonitrile/ water (90/10; v/v)	None	MS/MS	AB API 365
2010	150	No	No	N/A	N/A	N/A	HILIC	SeQuant ZIC-HILIC, 5 um (2.1 x 150 mm; 5 µm)	acetonitrile and 60 mM formic acid 60/40 (v/v), isocratic flow	None	MS/MS	Thermo Finnigan HPLC, Finnigan LCQ Deca XP Plus
2010	150	Yes	Yes	N/A	N/A	N/A	RPLC	Waters C18 AccQ-Tag Ultra (2.1 x 100 mm)	acetonitrile and 0.05% TFA; 70/30 (v/v), isocratic flow	AQC	MS/MS	Finnigan TSQ Quantum Ultra AM
2010	150	No	No	N/A	N/A	N/A	HILIC	Tosoh TSKgel amide-80 (2.0 x 250 mm; 5 µm)	acetonitrile and 0.05% TFA; 70/30 (v/v), isocratic flow	none	MS/MS	Micromass Quattro Ultima Pt
2010	66	Yes	Yes	Yes	N/A	N/A	RPLC	Thermo Hypersil GOLD (2.1 x 100 mm; 3 µm)	acetonitrile/ formic acid	AQC	MS/MS	AB API 2000 Triple Quadrupole

2010	74	Yes	Yes	Yes	Yes	N/A	RPLC	Thermo Hypersil GOLD (2.1 x 100 mm; 3 µm)	acetonitrile/formic acid	AQC	MS/MS	AB API 2000 Triple Quadrupole
2010	27	Yes	Yes	Yes	N/A	N/A	RPLC	Waters Nova-Pak C18 (3.9 x 300 mm)	140 mM sodium acetate, 5.6 mM triethylamine (pH 5.2)/60% acetonitrile	AQC	FLD	Waters 2487 Dual-FLD
2010	27	Yes	Yes	Yes	N/A	N/A	RPLC	Thermo Hypersil GOLD (2.1 x 100 mm; 3 µm)	acetonitrile/formic acid	AQC	MS/MS	Thermo TSQ Quantum Discovery Max
2009	151	No	N/A	N/A	N/A	No	GC	Restek film RTX-5MS (60 m x 0.25 mm)	helium gas	ECF	MS	HP 5971A GC/MS
2009	152	Yes	Yes	N/A	N/A	N/A	HILIC	SeQuant ZIC-HILIC (2.1 x 150 mm; 5 µm)	acetonitrile/formic acid	None	MS/MS	Agilent G6410A
2009	59	Yes	N/A	Yes	N/A	N/A	RPLC	Waters AccQTag Ultra (2.1 x 100 mm)	0.1% formic acid in water/0.1% formic acid in acetonitrile	AQC	MS/MS	Thermo TSQ Quantum Discovery Max
2009	59	Yes	N/A	Yes	N/A	N/A	RPLC	Waters AccQTag Ultra (2.1 x 100 mm)	Waters AccQ-Tag Ultra Eluents A& B	AQC	MS	Waters EMD 1000 Single Quad

2009	59	Yes	N/A	Yes	N/A	N/A	RPLC	Waters Nova-Pak C18 column (3.9 x 300 mm)	140 mM sodium acetate, 5.6 mM triethylamine (pH 5.2)/60% acetonitrile	AQC	FLD	Waters 2487 Dual-I FLD,
2009	59	Yes	N/A	Yes	N/A	N/A	RPLC	Waters AccQTag Ultra (2.1 x 100 mm)	Waters AccQ-Tag Ultra Eluents A&B	AQC	UV	Acquity UV Detector
2009	153	Yes	Yes	N/A	N/A	N/A	RPLC	Thermo Hypersil GOLD (2.1 x 100 mm; 3 µm)	0.1% formic acid in water/0.1% formic acid in acetonitrile	AQC	MS/MS	Thermo TSQ Quantum Discovery Max
2009	154	Yes	Yes	N/A	N/A	N/A	RPLC	2 serially coupled Chromolith Perform RP-18e (3 mm)	20 mM sodium acetate (pH 5.5)/50/50 acetonitrile/water	AQC	FLD	Hitachi LaChrom FLD L-7485
2009	67	Yes	Yes	N/A	N/A	N/A	RPLC	Waters AccQTag Ultra (2.1 x 100 mm)	0.1% formic acid in water/0.1% formic acid in acetonitrile	AQC	MS	Waters EMD 1000 Single Quad
2009	153	Yes	Yes	N/A	N/A	N/A	RPLC	Waters Nova-Pak C18 (3.9 x 300 mm)	140 mM sodium acetate, 5.6 mM triethylamine (pH 5.2)/60% acetonitrile	AQC	FLD	Waters 2487 Dual-I Fluorescence Detector, Waters 2488 UV detector
2009	29	Yes	N/A	N/A	N/A	Yes	RPLC	Thermo Hypersil GOLD (2.1 x 100 mm; 3	0.1% formic acid in water/0.1% formic acid	AQC	MS/MS	Thermo TSQ Quantum Discovery

								µm)	in acetonitrile			Max
2009	29	Yes	N/A	N/A	N/A	Yes	RPLC	Waters Nova-Pak C18 (3.9 x 300 mm)	140 mM sodium acetate, 5.6 mM triethylamine (pH 5.2)/60% acetonitrile	AQC	FLD	Waters 2487 Dual-FLD, Waters 2488 UV Detector
2009	155	Yes	Yes	N/A	N/A	N/A	RPLC	Waters AccQTag Ultra (2.1 x 100 mm)	0.1% formic acid in water/0.1% formic acid in acetonitrile	AQC	MS/MS	Thermo Finnigan Ultra AM
2009	83	Yes	Yes	N/A	N/A	N/A	RPLC	Waters AccQTag Ultra (2.1 x 100 mm)	0.1% formic acid in water/0.1% formic acid in acetonitrile	AQC	MS/MS	Thermo Finnigan Triple Quadrupole MS (model unspecified)
2009	156	No	No	N/A	N/A	N/A	RPLC	Supelco LC-18 (4.6 x 150 mm; 5 µm) with Zorbax C18 guard (4.6 x 12.5 mm; 5µm)	0.1 M ammonium formate (pH 4.0)/acetonitrile	FMOG	FD	Agilent 1100 LC-FLD
2009	157	Yes	N/A	Yes	N/A	No	RPLC	Agilent XDB C18 (2.1 x 50 mm; 1.8 µm)	10 mM formic acid/10 mM formic acid in acetonitrile	iTRAQ	MS/MS	Sciex API4000
2009	158	Yes	N/A	N/A	N/A	N/A	RPLC	Waters Spherisorb ODS2 (4.6 x 250 mm; 5 µm)	acetonitrile /formic acid	None	MS	Thermo LCQ DecaXP Plus Ion Trap

2008	75	Yes	Yes	N/A	N/A	N/A	GC	ZB-AAA GC (10 m x 0.25 mm)	helium gas	ECF	EI-MS	Thermo Finnigan Trace MS Plus
2008	65	No	No	N/A	N/A	N/A	HILIC	Tosoh TSK-gel Amide-80 (2.0 x 250; 5 µm)	acetonitrile/water	None	MS	Shimadzu LCMS 2010A
2008	54	No	No	N/A	N/A	N/A	HILIC	SeQuant ZIC-HILIC (2.1 x 50; 5 µm/2.1 x 150; 5 µm)	acetonitrile/60 mM formic acid	None	MS/MS	Micromass Quattro Ultima or API 4000 QTRAP
2008	58	Yes	Yes	N/A	N/A	N/A	RPLC	Waters AccQTag Ultra (2.1 x 100 mm)	0.1% formic acid in water/0.1% formic acid in acetonitrile	AQC	MS	Waters EMD 1000 Single Quad
2008	58	Yes	Yes	N/A	N/A	N/A	RPLC	Waters AccQTag Ultra (2.1 x 100 mm)	Waters AccQ-Tag Ultra Eluents A& B	AQC	UV	Acquity UV Detector
2008	58	Yes	Yes	N/A	N/A	N/A	RPLC	Waters Nova-Pak C18 (3.9 x 300 mm)	140 mM sodium acetate, 5.6 mM triethylamine (pH 5.2)/60% acetonitrile	AQC	FLD	Waters 2487 Dual-I FLD
2008	58	Yes	Yes	N/A	N/A	N/A	RPLC	Thermo Hypersil GOLD (2.1 x 100 mm; 3 µm)	0.1% formic acid in water/0.1% formic acid in acetonitrile	AQC	MS/MS	Thermo TSQ Quantum Discovery Max

2008	159	Yes	Yes	N/A	N/A	N/A	RPLC	Thermo Hypersil GOLD (2.1 x 100 mm; 3 µm)	0.1% formic acid in water/0.1% formic acid in acetonitrile	AQC	MS/MS	Thermo TSQ Quantum Discovery Max
2008	159	Yes	Yes	N/A	N/A	N/A	RPLC	Waters Nova-Pak C18 (3.9 x 300 mm)	140 mM sodium acetate, 5.6 mM triethylamine (pH 5.2)/60% acetonitrile	AQC	FLD	Waters 2487 Dual-FLD, Waters 2488 UV Detector
2007	52	Yes	Yes	N/A	N/A	N/A	IEX	Hitachi Reaction with Li-form resin & Ion Exchange ammonia filter	proprietary eluents (Hitachi)	Ninhydrin	FLD	Hitachi Amino Acid Analyzer
2007	52	Yes	Yes	N/A	N/A	N/A	RPLC	Waters AccQTag Ultra (2.1 x 100 mm)	0.1% formic acid in water/0.1% formic acid in acetonitrile	AQC	MS/MS	Thermo TSQ Quantum Discovery Max
2007	52	Yes	Yes	N/A	N/A	N/A	RPLC	Waters AccQTag Ultra (2.1 x 100 mm)	Waters AccQ-Tag Ultra Eluents A& B	AQC	MS	Waters EMD 1000 Single Quad
2007	52	Yes	Yes	N/A	N/A	N/A	RPLC	Waters Nova-Pak C18 (3.9 x 300 mm)	140 mM sodium acetate, 5.6 mM triethylamine (pH 5.2)/60% acetonitrile	AQC	FLD	Waters 2487 Dual-FLD,

2007	52	Yes	Yes	N/A	N/A	N/A	RPLC	Waters AccQTag Ultra (2.1 x 100 mm)	Waters AccQ-Tag Ultra Eluents A& B	AQC	UV	Acquity UV Detector
2006	23	Yes	N/A	Yes	N/A	N/A	RPLC	Waters Nova-Pak C18 (3.9 x 300 mm)	140 mM sodium acetate, 5.6 mM triethylamine (pH 5.2)/60% acetonitrile	AQC	MS	Agilent SL Single Quad, DAD
2006	23	Yes	N/A	Yes	N/A	N/A	RPLC	Waters Nova-Pak C18 (3.9 x 300 mm)	140 mM sodium acetate, 5.6 mM triethylamine (pH 5.2)/60% acetonitrile	AQC	FLD	Waters 2487 Dual-I FLD, Waters 2488 UV Detector
2005	22	Yes	Yes	Yes	N/A	N/A	RPLC	Waters Nova-Pak C18 (3.9 x 300 mm)	140 mM sodium acetate, 5.6 mM triethylamine (pH 5.2)/60% acetonitrile	AQC	MS	Agilent SL Single Quad, DAD
2005	22	Yes	Yes	Yes	N/A	N/A	RPLC	Waters Nova-Pak C18 (3.9 x 300 mm)	140 mM sodium acetate, 5.6 mM triethylamine (pH 5.2)/60% acetonitrile	AQC	FLD	Waters 2487 Dual-I FLD, Waters 2488 UV Detector

2005	77	No	N/A	N/A	N/A	No	RPLC	Unspecified	Unspecified	FMOc	FLD	Unspecified
2004	28	Yes	N/A	N/A	N/A	Yes	RPLC	Waters Nova-Pak C18 (3.9 x 300 mm)	140 mM sodium acetate, 5.6 mM triethylamine (pH 5.2)/60% acetonitrile	AQC	MS	Agilent SL Single Quad, DAD
2004	160	Yes	Yes	Yes	N/A	Yes	RPLC	Waters Nova-Pak C18 (3.9 x 300 mm)	140 mM sodium acetate, 5.6 mM triethylamine (pH 5.2)/60% acetonitrile	AQC	FLD	Waters 2487 Dual-FLD, Waters 2488 UV Detector
2004	160	Yes	Yes	Yes	N/A	Yes	RPLC	Waters Symmetry	acetonitrile/water	AQC	MS, DAD	Agilent SL Single Quad, DAD
2003	20	Yes	Yes	Yes	N/A	Yes	RPLC	Waters SymmetryShield RP-18	acetonitrile/water	AQC	MS / UV	Agilent SL Single Quad, DAD
2003	161	Yes	N/A	Yes	N/A	N/A	RPLC	Waters Nova-Pak C18 (3.9 x 300 mm)	140 mM sodium acetate, 5.6 mM triethylamine (pH 5.2)/60% acetonitrile	AQC	FLD	Waters 2487 Dual-FLD,

2003	21	Yes	N/A	Yes	N/A	N/A	RPLC	Waters Nova-Pak C18 (3.9 x 300 mm)	140 mM sodium acetate, 5.6 mM triethylamine (pH 5.2)/60% acetonitrile	AQC	FLD	Waters 2487 Dual-I FLD,
------	----	-----	-----	-----	-----	-----	------	------------------------------------	---	-----	-----	-------------------------

Table S2. Comparison of non-chromatography method

s used for detection of BMAA. ¹Includes cyanobacterial cultures grown *in-vitro*, and cyanobacterial samples collected from natural habitats. ²Includes BMAA bioaccumulated in higher trophic level organisms such as traditional Chamorro dietary items such as cycad flour and flying foxes and seafoods such as mussels, lobster, fish and crab. ³Phytoplankton include diatoms and dinoflagellates. ⁴Includes human brain tissue and urine. Studies that did not analyze the specified sample type are labelled N/A.

Year	Reference	BMAA Detected?	BMAA Detected in Cyanobacteria ¹ ?	BMAA Detected in Bioaccumulated Sources ² ?	BMAA Detected in Phytoplankton or Zooplankton ³ ?	BMAA Detected in Human Tissues/Waste ⁴ ?	Separation	Separation Device	Elution Buffer(s)/ BGE	Derivative	Detection Method	Detector
2018	104	Yes	Yes	N/A	N/A	N/A	ELISA	Abraxis BMAA ELISA kit (PN 520040)	N/A	none	Spectrophotometer	Sunrise Absorbance Reader
2018	103	Yes	N/A	Yes	N/A	N/A	Chip-based CZE	908 Devices ZipChip HS Chip	2% formic acid in 50% methanol in water	none	MS/MS	Q Exactive HF MS (Thermo Fisher)
2018	162	Yes	N/A	N/A	N/A	N/A	ELISA	Abraxis BMAA ELISA kit (PN 520040)	N/A	none	Spectrophotometer	BioTek ELx800 Absorbance Microplate Reader
2017	80	Yes	N/A	N/A	N/A	Yes	ELISA	Abraxis BMAA ELISA kit (PN 520040)	N/A	none	Spectrophotometer	BioTek Power Wave Spectrometer

2017	163	Yes	No	Yes	N/A	N/A	CE	Agilent G1600A CE System with custom interface	5 M formic acid with 10% acetonitrile	none	MS/MS	Sciex API-4000 MS
2016	97	No	N/A	No	N/A	N/A	ELISA	Abraxis BMAA ELISA kit (PN 520040)	N/A	none	Spectrophotometer	Microplate ELISA Photometer
2016	92	Yes	Yes	N/A	N/A	N/A	ELISA	Abraxis BMAA ELISA kit (PN 520040)	N/A	none	Spectrophotometer	Biotek Synergy HT
2013	57	Yes	N/A	N/A	N/A	N/A	ELISA	Abraxis BMAA ELISA kit (PN 520040)	N/A	none	MTP Reader	Biotek Synergy HT
2011	56	Yes	Yes	N/A	N/A	N/A	CE	fused-silica (50 cm x 75 µm)	5 mM sodium tetraborate (pH 9)	none	UV	Beckman Coulter P/ACE MDQ system
2009	55	Yes	Yes	N/A	N/A	N/A	N/A	N/A	N/A	none	¹ H NMR	Bruker DPX 300

References

- 1 H. W. Paerl and T. G. Otten, *Microb. Ecol.*, 2013, **65**, 995–1010.

- 2 W. W. Carmichael and G. L. Boyer, *Harmful Algae*, 2016, **54**, 194–212.
- 3 M. Rutkowska, J. Piotka-Wasyłka, T. Majchrzak, W. Wojnowski, H. Mazur-Marzec and J. Namiesnik, *Trends Anal. Chem.*, 2019, **112**, 112–122.
- 4 K. Manolidi, T. M. Triantis, T. Kaloudis and A. Hiskia, *J. Hazard. Mater.*, 2019, **365**, 346–365.
- 5 D. Réveillon, V. Séchet, P. Hess and Z. Amzil, *Harmful Algae*, 2016, **58**, 45–50.
- 6 D. Réveillon, E. Abadie, V. Séchet, E. Masseret, P. Hess and Z. Amzil, *Mar. Environ. Res.*, 2015, **110**, 8–18.
- 7 S. Lage, A. Burian, U. Rasmussen, P. R. Costa, H. Annadotter, A. Godhe and S. Rydberg, *Environ. Sci. Pollut. Res.*, 2016, **23**, 338–350.
- 8 N. Torbick, B. Ziniti, E. Stommel, E. Linder, A. Andrew, T. Caller, J. Haney, W. Bradley, P. L. Henegan and X. Shi, *Neurotox. Res.*, 2018, **33**, 199–212.
- 9 M. G. Whiting, *Econ. Bot.*, 1963, **17**, 270–302.
- 10 A. Arnold, D. C. Edgren and V. S. Palladino, *J. Nerv. Ment. Dis.*, 1953, 117, 135–139.
- 11 G. L. Laqueur, O. Mickelsen, M. G. Whiting and L. T. Kurland, *Carcinogenic properties of nuts from cycas circinalis L. indigenous to guam*, 1963, vol. 31.
- 12 D. Reed, D. Labarthe, K. M. Chen and R. Stallones, *Am. J. Epidemiol.*, 1987, **125**, 92–100.
- 13 A. Vega and E. A. Bell, *Phytochemistry*, 1967, **6**, 759–762.
- 14 A. Vega, E. a. Bell and P. B. Nunn, *Phytochemistry*, 1968, **7**, 1885–1887.
- 15 P. S. Spencer, P. B. Nunn, J. Hugon, A. C. Ludolph, M. Stephen, D. N. Roy, R. C. Robertson, P. S. Spencer, P. B. Nunn, J. Hugon, A. C. Ludolph, S. M. Ross, D. N. Roy and R. C. Robertson, 1987, **237**, 517–522.
- 16 M. Duncan, I. Kopin, R. Garruto, L. Lavine and S. Markey, *Lancet*, 1988, **332**, 631–632.
- 17 P. A. Cox and O. W. Sacks, *Neurology*, 2002, **58**, 956–959.
- 18 C. S. Monson, S. A. Banack and P. A. Cox, *Conserv. Biol.*, 2003, **17**, 678–686.
- 19 S. J. Murch, P. A. Cox and S. A. Banack, *Proc. Natl. Acad. Sci. U. S. A.*, 2004, **101**, 12228–31.
- 20 P. A. Cox, S. A. Banack and S. J. Murch, *Proc. Natl. Acad. Sci.*, 2003, **100**, 13380–13383.
- 21 S. A. Banack and P. A. Cox, *Neurology*, 2003, **61**, 387–389.
- 22 P. A. Cox, S. A. Banack, S. J. Murch, U. Rasmussen, G. Tien, R. R. Bidigare, J. S. Metcalf, L. F. Morrison, G. A. Codd and B. Bergman, *Proc. Natl. Acad.*

Sci. U. S. A., 2005, **102**, 5074–5078.

- 23 S. A. Banack, S. J. Murch and P. A. Cox, *J. Ethnopharmacol.*, 2006, **106**, 97–104.
- 24 L. Jiang, N. Kiselova, J. Rosén and L. L. Ilag, *Sci. Rep.*, 2014, **4**, 6931.
- 25 D. G. Beach, E. S. Kerrin, S. D. Giddings, M. A. Quilliam and P. McCarron, *Sci. Rep.*, 2018, **8**, 1–11.
- 26 T. C. Baker, F. J. M. Tymms and S. J. Murch, *Neurotox. Res.*, 2018, **33**, 43–54.
- 27 L. E. Brand, J. Pablo, A. Compton, N. Hammerschlag and D. C. Mash, *Harmful Algae*, 2010, **9**, 620–635.
- 28 S. J. Murch, P. A. Cox, S. A. Banack, J. C. Steele and O. W. Sacks, *Acta Neurol. Scand.*, 2004, **110**, 267–269.
- 29 J. Pablo, S. A. Banack, P. A. Cox, T. E. Johnson, S. Papapetropoulos, W. G. Bradley, A. Buck and D. C. Mash, *Acta Neurol. Scand.*, 2009, **120**, 216–225.
- 30 S. D. Rao, S. A. Banack, P. A. Cox and J. H. Weiss, *Exp. Neurol.*, 2006, **201**, 244–252.
- 31 P. S. Spencer, P. B. Nunn, J. Hugon, A. C. Ludolph, S. M. Ross, D. N. Roy and R. C. Robertson, *Science (80-.)*, 1987, **237**, 517–522.
- 32 W. B. Glover, D. C. Mash and S. J. Murch, *Amino Acids*, 2014, **46**, 2553–2559.
- 33 R. A. Dunlop, P. A. Cox, S. A. Banack and K. J. Rodgers, *PLoS One*, 2013, **8**, 1–8.
- 34 P. A. Cox, D. A. Davis, D. C. Mash, J. S. Metcalf and S. A. Banack, *Proc. R. Soc. B Biol. Sci.*, 2016, **283**, 1–10.
- 35 D. A. Davis, K. Mondo, E. Stern, A. K. Annor, S. J. Murch, T. M. Coyne, L. E. Brand, M. E. Niemeyer, S. Sharp, W. G. Bradley, P. A. Cox and D. C. Mash, *PLoS One*, 2019, **14**, 1–18.
- 36 O. Karlsson, C. Berg, E. B. Brittebo and N. G. Lindquist, *Pigment Cell Melanoma Res.*, 2009, **22**, 120–130.
- 37 M. Arif, S. F. Kazim, I. Grundke-Iqbal, R. M. Garruto and K. Iqbal, *Proc. Natl. Acad. Sci.*, 2014, **111**, 1144–1149.
- 38 A. Rauk, *J. Phys. Chem. B*, 2018, **122**, 4472–4480.
- 39 W. B. Glover, C. M. Liberto, W. S. McNeil, S. A. Banack, P. R. Shipley and S. J. Murch, *Anal. Chem.*, 2012, **84**, 7946–7953.
- 40 P. Diaz-Parga, J. J. Goto and V. V. Krishnan, *Neurotox. Res.*, 2018, **33**, 76–86.
- 41 S. A. Banack and S. J. Murch, *Neurotox. Res.*, 2018, **33**, 184–191.
- 42 S. A. Banack, J. S. Metcalf, Z. Spáčil, T. G. Downing, S. Downing, A. Long, P. B. Nunn and P. A. Cox, *Toxicon*, 2011, **57**, 730–738.

- 43 S. A. Banack, J. S. Metcalf, L. Jiang, D. Craighead, L. L. Ilag and P. A. Cox, *PLoS One*, 2012, **7**, 1–4.
- 44 S. L. Bishop, J. K. Kerkovius, F. Menard and S. J. Murch, *Neurotox. Res.*, 2018, **33**, 133–142.
- 45 D. G. Beach, E. S. Kerrin and M. A. Quilliam, *Anal. Bioanal. Chem.*, 2015, **407**, 8397–8409.
- 46 L. Jiang, B. Aigret, W. M. De Borggraeve, Z. Spacil and L. L. Ilag, *Anal. Bioanal. Chem.*, 2012, **403**, 1719–1730.
- 47 A. Li, Y. Hu, J. Song, S. Wang and L. Deng, *Toxicol.*, 2018, **151**, 129–136.
- 48 A. J. Foss, N. Chernoff and M. T. Aubel, *Toxicol.*, 2018, **152**, 150–159.
- 49 D. Réveillon, V. Séchet, P. Hess and Z. Amzil, *Toxicol.*, 2016, **110**, 35–46.
- 50 G. E. Kisby, D. N. Roy and P. S. Spencer, *J. Neurosci. Methods*, 1988, **26**, 45–54.
- 51 M. Esterhuizen-Londt, S. Downing and T. Downing, *Water SA*, 2011, **37**, 133–139.
- 52 S. A. Banack, H. E. Johnson, R. Cheng and P. A. Cox, *Mar. Drugs*, 2007, **5**, 180–196.
- 53 M. L. Salomonsson, A. Hansson and U. Bondesson, *Anal. Methods*, 2013, **5**, 4865–4874.
- 54 J. Rosén and K. E. Hellenäs, *Analyst*, 2008, **133**, 1785–1789.
- 55 S. Moura, M. de A. Ultramari, D. M. L. de Paula, M. Yonamine and E. Pinto, *Toxicol.*, 2009, **53**, 578–583.
- 56 M. S. Baptista, R. C. C. Cianca, V. R. Lopes, C. M. R. Almeida and V. M. Vasconcelos, *Toxicol.*, 2011, **58**, 410–414.
- 57 E. J. Faassen, W. Beekman and M. Lüring, *PLoS One*, 2013, **8**, 1–8.
- 58 H. E. Johnson, S. R. King, S. A. Banack, C. Webster, W. J. Callanaupa and P. A. Cox, *J. Ethnopharmacol.*, 2008, **118**, 159–165.
- 59 R. Cheng and S. A. Banack, *Amyotroph. Lateral Scler.*, 2009, **10**, 41–43.
- 60 E. J. Faassen, F. Gillissen and M. Lüring, *PLoS One*, 2012, **7**, 1–8.
- 61 M. W. Duncan, *Adv. Neurol.*, 1991, **56**, 301–310.
- 62 M. Duncan, I. Kopin, J. S. Crowley, S. M. Jones and S. P. Markey, *J. Anal. Toxicol.*, 1989, **13**, 169–175.
- 63 G. E. Kisby, M. Ellison and P. S. Spencer, *Neurology*, 1992, **42**, 1336–1340.

- 64 I. Khabazian, J. S. Bains, D. E. Williams, J. Cheung, J. M. B. Wilson, B. A. Pasqualotto, S. L. Pelech, R. J. Andersen, Y. T. Wang, L. Liu, A. Nagai, S. U. Kim, U. K. Craig and C. A. Shaw, *J. Neurochem.*, 2002, **82**, 516–528.
- 65 T. Kubo, N. Kato, K. Hosoya and K. Kaya, *Toxicol.*, 2008, **51**, 1264–1268.
- 66 Z. Spáčil, J. Eriksson, S. Jonasson, U. Rasmussen, L. L. Ilag and B. Bergman, *Analyst*, 2010, **135**, 127–132.
- 67 B. R. Roney, L. Renhui, S. A. Banack, S. Murch, R. Honegger and P. A. Cox, *Amyotroph. Lateral Scler.*, 2009, **10**, 44–49.
- 68 M. Esterhuizen-Londt and T. Downing, *Water SA*, 2011, **37**, 523–528.
- 69 B. Yan, Z. Liu, R. Huang, Y. Xu, D. Liu, T. F. Lin and F. Cui, *Anal. Chem.*, 2017, **89**, 10991–10998.
- 70 D. G. Beach, E. S. Kerrin, S. D. Giddings, M. A. Quilliam and P. McCarron, *Sci. Rep.*, 2018, **8**, 1–11.
- 71 W. B. Glover, T. C. Baker, S. J. Murch and P. N. Brown, *J. AOAC Int.*, 2015, **98**, 1559–1565.
- 72 AOAC, *AOAC Off. Methods Anal.*, 2013, 1–32.
- 73 D. Moher, A. Liberati, J. Tetzlaff, D. G. Altman, D. Altman, G. Antes, D. Atkins, V. Barbour, N. Barrowman, J. A. Berlin, J. Clark, M. Clarke, D. Cook, R. D'Amico, J. J. Deeks, P. J. Devereaux, K. Dickersin, M. Egger, E. Ernst, P. C. Gøtzsche, J. Grimshaw, G. Guyatt, J. Higgins, J. P. A. Ioannidis, J. Kleijnen, T. Lang, N. Magrini, D. McNamee, L. Moja, C. Mulrow, M. Napoli, A. Oxman, B. Pham, D. Rennie, M. Sampson, K. F. Schulz, P. G. Shekelle, D. Tovey and P. Tugwell, *J. Chinese Integr. Med.*, 2009, **7**, 889–896.
- 74 S. Jonasson, J. Eriksson, L. Berntzon, Z. Spáčil, L. L. Ilag, L.-O. Ronnevi, U. Rasmussen and B. Bergman, *Proc. Natl. Acad. Sci. U. S. A.*, 2010, **107**, 9252–9257.
- 75 M. Esterhuizen and T. G. A. Downing, *Ecotoxicol. Environ. Saf.*, 2008, **71**, 309–313.
- 76 K. Mondo, W. Broc Glover, S. J. Murch, G. Liu, Y. Cai, D. A. Davis and D. C. Mash, *Food Chem. Toxicol.*, 2014, **70**, 26–32.
- 77 T. J. Montine, K. Li, D. P. Perl and D. Galasko, *Neurology*, 2005, **65**, 768–769.
- 78 L. Berntzon, L. O. Ronnevi, B. Bergman and J. Eriksson, *Neuroscience*, 2015, **292**, 137–147.
- 79 D. Bhandari, B. A. Bowman, A. B. Patel, D. M. Chambers, V. R. De Jesús and B. C. Blount, *J. Chromatogr. B Anal. Technol. Biomed. Life Sci.*, 2018, **1083**, 86–92.
- 80 L. Bláhová, J. Kohoutek, E. Kadlecová, L. Kozáková and L. Bláha, *Toxicol.*, 2017, **133**, 48–57.

- 81 S. Downing, L. L. Scott, N. Zguna and T. G. Downing, *Toxins (Basel)*, 2018, **10**, 1–15.
- 82 S. A. Banack, T. G. Downing, Z. Spáčil, E. L. Purdie, J. S. Metcalf, S. Downing, M. Esterhuizen, G. A. Codd and P. A. Cox, *Toxicon*, 2010, **56**, 868–879.
- 83 D. Craighead, J. S. Metcalf, S. A. Banack, L. Amgalan, H. V. Reynolds and M. Batmunkh, *Amyotroph. Lateral Scler.*, 2009, **10**, 96–100.
- 84 A. Mlinarić, M. Horvat and V. Š. Smolčić, *Biochem. Medica*, 2017, **27**, 1–6.
- 85 J. P. Meneely, O. P. Chevallier, S. Graham, B. Greer, B. D. Green and C. T. Elliott, *Sci. Rep.*, 2016, **6**, 1–9.
- 86 M. W. Duncan, *J. Mass Spectrom.*, 2012, **47**, 795–809.
- 87 E. J. Faassen, *Toxins (Basel)*, 2014, **6**, 1109–1138.
- 88 D. S. S. Lim, B. E. Laval, G. Slater, D. Antoniadis, A. L. Forrest, W. Pike, R. Pieters, M. Saffari, D. Reid, D. Schulze-Makuch, D. Andersen and C. P. Mckay, *Fundam. Appl. Limnol.*, 2009, **173**, 329–351.
- 89 S. Downing, M. van de Venter and T. G. Downing, *Microb. Ecol.*, 2012, **63**, 149–156.
- 90 M. Monteiro, M. Costa, C. Moreira, V. M. Vasconcelos and M. S. Baptista, *J. Appl. Phycol.*, 2017, **29**, 879–888.
- 91 H. S. Yin, B. C. Li, Y. L. Zhou, H. Y. Wang, M. H. Wang and S. Y. Ai, *Biosens. Bioelectron.*, 2017, **96**, 106–112.
- 92 E. Pip, K. Munford and L. Bowman, *Environ. Pollut.*, 2016, **5**, 110–118.
- 93 Y. Jiao, Q. Chen, X. Chen, X. Wang, X. Liao, L. Jiang, J. Wu and L. Yang, *Sci. Total Environ.*, 2014, **468–469**, 457–463.
- 94 E. J. Faassen, M. G. Antoniou, W. Beekman-Lukassen, L. Blahova, E. Chernova, C. Christophoridis, A. Combes, C. Edwards, J. Fastner, J. Harmsen, A. Hiskia, L. L. Ilag, T. Kaloudis, S. Lopacic, M. Lüring, H. Mazur-Marzec, J. Meriluoto, C. Porojan, Y. Viner-Mozzini and N. Zguna, *Mar. Drugs*, 2016, **14**, 1–12.
- 95 A. Li, H. Fan, F. Ma, P. McCarron, K. Thomas, X. Tang and M. A. Quilliam, *Analyst*, 2012, **137**, 1210.
- 96 E. J. Faassen, in *Handbook of Cyanobacterial Monitoring and Cyanotoxin Analysis*, eds. J. Meriluoto, L. Spoof and G. A. Codd, John Wiley & Sons, West Sussex, 2017, pp. 439–446.
- 97 M. T. Clausi, V. Vita, M. Bruno, C. Franchino, G. Trifirò, M. P. Palumbo, F. Floridi and R. De Pace, *Int. J. Environ. Anal. Chem.*, 2016, **96**, 1290–1299.
- 98 W. J. Youden, *Anal. Chem.*, 1948, **20**, 1136–1140.
- 99 P. B. Nunn, P. O'Brien, L. D. Pettit and S. I. Pyburn, *J. Inorg. Biochem.*, 1989, **37**, 175–183.

- 100 J. Zurita, N. Zguna, R. Andrýs, A. Strzelczak, L. Jiang, G. Thorsen and L. L. Ilag, *Anal. Methods*, 2019, **11**, 432–442.
- 101 D. Réveillon, E. Abadie, V. Séchet, L. Brient, V. Savar, M. Bardouil, P. Hess and Z. Amzil, *Mar. Drugs*, 2014, **12**, 5441–5467.
- 102 E. S. Kerrin, R. L. White and M. A. Quilliam, *Anal. Bioanal. Chem.*, 2017, **409**, 1481–1491.
- 103 J. Beri, K. I. Kirkwood, D. C. Muddiman and M. S. Bereman, *Anal. Bioanal. Chem.*, 2018, **410**, 2597–2605.
- 104 H. Almuhtaram, Y. Cui, A. Zamyadi and R. Hofmann, *Toxins (Basel)*, 2018, **10**, 1–15.
- 105 J. Rosén, E. Westerberg, S. Schmiedt and K. E. Hellenäs, *Toxicon*, 2016, **109**, 45–50.
- 106 J. P. Violi, S. M. Mitrovic, A. Colville, B. J. Main and K. J. Rodgers, *Ecotoxicol. Environ. Saf.*, 2019, **172**, 72–81.
- 107 M. Esterhuizen-Londt and S. Pflugmacher, *Environ. Res.*, 2019, **169**, 357–361.
- 108 B. Li, S. Yu, G. Li, X. Chen, M. Huang, X. Liao, H. Li, F. Hu and J. Wu, *Chemosphere*, 2019, **219**, 997–1001.
- 109 J. S. Metcalf, S. Anne, J. T. Powell, F. J. M. Tymms, S. J. Murch, L. E. Brand and P. A. Cox, *Water Policy*, 2018, **20**, 919–932.
- 110 A. D. Jungblut, J. Wilbraham, S. A. Banack, J. S. Metcalf and G. A. Codd, *Eur. J. Phycol.*, 2018, **53**, 115–121.
- 111 A. D. Chatziefthimiou, E. J. Deitch, W. B. Glover, J. T. Powell, S. A. Banack, R. A. Richer, P. A. Cox and J. S. Metcalf, *Neurotox. Res.*, 2018, **33**, 143–152.
- 112 B. J. Main and K. J. Rodgers, *Neurotox. Res.*, 2018, **33**, 33–42.
- 113 L. L. Scott, S. Downing and T. Downing, *Toxicon*, 2018, **150**, 261–266.
- 114 P. Rzymiski, B. Poniedziałek, J. Mankiewicz-Boczek, E. J. Faassen, T. Jurczak, I. Gągała-Borowska, A. Ballot, M. Lürling and M. Kokociński, *Algal Res.*, 2017, **24**, 72–80.
- 115 A. C. Braga, S. Lage, M. Pacheco, S. Rydberg and P. R. Costa, *Mar. Environ. Res.*, 2017, **129**, 147–155.
- 116 J. S. Metcalf, D. Lobner, S. A. Banack, G. A. Cox, P. B. Nunn, P. B. Wyatt and P. A. Cox, *Amino Acids*, 2017, **49**, 1427–1439.
- 117 A. Roy-Lachapelle, M. Sollicec, M. F. Bouchard and S. Sauvé, *Toxins (Basel)*, 2017, **9**, 1–18.
- 118 J. Rosén, E. Westerberg, K. E. Hellenäs and M. L. Salomonsson, *Toxicon*, 2016, **121**, 105–108.
- 119 A. Li, J. Song, Y. Hu, L. Deng, L. Ding and M. Li, *Mar. Drugs*, 2016, **14**, 1–12.
- 120 N. Hammerschlag, D. Davis, K. Mondo, M. Seely, S. Murch, W. Brocglower, T. Divoll, D. Evers and D. C. Mash, *Toxins (Basel)*, 2016, **8**, 1–14.

- 121 A. D. Chatziefthimiou, J. S. Metcalf, W. B. Glover, S. A. Banack, S. R. Dargham and R. A. Richer, *Toxicon*, 2016, **114**, 75–84.
- 122 H. Fan, J. Qiu, L. Fan and A. Li, *Environ. Sci. Pollut. Res.*, 2015, **22**, 5943–5951.
- 123 S. A. Banack, T. Caller, P. Henegan, J. Haney, A. Murby, J. S. Metcalf, J. Powell, P. Alan and E. Stommel, *Toxins (Basel)*, 2015, **7**, 322–336.
- 124 R. Richer, S. A. Banack, J. S. Metcalf and P. A. Cox, *J. Arid Environ.*, 2015, **112**, 134–139.
- 125 R. Andrýs, J. Zurita, N. Zguna, K. Verschueren, W. M. De Borggraeve and L. L. Ilag, *Anal. Bioanal. Chem.*, 2015, **407**, 3743–3750.
- 126 M. L. Salomonsson, E. Fredriksson, A. Alfjorden, M. Hedeland and U. Bondesson, *Toxicol. Reports*, 2015, **2**, 1473–1481.
- 127 A. Roy-Lachapelle, M. Sollicec and S. Sauv e, *Anal. Bioanal. Chem.*, 2015, **407**, 5487–5501.
- 128 J. S. Metcalf, S. A. Banack, R. Richer and P. A. Cox, *J. Arid Environ.*, 2015, **112**, 140–144.
- 129 P. McCarron, A. C. Logan, S. D. Giddings and M. A. Quilliam, *Aquat. Biosyst.*, 2014, **10**, 1–7.
- 130 A. Combes, S. El Abdellaoui, J. Vial, E. Lagrange and V. Pichon, *Anal. Bioanal. Chem.*, 2014, **406**, 4627–4636.
- 131 M. A. Al-Sammak, K. D. Hoagland, D. Cassada and D. D. Snow, *Toxins (Basel)*, 2014, **6**, 488–508.
- 132 S. A. Banack, J. S. Metcalf, W. G. Bradley and P. A. Cox, *Toxicon*, 2014, **90**, 167–173.
- 133 S. Lage, P. R. Costa, T. Moita, J. Eriksson, U. Rasmussen and S. J. Rydberg, *Aquat. Toxicol.*, 2014, **152**, 131–138.
- 134 L. L. Scott, S. Downing, R. R. Phelan and T. G. Downing, *Toxicon*, 2014, **87**, 1–5.
- 135 S. Downing, V. Contardo-Jara, S. Pflugmacher and T. G. Downing, *Ecotoxicol. Environ. Saf.*, 2014, **101**, 51–58.
- 136 A. Combes, S. El Abdellaoui, C. Sarazin, J. Vial, A. Mejean, O. Ploux and V. Pichon, *Anal. Chim. Acta*, 2013, **771**, 42–49.
- 137 M. A. Al-Sammak, K. D. Hoagland, D. D. Snow and D. Cassada, *Toxicon*, 2013, **76**, 316–325.
- 138 E. Masseret, S. Banack, F. Boum di ne, E. Abadie, L. Brient, F. Pernet, R. Juntas-Morales, N. Pageot, J. Metcalf, P. Cox, W. Camu, G. Besson, V. Bonneterre, P. Couratier, M. Druet-Cabanac, E. Lagrange, B. Marin, A. Mejean and P. M. Preux, *PLoS One*, 2013, **8**, 1–10.
- 139 J. S. Metcalf, S. A. Banack, K. Kotut, L. Krienitz and G. A. Codd, *Chemosphere*, 2013, **90**, 835–839.
- 140 L. Jiang, E. Johnston, K. M.  berg, U. Nilsson and L. L. Ilag, *Anal. Bioanal. Chem.*, 2013, **405**, 1283–1292.
- 141 N. C. Field, J. S. Metcalf, T. A. Caller, S. A. Banack, P. A. Cox and E. W. Stommel, *Toxicon*, 2013, **70**, 179–183.

- 142 K. Mondo, N. Hammerschlag, M. Basile, J. Pablo, S. A. Banack and D. C. Mash, *Mar. Drugs*, 2012, **10**, 509–520.
- 143 R. C. Cervantes Cianca, M. S. Baptista, L. P. Da Silva, V. R. Lopes and V. M. Vasconcelos, *Toxicon*, 2012, **59**, 379–384.
- 144 R. C. Cervantes Cianca, M. S. Baptista, V. R. Lopes and V. M. Vasconcelos, *Amino Acids*, 2012, **42**, 2473–2479.
- 145 S. J. Christensen, T. K. Hemscheidt, H. Trapido-Rosenthal, E. A. Laws and R. R. Bidigare, *Limnol. Oceanogr. Methods*, 2012, **10**, 891–898.
- 146 K. de C. Mariotti, F. Barreto, G. C. Schmitt, I. Zancanaro, E. Dallegrove, R. B. Singer, M. B. Leal and R. P. Limberger, *Brazilian J. Pharm. Sci.*, 2011, **47**, 623–628.
- 147 S. Downing, S. A. Banack, J. S. Metcalf, P. A. Cox and T. G. Downing, *Toxicon*, 2011, **58**, 187–194.
- 148 T. E. Marler, L. R. Snyder and C. A. Shaw, *Toxicon*, 2010, **56**, 563–568.
- 149 T. Krüger, B. Mönch, S. Oppenhäuser and B. Luckas, *Toxicon*, 2010, **55**, 547–557.
- 150 A. Li, Z. Tian, J. Li, R. Yu, S. A. Banack and Z. Wang, *Toxicon*, 2010, **55**, 947–953.
- 151 L. R. Snyder, R. Cruz-Aguado, M. Sadilek, D. Galasko, C. A. Shaw and T. J. Montine, *Neurology*, 2009, **72**, 1360–1361.
- 152 E. J. Faassen, F. Gillissen, H. A. J. Zweers and M. Lrling, *Amyotroph. Lateral Scler.*, 2009, **10**, 79–84.
- 153 P. A. Cox, R. Richer, J. S. Metcalf, S. A. Banack, G. A. Codd and W. G. Bradley, *Amyotroph. Lateral Scler.*, 2009, **10**, 109–117.
- 154 J. Eriksson, S. Jonasson, D. Papaefthimiou, U. Rasmussen and B. Bergman, *Amino Acids*, 2009, **36**, 43–48.
- 155 T. A. Caller, J. W. Doolin, J. F. Haney, A. J. Murby, K. G. West, H. E. Farrar, A. Ball, B. T. Harris and E. W. Stommel, *Amyotroph. Lateral Scler.*, 2009, **10**, 101–108.
- 156 P. M. Scott, B. Niedzwiadek, D. F. K. Rawn and B. P.-Y. Lau, *J. Food Prot.*, 2009, **72**, 1769–1773.
- 157 M. M. Kushnir and J. Bergquist, *Eur J Mass Spectrom (Chichester, Eng)*, 2009, **15**, 439–443.
- 158 R. R. Bidigare, S. J. Christensen, S. B. Wilde and S. A. Banack, *Amyotroph. Lateral Scler.*, 2009, **10**, 71–73.
- 159 J. S. Metcalf, S. A. Banack, J. Lindsay, L. F. Morrison, P. A. Cox and G. A. Codd, *Environ. Microbiol.*, 2008, **10**, 702–708.
- 160 S. J. Murch, P. A. Cox and S. A. Banack, *Proc. Natl. Acad. Sci.*, 2004, **101**, 12228–12231.
- 161 S. A. Banack and P. A. Cox, *Bot. J. Linn. Soc.*, 2003, **143**, 165–168.

- 162 D. Wiltsie, A. Schnetzer, J. Green, M. Vander Borgh and E. Fensin, *Toxins (Basel)*., 2018, **10**, 1–23.
- 163 E. S. Kerrin, R. L. White and M. A. Quilliam, *Anal. Bioanal. Chem.*, 2017, **409**, 1481–1491.